# ドラッグ・リポジショニング(DR)による医薬品開発

## DR による大腸がん予防の有用性

ーアスピリンによる大腸ポリープ再発抑制ー

Potential of Drug Repositioning for Colorectal Cancer Prevention: Inhibition of Colorectal Polyp Recurrence by Aspirin

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有望ながん化学予防剤とその予防介入試験について,これまでの基礎資料を提示し,実 現可能性の観点からドラッグ・リポジショニングが今後のがん予防に有用であることを述 べる。具体的には,アスピリンのがん予防介入試験を題材にして,試験対象者数や研究デ ザイン,さらには政策への提言に関して考察する。

#### 1. はじめに

「がん予防」というテーマは、一般になじみ深 く、興味も高い。しかし我が国には、がん化学予 防「薬」はない。我が国においても、多くの基礎 研究の成果によりがん化学予防「剤」が見出され てきたのは事実であるが、がん予防「剤」が「薬」 として実用化されるためには克服すべき問題が多 く残っている。疾患名のない状態には原則として 国民皆保険制度は適用されず、健康な状態を対象 とした予防薬は保健の適用外と考えられている (そのため、胃がん予防に有効と考えられるピロ リ菌の除菌は感染症由来の別疾患として保険適用 を行い、また子宮頸がん予防に有効な HPV ワク チンは保険適用でなく、公的補助のみである)。 しかし、対象がしっかりと絞られており、エンド ポイントもはっきりとした臨床試験における有効 性を示せるデータがあれば, 公的補助やオーファ ンドラッグ申請など, 予防薬が公的に認められる 将来像を描くことは十分に可能である。

日本人における真のがん予防を目指していくためには、日本人を対象とした予防介入試験による結果からの裏付けが必須であると言える。不十分とはいえ、ある程度の発がんメカニズムの解明も進み<sup>1,2)</sup>、proof of concept を担保しつつがん予防介入試験を行うことが可能である時代になってきた。したがって、日本人において実現可能性の高いがん予防介入試験に関する体制整備(基盤整備)を必要予算の概略を含めて提示していくことがこれからの重要な課題である。

筆者らはアスピリンを用いたがん予防臨床介入 試験におけるアジアで初めての成功を本年発表した<sup>3)</sup>。ランダム化比較試験(RCT)レベルのがん 予防臨床試験は施行が難しく、その成果は、数十年に一度出るかというまれな出来事であり、今回 の成果はがん化学予防剤開発のマイル・ストーンとなる成果であると自負している。食品成分ではなく合成化合物としては、1996年非環式レチノイドによる肝がん再発予防(岐阜大学 武藤教授)が我が国で初めて報告されたがん化学予防の

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成功例である<sup>4)</sup>。この報告から今回の成功まで約 18年の歳月が必要であったことを考えると実務 的開発の難しさが想像される。

製薬企業は新薬開発に苦しんでいるが、この課題を克服する手段としてドラッグ・リポジショニング (DR) の概念がある。今回のアスピリン臨床試験はまさにその成功例であり、開発を促す追い風となると考えられる (探索レベルからがん予防剤を開発するのでは間に合わないほど、高齢化によりがん罹患者数は増加している)。 医療経済的な意義を鑑みても、予防薬が公的に認められる可能性は十分にあると考えており、将来的ながん予防介入試験の成果からその実用への道筋を示せ

たら,企業に新たなフロンティアを示せ,日本発の薬剤開発成功例として,経済発展に大きく貢献できる可能性もある。そこで本稿では生活習慣に起因するがん種であり、日本人の罹患数の多いがんとして大腸がんを取り上げ,がん予防介入試験に向けた現状の整理と実現化に関して,アスピリン介入試験を中心に解説する。

# 2. 基礎研究において予防効果の確認されている大腸がん化学予防剤

1976年に M. Sporn 博士ががん化学予防の定義 (化合物を用いたがんの増殖抑制)を発表してか ら既に 38.年が経ち<sup>5)</sup>, 多くのがん化学予防剤の

表 1 動物実験において大腸がん予防効果が確認されている主な物質(合成化合物)

カテゴリー		標的	化合物名	
阻害剤	酵素	Carbonic anhydrase	Acetazolamide	
		Cyclooxygenase	Aspirin	
			Celecoxib	
			JTE-523	
		,	Indomethacin	
			Mofezolac	
			Nimesulide	
	•		Piroxicam	
			Sulindac	
		Farnesyltransferase	Manumycin A	
		HMG-CoA reductase	Pitavastatin	
		iNOS	Aminoguadine	
			L-NAME	
			GOFA-L-NAME	
			ONO-1714	
			SG-51	
		NADPH oxidase	Apocynin	
		ODC	Difluoromethylornithine (DFMO)	
		PAI-1	SK-116, SK-216	
	受容体	Angiotensin I	Candesartan	
			Losartan	
		EGF	EKB-785	
			Pelitinib/EKB-569	
		Histamine	Cimetidine (H2)	
		Prostaglandin E <sub>2</sub>	ONO-8711 (EP1)	
			ONO-AE2-227 (EP4)	
舌性化剤		AMPK	Metformin	
		LPL	NO-1886	
		PPAR	Pioglitazone	

候補が挙げられた。しかし、臨床応用がされているものは乳がんに対するタモキシフェンなどごく少数に限られ、日本において臨床応用例はない。ここでは Apc 遺伝子変異動物や化学発がん実験系を含むげっ歯類の動物実験において、大腸発がん予防効果が確認されている合成低分子化合物および天然物をそれぞれ表 1 および表 2 にまとめてみた。

表 2 動物実験において大腸がん予防効果が確認 されている主な物質(天然物)

カテゴリー	化合物名等					
Coffee	Chlorogenic acids					
	Cafestol					
Fat	Docosahexaenoic acid (DHA)					
	Fish oil					
Fiber	Cellulose					
	Fructo-oligosacharide					
	Guar gum					
	Wheat bran					
Phytochemical	Catechin					
	Chafuroside					
	Croein					
	Curcumin					
	Resveratrol					
	Soy isoflavones					
	Tea extract (green)					
Vitamin	Folate (Vit. B)					
Other	BCAA					
	Bovine lactoferrin					
	Magnesium					
	Selenium					
	Sphingomyelin-ceramides					

#### 2.1 大腸発がん予防効果が確認されている合成 低分子化合物

合成化合物は大きく分子の阻害剤と活性化剤に 分類される。一見して、活性化を促す低分子化合 物は少なく. 多くは阻害剤であることが分かる。 また、阻害剤は酵素と受容体とを標的としている。 疾患等のないヒトを対象としていることから、が ん化学予防剤に求められる最も大切な条件として 副作用がほとんどないことが求められる。抗がん 剤に散見される DNA 合成阻害効果ではなく酵素 活性阻害を標的としている候補物質が多いこと は、そのことを物語っている。また、標的酵素の 多くは炎症に関わり、標的受容体としては増殖因 子に関わるものが多くを占めている。D. Hanahan 博士と R. A. Weinberg 博士が指摘して いるように、発がんの初期段階において重要な役 割を演じているのが、細胞増殖やアポトーシスに 関わる分子である1,2)。現在の分子生物学・遺伝学 の進歩を受け、がん化学予防剤の開発も具体的分 子を標的とした分子標的予防薬の開発が中心と なってきていると言える。アスピリンなどのシク ロオキシゲナーゼ (COX) の阻害 (図1) やプロ スタグランジンの受容体の阻害はメカニズム的に 同様な増殖シグナルや抗アポトーシスの阻害を意 味している。また、HMG-CoA 還元酵素阻害と farnesyl transferase 阻害もほぼ同様なメカニズ ム阻害に由来していると考えられる<sup>6,7)</sup>。

### 2.2 大腸発がん予防効果が確認されている天然物 天然物としてがん予防効果が検討されている物

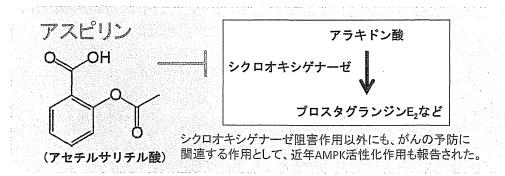


図1 アスピリンによるシクロオキシゲナーゼの阻害

質は、疫学研究によりその有効性が示唆されているものが中心である(表 2)。発がんモデル動物を用いる実験によりその成分ごとの解析も行われている。例えばコーヒー成分であるクロロゲン酸、 $\omega$ -3 系脂肪酸であるドコサヘキサエン酸(DHA)、フレンチパラドックスとして有名な赤ワイン成分レスベラトロールなどが有効成分として挙げられる。これらの想定作用機序は多岐に渡っており、多面的な効果が見出されている。しかしそれが逆に科学的な投与方法の確立やがん予防機構の解明を複雑にしているという一面がある。

# 3. RCT レベルで予防効果の確認されているがん化学予防剤

実際の介入試験に基礎データをつなげるためには多くの疫学的観察研究結果の蓄積が必要である。また、何にもまして優先順位の決定には、先にも述べたようにがん予防効果の強弱よりも副作用がないことが最優先されることを留意すべきである。そのような視点に立つと、介入試験に入る優先順位の高いがん化学予防剤の候補は、すでに市場に出ている薬剤の転用(DR)により開発するのが妥当と思われる。RCTレベルで大腸腺腫又は大腸がんの予防効果の報告されている主な物質を表3にまとめた。

表3 ランダム化比較試験にて大腸腺腫/がん予防効果 が報告されている主な物質

カテゴリー	化合物名等	対象
合成化合物	Aspirin	Lynch 症候群 家族性大腸腺腫症 大腸腺腫の既往者
天然物	Celecoxib Sulindac DFMO + Sulindac Eicosapentaenoic	家族性大腸腺腫症 家族性大腸腺腫症 大腸腺腫の既往者 家族性大腸腺腫症
	acid (EPA) Bovine lactoferrin	大腸腺腫症

#### 3.1 大腸発がん予防効果が確認されている合成 低分子化合物

がん予防効果の確認されている薬剤を見ると, 多くは抗炎症作用を主な機序としている。しか し、大腸がん予防薬として最も期待されていたコ キシブ系薬剤(COX-2選択的阻害剤)に重篤な 心血管系障害の副作用が見つかり、開発は振り出 しに戻っていた<sup>8)</sup>。その一方、アスピリンは世に 出てから今年で115年目を迎え、副作用などが十 分に分かっているため、近年再注目されている。 本年公表した筆者らのデータではアスピリンは大 腸腫瘍の再発リスクを40%も減弱させることを 示唆するデータが得られた<sup>3)</sup>。一方,糖尿病薬で あるメトホルミンの大腸がんに対する予防効果 は、十分に示されていない。その他、最近の臨床 試験データベースを見ると, synthetic lethal な 考え方を考慮して2剤以上の多剤併用投与が増え てきている。

#### 3.2 大腸発がん予防効果が確認されている天然物

天然物としてはアジア人では魚油のエイコサペンタエン酸(EPA)の有効性が示されている<sup>9</sup>。ラクトフェリンに関しては動物実験ほどの強烈ながん予防効果がヒトを対象とした臨床試験では示されなかった<sup>10</sup>。その他、葉酸はがん高危険度群での検討はなく、13のRCTを集めたメタ解析でそのがん予防効果が否定されている。カルシウムやセレニウムの効果は報告により結果が異なり、レスベラトロールやクルクミンは安全性試験が終了、現在進行中の試験が多い。

# 4. 想定される段階別の臨床試験方法および対象集団

がんの予防をエンドポイントとした介入試験は 主に欧米で行われており、我が国にはほとんどな い。しかし、超高齢化社会を迎えて国民の医療へ の関心が高くなり、医療経済への対応が迫られる 中、先制医療を目指した予防薬開発のニーズは高 まってきている。そこで、がん化学予防剤のサイ ズ別の集団予防への適用性や予防介入試験のデザ

表 4 想定される大腸がん化学予防剤の臨床試験のパターン

対象者数	対象者	デザイン	試験期間	目標とする政策	その他
1~数十人	遺伝性疾患等のがん高危険群 (lifetime risk 40-100%) 例) FAP, Lynch 症候群	症例検討 Phase I			企業参画のための エビデンス
~100 人	遺伝性疾患等のがん高危険群	RCT	半年程度	オーファン・ドラッグ 申請	
~数百人	がん中危険群 (lifetime risk 10-20%) 例) 腺腫既往者/がんサバイバー	RCT	2年程度	保険収載 保険適用拡大	診療ガイドライン への影響
~数千人 ~数万人	一般集団 (lifetime risk <10%)	単一介入	10 年程度	保険収載 保険適用拡大	

インを,アスピリンの臨床介入試験を例として以下に考察する(**表 4**)。

#### 4.1 がんの高危険群に対するがん予防介入試験

1人~数十人を対象とした症例検討では学術的 なエビデンスは低いが、よりエビデンスの高い試 験を施行するための、また企業との連携提案材料 としての利用を考えて実施することができる。生 涯に大腸がんに罹患する可能性により40~100% の集団をがんの高危険群と想定すると, 家族性大 腸腺腫症(FAP)患者や Lynch 症候群の患者が 該当することになる。また、100人レベルでは RCT の臨床試験を行うことができ、その試験期 間も半年程度である110。しかし、逆に該当患者数 も少なく、そのため全国規模の多施設の協力が必 要となる (例えば、 Tuvenile polyposis、 Cowden syndrome, Peutz-Jeghers syndrome などは患者 数が少なすぎて、RCTの施行は難しい)。一方、 これらがんの高危険群に対するオーファンドラッ グとしてのがん化学予防剤の申請は価値のあるも のになると考えられる。

#### 4.2 がんの中危険群に対するがん予防介入試験

生涯に大腸がんに罹患する可能性が10~20%の 集団をがんの中危険群と想定すると、大腸腺腫の 既往があり腺腫を取った患者や、大腸がん治療後 のがんサバイバーにおける再発(3次予防の対象 者)はその中危険群と考えられる。試験としては,数百人レベルでRCTを行うことができ,その試験期間も数年程度と考えられる。しかし,アスピリンの場合は,介入後に20年間観察すると介入後5年くらいから予防効果が強く見られてくることもあり<sup>12)</sup>,試験期間が終了してもさらなる長期的観察が必要なこともあるため,計画を立てる時にはフェールセーフ的に考えておく必要がある。がんの中危険群に対するがん化学予防剤に対し,既知の薬からのDRでは,保険適用拡大を目指すことになると思われるが,予防剤は薬ではないので,何らかの疾患名をつけた保険適用拡大を狙うか,公的補助という別の方向性の可能性も考える必要がある。

#### 4.3 一般集団に対するがん予防介入試験

一般化を目指すためには数万人レベルの臨床試験が必要となる。そのため薬剤の選定や対象者を考察すると、高齢化に伴う疾患(心血管疾患、痴呆等)や代謝疾患(糖尿病、脂質異常症、高血圧、骨粗鬆症など)など、薬適用者数の多い患者を対象とし、DR の考え方で利用できる薬剤を選定するのが現実的であろう。最近アスピリンの次に候補となるがん化学予防剤として、スタチン(脂質異常症治療薬)、メトホルミン(糖尿病治療薬)、ビスフォスフォネート(骨粗鬆症治療薬)が挙げられているが、この中でコホートデータが十分に

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揃っているのはやはりスタチンである<sup>13)</sup>。このレベルの試験のエンドポイントとしては複数想定されるが、大腸がんの発生にすると 10 年程度の試験期間が必要となる。集団における大腸がんの発生頻度からも RCT ではなく多施設単一介入試験が選ばれる可能性が高い。対照をどこに設定するかの問題はあるが、十数年の服用とその結果がポジティブな場合、十分なエビデンスとして保険収載を含めた政策決定に影響を与えるものと思われる。

#### 5. おわりに

私見ではあるが、DHA の基礎的動物データの報告年<sup>14)</sup> と魚油の臨床介入試験の報告年<sup>9)</sup> を考えると、臨床介入試験まで持っていくためには、現在においても約10年の年月が必要と推察される。また、割合を考えると動物試験で得られた100候補のうち数個がヒトでの臨床試験で検討できれば良い方と考えられ、RCT レベルまで検討できている薬剤は約40年の歴史の中でも海外を含めて数えるほどでしかない。臨床レベルに持っていける薬剤数もまだ十分ではなく、年間数例の有望ながん化学予防剤の候補物質が、動物レベル実験で精度良く実施・提案される必要がある。そしてこの選別を経た有力な候補物質を中規模な臨床介入試験で効果確認した上で大規模な臨床介入試験で効果確認した上で大規模な臨床介入試験へと進めることが重要と考えられる。

「がん予防」の研究は、ビジネスとしての展開が開けていないことから、現時点では企業等の参入が難しい領域で、ほとんど全てのがん予防研究は公的研究費で行われている。直接的/実際的に役立つ結果を DR の観点より積み上げて、実務的

な価値判断を一般消費者や行政当局に訴えて行く 必要がある。がん予防研究推進のためにも世論喚 起を引き続き行い、皆様のご協力を期待したい。

[謝辞] アスピリンを用いたがん予防臨床介入試験の功労者は19施設の研究グループの各先生方であり、特に研究を中心的に推進した石川秀樹 京都府立医大特任教授の献身的な活動があっての成果です。また本稿は、厚生労働省 第3次対がん総合戦略事業 (H22-3次がん一般-014) および国立がん研究センター がん研究開発費 (25-A-15) による成果です。

#### 文 献

- D. Hanahan & R. A. Weinberg, Cell, 100, 57 (2000)
- D. Hanahan & R. A. Weinberg, Cell, 144, 646 (2011)
- 3) H. Ishikawa et al., Gut, 63, 1755 (2014)
- 4) Y. Muto et al., N. Engl. J. Med., 334, 1561 (1996)
- 5) M. B. Sporn, Canc. Res., 36, 2689 (1976)
- 6) N. Teraoka et al., Int. J. Can., 129, 528 (2011)
- 7) S. F. Nielsen *et al.*, *N. Engl. J. Med.*, **367**, 1792 (2012)
- 8) S. D. Solomon *et al.*, *N. Engl. J. Med.*, **352**, 1071 (2005)
- 9) N. J. West et al., Gut, 59, 918 (2010)
- 10) T. Kozu et al., Canc. Prev. Res., 2, 975 (2009)
- 11) H. Ishikawa et al., Canc. Med., 2, 50 (2013)
- 12) J. Burn et al., Lancet, 378, 2081 (2011)
- 13) N. Gronich & G. Rennert, *Nat. Rev. Clin. Oncol.*, **10**, 625 (2013)
- 14) M. Takahashi et al., Canc. Res., 53, 2786 (1993)

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REVIEV

## Potential ability of xanthophylls to prevent obesityassociated cancer

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#### **Abstract**

Obesity-associated cancers, including colon cancer and breast cancer, are increasing in Asian countries with Westernized lifestyles as exemplified by reduced physical activity and increased fat/sugar consumption. An excessive accumulation of visceral adipose tissue causes insulin resistance, dyslipidemia and adipocytokine imbalance, and these factors are suggested to be involved in cancer promotion. To prevent obesityassociated cancers, researcher attention is increasing on the so-called "functional foods". In addition, new approaches to cancer control are in high demand, and using "functional foods" as supplemental or adjuvant agents in chemotherapy is thought to be a promising approach. One of these functional ingredients is xanthophylls, which are natural fat-soluble pigments found in fruits, vegetables, algae and other plants. Xanthophylls belong to the carotenoid class and have structures containing oxygen. Some studies have revealed that xanthophylls improve the inflammation status, serum triglyceride levels, blood pressure levels and liver function test values. Furthermore, recent studies show that xanthophylls possess high anti-cancer, anti-diabetic, anti-obesity and anti-oxidant properties. In this review, we highlight the recent findings for five xanthophylls, namely astaxanthin,  $\beta$ -cryptoxanthin, fucoxanthin, neoxanthin and zeaxanthin/lutein, and their relevance to cancer prevention.

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Key words: Cancer prevention; Xanthophylls

Core tip: Xanthophylls belong to the class of carotenoids, and are natural fat-soluble pigments found in fruits, vegetables, algae and so on. It has been shown that the versatile functions of xanthophylls have great potential for the prevention of metabolic syndrome and cancers. Xanthophylls have proved safety, and several xanthophylls provide other health benefits, including improvement of inflammation, dyslipidemia, hypertension and liver function. These findings indicate that xanthophylls could be useful to prevent obesity-associated cancer.

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#### INTRODUCTION

Obesity has recently attracted much interest as a risk factor for several cancers, such as breast cancer and colorectal cancer<sup>[1,2]</sup>. Both metabolic syndrome that is characterized by obesity, hyperlipidemia, type 2 diabetes and hypertension

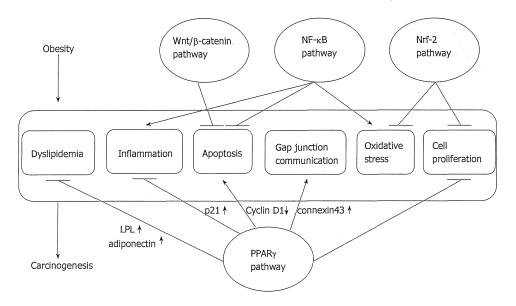


Figure 1 Possible mechanisms for obesity-associated cancer prevention. LPL: Lipoprotein lipase; NF-κB: Nuclear factor kappa B; Nrf2: Nuclear factor-erythroid 2 related factor 2; PPAR: Peroxisome proliferator-activated receptor.

# Table 1 Obesity-associated cancers Type of cancer Breast (postmenopausal) Colorectum Endometrium Esophagus Gallbladder Kidney Pancreas Thyroid

and obesity-associated cancers (Table 1) are extremely common in Western countries, and they are currently increasing in Eastern countries, including Japan. The factors linking obesity and cancer are becoming apparent, and they are insulin resistance, dyslipidemia and a subsequent adipocytokine imbalance (Figure 1)<sup>[1,2]</sup>. Carotenoid intake is reported to be inversely associated with obesity and with the risk of many cancers<sup>[3-6]</sup>.

Carotenoids are fat-soluble pigments found in fruits, vegetables, algae and other plants. Humans cannot synthesize carotenoids, and we should therefore consume them as part of our diet. Carotenoids belong to the tetraterpenoid category, and they can be divided into xanthophylls and carotenes according to whether the structure contains oxygen or not. Carotenoids with structures containing oxygen are xanthophylls. As the name indicates, the color of xanthophylls is usually yellow, and they are usually lipophilic because of the long unsaturated aliphatic chain in their structure.

Because conventional chemotherapy has failed to reduce the mortality rates of common cancers, including obesity-associated cancers, new approaches to controlling the development of cancer are in great demand<sup>[7]</sup>. One approach is the use of functional foods/plant-derived agents as supplemental or adjuvant agents in chemo-

therapy<sup>[8,9]</sup>. Another approach is chemoprevention for the control of cancer development<sup>[8,9]</sup>. In both methods, using xanthophylls seems to be an attractive approach. As shown in this review, xanthophylls provided health benefits, such as improvements in inflammation, dyslipidemia, hypertension and liver function. Moreover, the biological significance of xanthophylls as important candidates for the chemoprevention of cancer is becoming clearer, and the safety of xanthophylls has been affirmed, as described in this review. Another candidate called - carotene is the most abundant dietary carotenoid, and long-term supplementation with this compound has been shown to be ineffective for cancer chemoprevention in several recent large-scale intervention trials<sup>[10-12]</sup>.

In this review, we focus on recent findings for five xanthophylls as follows: astaxanthin,  $\beta$ -cryptoxanthin, fucoxanthin, neoxanthin and zeaxanthin/lutein, and their relevance to cancer prevention (Figure 2).

#### **ASTAXANTHIN**

#### Distribution and nature of astaxanthin

Astaxanthin (AX) is a natural fat-soluble red pigment and belongs to the xanthophyll subclass of carotenoids. Dietary sources of AX are eggs of salmon and trout, skin of red sea bream, crabs, shrimps and lobsters. AX is synthesized in microalgae (*Chlorella zofingiensis*, *Chlorococcum* and *Haematococcus pluvialis*). Krills (*Euphausia superba*) feed on the microalgae and in turn are fed upon by fishes. The microalga, *H. pluvialis*, is the main source of natural AX and is able to accumulate up to 4%AX on dry weight basis<sup>[13-15]</sup>. AX extracted from *H. pluviali* is used as a food dye in many countries. AX exists in stereoisomers and geometric isomers. *H. pluvialis* biosynthesizes the (3S, 3'S)-isomer, meanwhile *P. rhodozyma* biosynthesizes the (3R, 3'R)-isomer. AX has two hydroxyl groups and is able to react with fatty acids and proteins. AX is found as free, mono- and di-ester forms in organisms<sup>[13]</sup>.

Figure 2 Structure of xanthophylls. A: Astaxanthin; B: β-cryptoxanthin; C: Fucoxanthin; D: 9'-cis-neoxanthin; E: Lutein; F: Zeaxanthin.

AX can take to transverse cell membrane orientation, and shows strong antioxidative activity<sup>[13,15]</sup>. After oral administration of AX, AX changes to all-*E*, 9*Z*-, 13*Z*-geometrical isomers and 3*R*,3'*R*-, 3*R*,3'*S* meso-, 3*S*,3'*S*-optical isomers, all of which can be detected in human blood<sup>[16]</sup>.

#### Safety profile

Many experimental and clinical studies have demonstrated the safety of AX<sup>[13,17]</sup>. In a subchronic toxicity study in rats, feeding AX-rich microalgae biomass corresponding to doses of 465 and 557 mg AX/kg per day for 90 d in male and female rats, respectively, revealed no adverse events<sup>[18]</sup>. A randomized, double-blind, placebocontrolled study has demonstrated that it is safe to administrate 6 mg/d AX in healthy adults for 8 wk<sup>[19]</sup>, and a significant decrease of triglycerides and increase of adiponectin and high density lipoprotein cholesterol in participants with mild dyslipidemia by administration of AX at doses of 12 mg/d and 18 mg/d for 12 wk<sup>[20]</sup>.

#### Preclinical studies and anti-cancer mechanisms

Oxidative stress and inflammation are closely related to carcinogenesis (Figure 1), and many antioxidants, including carotenoids have been demonstrated to decrease cancer development in experimental animal models<sup>[14]</sup>. There are papers on preventive effects of AX on urinary bladder<sup>[21]</sup>, oral<sup>[22,23]</sup>, and colorectal<sup>[24,25]</sup> carcinogenesis. In mouse urinary bladder cancer induced by N-butyl-N(4-hydroxybutyl)nitrosamine (OH-BBN), AX administration at a dose of 50 ppm in water for 20 wk after OH-BBN exposure for 20 wk resulted in a decrease in the incidences of precancerous lesions and bladder cancer<sup>[21]</sup>. In rat oral carcinogenesis induced by 4-nitroquinoline 1-oxide (4-NQO), the incidence of oral precancerous le-

sions in rats treated with 20 ppm 4-NQO and 100 ppm AX was smaller compared to those of the non-treatment group, and oral neoplasms did not observed in rats fed AX among the 4-NQO exposure<sup>[22]</sup>. In these studies, AX decreased cell growth activity in the non-cancerous epithelial tissues of 4-NQO-exposed animals<sup>[21,22]</sup>. AX has also been demonstrated to show preventive effects in 7,12-dimethylbenz[a]anthracene (DMBA)-induced hamster buccal pouch carcinogenesis via nuclear factorerythroid 2 related factor 2 (Nrf2) activation<sup>[23]</sup>. Moreover, AX has been shown to inhibit nuclear factor kappa B (NF-κB) and Wnt/β-catenin signaling pathways [26]. Related to colorectal carcinogenesis, AX at 500 ppm in diet significantly decreased the development of aberrant crypt foci (ACF) and the incidence and multiplicity of colorectal tumors induced by azoxymethane (AOM)[24]. AX at 200 ppm in diet also suppressed mucosal ulcers induced by dextran sulfate sodium (DSS), and development of dysplastic ACF and colonic adenocarcinoma induced by both treatment of DSS and AOM<sup>[25]</sup>. In addition, AX reduced the number and size of aflatoxin B1-induced liver preneoplastic foci in rats<sup>[27]</sup>. Growth of WAZ-2T cells, mammary tumor cells, inoculated into the mice mammary fat pad was also inhibited by AX at 100 ppm or 400 ppm in diet<sup>[28]</sup>. Lipid peroxidation activity in tumors was reduced in tumors treated with 400 ppm<sup>[28]</sup>. AX markedly attenuated the promotion of hepatic metastasis of P815 mastocytoma cells in a syngenic graft model under restraint stress<sup>[29]</sup>. In in vitro cell culture systems, AX suppressed invasion of rat ascites hepatoma AH109A cells<sup>[30]</sup>. AX inhibited cell proliferation and decreased cell viability of leukemia K562 cells via induction of apoptosis along with up-regulation of peroxisome proliferatoractivated receptor (PPAR)y and p21, and down-regulation

of cyclin D1 [31]. Induction of connexin 43, gap junction protein, through activation of PPARy is suggested to be one of the anti-tumor mechanisms of AX<sup>[32,33]</sup>. Up-regulation of the Nrf2 pathway is also involved in antioxidant activity of AX<sup>[23,31,34]</sup>, and may improve mitochondrial function<sup>[35]</sup>. However, the role of Nrf2 activation in antitumorigenesis is controversial. The oncogenic K-ras gene induces Nrf2 expression, and detoxification of reactive oxygen species promotes tumor growth [36]. Deficiency of Nrf2 has been reported to increase induction of tumors in urethane-induced mouse lung carcinogenesis, but reduce the number of malignant tumors harboring activated mutation in the K-ras gene, indicating that Nrf2 prevents initiation but accelerates progression under the activation of the K-ras signaling pathway<sup>[37]</sup>. Indeed, there is a report that effects of AX differ at the stages of initiation and the stage of promotion in mammary tumors. AX fed before tumor initiation delayed mammary tumor growth and modulated immune response, but AX supplementation after tumor initiation resulted in more rapid tumor growth [38]. Thus, use of antioxidants for cancer prevention is considered to be useful at the time before tumor initiation, but more caution is required in using them after the stage accompanied with activated K-ras signaling.

#### Clinical studies

A randomized, double-blind, placebo-controlled study has demonstrated that AX reduces oxidation of fatty acids<sup>[39]</sup>, decreases oxidative stress markers<sup>[40]</sup> and inflammation<sup>[41]</sup>, and improves dyslipidemia<sup>[20]</sup> and age-related dysfunction of eyes<sup>[42,43]</sup> and brain<sup>[44]</sup>. However, human cancer prevention studies using AX have not yet been reported.

#### **β-CRYPTOXANTHIN**

#### Distribution and nature of $\beta$ -cryptoxanthin

β-cryptoxanthin (β-CRX) is one of the naturally occurring carotenoid pigments, and is also classified as a xanthophyll. Its unique character is that it is found in specific fruits and vegetables such as mango, papaya, loquat, Japanese persimmon, peach, sweet red peppers and citrus fruits of the mandarin family<sup>[45,46]</sup>. Satsuma mandarin, *Citrus unshiu*, is one of the most β-CRX rich fruits in Japan. The content of β-CRX in *C. unshiu* reaches several mg/100g wet weight. The level of β-CRX in Valencia orange is very low and grapefruit has been found to be devoid of it.

In the human body,  $\beta$ -CRX is easily converted to vitamin A (retinol) and is therefore considered as a provitamin A. It is also known that  $\beta$ -CRX might be easily absorbed<sup>[47]</sup>, and is accumulated in various organs<sup>[48]</sup>. Moreover, it can be stored for several months in the human body<sup>[49]</sup>. Serum  $\beta$ -CRX concentration could be around 96  $\mu$ g/dL<sup>[50]</sup>. It is also reported that  $\beta$ -CRX concentration in Japanese mother's milk and serum are nearly parallel with their intake of the Satsuma mandarin, and

are higher than other countries [51,52].

#### Epidemiologic studies

Many epidemiological studies showed the intake of β-CRX was significantly associated with reduced risks of type 2 diabetes [relative risk (RR) = 0.58]<sup>[53]</sup> and rheumatoid arthritis (RR = 0.59)<sup>[54]</sup>.  $\beta$ -CRX supplementation significantly decreased cigarette smoke-induced lung squamous metaplasia and inflammation<sup>[55]</sup>. Regarding cancer risk, several observational epidemiologic studies suggest that B-CRX could potentially prevent cancer development. The demonstrated cancer risks for lung, esophageal and bladder were 0.76 (RR), 0.16 [odds ratio (OR)] and 0.74 (RR), respectively, comparing the highest to lowest quintile of intake<sup>[56-58]</sup>. A greater intake of β-CRX was also inversely associated with developing undetermined cervical atypical squamous cells (OR = 0.4)<sup>[59]</sup>. Interestingly, the serum level of  $\beta$ -CRX is lower in the patients of liver cancer than that in healthy counterparts [60]. These results suggest that a high serum β-CRX concentration or intake of  $\beta$ -CRX is beneficial to human health.

#### Safety profile

The scientific panel on additives and products or substances used in animal feed (FEEDAP) panel members considered  $\beta\text{-CRX}$  to appear not to be mutagenic and show no clastogenic activity [61]. In subchronic studies, The FEEDAP panel could not find any adverse effects [61]. Also an acceptable daily intake has not been determined [61]. Previously, we have reported the chemoprevention effect of  $\beta\text{-CRX}$  against chemically-induced bladder carcinogenesis in ICR mice [62]. Mice were fed with 1, 5 and 25 ppm of  $\beta\text{-CRX}$  for 24 wk, and no clinical signs of toxicity and poor condition, low survival or histopathological changes were found [62]. Many epidemiological studies [53-60,63-68] indicated that administration of  $\beta\text{-CRX}$  is safe for human health.

#### Preclinical studies and anti-cancer mechanisms

Various functions of  $\beta$ -CRX have been reported recently.  $\beta$ -CRX is an antioxidant phytochemical and may help prevent oxidative damage<sup>[69]</sup>. Thus, it is believed that  $\beta$ -CRX has health benefits for people with risk of chronic diseases.

Numerous possible mechanisms for the anti-carcinogenic potential of  $\beta$ -CRX have been proposed. These include the antioxidant function that is associated with the enhancement of DNA repair<sup>[55,69]</sup>, suppression of efficacy of key proinflammatory cytokine expression, such as tumor necrosis factor- $\alpha$ <sup>[55]</sup> and an apoptotic induction effect<sup>[70]</sup>. Also,  $\beta$ -CRX is known to stimulate the expression of the RB gene (a tumor-suppressor gene) and p73 gene (a p53-related gene)<sup>[71]</sup> and reduce the expression of NF- $\kappa$ B and activator protein-1 (AP-1), that induces numerous genes including inflammation, cell proliferation, and apoptosis<sup>[55]</sup>. These mechanisms indicate that  $\beta$ -CRX may be a promising chemopreventive agent against cancer. Indeed,  $\beta$ -CRX exerts an anti-tumor promoter action *in vitro*<sup>[72]</sup> and

inhibits chemically induced carcinogenesis in vivo<sup>[62,71,73,74]</sup>. Previously, we investigated the effects of β-CRX extracted from C. unshiu oranges on OH-BBN-induced urinary bladder carcinogenesis in male ICR mice<sup>[62]</sup>. OH-BBN-exposed mice were fed with 1, 5 and 25 ppm of β-CRX for 24 wk starting 1 wk after the cessation of OH-BBN exposure. Feeding with  $\beta$ -CRX decreased the incidence and multiplicity of precancerous and cancerous urinary bladder lesions. Especially, 25 ppm β-CRX markedly reduced the occurrence of bladder cancer. Meanwhile, β-CRX is also reported to reduce mouse skin<sup>[71]</sup>, mouse lung<sup>[74]</sup> and rat colon<sup>[71]</sup> carcinogenesis. In our report, β-CRX lowered ratios of cyclin D1-positive cell in various urinary bladder lesions, meaning that reduction in the incidence of precancerous and cancerous urinary bladder lesions is due to reduced cell cycle progression<sup>[62]</sup>.

#### Clinical studies

The efficacy of β-CRX supplementation on obesity have been investigated<sup>[75]</sup>. Seventeen postmenopausal obese women were provided 200 mL of a beverage containing β-CRX (1.56 mg/serving and 4.7 mg/d) for 3 wk<sup>[75]</sup>. As a result, the levels of serum β-CRX were significantly elevated from 0.28 (initial period) to 1.15 mg/mL, and high molecular weight-adiponectin was also elevated from 9.8 to 11.1 mg/mL<sup>[75]</sup>. At the end of the study, the levels of serum triglyceride (P = 0.057) and total plasminogen activator inhibitor-1 (PAI-1) (P = 0.052) tended to decrease. Nishino et al601 reported an intervention study where β-CRX-rich mandarin orange juice (3 mg β-CRX in 80 mL) was provided for 12 wk to obese men or obese men with elevated serum  $\gamma$ -glutamyl transpeptidase ( $\gamma$ GTP) levels [60]. After drinking  $\beta$ -CRX for 12 wk, body weight (P < 0.001), BMI (P < 0.001) and β-GTP levels (P < 0.005) were decreased.

An intervention trial regarding prevention of liver cancer has also been reported<sup>[60]</sup>. Viral hepatitis with cirrhosis patients were randomly assigned into two groups in the trial. The treatment group was administered mandarin orange juice enriched with  $\beta$ -CRX and with the carotenoids mixture (lycopene,  $\beta$ -carotene and  $\alpha$ -carotene). Patients in the control group were administered a carotenoids mixture alone. At year 2.5, cumulative incidence of liver cancer/hepatocellular carcinoma development in the mandarin orange juice group was lower than that of the carotenoids mixture alone group (P = 0.05). The combinational use of natural carotenoids containing  $\beta$ -CRX might be valuable for the prevention of liver cancer in hepatitis virus infected patients with cirrhosis.

#### **FUCOXANTHIN**

#### Distribution and nature of fucoxanthin

Brown seaweeds include *Undaria pinnatifida* (wakame), *Hizikia fusiforme* (hijiki), *Laminaria japonica* (ma-kombu) and *Sargassum fulvellum*. The Japanese have been estimated to intake wakame at 1 g/d<sup>[76]</sup>. Brown seaweeds are known to contain many bioactive components, *i.e.*,

fucoxanthin (FX), fucoidan, vitamins, minerals, dietary fibers, proteins, ω-3 polyunsaturated fatty acids (PUFAs), polysaccharides, other carotenoids and various functional polyphenols. Fucoidan is a sulfated polysaccharide that is one of the major bioactive components in seaweed[77,78], but we would like to focus on FX in this review. FX is a xanthophyll belonging to non-provitamin A carotenoids, constructed with an unusual allenic bond, an epoxide group, and a conjugated carbonyl group in a polyene chain<sup>[79]</sup>. Some reports demonstrated that the FX content of *U. pinnatifida* is approximately 1.0-3.0 mg/g dry weight through one life cycle [80,81]. It has been proven that mice convert FX into keto-carotenoids by oxidation of the secondary hydroxyl groups (FX +  $H_2O \rightarrow FuOH$ ;  $FuOH + NAD^{+} \rightarrow amarouciaxanthin A + NADH)^{[79]}$ . On the other hand, oral administration of kombu extract containing FX in humans revealed that the FuOH and the as-isomer of FuOH could be found in the serum, detected by HPLC<sup>[82]</sup>.

#### Safety profile

FX has been proved to be safe with no side effects by single (1000 or 2000 mg/kg BW) and repeated (500 or 1000 mg/kg BW for 30 d) oral dose toxicity studies in male and female mice<sup>[83]</sup>. In the repeated doses study, histological examination of the gonadal tissues, kidneys, liver and spleen revealed no abnormal changes<sup>[83]</sup>. In rats, 13-wk oral subchronic toxicity studies suggested that more than 2000 mg/kg BW of microalgal FX oil induce the 50% lethal<sup>[84]</sup>.

#### Preclinical studies and anti-cancer mechanisms

Many studies suggested FX possesses anti-cancer potential, especially shown in colon cancer cell lines (Caco-2, DLD-1 and HT-29), liver cell lines (HepG2), prostate cancer cell lines (DU 145, LNCaP and PC-3) and urinary bladder<sup>[85-88]</sup>. The main biomolecules involved in anti-cancer mechanism is assumed to be the biomolecules related to apoptosis and cell cycle<sup>[89,90]</sup> and those may associate with antioxidant activity through their free radical scavenging action<sup>[91]</sup>. Moreover, inhibition of PI3K/Akt and NF-<sub>K</sub>B signals were reported in human cervical and breast cancer cells, respectively<sup>[92,93]</sup>.

Its metabolite fucoxanthinol (FuOH) also has inhibitory effects on cancer cell growth [94,95], and 1,2-dimethylhydrazine-induced formation of colonic ACF in mice and AOM/DSS-induced colon carcinogenesis [25,96]. To find new cancer prevention approaches, we investigated the combination effect of FuOH and 1α,25-dihydroxyvitamin D<sub>3</sub> (1α,25(OH)<sub>2</sub>D<sub>3</sub>), and found inhibition of cell viability and induction of apoptosis in DLD-1 and HT-29 cells [97]. Down-regulation of PPARγ and NF-κB p52 were suggested to be involved in the inhibition of cell viability due to the combination of FuOH and 1α,25(OH)<sub>2</sub>D<sub>3</sub>. It has been shown that activation of PPARγ suppresses intestinal polyp development in *Apc*-mutant mice and AOM-induced colonic ACF development in obese KK-A mice [98,99].

#### Clinical studies

FX has been reported to provide health benefits in humans, such as improvement of obesity, reduction of inflammation, healthy triglyceride levels, and improvements in blood pressure levels [100,101].

After daily intake of *U. pinnatifida*, FuOH is detectable in human plasma<sup>[82]</sup>. Although metabolites of FX could be measured as a marker of exposure, effects of FX or FuOH in human carcinogenesis have not been reported to date. From the aspect of obesity-associated cancer, we here introduce one study that has been conducted to assess the effects of FX supplementation on weight loss. FX supplementation on obese patients with nonalcoholic fatty liver disease results in the improvement of liver inflammatory markers, such as alanine aminotransferase, aspartate aminotransferase, C-reactive protein, γ-glutamyltransferase (γGT, GGT)<sup>[101]</sup>. Of note, it has been demonstrated that increased of GGT plasma levels are associated with an increased risk of pancreatic cancer<sup>[102,103]</sup>, nevertheless GGT has no causative role itself.

It is also interesting to mention that intake of 5 g/d *U. pinnatifida* stimulated a significant 50% reduction in urinary urokinase-like plasminogen activator receptor (uPAR) proteins in postmenopausal women. uPAR, is the membrane receptor for uPA, responsible for extracellular membrane proteins degradation and PAI-1, responsible for the inhibition of plasminogen activation<sup>[104]</sup>. Generally, uPAR is known to be higher in postmenopausal women as well as in breast cancer patients<sup>[105]</sup>. Moreover, it has been reported that uPA and/or PAI-1 is positively correlated with poor prognosis in patients with breast cancer, *i.e.*, correlation with cancer metastatic potential<sup>[106,107]</sup>. Thus, uPA, PAI-1and uPAR might be used as prognostic markers for breast cancer<sup>[108]</sup>, and FX may reduce such a tumor marker.

#### **NEOXANTHIN**

#### Distribution and nature of Neoxanthin

Neoxanthin (NX), a non-provitamin A carotenoid, has an unusual allenic bond and a 5,6-monoepoxide as well as FX. NX is widely present in terrestrial and marine biota and the occurrence of two geometric cis/trans isomers is known to be species dependent<sup>[109-111]</sup>. The 9'-cis form of NX (9'-cis NX) is mainly localized and used in the photosynthetic organs of spinach leaves and marine algae such as Euglenophyta. It is also used as a precursor of abscisic acid, a plant hormone [112,113]. Whereas the all-trans form of NX is predominant in the petals of globeflower and yellow rose, this xanthophyll is not involved in the photosynthetic system<sup>[111,114]</sup>. We mainly obtain the 9'-as NX from leafy green vegetables. Fresh spinach contains 9'-as NX around 5 mg/100 g in fresh leaf [110]. It has been estimated that 9'-ais NX exists at 0.95 µmol/L in digested fluid (9 L/d), when we ingest 100 g/d spinach.

The 5,6-monoepoxide moiety in 9'-cis NX is easily isomerized to 5,8-epoxide under the acidic conditions of the stomach and generates almost equal amounts of

(8'-R/S)-neochrome [115,116]. After a 1-wk spinach intervention (3 mg 9'-cis NX/day), highly hydrophilic xanthophylls of 9'-cis NX and (8'-R and 8'-S)-neochromes appeared at a very low level in human plasma (about 1 nmol/L)[117]. It is known that the uptake of various carotenoids by human colon cancer cells (Caco-2 cells) positively correlates with the lipophilicity of the carotenoids [118]. The highly hydrophilic xanthophylls such as NX, FX and violaxanthin could be detected slightly in human plasma, when we intake purified forms and food matrices [79,101,117,119-121]. Because of the poor intestinal absorption of NX, a considerable amount of ingested 9'-cis NX and (8'-R and 8'-S)-neochrome would be delivered to the colon, and even if absorbed in the small intestine, they would be metabolized easily.

#### Preclinical studies and anti-cancer mechanisms

It has been reported that both 9'-cis NX and all-trans NX possess strong potential of cell growth inhibition and apoptosis induction in human prostate cancer cells [87,94,115,122], human colon cancer<sup>[122-124]</sup>, mouse melanoma<sup>[122]</sup> and mouse embryonic mesenchymal cells<sup>[125]</sup>. In addition, several researchers have reported that 9'-cis NX, all-trans NX and (8' R/S)-neochrome have cancer preventive effects<sup>[126]</sup>, and also anti-tumor promoter functions<sup>[70]</sup>. Moreover, induction of cell cycle arrest<sup>[115]</sup>, anti-oxidant properties<sup>[127]</sup> and antiobesity properties [128] have been reported. Recently, we additionally demonstrated that 9'-cis NX rapidly accumulated in the mitochondria, caused mitochondria  $\Delta \Psi$  loss and thereafter the release of cytochrome c and production of apoptosis-inducing factor in human colon cancer cells [123]. It is regrettable that there is little information about the anticancer mechanisms of dietary NX in mammals, except for that described above.

#### Safety profile and clinical studies

No safety profile and clinical studies have been reported on 9'-cisNX, any -trans NX and (8'R/S)-neochromes. However, epidemiological data show that higher intake of fruits and vegetables, rich in highly hydrophilic epoxyxanthophylls such as NX, is associated with a lower risk of colorectal cancer<sup>[129,130]</sup>. Further studies are required to elucidate the clinical beneficial properties of NX.

#### **ZEAXANTHIN/LUTEIN**

#### Distribution and nature of zeaxanthin / lutein

Zeaxanthin (ZX) and lutein also belong to the xanthophyll family. Their unique character is that they are the only carotenoid among more than 600 species of carotenoid existing in eye tissue, especially in the retina<sup>[131]</sup>. Lutein can be photochemically transformed to meso-ZX. They are stereoisomer of each other, differ by the location of a double bond. Lutein is abundant in egg yolk, and in dark-green leafy vegetables, such as broccoli, brussels sprouts, kale and spinach<sup>[132]</sup>. In the human body, lutein is distributed at the skin, breasts, cervix uteri, and also found in serum in high amounts. Serum lutein and

ZX levels are reported to be around 180 and 20 ng/mL, respectively<sup>[133]</sup>. They are assumed to play a critical role in ocular health because they act as strong anti-oxidants and filtered out high-energy blue light<sup>[134]</sup>. Of note, no correlation between plasma concentrations of lutein/zeaxanthin and BMI or insulin resistance has been reported<sup>[135]</sup>.

#### Epidemiologic studies

In many papers, target organs for lutein are reported to be the eyeballs, the skin and the heart. Regarding ocular conditions, age-related macular degeneration, cataracts, and retina pigmentosa have been reported to have some correlation with lutein. Lutein also possesses a preventive function of cardiovascular diseases/stroke<sup>[131,134,136,137]</sup>.

Regarding lung cancer, some epidemiologic studies state lutein has an important cancer preventive function<sup>[4,14]</sup>. A ten-year study of 120000 United States people revealed that lung cancer incidence was significantly reduced in those who ingested a high amount of total carotenoids, including lutein and ZX<sup>[138]</sup>. Similar relationships were found in Fijians, when compared to the other South Pacific islands' people. Fijians intake 25 mg lutein daily on an average (200 g dark greens), whereas other 20 South Pacific countries intake less lutein in diets<sup>[139]</sup>. Thus, there was a clear inverse association with lutein intake and lung cancer incidence.

Regarding colorectal cancer, inverse associations with dietary lutein intake have been reported [124], and serum ZX concentration by Okuyama  $et\ al^{[140]}$ . However, no association has been detected between the levels of plasma lutein and the risk of gastric cancer [141].

Regarding skin cancer risk, the specific effects of lutein are not fully known. The only reported data is that a combination of carotenoids may protect erythema development in human skin<sup>[142]</sup>, and that may be correlated with the presence of skin cancer or precancerous lesions<sup>[124]</sup>.

Regarding breast cancer, there is some possibility for protective effects of lutein [6,14,143]. Intake of lutein-rich foods significantly lowered the risk of premenopausal breast cancer. The Nurse's Health Study demonstrated a weak inverse association, but significant, between lutein and ZX intake and the breast cancer risk among premenopausal women<sup>[6]</sup>. Of note, the protective effect of lutein and ZX was strongest in patients have a family history of breast cancer. Also there is a report that increasing serum levels of lutein and ZX were associated with a reduced breast cancer risk, but the trend was only marginally significant in a case-control study<sup>[143]</sup>. There is a report comparing biopsy samples from breast cancer tissue and benign mammary tissue. In this report, increasing lutein and ZX concentrations tends to decrease the risk of breast cancer<sup>[144]</sup>. Meanwhile, Other studies have shown that there are no differences of lutein and ZX concentrations in mammary adipose tissue between benign breast tumors and breast cancer<sup>[145]</sup>. New York University Women's Health Study, a nested case-control prospective study, demonstrated an inverse relationship

between plasma levels of lutein, but not ZX, and risk of breast cancer<sup>[146]</sup>.

Regarding other cancers, significant inverse relations were observed for lutein and ZX in oral cavity and pharyngeal cancer<sup>[147]</sup>.

#### Safety profile

No toxicities or adverse reactions for intake of lutein/ZX have been reported at doses up to 40 mg/d for 2 mo<sup>[131,148]</sup>. High doses of  $\beta$ -carotene supplements (> 30 mg/d) are well known to be associated with carotenodermia<sup>[149]</sup>, and the same could happen when we consume high doses of lutein and ZX. Also it has been demonstrated that lutein has no mutagenic effect in the Ames test<sup>[150]</sup>.

#### Preclinical study and anticancer mechanism

Lutein/ZX is thought to have a superior anti-oxidant ability to scavenge free radicals than other carotenoids. An *in vitro* study showed that lutein could quench peroxy radicals and play a guarding role against oxidative injury<sup>[151,152]</sup>. In this experiment, a synergistic antioxidant effect was obtained with a combination of lutein and lycopene<sup>[153]</sup>. Carotenoids also show a superb function for immune response<sup>[154]</sup>.

Lutein could also function as an anti-carcinogenic reagent, such as a modulator of cell growth and apoptosis signaling. Lutein induces cell cycle arrest in human prostate and esophageal cancer cell<sup>[155,156]</sup>. Lutein induces apoptosis in transformed cancer cells but do not induce apoptosis in normal human mammary cells through modulating the ratio of Bcl-xL/Bax protein expression<sup>[157]</sup>. Meanwhile, ZX, structural isomer of lutein, induced cell cycle arrest in human beast cancer cells[158]. Lutein stimulates some genes involved in T-cell transformations activated by antigens, cytokines and mitogens<sup>[159]</sup>. Lutein interacts with carcinogens such as 1-nitropyrene and aflatoxin B1, and lowered its carcinogenetic activity<sup>[150,160]</sup>. In a recent report, female BALB/c mice were fed a diet containing lutein for 14 d, and then inoculated with 0 to  $2.5 \times 10^3$  mammary tumor cells. The results demonstrated that 0.002% and 0.02% lutein lowered both mammary tumor incidence and tumor growth [161].

#### **FUTURE ASPECTS**

The versatile functions of xanthophylls have shown great potential for the prevention of metabolic syndrome and cancers, both *in vitro* and *in vivo*. Xanthophylls have been verified as safe with no side events, and several xanthophylls provide other health benefits, including improvements in inflammation, dyslipidemia, hypertension and liver function, as shown in this review. The accumulated evidence indicates the functionality of xanthophylls as anti-obesity and anti-insulin-resistance functional foods, implying that xanthophylls could be useful in preventing obesity-associated cancer.

The chemical synthesis of each xanthophyll is not impossible, but it may be very expensive. However, the

promising results obtained from *in vivo* studies encourage researchers to undertake more clinical studies in humans. We have some information about xanthophylls trials, and we should further promote human clinical studies to obtain information about the adequate dosage of xanthophylls needed to prevent cancers.

#### **REFERENCES**

- Fujii G, Yamamoto M, Takahashi M, Mutoh M. Role of adipocytokines in colorectal carcinogenesis. Curr Res in Cancer 2011; 5: 39-48
- 2 Ishino K, Mutoh M, Totsuka Y, Nakagama H. Metabolic syndrome: A novel high-risk state for colorectal cancer. *Cancer Lett* 2013; **334**: 56-61 [PMID: 23085010 DOI: 10.1016/j.canlet.2012.10.012]
- 3 Eastwood MA. Interaction of dietary antioxidants in vivo: how fruit and vegetables prevent disease? *QJM* 1999; 92: 527-530 [PMID: 10627873 DOI: 10.1093/qjmed/92.9.527]
- 4 Holick CN, Michaud DS, Stolzenberg-Solomon R, Mayne ST, Pietinen P, Taylor PR, Virtamo J, Albanes D. Dietary carotenoids, serum beta-carotene, and retinol and risk of lung cancer in the alpha-tocopherol, beta-carotene cohort study. Am J Epidemiol 2002; 156: 536-547 [PMID: 12226001 DOI: 10.1093/aje/kwf072]
- 5 Rock CL. Carotenoid update. J Am Diet Assoc 2003; 103: 423-425 [PMID: 12668998 DOI: 10.1016/S0002-8223(03)00164-0]
- 6 Zhang S, Hunter DJ, Forman MR, Rosner BA, Speizer FE, Colditz GA, Manson JE, Hankinson SE, Willett WC. Dietary carotenoids and vitamins A, C, and E and risk of breast cancer. J Natl Cancer Inst 1999; 91: 547-556 [PMID: 10088626 DOI: 10.1093/jnci/91.6.547]
- 7 Sporn MB, Suh N. Chemoprevention: an essential approach to controlling cancer. *Nat Rev Cancer* 2002; 2: 537-543 [PMID: 12094240 DOI: 10.1038/nrc844]
- 8 Ferguson LR, Schlothauer RC. The potential role of nutritional genomics tools in validating high health foods for cancer control: broccoli as example. Mol Nutr Food Res 2012; 56: 126-146 [PMID: 22147677 DOI: 10.1002/mnfr.201100507]
- 9 Temraz S, Mukherji D, Shamseddine A. Potential targets for colorectal cancer prevention. *Int J Mol Sci* 2013; 14: 17279-17303 [PMID: 23975167 DOI: 10.3390/ijms140917279]
- Heinonen OP, Albanes D. The effect of vitamin E and beta carotene on the incidence of lung cancer and other cancers in male smokers. The Alpha-Tocopherol, Beta Carotene Cancer Prevention Study Group. N Engl J Med 1994; 330: 1029-1035 [PMID: 8127329 DOI: 10.1056/NEJM199404143301501]
- Hennekens CH, Buring JE, Manson JE, Stampfer M, Rosner B, Cook NR, Belanger C, LaMotte F, Gaziano JM, Ridker PM, Willett W, Peto R. Lack of effect of long-term supplementation with beta carotene on the incidence of malignant neoplasms and cardiovascular disease. N Engl J Med 1996; 334: 1145-1149 [PMID: 8602179 DOI: 10.1056/NEJM199605023341801]
- 12 Omenn GS, Goodman GE, Thornquist MD, Balmes J, Cullen MR, Glass A, Keogh JP, Meyskens FL, Valanis B, Williams JH, Barnhart S, Hammar S. Effects of a combination of beta carotene and vitamin A on lung cancer and cardiovascular disease. N Engl J Med 1996; 334: 1150-1155 [PMID: 8602180 DOI: 10.1056/NEJM199605023341802]
- Ambati RR, Phang SM, Ravi S, Aswathanarayana RG. Astaxanthin: sources, extraction, stability, biological activities and its commercial applications—a review. *Mar Drugs* 2014; **12**: 128-152 [PMID: 24402174 DOI: 10.3390/md12010128]
- 14 Tanaka T, Shnimizu M, Moriwaki H. Cancer chemoprevention by carotenoids. *Molecules* 2012; 17: 3202-3242 [PMID: 22418926 DOI: 10.3390/molecules17033202]
- 15 Kidd P. Astaxanthin, cell membrane nutrient with diverse clinical benefits and anti-aging potential. Altern Med Rev

- 2011; 16: 355-364 [PMID: 22214255]
- 16 Coral-Hinostroza GN, Ytrestøyl T, Ruyter B, Bjerkeng B. Plasma appearance of unesterified astaxanthin geometrical E/Z and optical R/S isomers in men given single doses of a mixture of optical 3 and 3'R/S isomers of astaxanthin fatty acyl diesters. Comp Biochem Physiol C Toxicol Pharmacol 2004; 139: 99-110 [PMID: 15556071 DOI: 10.1016/j.cca.2004.09.011]
- 17 Fassett RG, Coombes JS. Astaxanthin in cardiovascular health and disease. *Molecules* 2012; 17: 2030-2048 [PMID: 22349894 DOI: 10.3390/molecules17022030]
- Stewart JS, Lignell A, Pettersson A, Elfving E, Soni MG. Safety assessment of astaxanthin-rich microalgae biomass: Acute and subchronic toxicity studies in rats. Food Chem Toxicol 2008; 46: 3030-3036 [PMID: 18588938 DOI: 10.1016/ j.fct.2008.05.038]
- 19 Spiller GA, Dewell A. Safety of an astaxanthin-rich Haematococcus pluvialis algal extract: a randomized clinical trial. J Med Food 2003; 6: 51-56 [PMID: 12804020 DOI: 10.1089/1096 62003765184741]
- Yoshida H, Yanai H, Ito K, Tomono Y, Koikeda T, Tsukahara H, Tada N. Administration of natural astaxanthin increases serum HDL-cholesterol and adiponectin in subjects with mild hyperlipidemia. *Atherosclerosis* 2010; 209: 520-523 [PMID: 19892350 DOI: 10.1016/j.atherosclerosis.2009.10.012]
- 21 Tanaka T, Morishita Y, Suzui M, Kojima T, Okumura A, Mori H. Chemoprevention of mouse urinary bladder carcinogenesis by the naturally occurring carotenoid astaxanthin. Carcinogenesis 1994; 15: 15-19 [PMID: 8293542 DOI: 10.1093/carcin/15.1.15]
- 22 Tanaka T, Makita H, Ohnishi M, Mori H, Satoh K, Hara A. Chemoprevention of rat oral carcinogenesis by naturally occurring xanthophylls, astaxanthin and canthaxanthin. Cancer Res 1995; 55: 4059-4064 [PMID: 7664280]
- 23 Kavitha K, Thiyagarajan P, Rathna Nandhini J, Mishra R, Nagini S. Chemopreventive effects of diverse dietary phytochemicals against DMBA-induced hamster buccal pouch carcinogenesis via the induction of Nrf2-mediated cytoprotective antioxidant, detoxification, and DNA repair enzymes. *Biochimie* 2013; 95: 1629-1639 [PMID: 23707664 DOI: 10.1016/j.biochi.2013.05.004]
- 24 Tanaka T, Kawamori T, Ohnishi M, Makita H, Mori H, Satoh K, Hara A. Suppression of azoxymethane-induced rat colon carcinogenesis by dietary administration of naturally occurring xanthophylls astaxanthin and canthaxanthin during the postinitiation phase. Carcinogenesis 1995; 16: 2957-2963 [PMID: 8603470 DOI: 10.1093/carcin/16.12.2957]
- Yasui Y, Hosokawa M, Mikami N, Miyashita K, Tanaka T. Dietary astaxanthin inhibits colitis and colitis-associated colon carcinogenesis in mice via modulation of the inflammatory cytokines. *Chem Biol Interact* 2011; 193: 79-87 [PMID: 21621527 DOI: 10.1016/j.cbi.2011.05.006]
- 26 Kavitha K, Kowshik J, Kishore TK, Baba AB, Nagini S. Astaxanthin inhibits NF-κB and Wnt/β-catenin signaling pathways via inactivation of Erk/MAPK and PI3K/Akt to induce intrinsic apoptosis in a hamster model of oral cancer. Biochim Biophys Acta 2013; 1830: 4433-4444 [PMID: 23726989 DOI: 10.1016/j.bbagen.2013.05.032]
- 27 Gradelet S, Le Bon AM, Bergès R, Suschetet M, Astorg P. Dietary carotenoids inhibit aflatoxin B1-induced liver preneoplastic foci and DNA damage in the rat: role of the modulation of aflatoxin B1 metabolism. *Carcinogenesis* 1998; 19: 403-411 [PMID: 9525273 DOI: 10.1093/carcin/19.3.403]
- 28 Chew BP, Park JS, Wong MW, Wong TS. A comparison of the anticancer activities of dietary beta-carotene, canthaxanthin and astaxanthin in mice in vivo. Anticancer Res 1999; 19: 1849-1853 [PMID: 10470126]
- 29 Kurihara H, Koda H, Asami S, Kiso Y, Tanaka T. Contribution of the antioxidative property of astaxanthin to its protective effect on the promotion of cancer metastasis in mice treated with restraint stress. *Life Sci* 2002; 70: 2509-2520

- [PMID: 12173414]
- 30 Kozuki Y, Miura Y, Yagasaki K. Inhibitory effects of carotenoids on the invasion of rat ascites hepatoma cells in culture. *Cancer Lett* 2000; 151: 111-115 [PMID: 10766430 DOI: 10.1016/S0304-3835(99)00418-8]
- 31 Zhang X, Zhao WE, Hu L, Zhao L, Huang J. Carotenoids inhibit proliferation and regulate expression of peroxisome proliferators-activated receptor gamma (PPARγ) in K562 cancer cells. Arch Biochem Biophys 2011; 512: 96-106 [PMID: 21620794 DOI: 10.1016/j.abb.2011.05.004]
- 32 Hix LM, Lockwood SF, Bertram JS. Bioactive carotenoids: potent antioxidants and regulators of gene expression. Redox Rep 2004; 9: 181-191 [PMID: 15479561]
- 33 Vine AL, Bertram JS. Upregulation of connexin 43 by retinoids but not by non-provitamin A carotenoids requires RARs. *Nutr Cancer* 2005; 52: 105-113 [PMID: 16091010 DOI: 10.1207/s15327914nc5201\_13]
- 34 Saw CL, Yang AY, Guo Y, Kong AN. Astaxanthin and omega-3 fatty acids individually and in combination protect against oxidative stress via the Nrf2-ARE pathway. Food Chem Toxicol 2013; 62: 869-875 [PMID: 24157545 DOI: 10.1016/j.fct.2013.10.023]
- Wolf AM, Asoh S, Hiranuma H, Ohsawa I, Iio K, Satou A, Ishikura M, Ohta S. Astaxanthin protects mitochondrial redox state and functional integrity against oxidative stress. *J Nutr Biochem* 2010; 21: 381-389 [PMID: 19423317 DOI: 10.1016/j.jnutbio.2009.01.011]
- 36 DeNicola GM, Karreth FA, Humpton TJ, Gopinathan A, Wei C, Frese K, Mangal D, Yu KH, Yeo CJ, Calhoun ES, Scrimieri F, Winter JM, Hruban RH, Iacobuzio-Donahue C, Kern SE, Blair IA, Tuveson DA. Oncogene-induced Nrf2 transcription promotes ROS detoxification and tumorigenesis. Nature 2011; 475: 106-109 [PMID: 21734707 DOI: 10.1038/nature10189]
- 37 Satoh H, Moriguchi T, Takai J, Ebina M, Yamamoto M. Nrf2 prevents initiation but accelerates progression through the Kras signaling pathway during lung carcinogenesis. Cancer Res 2013; 73: 4158-4168 [PMID: 23610445 DOI: 10.1158/0008-5472.CAN-12-4499]
- 38 Nakao R, Nelson OL, Park JS, Mathison BD, Thompson PA, Chew BP. Effect of dietary astaxanthin at different stages of mammary tumor initiation in BALB/c mice. Anticancer Res 2010; 30: 2171-2175 [PMID: 20651366]
- 39 Karppi J, Rissanen TH, Nyyssönen K, Kaikkonen J, Olsson AG, Voutilainen S, Salonen JT. Effects of astaxanthin supplementation on lipid peroxidation. *Int J Vitam Nutr Res* 2007; 77: 3-11 [PMID: 17685090]
- 40 Choi HD, Kim JH, Chang MJ, Kyu-Youn Y, Shin WG. Effects of astaxanthin on oxidative stress in overweight and obese adults. *Phytother Res* 2011; 25: 1813-1818 [PMID: 21480416 DOI: 10.1002/ptr.3494]
- 41 Park JS, Chyun JH, Kim YK, Line LL, Chew BP. Astaxanthin decreased oxidative stress and inflammation and enhanced immune response in humans. *Nutr Metab* (Lond) 2010; 7: 18 [PMID: 20205737 DOI: 10.1186/1743-7075-7-18]
- 42 Parisi V, Tedeschi M, Gallinaro G, Varano M, Saviano S, Piermarocchi S. Carotenoids and antioxidants in age-related maculopathy italian study: multifocal electroretinogram modifications after 1 year. Ophthalmology 2008; 115: 324-333. e2 [PMID: 17716735 DOI: 10.1016/j.ophtha.2007.05.029]
- 43 Piermarocchi S, Saviano S, Parisi V, Tedeschi M, Panozzo G, Scarpa G, Boschi G, Lo Giudice G. Carotenoids in Age-related Maculopathy Italian Study (CARMIS): two-year results of a randomized study. Eur J Ophthalmol 2012; 22: 216-225 [PMID: 22009916 DOI: 10.5301/ejo.5000069]
- 44 Katagiri M, Satoh A, Tsuji S, Shirasawa T. Effects of astaxanthin-rich Haematococcus pluvialis extract on cognitive function: a randomised, double-blind, placebo-controlled study. J Clin Biochem Nutr 2012; 51: 102-107 [PMID: 22962526 DOI: 10.3164/jcbn.11-00017]

- 45 Yano M, Kato M, Ikoma Y, Kawasaki A, Fukazawa Y, Sugiura M, Matsumoto H, Oohara Y, Nagao A, Ogawa K. Quantitation of carotenoids in raw and processed fruits in Japan. Food Sci Technol Res 2005; 11: 13-18 [DOI: 10.3136/fstr.11.13]
- 46 Holden JM, Eldridge AL, Beecher GR, Buzzard IM, Bhagwat S, Davis CS, Douglass LW, Gebhardt S, Haytowitz D, Schakel S. Carotenoid content of US foods: An update of the database. J Food Comp Anal 1999; 12: 169-196 [DOI: 10.1006/jfca.1999.0827]
- 47 Wahlqvist ML, Wattanapenpaiboon N, Macrae FA, Lambert JR, MacLennan R, Hsu-Hage BH. Changes in serum carotenoids in subjects with colorectal adenomas after 24 mo of beta-carotene supplementation. Australian Polyp Prevention Project Investigators. Am J Clin Nutr 1994; 60: 936-943 [PMID: 7985637]
- 48 Sugiura M, Ogawa K, Yano M. Absorption, storage and distribution of β-cryptoxanthin in rat after chronic administration of Satsuma mandarin (Citrus unshiu MARC.) juice. Biol Pharm Bull 2013; 36: 147-151 [PMID: 23302648 DOI: 10.1248/bpb.b12-00836]
- 49 Sugiura M, Kato M, Matsumoto H, Nagao A, Yano M. Serum concentration of beta-cryptoxanthin in Japan reflects the frequency of Satsuma mandarin (Citrus unshiu Marc.) consumption. *J Health Sci* 2002; 48: 350-353 [DOI: 10.1248/jhs.48.350]
- 50 Sugiura M, Matsumoto H, Kato M, Ikoma Y, Yano M, Nagao A. Seasonal changes in the relationship between serum concentration of beta-cryptoxanthin and serum lipid levels. J Nutr Sci Vitaminol (Tokyo) 2004; 50: 410-415 [PMID: 15895516]
- 51 Sugiura M, Matsumoto H, Kato M, Ikoma Y, Yano M, Nagao A. Multiple linear regression analysis of the seasonal changes in the serum concentration of beta-cryptoxanthin. J Nutr Sci Vitaminol (Tokyo) 2004; 50: 196-202 [PMID: 15386932]
- 52 Canfield LM, Clandinin MT, Davies DP, Fernandez MC, Jackson J, Hawkes J, Goldman WJ, Pramuk K, Reyes H, Sablan B, Sonobe T, Bo X. Multinational study of major breast milk carotenoids of healthy mothers. *Eur J Nutr* 2003; 42: 133-141 [PMID: 12811470]
- 53 Montonen J, Knekt P, Järvinen R, Reunanen A. Dietary antioxidant intake and risk of type 2 diabetes. *Diabetes Care* 2004; 27: 362-366 [PMID: 14747214]
- 54 Cerhan JR, Saag KG, Merlino LA, Mikuls TR, Criswell LA. Antioxidant micronutrients and risk of rheumatoid arthritis in a cohort of older women. *Am J Epidemiol* 2003; 157: 345-354 [PMID: 12578805 DOI: 10.1093/aje/kwf205]
- 55 Liu C, Bronson RT, Russell RM, Wang XD. β-Cryptoxanthin supplementation prevents cigarette smoke-induced lung inflammation, oxidative damage, and squamous metaplasia in ferrets. Cancer Prev Res (Phila) 2011; 4: 1255-1266 [PMID: 21421799 DOI: 10.1158/1940-6207.CAPR-10-0384]
- Männistö S, Smith-Warner SA, Spiegelman D, Albanes D, Anderson K, van den Brandt PA, Cerhan JR, Colditz G, Feskanich D, Freudenheim JL, Giovannucci E, Goldbohm RA, Graham S, Miller AB, Rohan TE, Virtamo J, Willett WC, Hunter DJ. Dietary carotenoids and risk of lung cancer in a pooled analysis of seven cohort studies. Cancer Epidemiol Biomarkers Prev 2004; 13: 40-48 [PMID: 14744731 DOI: 10.1158/1055-9965.EPI-038-3]
- 57 De Stefani E, Brennan P, Boffetta P, Ronco AL, Mendilaharsu M, Deneo-Pellegrini H. Vegetables, fruits, related dietary antioxidants, and risk of squamous cell carcinoma of the esophagus: a case-control study in Uruguay. *Nutr Cancer* 2000; 38: 23-29 [PMID: 11341040 DOI: 10.1207/S15327914NC381\_4]
- 58 Zeegers MP, Goldbohm RA, van den Brandt PA. Are retinol, vitamin C, vitamin E, folate and carotenoids intake associated with bladder cancer risk? Results from the Netherlands Cohort Study. Br J Cancer 2001; 85: 977-983 [PMID: 11592769 DOI: 10.1054/bjoc.2001.1968]

- 59 Goodman MT, McDuffie K, Hernandez B, Hankin JH, Wilkens LR, Franke AA, Kolonel LN, Kuypers J, Kiviat N, Bertram CC, Kessel B, Sunoo C, Nakamura J, Killeen J. The Association of Plasma Micronutrients with the Risk of Cervical Atypical Squamous Cells of Undetermined Significance (ASCUS). Asian Pac J Cancer Prev 2000; 1: 337-345 [PMID: 12716311]
- 60 Nishino H, Murakoshi M, Satomi Y. Health promotion by antioxidants. Functional Foods in Health and Disease 2011; 1: 574-581. Available from: URL: http://www.functionalfoodscenter.net/files/48097641.pdf
- 61 Opinion of the scientific panel on additives and products or substances used in animal feed on the request from the commission on the safety of use of colouring agents in animal nutrition. *The EFSA Journal* 2006; 386: 1-40. Available from: URL: http://www.efsa.europa.eu/en/efsajournal/ doc/320.pdf
- 62 Miyazawa K, Miyamoto S, Suzuki R, Yasui Y, Ikeda R, Kohno H, Yano M, Tanaka T, Hata K, Suzuki K. Dietary beta-cryptoxanthin inhibits N-butyl-N-(4-hydroxybutyl)nitrosamine-induced urinary bladder carcinogenesis in male ICR mice. *Oncol Rep* 2007; 17: 297-304 [PMID: 17203164 DOI: 10.3892/or.17.2.297]
- 63 Sugiura M, Nakamura M, Ikoma Y, Yano M, Ogawa K, Matsumoto H, Kato M, Ohshima M, Nagao A. Serum carotenoid concentrations are inversely associated with serum aminotransferases in hyperglycemic subjects. *Diabetes Res Clin Pract* 2006; 71: 82-91 [PMID: 16005096 DOI: 10.1016/j.atherosclerosis.2005.04.006]
- 64 Sugiura M, Nakamura M, Ikoma Y, Yano M, Ogawa K, Matsumoto H, Kato M, Ohshima M, Nagao A. High serum carotenoids are inversely associated with serum gammaglutamyltransferase in alcohol drinkers within normal liver function. J Epidemiol 2005; 15: 180-186 [PMID: 16195638]
- Nakamura M, Sugiura M, Aoki N. High beta-carotene and beta-cryptoxanthin are associated with low pulse wave velocity. Atherosclerosis 2006; 184: 363-369 [PMID: 15936762]
- 66 Sugiura M, Nakamura M, Ikoma Y, Yano M, Ogawa K, Matsumoto H, Kato M, Ohshima M, Nagao A. The homeostasis model assessment-insulin resistance index is inversely associated with serum carotenoids in non-diabetic subjects. *J Epidemiol* 2006; 16: 71-78 [PMID: 16537987 DOI: 10.2188/jea.16.71]
- 67 Sugiura M, Nakamura M, Ogawa K, Ikoma Y, Matsumoto H, Ando F, Shimokata H, Yano M. Associations of serum carotenoid concentrations with the metabolic syndrome: interaction with smoking. *Br J Nutr* 2008; 100: 1297-1306 [PMID: 18445303 DOI: 10.1017/S0007114508978302]
- 68 Sugiura M, Nakamura M, Ogawa K, Ikoma Y, Ando F, Shimokata H, Yano M. Dietary patterns of antioxidant vitamin and carotenoid intake associated with bone mineral density: findings from post-menopausal Japanese female subjects. Osteoporos Int 2011; 22: 143-152 [PMID: 20480147 DOI: 10.1007/s00198-010-1239-9]
- 69 Lorenzo Y, Azqueta A, Luna L, Bonilla F, Domínguez G, Collins AR. The carotenoid beta-cryptoxanthin stimulates the repair of DNA oxidation damage in addition to acting as an antioxidant in human cells. *Carcinogenesis* 2009; 30: 308-314 [PMID: 19056931 DOI: 10.1093/carcin/bgn270]
- 70 Uchiyama S, Yamaguchi M. Beta-cryptoxanthin stimulates apoptotic cell death and suppresses cell function in osteoclastic cells: change in their related gene expression. J Cell Biochem 2006; 98: 1185-1195 [PMID: 16514646 DOI: 10.1002/ jcb.20824]
- 71 Nishino H, Tokuda H, Murakoshi M, Satomi Y, Masuda M, Onozuka M, Yamaguchi S, Takayasu J, Tsuruta J, Okuda M, Khachik F, Narisawa T, Takasuka N, Yano M. Cancer prevention by natural carotenoids. *Biofactors* 2000; 13: 89-94 [PMID: 11237205]
- 72 Tsushima M, Maoka T, Katsuyama M, Kozuka M, Matsuno

- T, Tokuda H, Nishino H, Iwashima A. Inhibitory effect of natural carotenoids on Epstein-Barr virus activation activity of a tumor promoter in Raji cells. A screening study for antitumor promoters. *Biol Pharm Bull* 1995; **18**: 227-233 [PMID: 7742789]
- 73 Narisawa T, Fukaura Y, Oshima S, Inakuma T, Yano M, Nishino H. Chemoprevention by the oxygenated carotenoid beta-cryptoxanthin of N-methylnitrosourea-induced colon carcinogenesis in F344 rats. *Jpn J Cancer Res* 1999; 90: 1061-1065 [PMID: 10595732]
- 74 Iskandar AR, Liu C, Smith DE, Hu KQ, Choi SW, Ausman LM, Wang XD. β-cryptoxanthin restores nicotine-reduced lung SIRT1 to normal levels and inhibits nicotine-promoted lung tumorigenesis and emphysema in A/J mice. Cancer Prev Res (Phila) 2013; 6: 309-320 [PMID: 23275008 DOI: 10.1158/1940-6207.CAPR-12-0368]
- 75 Iwamoto M, Imai K, Ohta H, Shirouchi B, Sato M. Supplementation of highly concentrated β-cryptoxanthin in a satsuma mandarin beverage improves adipocytokine profiles in obese Japanese women. *Lipids Health Dis* 2012; 11: 52 [PMID: 22584034 DOI: 10.1186/1476-511X-11-52]
- 76 Ministry of Health. Labour and Welfare Japan (2004) The National Nutrition Survey in Japan [in Japanese]. Tokyo, Japan: Dai-ichi shuppan, 2002
- 77 Cumashi A, Ushakova NA, Preobrazhenskaya ME, D' Incecco A, Piccoli A, Totani L, Tinari N, Morozevich GE, Berman AE, Bilan MI, Usov AI, Ustyuzhanina NE, Grachev AA, Sanderson CJ, Kelly M, Rabinovich GA, Iacobelli S, Nifantiev NE. A comparative study of the anti-inflammatory, anticoagulant, antiangiogenic, and antiadhesive activities of nine different fucoidans from brown seaweeds. Glycobiology 2007; 17: 541-552 [PMID: 17296677 DOI: 10.1093/glycob/cwm014]
- 78 Shibata H, Iimuro M, Uchiya N, Kawamori T, Nagaoka M, Ueyama S, Hashimoto S, Yokokura T, Sugimura T, Wakabayashi K. Preventive effects of Cladosiphon fucoidan against Helicobacter pylori infection in Mongolian gerbils. Helicobacter 2003; 8: 59-65 [PMID: 12603617 DOI: 10.1046/j.1523-5378.2003.00124.x]
- 79 Yonekura L, Kobayashi M, Terasaki M, Nagao A. Ketocarotenoids are the major metabolites of dietary lutein and fucoxanthin in mouse tissues. *J Nutr* 2010; **14**0: 1824-1831 [PMID: 20739451 DOI: 10.3945/jn.110.126466]
- 80 Campbell SJ, Bité JS, Burridge TR. Seasonal patterns in the photosynthetic capacity, tissue pigment and nutrient content of different developmental stages of Undaria pinnatifida (Phaeophyta: Laminariales) in port phillip bay, south-eastern Australia. Bot Mar 1999; 42: 231-242 [DOI: 10.1515/BOT.1999.027]
- 81 Terasaki M, Narayan B, Kamogawa H, Nomura M, Stephen NM, Kawagoe C, Hosokowa M, Miyashita K Carotenoid profile of edible Japanese seaweeds: An improved HPLC method for separation of major carotenoids. *J Aquatic Food Prod Tech* 2012; 21: 468-479 [DOI: 10.1080/10498850.2011.610025]
- 82 **Hashimoto** T, Ozaki Y, Mizuno M, Yoshida M, Nishitani Y, Azuma T, Komoto A, Maoka T, Tanino Y, Kanazawa K. Pharmacokinetics of fucoxanthinol in human plasma after the oral administration of kombu extract. *Br J Nutr* 2012; **107**: 1566-1569 [PMID: 21920061 DOI: 10.1017/S0007114511004879]
- 83 Beppu F, Niwano Y, Tsukui T, Hosokawa M, Miyashita K. Single and repeated oral dose toxicity study of fucoxanthin (FX), a marine carotenoid, in mice. J Toxicol Sci 2009; 34: 501-510 [PMID: 19797858 DOI: 10.2131/jts.34.501]
- 84 **Iio K**, Okada Y, Ishikura M. [Single and 13-week oral toxicity study of fucoxanthin oil from microalgae in rats]. *Shokuhin Eiseigaku Zasshi* 2011; **52**: 183-189 [PMID: 21720124 DOI: http://dx.odi.org/10.3358/shokueishi.52.183]
- 85 Hosokawa M, Kudo M, Maeda H, Kohno H, Tanaka T, Miyashita K. Fucoxanthin induces apoptosis and enhances the antiproliferative effect of the PPARgamma ligand, troglitazone, on colon cancer cells. Biochim Biophys Acta 2004;

- **1675**: 113-119 [PMID: 15535974]
- 86 Das SK, Hashimoto T, Kanazawa K. Growth inhibition of human hepatic carcinoma HepG2 cells by fucoxanthin is associated with down-regulation of cyclin D. *Biochim Biophys* Acta 2008; 1780: 743-749 [PMID: 18230364 DOI: 10.1016/ j.bbagen.2008.01.003]
- 87 Kotake-Nara E, Kushiro M, Zhang H, Sugawara T, Miyashita K, Nagao A. Carotenoids affect proliferation of human prostate cancer cells. *J Nutr* 2001; 131: 3303-3306 [PMID: 11739884]
- 88 Zhang Z, Zhang P, Hamada M, Takahashi S, Xing G, Liu J, Sugiura N. Potential chemoprevention effect of dietary fucoxanthin on urinary bladder cancer EJ-1 cell line. *Oncol Rep* 2008; 20: 1099-1103 [PMID: 18949407 DOI: 10.3892/or\_00000115]
- 89 Miyashita K, Nishikawa S, Beppu F, Tsukui T, Abe M, Hoso-kawa M. The allenic carotenoid fucoxanthin, a novel marine nutraceutical from brown seaweeds. *J Sci Food Agric* 2011; 91: 1166-1174 [PMID: 21433011 DOI: 10.1002/jsfa.4353]
- 90 Liu CL, Huang YS, Hosokawa M, Miyashita K, Hu ML. Inhibition of proliferation of a hepatoma cell line by fucoxanthin in relation to cell cycle arrest and enhanced gap junctional intercellular communication. *Chem Biol Interact* 2009; 182: 165-172 [PMID: 19737546 DOI: 10.1016/j.cbi.2009.08.017]
- 91 Liu CL, Chiu YT, Hu ML. Fucoxanthin enhances HO-1 and NQO1 expression in murine hepatic BNL CL.2 cells through activation of the Nrf2/ARE system partially by its pro-oxidant activity. J Agric Food Chem 2011; 59: 11344-11351 [PMID: 21919437 DOI: 10.1021/jf2029785]
- 92 Ye G, Lu Q, Zhao W, Du D, Jin L, Liu Y. Fucoxanthin induces apoptosis in human cervical cancer cell line HeLa via PI3K/Akt pathway. *Tumour Biol* 2014; 35: 11261-11267 [PMID: 25113250 DOI: 10.1007/s13277-014-2337-7]
- 93 Rwigemera A, Mamelona J, Martin LJ. Inhibitory effects of fucoxanthinol on the viability of human breast cancer cell lines MCF-7 and MDA-MB-231 are correlated with modulation of the NF-kappaB pathway. Cell Biol Toxicol 2014; 30: 157-167 [PMID: 24760606 DOI: 10.1007/s10565-014-9277-2]
- 94 Kotake-Nara E, Asai A, Nagao A. Neoxanthin and fucoxanthin induce apoptosis in PC-3 human prostate cancer cells. Cancer Lett 2005; 220: 75-84 [PMID: 15737690 DOI: 10.1016/j.canlet.2004.07.048]
- 95 Tafuku S, Ishikawa C, Yasumoto T, Mori N. Anti-neoplastic effects of fucoxanthin and its deacetylated product, fucoxanthinol, on Burkitt's and Hodgkin's lymphoma cells. Oncol Rep 2012; 28: 1512-1518 [PMID: 22859062 DOI: 10.3892/ or.2012.1947]
- 96 Kim JM, Araki S, Kim DJ, Park CB, Takasuka N, Baba-Toriyama H, Ota T, Nir Z, Khachik F, Shimidzu N, Tanaka Y, Osawa T, Uraji T, Murakoshi M, Nishino H, Tsuda H. Chemopreventive effects of carotenoids and curcumins on mouse colon carcinogenesis after 1,2-dimethylhydrazine initiation. Carcinogenesis 1998; 19: 81-85 [PMID: 9472697 DOI: 10.1093/carcin/19.1.81]
- 97 Terasaki M, Nagao A, Maeda H, Miyashita K, Masuda S. Combined antiproliferative effect of dietary PPARγ suppressing lipids fucoxanthinol and 1α,25-dihydroxyvitamin D3 in human colon cancer cells. (Proceeding of The Japanese Society for Carotenoid Research) Carotenoid Science 2012; 17: 40-43
- 98 Mutoh M, Niho N, Wakabayashi K. Concomitant suppression of hyperlipidemia and intestinal polyp formation by increasing lipoprotein lipase activity in Apc-deficient mice. *Biol Chem* 2006; 387: 381-385 [PMID: 16606335 DOI: 10.1515/BC.2006.051]
- 99 Ueno T, Teraoka N, Takasu S, Nakano K, Takahashi M, Yamamoto M, Fujii G, Komiya M, Yanaka A, Wakabayashi K, Mutoh M. Suppressive effect of pioglitazone, a PPAR gamma ligand, on azoxymethane-induced colon aberrant crypt foci in KK-Ay mice. Asian Pac J Cancer Prev 2012; 13: 4067-4073 [PMID: 23098518]

- 100 Maeda H, Hosokawa M, Sashima T, Funayama K, Miyashita K. Effect of medium-chain triacylglycerols on anti-obesity effect of fucoxanthin. *J Oleo Sci* 2007; **56**: 615-621 [PMID: 17992001 DOI: 10.5650/jos.56.615]
- 101 Abidov M, Ramazanov Z, Seifulla R, Grachev S. The effects of Xanthigen in the weight management of obese premenopausal women with non-alcoholic fatty liver disease and normal liver fat. *Diabetes Obes Metab* 2010; 12: 72-81 [PMID: 19840063 DOI: 10.1111/j.1463-1326.2009.01132.x]
- 102 Penn R, Worthington DJ. Is serum gamma-glutamyltransferase a misleading test? *Br Med J* (Clin Res Ed) 1983; **286**: 531-535 [PMID: 6130816]
- 103 Hori M, Takahashi M, Hiraoka N, Yamaji T, Mutoh M, Ishigamori R, Furuta K, Okusaka T, Shimada K, Kosuge T, Kanai Y, Nakagama H. Association of pancreatic Fatty infiltration with pancreatic ductal adenocarcinoma. Clin Transl Gastroenterol 2014; 5: e53 [PMID: 24622469 DOI: 10.1038/ctg.2014.5]
- 104 Eden G, Archinti M, Furlan F, Murphy R, Degryse B. The urokinase receptor interactome. *Curr Pharm Des* 2011; 17: 1874-1889 [PMID: 21711237 DOI: 10.2174/138161211796718215]
- 105 Teas J, Vena S, Cone DL, Irhimeh M. The consumption of seaweed as a protective factor in the etiology of breast cancer: proof of principle. J Appl Phycol 2013; 25: 771-779 [PMID: 23678231 DOI: 10.1007/s10811-012-9931-0]
- 106 Jänicke F, Prechtl A, Thomssen C, Harbeck N, Meisner C, Untch M, Sweep CG, Selbmann HK, Graeff H, Schmitt M. Randomized adjuvant chemotherapy trial in high-risk, lymph node-negative breast cancer patients identified by urokinase-type plasminogen activator and plasminogen activator inhibitor type 1. J Natl Cancer Inst 2001; 93: 913-920 [PMID: 11416112 DOI: 10.1093/jnci/93.12.913]
- 107 Look MP, van Putten WL, Duffy MJ, Harbeck N, Christensen IJ, Thomssen C, Kates R, Spyratos F, Fernö M, Eppenberger-Castori S, Sweep CG, Ulm K, Peyrat JP, Martin PM, Magdelenat H, Brünner N, Duggan C, Lisboa BW, Bendahl PO, Quillien V, Daver A, Ricolleau G, Meijer-van Gelder ME, Manders P, Fiets WE, Blankenstein MA, Broët P, Romain S, Daxenbichler G, Windbichler G, Cufer T, Borstnar S, Kueng W, Beex LV, Klijn JG, O'Higgins N, Eppenberger U, Jänicke F, Schmitt M, Foekens JA. Pooled analysis of prognostic impact of urokinase-type plasminogen activator and its inhibitor PAI-1 in 8377 breast cancer patients. J Natl Cancer Inst 2002; 94: 116-128 [PMID: 11792750 DOI: 10.1093/jnci/94.2.116]
- 108 Harris L, Fritsche H, Mennel R, Norton L, Ravdin P, Taube S, Somerfield MR, Hayes DF, Bast RC. American Society of Clinical Oncology 2007 update of recommendations for the use of tumor markers in breast cancer. *J Clin Oncol* 2007; 25: 5287-5312 [PMID: 17954709]
- 109 Goodwin TW. The biochemistry of the carotenoids, Vol. I. Plants. 2nd ed. New York, NY: Chapman and Hall, 1980
- 110 Khachik F, Beecher GR, Whittaker NF. Separation, identification, and quantification of the major carotenoid and chlorophyll constituents in extracts of several green vegetables by liquid chromatography. J Agric Food Chem 1986; 34: 603-616 [DOI: 10.1021/jf00070a006]
- 111 Takaichi S, Mimuro M. Distribution and geometric isomerism of neoxanthin in oxygenic phototrophs: 9'-cis, a sole molecular form. *Plant Cell Physiol* 1998; 39: 968-977 [DOI: 10.1093/oxfordjournals.pcp.a029461]
- 112 Ruban AV, Lee PJ, Wentworth M, Young AJ, Horton P. Determination of the stoichiometry and strength of binding of xanthophylls to the photosystem II light harvesting complexes. J Biol Chem 1999; 274: 10458-10465 [PMID: 10187836 DOI: 10.1074/jbc.274.15.10458]
- 113 Seo M, Koshiba T. Complex regulation of ABA biosynthesis in plants. *Trends Plant Sci* 2002; 7: 41-48 [PMID: 11804826]
- 114 Marki-Fischer E, Eugster CH Neoflor and 6-epineoflor from flowers of Trollius europaeus; highfield 1H-NMR spectra of (all-E)-neoxanthin and (9'Z)-neoxanthin. *Helv Chim Acta* 1990; 73: 1637-1643 [DOI: 10.1002/hlca.19900730608]

- 115 **Asai A**, Terasaki M, Nagao A. An epoxide-furanoid rearrangement of spinach neoxanthin occurs in the gastrointestinal tract of mice and in vitro: formation and cytostatic activity of neochrome stereoisomers. *J Nutr* 2004; **134**: 2237-2243 [PMID: 15333710]
- 116 Eugster CH (1995) Chemical derivatization: microscale tests for the presence of common functional groups in carotenoids. In: Carotenoids Vol. 1A: Isolation and Analysis. Britton G, Liaaen-Jensen S, Pfander H, editors. Birkhäuser Verlag, Basel, Switzerland, 1995: 71-80
- 117 Asai A, Yonekura L, Nagao A. Low bioavailability of dietary epoxyxanthophylls in humans. *Br J Nutr* 2008; 100: 273-277 [PMID: 18186952 DOI: 10.1017/S0007114507895468]
- 118 Sugawara T, Kushiro M, Zhang H, Nara E, Ono H, Nagao A. Lysophosphatidylcholine enhances carotenoid uptake from mixed micelles by Caco-2 human intestinal cells. *J Nutr* 2001; 131: 2921-2927 [PMID: 11694619]
- 119 Barua AB, Olson JA. Xanthophyll epoxides, unlike betacarotene monoepoxides, are not detectibly absorbed by humans. J Nutr 2001; 131: 3212-3215 [PMID: 11739868]
- 120 Hashimoto T, Ozaki Y, Taminato M, Das SK, Mizuno M, Yoshimura K, Maoka T, Kanazawa K. The distribution and accumulation of fucoxanthin and its metabolites after oral administration in mice. *Br J Nutr* 2009; **102**: 242-248 [PMID: 19173766 DOI: 10.1017/S0007114508199007]
- 121 **Pérez-Gálvez A**, Martin HD, Sies H, Stahl W. Incorporation of carotenoids from paprika oleoresin into human chylomicrons. *Br J Nutr* 2003; **89**: 787-793 [PMID: 12828795]
- 122 **Kotake-Nara** E, Sugawara T, Nagao A. Antiproliferative effect of neoxanthin and fucoxanthin on culture cells. *Fish Sci* 2005; 71: 459-461. Available from: URL: http://link.springer.com/article/10.1111/j.1444-2906.2005.00986.x#page-1
- 123 Terasaki M, Asai A, Zhang H, Nagao A. A highly polar xanthophyll of 9'-cis-neoxanthin induces apoptosis in HCT116 human colon cancer cells through mitochondrial dysfunction. *Mol Cell Biochem* 2007; 300: 227-237 [PMID: 17186379 DOI: 10.1007/s11010-006-9387-0]
- 124 Ugocsai K, Varga A, Molnár P, Antus S, Molnár J. Effects of selected flavonoids and carotenoids on drug accumulation and apoptosis induction in multidrug-resistant colon cancer cells expressing MDR1/LRP. *In Vivo* 2005; 19: 433-438 [PMID: 15796208]
- 125 Chang JM, Lin JK. Isolation of neoxanthin from spinach and its prevention on lipid peroxidation. J Chin Med 1993; 4: 235-245. Available from: URL: http://tao.wordpedia.com/show\_pdf. ashx?sess=m3o4bi2vfuk2zou5qjretbmz&file\_name=JO00000295\_4-3\_235-245&file\_type=r
- 126 Chang JM, CHen WC, Hong D, Lin JK. The inhibition of DMBA-induced carcinogenesis by neoxanthin in hamster buccal pouch. *Nutr Cancer* 1995; 24: 325-333 [PMID: 8610051 DOI: 10.1080/01635589509514421]
- 127 Murakami A, Nakashima M, Koshiba T, Maoka T, Nishino H, Yano M, Sumida T, Kim OK, Koshimizu K, Ohigashi H. Modifying effects of carotenoids on superoxide and nitric oxide generation from stimulated leukocytes. *Cancer Lett* 2000; 149: 115-123 [PMID: 10737715]
- 128 Okada T, Nakai M, Maeda H, Hosokawa M, Sashima T, Miyashita K. Suppressive effect of neoxanthin on the differentiation of 3T3-L1 adipose cells. J Oleo Sci 2008; 57: 345-351 IPMID: 184694971
- 129 Mayne ST. Beta-carotene, carotenoids, and disease prevention in humans. *FASEB J* 1996; 10: 690-701 [PMID: 8635686]
- 130 Slattery ML, Benson J, Curtin K, Ma KN, Schaeffer D, Potter JD. Carotenoids and colon cancer. Am J Clin Nutr 2000; 71: 575-582 [PMID: 10648274]
- 131 Lutein and zeaxanthin. Monograph. Altern Med Rev 2005; 10: 128-135 [PMID: 15989382]
- 132 **Sommerburg O**, Keunen JE, Bird AC, van Kuijk FJ. Fruits and vegetables that are sources for lutein and zeaxanthin: the macular pigment in human eyes. *Br J Ophthalmol* 1998;

- 82: 907-910 [PMID: 9828775 DOI: 10.1136/bjo.82.8.907]
- 133 Kelly ER, Plat J, Haenen GR, Kijlstra A, Berendschot TT. The effect of modified eggs and an egg-yolk based beverage on serum lutein and zeaxanthin concentrations and macular pigment optical density: results from a randomized trial. *PLoS One* 2014; 9: e92659 [PMID: 24675775 DOI: 10.1371/journal.pone.0092659]
- 134 Landrum JT, Bone RA. Lutein, zeaxanthin, and the macular pigment. *Arch Biochem Biophys* 2001; 385: 28-40 [PMID: 11361022 DOI: 10.1006/abbi.2000.2171]
- 135 Ben Amara N, Tourniaire F, Maraninchi M, Attla N, Amiot-Carlin MJ, Raccah D, Valéro R, Landrier JF, Darmon P. Independent positive association of plasma β-carotene concentrations with adiponectin among non-diabetic obese subjects. Eur J Nutr 2014 [PMID: 24906472]
- 136 Kritchevsky SB. beta-Carotene, carotenoids and the prevention of coronary heart disease. J Nutr 1999; 129: 5-8 [PMID: 9915867]
- 137 Ascherio A, Rimm EB, Hernán MA, Giovannucci E, Kawachi I, Stampfer MJ, Willett WC. Relation of consumption of vitamin E, vitamin C, and carotenoids to risk for stroke among men in the United States. *Ann Intern Med* 1999; 130: 963-970 [PMID: 10383366 DOI: 10.7326/0003-4819-130-12-19 9906150-00003]
- 138 **Michaud DS**, Feskanich D, Rimm EB, Colditz GA, Speizer FE, Willett WC, Giovannucci E. Intake of specific carotenoids and risk of lung cancer in 2 prospective U.S. cohorts. *Am J Clin Nutr* 2000; **72**: 990-997 [PMID: 11010942]
- 139 Le Marchand L, Hankin JH, Bach F, Kolonel LN, Wilkens LR, Stacewicz-Sapuntzakis M, Bowen PE, Beecher GR, Laudon F, Baque P. An ecological study of diet and lung cancer in the South Pacific. *Int J Cancer* 1995; 63: 18-23 [PMID: 7558446]
- 140 Okuyama Y, Ozasa K, Oki K, Nishino H, Fujimoto S, Watanabe Y. Inverse associations between serum concentrations of zeaxanthin and other carotenoids and colorectal neoplasm in Japanese. *Int J Clin Oncol* 2014; 19: 87-97 [PMID: 23380957 DOI: 10.1007/s10147-013-0520-2]
- 141 **Tsubono Y**, Tsugane S, Gey KF. Plasma antioxidant vitamins and carotenoids in five Japanese populations with varied mortality from gastric cancer. *Nutr Cancer* 1999; 34: 56-61 [PMID: 10453442]
- 142 Stahl W, Heinrich U, Jungmann H, Sies H, Tronnier H. Carotenoids and carotenoids plus vitamin E protect against ultraviolet light-induced erythema in humans. *Am J Clin Nutr* 2000; 71: 795-798 [PMID: 10702175]
- 143 Dorgan JF, Sowell A, Swanson CA, Potischman N, Miller R, Schussler N, Stephenson HE. Relationships of serum carotenoids, retinol, alpha-tocopherol, and selenium with breast cancer risk: results from a prospective study in Columbia, Missouri (United States) Cancer Causes Control 1998; 9: 89-97 [PMID: 9486468]
- 144 Zhang S, Tang G, Russell RM, Mayzel KA, Stampfer MJ, Willett WC, Hunter DJ. Measurement of retinoids and carotenoids in breast adipose tissue and a comparison of concentrations in breast cancer cases and control subjects. Am J Clin Nutr 1997; 66: 626-632 [PMID: 9280184]
- 145 Yeum KJ, Ahn SH, Rupp de Paiva SA, Lee-Kim YC, Krinsky NI, Russell RM. Correlation between carotenoid concentrations in serum and normal breast adipose tissue of women with benign breast tumor or breast cancer. *J Nutr* 1998; 128: 1920-1926 [PMID: 9808643]
- 146 **Toniolo P**, Van Kappel AL, Akhmedkhanov A, Ferrari P, Kato I, Shore RE, Riboli E. Serum carotenoids and breast cancer. *Am J Epidemiol* 2001; **153**: 1142-1147 [PMID: 11415946 DOI: 10.1093/aje/153.12.1142]
- 147 Bravi F, Bosetti C, Filomeno M, Levi F, Garavello W, Galimberti S, Negri E, La Vecchia C. Foods, nutrients and the risk of oral and pharyngeal cancer. Br J Cancer 2013; 109: 2904-2910 [PMID: 24149181 DOI: 10.1038/bjc.2013.667]

- 148 Dagnelie G, Zorge IS, McDonald TM. Lutein improves visual function in some patients with retinal degeneration: a pilot study via the Internet. *Optometry* 2000; 71: 147-164 [PMID: 10970259]
- 149 Granado F, Olmedilla B, Gil-Martínez E, Blanco I. Lutein ester in serum after lutein supplementation in human subjects. Br J Nutr 1998; 80: 445-449 [PMID: 9924266 DOI: 10.1017/S0007114598001512]
- 150 **González de Mejía** E, Ramos-Gómez M, Loarca-Piña G. Antimutagenic activity of natural xanthophylls against aflatoxin B1 in Salmonella typhimurium. *Environ Mol Mutagen* 1997; 30: 346-353 [PMID: 9366914 DOI: 10.1002/(SICI)1098-2 280(1997)30: 3<346: AID-EM14>3.0.CO; 2-D]
- 151 Kawashima T. A marine carotenoid, fucoxanthin, induces regulatory T cells and inhibits Th17 cell differentiation in vitro. *Biosci Biotechnol Biochem* 2011; 75: 2066-2069 [PMID: 21979096]
- 152 Iannone A, Rota C, Bergamini S, Tomasi A, Canfield LM. Antioxidant activity of carotenoids: An electron-spin resonance study on β-carotene and lutein interaction with free radicals generated in a chemical system. J Biochem Mol Toxicol 1998; 1: 299-304 [PMID: 9664236 DOI: 10.1002/(SICI)1099-0461(1998)12:5<299::AID-JBT6>3.0.CO;2-G]
- 153 Sujak A, Gabrielska J, Grudzinski W, Borc R, Mazurek P, Gruszecki WI. Lutein and zeaxanthin as protectors of lipid membranes against oxidative damage: The structural aspects. Arch Biochem Biophys 1999; 371: 301-307 [PMID: 10545218 DOI: 10.1006/abbi.1999.1437]
- 154 **Stahl W**, Junghans A, de Boer B, Driomina ES, Briviba K, Sies H. Carotenoid mixtures protect multilamellar li-

- posomes against oxidative damage: Synergistic effects of lycopene and lutein. *FEBS Lett* 1998; **427**: 305-308 [PMID: 9607334 DOI: 10.1016/S0014-5793(98)00434-7]
- 155 Rafi MM, Kanakasabai S, Gokam SV, Krueger EG, Bright JJ. Dietary Lutein Modulates Growth and Survival Genes in Prostate Cancer Cells. *J Med Food* 2014 Aug 27; Epub ahead of print [PMID: 25162762 DOI: 10.1089/jmf.2014.0003]
- 156 Pei YX, Heng ZC, Duan GC, Wang MC. [The mechanisms and effects of lutein on inducing the cell differentiation of human esophagus cancer EC9706]. Sichuan Daxue Xuebao Yixueban 2007; 38: 629-632 [PMID: 17718427]
- 157 Sumantran VN, Zhang R, Lee DS, Wicha MS. Differential regulation of apoptosis in normal versus transformed mammary epithelium by lutein and retinoic acid. *Cancer Epidemiol Biomarkers Prev* 2000; 9: 257-263 [PMID: 10750663]
- 158 Li Z, Wang Y, Mo B. [The effects of carotenoids on the proliferation of human breast cancer cell and gene expression of bcl-2]. *Zhonghua Yufangyixue Zazhi* 2002; **36**: 254-257 [PMID: 12411207]
- 159 Park JS, Chew BP, Wong TS, Zhang JX, Magnuson NS. Dietary lutein but not astaxanthin or beta-carotene increases pim-1 gene expression in murine lymphocytes. *Nutr Cancer* 1999; 33: 206-212 [PMID: 10368818 DOI: 10.1207/S15327914NC330214]
- 160 Gonzalez de Mejia E, Loarca-Pina G, Ramos-Gomez M. Antimutagenicity of xanthophylls present in Aztec Marigold (Tagetes erecta) against 1-nitropyrene. Mutat Res 1997; 389: 219-226 [PMID: 9093387 DOI: 10.1016/S1383-5718(96)00151-9]
- 161 Park JS, Chew BP, Wong TS. Dietary lutein from marigold extract inhibits mammary tumor development in BALB/c mice. J Nutr 1998; 128: 1650-1656 [PMID: 9772131]

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Treatment strategy of diminutive colorectal polyp <5 mm in size - Should it be removed and discarded without pathologic assessment?

## Endoscopic features and management of diminutive colorectal submucosal invasive carcinoma

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Background and Aim: The vast majority of diminutive (~5 mm) colorectal tumors consist of a very low prevalence of advanced neoplasia, and a predict-resect-and-discard policy has been proposed recently in Western countries. The histology of some diminutive colorectal tumors reveals carcinoma, not adenoma, although the frequency is relatively low. Clarifying the endoscopic features of diminutive submucosal invasive colorectal carcinoma (CRC) during colonoscopy is important for managing diminutive lesions.

Methods: A total of 111 cases of submucosal invasive CRC ≤10 mm were analyzed. The incidence of submucosal invasion in early CRC per gross type, size, location, pit pattern diagnosis, and rate of lymph node (LN) metastasis was evaluated.

Results: In diminutive tumors, the overall submucosal invasion rate in early CRC was 9.6%; however, depressed tumors had a significantly higher frequency of submucosal invasion than protruded or flat elevated tumors. There were no significant differences in the distribution of submucosal invasive CRC between the diminutive tumors and those that were 6-10 mm. The pit pattern diagnosis of diminutive submucosal invasive CRC was type V<sub>1</sub> pit pattern in all cases. Each case of submucosal invasive CRC was completely resected by en bloc endoscopic resection, and there were no cases of LN metastasis.

Conclusion: Diminutive tumors with depression have a high frequency of submucosal invasive CRC and an initial indication for endoscopic resection.

Key words: colon and rectum, diminutive tumor, endoscopic feature, pit pattern, submucosal invasive carcinoma

#### INTRODUCTION

IMINUTIVE (~5 MM) COLORECTAL lesions are commonly encountered during colonoscopy. The vast majority of diminutive colorectal lesions have a very low prevalence of advanced neoplasia. 1,2 In addition, advanced endoscopic imaging, such as pit pattern diagnosis and narrow band imaging, has made it possible to differentiate between non-neoplastic and neoplastic, as well as between non-invasive and invasive neoplastic, lesions.3-6 Recently, in Western countries, a predict-resect-and-discard policy has been proposed in order to eliminate the pathological examination.7-9 In Japan, diminutive sessile colorectal tumors, which are confirmed as low-grade adenoma by detailed endoscopic observations, are not currently an indication for endoscopic resection (ER), because tumor size is generally unchanging during surveillance colonoscopy. 10

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However, the natural history and malignant potential of diminutive colorectal tumors have not been clarified.

Some diminutive colorectal tumors have submucosal invasion, although the frequency is extremely low.<sup>1,2</sup> In particular, submucosal invasive colorectal carcinoma (CRC) has the possibility of lymph node (LN) metastasis regardless of size. 11,12 To confirm a precise pathological diagnosis, it is necessary to completely resect the tumor with negative horizontal and vertical margins. Therefore, the use of a hot biopsy or a resect-and-discard strategy should be prohibited for diminutive colorectal tumors that are suspected to have submucosal invasion from endoscopic findings. In the present study, to clarify the indications for ER, we analyzed the endoscopic features of diminutive submucosal invasive CRC detected in our institution.

#### **METHODS**

THE PRESENT STUDY involved 12 521 colorectal L tumors (11 771 adenomas and 750 early CRC)  $\leq$ 10 mm that had been consecutively resected endoscopically or surgically at Hiroshima University Hospital between January