

ARTICLE OPEN



Identification of novel *STAT5B* mutations and characterization of TCR β signatures in CD4⁺ T-cell large granular lymphocyte leukemia

Dipabarna Bhattacharya^{1,2,9}, Antonella Teramo^{3,9}, Vanessa Rebecca Gasparini^{3,9}, Jani Huuhtanen^{1,2,4,9}, Daehong Kim^{1,2}, Jason Theodoropoulos^{1,2,4}, Gianluca Schiavoni⁵, Gregorio Barilà³, Cristina Vicenzetto³, Giulia Calabretto³, Monica Facco³, Toru Kawakami⁶, Hideyuki Nakazawa⁶, Brunangelo Falini⁵, Enrico Tiacci⁵, Fumihiko Ishida⁷, Gianpietro Semenzato³, Tiina Kelkka^{1,2,10}, Renato Zambello^{3,10} and Satu Mustjoki^{1,2,8,10}✉

© The Author(s) 2022

CD4⁺ T-cell large granular lymphocyte leukemia (T-LGLL) is a rare subtype of T-LGLL with unknown etiology. In this study, we molecularly characterized a cohort of patients ($n = 35$) by studying their T-cell receptor (TCR) repertoire and the presence of somatic *STAT5B* mutations. In addition to the previously described gain-of-function mutations (N642H, Y665F, Q706L, S715F), we discovered six novel *STAT5B* mutations (Q220H, E433K, T628S, P658R, P702A, and V712E). Multiple *STAT5B* mutations were present in 22% (5/23) of *STAT5B* mutated CD4⁺ T-LGLL cases, either coexisting in one clone or in distinct clones. Patients with *STAT5B* mutations had increased lymphocyte and LGL counts when compared to *STAT5B* wild-type patients. TCR β sequencing showed that, in addition to large LGL expansions, non-leukemic T cell repertoires were more clonal in CD4⁺ T-LGLL compared to healthy. Interestingly, 25% (15/59) of CD4⁺ T-LGLL clonotypes were found, albeit in much lower frequencies, in the non-leukemic CD4⁺ T cell repertoires of the CD4⁺ T-LGLL patients. Additionally, we further confirmed the previously reported clonal dominance of TRBV6-expressing clones in CD4⁺ T-LGLL. In conclusion, CD4⁺ T-LGLL patients have a typical TCR and mutation profile suggestive of aberrant antigen response underlying the disease.

Blood Cancer Journal (2022)12:31 ; <https://doi.org/10.1038/s41408-022-00630-8>

INTRODUCTION

T-cell large granular lymphocyte leukemia (T-LGLL) is a rare lymphoproliferative disease characterized by chronic expansion of clonal, mature cytotoxic T cells in the peripheral blood and bone marrow [1]. Two subtypes of T-LGLL are commonly recognized: the most common CD8⁺ T-LGLL (70% of cases) and the less frequent CD4⁺ T-LGLL (30% of cases) [1]. Clinically, CD4⁺ T-LGLL is usually an indolent disease and unlike in its CD8⁺ counterpart, these patients rarely have cytopenias or autoimmune symptoms. However, CD4⁺ T-LGLL has been reported more frequently associated with secondary neoplasms such as monoclonal B-cell lymphocytosis and plasma cell disorders [2, 3]. In CD4⁺ T-LGLL, the leukemic T-LGLs express CD4 (either alone or together with CD8) and the α/β T-cell receptor (TCR) together with a typical mature cytotoxic (Granzyme B⁺, CD56⁺, CD57⁺) and activated/memory T-cell (CD2^{bright}, CD7^{dim}, CD11a^{bright}, CD28⁻, CD62L⁻) phenotype [3].

Up to 55% of CD4⁺ T-LGLL patients have been shown to harbor *STAT5B* mutations [4, 5]. In CD8⁺ T-LGLL, the most common mutated gene is *STAT3* [6], whereas *STAT5B* mutations are rare and often associated with an aggressive disease form [7–9]. All reported *STAT5B* mutations in CD4⁺ T-LGLL are point mutations within the SH2 or transactivation domains of *STAT5B*. N642H and Y665F are the most common *STAT5B* mutations, and they both have been shown to increase *STAT5B* protein activity [4, 10, 11].

The etiology of CD4⁺ T-LGLL remains unknown. An initial antigen-driven expansion of CD4⁺ T cells, followed by the occurrence of oncogenic events (i.e., somatic mutations), has been suggested to lead to the persistence of abnormal T-cell clones [11]. Non-self-antigen(s) instead of autoantigens [5] are proposed targets of CD4⁺ T-LGL clones as CD4⁺ T-LGLL is not associated with autoimmune diseases. In some earlier reports, CD4⁺ T-LGL clones have been implied to recognize cytomegalovirus (CMV) antigens [12, 13]. In CD4⁺ T-LGLL [3], the enrichment of the V β 13.1 gene usage has also been reported, differentiating it from

¹Hematology Research Unit Helsinki, University of Helsinki and Helsinki University Hospital Comprehensive Cancer Center, Helsinki, Finland. ²Translational Immunology Research Program and Department of Clinical Chemistry and Hematology, University of Helsinki, Helsinki, Finland. ³Department of Medicine, Hematology and Clinical Immunology Branch, University of Padova and Veneto Institute of Molecular Medicine (VIMM), Padova, Italy. ⁴Department of Computer Science, Aalto University, Espoo, Finland. ⁵Institute of Hematology and Center for Hemato-Oncology Research, University and Hospital of Perugia, Perugia, Italy. ⁶Department of Internal Medicine, Division of Hematology, Shinshu University School of Medicine, Matsumoto, Japan. ⁷Department of Biomedical Laboratory Sciences, Shinshu University School of Medicine, Matsumoto, Japan. ⁸ICAN Digital Precision Cancer Medicine Flagship, Helsinki, Finland. ⁹These authors contributed equally: Dipabarna Bhattacharya, Antonella Teramo, Vanessa Rebecca Gasparini, Jani Huuhtanen. ¹⁰These authors jointly supervised this work: Tiina Kelkka, Renato Zambello, Satu Mustjoki. ✉email: satu.mustjoki@helsinki.fi

Received: 10 September 2021 Accepted: 20 January 2022

Published online: 24 February 2022