(H30-化学-一般-001) 厚生労働科学研究費補助金(化学リスク研究事業) 令和元年度総括研究年度終了報告書

化学物質の動物個体レベルの免疫毒性データ集積とそれに基づくMulti-ImmunoTox assay (MITA)による予測性試験法の確立と国際標準化

(30210101)

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研究要旨

本課題においては、これまでに1)我々が開発した多項目免疫毒性評価 系 Multi-ImmunoTox Assay (MITA)の免疫毒性化学物質評価法としての |OECD||テストガイドライン化に向けて国際 validation 試験ならびに 2) 免疫毒性化学物質のデータベース作成を行ってきた。1)においては, 既に MITA を構成する試験法の一つである IL-2 Luc assay に関して validation 試験を終了し,それに基づき validation report を作成し peer review panel の評価を受けている。また IL-1 Luc assay に関し ても phase I, phase II の validation 試験を終了し, 2020年1月に行 われる海外からの liaison 委員を交えた validation management team (VMT)会議にて予測性を除いた試験結果の評価がなされる施設内施設間 再現生結果が承認された。一方,2)においては,上記validation 試 験にて評価した 50 化学物質, validation report 作成にあたり MITA に て評価した 60 化学物質に関して免疫毒性データを収集し免疫毒性デー タベースを構築した。また MITA の OECD テストガイドライン申請に向け て, in vitro 免疫毒性試験法の現状と MITA の有用性に関して detailed review paper を作成し OECD に提出する準備を始めた。

研究分担者氏名・所属研究機関名及び所属研究機関 における職名

小島 肇・国立医薬品食品衛生研究所安全性 生物試験研究センター薬理部・ 室長

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A.研究目的

研究背景:

アレルギー、自己免疫、免疫抑制など、人体に有害な影響を及ぼす化学物質による免疫毒性は、消費者、生産者はもとより厚生労働行政にとっても重大な課題となっている。現在、免疫毒性評価のゴールドスタンダードは動物実験であるが、数万ともいわれる化学物質を網羅的に評価、管理するには、*in vitro* high throughput 評価系や *in silico*評価系の構築が不可欠である。そのためには、化学物質のアレルギー発症、易感染性など個体レベルの免疫毒性データの集積、その分子メカニズムの解析、さらにはそれらに基づいた adverse outcome pathway の作成が不可欠である。

我々は,平成18-22年NEDO「高機能簡易型有害性評価手法の開発」プロジェクトにおいて、化学物質の免疫毒性多項目評価システム(Multi-ImmunoToxicity assay;MITA)を構築し国内外の特許を取得している。

また平成24年度から平成26年度の3年間にわたる厚生労働科学研究費補助金事業「多色発光細胞を用いたhigh-throughput免疫毒性評価試験法の開発」においては、作用機序の明らかな種々の免疫抑制剤をMITAにより評価するなかで、化学物質免疫毒性評価におけるMITAのプロトコールを作成し、そのプロトコールに基づいて薬剤の免疫毒性評価を行った。その結果、代表的な免疫抑制剤であるデキサメサゾン(Dex)、サイクロスポリン

(CyA)、タクロリムス (Tac)のT細胞とマクロファージ/樹状細胞に対する薬理効果をMITAが予測できることを明らかにした[1,2]。

さらに平成27年度以降は、皮膚感作性試験法IL-8 Luc assayとMITAを組み合わせたmodified MITAを構築し60種類の化学物質を評価しdata set を作成した。また、そのdata setを基に化学物質のclusteringを行い、化学物質が免疫毒性のprofileの違いにより6つのグループに分類できることを示した[3]。さらに、研究期間中にIL-8 Luc assayをOECDテストガイドライン化することができた(OECD442E)[4,5]。

計画全体の目的:

1)既にOECD テストガイドライン(442E)に承認されているIL-8 Luc assay に加え、MITA を構成する IL-2 転写活性抑制評価試験(IL-2 Luciferase reporter assay; IL-2 Luc assay)と IL- 転写活性抑制評価試験(IL-1 luciferase reporter assay; IL-1 Luc assay)の国際 validation study を行い、MITA の多項目免疫毒性評価系として OECD テストガイドライン化を目指す。

2)National Toxicology Program (NTP)のDori Germolec 博士とミラノ大学の Emanuela Corsini 博士の協力を仰ぎ、NTP ならびに European Centre for Ecotoxicology and Toxicology of Chemicals のデータベースおよび PubMed を利用した文献検索に基づき免疫毒性のデーターベースを構築する。

3)上記データベースに基づき ,MITA(図2)を用いた化学物質の免疫毒性別クラスター分類における各クラスター免疫毒性の特性を明らかにする。

2019年度

IL-2転写活性抑制試験 (IL-2 Luc assay)に関する validation report に対する peer review panelによる評価とそれに対する対応

IL-1 転写活性抑制試験(IL-1 Luc assay) に関するPhase I, Phase II validation試 験とValidation management teamによる最 終評価

IL-1 Luc assay, IL-2 Luc assay により多種類の化学物質を評価し data set を作成する。

免疫毒性化学物質のデータベース作成

MITAによる免疫毒性 clustering の有用性の 検討

MITA を用いた免疫毒性評価系国際化へ向けて, detailed review paper 作成を目的とした国際会議の開催

B. 研究方法

IL-2 Luc assay validation reportに対するpeer review panelによるコメントとそれに対する対応

以下の会議を開催し, peer review panelから IL-2 Luc assay validation reportに対するコ メントが提出され, それらに対応した。

 1. 1st International peer review panel meeting on Multi-Immunotoxicity Test Assay (MITA))

2019年2月27-28日,品川

Peer review panel: Henk van Loveren, Haley LaNef Ford, Barbara Kaplan, Sang-Hyun Kim, Fujio Kayama, Takao Ashikaga,

Xingchao Geng

参加者:Hajime Kojima, Yutaka Kimura, Setsuva Aiba

2. 2nd International peer review panel meeting on Multi-Immunotoxicity Test Assay (MITA)(Webex)

2019年10月1日(火)

Peer review panel: Henk van Loveren, Haley Neff-LaFord, Barbara Kaplan, Fujio Kayama, Takao Ashikaga

参加者:Hajime Kojima, Yutaka Kimura, Setsuya Aiba

3. 3rd International peer review panel meeting on Multi-Immunotoxicity Test Assay (MITA) (Webex)

2019年11月18日(月)

Peer review panel: Henk van Loveren, Haley Neff-LaFord, Barbara Kaplan, Lin Shi, Xingchao Geng, Fujio Kayama, Takao Ashikaga 参加者: Hajime Kojima, Yutaka Kimura, Setsuya Aiba

IL-1 Luc assay Phase IならびにPhase II validation試験

Phase I試験においては,国際バリデーション実行委員会 (VMT)にて選定された5化学物質をコ

ード化し、東北大学,産業技術総合研究所バイオメディカル研究部門,産業技術総合研究所工学研究部門の参加3施設においてMulti-ImmunoTox Assay protocol for TGCHAC-A4 ver. 008Eにのっとり各物質3回繰り返し1セットの試験を3セットと実施した。

Phase II試験においては、VMTにより選定された20化学物質をコード化し、東北大学、産業技術総合研究所バイオメディカル研究部門、産業技術総合研究所工学研究部門の参加3施設において Multi-ImmunoTox Assay protocol for TGCHAC-A4 ver. 008Eにのっとり各物質3回繰り返し1セットを実施した。

また, validation試験を遂行にあたり以下の VMT会議を行った。

1. 2019年度第1回MITAバリデーション電話会議 (スカイプ)

2019年4月5日(金)9:30-11:00

参加者:大森、髙木、小島、足利、相場、木村 2. 2019年度第2回MITAバリデーション電話会 議 (スカイプ)

2019年5月2日(木)10:00-12:00

参加者:大森、小島、安野、中島、相場、木村、 藤村

3. Conference call for the MITA assay (Webex)

2019年6月26日(水)20:00-

参加者:Corsini, E., Roggen, E., Germolec, D., Inoue, T., Aiba, S., Kimura, Y., Omori, T., Kojima, H.

4.5th meeting for the MITA Validation study 2020年1月30日 (水) 10:00-17:00

2020年1月31日(金)10:00-13:00

参加者:Corsini, E., Germolec, D., Inoue, T., Aiba, S., Kimura, Y., Omori, T., Kojima, H., Yasuno, R., Nakajima, Y.

IL-2 Luc assay, IL-1 Luc assayのdata set 作成

Validation試験で評価した化学物質以外の化学物質もIL-1 Luc assay、IL-2 Luc assayにて評価し,これらの試験法のdata setを作成した。

免疫毒性物質データベースの作成

National Toxicology Program (NTP)のDori Germolec 博士とミラノ大学の Emanuela Corsini 博士の協力を仰ぎ、NTP ならびに European Centre for Ecotoxicology and Toxicology of Chemicalsのデータベースおよび PubMed を利用した文献検索に基づき, validation 試験で用いた化学物質, data set に際して評価した化学物質を中心に免疫毒性データベースを構築した。

MITAによる免疫毒性 clustering の有用性の 検討

一方、我々はこれまでに60種類の化学物質をMITA の複数項目に関して効果発現最低濃度 (Lowest observed effect level; LOWEL)を基にクラスター分類することにより、免疫毒性物質が6種類のクラスターに分類できることを明らかにした[3]。そこで、さらに改訂された上記データベースを参考に MITA によりクラスター分類を再検討する。

MITA を用いた免疫毒性評価系国際化へ向けての国際評価会議の開催

皮膚感作性試験法を除いては, in vitro 免疫毒性試験法は OECD テストガイドラインに存在しない。そこで, OECD 免疫毒性試験評価者の in vitro 免疫毒性評価系の現状と MITA の有用性の理解の促進を図る目的で, in vitro 免疫毒性評価法に関する detailed review paper (DRP)の作成を計画し以下の会議を開催した。

1.1st call for DRP in vitro immunotoxicity (Webex)

2019年9月18日(水)、20時

Emanuela Corsini, Erwin Roggen, Dori Germolec, Henk van Loveren, Barbara Kaplan, Setsuya Aiba, Yutaka Kimura, Takayuki Yoshimoto, Hajime Kojima, Steve Venti

2. 2nd call for DRP in vitro immunotoxicity (Webex)

2019年10月28日(水)、20時

Emanuela Corsini, Erwin Roggen, Dori Germolec, Henk van Loveren, Barbara Kaplan, Setsuya Aiba, Yutaka Kimura, Takayuki Yoshimoto, Hajime Kojima, Steve Venti

3.3rd meeting for OECD DRP on in vitro immunotoxicity.

2020年1月28日 9:00-17:30 2020年1月29日 9:00-15:00

Emanuela Corsini, Dori Germolec, Henk van Loveren, Barbara Kaplan, Setsuya Aiba, Yutaka Kimura, Takayuki Yoshimoto, Hajime Kojima, Steve Venti

(倫理面への配慮)

健常人からの採血に際しては、研究内容、採血における危険性、得られた検査結果により本人の人権が損なわれることのないこと、得られた検査結果は守秘され個人のプライバシーを侵害する可能性がないこと、研究に協力することに同意した後もいつでも自由に辞退できること、この研究によって生じる知的財産権は被験者には帰属しないことについて説明し、本人より同意書を取得している。

C. 研究結果

IL-2 Luc assay validation reportに対する peer review panelによるコメントとそれに対する対応

今回IL-2 Luc assay validation reportを作 成するにあたり,施設内,施設間再現生は試験 開始前の目標値であった80%を達成した。しかし 予測性に関しては, そもそも医薬品を除く多く の化学物質の免疫毒性評価が必ずしも定まって いないため確定できないでいた。またpeer review pane会議にて, IL-2 Luc assayは免疫毒 性一般を評価する試験系ではなく、T細胞を一次 標的として免疫毒性を惹起する免疫毒性物質の 評価系であり、それを加味して予測性を決定す るように指導された。そこで,本試験において, NTPのLusterら[6-9]が51種類の化学物質の免疫 毒性を動物実験を用いて評価した際の判定基準 を参考にT細胞を標的とした化学物質の免疫毒 性を評価する分類法を提案し、peer review panelにより了承された。分類方法は添付資料1 を参照。これによりIL-2 Luc assayの予測性が 決定した(添付資料2)。それに基づき validation reportを作成し提出した(添付資料 3 抜粋)。

我々が提出したvalidation reportに対して, 1st International peer review panel meeting on Multi-Immunotoxicity Test Assay (MITA)) にて添付資料4のaction items (簡略版)が提案された。それに対して,添付資料5で対応した。さらに我々の回答に対して,2nd International peer review panel meeting on Multi-Immunotoxicity Test Assay (MITA) (Webex)で

は,添付資料6のaction itemsが提案され,それに対して添付資料7で対応した。

更に,3rd International peer review panel meeting on Multi-Immunotoxicity Test Assay (MITA) (Webex)では,添付資料8のaction items が提案され,それに対して添付資料8の赤字にて回答した。

IL-1 Luc assay Phase IならびにPhase II validation試験

IL-1 Luc assay Phase I試験を実施した。添付資料 9 に結果を示すが, within laboratory reproducibility, between laboratory reproducibility いずれも100%と極めて良好な結果が得られた。この結果に関して以下の会議を開催した。

2019年度第1回MITAバリデーション電話会議 (スカイプ)

2019年4月5日(金)9:30-11:00

参加者:大森、髙木、小島、足利、相場、木村

2019年度第2回MITAバリデーション電話会議 (スカイプ)

2019年5月2日(木)10:00-12:00

参加者:大森、小島、安野、中島、相場、木村、 藤村

第1回VMT会議 Conference call for the MITA assay (Webex)

2019年6月26日(水))20:00-

参加者:Corsini, E., Roggen, E., Germolec, D., Inoue, T., Aiba, S., Kimura, Y., Omori, T., Kojima, H.

以上の会議で,予測性に関しての最終評価は定まっていないが,さらに20化学物質を用いて施設間再現性を評価するPhase II 試験を行う事が了承された。そこで,3施設でPhase II 試験を実施し2019年12月までに全ての施設が試験を完了した。そこで以下の会議で試験結果が検討された。その結果,施設間再現性はPhase II 試験のみの結果で80%(資料10),Phase I の施設間再現性と共に試験開始前に想定していた採択基準をクリーアした。しかし,IL-1 Luc assayの再現性に関しては更に議論が必要と言うことになり,最終結論は次回のMVT会議に持ち越された。

第2回VMT会議

2020年1月31日(水))

会場:国立医薬品食品衛生研究所

参加者:Corsini, E., Roggen, E., Germolec, D., Inoue, T., Aiba, S., Kimura, Y., Omori, T., Kojima, H.

IL-1 Luc assay, IL-2 Luc assayのdata set 作成

IL-1 Luc assay, IL-2 Luc assayおよびIL-8 Luc assay に関して,それぞれの試験法の最終判定基準に則りdata setを作成した(添付資料11)

免疫毒性物質データベースの作成

IL-2 Luc assayのvalidationに用いた25化学物 質, IL-2 Luc assayのdata set作成に用いた化 学物質に関して免疫毒性データベースを作成し た。(添付資料12,添付資料13)データベースで は、化学物質の毒性データをin vivo、ex vivo、 in vitroデータの3種類に分類した。具体的には、 in vivo データの中には、免疫臓器の重量変化 遅延型過敏症,易感染性,移植腫瘍に対する抵 抗性が、ex vivo データには、化学物質を投与 された個体から採取した免疫担当細胞を用いて in vitroで化学物質の影響を評価するサイトカ イン産生試験 ,T細胞依存性性抗体産生試験 (Tcell dependent antibody response; TDAR)が、 in vitroデータには、個体から採取した免疫担 当細胞に、in vitroで化学物質を加えてそのサ イトカイン産生能の変化を評価するサイトカイ ン産生試験 ,T細胞の増殖能を評価する細胞増殖 試験などを含めた。この作成に当たっては、 National Toxicology Program (NTP)の協力を仰 いだ。

MITA による免疫毒性 clustering の有用性の 検討

あらたに得られたデータセットをもとに IL-8 Luc assay と組み合わせた MITA により化学物質の clustering を実施した。その結果を添付資料13 に示す。しかし,IL-1 Luc assay,IL-2 Luc assay,IL-8 Luc assayの組み合わせでは,以前論文で報告した IL-2 Luc assay,IL-8 promoter assay,IL-8 Luc assayの組み合わせで行ったようには綺麗に clustering できなかった。また残念ながら、MITA では、一部の DNA合成、細胞増殖抑制機序に基づく免疫毒性物質が評価できないことも明らかになった。

MITA を用いた免疫毒性評価系国際化へ向けての国際評価会議の開催

MITA のテストガイドライン化に向けて in vitro 免疫毒性評価法に関する detailed review paper (DRP)の作成を計画し以下の会議を開催した。

1.1st call for DRP in vitro immunotoxicity (Webex)

2019年9月18日(水)、20時

2. 2nd call for DRP in vitro immunotoxicity (Webex)

2019年10月28日(水)、20時

Emanuela Corsini, Erwin Roggen, Dori Germolec, Henk van Loveren, Barbara Kaplan, Setsuya Aiba, Yutaka Kimura, Takayuki Yoshimoto, Hajime Kojima, Steve Venti

上記会議において,以下の様な項目と執筆担当者が決定した添付資料 14。さらに下記の会議にて draft 案が提案され,それの修正を行った。修正後の draft を添付する(資料 15)

3.3rd meeting for OECD DRP on in vitro immunotoxicity.

2020年1月28日9:00-17:30

2020年1月29日9:00-15:00

Emanuela Corsini, Dori Germolec, Henk van Loveren, Barbara Kaplan, Setsuya Aiba, Yutaka Kimura, Takayuki Yoshimoto, Hajime Kojima, Steve Venti

E. 考察

臨床的に使われる免疫抑制剤を除くと,化学物質の免疫毒性,特にヒトに対する免疫毒性の評価は定まっていない。確かに,個々の化学物質に関して,幾つかの免疫毒性評価試験を行った報告は多数存在するが,それらを総括して化学物質の免疫毒性の有無を総括した報告は我々が調べた限り存在しない。この問題は,免疫毒性試験法のvalidation試験を行う際に大きな障害となった。

そこで本課題において、化学物質の免疫毒性に関する文献資料を基に免疫毒性の有無を判定するクライテリアを提案した。幸い、本課題においてはvalidation試験と並行して行ってきた

免疫毒性データベースが存在し、それをもとに 分類することを検討した。その際に, Lusterら [6-9]が報告した免疫毒性分類法を参考にした。 この方法では,51種類の化学物質をマウスに投 与し,その動物を種々の免疫毒性試験法で評価 し免疫毒性の有無を判定するクライテリアを提 案している。またそのクライテリアの判定結果 とマウス感染実験から得られた易感染性の有無 との相関も検討している。IL-2 Luc assavの予 測性の評価においても,ほぼLusterらのクライ テリアを参考に,作成した化学物質免疫毒性デ ータベースをもとに評価化学物質の免疫毒性の 有無を決定した。この妥当性は, peer review panelにからも承認された。この評価法に基づく と, Phase I、IIをまとめたpredictivityは75% となった。以上の結果をもとにvalidation reportを提出し現在peer review panelからの コメントに対応している。

また , 上記のように IL-2 Luc assayの validation試験の予測性評価を通して本課題で作成した免疫毒性データベースの有用性が確認された。

IL-1 Luc assayに関しては,これまでに順調に Phase I,Phase II試験を終了し,2020年1月に 行われるVMT会議で良好な施設内,施設間再現性 が評価され,現在予測性に関して検討中である。

最後に、IL-1 Luc assay、IL-2 Luc assayと免疫毒性評価法をOECDテストガイドライン化を進めるにあたり、detailed review paperを提出することにし既にOECDにSPSFを提出した。さらに、その中に含まれる項目と執筆担当者を決定した。さらに2020年1月において、draft案が提案され、それの修正を行った。修正後のdraftを添付する担当者が一同に介する会議を東京にて開催予定である。

一方,本課題のもう一つのテーマである化学物質の免疫毒性データベースの作成をNTPの協力を得て行った。25種類の化学物質の入手可能な免疫毒性データを網羅し、それらをin vivo, ex vivo、in vitroデータに分類し、さらにそれらを添付資料11,12にまとめた。その結果、各化学物質の大凡の免疫毒性profileが俯瞰可能となった。

IL-2 Luc assayのpredictivityに関しては、2019年2月27日から28日まで、東京にて開催予定のMITAのOECDガイドライン化に向けての国際評価会議にて検討する予定である。

E. 結論

本課題においては、これまで我々が開発した 多項目免疫毒性評価系 Multi-ImmunoTox Assay (MITA)の免疫毒性化学物質評価法としての OECD テストガイドライン化に向けて国際的 validation 試験を行ってきた。2019 年度までに MITA を構成する試験法の一つである IL-2 Luc assav に関しては validation 試験を終了し, そ れに基づき validation report を作成し peer review panel の評価を受けている。また IL-1 Luc assav に関しても phase I, phase II の validation 試験を終了し,2020年1月に行われ る海外からの liaison 委員を交えた validation management team (VMT)会議にて施設内施設間 再現生結果は承認された。また MITA の OECD テ ストガイドライン化にむけて,最終年度に提出 する予定の in vitro 免疫毒性試験に関する detailed review paper O standard project submission form (SPSF)を提出した。また validation 試験において評価した化学物質, MITA の data set の中に含まれる化学物質に関 して,既知の免疫毒性特性を文献的に収集し、本 課題のもう一つのテーマである化学物質免疫毒 性データベースの構築を進めた。

引用文献

- Kimura, Y., Fujimura, C., Ito, Y., Takahashi, T., Aiba, S. Evaluation of the Multi-ImmunoTox Assay composed of 3 human cytokine reporter cells by examining immunological effects of drugs. Toxicol In Vitro, 2014; 28: 759-768
- Saito, R., Hirakawa, S., Ohara, H., Yasuda, M., Yamazaki, T., Nishii, S., et al. Nickel differentially regulates NFAT and NF-kappaB activation in T cell signaling. Toxicol Appl Pharmacol, 2011; 254: 245-255
- 3. Kimura, Y., Fujimura, C., Ito, Y., Takahashi, T., Terui, H., Aiba, S. Profiling the immunotoxicity of chemicals based on in vitro evaluation by a combination of the Multi-ImmunoTox assay and the IL-8 Luc assay. Arch Toxicol, 2018; 92: 2043-2054
- 4. Takahashi, T., Kimura, Y., Saito, R., Nakajima, Y., Ohmiya, Y., Yamasaki, K., et al. An in vitro test to screen skin sensitizers using a stable THP-1-derived IL-8 reporter cell line, THP-G8. Toxicol Sci, 2011; 124: 359-369

- 5. Kimura, Y., Fujimura, C., Ito, Y., Takahashi, T., Nakajima, Y., Ohmiya, Y., Aiba, S. Optimization of the IL-8 Luc assay as an in vitro test for skin sensitization. Toxicol In Vitro, 2015: 29: 1816-1830
- 6. Luster, M.I., Munson, A.E., Thomas, P.T., et al. Development of a testing battery to assess chemical-induced immunotoxicity: National Toxicology Program's guidelines for immunotoxicity evaluation in mice. Fundam Appl Toxicol, 1988; 10: 2-19
- 7. Luster, M.I., Pait, D.G., Portier, C., et al. Qualitative and quantitative experimental models to aid in risk assessment for immunotoxicology. Toxicol Lett, 1992a; 64-65 Spec No: 71-78
- 8. Luster, M.I., Portier, C., Pait, D.G., et al. Risk assessment in immunotoxicology. I. Sensitivity and predictability of immune tests. Fundam Appl Toxicol, 1992b; 18: 200-210
- 9. Luster, M.I., Portier, C., Pait, D.G., et al. Risk assessment in immunotoxicology. II. Relationships between immune and host resistance tests. Fundam Appl Toxicol, 1993; 21: 71-82

F. 研究発表

1.論文発表

- Kimura, Y., Yasuno, R., Watanabe, M., Kobayashi, M., Iwaki, T., Fujimura, C., Ohmiya, Y., Yamakage, K., Nakajima, Y., Kobayashi, M., Mashimo, N., Takagi, Y., Omori, T., Corsini, E., Germolec, D., Inoue, T., Rogen, E.L., Kojima, H., Aiba, S. An international validation study of the IL-2 Luc assay for evaluating the potential immunotoxic effects of chemicals on T cells and a proposal for reference data for immunotoxicchemicals. Toxicol In Vitro, 2020; in press.
- 2. Hidaka, T., Fujimura, T., <u>Aiba, S.</u> Aryl hydrocarbon receptor modulates carcinogenesis and maintenance of skin cancers. Frot Med, 2019: 6: 194-

2. 学会発表

 15th International Congress of Toxicology, Hawaii convention center, July 15, 2019. Immunotoxicological Profiling of Chemicals Using Novel In Vitro Assays. Setsuya Aiba 2. 木村裕他: Multi-ImmunoTox Assay (MITA) の予測性評価に必要な文献に基づく化学物質免疫毒性分類の試み 日本動物実験代替法学会 第 32 回大会(つくば) 2019 年 11月

H.知的財産権の出願・登録状況 (予定を含む。)

- 1. 特許取得
- 1. 相場節也 齋藤るみ子 木村裕 近江谷克 裕 中島芳浩 西井重明 山崎友実 安田 真琴;特許第 5999644 号(2016);多色発光 細胞を用いた免疫毒性評価システム
- 2. <u>相場節也</u> 木村裕 近江谷克裕 西井重明;特開 2014-3939;免疫毒性評価細胞を用いたTNF 阻害活性を定量化するシステム
- 3. 木村裕 <u>相場節也</u>;特開 2016-208851; T L R 刺激物質の検出方法

添付資料1.化学物質免疫毒性評価基準 (Criteria to determine immunotoxicity of chemicals induced by directly targeting T cells) (IL-2 Luc assay validation report から抜粋)

To determine the performance of the IL-2 Luc assay, it is crucial to understand the immunotoxicological characteristics of the chemicals used in the study. Since the IL-2 Luc assay focuses on the effects of chemicals on IL-2 transcription by T cells, we attempted to classify the chemicals into two categories: (i) immunotoxic chemicals which target T cells (TTCs), which include chemicals that directly affect T cell viability, T cell proliferation or T cell function and (ii) others (NTTCs), which include chemicals that do not directly affect T cell viability, T cell proliferation or T cell function. In this assay, to define TTCs, we first surveyed the literature and collected the following six findings regarding each of the chemicals proposed for use in the study (Table 1). Using these six findings, we defined TTCs by the 4 criteria according to the rationale for classifying immunotoxic chemicals reported by Luster et al (Luster et al., 1992) (Table 2). Namely, if chemicals satisfy one of 4 criteria, they are considered as TTCs. Then, by comparing the results of the IL-2 Luc assay (positive or no effect) with the classification of the chemicals (TTC or NTTC), we calculated the accuracy, sensitivity and specificity of the IL-2 Luc assay in the validation study.

Table 1. The immunotoxicological data obtained from the literature

Findings	Information
Finding 1	Decreased thymus weight
Finding 2	Increased or decreased IL-2, IFN-g, IL-4 or other T cell-
	specific cytokine mRNA expression or protein production
	by T cells ex vivo.
Finding 3	Increased or decreased IL-2, IFN-g, IL-4 or other T cell-
	specific cytokine mRNA expression or protein production
	by T cells in vitro.
Finding 4	Suppressed T cell proliferation
Finding 5	Suppressed cytotoxic T cell response
Finding 6	The NTP data clearly indicate that one of the
	immunotoxic mechanism of chemicals are attributed to
	its effect on T cells.

Table 2. The criteria to classify immunotoxic chemicals by affecting T cells.

Criteria	Definition
Criterion 1	If chemicals are demonstrated to decrease thymus weight, one finding among Finding 2 to Finding 5
Criterion 2	There are multiple reports of Finding 2 or Finding 3.
Criterion 3	There are reports of increased or decreased mRNA expression or protein production in two or more cytokines for Finding 2 or Finding 3.
Criterion 4	The presence of the NTP data including Finding 6.

添付資料 2. IL-2 Luc assay バリデーション試験最終結果 (IL-2 Luc assay validation report から抜粋)

Chemical	CAS	Lab.A	Lab.B	Lab.C	concordance	T cell targeting
		Pha	se I			
Dibutyl phthalate	84-74-2	PPP	PPP	PPP	1	Yes
Hydrocortisone	50-23-7	PNN	PPP	PPN	0	Yes
Lead(II) acetate	6080-56-4	PPP	PPP	PPP	1	Yes
Nickel(II) sulfate	10101-97-0	PPP	PPP	PPP	1	Yes
DMDTC	137-30-4	NNN	NNN	NNN	1	No
Phase II						
2,4-Diaminotoluene	95-80-7	N	N	N	1	No
Benzo(a)pyrene	50-32-8	Р	Р	Р	1	Yes
Cadmium chloride	10108-64-2	N	N	N	1	Yes
Dibromoacetic acid	631-64-1	Р	Р	N	0	Yes
Diethylstilbestol	56-53-1	Р	Р	Р	1	Yes
Diphenylhydantoin	630-93-3	N	N	N	1	Yes
o-Benzyl-p-chorolophenol	120-32-1	Р	Р	Р	1	No

Ethylene dibromide	106-93-4	N	N	N	1	Yes
Glycidol	556-52-5	Р	Р	Р	1	No
Indomethacin	53-86-1	Р	Р	Р	1	Yes
Isonicotinic Acid Hydrazide	54-85-3	Р	N	Р	0	Yes
Nitrobenzene	98-95-3	N	S	N	0	Undetermine d
Urethane, Ethyl carbamate	51-79-6	Р	Р	Р	1	Yes
Tributyltin chloride	1461-22-9	Р	Р	Р	1	Yes
Perfluorooctanoic acid	335-67-1	Р	Р	Р	1	Yes
Dichloracetic acid	79-43-6	Р	Р	Р	1	Yes
Toluene	108-88-3	N	N	N	1	No
Acetonitril	75-05-8	N	N	N	1	No
Mannitol	69-65-8	N	N	N	1	No
Vanadium pentoxide	1314-62-1	N	N	N	1	No
o-Benzyl-p- chorolophenol	120-32-1	Р	Р	Р	1	No

Within-laboratory	80 (4/5)	100 (5/5)	80 (4/5)		
reproducibilities (%)	Average 86.7 (13/15)				
Between-laboratory reproducibilities (%) (Based on majority for Phase I)			80 (20/25)		
	75.0	75.0	75.0 (12/16)		
Sensitivity (%)	(12/16)	(12/16) Average 75.0 (36/48)			
	75.0	75.0	75.0 (6/8)		
Specificity (%)	(6/8)	(6/8) Average 75.0 (18/24)			
Accuracy (%)	75.0	75.0	75.0		
	(18/24)	(18/24) Average 75.0 (54/72)	(18/24)		

添付資料 3. IL-2 Luc assay バリデーションレポート (目次のみ抜粋)

Report on a Validation Study of the IL-2 Luc Assay for Evaluating the Potential Immunotoxic Effects of Chemicals on T-Cells

Validation Management Team

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添付資料4.IL-2 Luc assay validation report に対する Peer review panel からのコメント (1)

201902

Action Items to peer reviewers for the validation report on the IL-2 Luc assay

Evaluation Criterion 1: A rationale for the test method should be available, including a description of the human health effect, a clear statement of scientific need, and regulatory application.

PRP Comment: Together with a new title, the rationale needs to be stated clearly to be T-cell targeting.

Evaluation Criterion 2: The toxicological mechanisms and the relationship between the test method endpoint(s) with the biological effect as well as the toxicity of interest should be addressed, describing limitations of the test method.

PRP Comment: Needs to focus on IL-2, including the limitations described in the meeting minutes. The introduction needs to focus solely on IL-2 and the IL-2 Luc Assay. Discussion about its part in MITA should be left until the discussion section.

Evaluation Criterion 3: A detailed test method protocol should be available PRP Comment: The commercial availability of the #2H4 cell line needs to be described.

Evaluation Criterion 4: The within and between laboratory reproducibility of the test method should be demonstrated

PRP Comment: Acceptable

Evaluation Criterion 5: Demonstration of the test method's performance should be based on testing of representative, preferably coded reference chemicals

PRP Comment: We think only four or five negatives is not enough, so we suggest that some additional testing of negatives be performed.

Evaluation Criterion 6: Predictive capacity should be demonstrated using representative chemicals.

PRP Comment: Predictive capacity needs to be reassessed based on today's proposed definition of T-cell–targeting chemicals.

Evaluation Criterion 7: All data should adequately support the assessment of the validity of the test method for peer review.

PRP Comment: A clear definition of the 35% threshold and a clear explanation of Criteria 5 and how it was developed is needed. Should the table in Appendix 8 include the test judgment? Also, delete DTH, tumor, infection, and NK activity but specify T-cell proliferation in the table in Appendix 8.

Evaluation Criterion 8: All data from the validation study supporting the validity of a test method should be obtained in accordance with the principles of Good Laboratory Practice (GLP)

PRP Comment: The report needs to explain clearly and in detail what is meant by the phrase "in the spirit of GLP" and whether or not each laboratory performed their work in this spirit.

Evaluation Criterion 9: Applicability domain of the test method should be defined PRP Comment: We recommend that the applicability domain be more clearly defined as noted in the PRP meeting minutes.

Evaluation Criterion 10: Proficiency chemicals should be set up in the proposed protocol

PRP Comment: None

Evaluation Criterion 11: Performance standards should be set up with the proposed protocol

PRP Comment: If performance standards are understood to be assay controls, then the use of three-fold stimulation of IL-2 Luc by PMA/IO and inhibition of stimulated IL-2 Luc by DEX and CYA are sufficient. We suggest that acceptance criteria for variability within test replicates be defined.

Evaluation Criterion 12: Advantages in terms of time, cost and animal welfare

PRP Comment: We suggest that the conclusion leave out mention of in vivo testing to

confirm T-cell immunotoxicity and include discussion of the use of IL-2 Luc assay

within MITA.

Evaluation Criterion 13: Completeness of all data and documents supporting the

assessment of the validity of the test method.

PRP Comment: We suggest that data be redone to reassess predictive capacity based

on today's proposed definition of T-cell-targeting chemicals. Also, a critical

assessment of the 35% threshold in the context of the new definition of T-cell

targeting is necessary.

Evaluation Criterion 14: Validation Study Management and Conduct

PRP Comment: None

Other considerations

PRP Comment: None

Conclusion

PRP Comment: We look forward to seeing a revised report based on our comments.

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添付資料 5. Peer review panel からのコメント (1)に対する対応 Dear the PRP:

Thank you for your kind and constructive comments and suggestions. We responded to each comment below and revised the VR taking the PRP comments into consideration. We used red fonts in the revised or newly added parts.

Evaluation Criterion 1: A rationale for the test method should be available, including a description of the human health effect, a clear statement of scientific need, and regulatory application.

PRP Comment: Together with a new title, the rationale needs to be stated clearly to be *T-cell targeting*.

The title was revised and changed to "Report on a Validation Study of the IL-2 Luc Assay for Evaluating the Potential Effect of Chemicals on T-Cells".

The rationale to judge chemicals whether they were T-cell targeting or not was described in 10-3-1.

10-3-1. Rationale to determine the predictivity of the IL-2 Luc assay by the concordance between positive effects and the immunotoxic effects targeting T cell response

A well-functioning immune system is essential in maintaining the integrity of the organism. Therefore, immune dysregulation caused by chemicals, i.e., immunotoxic effects of chemicals, may make serious impacts on human health. It ranges from reduced resistance to infection and neoplasia to allergic and autoimmune conditions. The immune system comprises innate and adaptive immunity (Fig. 2). Both arms of the immune response function differently and are driven by different populations of cells. Chemicals can potentially affect the immune system by targeting either the innate immune system or the acquired immune system (Fig. 2 and Fig. 3). Therefore, in vitro test methods to detect immunotoxic effects of chemicals are needed to adequately assess their effects on both arms of immune system. However, it is impossible to predict the toxic effects of chemicals on the whole aspects of immune system by a single in vitro assay. Consequently, to accomplish the final goal of in vitro immunotoxicity tests that cover the whole aspects of immune system, it is indispensable to develop an integrated approach composed of multiple in vitro immunotoxic tests evaluating different aspects of immune responses. The MITA including the IL-2 Luc assay was developed to be components of the integrated approach.

Among various immune responses, one of pivotal responses is the development of antigen-specific effector T-helper subtypes, such as, Th1 cells, Th2 cells, Th17 cells, and regulatory T cells (Treg cells) that are associated with the clinical features and disease progression (reviewed by [1]). Therefore, the in vitro assay to clarify the effects

of chemicals on the development of these T-helper subtypes is one of the critical components of the integrated approach.

Now it is known that IL-2 exerts pleiotropic actions on CD4+ T cell differentiation via its modulation of cytokine receptor expression. It promotes Th1 differentiation by inducing IL-12Rb2 (and IL-12Rb1), promotes Th2 differentiation by inducing IL-4Ra, inhibits Th17 differentiation by inhibiting gp130 (and IL-6Ra), and drives Treg differentiation by inducing IL-2Ra. IL-2 also potently represses IL-7Ra, which decreases survival signals that normally promote cell survival and memory cell development (reviewed by [2]). Therefore, it is conceivable that chemicals, which affect IL-2 release by T cells, give significant impact on the development of Th cells.

When immunotoxic information of chemical is collected from the literature, however, most of the published data are not focusing on the effects of chemicals on the development of Th subsets. To overcome this problem, in this study, the predictivity was evaluated by the criteria whether chemicals affect T cell functions, namely T cell targeting, or not. To determine T cell targeting chemicals (TTCs), we collected the following 6 components in the literature.

- #1. The decreased thymus weight
- #2. The increased or decreased IL-2, IFN-g, or IL-4 mRNA expression or production by T cells in ex vivo.
- #3. The increased or decreased IL-2, IFN-g, or IL-4 mRNA expression or production by T cells in vitro.
- #4. The suppression of T cell proliferation
- #5. The suppression of cytotoxic T cell response
- #6. There is a clear statement in the NTP data that one of the immunotoxic mechanism of chemicals are attributed to its effect on T cells.

Then, we determined TTCs as chemicals that satisfied one of the following criteria

- 1) The combination of more than two components among #1 to #5 components
- 2) Multiple reports on #2 or #3
- 3) #2 or #3 on two or more cytokines
- 4) #5

Evaluation Criterion 2: The toxicological mechanisms and the relationship between the test method endpoint(s) with the biological effect as well as the toxicity of interest should be addressed, describing limitations of the test method.

PRP Comment: Needs to focus on IL-2, including the limitations described in the meeting minutes. The introduction needs to focus solely on IL-2 and the IL-2 Luc Assay. Discussion about its part in MITA should be left until the discussion section.

The limitation of this assay was described in the applicability domain (10-6).

10-6. Limitations and drawback, and applicability domain of the IL-2 Luc assay

Since the 2H4 cell line used in the IL-2 Luc assay is derived from Jurkat cells, it is
conceivable that this cell line is more resistant to the cytotoxic effects of chemicals than
bone marrow cells. Indeed, our study demonstrated that the IL-2 Luc assay cannot
evaluate the immunotoxic effects of some immunosuppressive drugs which act by
inhibiting DNA synthesis leading to myelotoxicity [3]. Thus, these chemicals in
addition to chemicals that need metabolic activation should be outside the applicability
domain. To overcome this drawback at present, the IL-2 Luc assay must be combined
with assays capable of detecting myelotoxicity, such as the conventional 28-day
subacute toxicity test [4] or *in vitro* myelotoxicity tests [5]. Similar to other *in vitro* test
methods, poor water soluble chemicals are not suitable for this assay.

The introduction was revised according to the PRP comment. The detailed discussion on the MITA was moved to the Discussion.

Evaluation Criterion 3: A detailed test method protocol should be available PRP Comment: The commercial availability of the #2H4 cell line needs to be described.

2H4 cells will be obtained from the GPC laboratory, Tottori, Japan after this assay is accepted as the test guideline.

Evaluation Criterion 4: The within and between laboratory reproducibility of the test method should be demonstrated

PRP Comment: Acceptable

Evaluation Criterion 5: Demonstration of the test method's performance should be based on testing of representative, preferably coded reference chemicals

PRP Comment: We think only four or five negatives is not enough, so we suggest that some additional testing of negatives be performed.

We reconsidered the immunotoxic characteristics of chemicals evaluated in Phase I and II studies. Finally, these two studies contained 7 negative chemicals (Appendix 8).

Evaluation Criterion 6: Predictive capacity should be demonstrated using representative chemicals.

PRP Comment: Predictive capacity needs to be reassessed based on today's proposed definition of T-cell-targeting chemicals.

We admit that it is crucial to more clearly define the criteria to classify chemicals into T cell-targeting chemical (TTC) and non-T cell-targeting chemical (NTTC). So, we proposed the new criteria with the international expert members, Dr. Emanuela Corsini and Dr. Dori Germolec taking PRP's proposal into consideration. The following was the revised session of predictivity (Revised VR 9-1-3).

9-1-3. Predictivity

To determine the predictivity of the IL-2 Luc assay, it is crucial to understand the immunotoxic characteristics of chemicals used in the study. Since the IL-2 Luc assay focuses on the effects of chemicals on IL-2 transcription by T cells, we tried to classify chemicals into those that affect T cell function, i.e., T cell-targeting chemical (TTC) and those that do not directly affect T cell function, i.e., non-T cell-targeting chemicals (NTTC). In this assay, to define TTCs, we collected the following 6 components in the literature.

- #1. The decreased thymus weight
- #2. The increased or decreased IL-2, IFN-g, or IL-4 mRNA expression or production by T cells in ex vivo.
- #3. The increased or decreased IL-2, IFN-g, or IL-4 mRNA expression or production by T cells in vitro.
- #4. The suppression of T cell proliferation
- #5. The suppression of cytotoxic T cell response
- #6. There is a clear statement in the NTP data that one of the immunotoxic mechanism of chemicals are attributed to its effect on T cells.

Then, we defined TTCs as chemicals that satisfy one of the following criteria

- 1) The combination of more than two components among #1 to #5 components
- 2) Multiple reports on #2 or #3
- 3) #2 or #3 on two or more cytokines
- 4) #5

To classify 25 chemicals used in the Phase I and II studies, we used the chemical information kindly provided by the National Toxicology Program (NTP). The immunotoxic characteristics of each chemical are shown in Appendix 7 and their summarized data are shown in Appendix 8. The table in Appendix 8 is the combined data of the NTP data and the data collected by the VMT member. As already described, IL-2 exerts pleiotropic actions on CD4+ T cell differentiation via its modulation of cytokine receptor expression. Indeed, IL-2 promotes Th1 and Th2 differentiation, while it also drives Treg differentiation. Therefore, it suggests that the augmentation of IL-2 transcription can lead to either immunostimulation or immunosuppression depending on surrounding tissue environment *in vivo*. Therefore, in this assay, if chemicals were judged as either augmentation or suppression, they were both considered as positive (P) and if not, they were judged as negative (N). Then we examined concordance between positive judgment and TTC.

Based on the new criterial for chemical classification, the predictivity of the Phase I and Phase II studies was summarized in 10-3-2.

10-3-2. The predictivity of the Phase I and Phase II studies

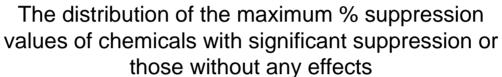
To classify 25 chemicals used in the Phase I and II studies, we used the chemical information kindly provided by the National Toxicology Program (NTP) and those collected by the VMT members. The immunotoxic characteristics of each chemical are shown in Appendix 7 and their summarized data are shown in Appendix 8. Based on the criteria, the 25 chemicals were classified into 14 TTCs, 9 NTTCs, and 2 unclassified chemicals that could not be classified because of insufficient data. According to this classification, the sensitivities of the assays as conducted by Lab A, Lab B, Lab C, and their average in the combined data of the Phase I and II studies are 80.0%, 80.0%, 73.3% and 77.7%, respectively. The specificities of the assays as conducted by Lab A, Lab B, Lab C, and their average are 75.0%, 75.0%, 75.0%, and 75.0%, respectively. The accuracies of the assays conducted by Lab A, Lab B, Lab C, and their average are 78.2%, 78.2%, 73.9%, and 76.8%, respectively.

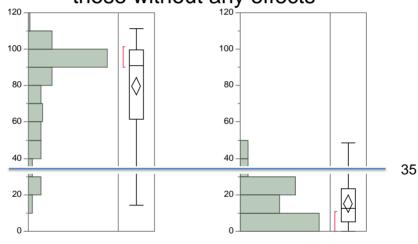
Evaluation Criterion 7: All data should adequately support the assessment of the validity of the test method for peer review.

PRP Comment: A clear definition of the 35% threshold and a clear explanation of Criteria 5 and how it was developed is needed. Should the table in Appendix 8

include the test judgment? Also, delete DTH, tumor, infection, and NK activity but specify T-cell proliferation in the table in Appendix 8.

To determine the optimum threshold, we first potted the maximum % suppression values of chemicals with statistically significant suppression or those without any effects. The comparison of these two graphs showed that the threshold 35 can divide chemicals with significant suppression and those without any effects with minimum false positive or negative results.





Chemicals without any significant effects

The values of the maximum % suppression were derived from the data set made by the lead laboratory in our recent publication in Arch Toxicol (see the attached file)

We revised Appendix 8. As suggested, we deleted test judgment, DTH, infection, tumor rejection, and NK activity, and specified T cell proliferation.

Chemicals with significant suppression

Evaluation Criterion 8: All data from the validation study supporting the validity of a test method should be obtained in accordance with the principles of Good Laboratory Practice (GLP)

PRP Comment: The report needs to explain clearly and in detail what is meant by the phrase "in the spirit of GLP" and whether or not each laboratory performed their work in this spirit.

Evaluation Criterion 9: Applicability domain of the test method should be defined PRP Comment: We recommend that the applicability domain be more clearly defined as noted in the PRP meeting minutes.

We described the applicability domain more precisely, taking the PRP comments into consideration in 10-6.

10-6. Limitations and drawback, and applicability domain of the IL-2 Luc assay

Since the 2H4 cell line used in the IL-2 Luc assay is derived from Jurkat cells, it is
conceivable that this cell line is more resistant to the cytotoxic effects of chemicals than
bone marrow cells. Indeed, our study demonstrated that the IL-2 Luc assay cannot
evaluate the immunotoxic effects of some immunosuppressive drugs which act by
inhibiting DNA synthesis leading to myelotoxicity [3]. Thus, these chemicals in
addition to chemicals that need metabolic activation should be outside the applicability
domain. To overcome this drawback at present, the IL-2 Luc assay must be combined
with assays capable of detecting myelotoxicity, such as the conventional 28-day
subacute toxicity test [4] or *in vitro* myelotoxicity tests [5]. Similar to other *in vitro* test
methods, poor water soluble chemicals are not suitable for this assay.

Evaluation Criterion 10: Proficiency chemicals should be set up in the proposed protocol

PRP Comment: None

Evaluation Criterion 11: Performance standards should be set up with the proposed protocol

PRP Comment: If performance standards are understood to be assay controls, then the use of three-fold stimulation of IL-2 Luc by PMA/IO and inhibition of stimulated IL-2 Luc by DEX and CYA are sufficient. We suggest that acceptance criteria for variability within test replicates be defined.

Based on the PRP comments, we added the performance standard in the revised VR, Appendix 15.

Evaluation Criterion 12: Advantages in terms of time, cost and animal welfare PRP Comment: We suggest that the conclusion leave out mention of in vivo testing to confirm T-cell immunotoxicity and include discussion of the use of IL-2 Luc assay within MITA.

In the revised VR, we deleted the description of requirement of in vivo testing. In addition, we described the potential of the IL-2 Luc assay (10-7).

10-7. Potential of the IL-2 Luc assay

The IL-2 Luc assay evaluates the effects of chemicals on IL-2 transcription by Jurkat T cells stimulated with PMA and CI. The simultaneous stimulation of PMA and calcium ionophore or ionomycin surrogates the stimulation by T cell receptor (TCR) and CD28 [6, 7]. The downstream signaling after the stimulation by TCR/CD28 is shown in Fig. 17. It indicates that the signaling required for IL-2 transcription after TCR/CD28 or PMA/CI stimulation involves the pathways leading the activation of AP1/2, mTOR, NF-kB, and NFAT. The immune system is composed of innate immune system and acquired immune system at least. The innate immune systems are activated by pathogen-associated molecular patterns (PAMPs) or damage-associated molecular patters via Toll-like receptors (TLRs), RIG-I-like receptors (RLRs), Nod-like receptors (NLRs), or cytokine receptors for IL-1 family or TNF family. Most of the downstream signaling after the stimulation of these receptors involves NF-kB and AP1/2 pathways [8]. In the acquired immune system, in addition to the process of T cell activation, B cell activation after B cell receptor stimulation and the signaling of various cytokines also involves NF-kB pathway (reviewed by Zhang and Sun [9]. Therefore, it is conceivable that the effects of chemicals on quite a few aspects of immune responses can be detected by the IL-2 Luc assay.

Evaluation Criterion 13: Completeness of all data and documents supporting the assessment of the validity of the test method.

PRP Comment: We suggest that data be redone to reassess predictive capacity based on today's proposed definition of T-cell—targeting chemicals. Also, a critical assessment of the 35% threshold in the context of the new definition of T-cell targeting is necessary.

In the revised validation report, we clearly defined the T cell-targeting chemicals. Based on the definition, we classified chemicals into T cell-targeting chemicals (TTCs) or non-T cell targeting chemicals (NTTCs). According to this classification, we recalculated the sensitivity, specificity, and accuracy of the Phase I and II studies.

Evaluation Criterion 14: Validation Study Management and Conduct PRP Comment:None

Other considerations

PRP Comment:None

Conclusion

PRP Comment: We look forward to seeing a revised report based on our comments.

References

- [1] Kaiko GE, Horvat JC, Beagley KW, Hansbro PM: Immunological decision-making: how does the immune system decide to mount a helper T-cell response? Immunology 123: 326-338, 2008.
- [2] Liao W, Lin JX, Wang L, Li P, Leonard WJ: Modulation of cytokine receptors by IL-2 broadly regulates differentiation into helper T cell lineages. Nat Immunol 12: 551-559, 2011.
- [3] Kimura Y, Fujimura C, Ito Y, Takahashi T, Aiba S: Evaluation of the Multi-ImmunoTox Assay composed of 3 human cytokine reporter cells by examining immunological effects of drugs. Toxicol In Vitro 28: 759-768, 2014.
- [4] Investigators TIG: Report of validation study of assessment of direct immunotoxicity in the rat. The ICICIS Group Investigators. International Collaborative Immunotoxicity Study. Toxicology 125: 183-201, 1998.
- [5] Pessina A, Albella B, Bayo M, Bueren J, Brantom P, Casati S, et al.: Application of the CFU-GM assay to predict acute drug-induced neutropenia: an international blind

- trial to validate a prediction model for the maximum tolerated dose (MTD) of myelosuppressive xenobiotics. Toxicol Sci 75: 355-367, 2003.
- [6] Truneh A, Albert F, Golstein P, Schmitt-Verhulst AM: Calcium ionophore plus phorbol ester can substitute for antigen in the induction of cytolytic T lymphocytes from specifically primed precursors. J Immunol 135: 2262-2267, 1985.
- [7] Kumagai N, Benedict SH, Mills GB, Gelfand EW: Requirements for the simultaneous presence of phorbol esters and calcium ionophores in the expression of human T lymphocyte proliferation-related genes. J Immunol 139: 1393-1399, 1987.
- [8] Newton K, Dixit VM: Signaling in innate immunity and inflammation. Cold Spring Harb Perspect Biol 4, 2012.
- [9] Zhang H, Sun SC: NF-kappaB in inflammation and renal diseases. Cell Biosci 5: 63, 2015.

添付資料 6. Peer review panel との teleconferance の議事録

Teleconference for IL-2 PRP

October 1, 2019

Peer Review Panel: Henk van Loveren, Haley Neff-LaFord, Barbara Kaplan, Fujio Kayama, Takao Ashikaga

VMT: Hajime Kojima

Observers: Steve Venti (meeting minutes)

Kojima:	In this meeting, we will discuss the revised validation report and the schedule going forward.
	I will explain the changes in the report, which are shown in red.
	One important point is Appendix 7. It has 290 pages and discusses the data available on
	immunotoxic effects of chemicals. Mainly, the figures for predictivity and the summary were revised. I heard Dr. Aiba is on-
Kaplan:	going to revise minorly. After the meeting, I ill share the newest Validation report. This summary is in line with what we discussed at the FTF meeting.
Kapian. Kojima:	Does everyone accept this summary?
Everyone:	Yes.
Kojima:	Section 9-1-3 addresses predictivity and describes the effects of chemicals on T-cells. And
Kojima.	there is a definition of T-cell targeting chemicals (TTCs).
Kaplan:	Criterion 3 says "#2 or #3 on two or more cytokines." Does that refer only to the three
Tapian.	cytokines mentioned in #2 and #3? For example, is IL-17 excluded? This is not clear. If there is
	a report for other cytokines, would they be considered TTCs?
Kojima:	I can't answer at the moment, but I will ask Dr. Aiba.
Kaplan:	This is an improvement over the original report. Once we have some clarification on Criterion
	3, I think that these criteria are acceptable.
van Loveren:	Although I think it would be good to extend this to other cytokines, not just the ones listed.
Kojima:	(Brief review of other changes in red. Please see revised Validation Study Report.)
	If you are happy with this report, then we can move on to reviewing the PRP Evaluation
	Criteria and creating the PRP report.
Kaplan:	Do we need to read this and provide comments? What do you need from the PRP to submit to
	the OECD?
Kojima:	If you feel that the Validation Study Report satisfies the 14 PRP Evaluation Criteria, then you
	can prepare a Peer Review Report of about 12 pages with a comment about each criterion. And
	then the Validation Study Report and the Peer Review Report will be reviewed by an OECD
	expert working group.
van Loveren:	Are there specific places we should comment on?
Kojima:	We revised the Validation Report based on the comments from the PRP.
Kaplan:	So we have already covered the critical issues. But if there is anything specific you want us to
	look at, please tell us now.
van Loveren:	Is there any issue we need to address now?
Kojima:	I will share these documents with you, and after we have your comments, Dr. Kayama will
NI CCI E 1	write the final PRP report.
Neff-LaFord:	Once you see the documents, it is pretty easy to follow what has been changed, so we should
Valler	be able to follow it.
Kojima:	The deadline for comments if possible, would be by the end of October and then we can have
	another teleconference in early or mid-November.
	OK, I will send you meeting minutes, the newest validation Study Report, and the evaluation

添付資料 7. Teleconferance のコメントに対する対応

October 4th, 2019

The response to the reviewers' comments:

Thank you for your kind consideration and important suggestions to the validation report. We revised the validation report according to the reviewers' comments. In addition, we corrected the values of the predictivity of this assay because there was one calculation error and we changed the classification of chemicals based on several references we found. The modified part was as follows. All the modified parts were written in red.

- We modified the criteria to classify immunotoxic chemicals according to the reviewers' comments. (9-1-3. Predictivity in Page 61 and 10-3-1. Rationale to determine...... in Page 82)
- 2. We recalculated the predictivity. Consequently, the predictivity of the Phase II study, the combined Phase I and Phase II studies, and the data set was slightly changed. Briefly, the average predictivity of the Phase II was changed from 74.0% (40/54) to 70.2% (40/57). The average predictivity of the combined Phase I and Phase II studies was changed from 76.8% (53/69) to 75.0% (54/72). The predictivity of 60 chemicals was not changed. These changes were precisely described in Abstract, 9-4-3. Predictivity in the Phase II study, Table 22, 9-6-3. Predictivity in the Phases I and II studies, Table 23, 10-3-2. The predictivity of the Phase I and Phase II studies, 10-4. IL-2 Luc assay data set for 60 chemicals, and Table 24.
- 3. While revising the VR, we found a very crucial report by Luster et al, 1992. In their manuscript (Luster et al., 1992b), they proposed the rationale for immunotoxic classification. Namely, their proposal was that a positive was established on the basis that the test material either produced significant dose-response effect in the immune tests or significantly altered two or more test results at the highest dose of chemical tested. Furthermore, they classified chemicals based on their results of immune tests according to this rationale and found that there was a significant correlation between the judgment of immunotoxic chemicals and the host resistance (Luster et al., 1993). Therefore, we referred to their paper in 9-1-3. Predictivity and 10-3-1. Rationale to determine.....

4. We also added the comparison between the predictivity of the IL-2 Luc assay and that reported by Luster et al. (Luster et al., 1992a; Luster et al., 1993; Luster et al., 1992b)and between the predictivity of the IL-2 Luc assay and that of the human whole blood cytokine release assay by Langezaal et al. (Langezaal et al., 2002) in 10.4. IL-2 Luc assay data set for 60 chemicals.

References

- Langezaal, I., Hoffmann, S., Hartung, T., et al., 2002. Evaluation and prevalidation of an immunotoxicity test based on human whole-blood cytokine release. Alternatives to laboratory animals: ATLA 30, 581-595.
- Luster, M.I., Pait, D.G., Portier, C., et al., 1992a. Qualitative and quantitative experimental models to aid in risk assessment for immunotoxicology. Toxicol Lett 64-65 Spec No, 71-78.
- Luster, M.I., Portier, C., Pait, D.G., et al., 1993. Risk assessment in immunotoxicology. II. Relationships between immune and host resistance tests. Fundam Appl Toxicol 21, 71-82.
- Luster, M.I., Portier, C., Pait, D.G., et al., 1992b. Risk assessment in immunotoxicology. I. Sensitivity and predictability of immune tests. Fundam Appl Toxicol 18, 200-210.

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添付資料8.

Teleconference for IL-2 PRP

November 11, 2019

Peer Review Panel: Henk van Loveren, Barbara Kaplan, Haley Neff-LaFord, Fujio Kayama,

Takao Ashikaga, Lin Shi, Xingchao Geng

VMT: Hajime Kojima, Setsuya Aiba, Takuya Kimura

Observers: Steve Venti (meeting minutes)

Kojima:	In this meeting, we will discuss the revised validation report prior to discus
	items. We revised the report based on your comments. After the previous
	we received it in accordance with the comments from Barbara, and you have
	comments that have not been reflected yet, so I think we need to discuss the
Kaplan:	I think these revisions are fine as long as things are separated into a table o
_	intelligible.
Aiba:	I don't know who made this table, but it presents what I wanted to say, so I
	this if the PRP agrees.
Kojima:	Dr. Aiba will calculate predictive capacity based on this table, so the most i
	that the PRP finds this table acceptable.
Kayama:	I think these criteria are easier to understand as presented in the table.
van Loveren:	I am still concerned that the introduction is confusing to a naïve reader. We
	understand that MITA is the context, <i>not</i> the aim, of this study. But the intro
	clear statement at the start of the introduction that the aim of this validation
	not MITA in general. Mentioning MITA in the introduction is fine, but you
	at the start of the introduction. The introduction must begin with the aim of
	IL-2.
	According to the reviewer's suggestion, I changed the abstract and began it
	this study.
Kaplan:	The first time I read this introduction, I thought that you were validating the
	later I realized that is not the case. The goal is to validate the IL-2 assay. I ϵ
	Haley that the goal of the validation needs to be stated clearly at the start of
	Even just one sentence is enough. Just clearly state that the goal is to validate
	As described in the response to Dr. van Loveren's comment, I changed the
	it with the purpose of this study.
Neff-LaFord:	Yes, just more section 3 up higher.
	As suggested by the reviewer, we moved the objective of the study to section
van Loveren:	We need to say "proposed AOP" because this AOP has not yet been acceptor
	As suggested, we added "proposed " in 3-9. The proposed Adverse Outcom
	of chemicals that affect IL-2 transcription.
Neff-LaFord:	The expression "IL-2 LA" appears to mean the same thing as "IL-2 Luc As
	intended to mean something different, then this needs to be spelled out mor
	According to the reviewer's comment, we modified Table 3. Definition of t

Aiba:	Yes, I will clarify that.
van Loveren:	On page seven in introduction, I have suggested a revision, but perhaps the
	the applicability range that I deleted needs to be added back.
Kaplan:	I think that in context, the meaning of "applicability domain" is clear enough
	the word "however" should be removed for clarity.
	As suggested by several reviewers, we deleted "however".
van Loveren:	The applicability domain is discussed in the preceding paragraph, so maybe
	Haley's suggestion as is.
Kojima:	In section 9-5, I will inform you the detailed records collected in the principal section 9-5.
Neff-LaFord:	1
	We changed "comparable" to "similar to", which is now in section 10-7.
Kaplan:	Given the emphasis on comparing IL-2 results with the results of other test
	section needs to be expressed more clearly. I think this information is impo
	it should be described more clearly.
	In the revised VL, we tried to describe more clearly the following sections,
Ashikaga:	I couldn't find any description about regulatory application in the report.
	We added a new section describing the regulatory application (10-9)
Aiba:	Do I need to respond to each of these comments one by one?
Ashikaga:	Why is SFO-luciferase activity measured in this assay?
	We made a comment for the reason to ignore SLO-luciferase activity or IFI
Aiba:	It is automatically measured but it is not necessary for this assay.
Kaplan:	This is related to what we were talking about before. This report contains a
	that is only incidentally related to IL-2, which confuses the reader.
Ashikaga:	I could not find a list of proficiency chemicals. Shouldn't the developer sub-
Aiba:	Yes. Appendix 14 and 15 have a list of proficiency chemicals.
Kojima:	Are there any other comments?
Xingchao:	I agree with the comments and I think the report is improved.
Lin:	(inaudible)
Aiba:	(inaudible)

The applicability domain does not seem to be defined anywhere. Where is applicability domain? All the information is there, but there is no single cle
could rename 10-6 and start with a simple explanation of the applicability (
According to the reviewers' suggestion, we changed the name of 10-6 to th
domain and the limitation of the IL-2 Luc assay and added a simple explan
applicability domain.
This is a good point. We have defined a T-cell target, so we need to say that
applicability domain.
We have answered to Dr. van Loveren's comment.
OK, I will provide a clear definition of what the applicability domain is.
I will share the minutes of this meeting, and then Dr. Aiba and the VMT wi
validation report to share with the PRP. Perhaps you can then submit your of
Kayama within one month and to be created the PRP report by Dr. Kayama
The most important comment today is Henk's last comment.
I'd like to ask Dr. Kayama to summarize the PRP comments, because I alre
original comments. I would like to know what I should respond to.
Will the PRP report be incorporated into the validation report or separately
Separately attached.

添付資料 9. IL-1 Luc assay Phase I validation 試験結果

The results of the Phase I study

Line25 judge		25									
	LabA 1	LabA Tohoku LabB AIST tsukuba LabC AIST shikoku								T shikoku	
setNo.	code No.			setNo.	code No.			setNo.	code No.		
Set1	MITA103	MITA103	S	Set1	MITB402	MITB402	S	Set1	MITC704	MITC704	S
Set2	MITA203	MITA203	S	Set2	MITB501	MITB501	S	Set2	MITC803	MITC803	S
Set3	MITA304	MITA304	S	Set3	MITB605	MITB605	S	Set3	MITC902	MITC902	S
Set1	MITA101	MITA101	N	Set1	MITB404	MITB404	N	Set1	MITC701	MITC701	N
Set2	MITA205	MITA205	N	Set2	MITB505	MITB505	N	Set2	MITC802	MITC802	N
Set3	MITA305	MITA305	N	Set3	MITB603	MITB603	N	Set3	MITC905	MITC905	N
Set1	MITA104	MITA104	N	Set1	MITB403	MITB403	N	Set1	MITC705	MITC705	N
Set2	MITA202	MITA202	N	Set2	MITB502	MITB502	N	Set2	MITC805	MITC805	N
Set3	MITA303	MITA303	N	Set3	MITB601	MITB601	N	Set3	MITC901	MITC901	N
Set1	MITA105	MITA105	S	Set1	MITB401	MITB401	S	Set1	MITC702	MITC702	S
Set2	MITA204	MITA204	S	Set2	MITB503	MITB503	S	Set2	MITC801	MITC801	S
Set3	MITA301	MITA301	S	Set3	MITB602	MITB602	S	Set3	MITC904	MITC904	S
Set1	MITA102	MITA102	N	Set1	MITB405	MITB405	N	Set1	MITC703	MITC703	N
Set2	MITA201	MITA201	N	Set2	MITB504	MITB504	N	Set2	MITC804	MITC804	N
Set3	MITA302	MITA302	N	Set3	MITB604	MITB604	N	Set3	MITC903	MITC903	N

Within laboratory reproducibility: Lab A: 100% (5/5), Lab B: 100% (5/5), Lab C 100% (5/5) Between laboratory reproducibility: 100% (5/5)

添付資料 10. IL-1 Luc assay Phase II validation 試験結果

	LabA T	ohoku	LabB T	sukuba	LabC AIS	T Shikoku	Between-labolatory
Chem No.	Code No.	Judge	Code No.	Judge	Code No.	Judge	concodance or disconcodance
2	MTA117	S	MIB221	S	MTC305	S	concodance
3	MTA105	N	MIB220	N	MTC301	N	concodance
4	MTA120	Ν	MIB203	N	MTC318	S	disconcodance
5	MTA115	Ν	MIB211	N	MTC307	S	disconcodance
6	MTA111	N	MIB224	N	MTC302	N	concodance
7	MTA112	N	MIB208	N	MTC312	N	concodance
8	MTA125	S	MIB214	S	MTC303	S	concodance
11	MTA110	N	MIB218	N	MTC322	N	concodance
12	MTA124	S	MIB217	S	MTC313	S	concodance
13	MTA102	N	MIB206	N	MTC317	N	concodance
14	MTA121	N	MIB205	N	MTC324	N	concodance
15	MTA116	N	MIB223	N	MTC309	N	concodance
16	MTA118	Ν	MIB202	S	MTC316	Ν	disconcodance
17	MTA108	S	MIB204	S	MTC315	S	concodance
20	MTA113	S	MIB219	S	MTC323	S	concodance
22	MTA107	S	MIB222	S	MTC314	S	concodance
23	MTA119	Ν	MIB201	N	MTC306	S	disconcodance
25	MTA104	N	MIB210	N	MTC311	N	concodance
26	MTA114	S	MIB216	S	MTC304	S	concodance
27	MTA127	N	MIB227	N	MTC327	N	concodance
		Betwee	n-labolator	y concoda	nce rate		80% (16/20)

添付資料 11. IL-1 Luc assay, IL-2 Luc assay, IL-8 Luc assay data set

Chemicals	IL-2		IL-1β		IL-8 Luc
	Judge	LOEL (ug/mL)	Judge	LOEL (ug/mL)	Judge
FK506	S	0.00			N
Cyclosporine A	S	0.00			N
Actinomycin D	S	0.02		0.13	
Digoxin	S	0.07		0.59	
Colchicine	S	0.27			Р
FR167653	S	1.30	S	0.49	
Benzethonium chloride	S	1.63	N		Р
Mercuric chloride	S	1.95	S	1.95	Р
Chlorpromazine	S	1.95	S	3.91	Р
Dibutyl phthalate	S	2.60	S	15.63	N
Amphoterycin B	S	2.60	S	1.17	Р
2-Aminoanthracene	s	5.86	S	11.72	Р
sophorone diisocyanate	s	7.81	S	3.91	Р
Formaldehyde	s	7.81	N		Р
Pyrimethamine	s	7.81	N		Р
Cobalt chloride	S	16.93	s	46.88	Р
Cisplatin	s	16.93	s	46.88	Р
Chloroquine	S	17.83		39.06	
Minocycline	S	18.52		62.50	
Mitomycin C	S	20.00		12.00	P
Hydrogen peroxide	S	23.44		375.00	
Citral	s	25.00		4.88	
Dexamethasone	S	41.67		0.98	
Pentamidine isethionate	S	52.08		64.45	
Lead(II) acetate	S	57.29		04.43	N N
	S	58.48		41.55	
Azathioprine Diesel exhaust particles		62.50		39.06	
· · · · · · · · · · · · · · · · · · ·	S S				
Sodium dodecyl sulfate		62.50		62.50	
Dapsone	S	72.92		125.00	
o-Nitroaniline	S	83.33		125.00	
Nitrofurazone	S	83.33			P
Sulfasalazine	S	92.94		44.81	
Nickel sulfate	S	104.17		375.00	
Aluminum chloride	S	104.17			N
Chloroplatinicacid	S	250.00		23.44	
Diethanolamin	S	250.00		333.33	
Sodium bromate	S	500.00		500.00	
Histamine	S	750.00			Р
soniazid	S	1000.00	N		N
Triethanolamine	S	1333.33		1000.00	
Magnesium sulfate	S	2000.00	N		N
Warfarin	N		N		N
Hydrocortisone	N		N		N
Lithium carbonate	N		N		Р
2,4-Diaminotoluene	N		N		N
Dibenzopyrene	N		N		N
Cyclophosphamide	N		N		Р
Ethanol	N		N		N
Methanol	N		N		N
Hexach l orobenzene	N		N		N
Trichloroethylene	N		N		N
Methotrexate	N		N		Р
Rapamycin	N		N		N
Mizoribine	N		N		N
Mycophenolicacid	A	0.40		72.00	
2-Mercaptobenzothiazole	A	16.11		93.75	
Ribavirin	A	26.04		750.00	
Acetaminophen	A	100.00		755.00	N
Nicotinamide	A	288.07			N
1.00tillarilla0	^	200.07			. *

添付資料 12. IL-2 Luc assay Phase I, Phase II 化学物質の免疫毒性データーベース

					NTP data			
	Immunotoxicit	y classification	In vivo	Ex vivo		In vitro		
Chemical name	Classification	Rationale	sytem organ weight	cytokine production	TDAR	cytokine production	T cell proliferation	Mode of action
Phase I study						S (IL-2, 4, IFN-g)(H)		This compound then is proposed to modulate
Dibutyl phthalate	TTC	3), 4)	A (spleen)			A (IL-1b)(H) x 3 S (IL-1b)		cytokine secretion from both monocytes/macrophages and T cells.
Hydrocortisone	TTC	1)	S (thymus) x 2 S (spleen)		N	S (IFN-a)		
Lead(II) acetate	ттс	1)	A(thymus)		S N	S (IFN-g, IL-1b)(H) A (IL-4)(H)	S(H)	
Nickel(II) sulfate	ттс	1)	N S (thymus)		N	A (IL-4, IFN-g)(H) S (IL-2) S (IFN-g)		
dimethyldithiocarba mate (DMDTC) Phase II study	NTTC					S (IL-1b)	N(H)	
2.4-diaminotoluene	NTTC		N (spleen) A (spleen)		s	=	-	
Benzo(a)pyrene	TTC	2), 3)	A (spiecii)	S(IL-2)	S x 5	A (IL-4)(H) N (IFNγ)(H) N (IL-2)(H)	S (H) x 2 S x 6	Disruption of T-cell activities has been associated with B(a)P induced immunotoxic effects (Urso et al. 1986).
Cadmium Chloride	ттс	2), 3)	A (spleen) S (spleen)	A (IL-2) N (IFN-γ)	Sx4	S (IL-2, 4, IFN-g) A (IFN-g)(H) S (IL-2, IFN-g) A (IFN-g) S (IL-2) A (IL-2)	S	
Dibromoacetic acid (DBAA)	ттс	1), 4)	A (spleen) S (thymus) x 2		N	S (IL-2, 4)	s	Overall, studies suggest that DBAA produces immunotoxic effects through modulation of T-cell mediated cell immunity. T-cell apoptosis, through extrinsic and intrinsic pathways, are proposed to play a role in the mode of action.
Diethylstilbestrol (DES)	ттс	1), 2), 4)	S (thymus) x 4 A (thymus) x 2 A (spleen)	A (IFN-g) x 3	s	A (IL-1) A (IL-2)		DES exposure was associated with down-regulation of gene expression in the TCR complex, and the TCR and CD28 signaling pathways.
Diphenylhydantoin	ттс	2), 3), 4)		A (IL-4) S (IFN-γ, IL-2) S (IL-1α) N (IL-6, 12)	S Ax2	-	-	DPH treatment can lead to a decrease of suppressor T cells
Ethylene Dibromide (EDB)	ттс	1)	S (thymus) S (spleen) N		A	-	s	
Glycidol	NTTC		N		s	-	-	Studies suggest that glycidol modulates B-cell function, and NK cell and macrophage activities.111 and decreased cytotoxic T cell activity
Indomethacin	TTC	3), 4)	N A (spleen)		Sx3 Ax1	A (IL-2)(H) A (IFN-g)(H)	A (H) x 4 S A x 3	indomethacin inhibition of prostaglandin synthesis leads to altered T-cell function,
Isonicotinic Acid Hydrazide (IAH)	TTC	2)	N x 2			S (IL-2)(H) A (IL-2)(H) S (IL-1)(H)	S (H) x 3 A (H) x 6 A N	
Nitrobenzene	Undetermined		A (spleen) x 3 A (thymus) x 2		S N	-		effects on T-cell function may play a role in increased susceptibility to L. monocytogenes (Burns et al. 1994).
Urethane, Ethyl carbamate	ттс	1)	S (thymus) x2 S (spleen) x 2 N A (thymus) A (spleen)	N (IL-2)	Sx2 N	N (IL-2, 4, IFN-g)(H) A (IFN-g)(H) S (IFN-g)(H)	Nx2	
Tributyltin Chloride (TBTC)	TTC	1)	S (thymus) x4 S (spleen) x 3		N S	A (INF-g)(H) N (IL-2, 4)(H) S (IFN-g)(H)	S (H) S x 3	
Perflouorooctanoic Acid (PFOA)	TTC	1)	S (thymus) x2 S (spleen) x 2	N (IFN-g)		S (IL-4)(H) N (IL-2)(H)	A (H) S (H) N (H)	Direct modulation of NF-kB has been implicated in modulation of cytokine production and secretion (Corsini et al. 2012).
Dichloroacetic Acid (DCAA)	ттс	2), 3)	A(spleen)	N (IL-2) A (IFN-γ) x 3 S (IL-4) x 2 S (IL-2)	N	A (IL-2)(H) A (IL-2, IFN-g)		T-cell activation was one proposed mode of action for DCAA.
Toluene	NTTC		N		N S		N	
Acetonitrile	NTTC		S(thymus)		S	•	-	
Mannnitol Vanadium	NTTC		N				N (H)	
Pentoxide	NTTC		A (spleen)			N	N	
o-Benzyl-p- chlorophenol (BCP)	NTTC		N		47	-	-	

Appendix 8 Table. The summary of immunotoxicological data of 25 chemicals (continue

			vitro offcat an "	•	The dat		ed by the VMT				itro offort "	4	
		In	vitro effect on IL-	-2 		ln v	/itro effect on IFN-γ		In vitro effect on IIL-4				
Chemical name	Effect	Animal	in vitro (method)	References	Effect	Animal	in vitro (method)	References	Effect	Animal	in vitro (method)	References	
Phase I study Dibutyl phthalate					S	human	T cells (in vitro)	Hansen et al. 2015	S	human	T cells (in vitro)	Hansen et al. 2015 (0.0278~27.8 ug/mL)	
Hydrocortisone	s s	human	lymphocyte (in vitro) PBL (in vitro)	Chikanza and Panayi 1993 Goodwin et al. 1986									
		noman	i DE (iii iiii o)	OGGAMITOTAL 1000	s	mice	splenocyte (ex vivo)	Fernandez-Cabezudo et al. 2007	A no effect	mice	splenocyte (ex vivo)	Femandez- Cabezudo et al.	
Lead(II) acetate					no effect S	mice human	cell line (EL-4) PBMC	Wagner et al. 2006 Hemdan et al. 2005	A A	mice human rat	cell line (EL-4) PBMC (in vitro) ?	2007 Wagner et al. 2006 Hemdan et al. 2005 Chen et al. 2004	
Nickel(II) sulfate					A A (NiCl2) A A	mice mice human rat	spleen cell (in vitro) cell line (EL-4) PBMC (in vitro) lymphoid lung cell (ex vivo)	Kim et al. 2009 Wagner et al. 2006 Thomas et al. 2003 Goutet et al. 2000	A, S A (NiCl2) A	mice mice human	spleen cell (in vitro) cell line (EL-4) PBMC (in vitro)	Kim et al. 2009 Wagner et al. 2006 Thomas et al. 2003	
dimethyldithiocarba mate (DMDTC)													
Phase II study								+					
2.4-diaminotoluene													
Benzo(a)pyrene													
Cadmium Chloride					N (ex vivo), A (in vitro) S S (IC50=7.05E- 05 M) S	rat rat human mice	splenocyte (ex vivo, in vitro) spleen cell (ex vivo) PBMC (in vitro) thymocyte, splenocyte (in vitro)	Wang et al. 2017 Demenesku et al. 2014 Kooijiman et al. 2010 Pathak and Khandelwal 2008	no effect	rat	spleen cell (ex vivo)	Demenesku et al. 2014	
Dibromoacetic acid (DBAA)													
Diethylstilbestrol (DES)													
Diphenylhydantoin													
Ethylene Dibromide (EDB)													
Glycidol													
Indomethacin		human	PBMC (in vitro), cell	Tsuboi et al. 1995									
Isonicotinic Acid Hydrazide (IAH)	A		line (Jurkat)										
Nitrobenzene													
Urethane, Ethyl carbamate													
Tributyltin Chloride (TBTC)					no effect (TBTO)	mice	cell line (EL-4)	Ringerike et al. 2005					
Perflouorooctanoic Acid (PFOA)													
Dichloroacetic Acid (DCAA)													
Toluene													
Acetonitrile													
Mannitol Vanadium Pentoxide													
o-Benzyl-p- chlorophenol (BCP)			et, (H) humana study,										

S: Suppression, A: Augumentation, N: No effect, (H) humana study,

^{#:} The criterion number used to define immunotoxicity

- 引用文献の記されていないデータは NTP の好意により作成して頂いた免疫毒性 データベースに基づいている(昨年度の成果報告書に記載)。引用文献が書か れている文献は以下の通りである。
 - Chen, S., Golemboski, K., Piepenbrink, M., et al., 2004. Developmental immunotoxicity of lead in the rat: influence of maternal diet. J Toxicol Environ Health A 67, 495-511.
 - Chikanza, L.C., Panayi, G.S., 1993. The effects of hydrocortisone on in vitro lymphocyte proliferation and interleukin-2 and -4 production in corticosteroid sensitive and resistant subjects. Eur J Clin Invest 23, 845-850.
 - Demenesku, J., Mirkov, I., Ninkov, M., et al., 2014. Acute cadmium administration to rats exerts both immunosuppressive and proinflammatory effects in spleen.

 Toxicology 326, 96-108.
 - Fernandez-Cabezudo, M.J., Ali, S.A., Ullah, A., et al., 2007. Pronounced susceptibility to infection by Salmonella enterica serovar Typhimurium in mice chronically exposed to lead correlates with a shift to Th2-type immune responses. Toxicol Appl Pharmacol 218, 215-226.
 - Goodwin, J.S., Atluru, D., Sierakowski, S., et al., 1986. Mechanism of action of glucocorticosteroids. Inhibition of T cell proliferation and interleukin 2 production by hydrocortisone is reversed by leukotriene B4. J Clin Invest 77, 1244-1250.
 - Goutet, M., Ban, M., Binet, S., 2000. Effects of nickel sulfate on pulmonary natural immunity in Wistar rats. Toxicology 145, 15-26.
 - Hansen, J.F., Nielsen, C.H., Brorson, M.M., et al., 2015. Influence of phthalates on in vitro innate and adaptive immune responses. PLoS One 10, e0131168.
 - Hemdan, N.Y., Emmrich, F., Adham, K., et al., 2005. Dose-dependent modulation of the in vitro cytokine production of human immune competent cells by lead salts. Toxicol Sci 86, 75-83.
 - Iavicoli, I., Marinaccio, A., Castellino, N., et al., 2004. Altered cytokine production in mice exposed to lead acetate. Int J Immunopathol Pharmacol 17, 97-102.
 - Kim, J.Y., Huh, K., Lee, K.Y., et al., 2009. Nickel induces secretion of IFN-gamma by splenic natural killer cells. Exp Mol Med 41, 288-295.
 - Kooijman, R., Devos, S., Hooghe-Peters, E., 2010. Inhibition of in vitro cytokine production by human peripheral blood mononuclear cells treated with

- xenobiotics: implications for the prediction of general toxicity and immunotoxicity. Toxicol In Vitro 24, 1782-1789.
- Metushi, I.G., Uetrecht, J., 2014. Isoniazid-induced liver injury and immune response in mice. J Immunotoxicol 11, 383-392.
- Pathak, N., Khandelwal, S., 2008. Comparative efficacy of piperine, curcumin and picroliv against Cd immunotoxicity in mice. Biometals 21, 649-661.
- Ringerike, T., Ulleras, E., Volker, R., et al., 2005. Detection of immunotoxicity using T-cell based cytokine reporter cell lines ("Cell Chip"). Toxicology 206, 257-272.
- Thomas, P., Barnstorf, S., Summer, B., et al., 2003. Immuno-allergological properties of aluminium oxide (Al2O3) ceramics and nickel sulfate in humans.

 Biomaterials 24, 959-966.
- Tsuboi, I., Tanaka, H., Nakao, M., et al., 1995. Nonsteroidal anti-inflammatory drugs differentially regulate cytokine production in human lymphocytes: up-regulation of TNF, IFN-gamma and IL-2, in contrast to down-regulation of IL-6 production. Cytokine 7, 372-379.
- Wagner, W., Walczak-Drzewiecka, A., Slusarczyk, A., et al., 2006. Fluorescent Cell Chip a new in vitro approach for immunotoxicity screening. Toxicol Lett 162, 55-70.
- Wang, P., Wang, J., Sun, Y.J., et al., 2017. Cadmium and chlorpyrifos inhibit cellular immune response in spleen of rats. Environ Toxicol 32, 1927-1936.

添付資料 13. IL-2 data set 化学物質の免疫毒性データベース

Appendix 9 Table . The summary of immunotoxicological data of 60 chemicals in the IL-2 Luc assay data set.

Chemical name		classification		Thymus				Ex Vivo effect on ex vivo			
Chemical name	Classification	Rationale*	Weight	Animal	Reference	Effect	Animal	(method)	Reference		
FK506	ттс	1,3	decrease decrease	rat rat	Nalesnik et al. 1987 Takai et al. 1990						
Cyclosporine A	TTC	1,3	decrease no effect decrease decrease	mice mice rat mice	Auli et al. 2012 Kanariou et al. 1989 Beschorner et al. 1987 Hattori et al. 1987						
Actinomycin D	TTC	3									
Digoxin	ттс	2, 3									
Colchicine	ттс	2,3				A	human	PBMC (ex vivo)	Freed et al. 1989		
FR167653	Undetermined	2, 3									
Benzethonium chloride	Undetermined	1	decrease		National Toxicology Program 1995						
Mercuric chloride	ттс	1,3	decrease	mice	Dieter et al. 1983						
Chlorpromazine	ттс	1,3	decrease decrease	mice rat	Auli et al. 2012 Silvestrini et al. 1967						
Amphotericin B	Undetermined	1	decrease	mice	Blanke et al. 1977						
Dibutyl phthalate	TTC	3	no effect no effect	rat rat	Zhang et al. 2013 Salazar et al. 2004						
2-Aminoanthracene	Undetermined		no onocc	iat							
Formaldehyde	TTC	2,3	no effect	rat	Vargova et al. 1993						
Pyrimethamine	Undetermined										
sophorone diisocyanate	Undetermined										
Cisplatin	ттс	1,2,3	decrease decrease	mice mice	Kouchi et al. 1996 Sugiyama et al. 1995	S	mice	Spleen cell (ex vivo)	Kim et al. 2019		
Cobalt chloride	TTC	1, 3	decrease	rat	Chetty et al. 1979						
Chloroquine	ттс	1,3	decrease	human	Garly et al. 2008						
Minocycline	TTC	3									
Mitomycin C	Undetermined										
Hydrogen peroxide	TTC	3									

Appendix 9 Table . The summary of immunotoxicological data of 60 chemicals in the IL-2 Luc assay data set.(continue)

Chemical name	=		In vitro effect on IL-2				n vitro effeon on IFN-	
	Effect	Animal	in vitro (method)	Reference	Effect	Animal	in vitro (method)	Reference
	s s	mice rat	cell line (EL-4) primary astrocyte cell (in vitro)	Wagner et al. 2006 Gabryel et al. 2004	S	mice	cell line (EL-4)	Wagner et al. 2006
FK506	S S	human human	cell line (Jurkat, Hut-	Henderson et al. 1991 Yoshimura et al. 1989				
			PBMC					
	S	mice	cell line (3A9 Tcell hybridoma)	Lehmann and Williams 2018	IC50=5.00E-08 M	human mice	PBMC (in vitro) cell line (EL-4)	Kooijman et al. 2010 Wagner et al. 2006
	S	mice	cell line (EL-4)		S	mice	cell line (EL-4)	Ringerike et al. 200
Cyclosporine A	S	rat	primary astrocyte cell	Ringerike et al. 2005	S			
	s	human	(in vitro) cell line (Jurkat, Hut-	Gabryel et al. 2004				
			78)	Henderson et al. 1991				
Actinomycin D	S	mice	cell line (EL-4)	Wagner et al. 2006	no effect	mice	cell line (EL-4)	Wagner et al. 2006
	S	human	PBMC (in vitro)	Wang et al. 1984				
	S	human	cell line (HepG2),	Karas et al. 2018, He	S (ex vivo), no		spleen cell (ex vivo, in	
Digoxin	no effect S	human	Th17 cell, thymocytes PBMC (in vitro) PBMC (in vitro)	et al. 1998 Sheikhi et al. 2007	effect (in vitro) S (IC50=4.31E- 07 M)	human	vitro) PBMC (in vitro)	Kooijman et al. 201
		IIdiliali	DIVIO (III VILIO)	Gentile et al. 1997	07 101)			
	А	human	cell line (Jurkat)	Dupuis et al. 1993	N	human	PBMC (in vitro)	Kooijman et al. 201
					(IC50>5.00E-			
					04 M(=200	mice	spleen cell (in vitro)	Sosroseno 2009
Colchicine					ug/mL))	human	PBMC (in vitro)	Tzortzaki et al. 200
					S (in vitro)	human		Altindag et al. 1997
					A			
					S			
ED407050	no effect	mice	cell line (EL-4)	Wagner et al. 2006	no effect	mice	cell line (EL-4)	Wagner et al. 2006
FR167653	no effect	human	lymphocyte (in vitro)	Yamamoto et al. 1996	S	mice	spleen cell (ex vivo)	Ando et al. 2004
	no effect	mice	cell line (EL-4)	Wagner et al. 2006	no effect no effect	human mice	lymphocyte (in vitro) cell line (EL-4)	Yamamoto et al. 19
Benzethonium chloride	no enect	mice	cell lille (EL-4)	wagner et al. 2000	no enect	mice	cell lille (EL-4)	Wagner et al. 2006
	S	mice	plasma (in vivo)	Santarelli et al. 2006	S	human	PBMC (in vitro)	Kooijman et al. 201
	no effect	mice	cell line (EL-4)	Wagner et al. 2006	(IC50=3.06E-	mice	cell line (EL-4)	Wagner et al. 2006
Mercuric chloride	Α	mice	spleen cell	Hu et al. 1997	` 06 M)	mice	cell line (EL-4)	Ringerike et al. 200
			'		A		, ,	
					S			
	Α	human	whole blood (in vitro)	Himmerich et al. 2011	S	human	thymocytes (in vitro)	Schleuning et al. 19
	S	rat	mixed glial and	Labuzek et al. 2005	S	mice	Spleen cell (in vitro)	Johnson et al. 1985
Chlorpromazine			microglial cell cultures					
			(in vitro)					
Amphotericin B	S	human	thymocytes (in vitro)	Schleuning et al. 1989				
•	S	human	T cell (in vitro)	Hansen et al. 2015	S	human	T cells (in vitro)	Hansen et al. 2015
Dibutyl phthalate	_		()				()	
2-Aminoanthracene	А	mice	cell line (EL-4)	Wagner et al. 2006	А	mice	cell line (EL-4)	Wagner et al. 2006
F							T cell (in vitro)	Sasaki et al. 2009
Formaldehyde					protein)	mice	spleen cell (ex vivo)	Fujimaki et al. 2004
	A	mice	cell line (EL-4)	Wagner et al. 2006	A no effect	mice	cell line (EL-4)	Wagner et al. 2006
Pyrimethamine	no effect	human	lymphocyte (in vitro)	Bygbjerg et al. 1987	011000		(LL ¬)	
,	(<loel)< td=""><td></td><td></td><td>30, 3</td><td></td><td></td><td></td><td></td></loel)<>			30, 3				
ophorone diisocyanate					no effect	mice	Lymph node (ex vivo)	Selgrade et al. 200
,	no offert	mine	coll line (EL 4)	Wagner et al. 2006		minc	Sploop cell (av vive)	Kim et al. 2010
	no effect	mice	cell line (EL-4)	Wagner et al. 2006	S	mice	Spleen cell (ex vivo)	Kim et al. 2019
Cisplatin	(<loel)< td=""><td>human</td><td>PBL (in vitro)</td><td>Riesbeck 1999</td><td>А</td><td>mice</td><td>cell line (EL-4)</td><td>Wagner et al. 2006</td></loel)<>	human	PBL (in vitro)	Riesbeck 1999	А	mice	cell line (EL-4)	Wagner et al. 2006
	A S	human	PBL (in vitro)	Sfikakis et al. 1996				
0-1-11-11-11	S	mice	cell line (EL-4)	Wagner et al. 2006	A	mice	cell line (EL-4)	Wagner et al. 2006
Cobalt chloride								
Chloroquine	S	human	Synovial T cell clones	Landewe et al. 1995	A S	mice human	? (ex vivo) T cell clone	Rosa et al. 1999 Landewe et al. 199
	S	human	PBMC (in vitro)	Maeda et al. 2010	no effect	mice	splenocyte (ex vivo)	Chen et al. 2010
Minocycline	S	human	T cell clones (in vitro)	Kloppenburg et al.				
•			'	1995	S	human	T cell clones (in vitro)	Kloppenburg et al.
	no effect	mice	cell line (EL-4)	Wagner et al. 2006	no effect	mice	cell line (EL-4)	Wagner et al. 2006
Mitomycin C	(<loel)< td=""><td>human</td><td>mononuclear</td><td>Roche et al. 1988</td><td></td><td></td><td></td><td></td></loel)<>	human	mononuclear	Roche et al. 1988				
	S		leukocyte (in vitro)					
Hydrogen peroxide	А	mice	cell line (EL-4)	Wagner et al. 2006	Α	mice	cell line (EL-4)	Wagner et al. 2006
	s	human	PBMC (in vitro)	Freed et al. 1987				
Try arogon poroxido	1 3		,					

			In vitro effect on IL	-4	
Chemical name	Effect	Animal	in vitro (method)	Reference	
	S	mice	cell line (EL-4)	Wagner et al. 2006	
FK506					
	S	mice	cell line (EL-4)	Wagner et al. 2006	
	S S	mice human	cell line (EL-4) cell line (D10.G4.1)	Ringerike et al. 2005 Schmidt et al. 1994	
Cyclosporine A					
	A	mino	coll line (EL 4)	Wagner et al. 2006	
Actinomycin D	_ ^	mice	cell line (EL-4)	wagner et al. 2006	
Digoxin					
	A (in vitro)	mice	spleen cell (in vitro)	Sosroseno 2009	
Colchicine					
		unia a	sall line (FL 4)	Managar et al. 2006	
FR167653	S no effect	mice mice	cell line (EL-4) spleen cell (ex vivo)	Wagner et al. 2006 Ando et al. 2004	
	A	mice	cell line (EL-4)	Wagner et al. 2006	
enzethonium chloride	_ ^	mice	Cell lille (LL-4)	wagner et al. 2000	
	A	mice	cell line (EL-4)	Wagner et al. 2006	
Mercuric chloride					
	S	mice	splenic lymphocyte (Pei et al. 2014	
Chlorpromazine	A	human	in vitro) whole blood (in vitro)	Himmerich et al. 2011	
·			, ,		
Amphotericin B					
Dibutyl phthalate	S	human	T cells (in vitro)	Hansen et al. 2015	
2-Aminoanthracene	A	mice	cell line (EL-4)	Wagner et al. 2006	
Campa al dalah i ida	no effect	human	T cell (in vitro)	Sasaki et al. 2009	
Formaldehyde					
Pyrimethamine	А	mice	cell line (EL-4)	Wagner et al. 2006	
r ymmethamme					
ophorone diisocyanate					
	A no effect	mice mice	Spleen cell (ex vivo) cell line (EL-4)	Kim et al. 2019 Wagner et al. 2006	
Cisplatin	no enect	IIIICE	Cell lille (LL-4)	wagner et al. 2000	
	A	mice	cell line (EL-4)	Wagner et al. 2006	
Cobalt chloride		IIIICC		Wagner et al. 2000	
Chloroquine	no effect	mice	? (ex vivo)	Rosa et al. 1999	
	no effect	mice	splenocyte (ex vivo)	Chen et al. 2010	
Minocycline					
	no effect	mice	cell line (EL-4)	Wagner et al. 2006	
Mitomycin C					
Hydrogen peroxide	А	mice	cell line (EL-4)	Wagner et al. 2006	
, arogon peroxide	1				

Appendix 9 Table . The summary of immunotoxicological data of 60 chemicals in the IL-2 Luc assay data set.

Chemical name	IIIIIIIIIIIIII	classification		Thymus	weight					
Chemical name	Classification	Rationale*	Weight	Animal	Reference	Effect	Animal	ex vivo (method)	Reference	
Citral	Undetermined	1	decrease decrease	rat rat, mice	Ress et al. 2003 National Toxicology Program 2003					
Dexamethasone	TTC	1,3	decrease decrease decrease	mice mice rat	Auli et al. 2012 Munson et al. 1982 Exon et al. 1986					
Pentamidine isethionate	TTC	3								
Lead(II)acetate	ттс	1, 3	increase	rat	Bunn et al. 2001	no effect	rat	spleen cell (ex vivo) spleen cell (ex vivo)	Bunn et al. 2001 Miller et al. 1998	
Azathioprine	TTC	1,2, 3	decrease decrease	rat rat	De Waal et al. 1995 Vos and Van Loveren 1994	S S	mice, rat	lymphocyte, thymocyte (in vitro, ex vivo) PBMC (ex vivo)	Meredith and Scott 199 Dupont et al. 1985	
Diesel exhaust particle	TTC	1, 3	decrease	rat	Tsukue et al. 2001					
Sodium dodecyl sulfate	ттс	3								
Dapsone	ттс	3	No Effect	mice	https://ntp.nieh s.nih.gov/testing /types/imm/abs tracts/imm9001 5/index.html					
Nitrofurazone	NTTC		No Effect	mice	https://ntp.nieh s.nih.gov/testing /types/imm/abs tracts/imm9001 1/index.html					
p-Nitroaniline	TTC	1,3	increase,	mice	National Toxicology Program 1993b					
Sulfasalazine	TTC	1,3	decrease	rat	National Toxicology Program 1997					
Aluminium chloride	TTC	1,3	diminishe d thymic cellularity	mice	Synzynys et al. 2004					
Nickel sulfate	ттс	1, 3	no effect decrease decrease	rat	Knight et al. 1991 Haley et al. 1990 National Toxicology Program 1996					
Hydrocortisone	ттс	1,3	decrease decrease (PND 21), increase (PND 42)	mice rat	Van Dijk et al. 1979 El Fouhil et al. 1993a, El Fouhil et al.1993b, El Fouhil and Turkall 1993					
Diethanolamine	Undetermined	1	decrease	mice	https://ntp.niehs .nih.gov/testing/ types/imm/abstrac ts/imm20004/imm20 004.html					
Chloroplatinic acid	Undetermined				X					
Sodium bromate	Undetermined	1	No Effect	mice	https://ntp.niehs .nih.gov/testing/ types/imm/abstrac ts/imm98004/index .html					

Appendix 9 Table . The summary of immunotoxicological data of 60 chemicals in the IL-2 Luc assay data set.(continue)

			In vitro effect on IL-2				n vitro effeon on IFN-	
	Effect	Animal	in vitro (method)	Reference	Effect	Animal	in vitro (method)	Reference
Citral								X
	S	mice	cell line (3A9 Tcell	Lehmann and	S	human	PBL (in vitro)	Arya et al. 1984
			hybridoma)	Williams 2018	S	human	T cell (in vitro)	Reen and Yeh 1984
Dexamethasone	no effect	mice	cell line (EL-4)		S	mice	T cell clone (in vitro)	Kelso and Munck 19
	S	human	CBMC, PBMC (in	Wagner et al. 2006	S	mice	splenocyte (ex vivo)	Kunicka et al. 1993
			vitro)	Bessler et al. 1996	no effect	mice	cell line (EL-4)	Wagner et al. 2006
	s	mice	cell line (EL-4)	Ringerike et al. 2005	Α	mice	cell line (EL-4)	Wagner et al. 2006
Pentamidine isethionate	no effect	mice	cell line (EL-4)	Wagner et al. 2006	S	mice	cell line (EL-4)	Ringerike et al. 200
	no effect	human	whole blood (in vitro)	Van Wauwe et al.				
	(<loel)< td=""><td></td><td></td><td>1996</td><td></td><td></td><td></td><td></td></loel)<>			1996				
	S	mice	cell line (EL-4)	Wagner et al. 2006	S	mice	splenocyte (ex vivo)	Fernandez-Cabezu
								et al. 2007
Lead(II)acetate					no effect	mice	cell line (EL-4)	Wagner et al. 2006
,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,					S	human	PBMC	Hemdan et al. 2005
	S	mice	cell line (3A9 Tcell	Lehmann and	S	human	PBMC (ex vivo)	Weimar et al. 1995
			hybridoma)	Williams 2018	S	human	PBMC (ex vivo)	Dupont et al. 1985
Azathioprine	S	mice,	lymphocyte,					
		rat	thymocyte (in vitro, ex	Meredith et al. 1994				
			vivo)					
Diesel exhaust particle	Α	mice	cell line (EL-4)	Wagner et al. 2011	S	human	T cell (in vitro)	Sasaki et al. 2009
•	S	mice	cell line (EL-4)	Ringerike et al. 2005	S	human	PBMC (in vitro)	Kooijman et al. 201
Sodium dodecyl sulfate	1	111100	5511 IIII (LL-7)	gormo et al. 2000	(IC50=1.61E-	mice	cell line (EL-4)	Ringerike et al. 200
Juliani adadoyi bullate					04 M)		331 mio (EE 7)	go.inc ot al. 200
	S, A	mice	cell line (EL-4)	Wagner et al. 2006	S, A	mice	cell line (EL-4)	Wagner et al. 2006
	S	mice	splenocyte (in vitro)	Peterson et al. 1997	J 5,,, 1			
Dapsone			opionosyte (mae)					
	А	mice	cell line (EL-4)	Wagner et al. 2006	no effect	mice	cell line (EL-4)	Wagner et al. 2006
Nitrofurazone								
p-Nitroaniline	А	mice	cell line (EL-4)	Wagner et al. 2006	А	mice	cell line (EL-4)	Wagner et al. 2006
	s	mice	splenocyte (in vitro)	Fujiwara et al. 1990	S	human	BAL cell (in vitro)	Dobis et al. 2010
Sulfasalazine		111100	opionocyte (iii viiio)	r ajiwara ot ali. 1000	A	rat	CNS (in vivo)	Correale et al. 1991
	S	rat	lymphocyte (in vitro)	She et al. 2012		1	()	
Aluminium chloride								
	S (NiCl ₂)	human	Cell line (Jurkat)	Saito et al. 2011	Α	mice	spleen cell (in vitro)	Kim et al. 2009
	Α Α	mice	spleen cell (in vitro)	Kim et al. 2009	A (NiCl2)	mice	cell line (EL-4)	Wagner et al. 2006
Aliakal aute-4-	A (NiCl ₂)		cell line (EL-4)	Wagner et al. 2006	Α		PBMC (in vitro)	Thomas et al. 2003
Nickel sulfate	\			3 4 4 4 4 4 4 4 4	A	rat	lymphoid lung cell (ex	
							vivo)	
	+	human	lymphocyte (in vitro)	Chikanza and Panayi				
	1			1993	I			
	s	human	PBL (in vitro)	1993				
	S S	human human	PBL (in vitro) lymphocyte (in vitro)	Goodwin et al. 1986				
Hydrocortisone	1		' '					
Hydrocortisone	S	human	lymphocyte (in vitro)	Goodwin et al. 1986 Palacios and				
Hydrocortisone	s s	human	lymphocyte (in vitro)	Goodwin et al. 1986				
Hydrocortisone	s s	human	lymphocyte (in vitro)	Goodwin et al. 1986 Palacios and Sugawara 1982				
Hydrocortisone	s s	human	lymphocyte (in vitro)	Goodwin et al. 1986 Palacios and Sugawara 1982				
Hydrocortisone Diethanolamine	s s	human	lymphocyte (in vitro)	Goodwin et al. 1986 Palacios and Sugawara 1982 Northoff et al. 1980				
Diethanolamine	s s	human	lymphocyte (in vitro)	Goodwin et al. 1986 Palacios and Sugawara 1982 Northoff et al. 1980				
	s s	human	lymphocyte (in vitro)	Goodwin et al. 1986 Palacios and Sugawara 1982 Northoff et al. 1980				
Diethanolamine	s s	human	lymphocyte (in vitro)	Goodwin et al. 1986 Palacios and Sugawara 1982 Northoff et al. 1980				
Diethanolamine Chloroplatinic acid	s s	human	lymphocyte (in vitro)	Goodwin et al. 1986 Palacios and Sugawara 1982 Northoff et al. 1980				
Diethanolamine	s s	human	lymphocyte (in vitro)	Goodwin et al. 1986 Palacios and Sugawara 1982 Northoff et al. 1980				
Diethanolamine Chloroplatinic acid	s s	human	lymphocyte (in vitro)	Goodwin et al. 1986 Palacios and Sugawara 1982 Northoff et al. 1980				
Diethanolamine Chloroplatinic acid	S S S	human	lymphocyte (in vitro) PBMC (in vitro)	Goodwin et al. 1986 Palacios and Sugawara 1982 Northoff et al. 1980				

			In vitro effect on IL	-A	
Chemical name		Aurimont			
	Effect	Animai	in vitro (method)	Reference	
Citral					
Dexamethasone	A S S	mice human mice	cell line (EL-4) cell line (D10.G4.1) splenocyte (ex vivo)	Wagner et al. 2006 Schmidt et al. 1994 Kunicka et al. 1993	
Dexamethasone					
Pentamidine isethionate	A S	mice mice	cell line (EL-4)	Wagner et al. 2006 Ringerike et al. 2005	
	А	mice	splenocyte (ex vivo)	Fernandez-Cabezudo et	
Lead(II)acetate	no effect A A	mice human rat	cell line (EL-4) PBMC (in vitro) ?	al. 2007 Wagner et al. 2006 Hemdan et al. 2005 Chen et al. 2004	
Azathioprine					
Diocal aybayat =#-1-	no effect	human	T cell (in vitro)	Sasaki et al. 2009	
Diesel exhaust particle	IN GIRCE	naman	. oon (at vido)		
Sodium dodecyl sulfate					
	S	mice	cell line (EL-4)	Wagner et al. 2006	
Dapsone					
	no effect	mice	cell line (EL-4)	Wagner et al. 2006	
Nitrofurazone					
p-Nitroaniline	А	mice	cell line (EL-4)	Wagner et al. 2006	
Sulfasalazine	S	mice	mesangial cell (in vitro)	Tsai et al. 2000	
Aluminium chloride			,		
	A, S	mice	spleen cell (in vitro)	Kim et al. 2009	
Nickel sulfate	A (NiCl2) A	mice human	cell line (EL-4) PBMC (in vitro)	Wagner et al. 2006 Thomas et al. 2003	
Hydrocortisone					
Diethanolamine					
Chloroplatinic acid					
Sodium bromate					

Appendix 9 Table . The summary of immunotoxicological data of 60 chemicals in the IL-2 Luc assay data set.(continue)

Chemical name	<u> </u>	/ classification		,	weight	Ex Vivo effect on IL-2					
	Classification	Rationale#	Weight	Animal	Reference	Effect	Animal	ex vivo (method)	Reference		
Histamine	TTC	3									
Isoniazid	NTTC	1	No Effect	mice	https://ntp.niehs .nih.gov/testing/ types/imm/abstrac ts/imm96002/index .html						
Triethanolamine	Undetermined										
Magnesium sulfate	Undetermined										
Rapamycin	ттс	1, 3	decrease	rat	Lu et al. 2015						
	Undetermined										
Mizoribine											
Warfarin	TTC	3									
2,4-Diaminotoluene	NTTC	1	No Effect	mice	https://ntp.niehs .nih.gov/testing/ types/imm/abstrac ts/imm87034/index .html						
Cyclophosphamide	ттс	1	decrease decrease decrease	mice mice rat	Auli et al. 2012 https://ntp.niehs.ni h.gov/testing/types/ imm/abstracts/imm 90015/index.html Exon et al. 1986	S	mice	splenocyte (ex vivo)	Tabi et al. 1988		
Dibenzopyrene	Undetermined	3									
Ethanol	TTC	1, 3	decrease	mice	Kim and Park 2002						
Hexachlorobenzene	Undetermined	1,2	no effect decrease cortical atrophy	rat mice monkey	Vos et al. 1979 Loose et al. 1978 latropoulos et al. 1976	A A	rat	spleen cell (ex vivo) spleen cell (ex vivo)	Ezendam et al. 2004 Vandebriel et al. 1998		
Lithium carbonate	ттс	1,3	decrease	mice	https://ntp.niehs.ni h.gov/testing/types/ imm/abstracts/imm 85001/index.html						
Methanol	NTTC	1	decrease	rat	Parthasarathy et al. 2005						
Methotrexate	TTC	3									
Dimethyl sulfoxide	NTTC	1,3	no effect	mice	Caren et al. 1985						

Appendix 9 Table . The summary of immunotoxicological data of 60 chemicals in the IL-2 Luc assay data set.(continue)

Chemical name								
	Effect	Animal	in vitro (method)	Reference	Effect	Animal	in vitro (method)	Reference
Histamine	S S A, S	mice human mice	splenocyte (in vitro) PBMC (in vitro) spleen cell (in vitro)	Poluektova et al. 1999 Huchet and Grandjon 1988 Khan et al. 1985	no effect	mice	serum (in vivo)	Metushi and Uetrec 2014
Isoniazid	S (13.7, 137.1 ug/mL), A (0.0137~1. 37 ug/mL)	human	T cell (in vitro)	Kucharz and Sierakowski 1990				
Triethanolamine	,			X				
Magnesium sulfate	A, S A (0.0009ug/ mL), S (0.457ug/m L)	mice rat	cell line (EL-4) primary astrocyte cell (in vitro)	Ringerike et al. 2005 Gabryel et al. 2004	no effect	mice	cell line (EL-4)	Ringerike et al. 200
	S S	human	T cell (in vitro) cell line (Jurkat, Hut- 78)	Hanke et al. 1992 Henderson et al. 1991				
Mizoribine	S (>LOEL)	mice human	T cells (in vitro) peripheral blood T	Song et al. 2006 Turka et al. 1991				
	S	human	cells (in vitro) T cell (in vitro)	Bruserud and Lundin 1	S (IC50=3.16E-	human	PBMC (in vitro)	Kooijman et al. 201
Warfarin								
2,4-Diaminotoluene				X X				×
Cyclophosphamide	no effect (needs metabolizati on)	mice	cell line (3A9 Tcell hybridoma)	Lehmann and Williams 2018				
Dibenzopyrene	A	mice	cell line (EL-4)	Wagner et al. 2006	А	mice	cell line (EL-4)	Wagner et al. 2006
Ethanol	S	human	cell line (Jurkat), primary CD4+ T lymphocytes (in vitro)	Ghare et al. 2011	N (IC50>1.00E- 03 M)	human	PBMC (in vitro)	Kooijman et al. 201
Hexachlorobenzene					N (IC50>1.00E- 05 M)	human	PBMC (in vitro)	Kooijman et al. 201
Lithium carbonate	A A A	human human human	PBMC (in vitro) PBMC (in vitro) PBMC (in vitro)	Wilson et al. 1989 Parenti et al. 1988 Sztein et al. 1987	N (IC50>1.00E- 03 M)	human	PBMC (in vitro)	Kooijman et al. 201
Methanol			Wagner et al. 2006	N (IC50>1.00E- 03 M) no effect	human mice	PBMC (in vitro) cell line (EL-4)	Kooijman et al. 201 Wagner et al. 2006	
Methotrexate	S A	mice human	cell line (3A9 Tcell hybridoma) PBMC (in vitro)	Lehmann and Williams 2018	1.0 311000			
Dimethyl sulfoxide	S, A mice cell line (EL-4) Wag no effect (1 human PBMC (in vitro) de A		Cesario et al. 1984 Wagner et al. 2006 de Abreu Costa et al. 2017	no effect	mice	cell line (EL-4)	Wagner et al. 2006	

			In vitro effect on II	4	
Chemical name	Effect	Animal	in vitro (method)	Reference	
Histamine					
Isoniazid					
Triethanolamine					
Magnesium sulfate	S	mice	cell line (EL-4)	Ringerike et al. 2005	
Rapamycin					
Mizoribine					
Warfarin					
2,4-Diaminotoluene				Х	
Cyclophosphamide					
Dibenzopyrene	A	mice	cell line (EL-4)	Wagner et al. 2006	
Ethanol					
Hexachlorobenzene					
Lithium carbonate					
Methanol	A	mice	cell line (EL-4)	Wagner et al. 2006	
Methotrexate	no effect	human	cell line (D10.G4.1)	Schmidt et al. 1994	
Dimethyl sulfoxide	А	mice	cell line (EL-4)	Wagner et al. 2006	

	Immunotoxicity	classification		Thymus	weight	Ex Vivo effect on IL-2				
Chemical name	Classification	Rationale*	Weight	Animal	Reference	Effect	Animal	ex vivo (method)	Reference	
	NTTC	1	No Effect	mice, rat	https://ntp.niehs					
					.nih.gov/testing/					
					types/imm/abstrac					
					ts/imm20006/imm20					
Trichloroethylene					006. html					
					https://ntp.niehs					
					.nih.gov/testing/					
					types/imm/abstrac					
					ts/imm96007/imm96					
	Undetermined	4.0			007. html					
Mycophenolic acid	Undetermined	1, 3	decrease	rat	Pally et al. 2001					
Mercaptobenzothiazole	Undetermined									
	TTC	1, 3	decrease	mice	https://ntp.niehs					
					.nih.gov/testing/					
Ribavirin					types/imm/abstrac					
					ts/imm90010/index					
					. html					
Nicotinamide	Undetermined		. .		15 1 2000					
	Undetermined		no effect	mice	Kim and Park 2002					
Acataminanhan				rat, mice	National Toxicology					
Acetaminophen			(rat), no effect		Program 1993a					
			errect (mice)							
uppression, A: Augumenta	1 N N 00 1	an i								

#: The criterion number used to define immunotoxicity

Appendix 9 Table . The summary of immunotoxicological data of 60 chemicals in the IL-2 Luc assay data set.(continue)

			In vitro effect on IL-2	2	In vitro effeon on IFN-γ					
Chemical name	Effect	Animal	in vitro (method)	Reference	Effect	Animal	in vitro (method)	Reference		
Trichloroethylene										
Mycophenolic acid	no effect no effect	human mice	PBL (in vitro) spleen cell (in vitro)	Quemeneur et al. 2002 Lemster et al. 1992						
2-Mercaptobenzothiazole										
Ribavirin	A A	human human	PBMC (in vitro) T cells (in vitro)	Sookoian et al. 2004 Tam et al. 1999						
Nicotinamide										
Acetaminophen	A	mice	cell line (EL-4)	Wagner et al. 2006	A N (C50>5.00E- 04 M)	mice human	cell line (EL-4) PBMC (in vitro)	Wagner et al. 2006 Kooijman et al. 2010		
Suppression, A: Augumentation, N	N: No effect,	(H) humar	na study,							

Observation I conservation			In vitro effect on I		
Chemical name	Effect Animal in vitro (method)		Reference		
Trichloroethylene					
Mycophenolic acid					
Mercaptobenzothiazole					
Ribavirin					
Nicotinamide					
Acetaminophen	A	mice	cell line (EL-4)	Wagner et al. 2006	

#: The criterion number used to define immunotoxicity

						NTP	data	
	Immunotoxici	y classification	In vivo	Ex vivo		In vitro		
Chemical name	Classification	Rationale	immune sytem organ weight	cytokine production	TDAR	cytokine production	T cell proliferation	Mode of action
Phase I study			_					
Dibutyl phthalate	TTC	3), 4)	A (spleen)			S (IL-2, 4, IFN-g)(H) A (IL-1b)(H) x 3 S (IL-1b)		This compound then is proposed to modulate cytokine secretion from both monocytes/macrophages and T cells.
Hydrocortisone	TTC	1)	S (thymus) x 2 S (spleen)		N	S (IFN-a)		
Lead(II) acetate	ттс	1)	A(thymus)		S N	S (IFN-g, IL-1b)(H) A (IL-4)(H)	S(H)	
Nickel(II) sulfate	TTC	1)	N S (thymus)		N	A (IL-4, IFN-g)(H) S (IL-2) S (IFN-g)		
dimethyldithiocarba mate (DMDTC) Phase II study	NTTC					S (IL-1b)	N(H)	
2.4-diaminotoluene	NTTC		N (spleen) A (spleen)		s	-	-	
Benzo(a)pyrene	TTC	2), 3)		S(IL-2)	S x 5	A (IL-4)(H) N (IFNγ)(H) N (IL-2)(H) S (IL-2, 4, IFN-g)	S(H)x2 Sx6	Disruption of T-cell activities has been associated with B(a)P induced immunotoxic effects (Urso et al 1986).
Cadmium Chloride	TTC	2), 3)	A (spleen) S (spleen)	A (IL-2) N (IFN-γ)	Sx4	A (IFN-g)(H) S (IL-2, IFN-g) A (IFN-g) S (IL-2) A (IL-2)	S	
Dibromoacetic acid (DBAA)	ттс	1), 4)	A (spleen) S (thymus) x 2		N	S (II2, 4)	s	Overall, studies suggest that DBAA produces immunotoxic effects through modulation of T-cell mediated cell immunity. T-cell apoptosis, through extrinsic and intrinsic pathways, are proposed to play a role in the mode of action.
Diethylstilbestrol (DES)	TTC	1), 2), 4)	S (thymus) x 4 A (thymus) x 2 A (spleen)	A (IFN-g) x 3	s	A (IL-1) A (IL-2)		DES exposure was associated with down-regulation of gene expression in the TCR complex, and the TCR and CD28 signaling pathways.
Diphenylhydantoin	TTC	2), 3), 4)		A (IL-4) S (IFN-γ, IL-2) S (IL-1α) N (IL-6, 12)	S Ax2	-	-	DPH treatment can lead to a decrease of suppressor T cells
Ethylene Dibromide (EDB)	TTC	1)	S (thymus) S (spleen) N		A	-	s	
Glycidol	NTTC		N		s	-	-	Studies suggest that glycidol modulates B-cell function, and NK cell and macrophage activities.111 and decreased cytotoxic T cell activity
Indomethacin	TTC	3), 4)	N A (spleen)		S x 3 A x 1	A (IL-2)(H) A (IFN-g)(H)	A(H)x4 S Ax3	indomethacin inhibition of prostaglandin synthesis leads to altered T-cell function,
Isonicotinic Acid Hydrazide (IAH)	TTC	2)	N x 2			S (IL-2)(H) A (IL-2)(H) S (IL-1)(H)	S (H) x 3 A (H) x 6 A N	
Nitrobenzene	Undetermined		A (spleen) x 3 A (thymus) x 2		S N	-		effects on T-cell function may play a role in increased susceptibility to L. monocytogenes (Burns et al. 1994).
Urethane, Ethyl carbamate	ттс	1)	S (thymus) x2 S (spleen) x 2 N A (thymus) A (spleen)	N (IL-2)	Sx2 N	N (IL-2, 4, IFN-g)(H) A (IFN-g)(H) S (IFN-g)(H)	Nx2	
Tributyltin Chloride (TBTC)	TTC	1)	S (thymus) x4 S (spleen) x 3		N S	A (INF-g)(H) N (IL-2, 4)(H) S (IFN-g)(H)	S (H) S x 3	
Perflouorooctanoic Acid (PFOA)	TTC	1)	S (thymus) x2 S (spleen) x 2	N (IFN-g)		S (IL-4)(H) N (IL-2)(H)	A (H) S (H) N (H)	Direct modulation of NF-kB has been implicated in modulation of cytokine production and secretion (Corsini et al. 2012).
Dichloroacetic Acid (DCAA)	TTC	2), 3)	A(spleen)	N (IL-2) A (IFN-γ) x 3 S (IL-4) x 2 S (IL-2)	N	A (IL-2)(H) A (IL-2, IFN-g)		T-cell activation was one proposed mode of action for DCAA.
Toluene	NTTC		N		N		N	
Acetonitrile	NTTC		S(thymus)		S	-	-	
Mannitol	NTTC				S 1		N (H)	
Vanadium Pentoxide	NTTC		N A (spleen)	6	4	N	N(H)	
o-Benzyl-p-	NTTC		N		N	-	-	-

Appendix 8 Table. The summary of immunotoxicological data of 25 chemicals (continue)

		In	vitro effect on IL-	-2			ed by the VMT vitro effect on IFN-γ			In vi	itro effect on IIL	4
Chemical name	Effect	Animal	in vitro (method)	References	Effect	Animal	in vitro (method)	References	Effect	Animal	in vitro (method)	References
Phase I study					s	human	T cells (in vitro)	Hansen et al. 2015	S	human	T cells (in vitro)	Hansen et al. 2015
Dibutyl phthalate												(0.0278~27.8 ug/mL)
	S	human	lymphocyte (in vitro)	Chikanza and Panayi								
Hydrocortisone	s	human	PBL (in vitro)	1993 Goodwin et al. 1986								
					s	mice	splenocyte (ex vivo)	Fernandez-Cabezudo et al. 2007	A no effect	mice	splenocyte (ex vivo)	Fernandez- Cabezudo et al.
Lead(II) acetate					no effect S	mice human	cell line (EL-4) PBMC	Wagner et al. 2006 Hemdan et al. 2005	A A	mice	cell line (EL-4)	2007 Wagner et al. 2006 Hemdan et al. 2005
, ,						- idiridir	. Simo			human	PBMC (in vitro)	Chen et al. 2004
					Δ.	mice	spleen cell (in vitro)	Kim et al. 2009	A, S	rat mice	? spleen cell (in vitro)	Kim et al. 2009
Nickel(II) sulfate					A (NiCl2) A	mice human	cell line (EL-4) PBMC (in vitro)	Wagner et al. 2006 Thomas et al. 2003	A (NiCl2)	mice	cell line (EL-4)	Wagner et al. 2006 Thomas et al. 2003
					A	rat	lymphoid lung cell (ex vivo)	Goutet et al. 2000	A	human	PBMC (in vitro)	
limethyldithiocarba mate (DMDTC)												
Phase II study												
2.4-diaminotoluene												
Benzo(a)pyrene												
					N (ex vivo), A (in	rat	splenocyte (ex vivo, in vitro)	Wang et al. 2017	no effect	rat	spleen cell (ex vivo)	Demenesku et al.
					vitro) S	rat	spleen cell (ex vivo) PBMC (in vitro)	Demenesku et al. 2014				2014
Cadmium Chloride					S (IC50=7.05E- 05 M)	human	thymocyte, splenocyte (in	Kooijiman et al. 2010			1	
					s	mice	vitro)	Pathak and Khandelwal 2008				
Dibromoacetic acid												
(DBAA)												
Diethylstilbestrol (DES)												
Diphenylhydantoin												
Ethylene Dibromide												
(EDB)												
Chaidal												
Glycidol												
Indomethacin												
IROIR CHACH												
Isonicotinic Acid		human	PBMC (in vitro), cell line (Jurkat)	Tsuboi et al. 1995								
Hydrazide (IAH)	A											
Nitrobenzene												
. via obchizene												
Urethane, Ethyl carbamate												
Fributyltin Chloride					no effect (TBTO)	mice	cell line (EL-4)	Ringerike et al. 2005				
(TBTC)												
Perflouorooctanoic												
Acid (PFOA)												
Dichloroacetic Acid												
(DCAA)												
Toluene												
Acetonitrile												
Mannnitol					 							
Vanadium					6	5						
Pentoxide o-Benzyl-p-					 							
hlorophenol (BCP)			et, (H) humana study,					<u> </u>				

汝献

- 1993a. NTP Toxicology and Carcinogenesis Studies of Acetaminophen (CAS No. 103-90-2) in F344 Rats and B6C3F1 Mice (Feed Studies). Natl Toxicol Program Tech Rep Ser 394, 1-274.
- 1993b. NTP Toxicology and Carcinogenesis Studies of p-Nitroaniline (CAS No. 100-01-6) in B6C3F1 Mice (Gavage Studies). Natl Toxicol Program Tech Rep Ser 418, 1-203.
- 1995. NTP Toxicology and Carcinogenesis Studies of Benzethonium Chloride (CAS No. 121-54-0) in F344/N Rats and B6C3F1 Mice (Dermal Studies). Natl Toxicol Program Tech Rep Ser 438, 1-220.
- 1996. NTP Toxicology and Carcinogenesis Studies of Nickel Sulfate Hexahydrate (CAS No. 10101-97-0) in F344 Rats and B6C3F1 Mice (Inhalation Studies). Natl Toxicol Program Tech Rep Ser 454, 1-380.
- 1997. NTP Toxicology and Carcinogenesis Studies of Salicylazosulfapyridine (CAS No. 599-79-1) in F344/N Rats and B6C3F1 Mice (Gavage Studies). Natl Toxicol Program Tech Rep Ser 457, 1-327.
- 2003. NTP toxicology and carcinogenesiss studies of citral (microencapsulated) (CAS No. 5392-40-5) in F344/N rats and B6C3F1 mice (feed studies). Natl Toxicol Program Tech Rep Ser, 1-268.
- Almousa, L.A., Salter, A.M., Langley-Evans, S.C., 2018. Magnesium deficiency heightens lipopolysaccharide-induced inflammation and enhances monocyte adhesion in human umbilical vein endothelial cells. Magnes Res 31, 39-48.
- Auli, M., Domenech, A., Andres, A., et al., 2012. Multiparametric immunotoxicity screening in mice during early drug development. Toxicol Lett 214, 200-208.
- Beschorner, W.E., Namnoum, J.D., Hess, A.D., et al., 1987. Cyclosporin A and the thymus. Immunopathology. Am J Pathol 126, 487-496.
- Bessler, H., Straussberg, R., Gurary, N., et al., 1996. Effect of dexamethasone on IL-2 and IL-3 production by mononuclear cells in neonates and adults. Arch Dis Child Fetal Neonatal Ed 75, F197-201.
- Blanke, T.J., Little, J.R., Shirley, S.F., et al., 1977. Augmentation of murine immune responses by amphotericin B. Cell Immunol 33, 180-190.

- Bruserud, O., Lundin, K., 1987. The effect of drugs used in anticoagulation therapy on T lymphocyte activation in vitro. II. Warfarin inhibits T lymphocyte activation. J Clin Lab Immunol 23, 169-173.
- Bunn, T.L., Parsons, P.J., Kao, E., et al., 2001. Exposure to lead during critical windows of embryonic development: differential immunotoxic outcome based on stage of exposure and gender. Toxicol Sci 64, 57-66.
- Bygbjerg, I.C., Svenson, M., Theander, T.G., et al., 1987. Effect of antimalarial drugs on stimulation and interleukin 2 production of human lymphocytes. Int J Immunopharmacol 9, 513-519.
- Caren, L.D., Oven, H.M., Mandel, A.D., 1985. Dimethyl sulfoxide: lack of suppression of the humoral immune response in mice. Toxicol Lett 26, 193-197.
- Cesario, T.C., Slater, L.M., Kaplan, H.S., et al., 1984. Effect of antineoplastic agents on gamma-interferon production in human peripheral blood mononuclear cells. Cancer Res 44, 4962-4966.
- Chetty, K.N., Subba Rao, D.S., Drummond, L., et al., 1979. Cobalt induced changes in immune response and adenosine triphosphatase activities in rats. J Environ Sci Health B 14, 525-544.
- Chikanza, L.C., Panayi, G.S., 1993. The effects of hydrocortisone on in vitro lymphocyte proliferation and interleukin-2 and -4 production in corticosteroid sensitive and resistant subjects. Eur J Clin Invest 23, 845-850.
- de Abreu Costa, L., Henrique Fernandes Ottoni, M., Dos Santos, M.G., et al., 2017. Dimethyl Sulfoxide (DMSO) Decreases Cell Proliferation and TNF-alpha, IFN-gamma, and IL-2 Cytokines Production in Cultures of Peripheral Blood Lymphocytes. Molecules 22.
- De Waal, E.J., Timmerman, H.H., Dortant, P.M., et al., 1995. Investigation of a screening battery for immunotoxicity of pharmaceuticals within a 28-day oral toxicity study using azathioprine and cyclosporin A as model compounds. Regul Toxicol Pharmacol 21, 327-338.
- Dieter, M.P., Luster, M.I., Boorman, G.A., et al., 1983. Immunological and biochemical responses in mice treated with mercuric chloride. Toxicol Appl Pharmacol 68, 218-228.

- Dupont, E., Huygen, K., Schandene, L., et al., 1985. Influence of in vivo immunosuppressive drugs on production of lymphokines. Transplantation 39, 143-147.
- Dupuis, G., Martel, J., Bastin, B., et al., 1993. Microtubules are not an essential component of phytohemagglutinin-dependent signal transduction in Jurkat T lymphocytes. Cell Immunol 146, 38-51.
- el Fouhil, A.F., Iskander, F.A., Turkall, R.M., 1993a. Effect of alternate-day hydrocortisone therapy on the immunologically immature rat. II: Changes in T-and B-cell areas in spleen. Toxicol Pathol 21, 383-390.
- el Fouhil, A.F., Iskander, F.A., Turkall, R.M., 1993b. Effect of alternate-day hydrocortisone therapy on the immunologically immature rat. III: Changes in T-and B-cell areas in lymph nodes. Toxicol Pathol 21, 391-396.
- el Fouhil, A.F., Turkall, R.M., 1993. Effect of alternate-day hydrocortisone therapy on the immunologically immature rat. I: Effect on blood cell count, immunoglobulin concentrations, and body and organ weights. Toxicol Pathol 21, 377-382.
- Exon, J.H., Koller, L.D., Talcott, P.A., et al., 1986. Immunotoxicity testing: an economical multiple-assay approach. Fundam Appl Toxicol 7, 387-397.
- Ezendam, J., Hassing, I., Bleumink, R., et al., 2004. Hexachlorobenzene-induced Immunopathology in Brown Norway rats is partly mediated by T cells. Toxicol Sci 78, 88-95.
- Freed, B.M., Lempert, N., Lawrence, D.A., 1989. The inhibitory effects of Nethylmaleimide, colchicine and cytochalasins on human T-cell functions. Int J Immunopharmacol 11, 459-465.
- Freed, B.M., Rapoport, R., Lempert, N., 1987. Inhibition of early events in the human T-lymphocyte response to mitogens and alloantigens by hydrogen peroxide. Arch Surg 122, 99-104.
- Fujiwara, M., Mitsui, K., Yamamoto, I., 1990. Inhibition of proliferative responses and interleukin 2 productions by salazosulfapyridine and its metabolites. Jpn J Pharmacol 54, 121-131.
- Gabryel, B., Labuzek, K., Malecki, A., et al., 2004. Immunophilin ligands decrease release of pro-inflammatory cytokines (IL-1beta, TNF-alpha and IL-2 in rat

- astrocyte cultures exposed to simulated ischemia in vitro. Pol J Pharmacol 56, 129-136.
- Garly, M.L., Trautner, S.L., Marx, C., et al., 2008. Thymus size at 6 months of age and subsequent child mortality. J Pediatr 153, 683-688, 688.e681-683.
- Gentile, D.A., Henry, J., Katz, A.J., et al., 1997. Inhibition of peripheral blood mononuclear cell proliferation by cardiac glycosides. Ann Allergy Asthma Immunol 78, 466-472.
- Ghare, S., Patil, M., Hote, P., et al., 2011. Ethanol inhibits lipid raft-mediated TCR signaling and IL-2 expression: potential mechanism of alcohol-induced immune suppression. Alcohol Clin Exp Res 35, 1435-1444.
- Goodwin, J.S., Atluru, D., Sierakowski, S., et al., 1986. Mechanism of action of glucocorticosteroids. Inhibition of T cell proliferation and interleukin 2 production by hydrocortisone is reversed by leukotriene B4. J Clin Invest 77, 1244-1250.
- Haley, P.J., Shopp, G.M., Benson, J.M., et al., 1990. The immunotoxicity of three nickel compounds following 13-week inhalation exposure in the mouse. Fundam Appl Toxicol 15, 476-487.
- Hanke, J.H., Nichols, L.N., Coon, M.E., 1992. FK506 and rapamycin selectively enhance degradation of IL-2 and GM-CSF mRNA. Lymphokine Cytokine Res 11, 221-231.
- Hansen, J.F., Nielsen, C.H., Brorson, M.M., et al., 2015. Influence of phthalates on in vitro innate and adaptive immune responses. PLoS One 10, e0131168.
- Hattori, A., Kunz, H.W., Gill, T.J., 3rd, et al., 1987. Thymic and lymphoid changes and serum immunoglobulin abnormalities in mice receiving cyclosporine. Am J Pathol 128, 111-120.
- He, Y.W., Deftos, M.L., Ojala, E.W., et al., 1998. RORgamma t, a novel isoform of an orphan receptor, negatively regulates Fas ligand expression and IL-2 production in T cells. Immunity 9, 797-806.
- Henderson, D.J., Naya, I., Bundick, R.V., et al., 1991. Comparison of the effects of FK-506, cyclosporin A and rapamycin on IL-2 production. Immunology 73, 316-321.
- Himmerich, H., Schonherr, J., Fulda, S., et al., 2011. Impact of antipsychotics on cytokine production in-vitro. J Psychiatr Res 45, 1358-1365.

- Hu, H., Abedi-Valugerdi, M., Moller, G., 1997. Pretreatment of lymphocytes with mercury in vitro induces a response in T cells from genetically determined low-responders and a shift of the interleukin profile. Immunology 90, 198-204.
- Huchet, R., Grandjon, D., 1988. Histamine-induced regulation of IL-2 synthesis in man: characterization of two pathways of inhibition. Ann Inst Pasteur Immunol 139, 485-499.
- Iatropoulos, M.J., Hobson, W., Knauf, V., et al., 1976. Morphological effects of hexachlorobenzene toxicity in female rhesus monkeys. Toxicol Appl Pharmacol 37, 433-444.
- Kanariou, M., Huby, R., Ladyman, H., et al., 1989. Immunosuppression with cyclosporin A alters the thymic microenvironment. Clin Exp Immunol 78, 263-270.
- Karas, K., Salkowska, A., Sobalska-Kwapis, M., et al., 2018. Digoxin, an Overlooked Agonist of RORgamma/RORgammaT. Front Pharmacol 9, 1460.
- Khan, M.M., Melmon, K.L., Fathman, C.G., et al., 1985. The effects of autacoids on cloned murine lymphoid cells: modulation of IL 2 secretion and the activity of natural suppressor cells. J Immunol 134, 4100-4106.
- Kim, J.H., Park, J.S., 2002. Potentiation of the immunotoxicity of ethanol by acetaminophen in mice. Int Immunopharmacol 2, 15-24.
- Kim, J.Y., Huh, K., Lee, K.Y., et al., 2009. Nickel induces secretion of IFN-gamma by splenic natural killer cells. Exp Mol Med 41, 288-295.
- Kim, S.K., Kwon, D.A., Lee, H.S., et al., 2019. Preventive Effect of the Herbal Preparation, HemoHIM, on Cisplatin-Induced Immune Suppression. Evid Based Complement Alternat Med 2019, 3494806.
- Kloppenburg, M., Verweij, C.L., Miltenburg, A.M., et al., 1995. The influence of tetracyclines on T cell activation. Clin Exp Immunol 102, 635-641.
- Knight, J.A., Plowman, M.R., Hopfer, S.M., et al., 1991. Pathological reactions in lung, liver, thymus, and spleen of rats after subacute parenteral administration of nickel sulfate. Ann Clin Lab Sci 21, 275-283.
- Kouchi, Y., Maeda, Y., Ohuchida, A., et al., 1996. Immunotoxic effect of low dose cisplatin in mice. J Toxicol Sci 21, 227-233.

- Kucharz, E.J., Sierakowski, S.J., 1990. Studies on immunomodulatory properties of isoniazid. II. Effect of isoniazid on interleukin 2 production and interleukin 2receptor expression. J Hyg Epidemiol Microbiol Immunol 34, 207-211.
- Labuzek, K., Kowalski, J., Gabryel, B., et al., 2005. Chlorpromazine and loxapine reduce interleukin-1beta and interleukin-2 release by rat mixed glial and microglial cell cultures. Eur Neuropsychopharmacol 15, 23-30.
- Landewe, R.B., Miltenburg, A.M., Verdonk, M.J., et al., 1995. Chloroquine inhibits T cell proliferation by interfering with IL-2 production and responsiveness. Clin Exp Immunol 102, 144-151.
- Lee, J., Lim, K.T., 2012. SJSZ glycoprotein (38 kDa) modulates expression of IL-2, IL-12, and IFN-gamma in cyclophosphamide-induced Balb/c. Inflamm Res 61, 1319-1328.
- Lehmann, D.M., Williams, W.C., 2018. Development and utilization of a unique in vitro antigen presentation co-culture model for detection of immunomodulating substances. Toxicol In Vitro 53, 20-28.
- Lemster, B., Woo, J., Strednak, J., et al., 1992. Cytokine gene expression in murine lymphocytes activated in the presence of FK 506, bredinin, mycophenolic acid, or brequinar sodium. Transplant Proc 24, 2845-2846.
- Loose, L.D., Silkworth, J.B., Pittman, K.A., et al., 1978. Impaired host resistance to endotoxin and malaria in polychlorinated biphenyl- and hexachlorobenzene-treated mice. Infect Immun 20, 30-35.
- Lu, Z., Liu, F., Chen, L., et al., 2015. Effect of Chronic Administration of Low Dose Rapamycin on Development and Immunity in Young Rats. PLoS One 10, e0135256.
- Maeda, M., Ishii, H., Tanaka, S., et al., 2010. Suppressive efficacies of antimicrobial agents against human peripheral-blood mononuclear cells stimulated with T cell mitogen and bacterial superantigen. Arzneimittelforschung 60, 760-768.
- Meredith, C., Scott, M.P., 1994. Altered gene expression in immunotoxicology screening in vitro: Comparison with ex vivo analysis. Toxicol In Vitro 8, 751-753.
- Miller, L.C., Kaplan, M.M., 1992. Serum interleukin-2 and tumor necrosis factoralpha in primary biliary cirrhosis: decrease by colchicine and relationship to HLA-DR4. Am J Gastroenterol 87, 465-470.

- Miller, T.E., Golemboski, K.A., Ha, R.S., et al., 1998. Developmental exposure to lead causes persistent immunotoxicity in Fischer 344 rats. Toxicol Sci 42, 129-135.
- Munson, A.E., Sanders, V.M., Douglas, K.A., et al., 1982. In vivo assessment of immunotoxicity. Environ Health Perspect 43, 41-52.
- Nalesnik, M.A., Todo, S., Murase, N., et al., 1987. Toxicology of FK-506 in the Lewis rat. Transplant Proc 19, 89-92.
- Northoff, H., Carter, C., Oppenheim, J.J., 1980. Inhibition of concanavalin A-induced human lymphocyte mitogenic factor (Interleukin-2) production by suppressor T lymphocytes. J Immunol 125, 1823-1828.
- Palacios, R., Sugawara, I., 1982. Hydrocortisone abrogates proliferation of T cells in autologous mixed lymphocyte reaction by rendering the interleukin-2 Producer T cells unresponsive to interleukin-1 and unable to synthesize the T-cell growth factor. Scand J Immunol 15, 25-31.
- Pally, C., Tanner, M., Rizvi, H., et al., 2001. Tolerability profile of sodium mycophenolate (ERL080) and mycophenolate mofetil with and without cyclosporine (Neoral) in the rat. Toxicology 157, 207-215.
- Parenti, D.M., Simon, G.L., Scheib, R.G., et al., 1988. Effect of lithium carbonate in HIV-infected patients with immune dysfunction. J Acquir Immune Defic Syndr 1, 119-124.
- Parthasarathy, N.J., Kumar, R.S., Devi, R.S., 2005. Effect of methanol intoxication on rat neutrophil functions. J Immunotoxicol 2, 115-121.
- Peterson, K.P., Van Hirtum, M., Peterson, C.M., 1997. Dapsone decreases the cumulative incidence of diabetes in non-obese diabetic female mice. Proc Soc Exp Biol Med 215, 264-268.
- Poluektova, L.Y., Huggler, G.K., Patterson, E.B., et al., 1999. Involvement of protein kinase A in histamine-mediated inhibition of IL-2 mRNA expression in mouse splenocytes. Immunopharmacology 41, 77-87.
- Quemeneur, L., Flacher, M., Gerland, L.M., et al., 2002. Mycophenolic acid inhibits IL-2-dependent T cell proliferation, but not IL-2-dependent survival and sensitization to apoptosis. J Immunol 169, 2747-2755.

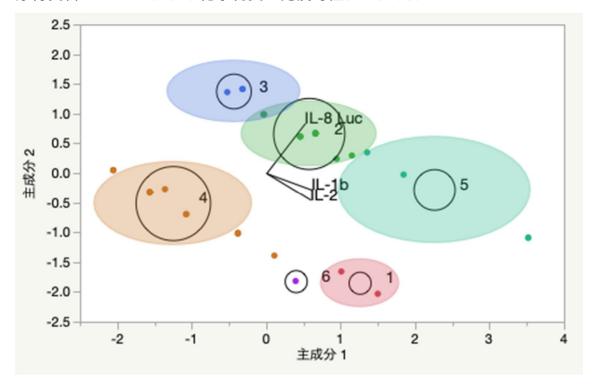
- Ress, N.B., Hailey, J.R., Maronpot, R.R., et al., 2003. Toxicology and carcinogenesis studies of microencapsulated citral in rats and mice. Toxicol Sci 71, 198-206.
- Riesbeck, K., 1999. Cisplatin at clinically relevant concentrations enhances interleukin-2 synthesis by human primary blood lymphocytes. Anticancer Drugs 10, 219-227.
- Ringerike, T., Ulleras, E., Volker, R., et al., 2005. Detection of immunotoxicity using T-cell based cytokine reporter cell lines ("Cell Chip"). Toxicology 206, 257-272.
- Roche, Y., Fay, M., Gougerot-Pocidalo, M.A., 1988. Enhancement of interleukin 2 production by quinolone-treated human mononuclear leukocytes. Int J Immunopharmacol 10, 161-167.
- Saito, R., Hirakawa, S., Ohara, H., et al., 2011. Nickel differentially regulates NFAT and NF-kappaB activation in T cell signaling. Toxicol Appl Pharmacol 254, 245-255.
- Salazar, V., Castillo, C., Ariznavarreta, C., et al., 2004. Effect of oral intake of dibutyl phthalate on reproductive parameters of Long Evans rats and pre-pubertal development of their offspring. Toxicology 205, 131-137.
- Santarelli, L., Bracci, M., Mocchegiani, E., 2006. In vitro and in vivo effects of mercuric chloride on thymic endocrine activity, NK and NKT cell cytotoxicity, cytokine profiles (IL-2, IFN-gamma, IL-6): role of the nitric oxide-L-arginine pathway. Int Immunopharmacol 6, 376-389.
- Schleuning, M.J., Duggan, A., Reem, G.H., 1989. Inhibition by chlorpromazine of lymphokine-specific mRNA expression in human thymocytes. Eur J Immunol 19, 1491-1495.
- Sfikakis, P.P., Souliotis, V.L., Katsilambros, N., et al., 1996. Downregulation of interleukin-2 and apha-chain interleukin-2 receptor biosynthesis by cisplatin in human peripheral lymphocytes. Clin Immunol Immunopathol 79, 43-49.
- She, Y., Wang, N., Chen, C., et al., 2012. Effects of aluminum on immune functions of cultured splenic T and B lymphocytes in rats. Biol Trace Elem Res 147, 246-250.
- Sheikhi, A., Jaberi, Y., Esmaeilzadeh, A., et al., 2007. The effect of cardiovascular drugs on pro-inflammatory cytokine secretion and natural killer activity of

- peripheral blood mononuclear cells of patients with chronic heart failure in vitro. Pak J Biol Sci 10, 1580-1587.
- Silvestrini, B., Lisciani, R., Barcellona, P.S., 1967. Anti-granuloma and thymolytic activity of certain drugs. Eur J Pharmacol 1, 240-246.
- Song, Y., Han, S., Kim, H., et al., 2006. Effects of mizoribine on MHC-restricted exogenous antigen presentation in dendritic cells. Arch Pharm Res 29, 1147-1153.
- Sookoian, S., Castano, G., Flichman, D., et al., 2004. Effects of ribavirin on cytokine production of recall antigens and phytohemaglutinin-stimulated peripheral blood mononuclear cells. (Inhibitory effects of ribavirin on cytokine production). Ann Hepatol 3, 104-107.
- Sugiyama, K., Ueda, H., Ichio, Y., et al., 1995. Improvement of cisplatin toxicity and lethality by juzen-taiho-to in mice. Biol Pharm Bull 18, 53-58.
- Synzynys, B.I., Sharetskii, A.N., Kharlamova, O.V., 2004. [Immunotoxicity of aluminum chloride]. Gig Sanit, 70-72.
- Sztein, M.B., Simon, G.L., Parenti, D.M., et al., 1987. In vitro effects of thymosin and lithium on lymphoproliferative responses of normal donors and HIV seropositive male homosexuals with AIDS-related complex. Clin Immunol Immunopathol 44, 51-62.
- Takai, K., Jojima, K., Sakatoku, J., et al., 1990. Effects of FK506 on rat thymus: time-course analysis by immunoperoxidase technique and flow cytofluorometry. Clin Exp Immunol 82, 445-449.
- Tam, R.C., Pai, B., Bard, J., et al., 1999. Ribavirin polarizes human T cell responses towards a Type 1 cytokine profile. J Hepatol 30, 376-382.
- Tsukue, N., Toda, N., Tsubone, H., et al., 2001. Diesel exhaust (DE) affects the regulation of testicular function in male Fischer 344 rats. J Toxicol Environ Health A 63, 115-126.
- Turka, L.A., Dayton, J., Sinclair, G., et al., 1991. Guanine ribonucleotide depletion inhibits T cell activation. Mechanism of action of the immunosuppressive drug mizoribine. J Clin Invest 87, 940-948.
- Van Dijk, H., Bloksma, N., Rademaker, P.M., et al., 1979. Differential potencies of corticosterone and hydrocortisone in immune and immune-related processes in the mouse. Int J Immunopharmacol 1, 285-292.

- Van Wauwe, J., Aerts, F., Van Genechten, H., et al., 1996. The inhibitory effect of pentamidine on the production of chemotactic cytokines by in vitro stimulated human blood cells. Inflamm Res 45, 357-363.
- Vandebriel, R.J., Meredith, C., Scott, M.P., et al., 1998. Effects of in vivo exposure to bis(tri-n-butyltin)oxide, hexachlorobenzene, and benzo(a)pyrene on cytokine (receptor) mRNA levels in cultured rat splenocytes and on IL-2 receptor protein levels. Toxicol Appl Pharmacol 148, 126-136.
- Vargova, M., Wagnerova, J., Liskova, A., et al., 1993. Subacute immunotoxicity study of formaldehyde in male rats. Drug Chem Toxicol 16, 255-275.
- Vos, J.G., van Logten, M.J., Kreeftenberg, J.G., et al., 1979. Hexachlorobenzene-induced stimulation of the humoral immune response in rats. Ann N Y Acad Sci 320, 535-550.
- Vos, J.G., Van Loveren, H., 1994. Developments of immunotoxicology methods in the rat and applications to the study of environmental pollutants. Toxicol In Vitro 8, 951-956.
- Wagner, W., Sachrajda, I., Pulaski, L., et al., 2011. Application of cellular biosensors for analysis of bioactivity associated with airborne particulate matter. Toxicol In Vitro 25, 1132-1142.
- Wagner, W., Walczak-Drzewiecka, A., Slusarczyk, A., et al., 2006. Fluorescent Cell Chip a new in vitro approach for immunotoxicity screening. Toxicol Lett 162, 55-70.
- Wang, Y., Walker, C., Stadler, B.M., et al., 1984. Transcription and translation dependent induction of interleukin 2 (IL-2) and IL-2 receptors. Immunol Lett 8, 227-231.
- Wilson, R., Fraser, W.D., McKillop, J.H., et al., 1989. The "in vitro" effects of lithium on the immune system. Autoimmunity 4, 109-114.
- Yamamoto, N., Sakai, F., Yamazaki, H., et al., 1996. Effect of FR167653, a cytokine suppressive agent, on endotoxin-induced disseminated intravascular coagulation. Eur J Pharmacol 314, 137-142.
- Yoshimura, N., Matsui, S., Hamashima, T., et al., 1989. Effect of a new immunosuppressive agent, FK506, on human lymphocyte responses in vitro. II. Inhibition of the production of IL-2 and gamma-IFN, but not B cell-stimulating factor 2. Transplantation 47, 356-359.

Zhang, W.Z., Yong, L., Jia, X.D., et al., 2013. Combined subchronic toxicity of bisphenol A and dibutyl phthalate on male rats. Biomed Environ Sci 26, 63-69.

添付資料 14. MITA による化学物質の免疫毒性プロフィル



添付資料 15. Detailed review paper content.

Potential title:

"In vitro immunotoxicity testing"

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Ver.2.1

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