





## Draft Genome Sequence of Campylobacter jejuni CAM970 and C. coli CAM962, Associated with a Large Outbreak of Foodborne Illness in Fukuoka, Japan, in 2016

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**ABSTRACT** Here, we report the draft genome sequences of *Campylobacter jejuni* CAM970 and *C. coli* CAM962, which were associated with a large outbreak of foodborne illness originating from undercooked chicken sushi in Fukuoka, Japan, in May 2016. Their genome sizes were 1,690,901 and 1,704,736 bp, with 22 and 23 rRNAs, 9 and 9 tRNAs, and  $411\times$  and  $419\times$  coverage for *C. jejuni* CAM970 and *C. coli* CAM962, respectively.

ampylobacter jejuni and C. coli are two of the leading causes of foodborne gastrointestinal illnesses, which, worldwide, number approximately 400 to 500 million cases annually (1, 2). In Japan, a total of 339 cases of foodborne campylobacteriosis were reported in 2016, which represented 29.7% of all foodborne cases (1,140 cases) reported that year (3). Epidemiological data have shown that chicken meat is one of the main sources for human campylobacteriosis (4). In May 2016, a large foodborne outbreak caused by Campylobacter spp. occurred; 266 patients suffered from gastroenteritis due to the consumption of undercooked chicken meat and vinegar rice (sushi) in Fukuoka city. Bacteriological surveillance determined that both C. jejuni and C. coli were associated with the outbreak. In order to characterize their genomic traits, we examined, and report here, the draft genome sequences of C. jejuni CAM970 and C. coli CAM962, which were isolated from patients treated during the outbreak. Genomic DNA of the two strains was sequenced by single-end sequencing with an Ion Torrent PGM sequencer (Thermo Fisher Scientific, Waltham, MA, USA), resulting in average coverages of 411 $\times$  and 419 $\times$ , respectively. Raw reads were trimmed and de novo assembled using CLC Genomics Workbench version 9.0 (Qiagen, Hilden, Germany). The parameters for trimming were as follows: ambiguous limit, 2; quality limit, 0.05; number of 5' terminal nucleotides, 20; and number of 3' terminal nucleotides, 5. The parameters for the de novo assembly were as follows: mapping mode, create simple contig sequences (fast); bubble size, 50; word size, 21; minimum contig length, 500 bp; perform scaffolding, no; and autodetect paired distances, yes. The draft genomes of C. jejuni CAM970 and C. coli CAM962 were assembled into 61 and 34 contigs with accumulated lengths of 1,690,901 and 1,704,736 bp ( $N_{50}$ , 92,774 and 156,200 bp) and average G+C contents of 30.2 or 31.3%, respectively. The genome was annotated with the RAST server (5). Annotation of these assemblies identified 1,813 and 1,820 coding sequences, 22 and 23 rRNAs, and 9 and 9 tRNAs, for C. jejuni CAM970 and C. coli CAM962, respectively.

The sequence types (STs) of these strains were also profiled with a multilocus sequence typing (MLST) scheme (6). The results showed that strains CAM970 and CAM962 belonged to ST5716 and ST1181, respectively, and both strains were found to associate with human infection based on the *Campylobacter MLST* database (https://

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pubmlst.org/campylobacter). The data provided here can aid in future efforts to identify the sources of infections, since the genome sequence-based framework has potential advantages for epidemiological studies of Campylobacter infections (7). With the increased application of whole-genome sequencing technologies to epidemiological studies of Campylobacter outbreaks (i.e., source tracking) (8, 9), further accumulation of genomic data of Campylobacter spp. associated with foodborne outbreaks will contribute to improving our understanding of host and geographic adaptation of this pathogen and lead to the development of preventive strategies.

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession numbers BDRZ01000000 (CAM970) and BDRY01000000 (CAM962). The versions described in this paper are the first versions, BDRZ01000000 (CAM970) and BDRY01000000 (CAM962).

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## **REFERENCES**

- 1. Kaakoush NO, Castaño-Rodríguez N, Mitchell HM, Man SM. 2015. Global epidemiology of Campylobacter infection. Clin Microbiol Rev 28: 687-720. https://doi.org/10.1128/CMR.00006-15.
- 2. Ruiz-Palacios GM. 2007. The health burden of Campylobacter infection and the impact of antimicrobial resistance: playing chicken. Clin Infect Dis 44:701-703. https://doi.org/10.1086/509936
- 3. Japan Ministry of Health, Labour and Welfare. 2017. Food poisoning statistics report. http://www.mhlw.go.jp/stf/seisakunitsuite/bunya/kenkou \_iryou/shokuhin/syokuchu/04.html.
- 4. Skarp CP, Hänninen ML, Rautelin Hl. 2016. Campylobacteriosis: the role of poultry meat. Clin Microbiol Infect 22:103-109. https://doi.org/10 .1016/i.cmi.2015.11.019.
- 5. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S. Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75.

- 6. Joky KA, Maiden MC. 2010. BIGSdb: scalable analysis of bacterial genome variation at the population level. BMC Bioinfomatics 11:595. https://doi .org/10.1186/1471-2105-11-595.
- 7. Llarena AK, Taboada E, Rossi M. 2017. Whole-genome sequencing in the epidemiology of Campylobacter jejuni infections. J Clin Microbiol 55: 1269-1275. https://doi.org/10.1128/JCM.00017-17.
- 8. Lahti E, Löfdahl M, Ågren J, Hansson I, Olsson Engvall E. 2017. Confirmation of a campylobacteriosis outbreak associated with chicken liver pâté using PFGE and WGS. Zoonoses Public Health 64:14-20. https:// doi.org/10.1111/zph.12272.
- 9. Clark CG, Berry C, Walker M, Petkau A, Barker DO, Guan C, Reimer A, Taboada EN. 2016. Genomic insights from whole genome sequencing of four clonal outbreak Campylobacter jejuni assessed within the global C. jejuni population. BMC Genomics 17:990. https://doi.org/10.1186/s12864 -016-3340-8.