

antiretroviral treatment. Out of these B cell clones, we identified 25 clones continuously producing MABs reactive to gp120 (Table 1). First, MABs were examined for their reactivity to the V3-peptide corresponding to the V3-region of the gp120_{JR-FL} (NNT20) and the reactive MABs, 0.5γ, 1D9, 2F8, 3E4, 3G8, 5G2, 717G2 and 16G6, were classified as anti-V3 antibodies. Later, using a set of peptides that have different V3 sequences and overlapping short peptides, we identified the minimum epitope and the cross-reactivity of these antibodies (Supplementary Table 1). The reactivity of these MABs to short peptides was decreased, and only 1D9 and 5G2 bound to 10 mer peptides. Although 2F8 did not recognize any overlapping short peptides, this MAB was capable to bind to the V3 peptide from NSI, correspondent to the CRF01_AE subtype.

MABs other than anti-V3 were classified in three groups according to the effect of soluble CD4 (sCD4) on the reactivity to gp120: CD4bs, CD4i and other epitopes (Figs. 1 and 2). MABs, 0.5δ, 4E3, 7B5, 42F9, 49G2 and 82D5, were classified as CD4bs antibodies according to the reduction of reactivity in the presence of sCD4. MABs, 4C11, 4E9C, 5D6S, 7F11, 916B2 and 917B11, were classified as CD4i antibodies according to the enhancement effect by sCD4. The rest of MABs, which did not react to the V3 peptide and did not show enhancement or inhibition by sCD4, were classified as MABs to other epitopes. Anti-V3 MAB, KD-247 (Eda et al., 2006a), CD4bs MAB, b12 (Burton et al., 1991) and CD4i MABs, 17b (Thali et al., 1993) were also analyzed as controls.

For the anti-V3 antibodies, no influence in their binding to monomeric gp120 was noted in the presence of sCD4 when analyzed by ELISA (Fig. 1). However, the enhanced reactivity by sCD4 was observed against Env on the cell surface in most of anti-V3 MABs (Fig. 2). This is consistent with the previous reports (Huang et al., 2005; Thali et al., 1993), which showed that the binding of CD4 to gp120 caused the V3 to protrude and became more available, particularly in the strains that use the CCR5 co-receptor. Although a reduction of reactivity was observed for the CD4bs MABs in ELISA, this reduction of reactivity against trimeric Env was not obvious in the MABs, such as 4E3 and 49G2 (Fig. 2).

The enhanced reactivity by sCD4 was observed in all the CD4i MABs, both in ELISA and flow cytometry analysis (Figs. 1 and 2) as previously reported for the CD4i MABs (Lusso et al., 2005; Mbah et al., 2001; Wu et al., 2008). The MABs to other epitopes did not show reactivity to trimeric Env, suggesting that these MABs recognized the unexposed region of the Env trimer (Fig. 2).

Genetic characterization of MABs

Generally, gp120 epitope reactivity is mediated by IgG1, although IgG2 can also be found (Banerjee et al., 2010). MABs predominantly consisted of IgG1. IgG3 was observed only in the “other epitope” group, and none of the antibodies isolated was IgG4 (Table 1). Representative MABs were genetically cloned, and gene usage, somatic mutation percentage and CDRH3 length was determined (Table 1). Anti-V3 antibodies showed a marked preference of the usage of the VH3-30 gene for the variable region of the heavy chain (VH) and VK2-28 for the variable region of the light chain (VL); and only the MAB 16G6 used the VH5-51 gene and λ light chain, which were previously reported as preferential gene usage by anti-V3 antibodies (Gorny et al., 2009). All the anti-V3 antibodies analyzed had the V to I substitution in position 55 (IMGT unique numbering) of FR2 (Supplementary Fig. 1). MABs, 1D9 and 5G2, used the same genes and the length of CDRH3 was also the same among them. These characteristics, as well as the close similarity in their sequences (Supplementary Fig. 1), suggested that these two MABs originated from a common ancestor. In contrast, 0.5γ, which used the same genes as 1D9 and 5G2, was considered as from another lineage, because 0.5γ had the CDRH3 different from 1D9 and 5G2 in terms of length and sequences (Table 1 and Supplementary Fig. 1). Two of the three CD4bs MABs used the same genes, VH1-18 and VL1-44, suggesting that these MABs had the same origin. All four of the CD4i MABs analyzed used the VH1-69 gene, which is consistent with previous reports (Gorny et al., 2009; Huang et al., 2004), but did not use the same light chain gene. This indicates that CD4i antibodies with the VH1-69 gene were

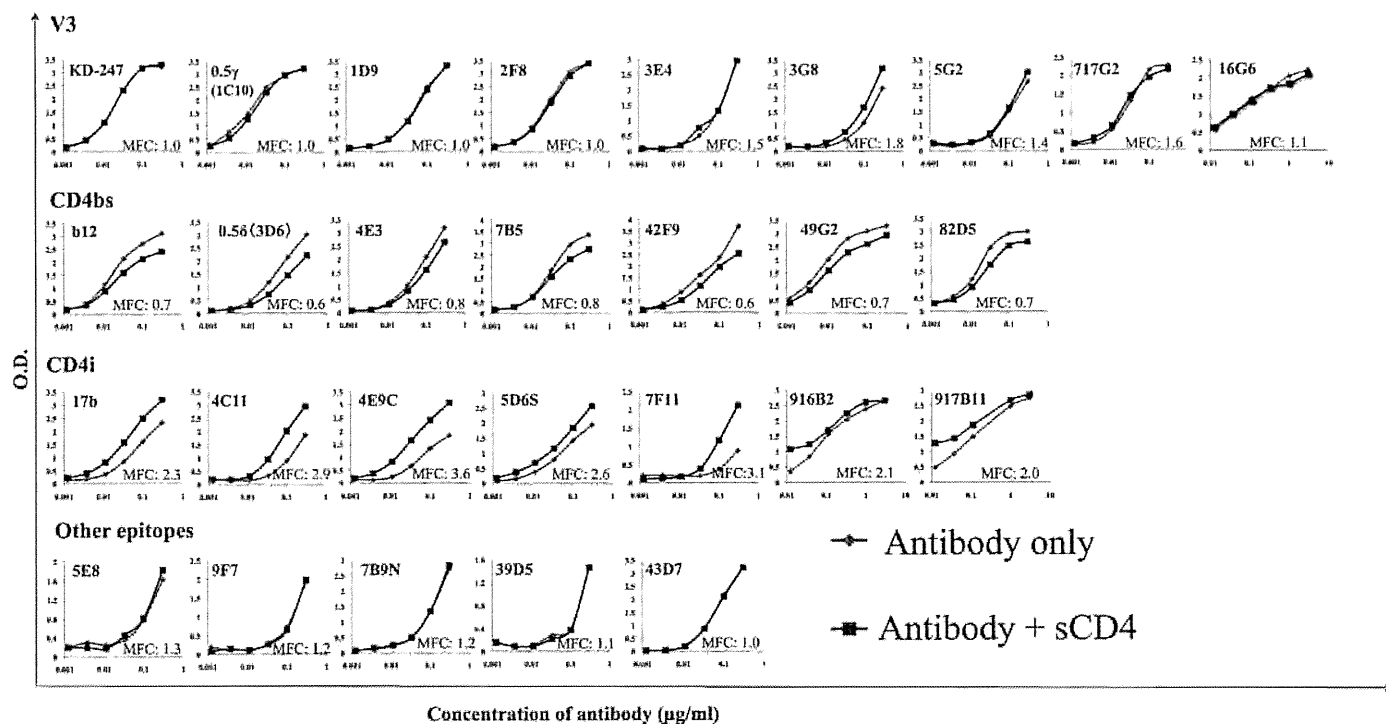


Fig. 1. Effect of sCD4 over binding of MABs isolated from patient ICTS376 to monomeric SF2_{gp120}. Reactivity to gp120 was examined for each MAB alone (gray diamond) or in the presence of sCD4 (black square) by capture ELISA assay. No effect was observed for MABs to V3 and “other epitopes”. Meanwhile, inhibition of binding was observed for MABs to CD4bs and enhancement of binding was observed for MABs to CD4i epitope. Maximum fold change (MFC) was calculated as follows: O.D. sCD4 positive/ O.D. sCD4 negative.

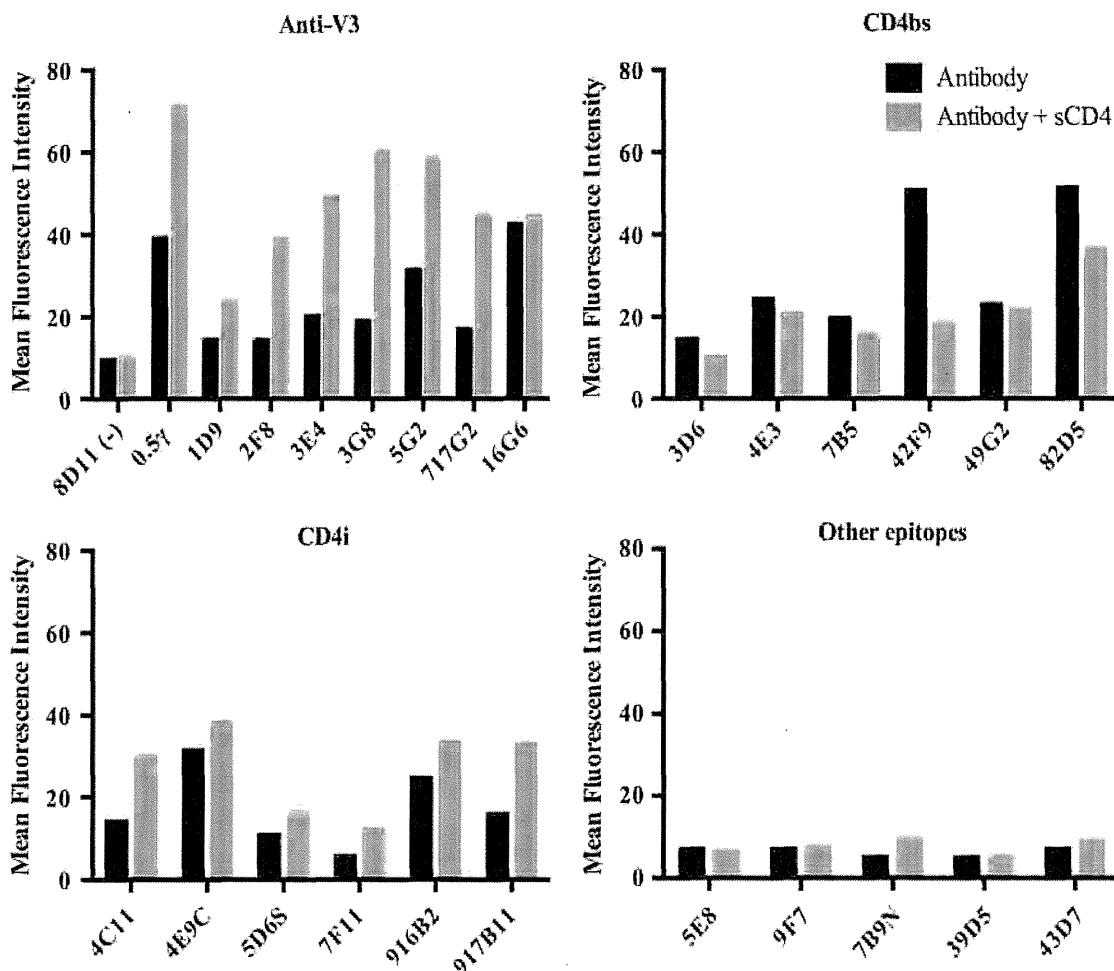


Fig. 2. Binding of MAbs from patient KTS376 to Env expressed on the surface of cells. Reactivity to PM1/CCR5 cells chronically infected with HIV-1_{JR-FL} was analyzed using flow cytometer. Mean fluorescence intensity is shown for each MAb in the presence (gray) or absence (black) of sCD4. Enhancement of binding was observed for most of the MAbs to V3 and all the CD4i MAbs when sCD4 was added. Inhibition of binding was observed for MAbs to CD4bs in the presence of sCD4. MAbs to other epitopes did not bind to trimeric Env on the cell surface.

frequently induced during infection. Abundance of tyrosines (Y) in CDHR3, which was reported previously in CD4i antibodies (Choe et al., 2003; Huang et al., 2004; Xiang et al., 2002) was present not only in CD4i MAbs but also in CD4bs MAbs. These results suggest that antibodies with preferential genes were abundantly induced in the patient KTS376, and that dominant antibody lineages were observed in the V3 and CD4bs antibodies.

Mostly, somatic hypermutation percentages were higher in VH than in VL, with the exceptions of 16G6 and 42F9 (Table 1). Overall somatic hypermutation percentages were relatively low (VH mean: 7.9% and VL mean: 4.0%) contrasting with the levels of somatic hypermutations reported for bNAbs. For example, mutation percentages of bNAbs HJ16, 2G12 and b12 were between 20 and 45% for VH and between 17 and 30% for VL; while the relatively less hypermutated bNAbs PG6 and PG9 showed percentages of 21.4% and 18.8% for VH and 21.2 and 14.1% for the VL, respectively (Klein et al., 2013; Sok et al., 2013; Wu et al., 2011; Zhou et al., 2010). The low somatic mutation percentages observed for the MAbs isolated from KTS376 corresponded to those reported for the “conventional antibodies” against HIV-1 (Zolla-Pazner, 2014).

Binding activities of the MAbs to various HIV-1 Envs

Cross-reactivity of MAbs was examined using Envs from various HIV-1 strains (Fig. 3). All the MAbs tested bound to Env from most of subtype B viruses including laboratory strains (89.6,

SF162 and YU2) and primary viruses including T/F viruses (WITO and RHPA), 6535.3 (SVPB5) and REJO4541.67 (SVPB16) (Fig. 3). The Env of the laboratory strain, NL4-3, was reactive to all the CD4bs and CD4i MAbs, but not to any of the V3 MAbs, consistent with the results observed in the V3-peptide ELISA (Supplementary Table 1 and Fig. 3). Cross-subtype binding activities against subtype A, CRF01_AE and subtype C were observed in most of CD4bs and CD4i MAbs, while most V3 MAbs exhibited significantly lower cross-subtype binding activities than CD4bs and CD4i MAbs (Fig. 3, Supplementary Fig. 2). Of the V3 antibodies, 16G6 bound all the subtype C viruses tested, and showed a considerable cross-reactivity compared with other V3 MAbs. This is consistent with the previous reports that antibodies encoded by the VH5-51/VL λ genes can recognize Envs from various subtypes (Gorny et al., 2011). These results demonstrated that these MAbs isolated from a single patient, as a set of MAbs, covered a wide range of viruses including B and other subtypes. The lack of reactivity of some MAbs was complemented with the activity of other MAbs, and at least two of the three epitopes tested (V3, CD4bs and CD4i) were recognized by these MAbs.

Neutralizing activities of the MAbs against various HIV-1 subtypes

To evaluate the neutralizing activity of these MAbs, we first used representative HIV-1 subtype B laboratory and primary isolates (Fig. 4). The neutralizing activity against SF162 and BaL

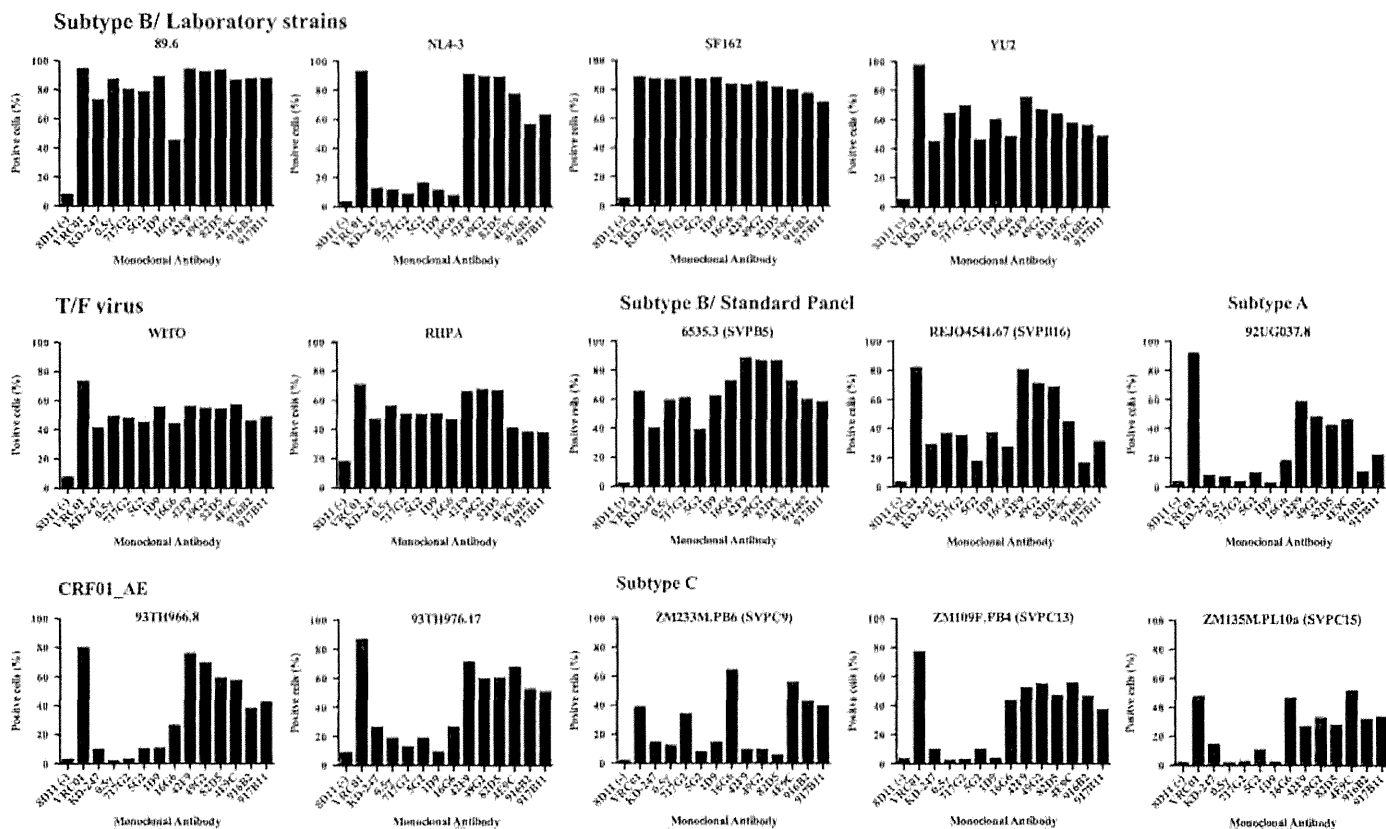


Fig. 3. Cross-reactivity of MABs from patient KTS376 to Env from various HIV-1 strains. Reactivity to Env from various HIV-1 strains, such as laboratory strains, transmitted/founder (T/F) viruses and viruses belonging to subtype A, B, C and CRF01_AE, was analyzed by flow cytometry using transfected 293T cells. Percentages of cells recognized by MABs are shown. Controls of the assay included the monoclonal IgG 8D11 (negative control), VRC01 and KD-247 (positive controls). MABs to V3 potentially bound to cells expressing Envs from all the subtype B viruses tested, with the exception of NL4-3. MABs to CD4bs and CD4i showed a greater cross-subtype reactivity more than MABs to V3 MABs.

was observed for all MABs tested, consistent with the results of binding activity. Most of the V3 MABs, with the exception of 16G6, showed a potent neutralization of laboratory strains other than IIB, and 0.5 γ , 717G2 and 1D9 were also effective against primary isolates. Although 16G6 reacted to many viruses (Fig. 3), the neutralizing activity of this MAB was weak and narrow. The CD4bs and CD4i MABs neutralized HIV-1 IIB, which was resistant to the neutralization by most of the V3 antibodies because of the two amino acid insertion adjacent to the V3-tip, while the potency and spectrum of the CD4bs and CD4i MABs were low and narrow, compared with those of V3 MABs. Especially, KTS376-96, the autologous virus from the patient that MABs were isolated, was sensitive to V3 MABs, but not to the CD4i or CD4bs MABs at all. These results suggest that the V3 antibodies mainly neutralized the viruses present in this patient.

To further evaluate the neutralizing activity of the MABs, we used a panel of pseudo-typed viruses named standard panel viruses subtype B (SVPB) and C (SVPC) (Li et al., 2006, 2005) (Fig. 4), and one subtype A and two CRF01_AE pseudoviruses. As shown in Fig. 4, 0.5 γ was highly effective against subtype B viruses, and 10 out of 12 SVPB viruses were neutralized by 0.5 γ with a half maximal inhibitory concentration (IC_{50}) below 150 μ g/ml. The coverage of subtype B viruses by 1D9 was lower than for 0.5 γ , but 1D9 showed a cross-subtype neutralization against ZM53M.PB12 (subtype C) and 92UG037.8 (subtype A). Consistent with the results of neutralization of laboratory and primary strains, the potency of CD4bs and CD4i MABs were low and narrow against standard panel viruses. However, THRO4156.18 (subtype B), which was reported as a neutralizing resistant virus (Li et al., 2005), was neutralized by the CD4bs MABs 49G2 and 42F9. In addition, TRO.11 (subtype B) and

CAAN5342.A2 (subtype B), which were not neutralized by the V3 MABs other than 0.5 γ , were sensitive to CD4bs MAB 49G2 and CD4i MAB, 4E9C, respectively. Moreover, cross-subtype neutralization activity was higher in the CD4bs and CD4i MABs, which neutralized 4 subtype C viruses, than the V3 MABs. CD4bs MAB 49G2 and CD4i MAB 4E9C significantly contributed to the cross-subtype neutralization, and five out of 12 SVPC viruses (42%) were neutralized by this set of MABs.

We also evaluated the neutralizing activity of these MABs against 10 T/F viruses of subtype B (Fig. 4) (Keele et al., 2008; Lee et al., 2009; Ochsenauber et al., 2012; Salazar-Gonzalez et al., 2009, 2008). Anti-V3 MABs neutralized 6 out of the 10 analyzed T/F viruses, and CD4i MAB 49G2 neutralized SUMA, which in contrast to V3 MABs did not neutralize. Interestingly, 5G2, which showed relatively weak and narrow neutralizing activity against other viruses, showed considerable neutralization against T/F viruses. As a result, MABs from a single patient provided coverage of 70% of the analyzed T/F viruses.

Plasma from KTS376 obtained in two different time points (2002 and 2005) was able to recapitulate the neutralizing activity of the monoclonal antibodies when confronted with representative viruses from subtype B, C and CRF01_AE (Supplementary Fig. 2).

These results demonstrated that MABs from patient KTS376 neutralized a broad range of viruses. Although the coverage of each MAB was limited, in combination they covered a considerable number of viruses from different subtypes and origins. Anti-V3 antibodies mainly provided potency and broadness against viruses inside subtype B, while CD4bs and CD4i MABs complementarily covered viruses that V3 antibodies did not neutralize, especially non-subtype B viruses.

| Viral strain | Sub-type | V3 | | | | | CD4bs | | | | CD4i | | | |
|---------------------------------------|----------|--------|-------|-------|--------|-------|-------|--------|-------|--------|-------|-------|-------|--------|
| | | KD-247 | 0.5γ | 5G2 | 717G2 | 1D9 | 16G6 | 0.56** | 42F9 | 49G2 | 82D5 | 4E9C | 916B2 | 917B11 |
| Laboratory and primary strains | | | | | | | | | | | | | | |
| IIIB | B | 16.46 | >150 | >150 | >150 | >150 | >150 | 2 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| 89.6 | B | 0.75 | <0.05 | 0.2 | <0.05 | <0.05 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | 33.44 |
| SF162 | B | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | 2.3 | 0.12 | 0.07 | 0.17 | 0.24 | 0.5 | 0.31 |
| Bal | B | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | >150 | 0.18 | 0.17 | 0.17 | <0.05 | 0.27 | <0.05 |
| JR-FL | B | 2.75 | 1.47 | 1.87 | 2.81 | 2.74 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 |
| YU2 | B | 12.33 | 1.20 | 24.8 | 11.74 | 7.61 | 13.67 | >150 | >150 | >150 | >150 | >150 | >150 | >150 |
| MOKW | B | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 | 0.42 | 0.3 | <0.05 | 0.66 | 0.1 | <0.05 | <0.05 |
| YKI | B | >150 | 13.78 | >150 | >150 | >150 | >150 | 2.4 | >150 | 132.2 | 15.82 | >150 | >150 | >150 |
| KKGO | B | >150 | 0.83 | >150 | 1.37 | 0.27 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 |
| KMT | B | >150 | 1.42 | >150 | 1.53 | <0.05 | >150 | 24 | 59.75 | 52.24 | >150 | >150 | >150 | >150 |
| KTS376-96* | B | 19.03 | 8.04 | 12.28 | 12.45 | 9.06 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 |
| Standard panel viruses | | | | | | | | | | | | | | |
| 6535.3 | B | 1.8 | 0.3 | 10.1 | 0.88 | 25.34 | 1.18 | 1.2 | 3.5 | 5.1 | 4 | 20 | 34.46 | 47.49 |
| QH0692.42 | B | >150 | 38 | >150 | 58.6 | 36.08 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 |
| SC422661.8 | B | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 |
| PVO.4 | B | >150 | 35 | >150 | >150 | 91.19 | >150 | >150 | >150 | >150 | >150 | 20 | >150 | >150 |
| TRO.11 | B | >150 | 82 | >150 | >150 | >150 | >150 | >150 | >150 | 7.2 | >150 | >150 | >150 | >150 |
| AC10.0.29 | B | >150 | 110 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 |
| RHPA4259.7 | B | >150 | 10.5 | >150 | 59.3 | 77.4 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 |
| THRO4156.18 | B | >150 | >150 | >150 | >150 | >150 | >150 | >150 | 71 | 45 | >150 | >150 | >150 | >150 |
| REJO4541.67 | B | >150 | 6.5 | >150 | 42 | 15.43 | 80.59 | 30 | 21 | 7.6 | 49.5 | >150 | >150 | 38.31 |
| TRJO4551.58 | B | 17 | 3.5 | 50 | >150 | 139.4 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 |
| WITO4160.33 | B | 28 | 1.1 | 11.5 | 51.8 | 87.04 | 104.1 | >150 | >150 | >150 | >150 | >150 | >150 | >150 |
| CAAN5342.A2 | B | >150 | 80 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | 50 | >150 | >150 |
| Du156.12 | C | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | NA | >150 | >150 | >150 |
| Du172.17 | C | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | NA | 130.1 | >150 | >150 |
| Du422.1 | C | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | NA | >150 | >150 | >150 |
| ZM197M.PB7 | C | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | NA | >150 | >150 | >150 |
| ZM214M.PL15 | C | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | NA | >150 | >150 | >150 |
| ZM233M.PB6 | C | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | 45 | NA | 42 | 10.26 | >150 |
| ZM249M.PL1 | C | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | NA | >150 | >150 | >150 |
| ZM53M.PB12 | C | >150 | >150 | >150 | >150 | 5.57 | >150 | >150 | >150 | >150 | NA | >150 | >150 | >150 |
| ZM109F.PB4 | C | >150 | >150 | >150 | >150 | >150 | >150 | 55 | >150 | 45 | NA | 42 | 35.01 | 48.65 |
| ZM135M.PL10a | C | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | 28 | NA | >150 | >150 | >150 |
| CAP45.2.00.G3 | C | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | NA | >150 | >150 | >150 |
| CAP210.2.00.E8 | C | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | >150 | NA | >150 | >150 | >150 |
| 92UG037.8 | A | >150 | >150 | >150 | >150 | 76.86 | >150 | NA | >150 | >150 | >150 | >150 | >150 | >150 |
| 93TH966.8 | CRF01_AE | >150 | >150 | >150 | >150 | >150 | >150 | NA | >150 | >150 | >150 | >150 | >150 | >150 |
| 93TH976.17 | CRF01_AE | >150 | >150 | >150 | >150 | >150 | >150 | NA | >150 | >150 | >150 | >150 | >150 | >150 |
| Transmitted/Founder viruses | | | | | | | | | | | | | | |
| WITO | B | >150 | 147.1 | >150 | >150 | 43.02 | >150 | NA | NA | >150 | >150 | >150 | >150 | >150 |
| CH058 | B | >150 | 62.86 | 84.67 | 126.65 | 25.19 | >150 | NA | NA | >150 | >150 | >150 | >150 | >150 |
| RHPA | B | 98.98 | 74.38 | 99.3 | >150 | >150 | >150 | NA | NA | >150 | >150 | >150 | >150 | >150 |
| REJO | B | >150 | >150 | >150 | >150 | 2.0 | >150 | NA | NA | >150 | >150 | >150 | >150 | >150 |
| TRJO | B | >150 | >150 | >150 | >150 | >150 | >150 | NA | NA | >150 | >150 | >150 | >150 | >150 |
| CH106 | B | >150 | >150 | >150 | >150 | >150 | >150 | NA | NA | >150 | >150 | >150 | >150 | >150 |
| CH077 | B | >150 | >150 | 16.23 | >150 | >150 | >150 | NA | NA | >150 | >150 | >150 | >150 | >150 |
| SUMA | B | >150 | >150 | >150 | >150 | >150 | >150 | NA | NA | 136.33 | >150 | >150 | >150 | >150 |
| CH040 | B | >150 | >150 | >150 | >150 | >150 | 58.4 | NA | NA | >150 | >150 | >150 | >150 | >150 |
| THRO | B | >150 | >150 | >150 | >150 | >150 | >150 | NA | NA | >150 | >150 | >150 | >150 | >150 |

Fig. 4. IC₅₀ (μg/ml) of MAbs from KTS376 against strains of HIV-1 using TZM-bl cells and single-round infection assay. The autologous virus (KTS376-96) was analyzed together with other laboratory and primary strains. Analysis of laboratory and primary strains for 0.58 was performed using the MTT assay of supernatant of infected PM1/CCR5 cells. Not available results are denoted as NA. Color code is as follows: Red: IC₅₀ 0.05–10; Orange: IC₅₀ > 10–50; Yellow: IC₅₀ > 50–150 μg/ml.

Synergistic effects of anti-V3 and CD4bs antibodies against HIV-1

Recent studies on functional trimers of envelopes suggest that exposure of V3-epitope and coreceptor binding sites including the CD4i epitope occurs following the interaction of gp120 with CD4 (Kwong et al., 2002, 1998; Liu et al., 2008; Mbah et al., 2001). In fact, sCD4-mediated enhancement was observed in the V3 and CD4i MABs (Fig. 2), as has been previously reported (Lusso et al., 2005; Thali et al., 1993; Wu et al., 2008). We then tested whether antibodies to CD4bs have the capacity to alter the envelope conformation and induce some critical epitopes accessible for the antibodies of other specificities. We first tested the binding activity of MABs in the presence or absence of the CD4bs MAB 0.5 δ by ELISA (Fig. 5). Marked enhancement of binding of 0.5 γ was observed in the presence of 0.5 δ . The enhancement was also observed for the other V3 antibody 3E4, although the effect was much higher in 0.5 γ than in 3E4. In contrast, suppression in the binding of 0.5 δ itself and CD4i MAB 4C11 was observed in the presence of 0.5 δ . This suggests that the 0.5 δ epitope may overlap with that of 4C11, or that the binding of either MAB may interfere with the other sterically.

Then, we tested the effect of MABs combinations, such as 0.5 γ (V3) and 0.5 δ (CD4bs), 1D9 (V3) and 49G2 (CD4i), and 16G6 (V3) and 17G2 (V3) over neutralization. The combinations of MABs showed an additive effect in most of cases, while the combination of 0.5 γ and 0.5 δ showed a synergistic effect in the neutralizing activity of HIV-1_{JR-FL} (Fig. 6). The combination of these MABs was shown to be more effective than either MAB alone and the synergy of neutralizing activity observed for the combination of 0.5 γ and

0.5 δ was up to 25% (Supplementary Table 3). This result suggests that these antibodies, besides their complementarity, can act synergistically. This phenomenon of synergism and complementarity may play an important role "in vivo" for suppressing viruses in a patient.

ADCC activities of the MABs

It has been reported that antibodies contribute to protection by its effector-mediated functions such as ADCC (Haynes et al., 2012a; Liao et al., 2013; Xiao et al., 2010). Then, we determined the ADCC activities of MABs from patient KTS376. First, we analyzed cells coated with gp120 from SF2 and cells infected with BaL and YU2 (Fig. 7, Supplementary Fig. 3). Significant ADCC activity was observed in most of the MABs from the three groups. The CD4i MAB 916B2 showed neutralizing activity against BaL, but did not show any statistically significant ADCC activity against BaL. The 916B2 epitope may require the CD4 binding to Env, as reported previously (Veillette et al., 2014). ADCC activity of MABs from patient KTS376 was also evaluated against cells infected with five T/F virus strains (Fig. 8, Supplementary Fig. 4). ADCC activity for the T/F viruses was relatively weak compared with the response observed for gp120, BaL and YU2. THRO was sensitive to all the MABs tested, while ADCC activity against CH106 was statistically significant in CD4bs MAB 49G2. Interestingly, the V3 MABs acted complementarily against CH040 and REJO. ADCC activity against CH040 was observed in MABs, 0.5 γ and 16G6, and the other V3 MABs were effective against REJO. Despite the fact that no neutralizing activity was observed for THRO, all the MABs showed

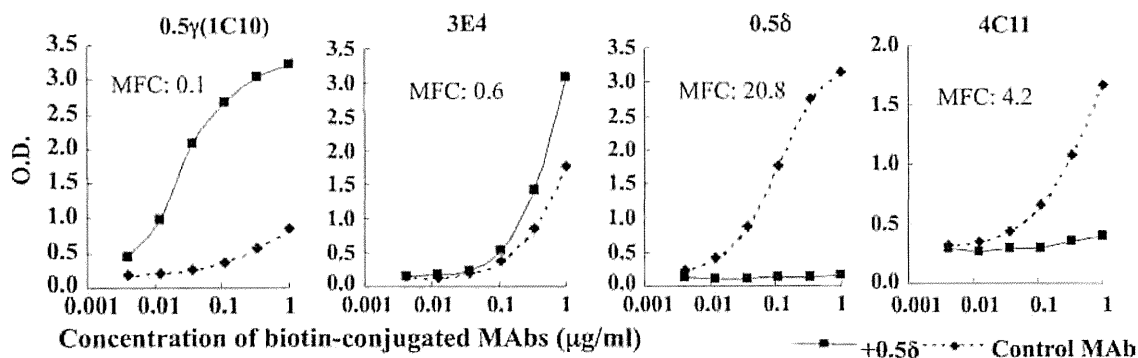


Fig. 5. Effect of CD4bs MAB 0.5 δ on reactivity of other MABs. Reactivity of MABs was analyzed by ELISA for biotin conjugated antibody alone (dotted line) or in combination with 0.5 δ (solid line). Marked enhancement of reactivity in the presence of 0.5 δ was observed in the MAB to V3 0.5 γ . Another anti-V3 MAB 3E4 showed only a slight enhancement by 0.5 δ , and abrogation of reactivity was evident for 0.5 δ and 4C11. Maximum fold change (MFC) was calculated as follows: O.D. 0.5 δ positive/ O.D. 0.5 δ negative.

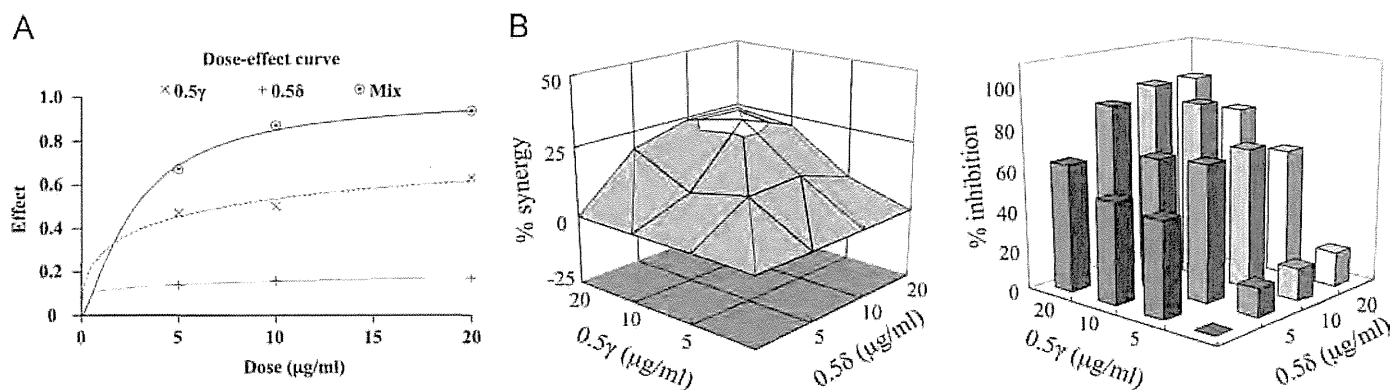


Fig. 6. Synergistic neutralizing activity of the anti-V3 MAB 0.5 γ and the CD4bs MAB 0.5 δ . Neutralization activity of 0.5 γ alone (X symbol), 0.5 δ alone (+ symbol) and 1:1 combination of both MABs (Circle and dot symbol) was evaluated and is shown as a dose-effect curve (A). The percentages of synergy (B) and viral inhibition (C) reached by combinations of 0.5 γ and 0.5 δ are shown. Neutralizing activity was analyzed against HIV-1_{JR-FL}. Synergy between 0.5 γ and 0.5 δ was calculated using the Chou–Talalay method and 3 dimensional analyses.

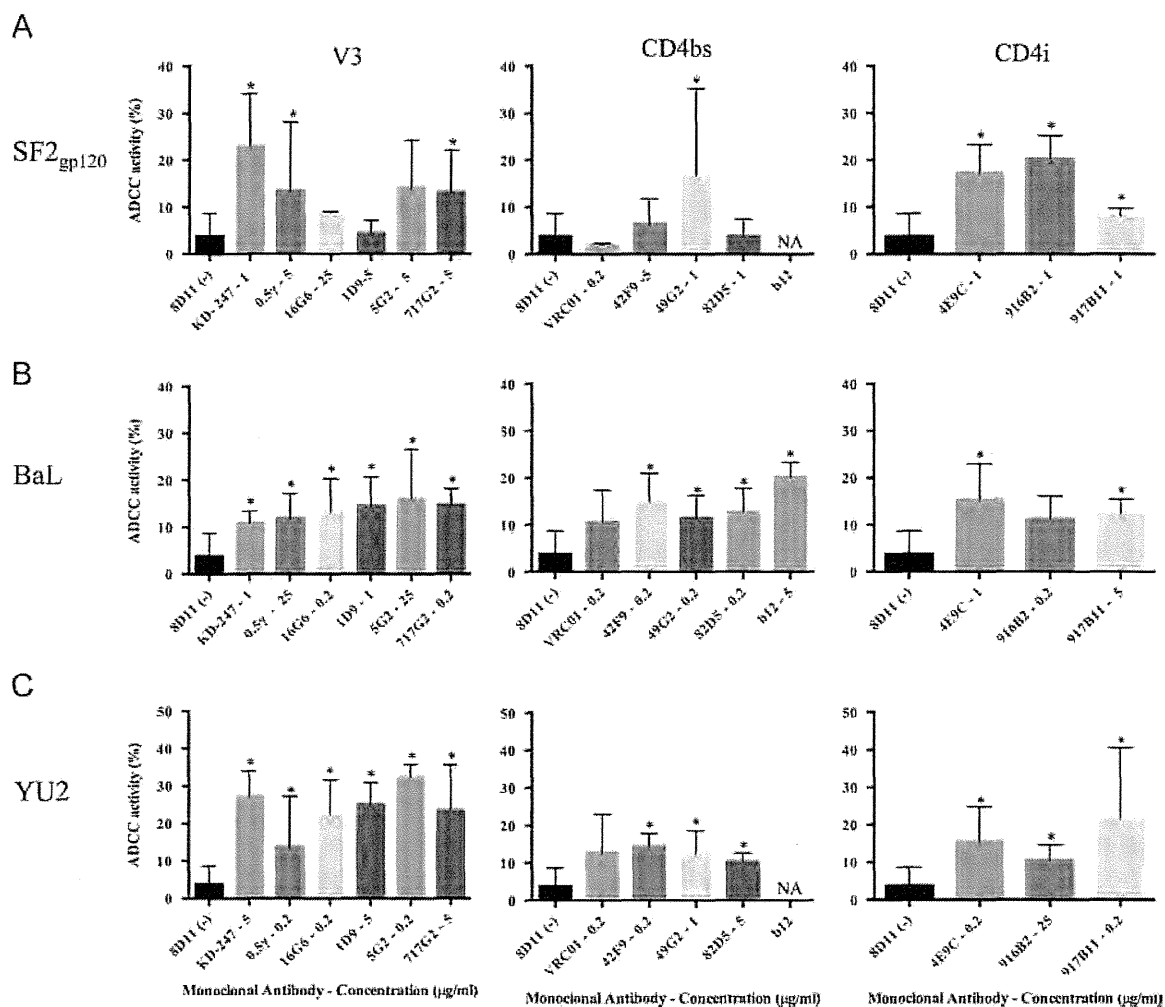


Fig. 7. ADCC activity of MABs against CEM.NKr.CCR5 cells coated with SF2_{gp120} (A), infected with BaL (B) and YU2 (C). ADCC activity was evaluated using CEM.NKr.CCR5 cells stained with CFSE and PKH-26 as target cells and PBMC (A and C) or NK-enriched PBMC (B) from healthy donors as effector cells. MAB concentration between 0.2 and 25 $\mu\text{g/ml}$ was analyzed, and the maximum percentages of killing are shown for each MAB. Concentration of MAB ($\mu\text{g/ml}$) is indicated together with MAB name. Asterisks correspond to values statistically different from the negative control ($p < 0.05$) calculated with the Mann-Whitney U test.

significant ADCC activity against this virus. This indicates that ADCC activity is not correlated with neutralizing activity. However, CH106 was resistant to both neutralization and ADCC.

These results demonstrated that MABs from a single donor presented ADCC activity against HIV-1 strains including T/F viruses. The coverage of a wide range of HIV-1 strains by ADCC supports the complementary character of these MABs from a single patient, not only for the neutralizing activities, but also for the non-neutralizing activities.

Presence of KTS376-like antibodies in plasma samples

In order to determine the presence of antibodies with the specificity similar to the MABs from patient KTS376, plasma samples from various patients were examined for the ability to block binding of 0.5 γ , 3E4, 0.5 δ and 4C11 (Fig. 9, Supplementary Fig. 5). Plasma samples from HIV-1-infected patients, which included progressors ($N=47$), slow progressors ($N=25$), long-term non-progressors, LTNP ($N=20$) and non-B-infected ($N=11$) patients, significantly suppressed the binding of CD4bs MAB 0.5 δ and CD4i MAB 4C11. This is consistent with a previous observation that most of the patients have antibodies to CD4bs and CD4i (Binley et al., 2008; Dhillon et al., 2007; Gnanakaran et al., 2010; Moulard et al., 2002; Sather et al., 2009). The V3 MABs, 0.5 γ and 3E4, were not inhibited significantly by plasma samples from non-

B-infected patients, and this maybe due to the poor cross-subtype reactivity of the V3 MABs (Fig. 3). Disease progression was associated with the low inhibition of MABs, and the plasma samples from progressors inhibited the binding of MABs less strongly than those from LTNP. Accordingly, the inhibition of 0.5 γ , which was weakly inhibited by plasma samples compared with other MABs, was not significant in progressors. However, considerably longer times of infection in the long-term non-progressor and the slow progressor groups may have contributed to affinity maturation of antibodies in these groups. This suggests that the presence of antibodies similar to those in patient KTS376 may contribute to the control of HIV-1 infection. On the other hand, KTS376-like antibodies were found in most of HIV-1-infected patients regardless of their disease status, suggesting that antibodies against V3, CD4bs and CD4i were induced in most HIV-1-infected patients similarly to patient KTS376.

Discussion

In the present study, we isolated 25 MABs from a patient, KTS376, who had suppressed HIV-1 infection for more than 25 years without any treatment, and further analyzed these MABs to clarify the mechanism to controlling HIV-1 infection. A wide coverage of viruses by a set of monoclonal antibodies from

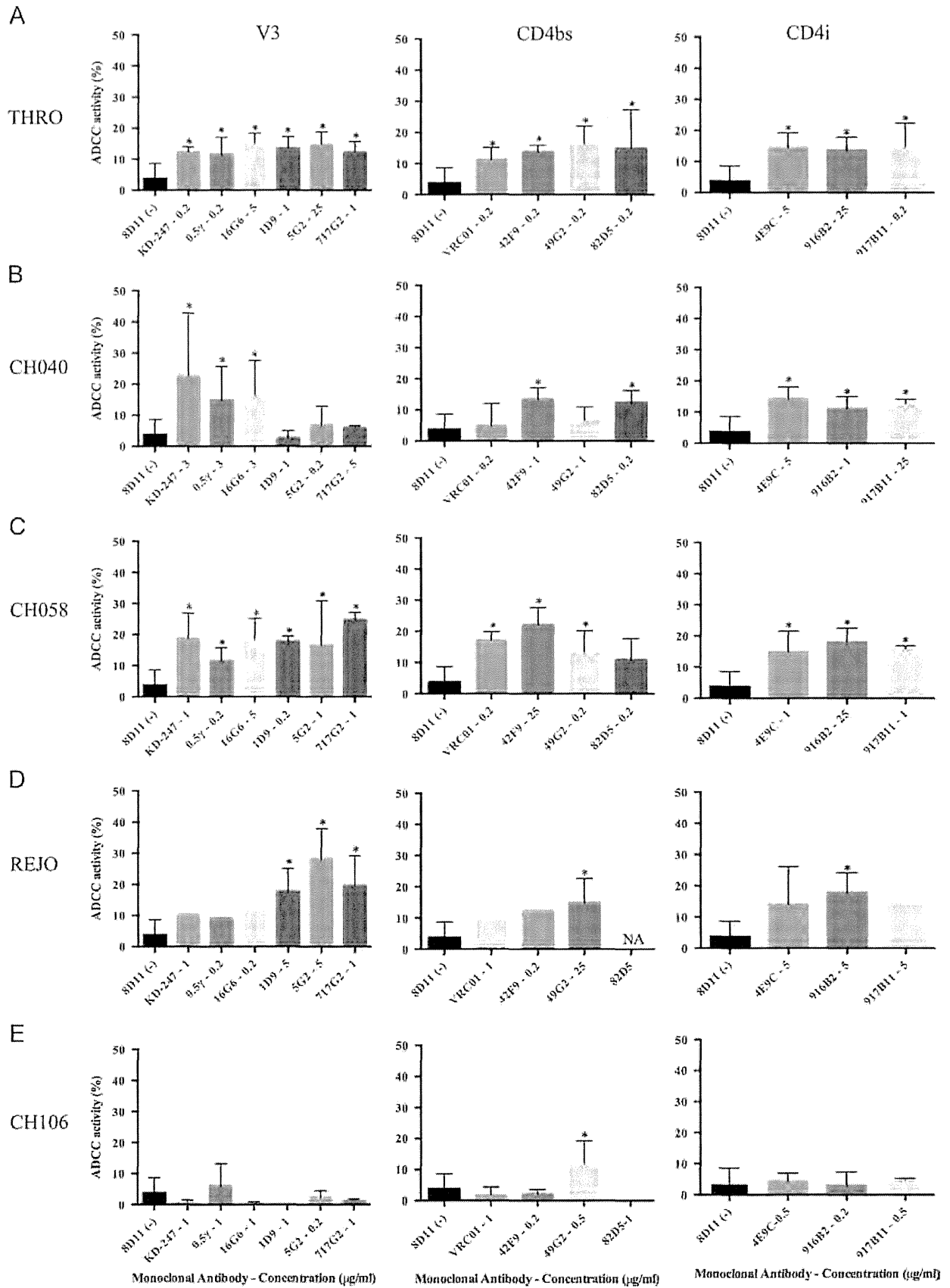


Fig. 8. ADCC activity of MAbs against CEM.NKr.CCR5 cells infected with T/F viruses, THRO (A), CH040 (B) CH058 (C) REJO (D) and CH106 (E). ADCC activity was evaluated using CEM.NKr.CCR5 cells stained with CFSE and PKH-26 as target cells and PBMC (C, D and E) or NK-enriched PBMC (A and B) from healthy donors as effector cells. MAb concentration between 0.2 and 25 µg/ml was analyzed, and the maximum percentages of killing are shown for each MAb. Concentration of MAb (µg/ml) is indicated together with MAb name. Asterisks correspond to values statistically different from the negative control ($p < 0.05$) calculated with the Mann–Whitney U test.

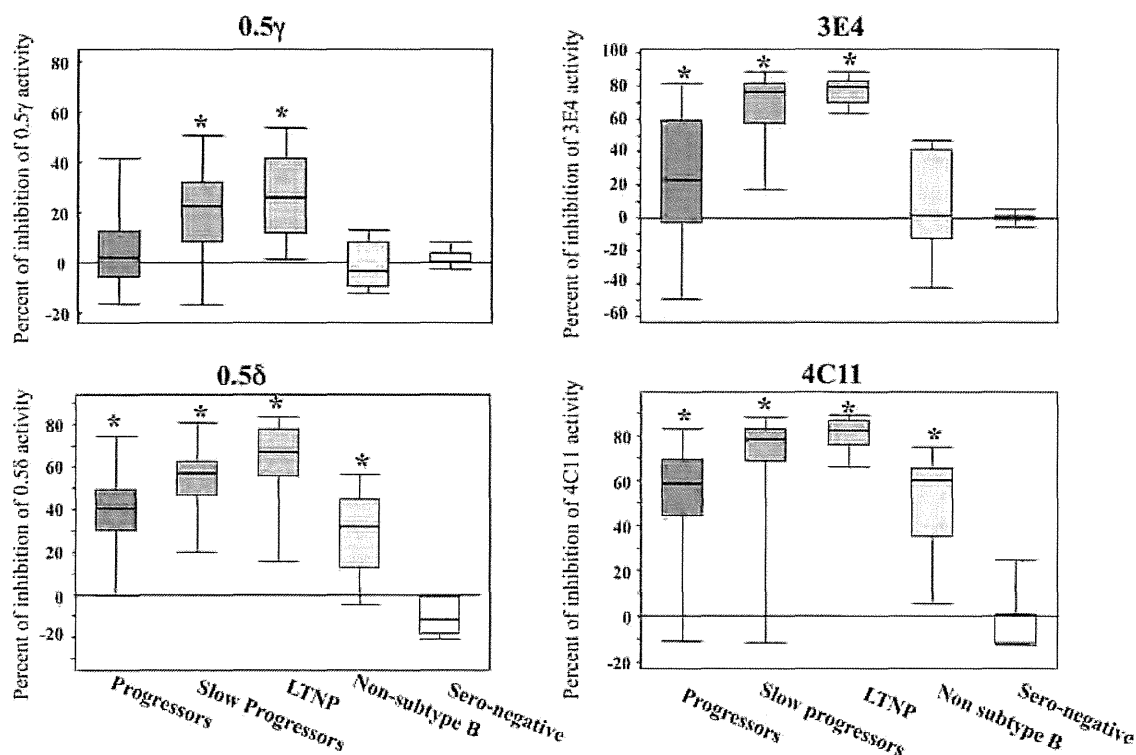


Fig. 9. Induction of antibodies with the specificity similar to MAbs from KTS376 in HIV-1-infected patients. Competition ELISA was performed using biotin-conjugated MAbs, 0.5 γ , 3E8, 0.5 δ and 4C11, and plasma samples (1:50 dilution) as a competitor. Plasma samples were obtained from progressors ($N=47$), slow progressors ($N=25$), Long-term non-progressors (LTNP, $N=20$), Non-subtype B infection ($N=11$) and sero-negative ($N=7$) individuals. Asterisks correspond to values statistically different from those of sero-negative individuals ($p < 0.05$) calculated by Student's t -test. The estimated time of infection with HIV-1 of patients from the long-term non-progressor group and the slow progressor group were 23 and 15 year, respectively. Time of infection for the other groups is not available.

KTS376 suggests that the humoral response played an important role in the long-term suppression of the patient. However, we cannot rule out other mechanisms, such as cellular response and viral fitness. The moderate neutralizing potency and breadth indicates that these MAbs are not classified as broad neutralizing antibodies, but rather as “conventional antibodies”. This type of antibodies was recently described by Zolla-Pazner, as antibodies that have limited breadth and potency in standard neutralization assays and are commonly induced in HIV-1-infected patients. Conventional antibodies are frequently elicited during infection, and induced by many vaccine candidates tested to date, but none have yet induced broadly reactive neutralizing antibodies (Zolla-Pazner, 2014). Although each conventional antibody has only a limited breadth in neutralization, the considerable cross-reactivity of MAbs from patient KTS376, indicates that conventional antibodies can cover a wide range of viruses.

Anti-V3 MAbs had the higher frequency and potency of neutralizing activity when compared with MAbs from other specificity, especially against subtype B viruses. V3 is highly immunogenic and it is estimated that about half of the antibody responses against HIV-1 Env in patient serum is directed against this region (Javaherian et al., 1989; Moore et al., 1994; Zolla-Pazner, 2004) and at a sequence level, V3 is more conserved in comparison with the other variable regions of gp120 (Jiang et al., 2010). The anti-V3 MAbs 0.5 γ and 1D9 showed the broadest neutralizing activities, comparable to that of other anti-V3 MAbs with cross-reactivity, as previously reported (Andrabi et al., 2013). MAb 16G6 has a remarkable cross-binding activity, consistent with reports of anti-V3 antibodies derived from the VH5-51 gene (Gorny et al., 2011). However, 16G6 has a weak neutralizing activity and this may be related with its low somatic mutation percentage. Despite the fact that anti-V3 MAbs frequently depend on the “GPGR” motif in the V3 tip, viruses with a different motif,

such as CH040 (GPGQ) and CH077 (GALR), were neutralized by these V3 MAbs. The induction of such antibodies with a relatively broad reactivity may require new approaches for designing V3-scaffold immunogens and adjuvants based on previous observations (Zolla-Pazner et al., 2011). In addition, the high frequency of 0.5 γ -like antibodies in LTNP suggests that maturation of B cells, which is much shorter than those for bNAbs, may be important to induce V3 antibodies with cross-reactivity.

CD4bs MAbs are reported as the earliest cross-neutralizing response in patients (Andrabi et al., 2013; Jiang et al., 2010; Zolla-Pazner et al., 2011) and is not rare to find them in HIV patients' sera. Patient KTS376 was not the exception and several CD4bs MAbs with cross-neutralizing activity, even against non-B subtype viruses, were isolated. However, as it is also observed for antibodies other than CD4bs, higher concentration of antibody was required for efficient neutralization. These types of antibodies could also contribute to enhancing the V3 MAbs response, as was observed when the anti-V3 0.5 γ was combined with the CD4bs MAb 0.5 δ , showing the complementary and synergistic potential of CD4bs MAb. Synergy of antibodies was reported to contribute to HIV-1 control in a vaccine study (Pollara et al., 2014), and in this study we showed that synergy was also observed in a combination of antibodies derived from a single donor. This suggests that this synergy phenomenon can contribute to suppression of HIV-1 in vivo.

CD4i MAbs are induced quite frequently in HIV-1-infected patients (Decker et al., 2005). CD4i MAbs isolated from patient KTS376 were not as potent as the V3 MAbs, but were able to neutralize viruses that were not covered by anti-V3 and CD4bs MAbs; thus providing a complementarity to the antibody set. Identification of at least four lineages of CD4i antibodies, which were revealed from a genetic analysis of light chain genes, indicates the vigorous induction of CD4i antibodies in this patient.

CD4i antibodies may not contribute significantly to protection from HIV-1 infection, but may play an important role in controlling HIV-1 replication after HIV-1 infection. Some CD4i antibodies bound to a hybrid epitope conformed by the CD4 and the gp120, and for these antibodies, the presence of CD4 is indispensable for binding (Lewis et al., 2011). However, our MABs were able to bind to gp120 even without CD4, which suggests that some part of the epitope may be exposed even before binding to CD4.

Low somatic hypermutation rates of MABs from patient KTS376 contrasts with those of the bNABs. A high mutation percentage is a consequence of the affinity maturation of antibodies, and is one of the main obstacles for induction of bNABs by vaccination. In fact, a positive correlation between the level of somatic hypermutation and the development of strength and breadth of the neutralizing activity of an antibody was reported (Sok et al., 2013). However, our data show that MABs even with low levels of affinity maturation are capable of binding and neutralizing a wide range of viruses, when antibodies targeting V3, CD4bs and CD4i acted complementarily and we observed also synergy. Synergy of neutralizing activity by two antibodies with different specificity has been already reported. Pollara et al. (2014) recently showed that C1 and V2 monoclonal antibodies isolated from RV144 vaccinees have synergistic activities for neutralization, infectious virus capture, and ADCC, and proposed that synergy among antibodies with different epitope specificities contributes to HIV-1 antiviral antibody responses. Although it is difficult to compare our results with the results reported by them due to the use of different experimental settings, they showed a 20- to 65-fold decrease of IC_{50} s by combination of antibodies.

Besides neutralizing activity of the antibodies, ADCC activity has been reported to have great importance in the protection and control of the HIV-1 infection (Haynes et al., 2012a; Rerks-Ngarm et al., 2009; Wren et al., 2012). Antibodies with ADCC activity tend to have cross- or broad- activity (Smalls-Mantey et al., 2012). In addition, ADCC-mediated natural killer cell activation was significantly broader and stronger in HIV controllers than in HIV progressors (Madhavi et al., 2014). We showed significant ADCC activities of MABs against various viruses, especially T/F viruses. The *in vitro* evaluation of ADCC presents certain obstacles, and one of them is a lack of suitable target cells. We used CEM.NKr.CCR5 cells as target cells, but the susceptibility of these cells to HIV-1 infection is not high due to the presence of restriction factors (Han et al., 2008). This limited the number of primary isolates evaluated, because infections could not be established for many viruses. The factors behind the presence or magnitude of the ADCC responses are not fully understood. It was reported that neutralizing and gp120 titers did not predict ADCC activity (Alpert et al., 2012). Although binding to Env is necessary for ADCC activity, we cannot observe a correlation between these two factors. Interaction of gp120 with CD4 was shown to changed ADCC activity (Veillette et al., 2014) and this could have important implications for the conformational status of Env in the infected cells. Possible causes may include the interactions between epitope and paratope and the differences in the level of epitope expression required for ADCC activity. However, the exact reasons behind this lack of correlation among binding, neutralizing and ADCC activities remains unclear, and will be a topic of future research.

A wide coverage of HIV-1 viruses by bNABs has been previously reported (Walker et al., 2011) and great effort is invested in developing this type of antibodies by vaccination (Zolla-Pazner, 2014); however, this study supports the rationale that “conventional antibodies” are capable to provide considerable levels of coverage, as we observed that the antibodies derived from a single patient had complementary and synergistic neutralizing activity and ADCC activity. The use of mixture of Env had been proposed as

a way to achieve the induction of polyclonal response in animal models (Nkolola et al., 2014; Seaman et al., 2007, 2005). Although the induction of antibodies to specific epitopes will require further studies, antibody induction to a specific epitope was demonstrated using the antigen containing epitope with scaffold (Zolla-Pazner et al., 2011). Elicitation of antibodies with characteristics similar to MABs from this patient, which is much easier than elicitation of highly mutated bNABs, could be a strategy for developing an effective vaccine against HIV-1.

Material and methods

Study participant

B cells were isolated from patient KTS376. During the period of B cell isolation, CD4 cells count was above 200/ μ l (mean of 17 visits; 297/ μ l) and viral load was below 200 copies/ml (mean of 17 visits; 44.3 copies/ml), and the time frame corresponded to 23 to 28 years after infection with HIV-1. Also, KTS376 did not receive any antiretroviral treatment and besides having Hepatitis C virus co-infection, did not present any symptoms or signs of immunodeficiency. Human blood samples were collected from KTS376 after signed informed consent in accordance with the study protocol and informed consent was reviewed and approved by the Ethics committee for clinical research & advanced medical technology at the Kumamoto University Medical School.

Cells, reagents and viruses

TZM-bl (Derdeyn et al., 2000; Lambotte et al., 2009; Platt et al., 1998; Takeuchi et al., 2008; Wei et al., 2002), 293A (Life Technologies, Carlsbad, CA) and 293T (Graham et al., 1977) cells were maintained in Dubelcco's modified Eagle medium (DMEM; Sigma, St. Louis, MO) and supplemented with 10% heat-inactivated fetal calf serum (FCS; Thermo Scientific, Waltham, MA). CEM.NKr.CCR5 cells (Howell et al., 1985; Lysterly et al., 1987; Trkola et al., 1999) were maintained in RPMI-1640 medium (Sigma, St. Luis MO) supplemented with 10% FCS. PM1/CCR5 cells (Monde et al., 2007) were maintained in RPMI-1640 supplemented with 10% FCS and 100 μ g/ml of G418 sulfate (Calbiochem-EMD Millipore, Billerica, MA). PM1/CCR5 cells chronically infected with HIV-1_{JR-FL} were maintained in RPMI-1640 supplemented with 10% FCS. Recombinant human soluble CD4 (sCD4) was purchased from R&D systems (Minneapolis, MN). The primary HIV-1 strains, MOKW, YKI, KKGO, KMT and KTS376-96 were isolated from drug-naïve Japanese patients (Maeda et al., 2001). These viruses were passaged in phytohemagglutinin (PHA)-activated peripheral blood mononuclear cells (PBMCs) and the culture supernatant was stored at -80°C until use. KD-247, an anti-V3 humanized mouse antibody, was produced by transferring the complementary region genes from the mouse hybridoma clone C25 into genes of the human V region as previously described (Eda et al., 2006b). CD4i MAb 17b was a kind gift from J. Robinson (Department of Pediatrics, Tulane University Medical Center, New Orleans, LA). CD4bs VRC01 was produced by Freestyle 293 expression system (Invitrogen, Life Technologies, Carlsbad, CA). Env plasmids of the Standard Virus Panel for subtype B and C strains (Li et al., 2006, 2005) were used for pseudovirus preparation and flow cytometry analysis. TZM-bl cells, CEM,NKr.CCR5 cells, VRC01 plasmid, SPVB and SPVC Env plasmids, and T/F plasmids were obtained through the NIH AIDS Reagent program, Division of AIDS, NIAID, NIH, by the kind contribution from Dr. John C. Kappes, Dr. Xiaoyun Wu, Tranzyme Inc., Dr. D. Montefiori, Dr. F. Gao, and Dr. Christina Ochsenbauer.

Pseudovirus and infectious molecular clones preparation

Pseudoviruses were prepared by transfecting exponentially dividing 293T cells with 5 µg of *rev/env* expression plasmid and 10 µg of an *env*-deficient HIV-1 backbone vector (pSG3ΔEnv), using Lipofectamin 2000 transfection reagent (Invitrogen, Life Technologies, Carlsbad, CA). Pseudovirus-containing culture supernatants were harvested 48 h after transfection, filtered (0.45 µm), and stored at –80 °C until use. Plasmids from the transmitted/founder viruses pWITO.c/2474, pCH058.c/2960, pRHPA.c/2635, pREJO.c/2864, pTRJO.c/2851, pCH106.c/2633, pCH040.c/2625, pCH077.t/2627, pSUMA.c/2821, and pTHRO.c/2626 (Keele et al., 2008; Lee et al., 2009; Salazar-Gonzalez et al., 2009, 2008) were used for producing infectious molecular clones by transfecting 5 µg of plasmid to 293T cells as described above.

Monoclonal antibody isolation

Transformation of B-cells with Epstein-Barr virus (EBV) and subsequent cloning were performed as previously described elsewhere (Matsushita et al., 1986). PBMC were separated from a patient with hemophilia A (KTS376) who had been infected with HIV-1 for more than 25 years without any antiretroviral treatment, by density gradient centrifugation of heparinized blood using Ficoll-paque (GE Healthcare, Cleveland, OH). Supernatants from individual clones was screened for the presence of IgG antibody that binds to the monomeric SF2 gp120 in a gp120 capture assay (Moore et al., 1994). Briefly, gp120 from SF2 (Austral Biologicals, San Ramon, CA) was captured onto solid phase via their carboxyl termini using sheep polyclonal antibody D7324 (Aalto Bioreagents, Dublin, Ireland). Supernatant samples were added to wells and IgG bound to gp120 was detected with alkaline phosphatase-conjugated goat anti-human IgG (Sigma, St. Luis MO) followed by addition of phosphatase substrate (Sigma, St. Luis MO). A₄₀₅ measurements were taken using a microplate reader (Biorad, Hercules, CA).

Construction of recombinant IgG

Recombinant IgG from antibodies 16G6, 1D9 and 7171G2 were constructed as described some lines below, and used exclusively in the posterior analysis. RNA was extracted from B cells of interest with the RNeasy mini kit (Qiagen, Venlo, Netherlands). First-strand cDNA was synthesized using oligo(dT)₂₀ (Toyobo, Osaka, Japan) and the SuperScript III Reverse transcriptase (Invitrogen, Life Technologies, Carlsbad, CA). The heavy chain variable regions were amplified using the following primers, LH1 (5'-TGG TGG CAG CCA CCG GTG CCC ACT CCC AGG TGC AG-3'), LH3 (5'-TGG TGG CAG CAG CCA CCG GTG TCC AGT GTG ARG TGC AG-3'), LH46 (5'-TGG TGG CAG CAG CCA CCC GGG TCC TGT CCC AGG TGC AG-3'), LH5 (5'-TGG TGG CAG CAG CCA CCG GAG TCT GTT CCG AGG TGC AG-3') and APS-R (5'-GGG GGA AGA CCG ATG GGC-3'). The light chain κ variable regions were amplified using primers, GKV1 (5'-GGT GCC TAC GGG GAT ATC CAG ATG ACC CAG TCT CC-3'), GKV24 (5'-GGT GCC TAC GGG GAT ATT GTG ATG ACY CAG TCT CC-3'), GKV3 (5'-GGT GCC TAC GGG GAT ATT GTG WTG ACR CAG TCT CC-3'), GKV5 (5'-GGT GCC TAC GGG GAT ACG ACA CTC ACG CAG TCT CC-3') and HCK5-B (5'-GAA GAC AGA TGG TGC AGC CAC AGT-3'). The light chain λ variable regions were amplified using primers, LLS1 (5'-ACT TTC TGC ACA GGC TCC TGG GCC CAG TCT GTG CTG-3'), LLS2 (5'-ACT TTC TGC ACA GGC TCC TGG GCC CAG TCT GCC CTG-3'), LLS3 (5'-ACT TTC TGC ACA GGC TCT GTG ACC TCC TAT GAG CTG-3'), LLS45 (5'-ACT TTC TGC ACA GGC TCT CTC TCS CAG CYT GTG CTG-3'), LLS6 (5'-ACT TTC TGC ACA GGC TCT TGG GCC AAT TTT ATG CTG-3'), LLS7 (5'-ACT TTC TGC ACA GGC TCC AAT TCY CAG GCT GTG GTG-3'), LLS8 (5'-ACT TTC TGC ACA GGC GTG GAT TCT CAG

ACT GTG GTG-3') and VL-R (5'-CAG TGT GGC CTT GTT GGC TTG-3'). Polymerase chain reaction (PCR) was performed using the following conditions by Platinum Taq DNA Polymerase High Fidelity (Invitrogen, Life Technologies, Carlsbad, CA): 94 °C for 30 s, followed by 35 cycles of 94 °C for 15 s, 55 °C for 15 s, and 68 °C for 60 s. The PCR products of the heavy chain, light chain κ and light chain λ were cloned into vectors pIgGH, pKVA2 and pLSH, respectively, using the GeneArt Seamless Cloning and Assembly kit (Invitrogen, Life Technologies, Carlsbad, CA). The vector pIgGH was basically constructed from pIgG (Rader et al., 2002), pHCG (Kuwata et al., 2011) and the plasmid to produce MAb 1C10 (0.5y) heavy chain fragment. Briefly, the VH gene was amplified from a B cell line producing MAb 1C10 with primers HFabVH3a-F and HFabVHJb-B (Barbas et al., 2001). The CH1 region was amplified using pCOMB3XTT and primers, HlgGCH1-F and dpseq (Barbas et al., 2001). These PCR products were combined by PCR with primers SgrVH135F (5'-GCC ACC GGT GCC CAC TCC SAG GTG CAG CTG KTG-3') and dpsp-R (5'-CAG TTT AAA CCT AAG AAG CGT AGT CCG GAA C-3'). The connected PCR product was digested with *SgrAI* and *Apal* and inserted into pHCG. The resulting p1C10Fab to produced the 1C10 heavy chain fragment. The *XbaI*-*Apal* fragment from pCOMB3XSS and the *Apal*-*PmeI* fragment from the PCR product, which was amplified using primers, HlgGH1-F (5'-GCC TCC ACC AAG GGC CCA TCG GTC-3') and CH-R (5'-AGG TTT ACT AGT ACC ACC ACA TGT TTT TAT CTC-3'), and pIgG as a template, were inserted into p1C10Fab. The resultant pIgGH has the signal peptide region and the constant regions of IgG, and was used to clone VH genes after removing the stuffer sequence by digestion with *SgrAI* and *Apal*. The vector pKVA2 was constructed by insertion of two PCR products into pcDNA3.1/Hygro(+) (Invitrogen, Life Technologies, Carlsbad, CA) and was used to clone VK genes after digestion with *EcoRV* and *AfeI*. The signal peptide region was amplified using the primers, SPKNH-F (5'-AAG CTA GCA TGG TGT TGC AGA C-3') and SPKEV-R (5'-AAG ATA TCC CCG TAG GCA CCA GAG-3'), and pIgG as a template, and the *NheI*-*EcoRV* fragment was used for the insertion. The light chain κ constant region was amplified using the primers, EcVAfe-F (5'-CCG ATA TCT TAG CGC TGC ACC ATC TGT CTT C-3') and BGH-R (5'-TAG AAG GCA CAG TCG AGG-3'), and p1C10L, which had the *HindIII*-*XbaI* fragment containing MAb 1C10 light chain in pcDNA3.1/Hygro(+). The vector pLSH was constructed by inserting the *HindIII*-*HpaI* fragment and the *HpaI*-*XbaI* fragment, which were amplified using p916B2L and primer pairs, T7-F and *HpaSfo*-R, and *Hpa*-F and BGH-R, respectively, into pcDNA3.1/Hygro(+). The plasmid p916B2L was constructed by inserting the *NheI*-*XbaI* fragment, which contained the signal peptide region from pLL-B404 to producing the light chain of MAb B404 (Kuwata et al., 2011) and the light chain λ gene from MAb 916B2, into pcDNA3.1/Hygro(+). Plasmids, pIgG, pCOMB3XSS and pCOMB3XTT were kindly provided from Dr. Barbas (The Scripps Research Institute).

Recombinant antibodies were obtained by co-transfection of heavy and light chain plasmids to 293A cells (Life Technologies, Carlsbad, CA) and cells stably expressing antibodies were selected with G418 (800 µg/ml) and Hygromycin (160 µg/ml). The nucleotide sequence of MABs was determined using a Big Dye Terminator, version 1.1 (Applied Biosystems, Life Technologies, Carlsbad, CA) and the genetic analyzer A&B 3500/3500xL (Applied Biosystems, Life Technologies, Carlsbad, CA). Sequences were aligned and analyzed using the CLC Sequence viewer 6 (CLCbio, Boston, MA). Evolutionary analyses were conducted in MEGA5 (Tamura et al., 2011).

Binding activity of MABs to HIV-1 Env by ELISA analysis

Capture ELISA described in the previous section and Flow cytometry were used to determine the binding of the MABs to

monomeric and trimeric gp120, respectively. The effect of sCD4 (2 µg/ml) over binding to gp120 and binding to V3 peptide allowed classifying the MABs into four groups: V3, CD4bs, CD4i and “other epitopes” MABs.

Reactivity of V3 MABs against a panel of known V3 epitopes was performed using a peptide-based ELISA as described elsewhere (Eda et al., 2006b). Briefly, synthetic V3 peptides in PBS (1 µg/ml) were pre-coated in polyvinyl chloride flexible 96 well plates (BD Falcon, Franklin Lakes, NJ) and incubated overnight at 4 °C; later plates were blocked with 2% bovine serum albumin-0.1% Tween 20 in PBS. V3 MABs (1 µg/ml) were added and incubated for 1 h at room temperature. IgG bound to V3 peptides was detected with alkaline phosphatase-conjugated goat anti-human IgG (Sigma, St. Luis MO) followed by addition of phosphatase substrate (Sigma, St. Luis MO). A_{405} measurements were taken using a microplate reader (Biorad, Hercules, CA).

Binding activity of MABs to HIV-1 Env by flow cytometry analysis

The ability of MABs to bind virus-infected cells was analyzed by flow cytometric analysis. Briefly, PM1/CCR5 cells were chronically infected with HIV-1_{JR-FL} as previously described (Yoshimura et al., 2006). Infected and uninfected cells were washed with PBS and adjusted to 5×10^6 cells/ml. For cell surface staining, 100 µl cells in PBS containing 0.2% BSA were incubated with 50 µl of 5 µg/ml MABs for 60 min at room temperature (RT). After washing with PBS containing 0.2% BSA, cells were incubated with 50 µl of allophycocyanin (APC)-conjugated AffiniPure F(ab')₂ Fragment Goat Anti-Human IgG (H+L) (Jackson ImmunoResearch, West Grove, PA) for 30 min at RT. Cells were fixed with PBS containing 10% formalin, and analyzed by FACSCalibur instrument (Becton Dickinson, Franklin Lakes, NJ). Data analysis was performed using FlowJo (TreeStar, San Carlos, CA).

Binding to Env from various HIV-1 strains was determined using the 293T cells transfected with plasmids that expressed both Env and enhanced green fluorescent protein (EGFP). The transfected cells were incubated with antibody (5 µg/ml) for 1 h at RT. The antibody binding was detected using APC-conjugated AffiniPure F(ab')₂ Fragment Goat Anti-Human IgG (H+L) and FACSCalibur as described above.

Determination of neutralization activity

Neutralization activity was measured with a modified version of an assay described previously (Montefiori, 2009). Briefly, serial dilutions of MABs and virus (400 Tissue Culture Infectious Dose [TCID]₅₀) were pre-incubated for 1 h. TZM-bl cells solution containing DEAE-dextran (25 µg/ml) were added and incubated for 48 h (37 °C and 5% CO₂), washed with PBS and lysed with lysis buffer (Galacto-star system, Life technologies, Carlsbad, CA). Lysate was transferred to an opaque plate containing β-galactosidase substrate (Galacto-star system, Life technologies, Carlsbad, CA) and incubated for 1 h. The β-galactosidase activity was measured in relative light units (RLU) using a Centro XS³ LB960 luminometer (Berthold technologies, Bad Wildbad, Germany). The reduction of infectivity was determined by comparing the RLU in the presence and absence of antibody and was expressed as a percentage of neutralization.

Antibody-dependent cell-mediated cytotoxicity (ADCC) assay

ADCC activity was addressed using the rapid fluorometric assessment of ADCC (RFADCC) previously reported elsewhere (Gómez-Román et al., 2006). Target cells were CEM.NKr.CCR5 cells which were chronically infected with different strains of HIV-1. The gp120 coated cells were prepared by incubating 5×10^6 cells

with 15 µg of gp120 for 1 h. Cells were washed twice in ice-cold medium and coating was confirmed by binding of VRC01 to Env using flow cytometry. The infected cells were prepared by infection 7 days prior to the assay by spinoculation (O'Doherty et al., 2000). Briefly, a mixture of 5×10^5 cells, viral inoculum and polybrene were centrifuged for 2 h at $1200 \times g$ at 25 °C. Then, cells were cultured in RPMI supplemented with 10% FCS and media was exchanged every 48 h. Envelope expression was confirmed by binding of VRC01 using flow cytometry. PBMC or NK enriched-PBMC were used as effector cells. The NK enriched-PBMC fraction was prepared by using the NK cell isolation kit (Miltenyi Biotec, Bergisch Gladbach, Germany). Target cells were stained with carboxyfluorescein succinimidyl ester (CFSE; (Invitrogen, Life Technologies, Carlsbad, CA) and PKH-26 (Sigma, St. Luis, MO) according to the manufacturer's instructions. Stained cells were re-suspended and 50 µl (5000 cells) were incubated with 100 µl of antibody for 1 h at room temperature. Different concentrations were tested for each antibody in order to determine the optimal concentration for each antibody-virus combination. Then, 50 µl of effector cells were added to reach an Effector:Target ratio of 50:1 for PBMC and 5:1 for NK-enriched PBMC. Plates were centrifuged for 3 min at $400 \times g$ and incubated for 6 h (37 °C and 5% CO₂). Finally, cells were washed with PBS, and fixed with PBS containing 10% formalin. Data was acquired with a FACSCalibur instrument and analyzed with FlowJo software. ADCC activity was calculated as follows: first PKH-26 positive cells were gated (target cells), later the percentage of death dead cells (i.e. that have lost the CFSE dye, CFSE-) was determined. In each experiment, the percentage of killing obtained from target and effector cells without antibody was denoted as “background killing” and was subtracted from all the samples. Representative dot plots can be found in the supplementary data section (Supplementary Figs. 3 and 4) Differences in the ADCC activity of the negative control (8D11) and the samples were evaluated using the Mann-Whitney *U* test (Wilcoxon rank-sum test) with a 95% confidence interval. Differences were considered significant if *p* was <0.05. The test was performed using the GraphPad Prism 6 software (GraphPad Software, La Jolla, CA).

Evaluation of MABs synergistic effect

We first tested the binding activity of MABs in the presence or absence of 0.5δ using a gp120 capture ELISA as described above. Briefly, plates covered with gp120 SF2 were incubated for 30 min with 50 µl of 0.5δ. Later, 50 µl of biotin-conjugated MABs at a concentration of 1 µg/ml were dispensed in each well. We also tested the combination effect of MABs over neutralization by using the neutralization assay described above. Briefly, serial dilutions of MABs and 0.5δ in 1:1 combination ratio and virus (400 TCID₅₀) were pre-incubated for 1 h. TZM-bl cells solution containing DEAE-dextran (25 µg/ml) were added and incubated for 48 h (37 °C and 5% CO₂), washed with PBS and lysed with lysis buffer (Galacto-star system, Life technologies, Carlsbad, CA). Lysate was transferred to an opaque plate containing β-galactosidase substrate (Galacto-star system, Life technologies, Carlsbad, CA) and incubated for 1 h. The β-galactosidase activity was measured in relative light units (RLU) using a Centro XS³ LB960 luminometer (Berthold technologies, Bad Wildbad, Germany). The reduction of infectivity was determined by comparing the RLU in the presence and absence of antibody and was expressed as a percentage of neutralization. Combination and synergistic effect of 0.5γ with 0.5δ was analyzed by the Chou and Talaly (1977) method and 3 dimensional analyses. Using the CalcuSyn version 2 software, combination indices (CI) were calculated using the MABs Inhibitory Concentrations (IC) and interpreted as follows: CI < 0.9

synergy, $0.9 < CI < 1.1$ additivity and $CI > 1.1$ antagonism (Maeda et al., 2001; Yoshimura et al., 2006).

Competitive binding assay

Selected MAbs 0.5 γ (V3), 3E4 (V3), 0.5 δ (CD4bs) and 4C11 (CD4i) were conjugated with biotin and subjected for evaluation of epitope-competing antibodies in plasma samples from HIV-1 infected patients, using a gp120 capture ELISA as described above. Briefly, plates covered with gp120 SF2 were incubated for 30 min with 50 μ l of 3 fold serial dilution of plasma samples. Later, 50 μ l of biotin-conjugated MAbs at a concentration of 1 μ g/ml were dispensed in each well. The competition in binding of each biotin-conjugated MAb was detected by ALP-conjugated Avidin (Sigma, St. Louis MO) and substrate.

We evaluated the existence of epitope-competing antibodies in patients' plasma samples including 47 from ordinary progressors, 25 from slow-progressors, 20 from non-progressors, 11 from non-subtype B virus infection and 7 from seronegative donors in this competition assay using 1:50 dilution of the plasma. Plasma samples of non-progressors were collected from hemophilic patients in Japan who kept CD4⁺ cells counts higher than 350/ μ l without antiviral treatment at 2005, the time frame estimated for 23 years of HIV-infection. Plasma samples of slow progressors were collected from hemophilic patients in Japan who started antiviral treatment with CD4⁺ cell counts at 350–200/ μ l after 1997, the time estimated for more than 15 years of HIV-infection. Ordinary progressors are patients with subtype B infection who had initiated antiviral treatment with CD4⁺ cells counts less than 200/ μ l.

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Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at <http://dx.doi.org/10.1016/j.virol.2014.11.011>.

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