



Figure 7. A Tandem model approach. **(a)** In general, a model represents a subset of the network (Model A) within an entire network (Model U) (Left). In this case, what is happening in practice is to implicitly consider Model A is interfacing Model \bar{A} which is a black box of the rest of the network. With proper parameter tuning and value range restrictions, the model can provide reasonably accurate prediction of behaviors of the subset of the network. **(b)** Three models represent overlapping part of the network, and validated for their accuracy. This implies that these models are tuned for their counter-part black box. Within predefined range of values, a tandem application of three models is expected to deliver qualitatively useful predictions on behaviors of the network.

often results in inconsistency in the kinetic parameter values and therefore, these values have to be re-tuned after model integration. The constraint here is that fine-tuning the kinetic parameters in an integrated model may not work owing to different levels of abstraction among models. Even if the level of abstraction and methods of model description have been made consistent, it is a computationally intensive and time-consuming task that often requires additional experimental data for accurate estimates of kinetic parameter values.

In this study, we applied an alternative approach that involves a tandem use of multiple models, each of which is validated to be consistent with the experiments. In general, computational models of biological networks are developed focusing on a subset of networks such as the MAPK and insulin receptor pathways and a model of the entire network is not created. Nevertheless, many of these models achieve a practically sufficient accuracy that can be used for further scientific studies and drug discovery.⁵¹ This degree of accuracy may be attributable to the modularity and robustness of the biological network. If the entire network is so tightly connected, perturbation of any part of the network affects the behaviors of all other parts. This results in an extremely unstable system, leading to an evolutionary dead-end.⁵² Thus, in practice, we can define a subset of the interaction network and consider the rest to be a ‘black box’ from which effects can be implicitly represented within a specific parameter setting of the model (Figure 7a). The success of this approach largely depends on appropriately defining the boundary of a focused network, so that strong interactions with a black box do not affect the dynamical behaviors of the model beyond the level adjustable by parameter settings. In this study, we used three models each representing a distinct functional pathway in the metabolic network, and they were validated experimentally (Figure 7b). Furthermore, the models were interfaced by the dose change of specific molecules. The output of the first model (glycogen metabolism and glycolysis) is a dose change of glucose and G6P that in turn became inputs for the second model

(pentose–phosphate pathway). The interface between the second model and the third model (glutathione metabolism) is NADP⁺ and NADPH. Fortunately, no obvious feedback loop exists between the models that could seriously alter the behaviors of an upstream model depending on the outcome of the downstream models. It is noteworthy that the purpose of the simulation was to understand the qualitative propensity of this part of the metabolic network following sunitinib administration, rather than computing the exact dynamics with quantitative precision. The results of this tandem simulation correctly predicted physiological responses to the administration of sunitinib. Further studies may reveal conditions for this tandem approach to be practically applicable, which would lay the foundation for scaling-up computational studies efficiently for larger networks. It should be noted that we used the model to understand how sunitinib administration may qualitatively influence the metabolic system, rather than to obtain precise numerical results. To this end, the models were only required to deliver the proper degree of change in output and therefore, could have different baseline variables. In other words, the dose of the interface molecule could be 100 units in one model, and the dose level of the same molecule could be represented as 1,000 units in the other model. This is because the only information required is the degree of change and not the absolute number in each model. In such cases, a combination of simulations using separate models is expected to provide reasonable predictions.

The results reported herein demonstrate that *in silico* prediction (with appropriately designed experimental validation) is an effective strategy to examine toxicological issues associated with kinase inhibitors. Results of this study have major implications for the use of sunitinib in clinical practice, as it is essential to overcome adverse reactions associated with sunitinib tolerability.¹³ Suppression of sunitinib-mediated thyroid dysfunction may improve patients’ quality of life by ameliorating subjective symptoms, such as fatigue. More importantly, since the sunitinib-mediated reduction in platelet counts is a dose-limiting toxicity, an improvement in thrombocytopenia will allow the continued use of higher doses, which is desirable in treating the primary disease. Indeed, the therapeutic efficacy of sunitinib in RCC correlates positively with the area under the plasma concentration curve for the drug.⁵³ The experiments conducted in the present study strongly indicate that the multi-organ toxicity suffered by patients receiving sunitinib may be alleviated by the concomitant administration of α -TN.

In conclusion, we demonstrated that a systems toxicological approach could be successfully utilized to elucidate molecular mechanisms underlying adverse reactions associated with kinase inhibitors. This approach may be applicable to other kinase inhibitors to maximize their clinical benefits.

ACKNOWLEDGEMENTS

This work was supported in part by a Grant-in-Aid for Scientific Research (B) 21390041 and a Grant-in-Aid for Scientific Research (A) 24249034 from the Japan Society for the Promotion of Science, and a Grant-in-Aid for Challenging Exploratory Research 26670265 and a Grant-in-Aid for Scientific Research on Innovative Areas ‘HD-physiology’ 22136015 from the Ministry of Education, Culture, Sports, Science and Technology. A part of this research was carried out under Memorandum of Understanding (MOU Number 225-12-8000) between The United States Food and Drug Administration (FDA) and the Systems Biology Institute.

COMPETING INTERESTS

The University of Tokyo has filed a patent application about concomitant use of sunitinib and anti-oxidants (PCT/JP2013/081210, inventors: Masashi Honma, Hiroshi Suzuki, Takahiro Amemiya and Haruki Kume).

REFERENCES

- Huang M, Shen A, Ding J, Geng M. Molecularly targeted cancer therapy: some lessons from the past decade. *Trends Pharmacol Sci* 2014; **35**: 41–50.
- Levitzi A. Tyrosine kinase inhibitors: views of selectivity, sensitivity, and clinical performance. *Annu Rev Pharmacol Toxicol* 2013; **53**: 161–185.
- Zamecnikova A. Novel approaches to the development of tyrosine kinase inhibitors and their role in the fight against cancer. *Expert Opin Drug Discov* 2014; **9**: 77–92.
- Brown RL. Tyrosine kinase inhibitor-induced hypothyroidism: incidence, etiology, and management. *Target Oncol* 2011; **6**: 217–226.
- Faivre S, Delbaldo C, Vera K, Robert C, Lozahic S, Lassau N et al. Safety, pharmacokinetic, and antitumor activity of SU11248, a novel oral multitarget tyrosine kinase inhibitor, in patients with cancer. *J Clin Oncol* 2006; **24**: 25–35.
- Vandyke K, Fitter S, Dewar AL, Hughes TP, Zannettino AC. Dysregulation of bone remodeling by imatinib mesylate. *Blood* 2010; **115**: 766–774.
- Hao D, Rowinsky EK. Inhibiting signal transduction: recent advances in the development of receptor tyrosine kinase and Ras inhibitors. *Cancer Invest* 2002; **20**: 387–404.
- Karaman MW, Herrgard S, Treiber DK, Gallant P, Atteridge CE, Campbell BT et al. A quantitative analysis of kinase inhibitor selectivity. *Nat Biotechnol* 2008; **26**: 127–132.
- Baka S, Clamp AR, Jayson GC. A review of the latest clinical compounds to inhibit VEGF in pathological angiogenesis. *Expert Opin Ther Targets* 2006; **10**: 867–876.
- Sablin MP, Dreyer C, Colichi C, Bouattour M, Delbaldo C, Faivre S et al. Benefits from pharmacological and pharmacokinetic properties of sunitinib for clinical development. *Expert Opin Drug Metab Toxicol* 2010; **6**: 1005–1015.
- Gotink KJ, Verheul HM. Anti-angiogenic tyrosine kinase inhibitors: what is their mechanism of action? *Angiogenesis* 2010; **13**: 1–14.
- Negrier S, Raymond E. Antiangiogenic treatments and mechanisms of action in renal cell carcinoma. *Invest New Drugs* 2012; **30**: 1791–1801.
- Ravaud A. Treatment-associated adverse event management in the advanced renal cell carcinoma patient treated with targeted therapies. *Oncol* 2011; **16** Suppl 2: 32–44.
- Facchini G, Perri F, Caraglia M, Pisano C, Striano S, Marra L et al. New treatment approaches in renal cell carcinoma. *Anti-cancer drugs* 2009; **20**: 893–900.
- Kariya Y, Honma M, Suzuki H. Systems-based understanding of pharmacological responses with combinations of multidisciplinary methodologies. *Biopharm Drug Dispos* 2013; **34**: 489–506.
- Iyengar R, Zhao S, Chung SW, Mager DE, Gallo JM. Merging systems biology with pharmacodynamics. *Sci Transl Med* 2012; **4**: 126ps127.
- Bollen M, Keppens S, Stalmans W. Specific features of glycogen metabolism in the liver. *Biochem J* 1998; **336**: 19–31.
- Hers HG. The control of glycogen metabolism in the liver. *Annu Rev Biochem* 1976; **45**: 167–189.
- Thomas PD, Campbell MJ, Kejariwal A, Mi H, Karlak B, Daverman R et al. PANTHER: a library of protein families and subfamilies indexed by function. *Genome Res* 2003; **13**: 2129–2141.
- Chelliah V, Laipe C, Le Novere N. BioModels database: a repository of mathematical models of biological processes. *Methods Mol Biol* 2013; **1021**: 189–199.
- Lu SC. Regulation of hepatic glutathione synthesis: current concepts and controversies. *FASEB J* 1999; **13**: 1169–1183.
- Lu SC. Regulation of glutathione synthesis. *Mol Aspects Med* 2009; **30**: 42–59.
- Funahashi A, Matsuoka Y, Akiya J, Morohashi M, Kikuchi N, Kitano H. CellDesigner 3.5: a versatile modeling tool for biochemical networks. *Proc IEEE* 2008; **96**: 1254–1265.
- Xu K, Morgan KT, Todd Gehris A, Elston TC, Gomez SM. A whole-body model for glycogen regulation reveals a critical role for substrate cycling in maintaining blood glucose homeostasis. *PLoS Comput Biol* 2011; **7**: e1002272.
- Sabate L, Franco R, Canela EI, Centelles JJ, Cascante M. A model of the pentose phosphate pathway in rat liver cells. *Mol Cell Biochem* 1995; **142**: 9–17.
- Reed MC, Thomas RL, Pavisic J, James SJ, Ulrich CM, Nijhout HF. A mathematical model of glutathione metabolism. *Theor Biol Med Model* 2008; **5**: 8.
- Wittig U, Kania R, Golebiewski M, Rey M, Shi L, Jong L et al. SABIO-RK—database for biochemical reaction kinetics. *Nucleic Acids Res* 2012; **40**: D790–D796.
- Hoops S, Sahle S, Gauges R, Lee C, Pahle J, Simus N et al. COPASI—a CComplex PATHway Simulator. *Bioinformatics* 2006; **22**: 3067–3074.
- Sekine S, Ito K, Horie T. Oxidative stress and Mrp2 internalization. *Free Rad Biol Med* 2006; **40**: 2166–2174.
- Chouhan JD, Zamarripa DE, Lai PH, Oramasionwu CU, Grabinski JL. Sunitinib (Sutent): a novel agent for the treatment of metastatic renal cell carcinoma. *J Oncol Pharm Pract* 2007; **13**: 5–15.
- Villarroel MC, Pratz KW, Xu L, Wright JJ, Smith BD, Rudek MA. Plasma protein binding of sorafenib, a multi kinase inhibitor: *in vitro* and in cancer patients. *Invest New Drugs* 2012; **30**: 2096–2102.
- Sanjo H, Kawai T, Akira S. DRAKs, novel serine/threonine kinases related to death-associated protein kinase that trigger apoptosis. *J Biol Chem* 1998; **273**: 29066–29071.
- Fitzgerald J, Bateman JF. Why mice have lost genes for COL21A1, STK17A, GPR145 and AHRI: evidence for gene deletion at evolutionary breakpoints in the rodent lineage. *Trends Genet* 2004; **20**: 408–412.
- Mao P, Hever MP, Niemaszzyk LM, Haghkerdar JM, Yanco EG, Desai D et al. Serine/threonine kinase 17A is a novel p53 target gene and modulator of cisplatin toxicity and reactive oxygen species in testicular cancer cells. *J Biol Chem* 2011; **286**: 19381–19391.
- Kearns AE, Donohue MM, Sanyal B, Demay MB. Cloning and characterization of a novel protein kinase that impairs osteoblast differentiation *in vitro*. *J Biol Chem* 2001; **276**: 42213–42218.
- Winchester JS, Rouchka EC, Rowland NS, Rice NA. *In Silico* characterization of phosphorylase kinase: evidence for an alternate intronic polyadenylation site in PHKG1. *Mol Genet Metab* 2007; **92**: 234–242.
- Maichele AJ, Burwinkel B, Maire I, Sovik O, Kilimann MW. Mutations in the testis/liver isoform of the phosphorylase kinase gamma subunit. PHKG2; cause autosomal liver glycogenosis in the gsd rat and in humans. *Nat Genet* 1996; **14**: 337–340.
- Pandolfi PP, Sonati F, Rivi R, Mason P, Grosveld F, Luzzatto L. Targeted disruption of the housekeeping gene encoding glucose 6-phosphate dehydrogenase (G6PD): G6PD is dispensable for pentose synthesis but essential for defense against oxidative stress. *EMBO J* 1995; **14**: 5209–5215.
- Wu G, Fang YZ, Yang S, Lupton JR, Turner ND. Glutathione metabolism and its implications for health. *J Nutr* 2004; **134**: 489–492.
- Maher JJ, Scott MK, Saito JM, Burton MC. Adenovirus-mediated expression of cytokine-induced neutrophil chemoattractant in rat liver induces a neutrophilic hepatitis. *Hepatology* 1997; **25**: 624–630.
- Bhojani N, Jeldres C, Patard JJ, Perrotte P, Suardi N, Hutterer G et al. Toxicities associated with the administration of sorafenib, sunitinib, and temsirolimus and their management in patients with metastatic renal cell carcinoma. *Eur Urol* 2008; **53**: 917–930.
- Degroot LJ, Niepomniszcze H. Biosynthesis of thyroid hormone: basic and clinical aspects. *Metabolism* 1977; **26**: 665–718.
- Takahashi S. Vascular endothelial growth factor (VEGF), VEGF receptors and their inhibitors for antiangiogenic tumor therapy. *Biol Pharm Bull* 2011; **34**: 1785–1788.
- Gibson RJ, Keefe DM, Lalla RV, Bateman E, Blijlevens N, Fijlstra M et al. Systematic review of agents for the management of gastrointestinal mucositis in cancer patients. *Support Care Cancer* 2013; **21**: 313–326.
- Andreyev J, Ross P, Donnellan C, Lennan E, Leonard P, Waters C et al. Guidance on the management of diarrhoea during cancer chemotherapy. *Lancet Oncol* 2014; **15**: e447–e460.
- Aapro MS, Bohlius J, Cameron DA, Dal Lago L, Donnelly JP, Kearney N et al. 2010update of EORTC guidelines for the use of granulocyte-colony stimulating factor to reduce the incidence of chemotherapy-induced febrile neutropenia in adult patients with lymphoproliferative disorders and solid tumours. *Eur J Cancer* 2011; **47**: 8–32.
- Voskens CJ, Goldinger SM, Loquai C, Robert C, Kaehler KC, Berking C et al. The price of tumor control: an analysis of rare side effects of anti-CTLA-4 therapy in metastatic melanoma from the ipilimumab network. *PLoS ONE* 2013; **8**: e53745.
- Judson RS, Martin MT, Egeghy P, Gangwal S, Reif DM, Kothiyi P et al. Aggregating data for computational toxicology applications: The U.S. Environmental Protection Agency. EPA; Aggregated Computational Toxicology Resource. ACToR; System. *Int J Mol Sci* 2012; **13**: 1805–1831.
- Stahl M, Guba W, Kansy M. Integrating molecular design resources within modern drug discovery research: the Roche experience. *Drug Discov Today* 2006; **11**: 326–333.
- Cases M, Briggs K, Steger-Hartmann T, Pognan F, Marc P, Kleinoder T et al. The eTOX data-sharing project to advance *in silico* drug-induced toxicity prediction. *Int J Mol Sci* 2014; **15**: 21136–21154.
- Schoeberl B, Pace EA, Fitzgerald JB, Harms BD, Xu L, Nie L et al. Therapeutically targeting ErbB3: a key node in ligand-induced activation of the ErbB receptor-P13K axis. *Sci Signal* 2009; **2**: ra31.
- Kirschner M, Gerhart J. Evolvability. *Proc Natl Acad Sci USA* 1998; **95**: 8420–8427.
- Houk BE, Bello CL, Poland B, Rosen LS, Demetri GD, Motzer RJ. Relationship between exposure to sunitinib and efficacy and tolerability endpoints in patients with cancer: results of a pharmacokinetic/pharmacodynamic meta-analysis. *Cancer Chemother Pharmacol* 2010; **66**: 357–371.



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

Supplementary Information accompanies the paper on the *npj Systems Biology and Applications* website (<http://www.nature.com/npjbsa>)

