

ALERT SIGNS		QUICKLY LOOK FOR	
		Signs, measurements	Diagnosis
Sudden onset of multisystem* symptoms (*respiratory, skin and mucosal)	Reduced blood pressure		Anaphylactic shock
Inspiratory dyspnea Dysphonia Sialorrhea			Laryngeal edema
Painful skin Atypical target lesions Erosions of mucosa (≥ 2 mucous membranes)	Skin blisters, bullae Nikolsky sign Blood count (leucopenia, thrombopenia) Renal function (↑urea, creatinin)		SJS/TEN
Fever > 38.5°C Skin extension > 50% Centrofacial edema	Lymphadenopathia (≥ 2 sites) Blood count (eosinophilia, atypical lymphocytes) Liver function tests (↑liver transaminases) Proteinuria		HSS/DRESS/DIHS
Purpuric infiltrated papules Necrosis	Blood count (exclude thrombocytopenia) Renal function (proteinuria, ↑urea, creatinin) Hypocomplementemia		Vasculitis

**Figure 2** Clinical and biological danger signs suggesting severe cutaneous and/or systemic reactions (created using data from (62)).

### Diagnosis

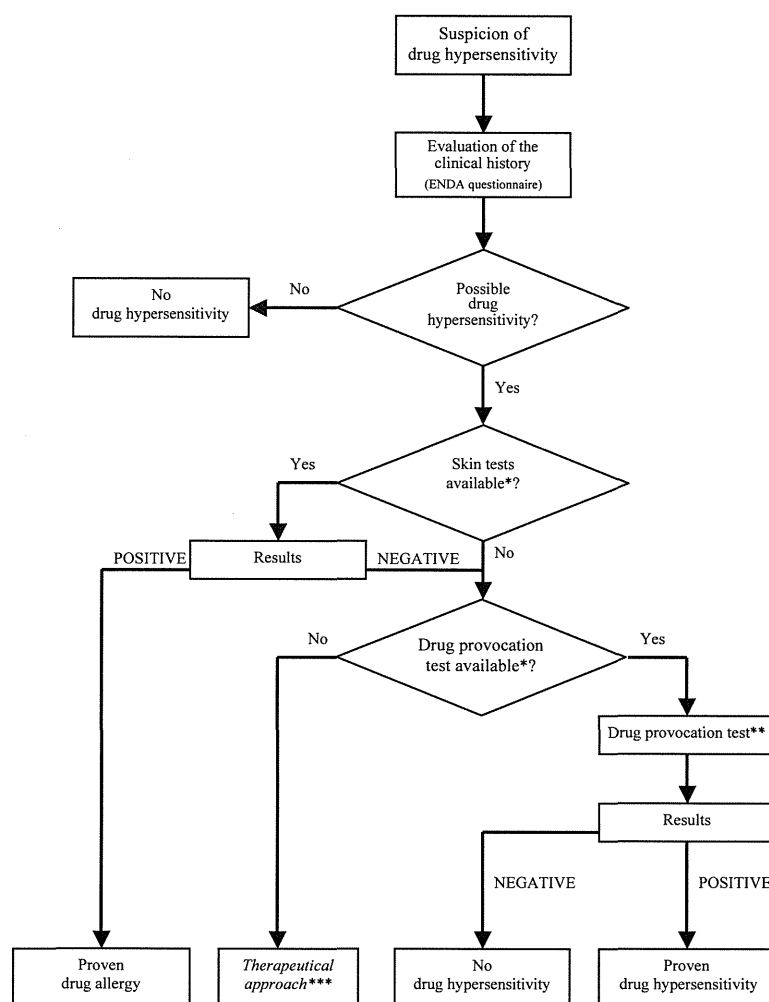
The diagnosis of DHRs requires knowledge of the scientific literature with access to Medline searches and to the Committee on Safety of Medicine and Embase Reports for the more recently introduced drugs. The lack of case studies involving a particular compound does not mean that it cannot induce a DHR, but for a widely used drug, it renders DHRs much less likely. The diagnosis is indeed based on history, on clinical manifestations, and if possible, on *in vivo* tests and some *in vitro* biological tests (Fig. 3) (78). However, only a few clinical and biological tools are available and fully validated. Moreover, a definitive diagnosis of such a reaction is preferred in order to institute proper preventive measures (Box 3).

#### Evaluation of the clinical history

Clinical history must be carefully obtained and should include the symptomatology (whether compatible with a DHR), the chronology of the symptoms (previous exposure, delay between the last dose and the onset of symptoms, effect of stopping treatment), other medications taken (both at the time of the reaction and other drugs of the same class taken since), and the medical background of the patient (any suggestion of a previous allergy, whether

associated with medication or not, or of a medical condition, such as chronic urticaria/chronic rhinosinusitis, that can be aggravated by the intake of certain drugs such as aspirin and noncyclooxygenase two selective NSAIDs). Data should ideally be recorded in a uniform format, and in order to harmonize the DHR diagnostic procedures, members of EAACI-DAIG/ENDA have developed a questionnaire (11) available in many different languages (Appendix S1 in the online Supporting Information). Diagnosis is more difficult when patients are not seen during the symptomatic phase, in which case photographs are helpful. When patients are seen during the reaction, the suspected drugs should be stopped after a benefit/risk balance analysis, especially if danger/severity signs are present (Fig. 2) (62).

A large number of reactions are presumed to be drug related and allergic in nature, but closer examination often reveals that they are not (3, 5). The history is often not reliable because different drugs are frequently taken simultaneously and each of them can account for the symptoms, although often with very different *a priori* probabilities. History can also be imprecise in many cases. Finally, the clinical picture of DHRs is very heterogeneous, mirroring many distinct pathophysiological events (Table 2). Thus, for the diagnosis of DHR, many healthcare professionals rely on history and various reference manuals. They do not attempt to prove the relationship between the drug intake and the symptoms or to clarify the underlying



**Figure 3** Flow chart when assessing DHRs (adapted from (78) with permission). \*Currently available biological tests to diagnose drug allergy lack sensitivity. \*\*In the absence of contraindications (Box 6).

\*\*\*If no alternative is available (e.g., NMBA, chemotherapeutic drugs), readministration of the drug is allowed under close surveillance, considering premedication and/or desensitization.

pathomechanism of the reaction. Such practice leads to a misunderstanding of the epidemiology and the pathophysiology of this highly relevant field. Members of the panel have listed situations in order to determine when to test and when not to test in suspected DHR (Boxes 4 and 5). An accurate diagnosis of DHRs allows implementation of the best measures required for prevention and treatment. For universal drugs such as  $\beta$ -lactams, NSAIDs, local anesthetics, simply avoiding the drug is not sufficient (Box 4). This procedure could lead to the contraindication of drugs which do not necessarily give rise to reactions and which are widely used. Besides, a false diagnosis can lead to a fake sense of security if other possible causes of serious reactions are not explored and excluded. However, this is a valid option until a specialist consultation can be scheduled.

**Box 3:** Key points regarding DHR diagnosis

- 1 A definitive diagnosis of a DHR is in many cases required in order to institute proper preventive measures.
- 2 Misclassification based on the DHR history alone may have consequences on individual treatment choices and be more detrimental for the patients than a complete drug allergy workup.
- 3 The clinical tools allowing a definitive diagnosis include a thorough clinical history, standardized skin tests, reliable *in vitro* tests, and drug provocation tests.
- 4 When properly performed in specialized centers, a reliable diagnosis is often possible and safe alternative medication can be administered.
- 5 Screening subjects without a prior history of allergic drug reactions is not recommended.

**Box 4:** DHR workup: When to evaluate?

- 1 When there is a history of prior DHR and the drug is required without an equally effective, structurally unrelated alternative, and if the risk/possible benefit ratio is positive:
  - a For the majority of patients with  $\beta$ -lactam, NSAIDs, local anesthetics DHRs.
  - b For others when drugs are required (depending on an individual medical needs).
- 2 When there is a history of prior severe DHR for other drugs (the best way to protect the patient is to find the culprit agents).

**Box 5:** DHR workup: When not to evaluate?

- 1 Cases with no drug allergy causality:
  - a Noncompatible symptomatology
  - b Noncompatible chronology
  - c Drug taken since with no reaction
  - d Reaction without having taken the drug
  - e Alternative diagnosis (e.g., herpesvirus eruption, chronic urticaria)
- 2 For drug provocation, every time the reaction was too severe: noncontrollable reaction and severe life-threatening reactions (Box 6)

The specific allergy workup should be carried out 4–6 weeks after the complete resolution of all clinical symptoms and signs (R2, Evidence D). How early testing can be made without results being falsely negative is unknown. On the other hand, after a time interval of more than 6–12 months, some drug tests may already have turned negative. These could be false-negative results (or true negative) depending on the results of the subsequent drug provocation test. According to the clinical presentations, a hypothesis on pathogenesis should be generated (Table 2) in order to select appropriate testing procedures (12, 62).

*Pharmacovigilance algorithms*

Pharmacovigilance algorithms for diagnosis are based principally on the clinical history (79); they are rarely specific for DHRs (80). They rarely produce a firm diagnosis of DHRs, and allergy testing is often necessary (79). Indeed, the symptoms are often suggestive, but not necessarily definitive in diagnosing DHR. The effect of discontinuation of the drug is not always conclusive (e.g., rebounds of urticaria after drug withdrawal is possible for a few hours) and no biological examination is reliable and specific. Often there is a lack of accurate information (imprecise chronology, exact name of drug or of corrective treatment not recalled by the patient), making drug causality assessment difficult to ascertain.

*Skin tests*

Skin tests are the most readily available means for confirming or excluding sensitization (22). Their diagnostic value has not

been fully evaluated for all drugs, and over the past decades, experience among different centers has rarely been exchanged in a systematic manner (22). These tests should follow standard procedures and should be performed by trained staff (6, 12). They should be performed 4–6 weeks after the reaction (R2, Evidence D). Skin tests have to be applied depending on the suspected pathomechanism of the DHR.

Skin prick tests and intradermal tests are particularly important for reactive haptens in order to demonstrate an IgE-dependent mechanism (62). Thus, for immediate DHRs, the prick test is recommended for initial screening due to its simplicity, rapidity, low cost, and high specificity. Intradermal tests (12) are undertaken when skin prick tests are negative. Compared to skin prick tests, they provide an enhanced sensitivity for drug-specific IgE (12). They should be performed with the intravenously injectable form of the drug whenever possible (22). Their sensitivity and predictive values vary, depending on the culprit drug and the clinical presentation. They appear to be 'good' for immediate DHRs to  $\beta$ -lactam antibiotics, NMBA, platin salts, and heparins, but moderate to low for most other drugs (R3, Evidence B) (22).

In order to demonstrate a T-cell-dependent mechanism for nonimmediate DHRs (manifesting by cutaneous symptoms such as a maculopapular exanthema occurring within hours after the last drug intake), patch tests and/or late-reading intradermal tests should be performed (15, 62). Unfortunately, apart from allergic reactions to several antibiotics and a few other drugs (81), for most drug allergens, standardized and validated test concentrations and vehicles have not been studied or are disputed in the literature. Sometimes the drug is not available in an adequately reactive form, generally because it is a metabolic derivative which is immunogenic and not the parent drug. In such cases, provocation tests are required to confirm the diagnosis. Available data have been summarized by EAACI-DAIG/ENDA experts (22).

Testing subjects without a prior history of an allergic drug reaction is not supported by available studies and therefore not recommended by any of the societies, in particular in preoperative settings (20).

While there is general agreement among guidelines on the importance of skin testing in the drug allergy workup, some discrepancies arise. The authors of the US Practice Parameters (6) consider that immediate DHRs to iodinated radiocontrast media (RCM) are all nonallergic (described as 'anaphylactoid') in nature and do not include skin testing in the management of a patient having experienced a previous DHR to iodinated RCM. This position is challenged by the multicenter study of EAACI-DAIG/ENDA (82), thus encouraging further studies (R4, Evidence C).

*Provocation tests*

A drug provocation test (DPT), also referred to as drug challenge, graded challenge, or test dosing, is the gold standard for the identification of the drug eliciting a DHR (R5, Evidence C). Whereas all the guidelines agree that the DPT

comes at the end of the stepwise approach in drug allergy (due to its inherent risks), it holds a slightly different meaning, depending on different guidelines. The authors of the US Practice Parameters (6) consider that the procedure is intended for patients who, after a full evaluation, are unlikely to be allergic to the given drug, that is, DPT performed to demonstrate tolerance to a less likely eliciting drug. The BSACI (8) guideline considers the primary aim of a DPT as a means to exclude DHR, but it can also be used to confirm a diagnosis. The EAACI-DAIG/ENDA guideline (13) addresses its role as a gold standard to establish or exclude the diagnosis of DHRs, but agrees that in some clinical practice situations, it might be more useful to look for safe alternatives instead of testing with a drug which was the definitive cause of the problem. It also mentions the altruistic and scientific value of the DPT (i.e., other patients might benefit from the obtained knowledge), but in these cases (and not in routine practice), approval by an ethical committee is mandatory.

The DPT is independent of the pathogenesis and consequently cannot differentiate between allergic from nonallergic DHRs. It takes individual factors such as the metabolism and genetic disposition of an individual into account. DPTs have the highest sensitivity, but should only be performed under the most rigorous surveillance conditions (Box 6). They are therefore usually restricted to certain specialist centers in which equipment, supplies, and personnel are present to manage serious reactions, and that personnel are well trained and experienced in performing this procedure in properly selected patients (13).

**Box 6:** Precautions and contraindications of performing DPTs

- 1** DPTs are contraindicated in noncontrollable and/or severe life-threatening DHRs:
  - a** Severe cutaneous reactions such as SJS, TEN, DRESS, vasculitis, AGEP
  - b** Systemic reactions such as DRESS, any internal organ involvement, hematological reactions
  - c** Anaphylaxis may be tested after risk/benefit analysis
- 2** DPTs are not indicated when:
  - a** The offending drug is unlikely to be needed and several structurally unrelated alternatives exist
  - b** Severe concurrent illness or pregnancy (unless the drug is essential for the concurrent illness or required during pregnancy or delivery)
- 3** DPTs should be performed under the highest safety conditions:
  - a** Trained staff: aware of the tests, ready to identify early signs of a positive reaction, and ready to manage a life-threatening reaction
  - b** With emergency resuscitative equipment available

These tests are particularly required for nonsteroidal anti-inflammatory drugs (23), local anesthetics, antibiotics other than  $\beta$ -lactams, and  $\beta$ -lactams when skin tests are negative. They should be performed after a certain time interval

following the DHR (at least 1 month) (R2, Evidence D) using, whenever possible, the same drug as in the initial reaction (13). Sometimes, when the clinical history has a favorable positive predictive value, performing DPT directly with an alternative drug seems more judicious (e.g., a cyclooxygenase-2 antagonist is typically tolerated uneventfully in the case of NSAID cross-reactors). Some authors evoke the option of prolonged DPTs (performed at home) in patients (children especially) with nonimmediate and nonsevere reactions (53, 83–85), sometimes without previous skin tests (53, 85). Recommendations have not yet echoed this strategy.

The route of administration depends on the suspected drug, which should in principle be administered in the same way as it was given when the initial reaction occurred. However, all the guidelines agree that the oral route is preferred whenever possible (R6, Evidence D). The precise challenge procedure varies a great deal from one team to another, and guidelines for the performance of DPTs have been proposed (13). A summary of DPT protocols has been reported in retrospective studies of more than one thousand consecutive patients (5, 85).

There is general consensus regarding the contraindications of DPT (see Box 6), with respect to the severity of the initial reaction and the availability of immediate treatment allowing complete and fast recovery (R7, Evidence D). The US Practice Parameters (6) state that rare exceptions to this may exist, such as treatment of a life-threatening illness, in which case the benefit of treatment outweighs the risk of a potentially life-threatening reaction. Arguments against a DPT would be if the offending drug is infrequently used and several alternatives exist. BSACI (8) and EAACI-DAIG/ENDA guidelines (13) mention that severe concurrent illness and pregnancy are generally considered as contraindications to DPT, unless the drug is essential for the concurrent illness (i.e., neurosyphilis and penicillin therapy, although desensitization may be considered first) or required during pregnancy or delivery (i.e., local anesthetics although it is not a classical DPT because subcutaneous injections are followed by a full dose of epidural anesthetic).

Despite the advantage of DPT over all the other test procedures, it has its limitations. First, the patient does not like to be re-exposed to a drug, which he or she considers harmful. Secondly, severe reactions are not amenable to DPTs (Box 6). Finally, a negative test does not prove tolerance to the drug in the future, but rather that there is no DHR at the time of the challenge and to the doses challenged. Nevertheless, a high negative predictive value (NPV) of  $\beta$ -lactam DPT of 94–98% was found in large studies involving both children and adults (86, 87), and most of the reactions reported by patients were both mild and nonimmediate reactions. Similarly, the NPV of DPT with NSAIDs also appears to be high (over 96%) whatever the NSAID (the one negatively tested or an alternative), and none of the false-negative patients described a life-threatening reaction (88). Desensitization by testing, as cause of false-negative DPT, is mentioned by the EAACI-DAIG/ENDA guideline (13) and the US Practice Parameters (6), but no reference to the existing literature is made. Resensitization by testing is addressed by EAACI-DAIG/ENDA (13) and BSACI (8) guidelines, with

respect to  $\beta$ -lactam allergy. Several studies have observed re-sensitization (i.e., a conversion to skin test positivity) after a negative DPT (followed by full therapeutic courses), with a frequency ranging from 0.9% (89) to 27.9% (90). Although this view is not mentioned in all guidelines and is not widely accepted, one approach might be to retest (2–4 weeks later) the patients who suffered severe immediate reactions and who displayed negative results at the first evaluation, which included a DPT (18) (R8, Evidence D).

### Biological tests

It would be highly advantageous to have discriminating biological tests available in order to establish the nature of the culprit agent. This would be helpful particularly for the patient receiving several drugs simultaneously and for severe life-threatening DHRs when skin tests are negative or not possible, and DPT contraindicated (Box 6). However, with some exceptions (e.g., major and minor determinants of penicillin G), the currently available biological methods to diagnose drug allergy lack sensitivity, although they are normally considered to be quite specific (>90%). There are no established methods to predict the immunogenic potential of a drug. It should also be remembered that the results need to be interpreted with caution. A negative test does not exclude the imputability of the drug, while a positive result shows a sensitivity to the drug, but does not reliably confirm its causality (R9, Evidence C).

*In vitro* assay for drug-specific IgE is not available for many allergenic drugs and, conversely, is offered for many drugs without evidence of validated assays. The demonstration of isolated drug-specific IgE (to penicillins (91), NMBA (92), chymopapain, or tetanus toxoid, for example) does not establish the diagnosis of a drug allergy. However, in conjunction with clinical findings (e.g., typical severe symptoms of rapid onset), an IgE-dependent mechanism can be assumed (particularly if the skin tests to the drug are also positive) (18, 91). Thus, EAACI-DAIG/ENDA advises that skin tests to antibiotics should be performed after IgE testing in severe immediate reactions (22). *In vitro* cross-reactivities between several drugs using quantitative inhibition may also be explored, knowing that its predicted clinical outcome is not fully validated (93). The absence of drug-specific circulating IgE does not rule out a diagnosis of immediate drug allergy (R9, Evidence C). Measurement of drug-specific IgM or IgG is of interest only in cases of drug-induced cytopenia, type III DHRs to vaccines or allergies to dextrans. However, the sensitivity of these tests is unknown and they are not widely available. *In vitro* histamine release from whole blood in the presence of the drug correlates well with skin tests and specific IgE for NMBA, but is not reliable for many other drugs (94). Moreover, it is costly and requires a high level of technical expertise. The usefulness of measuring sulfidopeptide leukotrienes produced *in vitro* by isolated peripheral blood leukocytes after allergenic drug stimulation still requires further validation in both IgE-dependent allergies and non-IgE-dependent DHRs (95). In cases of acute clinical reactions, blood measurements of

histamine or tryptase may confirm an involvement of basophils and mast cells whatever the cause of the degranulation (20, 96). Although tests for histamine are not widely commercially available, the test for tryptase is CAP FEIA (20). Basophil activation tests with flow cytometric reading hold promise and are currently undergoing validation for certain drugs (97–99).

For drug-induced type II and III allergic reactions, the following tests can be performed in some centers: Coombs' test, *in vitro* hemolysis test, determination of complement factors and circulating immune complexes. Assays involving T cells (lymphocyte transformation/activation tests) remain the domain of only a few laboratories with experience in DHRs, whereas results from commercial laboratories are generally not reliable (100). Searching for genetic markers may prove helpful, as several strong genetic associations between the expression of a particular HLA allele and the susceptibility to specific forms of DHRs have been recently discovered (Table 3) (50). For the drug abacavir, an association between B\*5701 expression and DRESS has prompted the development of predictive testing strategies (47) and labeling changes to drug information sheets. The same is now true for the drug carbamazepine in Han Chinese and the allele B\*1502 (101). The positive predictive value of the polymorphisms found so far varies widely (Table 3) and may not always lead to the simple and very successful predictive strategy of abacavir and B\*5701 (R10, Evidence A).

### Principles of drug allergy management

#### *Acute drug reactions*

Anaphylaxis must be treated promptly and appropriately (8), (102, 103), and all suspected drugs must be stopped (102, 104).

When patients experiencing nonanaphylactic reactions are examined during a reaction, the suspected drugs should be stopped if the risks of continuing the administration of the drug outweigh the benefits, and always if danger/severity signs are present (Fig. 2) (62). Indeed, during the acute phase of a severe delayed DHR, the putative drug as well as all 'less necessary' medication should be stopped with no delay in order to improve the prognosis (105).

Supportive treatment for delayed DHRs is not specifically covered by current drug allergy guidelines, but can be found in general reviews (58, 102, 106).

#### *Individual preventive measures*

A definitive diagnosis of DHRs allows more targeted preventive measures. Whatever the intensity of the clinical reaction, a state of hypersensitivity is shown toward the particular drug, with the possibility of a more serious reaction in the future. Individual measures include the issue of a written documentation specifying the culprit agent(s), the insertion of the allergy in the tab of the electronic medical record, the drawing up of a list of drugs to avoid, as well as a list of possible alternatives. The lists are only indicative and should be frequently updated (R11, Evidence D). The search for alternatives may require DPTs in a hospital setting when the alternatives belong to the same drug class (R12, Evidence C). The questioning (to elicit

any history of drug allergy) of every patient by every clinician prior to issuing a prescription is essential from both a medical and a medico-legal point of view (R13, Evidence D). The patient is also asked to make his 'allergies' known prior to all prescriptions and surgical operations.

Preventive measures by premedication (e.g., slow injection and pretreatment with glucocorticosteroids and H1-antihistamines) are useful mainly for nonallergic DHRs (for example to vancomycin, some NMBA, iodinated RCM, and chemotherapy drugs) (R14, Evidence C). Corticosteroids and H1-antihistamines may not reliably prevent IgE-dependent anaphylaxis (103).

#### *Desensitization*

Drug desensitization is defined as the induction of a temporary state of clinical unresponsiveness/tolerance to a compound responsible for a DHR (6, 19). Several other terms have been utilized in the past. To encompass classic IgE- and non-IgE-mediated drug desensitization, the Practice Parameters (6) introduced the term 'induction of drug tolerance'. Except for aspirin, the BSACI guidelines only propose desensitization related to an IgE-mediated mechanism (8).

The possibility of desensitization should always be considered when the offending drug is essential and when either no alternatives exist or they are unsatisfactory, as in the following cases (6, 19): sulfonamides in HIV-infected patients (107), quinolone allergies in some patients with cystic fibrosis, serious infections with allergy to  $\beta$ -lactams, antituberculosis drugs, allergy to tetanus vaccine, hemochromatosis with allergy to desferoxamine, taxanes, and platinum salt-based cancer chemotherapeutic agents (108), monoclonal antibodies utilized in several types of hematological and nonhematological neoplasms, aspirin and NSAID hypersensitivity in patients for whom the necessity for these drugs to treat either a cardiac (109) or rheumatic disease is clear.

There are no generally accepted protocols for drug desensitization in immediate DHRs, and guidelines (19) recommend referral to successfully applied existing protocols (R15, Evidence C). For nonimmediate DHRs, the literature is less extensive and more controversial. For EAACI-DAIG/ENDA experts, desensitization in delayed DHRs has to be restricted to uncomplicated exanthemas or fixed drug eruption, due to the unpredictability and limited therapeutic options in severe DHRs (110). Desensitization to aspirin, as a therapeutic intervention for aspirin-exacerbated respiratory disease or nasal polyps, is briefly mentioned by EAACI-DAIG/ENDA guidelines (19), whereas it is recommended in properly selected asthmatic patients by the US Practice Parameter (6), based on certain published data (111) (R16, Evidence D).

#### *General preventive measures*

General preventive measures include a declaration to the Committee on Safety of Medicine Reports. The reporting of DHRs leads to public health inquiries and decisions. Some successful examples of proper reports are the rules concerning the use of penicillins during animal feeding, the with-

drawal from the market of glafenine, the reformulation of propofol to eliminate the need for Cremophor EL (castor oil) and its replacement with other lipids, and the warnings concerning abacavir, carbamazepine, and nevirapine.

#### **Unmet needs**

##### *Unmet clinical needs*

Drug hypersensitivity reactions have a significant impact on clinical practice, drug development, and healthcare expenditures. However, epidemiological studies or research to increase understanding and to develop diagnostic and predictive tests has been limited. Epidemiologic risk factors for DHRs are not well characterized and may be influenced by regional/national differences in drug prescriptions and by genetic markers. All drugs can induce DHRs, but the incidence and risk factors for individual drugs remain a major unmet need. As an example, the co-medication of diclofenac with antiulcer medications may present a novel potentiating factor (112), as could the use of over-the-counter pholcodine regarding NMBA-induced anaphylaxis (113). The development of a network to increase the population size from which data on DHRs can be captured would be a major advance. This approach would aim to overcome the major limitation of spontaneous reporting, that is, under-reporting or non-proven case reporting, by engaging with interested clinicians and involving them in the network.

Physicians do not always have the confidence to clarify a suspected reaction. When they do so, and refer the patients to specialized centers, each one of them experiences a limited and partly biased spectrum of the disease (114). Although standardized diagnostic procedures have been published, validation of these clinical tests for all drugs does not exist and multicenter multinational studies are needed for this purpose. Current controversies and disagreements between the guidelines need to be addressed by further research (e.g., skin testing for iodinated RCM, NPV for penicillin skin testing, utility of skin testing for a variety of rare DHRs (steroids, preservatives, etc), and desensitization for delayed DHRs). Standardized diagnostic procedures should be tailored to specific drugs (e.g.,  $\beta$ -lactam antibiotics, non- $\beta$ -lactam antibiotics, NSAIDs, local anesthetics, radiocontrast media, chemotherapeutic agents, vaccines, biological agents), specific manifestations, and specific age groups (children vs adults). New diagnostic tools should be developed, in particular for the diagnosis of severe cutaneous DHRs, or DHRs those affecting internal organs including the liver, lungs, kidneys, and bone marrow. The development of tools for skin testing and biological diagnosis is indeed crucial for those cases where DPT is not possible. Standardized and widely accepted drug allergy procedures are crucial for both individual patient genotyping-phenotyping and epidemiological studies. There should be education in medical schools and residencies as well as postgraduate training programs that include aspects of DHR and its treatment, as well as funding for the postgraduate education of specialists.

The impact of DHRs on the quality of life of patients and their cost on the healthcare system, probably substantial, is

unknown. For this, one must take into account not only the direct costs (treatment of these reactions, hospitalizations, and prolongation of hospitalization), but also the indirect costs (sick leave, invalidity, excessive cost of the choice of alternatives which are not always medically satisfactory and which may lead to specific adverse effects including the induction of microbial resistance and reduced efficacy).

Additionally, most therapeutic recommendations, including new approaches such as drug desensitization, are mostly based on case reports or small case series. As we do not know the natural course of DHRs, it is not clear whether lifelong avoidance is really necessary. Specific research dedicated to the treatment for anaphylaxis should also be supported. DHR research has not been supported for a long time neither by the pharmaceutical industry nor by national programs. There is therefore a clear need for training, standardized criteria, and large, multicenter studies. The establishment of multinational, adequately resourced large DHR databases/registries would enable all observations to be collected, which would in turn facilitate epidemiologic, risk factor, pharmacovigilance, and research analyses.

#### *Unmet basic research needs*

The availability of tissue and serum samples from DHR patients is a prerequisite for basic research in the mechanism of DHRs, which may be allergic or nonallergic, with immunological or pharmacological recognition and with the allergenic and genetic determinants mostly unknown.

Evidence over the past ten years suggests that not all drugs need to bind covalently to the MCH in order to induce an immune response. Without undergoing the classical antigen processing and presentation pathway, some drugs may bind directly in a noncovalent fashion to immune receptors, triggering a drug-specific immune reaction (the p-i concept) and promoting an exchange of embedded peptides (38). The functional consequence of this peptide exchange should be further analyzed. This may explain the increased susceptibility of some patients and the frequency of non-IgE-mediated reactions that occur within hours of first exposure. Whether or not this mechanism is also involved in IgE-dependent reactions is not yet known. The prediction of such reactions may also be possible, but has not yet been fully evaluated. The importance here lies in future drug development, the prediction of which molecules may participate in such reactions, and the development of congeners which retain pharmacological activity, but do not cause immune reactions. For most drugs, the allergenic determinants are unknown. The lack of complete understanding of DHR mechanisms probably explains the low sensitivity of many skin tests and *in vitro* assays. There are many examples where existing tests are negative, and this is likely to be related to the use of an inappropriate antigen. Pinpointing the allergenic determinants is of crucial importance; this will allow a better prediction of cross-reactivities and will provide clinicians with tools for skin testing, biomarkers, and biological diagnosis. A better understanding of virus–drug interactions is also crucial. The availability and use of appropriate viral tests

are a prerequisite for a proper evaluation of the role of viral infections in DHRs.

Genetic differences can affect individual responses to drugs by influencing the way in which the drug is processed or acts in the body. They may explain why some drugs induce an immune reaction in only a minority of individuals. Genetic variation in the activity of enzymes and carrier substances can be responsible for changes in the absorption, transport, metabolism, and excretion of drugs. Some genetic variants in (i) drug-metabolizing enzymes (pharmacogenetics) interfering with oxidation, conjugation, and hydrolysis (cytochrome P450, glucuronyl transferase, and glutathione S transferase), acetylation; (ii) drug receptors and effector proteins; and (iii) genes controlling the immune response, especially in the MCH molecules (immunogenetics), have been associated with some DHRs (Table 3). This is an emerging field, which holds a great deal of promise for the development of individual predictive tests. However, this will only be possible if we can pool resources to identify and characterize a large cohort of patients with standardized phenotypic definitions to design studies with adequate statistical power (61). This will only be possible through collaboration.

To generate preclinical testing methods to assess the risk of potential DHRs in new drugs, research should encompass the characterization of drug-specific (chemical structure, metabolites, exposure), intrinsic (genetics), and extrinsic (viral infections, other danger signals) risk factors, complemented by preclinical prediction models (2, 115).

#### **Conclusions**

The diagnosis of DHRs is often challenging and requires the same careful approach, no matter which specific drug is involved. It remains largely clinical with the help of certain allergy tests that are available for some of the drug classes. Provocation tests are the gold standard for determining current tolerance, but require expertise, carry a certain amount of risk, and are limited to highly specialized centers when used to establish or rule out diagnosis. They cannot be applied for severe cutaneous reactions. New and validated biological tests for diagnosis, available to all clinicians, are necessary in order to improve care for these patients. Recently, HLA typing has provided an important tool for detecting susceptible patient populations. In view of the diagnostic uncertainty of most adverse drug reaction studies (1), the epidemiology of DHRs was not covered in this ICON document. However, understanding the epidemiology of adverse drug reactions in general and DHRs in particular remains an important future research priority. Finally, collaborative basic research into the pathophysiology of DHRs should be intensified in order to better understand this complex set of diseases associated with or induced by drug exposures and mediated (or not) by the immune system.

#### **Conflicts of interest**

The authors declare no conflicts of interest for this work.

## Supporting Information

Additional Supporting Information may be found in the online version of this article:

## Appendix S1. Drug hypersensitivity questionnaire (adapted from (11) with permission).

## References

- Gomes ER, Demoly P. Epidemiology of hypersensitivity drug reactions. *Curr Opin Allergy Clin Immunol* 2005;**5**:309–316.
- Demoly P, Pichler W, Pirmohamed M, Romano A. Important questions in Allergy 1–drug allergy/hypersensitivity. *Allergy* 2008;**63**:616–619.
- Gomes E, Cardoso MF, Praca F, Gomes L, Marino E, Demoly P. Self-reported drug allergy in a general adult Portuguese population. *Clin Exp Allergy* 2004;**34**:1597–1601.
- Mittmann N, Knowles SR, Gomez M, Fish JS, Cartotto R, Shear NH. Evaluation of the extent of under-reporting of serious adverse drug reactions: the case of toxic epidermal necrolysis. *Drug Saf* 2004;**27**:477–487.
- Messaad D, Sahla H, Benahmed S, Godard P, Bousquet J, Demoly P. Drug provocation tests in patients with a history suggesting an immediate drug hypersensitivity reaction. *Ann Intern Med* 2004;**140**:1001–1006.
- Joint Task Force on Practice Parameters; American Academy of Allergy AaIAcCoA, Asthma and Immunology; Joint Council of Allergy, Asthma and Immunology. Drug allergy: an updated practice parameter. *Ann Allergy Asthma Immunol* 2010;**105**:259–273.
- Kelso JM, Greenhawt MJ, Li JT, Nicklas RA, Bernstein DI, Blessing-Moore J et al. Adverse reactions to vaccines practice parameter 2012 update. *J Allergy Clin Immunol* 2012;**130**:25–43.
- Mirakian R, Ewan PW, Durham SR, Youlten LJ, Dugue P, Friedmann PS et al. BSACI guidelines for the management of drug allergy. *Clin Exp Allergy* 2009;**39**:43–61.
- Ewan PW, Dugue P, Mirakian R, Dixon TA, Harper JN, Nasser SM. BSACI guidelines for the investigation of suspected anaphylaxis during general anaesthesia. *Clin Exp Allergy* 2010;**40**:15–31.
- Przybilla B, Aberer W, Bircher AJ, Brehler R, Brockow K, Dickel H et al. Allergological approach to drug hypersensitivity reactions. *J Dtsch Dermatol Ges* 2008;**6**:240–243.
- Demoly P, Kropf R, Bircher A, Pichler WJ. Drug hypersensitivity: questionnaire. EAACI interest group on drug hypersensitivity. *Allergy* 1999;**54**:999–1003.
- Brockow K, Romano A, Blanca M, Ring J, Pichler W, Demoly P. General considerations for skin test procedures in the diagnosis of drug hypersensitivity. *Allergy* 2002;**57**:45–51.
- Aberer W, Bircher A, Romano A, Blanca M, Campi P, Fernandez J et al. Drug provocation testing in the diagnosis of drug hypersensitivity reactions: general considerations. *Allergy* 2003;**58**:854–863.
- Torres MJ, Blanca M, Fernandez J, Romano A, Weck A, Aberer W et al. Diagnosis of immediate allergic reactions to beta-lactam antibiotics. *Allergy* 2003;**58**:961–972.
- Romano A, Blanca M, Torres MJ, Bircher A, Aberer W, Brockow K et al. Diagnosis of nonimmediate reactions to beta-lactam antibiotics. *Allergy* 2004;**59**:1153–1160.
- Brockow K, Christiansen C, Kanny G, Clement O, Barbaud A, Bircher A et al. Management of hypersensitivity reactions to iodinated contrast media. *Allergy* 2005;**60**:150–158.
- Bousquet PJ, Demoly P, Romano A, Aberer W, Bircher A, Blanca M et al. Pharmacovigilance of drug allergy and hypersensitivity using the ENDA-DAHD database and the GALEN platform. The Galenda project. *Allergy* 2009;**64**:194–203.
- Blanca M, Romano A, Torres MJ, Fernandez J, Mayorga C, Rodriguez J et al. Update on the evaluation of hypersensitivity reactions to betalactams. *Allergy* 2009;**64**:183–193.
- Cernadas JR, Brockow K, Romano A, Aberer W, Torres MJ, Bircher A et al. General considerations on rapid desensitization for drug hypersensitivity - a consensus statement. *Allergy* 2010;**65**:1357–1366.
- Mertes PM, Malinovsky JM, Jouffroy L, Aberer W, Terreehorst I, Brockow K et al. Reducing the risk of anaphylaxis during anesthesia: 2011 updated guidelines for clinical practice. *J Investig Allergol Clin Immunol* 2011;**21**:442–453.
- Kowalski ML, Makowska JS, Blanca M, Bavbek S, Bochenek G, Bousquet J et al. Hypersensitivity to nonsteroidal anti-inflammatory drugs (NSAIDs) - classification, diagnosis and management: review of the EAACI/ENDA(®) and GA2LEN/HANNA\*. *Allergy* 2011;**66**:818–829.
- Brockow K, Garvey LH, Aberer W, Atanaskovic-Markovic M, Barbaud A, Bilo MB et al. Skin test concentrations for systemically administered drugs - an ENDA/EAACI Drug Allergy Interest Group position paper. *Allergy* 2013;**68**:702–712.
- Nizankowska-Mogilnicka E, Bochenek G, Mastalerz L, Swierczynska M, Picado C, Scadding G et al. EAACI/GA2LEN guideline: aspirin provocation tests for diagnosis of aspirin hypersensitivity. *Allergy* 2007;**62**:1111–1118.
- Thong B, Motala C, Vervloet D. Disease summaries - Drug allergies. Available from: [http://www.worldallergy.org/professional/allergic\\_diseases\\_center/drugallergy/](http://www.worldallergy.org/professional/allergic_diseases_center/drugallergy/). Last accessed 12 December 2013.
- Pichler WJ, Thong B. GLORIA module 11. Drug allergy. [cited 9 May 2012]; Available from: [http://www.worldallergy.org/educational\\_programs/gloria/international/materials.php](http://www.worldallergy.org/educational_programs/gloria/international/materials.php).
- Lotvall J, Pawankar R, Wallace DV, Akdis CA, Rosenwasser LJ, Weber RW et al. We call for iCAALL: International Collaboration in Asthma, Allergy and Immunology. *Allergy* 2012;**67**:449–450.
- Papadopoulos NG, Arakawa H, Carlsen KH, Custovic A, Gern J, Lemanske R et al. International consensus on (ICON) pediatric asthma. *Allergy* 2012;**67**:976–997.
- Harbour R, Miller J. A new system for grading recommendations in evidence based guidelines. *BMJ* 2001;**323**:334–336.
- Johansson SG, Bieber T, Dahl R, Friedmann PS, Lanier BQ, Lockey RF et al. Revised nomenclature for allergy for global use: Report of the Nomenclature Review Committee of the World Allergy Organization, October 2003. *J Allergy Clin Immunol* 2004;**113**:832–836.
- WHO. International drug monitoring: the role of national centres. Report of a WHO meeting. *Tech Rep Ser WHO* 1972;**498**:1–25.
- Davies DM, Ashton CH, Rao JG, Rawlins MD, Routledge PA, Savage RL et al. Comprehensive clinical drug information service: first year's experience. *Br Med J* 1977;**1**:89–90.
- Bircher AJ, Scherer Hofmeier K. Drug hypersensitivity reactions: Inconsistency in the use of the classification of immediate and nonimmediate reactions. *J Allergy Clin Immunol* 2012;**129**:263–264; author reply 265–266.
- Pichler WJ. Delayed drug hypersensitivity reactions. *Ann Intern Med* 2003;**139**:683–693.
- Park BK, Naisbitt DJ, Demoly P. Drug hypersensitivity. In: Holgate S, Church M,



- Broide D, Martinez F, editors. *Allergy*. New York: Elsevier Ltd, 2012: 321–330.
35. O'Connor N, Dargan PI, Jones AL. Hepatocellular damage from non-steroidal anti-inflammatory drugs. *QJM* 2003;**96**:787–791.
  36. Gallucci S, Matzinger P. Danger signals: SOS to the immune system. *Curr Opin Immunol* 2001;**13**:114–119.
  37. Chan KK, Vyas KH, Brandt KD. In vitro protein binding of diclofenac sodium in plasma and synovial fluid. *J Pharm Sci* 1987;**76**:105–108.
  38. Ostrov DA, Grant BJ, Pompeu YA, Sidney J, Harndahl M, Southwood S et al. Drug hypersensitivity caused by alteration of the MHC-presented self-peptide repertoire. *Proc Natl Acad Sci USA* 2012;**109**:9959–9964.
  39. Chung WH, Hung SI, Hong HS, Hsieh MS, Yang LC, Ho HC et al. Medical genetics: a marker for Stevens-Johnson syndrome. *Nature* 2004;**428**:486.
  40. Mehta TY, Prajapati LM, Mittal B, Joshi CG, Sheth JJ, Patel DB et al. Association of HLA-B\*1502 allele and carbamazepine-induced Stevens-Johnson syndrome among Indians. *Indian J Dermatol Venereol Leprol* 2009;**75**:579–582.
  41. Lochareonkul C, Loplumlert J, Limotai C, Korkij W, Desudchit T, Tongkobpetch S et al. Carbamazepine and phenytoin induced Stevens-Johnson syndrome is associated with HLA-B\*1502 allele in Thai population. *Epilepsia* 2008;**49**:2087–2091.
  42. McCormack M, Alfirevic A, Bourgeois S, Farrell JJ, Kasperaviciute D, Carrington M et al. HLA-A\*3101 and carbamazepine-induced hypersensitivity reactions in Europeans. *N Engl J Med* 2011;**364**:1134–1143.
  43. Ozeki T, Mushiroda T, Yowang A, Takahashi A, Kubo M, Shirakata Y et al. Genome-wide association study identifies HLA-A\*3101 allele as a genetic risk factor for carbamazepine-induced cutaneous adverse drug reactions in Japanese population. *Hum Mol Genet* 2011;**20**:1034–1041.
  44. Lonjou C, Thomas L, Borot N, Ledger N, de Toma C, LeLouet H et al. A marker for Stevens-Johnson syndrome.: ethnicity matters. *Pharmacogenomics J* 2006;**6**:265–268.
  45. Hung SI, Chung WH, Jee SH, Chen WC, Chang YT, Lee WR et al. Genetic susceptibility to carbamazepine-induced cutaneous adverse drug reactions. *Pharmacogenomics* 2006;**16**:297–306.
  46. Alfirevic A, Jorgensen AL, Williamson PR, Chadwick DW, Park BK, Pirmohamed M. HLA-B locus in Caucasian patients with carbamazepine hypersensitivity. *Pharmacogenomics* 2006;**7**:813–818.
  47. Mallal S, Phillips E, Carosi G, Molina JM, Workman C, Tomazic J et al. HLA-B\*5701 screening for hypersensitivity to abacavir. *N Engl J Med* 2008;**358**:568–579.
  48. Mallal S, Nolan D, Witt C, Masel G, Martin AM, Moore C et al. Association between presence of HLA-B\*5701, HLA-DR7, and HLA-DQ3 and hypersensitivity to HIV-1 reverse-transcriptase inhibitor abacavir. *Lancet* 2002;**359**:727–732.
  49. Stekler J, Maenza J, Stevens C, Holte S, Malhotra U, McElrath MJ et al. Abacavir hypersensitivity reaction in primary HIV infection. *AIDS* 2006;**20**:1269–1274.
  50. Pavlos R, Mallal S, Phillips E. HLA and pharmacogenetics of drug hypersensitivity. *Pharmacogenomics* 2012;**13**:1285–1306.
  51. Guglielmi L, Fontaine C, Gougat C, Avinens O, Eliaou JF, Guglielmi P et al. IL-10 promoter and IL4-Ralpha gene SNPs are associated with immediate beta-lactam allergy in atopic women. *Allergy* 2006;**61**:921–927.
  52. Barbaud A, Waton J, Herbeth B, Bursztejn AC, Bollaert M, Schmutz JL et al. Comparison of cytokine gene polymorphism in drug-induced maculopapular eruption, urticaria and drug reaction with eosinophilia and systemic symptoms (DRESS). *J Eur Acad Dermatol Venereol* 2014, doi: 10.1111/jdv.12130.
  53. Caubet JC, Kaiser L, Lemaitre B, Fellay B, Gervais A, Eigenmann PA. The role of penicillin in benign skin rashes in childhood: a prospective study based on drug rechallenge. *J Allergy Clin Immunol* 2011;**127**:218–222.
  54. Webster AW, Thompson RA. The ampicillin rash. Lymphocyte transformation by ampicillin polymer. *Clin Exp Immunol* 1974;**18**:553–564.
  55. Camous X, Calbo S, Picard D, Musette P. Drug Reaction with Eosinophilia and Systemic Symptoms: an update on pathogenesis. *Curr Opin Immunol* 2012;**24**:730–735.
  56. Descamps D, Collin G, Letourneur F, Apretre C, Damond F, Lousert-Ajaka I et al. Susceptibility of human immunodeficiency virus type 1 group O isolates to antiretroviral agents: in vitro phenotypic and genotypic analyses. *J Virol* 1997;**71**:8893–8898.
  57. Mardivirin L, Valeyrie-Allanore L, Brabant-Redon E, Beneton N, Jidar K, Barbaud A et al. Amoxicillin-induced flare in patients with DRESS (Drug Reaction with Eosinophilia and Systemic Symptoms): report of seven cases and demonstration of a direct effect of amoxicillin on Human Herpesvirus 6 replication in vitro. *Eur J Dermatol* 2010;**20**:68–73.
  58. Limsuwan T, Demoly P. Acute symptoms of drug hypersensitivity (urticaria, angioedema, anaphylaxis, anaphylactic shock). *Med Clin North Am* 2010;**94**:691–710.
  59. Mockenhaupt M. Severe drug-induced skin reactions: clinical pattern, diagnostics and therapy. *J Dtsch Dermatol Ges* 2009;**7**:142–160.
  60. Bircher AJ, Scherer K. Delayed cutaneous manifestations of drug hypersensitivity. *Med Clin North Am* 2010;**94**:711–725.
  61. Pirmohamed M, Friedmann PS, Molokhia M, Loke YK, Smith C, Phillips E et al. Phenotype standardization for immune-mediated drug-induced skin injury. *Clin Pharmacol Ther* 2011;**89**:896–901.
  62. Bircher AJ. Symptoms and danger signs in acute drug hypersensitivity. *Toxicology* 2005;**209**:201–207.
  63. Chiriac AM, Demoly P. Multiple drug hypersensitivity syndrome. *Curr Opin Allergy Clin Immunol* 2013;**13**:323–329.
  64. Sullivan T, Remedios C, Ong M, Gilliam L. Studies of the multiple drug allergy syndrome. *J Allergy Clin Immunol* 1989;**83**:270.
  65. Pichler WJ, Daubner B, Kawabata T. Drug hypersensitivity: flare-up reactions, cross-reactivity and multiple drug hypersensitivity. *J Dermatol* 2011;**38**:216–221.
  66. Patriarca G, Venuti A, Schiavino D, Romano A, Fais G, Di Rienzo V. The syndrome caused by multiple drug intolerance. *Recenti Prog Med* 1980;**68**:21–33.
  67. Schiavino D, Nucera E, Roncallo C, Pollastrini E, De Pasquale T, Lombardo C et al. Multiple-drug intolerance syndrome: clinical findings and usefulness of challenge tests. *Ann Allergy Asthma Immunol* 2007;**99**:136–142.
  68. Macy E, Ho NJ. Multiple drug intolerance syndrome: prevalence, clinical characteristics, and management. *Ann Allergy Asthma Immunol* 2012;**108**:88–93.
  69. Neukomm CB, Yawalkar N, Helbling A, Pichler WJ. T-cell reactions to drugs in distinct clinical manifestations of drug allergy. *J Investig Allergol Clin Immunol* 2001;**11**:275–284.
  70. Hari Y, Frutig-Schnyder K, Hurni M, Yawalkar N, Zanni MP, Schnyder B et al. T cell involvement in cutaneous drug eruptions. *Clin Exp Allergy* 2001;**31**:1398–1408.
  71. Gex-Collet C, Helbling A, Pichler WJ. Multiple drug hypersensitivity—proof of multiple drug hypersensitivity by patch and lymphocyte transformation tests. *J Investig Allergol Clin Immunol* 2005;**15**:293–296.
  72. Aihara Y, Ito S, Aihara M, Kobayashi Y, Yokota S. Different patterns of cytokines, ECP and immunoglobulin profiles at two adverse drug reactions in a patient. *Pediatr Int* 2005;**47**:616–621.
  73. Daubner B, Groux-Keller M, Hausmann OV, Kawabata T, Naisbitt DJ, Park BK et al. Multiple drug hypersensitivity: normal Treg cell function but enhanced in vivo activation of drug-specific T cells. *Allergy* 2012;**67**:58–66.

74. Blanca M, Torres MJ, Garcia JJ, Romano A, Mayorga C, de Ramon E et al. Natural evolution of skin test sensitivity in patients allergic to beta-lactam antibiotics. *J Allergy Clin Immunol* 1999;**103**:918–924.
75. Guttormsen AB, Johansson SG, Oman H, Wilhelmssen V, Nopp A. No consumption of IgE antibody in serum during allergic drug anaphylaxis. *Allergy* 2007;**62**:1326–1330.
76. Barbaud A, Collet E, Milpied B, Assier H, Staumont D, Avenel-Audran M et al. A multicentre study to determine the value and safety of drug patch tests for the three main classes of severe cutaneous adverse drug reactions. *Br J Dermatol* 2013;**168**:555–562.
77. Fernandez T, Torres MJ, R-Pena R, Fuentes MS, Robles S, Mayorga C et al. Decrease of selective immunoglobulin E response to amoxicillin despite repeated administration of benzylpenicillin and penicillin V. *Clin Exp Allergy* 2005;**35**:1645–1650.
78. Bousquet PJ, Gaeta F, Bousquet-Rouanet L, Lefrant JY, Demoly P, Romano A. Provocation tests in diagnosing drug hypersensitivity. *Curr Pharm Des* 2008;**14**:2792–2802.
79. Benahmed S, Picot MC, Dumas F, Demoly P. Accuracy of a pharmacovigilance algorithm in diagnosing drug hypersensitivity reactions. *Arch Intern Med* 2005;**165**:1500–1505.
80. Sassolas B, Haddad C, Mockenhaupt M, Dunant A, Liss Y, Bork K et al. ALDEN, an algorithm for assessment of drug causality in Stevens-Johnson Syndrome and toxic epidermal necrolysis: comparison with case-control analysis. *Clin Pharmacol Ther* 2010;**88**:60–68.
81. Barbaud A, Reichert-Penetrat S, Trechot P, Jacquin-Petit MA, Ehlinger A, Noirez V et al. The use of skin testing in the investigation of cutaneous adverse drug reactions. *Br J Dermatol* 1998;**139**:49–58.
82. Brockow K, Romano A, Aberer W, Bircher AJ, Barbaud A, Bonadonna P et al. Skin testing in patients with hypersensitivity reactions to iodinated contrast media - a European multicenter study. *Allergy* 2009;**64**:234–241.
83. Romano A, Gaeta F, Valluzzi RL, Alonzi C, Viola M, Bousquet PJ. Diagnosing hypersensitivity reactions to cephalosporins in children. *Pediatrics* 2008;**122**:521–527.
84. Padiá A, Antunez C, Blanca-Lopez N, Fernandez TD, Cornejo-García JA, Mayorga C et al. Non-immediate reactions to beta-lactams: diagnostic value of skin testing and drug provocation test. *Clin Exp Allergy* 2008;**38**:822–828.
85. Ponvert C, Perrin Y, Bados-Albiero A, Le Bourgeois M, Karila C, Delacourt C et al. Allergy to betalactam antibiotics in children: results of a 20-year study based on clinical history, skin and challenge tests. *Pediatr Allergy Immunol* 2011;**22**:411–418.
86. Ponvert C. Diagnosis of allergic and non-allergic hypersensitivity reactions to commonly used drugs and biological substances in children: diagnostic algorithm. *Arch Pediatr* 2011;**18**:486–492.
87. Demoly P, Romano A, Botelho C, Bousquet-Rouanet L, Gaeta F, Silva R et al. Determining the negative predictive value of provocation tests with beta-lactams. *Allergy* 2010;**65**:327–332.
88. Defrance C, Bousquet PJ, Demoly P. Evaluating the negative predictive value of provocation tests with nonsteroidal anti-inflammatory drugs. *Allergy* 2011;**66**:1410–1414.
89. Solensky R, Earl HS, Gruchalla RS. Lack of penicillin re-sensitization in patients with a history of penicillin allergy after receiving repeated penicillin courses. *Arch Intern Med* 2002;**162**:822–826.
90. Goldberg A, Confino-Cohen R. Skin testing and oral penicillin challenge in patients with a history of remote penicillin allergy. *Ann Allergy Asthma Immunol* 2008;**100**:37–43.
91. Fontaine C, Mayorga C, Bousquet PJ, Arnoux B, Torres MJ, Blanca M et al. Relevance of the determination of serum-specific IgE antibodies in the diagnosis of immediate beta-lactam allergy. *Allergy* 2007;**62**:47–52.
92. Gueant JL, Mata E, Monin B, Moneret-Vautrin DA, Kamel L, Nicolas JP et al. Evaluation of a new reactive solid phase for radioimmunoassay of serum specific IgE against muscle relaxant drugs. *Allergy* 1991;**46**:452–458.
93. Ebo DG, Sainte-Laudy J, Bridts CH, Mertens CH, Hagendorens MM, Schuerwegh AJ et al. Flow-assisted allergy diagnosis: current applications and future perspectives. *Allergy* 2006;**61**:1028–1039.
94. Demoly P, Lebel B, Messaad D, Sahla H, Rongier M, Daures JP et al. Predictive capacity of histamine release for the diagnosis of drug allergy. *Allergy* 1999;**54**:500–506.
95. Lebel B, Messaad D, Kvedariene V, Rongier M, Bousquet J, Demoly P. Cysteinyl-leukotriene release test (CAST) in the diagnosis of immediate drug reactions. *Allergy* 2001;**56**:688–692.
96. Watkins J, Wild G. Improved diagnosis of anaphylactoid reactions by measurement of serum tryptase and urinary methylhistamine. *Ann Fr Anesth Reanim* 1993;**12**:169–172.
97. Kvedariene V, Kamey S, Ryckwaert Y, Rongier M, Bousquet J, Demoly P et al. Diagnosis of neuromuscular blocking agent hypersensitivity reactions using cytofluorimetric analysis of basophils. *Allergy* 2006;**61**:311–315.
98. Sanz ML, Gamboa P, de Weck AL. A new combined test with flowcytometric basophil activation and determination of sulfidoleukotrienes is useful for in vitro diagnosis of hypersensitivity to aspirin and other nonsteroidal anti-inflammatory drugs. *Int Arch Allergy Immunol* 2005;**136**:58–72.
99. Torres MJ, Padiá A, Mayorga C, Fernandez T, Sanchez-Sabate E, Cornejo-García JA et al. The diagnostic interpretation of basophil activation test in immediate allergic reactions to betalactams. *Clin Exp Allergy* 2004;**34**:1768–1775.
100. Ebo DG, Leysen J, Mayorga C, Rozieres A, Knol EF, Terreehorst I. The in vitro diagnosis of drug allergy: status and perspectives. *Allergy* 2011;**66**:1275–1286.
101. Chen P, Lin JJ, Lu CS, Ong CT, Hsieh PF, Yang CC et al. Carbamazepine-induced toxic effects and HLA-B\*1502 screening in Taiwan. *N Engl J Med* 2011;**364**:1126–1133.
102. Simons FE, Arduoso LR, Bilo MB, Dimov V, Ebisawa M, El-Gamal YM et al. 2012 Update: World Allergy Organization Guidelines for the assessment and management of anaphylaxis. *Curr Opin Allergy Clin Immunol* 2012;**12**:389–399.
103. Simons FE, Arduoso LR, Bilo MB, El-Gamal YM, Ledford DK, Ring J et al. World Allergy Organization anaphylaxis guidelines: summary. *J Allergy Clin Immunol* 2011;**127**:587–593.
104. Simons FE, Arduoso LR, Bilo MB, El-Gamal YM, Ledford DK, Ring J et al. World allergy organization guidelines for the assessment and management of anaphylaxis. *World Allergy Organ J* 2011;**4**:13–37.
105. Garcia-Doval I, LeCleach L, Bocquet H, Otero XL, Roujeau JC. Toxic epidermal necrolysis and Stevens-Johnson syndrome: does early withdrawal of causative drugs decrease the risk of death? *Arch Dermatol* 2000;**136**:323–327.
106. Mockenhaupt M. The current understanding of Stevens-Johnson syndrome and toxic epidermal necrolysis. *Expert Rev Clin Immunol* 2011;**7**:803–813.
107. Demoly P, Messaad D, Sahla H, Fabre J, Faucherre V, Andre P et al. Six-hour trimethoprim-sulfamethoxazole-graded challenge in HIV-infected patients. *J Allergy Clin Immunol* 1998;**102**:1033–1036.
108. Castells MC. Hypersensitivity to antineoplastic agents. *Curr Pharm Des* 2008;**14**:2892–2901.
109. Gollapudi RR, Teirstein PS, Stevenson DD, Simon RA. Aspirin sensitivity: implications

- for patients with coronary artery disease. *JAMA* 2004;**292**:3017–3023.
110. Scherer K, Brockow K, Aberer W, Gooi JH, Demoly P, Romano A et al.; ENDA, the European Network on Drug Allergy and the EAACI Drug Allergy Interest Group. Desensitization in delayed drug hypersensitivity reactions – an EAACI position paper of the Drug Allergy Interest Group. *Allergy* 2011;**68**:844–852.
111. Berges-Gimeno MP, Simon RA, Stevenson DD. Long-term treatment with aspirin desensitization in asthmatic patients with aspirin-exacerbated respiratory disease. *J Allergy Clin Immunol* 2003;**111**:180–186.
112. Riemer AB, Gruber S, Pali-Scholl I, Kinaciyan T, Untersmayr E, Jensen-Jarolim E. Suppression of gastric acid increases the risk of developing immunoglobulin E-mediated drug hypersensitivity: human diclofenac sensitization and a murine sensitization model. *Clin Exp Allergy* 2010;**40**:486–493.
113. Florvaag E, Johansson SG, Irgens A, de Pater GH. IgE-sensitization to the cough suppressant pholcodine and the effects of its withdrawal from the Norwegian market. *Allergy* 2011;**66**:955–960.
114. Thong BY, Mirakian R, Castells M, Pichler W, Romano A, Bonadonna P et al. A World Allergy Organization international survey on diagnostic procedures and therapies in drug allergy/hypersensitivity. *World Allergy Organ J* 2011;**4**:257–270.
115. Adkinson NF Jr, Essayan D, Gruchalla R, Haggerty H, Kawabata T, Sandler JD et al. Task force report: future research needs for the prevention and management of immune-mediated drug hypersensitivity reactions. *J Allergy Clin Immunol* 2002;**109**:S461–S478.



## Fixed drug eruption

---

**Author**

Tetsuo Shiohara, MD, PhD  
Professor and Chairman,  
Department of Dermatology  
Kyorin University School of  
Medicine

**Section Editor**

Maja Mockenhaupt, MD, PhD  
Section Editor — Drug  
Eruptions  
Professor of Dermatology  
University Medical Center,  
Freiburg, Germany  
Disclosures:  
Grant/Research Support  
(unrestricted grants to the  
University of Freiburg):  
Cephalon USA;  
GlaxoSmithKline UK; Pfizer  
USA; MSD Sharp & Dohme  
Germany; Merck Germany;  
Tibotec Belgium (Severe  
cutaneous adverse  
reactions, analysis or search  
of RegiSCAR database).  
Consultant/Advisory Boards:  
Sanofi France; Boehringer  
Ingelheim; Merck USA;  
Pfizer USA (severe  
cutaneous adverse drug  
reactions).

**Deputy Editor**

Rosamaria Corona, MD, DSc  
Deputy Editor —  
Dermatology  
Disclosures: Employee of  
UpToDate, Inc.

**Document Version:** 2.0  
**Last Major Updated**  
**Date:** Jan 9, 2014

---

### Fixed drug eruption

**INTRODUCTION** — Fixed drug eruption (FDE) is a distinctive type of cutaneous drug reaction that characteristically recurs in the same locations upon reexposure to the offending drug. Acute FDE usually presents with a single or a small number of dusky red or violaceous plaques that resolve leaving postinflammatory hyperpigmentation ([picture 1A-C](#)). Rare severe atypical variants of FDE, including multiple, nonpigmenting, and generalized bullous variants, share clinical features with Stevens-Johnson syndrome/toxic epidermal necrolysis.

FDE will be discussed in this topic. Other types of drug eruptions are discussed separately. (See "[Drug eruptions](#)" and "[Exanthematous \(morbilliform\) drug eruption](#)" and "[Lichenoid drug eruption \(drug-induced lichen planus\)](#)" and "[Stevens-Johnson syndrome and toxic epidermal necrolysis: Pathogenesis, clinical manifestations, and diagnosis](#)".)

**EPIDEMIOLOGY** — Cutaneous skin reactions occur in approximately 2 to 3 percent of patients taking drugs. Fixed drug eruptions (FDE) are less common than exanthematous (morbilliform) eruptions, which are estimated to account for up to 95 percent of cutaneous drug reactions [1-3]. FDEs occur in both sexes and in all age groups; in children, FDEs account for 14 to 22 percent of cutaneous drug reactions [4,5].

## **ETIOLOGY AND PATHOGENESIS**

**Eliciting drugs** — Many drugs may induce fixed drug eruptions (FDE). The frequency with which individual drugs cause FDE varies over time and from country to country, depending upon drug availability and rates of consumption. Drugs most frequently associated with FDE include [6-8]:

- Antibacterial agents (trimethoprim-sulfamethoxazole, tetracyclines, penicillins, quinolones, dapsone)
- NSAIDs (acetylsalicylic acid, ibuprofen, naproxen, mefenamic acid)
- Acetaminophen (paracetamol)
- Barbiturates
- Antimalarials

**Immunologic mechanisms** — Intraepidermal CD8<sup>+</sup> T cells are thought to have a key role in mediating the localized epidermal lesion that characterizes FDE [9]. CD8<sup>+</sup> T cells with an effector-memory phenotype are abundantly detected along the dermoepidermal junction in established FDE lesions and persist in resting FDE lesions long after the clinical resolution (picture 2A-B) [9-13].

Sequential analysis of FDE lesions before and after rechallenge with the causative drug demonstrated that intraepidermal CD8<sup>+</sup> T cells are directly involved in epidermal damage (picture 3) [14]. CD8<sup>+</sup> T cells remain quiescent but in a primed state in healed FDE lesions unless the causative drug is readministered. Upon rechallenge, CD8<sup>+</sup> T cells are activated to release interferon gamma and cytotoxic granules such as granzyme B and perforin [12-14]. Mast cells also contribute to the activation of intraepidermal T cells via the induction of cell adhesion molecules on the surrounding keratinocytes, through the action of tumor necrosis factor alpha [13].

The activation of intraepidermal T cells is sufficient for triggering the reaction but insufficient to cause the extensive tissue damage observed in fully evolved lesions. Cytokines and/or adhesion molecule-mediated recruitment of CD4<sup>+</sup> T cells, CD8<sup>+</sup> T cells, and neutrophils may contribute to tissue damage in fully developed FDE lesions.

In the late stage of the immune response, CD4<sup>+</sup>CD25<sup>+</sup>Foxp3<sup>+</sup> regulatory T (Treg) cells are recruited into the lesion and participate in the homeostatic control of the immune reaction. The majority of the expanded or activated cell populations responsible for epidermal damage are eventually removed from tissue by apoptosis. However, a proportion of intraepidermal CD8<sup>+</sup> T cells is prevented from undergoing apoptosis by IL-15 secreted by basal keratinocytes and persists at the injury site as a memory T cell population [14,15].

**HISTOPATHOLOGY** — Histologically, FDE is the prototype of a lichenoid tissue reaction with pigmentary incontinence (accumulation of melanin in the upper dermis and dermal macrophages) [16]. In florid lesions, major features include hydropic degeneration of basal keratinocytes, dyskeratotic cells scattered in the epidermis, lymphocytic infiltration of the dermis, and dermal melanophages. The hair follicles and acrosyringia (the intraepidermal

portions of the sweat gland duct) also may be involved. In some lesions, the exocytosis of lymphocytes is marked, mimicking Pautrier microabscesses of mycosis fungoides.

Healed hyperpigmented lesions show pigmentary incontinence and a mild perivascular inflammatory infiltrate.

The pathologic changes occurring in a resting lesion after reexposure to the offending drug have been documented by sequential biopsies [14]:

- Resting FDE lesions are characterized by a small number of resident CD8<sup>+</sup> lymphocytes aligned along the epidermal side of the dermoepidermal junction (picture 2A-B). The overlying epidermis appears normal (picture 3).
- Two to three hours after rechallenge, the lymphocytes originally adhering to the basal layer move upward to the lower one-half of the epidermis while preserving the epidermal structures.
- Twenty-four to 48 hours after rechallenge, the typical changes of interface dermatitis can be seen, including vacuolar degeneration of keratinocytes, dermal lymphocytic infiltrate extending upward into the epidermis, and dermal pigmentary incontinence. The extent of epidermal damage is highly variable, ranging from intercellular edema and dyskeratotic cells scattered in the epidermis to confluent epidermal necrosis resembling TEN.

Several atypical histologic reaction patterns have been described in FDE, including a neutrophilic reaction, leukocytoclastic vasculitis, and nonpigmenting FDE (NPFDE) [17-20]. In NPFDE, the epidermal changes are mild or absent and a superficial perivascular infiltrate of lymphocytes and eosinophils without melanophages is seen in the upper reticular dermis. In generalized bullous FDE, the histologic features of the blister area closely resemble those seen in SJS/TEN (full-thickness necrosis of the epidermis without pigmentary incontinence) [21].

**CLINICAL MANIFESTATIONS** — Fixed drug eruption (FDE) typically presents with well demarcated, round to oval, dusky red to brown/black macules that may evolve into edematous plaques with or without vesiculation or blistering (picture 1A-B, 1D). Lesions are usually solitary, but may occur in small groups.

Systemic symptoms, such as fever and malaise, are usually absent. Pruritus and a burning or stinging sensation are common. However, the estimated frequency of these findings is not known because they are not consistently reported in case series.

FDE may occur anywhere on the body. Sites of predilection include the lips, genitalia, perianal area, hands, and feet [22]. On mucosal areas, erosions and ulcers may develop (picture 1E). FDE occasionally develops at the site of an antecedent trauma (eg, insect bite, burn, venipuncture) [23,24].

Acute lesions generally appear 30 minutes to 8 hours after drug administration, but can occur up to two weeks after drug exposure [7,25]. After discontinuation of the offending drug, lesions resolve spontaneously in 7 to 10 days, leaving a persistent gray/brown or slate gray postinflammatory hyperpigmentation (picture 1C).

Upon reexposure to the offending drug, lesions typically recur in the same site, but new lesions may develop elsewhere (picture 4). After one or more localized eruptions, FDE rarely

may evolve into a bullous generalized form mimicking Stevens-Johnson syndrome/toxic epidermal necrolysis ([picture 5](#)). (See '[Generalized FDE](#)' below and '[Generalized bullous FDE](#)' below.)

FDE can be induced by drugs with a chemical structure similar to the causative drug (cross-reactivity). There are isolated reports of FDE lesions reactivated by chemically unrelated drugs, a phenomenon known as polysensitivity [[26,27](#)].

**CLINICAL VARIANTS** — Fixed drug eruption (FDE) rarely may present with atypical features that mimic other skin diseases such as erythema multiforme, Stevens-Johnson syndrome/toxic epidermal necrolysis (SJS/TEN), cellulitis, paronychia, neutrophilic dermatosis, or large-plaque parapsoriasis.

**Erythema multiforme-like FDE** — FDE may present with targetoid lesions that mimic erythema multiforme ([picture 6](#)). In contrast to erythema multiforme, these lesions have only two concentric zones of color with a darker, dusky hue in the center. (See "[Pathogenesis, clinical features, and diagnosis of erythema multiforme](#)", section on '[Clinical manifestations](#)'.)

**Generalized FDE** — The typical lesions are multiple and disseminated and involve the trunk and extremities, sparing the mucosal and the semi-mucosal areas ([picture 7](#)). Generalized FDE may be difficult to distinguish clinically and histologically from erythema dyschromicum perstans ([picture 8](#)) [[28](#)].

**Generalized bullous FDE** — Generalized bullous fixed drug eruption (GBFDE) is an extremely rare form of FDE characterized by widespread red or brown macules or plaques with overlying large flaccid bullae ([picture 5](#)) [[29](#)]. Systemic symptoms, such as fever, malaise, or arthralgias may be present.

Patients with GBFDE can be misdiagnosed as having SJS/TEN, but in GBFDE mucosal involvement is usually absent or mild and the clinical course is favorable with rapid resolution in 7 to 14 days after drug discontinuation [[30-32](#)].

However, the prognosis of GBFDE may be unfavorable in older patients, particularly in those with comorbidities.

In a case-control study including patients with GBFDE (age range 68 to 84 years) and matched controls with SJS/TEN, the mortality rate among patients with GBFDE was 22 percent, similar to that observed among patients with SJS/TEN with the same amount of skin detachment [[33](#)]. (See "[Stevens-Johnson syndrome and toxic epidermal necrolysis: Pathogenesis, clinical manifestations, and diagnosis](#)", section on '[Clinical presentation](#)'.)

**Nonpigmenting FDE** — Nonpigmenting fixed drug eruptions (NPFDE) have been described in a small number of patients, most often in association with the administration of pseudoephedrine. NPFDE presents with large solitary or multiple, well-circumscribed erythematous plaques that resolve without postinflammatory hyperpigmentation [[17,34](#)].

A severe form of NPFDE, characterized by disseminated, large, symmetrical plaques and systemic symptoms may be misdiagnosed as SJS/TEN.

**Other variants** — Other rare forms of FDE have been described in isolated reports and include [[35-37](#)]:

- FDE presenting with acute paronychia
- FDE presenting with a linear distribution reminiscent of herpes zoster
- FDE mimicking large-plaque parapsoriasis

## DIAGNOSIS

**Clinical** — The diagnosis of fixed drug eruption (FDE) in its typical presentation is usually straightforward, based upon lesion morphology and history:

- Single or a small number of round or oval, demarcated, erythematous or hyperpigmented macules or plaques located most often on the lips, genitalia, and extremities ([picture 1A-B, 1E](#)).
- History of drug intake in the hours or days preceding the eruption. The medication history should be taken in great detail and include all types of medications and routes of administration, since patients tend to overlook medications that they have been taking sporadically over years. (See "[An approach to the patient with drug allergy](#)", section on 'Identification of the suspect drug'.)
- History of recurrence in the same sites following the administration of the same drug or a chemically related drug.

**Biopsy** — A skin biopsy for histopathologic examination may be warranted in the following circumstances:

- Unusual clinical presentation (eg, generalized or bullous FDE) that raises the suspicion of severe drug reaction
- Presence of systemic symptoms (eg, fever, malaise)
- When the diagnosis is uncertain (eg, negative medication history)

Histologic findings that support the diagnosis of FDE include hydropic degeneration of the basal layer, pigmentary incontinence, single necrotic keratinocytes (dyskeratotic cells) in the upper layers of the epidermis, and a dermal inflammatory infiltrate predominantly composed of lymphocytes. In generalized bullous FDE, the examination of a biopsy taken from the blister area reveals full-thickness necrosis of the epidermis without pigmentary incontinence. (See '[Histopathology](#)' above.).

**Provocation tests** — Systemic (oral challenge) and topical (patch testing) provocation tests can be performed to identify the culprit drug when history is unclear or multiple medications are suspected [[6,15](#)].

**Systemic provocation** — Oral challenge with the suspected drug induces the reactivation of a resting FDE lesion. In most cases, oral challenge is preferred to patch testing because it reproduces the conditions of exposure.

Oral challenge is contraindicated in patients with a history of generalized FDE. In patients without generalized FDE, oral challenge is considered relatively safe because the cutaneous response is localized and the risk of a severe reaction is low. In a series of 450 patients with FDE, oral challenge with the suspected drug produced pruritus in 10 percent of patients, fever in 0.9 percent, and generalized urticaria in 0.7 percent, but no more severe adverse effects [[6](#)].



However, neither the appropriate dose of the suspected drug sufficient to induce a mild reaction nor the timing of the test after the resolution of the initial eruption has been standardized. Our approach is to perform the test two to four weeks after the resolution of the eruption, starting with one-tenth of a single therapeutic dose and gradually increasing the amount every 12 to 24 hours up to one single therapeutic dose. Others may use a different approach. In a prospective series of 93 patients with FDE, oral challenge was started with one-half of the therapeutic dose; if no reaction was elicited, a full dose was given [38].

A flare-up reaction occurring within 30 minutes to 8 hours of the oral challenge within a resting FDE lesion is considered a positive test response ([picture 4](#)) [15].

**Patch testing** — Patch testing also can be used to confirm the diagnosis if oral testing cannot be performed (eg, patients with a history of generalized FDE) or if the patient or parent refuses an oral challenge. Patch testing can be performed by applying the suspected drug to an old FDE lesion to elicit a local reaction. Patch testing is generally considered to be safe because the reaction is localized.

There is no standardized method for patch testing in FDE. Various drug concentrations and application modalities (eg, single open, repeated open, or occlusive) have been used in several series of patients [7,8,39,40].

Our approach is to use the commercially available drug mixed in petrolatum or diluted in water at the concentration of approximately 10 to 20 percent. The drug is applied to the target area in open or occlusive modality. The development of erythema with or without induration that starts within 24 hours and lasts for at least 6 hours is considered a positive reaction ([picture 9](#)) [39].

False negative results may occur when testing drugs with limited ability to penetrate into the skin or when FDE is caused by a drug metabolite [15]. False positive reactions may be observed with drug concentrations higher than 20 percent due to patch test sensitization. (See "[Patch testing](#)", [section on 'Active sensitization'](#).)

**DIFFERENTIAL DIAGNOSIS** — The differential diagnosis of fixed drug eruption (FDE) includes:

**Erythema multiforme** – Erythema multiforme (EM) is rarely induced by drugs. The number, distribution, and morphology of lesions help to differentiate EM from FDE. Target lesions consisting of three components (a dusky central area or blister, a dark red inflammatory zone surrounded by a pale ring of edema, and an erythematous halo) are the hallmark of EM ([picture 10](#)). They are generally numerous and symmetrically distributed on the extremities. Mucosal lesions are present in many cases (EM majus) ([picture 11](#)). (See "[Pathogenesis, clinical features, and diagnosis of erythema multiforme](#)", [section on 'Clinical manifestations'](#).)

**Stevens-Johnson Syndrome/Toxic epidermal necrolysis** – Generalized bullous fixed drug eruption (GBFDE) may sometimes be difficult to differentiate from Stevens-Johnson syndrome/toxic epidermal necrolysis (SJS/TEN) ([picture 12](#)). However, in SJS/TEN, lesions have a less defined border than FDE lesions with a tendency to coalesce; mucous membranes (eg, oral, conjunctival) are involved in over 90 percent of cases ([picture 13](#)); and patients have systemic symptoms and rapid disease progression.

The histologic features in the early stage of SJS/TEN are similar to those of GBFDE [32]; however, as the disease progresses, full thickness epidermal necrosis with skin detachment above the basement membrane becomes evident. In most patients, GBFDE resolves rapidly in 7 to 14 days after drug discontinuation [30,31]. The course of SJS/TEN is generally longer than in GBFDE and complications and sequelae (eg, sepsis, organ failure, late ophthalmic and pulmonary complications) are frequent. (See "[Stevens-Johnson syndrome and toxic epidermal necrolysis: Management, prognosis, and long-term sequelae](#)", section on '[Long-term sequelae](#)' and "[Stevens-Johnson syndrome and toxic epidermal necrolysis: Pathogenesis, clinical manifestations, and diagnosis](#)".)

**Bullous pemphigoid** – Bullous pemphigoid (BP) typically occurs in patients older than 60 years and is frequently preceded by a prodromal phase characterized by pruritic inflammatory plaques resembling urticaria. The typical skin lesions of BP are large, tense blisters that rupture and heal spontaneously without scarring ([picture 14](#)). Histology reveals a subepidermal blister with a dermal infiltrate of eosinophils, neutrophils, and lymphocytes ([picture 15](#)). Direct immunofluorescence of perilesional skin shows linear IgG and/or linear C3 staining along the basement membrane zone ([picture 16](#)), which is absent in FDE. The diagnosis of BP can be confirmed by enzyme-linked immunosorbent assay demonstrating circulating autoantibodies against the bullous pemphigoid antigens BP180 NC16A domain and BP230. (See "[Clinical features and diagnosis of bullous pemphigoid and mucous membrane pemphigoid](#)", section on '[Clinical features](#)' and "[Clinical features and diagnosis of bullous pemphigoid and mucous membrane pemphigoid](#)", section on '[Diagnosis](#)'.)

**Large-plaque parapsoriasis** – Large-plaque parapsoriasis may resemble the pigmented patches of FDE ([picture 17A-B](#)). A skin biopsy is necessary to clarify the diagnosis. Histology reveals epidermal hyperplasia (or atrophy in poikilodermatous areas) and a dermal lymphocytic infiltrate that can become band-like at the dermal epidermal junction and show epidermotropism (exocytosis of small lymphocytes into the epidermis).

## MANAGEMENT

**Drug withdrawal and avoidance** — Discontinuation of the offending drug is the most important aspect of management of FDE. After drug discontinuation, lesions resolve without treatment in a few days leaving postinflammatory hyperpigmentation.

Patients should be advised to avoid the offending drug and chemically related drugs and be provided with a written list of the generic and brand names of the culprit drug and possibly of cross-reactive drugs.

**Symptomatic treatment** — The treatment of FDE is largely symptomatic and aimed at the relief of pruritus. The efficacy of symptomatic therapies for the treatment of FDE has not been evaluated in randomized trials. However, their use is based on clinical experience and indirect evidence of benefit in patients other pruritic skin conditions. (See "[Pruritus: Overview of management](#)".)

- For patients with single or a small number of lesions, we suggest medium to high potency topical corticosteroids (groups one to three ([table 1](#))) and systemic antihistamines. Topical corticosteroids are applied two times per day for 7 to 10 days. Oral H1 antihistamines are generally used, including:
  - Diphenhydramine – 25 to 50 mg orally every four to six hours for adults and children  $\geq 12$  years; 12.5 to 25 mg orally every four to six hours for children 6

to 11 years; and 6.25 mg orally every four to six hours for children 2 to 5 years. Diphenhydramine is continued until pruritus subsides.

- Hydroxyzine – 25 mg orally three to four times per day for adults and children ≥6 years; 2 mg/kg per day orally divided every six to eight hours for children <6 years. Hydroxyzine is continued until pruritus subsides.
- For patients with generalized FDE or generalized bullous FDE, particularly if systemic symptoms are present, a short course of moderate dose systemic corticosteroids (eg, prednisone 0.5 to 1 mg/kg per day for three to five days) may be beneficial.

## SUMMARY AND RECOMMENDATIONS

- Fixed drug eruption (FDE) is a cutaneous drug reaction that characteristically recurs in the same locations upon reexposure to the offending drug. Antibacterial sulfonamides, antibiotics, nonsteroidal antiinflammatory drugs, analgesics, and hypnotics are the most frequent causes of FDE. (See '[Introduction](#)' above and '[Eliciting drugs](#)' above.)
- FDE typically presents with solitary round to oval, dusky red to brown/black macules that may evolve into edematous plaques or bullae ([picture 1A-B, 1D](#)). Sites of predilection include the lips, genitalia, perianal area, and extremities. Acute lesions usually develop 30 minutes to 8 hours after drug administration and resolve spontaneously in 7 to 10 days, leaving a persistent gray/brown or slate gray postinflammatory hyperpigmentation ([picture 1C](#)). (See '[Clinical manifestations](#)' above.)
- In rare cases, FDE may present with atypical features, including generalized bullous FDE and nonpigmenting FDE that mimic more severe drug eruptions such as Stevens-Johnson syndrome/toxic epidermal necrolysis ([picture 5](#)). (See '[Clinical variants](#)' above.)
- The diagnosis of fixed drug eruption (FDE) in its typical presentation is usually straightforward, based upon lesion morphology and history. Histologic examination of a skin biopsy is helpful in establishing the diagnosis in cases with unusual clinical features. (See '[Clinical](#)' above and '[Biopsy](#)' above.)
- Systemic or topical provocation tests may be helpful in identifying the culprit drug when history is unclear or multiple medications are suspected. (See '[Provocation tests](#)' above.)
- Discontinuation and avoidance of the offending drug is the most important aspect of management of FDE. Symptomatic treatment of the acute eruption may include medium to high potency topical corticosteroids (groups one to three ([table 1](#))) and systemic antihistamines. (See '[Drug withdrawal and avoidance](#)' above and '[Symptomatic treatment](#)' above.)

## Reference

1	Bigby M, Jick S, Jick H, Arndt K. Drug-induced cutaneous reactions. A report from the Boston Collaborative Drug Surveillance Program on 15,438 consecutive inpatients, 1975 to 1982. JAMA 1986; 256:3358.
2	Bigby M. Rates of cutaneous reactions to drugs. Arch Dermatol 2001; 137:765.
3	Lee AY. Fixed drug eruptions. Incidence, recognition, and avoidance. Am J Clin Dermatol 2000; 1:277.

4	Khaled A, Kharfi M, Ben Hamida M, et al. Cutaneous adverse drug reactions in children. A series of 90 cases. <i>Tunis Med</i> 2012; 90:45.
5	Sharma VK, Dhar S. Clinical pattern of cutaneous drug eruption among children and adolescents in north India. <i>Pediatr Dermatol</i> 1995; 12:178.
6	Mahboob A, Haroon TS. Drugs causing fixed eruptions: a study of 450 cases. <i>Int J Dermatol</i> 1998; 37:833.
7	Brahimi N, Routier E, Raison-Peyron N, et al. A three-year-analysis of fixed drug eruptions in hospital settings in France. <i>Eur J Dermatol</i> 2010; 20:461.
8	Nnoruka EN, Ikeh VO, Mbah AU. Fixed drug eruption in Nigeria. <i>Int J Dermatol</i> 2006; 45:1062.
9	Shiohara T, Mizukawa Y. Fixed drug eruption: a disease mediated by self-inflicted responses of intraepidermal T cells. <i>Eur J Dermatol</i> 2007; 17:201.
10	Shiohara T, Moriya N. Epidermal T cells: their functional role and disease relevance for dermatologists. <i>J Invest Dermatol</i> 1997; 109:271.
11	Shiohara T, Mizukawa Y, Teraki Y. Pathophysiology of fixed drug eruption: the role of skin-resident T cells. <i>Curr Opin Allergy Clin Immunol</i> 2002; 2:317.
12	Komatsu T, Moriya N, Shiohara T. T cell receptor (TCR) repertoire and function of human epidermal T cells: restricted TCR V alpha-V beta genes are utilized by T cells residing in the lesional epidermis in fixed drug eruption. <i>Clin Exp Immunol</i> 1996; 104:343.
13	Mizukawa Y, Yamazaki Y, Teraki Y, et al. Direct evidence for interferon-gamma production by effector-memory-type intraepidermal T cells residing at an effector site of immunopathology in fixed drug eruption. <i>Am J Pathol</i> 2002; 161:1337.
14	Mizukawa Y, Yamazaki Y, Shiohara T. In vivo dynamics of intraepidermal CD8+ T cells and CD4+ T cells during the evolution of fixed drug eruption. <i>Br J Dermatol</i> 2008; 158:1230.
15	Shiohara T. Fixed drug eruption: pathogenesis and diagnostic tests. <i>Curr Opin Allergy Clin Immunol</i> 2009; 9:316.
16	Shiohara T, Mizukawa Y. The immunological basis of lichenoid tissue reaction. <i>Autoimmun Rev</i> 2005; 4:236.
17	Shelley WB, Shelley ED. Nonpigmenting fixed drug eruption as a distinctive reaction pattern: examples caused by sensitivity to pseudoephedrine hydrochloride and tetrahydrozoline. <i>J Am Acad Dermatol</i> 1987; 17:403.
18	Agnew KL, Oliver GF. Neutrophilic fixed drug eruption. <i>Australas J Dermatol</i> 2001; 42:200.
19	Ozkaya E, Büyükbabani N. Neutrophilic fixed drug eruption caused by naproxen: a real entity or a stage in the histopathologic evolution of the disease? <i>J Am Acad Dermatol</i> 2005; 53:178.
20	Harris A, Burge SM. Vasculitis in a fixed drug eruption due to paracetamol. <i>Br J Dermatol</i> 1995; 133:790.
21	Mockenhaupt M. Severe drug-induced skin reactions: clinical pattern, diagnostics and therapy. <i>J Dtsch Dermatol Ges</i> 2009; 7:142.
22	Korkij W, Soltani K. Fixed drug eruption. A brief review. <i>Arch Dermatol</i> 1984; 120:520.
23	Mizukawa Y, Shiohara T. Trauma-localized fixed drug eruption: involvement of burn scars, insect bites and venipuncture sites. <i>Dermatology</i> 2002; 205:159.
24	Shiohara T, Mizukawa Y. Recall phenomenon: some skin-resident cells remember previous insults. <i>Dermatology</i> 2003; 207:127.
25	Breathnach SM. Drug reactions. In: Rook's Textbook of Dermatology, 8th, Tony Burns, Stephen Breathnach, Neil Cox and Christopher Griffiths. (Eds), Wiley-Blackwell, 2010. Vol IV.
26	Ozkaya E. Polysensitivity in fixed drug eruption due to a novel drug combination-