

Figure 2. Secretion by activated $y\delta$ cells of multiple cytokines and their physiological roles.

Applications

γδ T-cell based cancer immunotherapy

In 1890, Paul Ehrlich proposed vaccines against cancer, which was the first suggestion using the immune system to cope with cancer. In fact, immune deficiency states, such as iatrogenic immune suppression, common variable immunodeficiency (CVID), severe combined immunodeficiency (SCID), and acquired immunodeficiency syndrome (AIDS), make patients more susceptible to various kinds of malignancies [117, 118]. The importance of immune surveillance against tumor emergence and progression was reinforced with the observation of different immune deficiency states. Targeting the immune system to combat tumors is a promising therapeutic strategy [119] although progress has been slow and success is limited.

Immunotherapeutic approaches for anti-tumor responses via stimulating the adaptive immune system rely on major histocompatibility complex (MHC)–restricted $\alpha\beta$ T cells. Although significant advances in the adaptive immunotherapies toward tumors have been made and vaccine-based strategies have been improved, $\alpha\beta$ T cell-mediated immunotherapies have met with limited success. Durable responses are scarce, and active immunotherapy has not achieved an established treatment modality. $\alpha\beta$ T cells require specific tumor-associated antigens (TAAs)

and suitable costimulatory molecules for activation. Once failure or loss of TAAs, MHC molecules, and/or costimulatory molecules occurs, it will render tumor cells less susceptible to $\alpha\beta$ T-cell-mediated cytotoxicity or induce anergy in specific $\alpha\beta$ T cells.

γδ T cells are considered to be a member of innate immune cells and exhibit an important role in immune-surveillance and immune defense against tumors, including melanomas [120], leukemias, lymphomas, neuroblastomas, and other types of carcinomas [102, 104, 121]. The antitumor activity of γδ T cells has been confirmed by expanding them $ex\ vivo$ followed by infusion to cancer patients [122, 123]. Recently, in vitro-activated γδ T cells have been shown to target a small number of colon cancer stem cells, which had been demonstrated to be attributable to the failure of conventional therapies. In addition, γδ T cells can kill chemotherapy (imatinib)-resistant chronic myelogenous leukemia lines.

 $\gamma\delta$ T cells can be utilized for antitumor therapies in an unconventional manner [64], because they exhibit a potent MHC-unrestricted lytic activity against a wide variety of tumor cells *in vitro*. In clinical studies, $\gamma\delta$ T cells have been shown to infiltrate into different kinds of tumors, including lung carcinomas [124], renal cell carcinomas [125], and breast carcinomas [126]. It is worthy of note that they have exhibited to specifically respond *in vitro* to tumors but not to normal cells. Moreover, a high level of $\gamma\delta$ T cells was found in disease-free survivors of acute leukemia

following allogeneic bone marrow transplantation [127]. Clinical evidence in the therapy of human malignancies using $\gamma\delta$ T cells stimulated with phosphoantigens or bisphosphonates [128] will further support the antitumor effects of this cell population *in vivo*.

Human $\gamma\delta$ T cells mediate anti-tumor immunity via several distinct pathways, such as the secretion of proinflammatory cytokines and pro-apoptotic molecules, and cell to cell contact-dependent lysis through an NK-like pathway or a TCR-dependent pathway [129]. Activated $\gamma\delta$ T cells secrete large amounts of cytokines, which act on tumor cells or their microenvironment [130]. IFN- γ , a major cytokine secreted by activated $\gamma\delta$ T cells, has multiple anti-tumor effects including direct inhibition of tumor growth, blocking angiogenesis, or stimulation of macrophages. IFN- γ , therefore, is a crucial cytokine in the $\gamma\delta$ T cell-mediated anti-tumor responses.

Many reports demonstrate that γδ T cell clones derived from either tumor sites or blood of cancer patients exhibit potent cytotoxicity against tumor cells ex vivo [20]. In addition, γδ T cells have a unique capacity to present antigens to both CD8 and CD4 cells and potentially elicit strong adaptive anti-tumor responses [131]. Furthermore, γδ T cells can respond to stress stimuli originated from transformed cells and exhibit MHC-unrestricted antigen recognition. Activated γδ T cells can exert an immediate and robust direct cytotoxic effect, and simultaneously induce secondary inflammation which attracts tumor specific effector T cells [132]. Adoptive transfer of large number of activated γδ T cells, either alone or along with CD8+ tumor specific T cells, therefore, can potentially boost the dysfunctional host immune system in many ways.

Several clinical trials including patients with advanced diseases refractory to conventional treatments have been performed to test the safety and efficacy of $\gamma\delta$ T-cell-based immunotherapy. Consequently, up to 50% efficacy has been observed in a most recent clinical trial using phosphostimTM for expanding the autologous $\gamma\delta$ T cells in patients with metastatic renal cell carcinomas [133].

Dokouhaki and his colleagues [134] have built a xenograft model of human non-small cell lung cancer (NSCLC) to study the *in vivo* function of $\gamma\delta$ T cells, and found that the lung cancer progression was remarkably inhibited after the infusion of $\gamma\delta$ T cells in the absence of cytokine co-administration. The results

indicate that the interaction between NKG2D and its ligand (s) may partially contribute to the anti-tumor effect of human $\gamma\delta$ T cells. Treatment with expanded and activated $\gamma\delta$ T cells for NSCLC is especially promising. Firstly, $\gamma\delta$ T cells reside in lung tissues and the number is increased in patients with NSCLC. Secondly, $\gamma\delta$ T cells can recognize NSCLC cells in spite of the loss of HLA expression found in 69% of the cases. Thirdly, infusion of *ex vivo* expanded $\gamma\delta$ T cells is feasible in patients with NSCLC [135].

To examine the anti-tumor activity of $\gamma\delta$ T cells in breast cancer, a phase I trial was conducted, in which Zol and low-dose IL-2 were administered to 10 advanced metastatic breast cancer patients who were therapeutically terminal [136]. The results suggest that the treatment was well tolerated and promoted the effector maturation of $V\gamma 9V\delta 2$ T cells in all patients. It is noteworthy that there was a statistically significant linkage between clinical outcome and the number of peripheral Vy9V82 T cells, as seven patients who failed to sustain Vy9V82 T cells showed progressive clinical deterioration, while three patients who sustained robust peripheral Vγ9Vδ2 cell populations displayed one instance of partial remission and two of stable disease, respectively. Consistent with the earlier clinical trial in prostate cancer, there was a strong correlation of Vγ9Vδ2 T cell status with reduced carcinoma progression, and Zol plus low-dose IL-2 provided a novel, safe, and feasible strategy to treat refractory patients with advanced breast cancer [136]. It has been reported that the treatment of patients with Vγ9Vδ2 T cells plus Zol could efficiently enhance the lysis of MCF-7 breast tumor cells and PC-3 prostate carcinoma cells in a γδ TCR-dependent manner at the effector to target ratios of 30:1 to 7.5:1 [126].

Recently, Kobayashi and his colleagues have reported a complete remission (CR) in a patient with advanced renal cell carcinomas who underwent 6 monthly cycles of autologous $\gamma\delta$ T cell therapy, in which the activation and expansion of $\gamma\delta$ T cells were performed *in vitro* using 2-methyl-3-butenyl diphosphate plus IL-2, followed by the infusion of the expanded $\gamma\delta$ T cells, Zol (4 mg) plus low-dose IL-2 (1.4×106 IU) [137]. The clinical responses were associated with a sharp increase in the number of IFN- γ -producing adoptively transferred V γ 9V δ 2 T cells. The CR patient has been disease free for more than 3 years without any additional treatment. Some important clinical trials on $\gamma\delta$ T-cell-based cancer immunotherapy are summarized in Table 1.

Table 1. Examples of the important clinical assays about γδ T-cell-based cancer immunotherapy.

Cancer Immunotherapy	Cancer Type	Results	Ref.
Treated with zoledronate+IL-2	Malignant melanomas	$\gamma\delta$ T cells and the cancer's stage are negatively correlated, result in target cell lysis and death	[120]
Treated with zoledronate+IL-2	Hormone-refractory prostate cancer	Partial remission and stable disease, aggregate increases in $\gamma\delta$ T cell numbers	[66]
Pretreated with pamidronate and zoledronate	MCF-7 breast tumor	Tumor cells were efficiently lysed, depended on the perforin-granzyme pathway.	[126]
Bone marrow grafts depleted of $\alpha\beta$ T cells	Acute lymphoblastic leukemia (ALL), Acute myeloid leukemia (AML)	Have a significant improvement in disease-free survival, a post-BMT absolute increase in $\gamma\delta$ T cells was significantly associated with $\alpha\beta$ T-cell depletion.	[127]
Autologous $\gamma\deltaT$ cells infusion alone or with IL-2	Renal cell carcinoma (metastatic)	The maximum-tolerated dose and safety of $\gamma\delta$ T cells is $8{\times}10^{9}$ cells.	[128]
Inoculation without cytokines	Non-small cell lung cancer	Progression is remarkablely inhibited.	[134]
In vitro proliferation of γδ T cells with pamidronate/IL-2	Non-Hodgkin lymphoma (relapsed and/or refractory) or multiple myeloma	Pamidronate/IL-2 was well tolerated, and no dose-limiting toxicity was observed.	
Autologous γδ T cells	Colon carcinoma	Recognition and efficient killing of autologous and allogeneic CCCL (Colon Carcinoma Cell Lines)	[139]
Treated with zoledronic acid	Pancreatic carcinoma	Tumor cells treated with zoledronic acid are more vulnerable against $\gamma\delta$ T cell attack.	[140]

Applications of $\gamma\delta$ T cells to treatment of patients with infectious diseases

Infectious diseases are a serious threat to human health and gradually increase in global morbidity and mortality in recent years. Current strategies to control infections mainly focus on the pathogens themselves, but neglect the host factors which may regulate the progression of the diseases. The frequent appearance of drug resistance in infectious pathogens often leads to costly but ineffective therapy. In addition, the efficiency of vaccines inducing adaptive immune responses could be impaired by the rapid immune evasion of pathogens through their frequent mutations. In conclusion, innate immune cells that recognize the conserved structural components of pathogens and elicit rapid responses against infections have great potential in anti-infection immunotherapy.

Human $\gamma\delta$ T cells are vital components of the innate immune system and play important roles in the early responses to invasive pathogens. Besides, some $\gamma\delta$ T cells, such as IL-17-producing $\gamma\delta$ T cells, have been demonstrated to be involved in the pathogenesis of transplantation rejection, autoimmune disease [141], inflammatory diseases [142] and allergy [143] in humans. The quality and quantity of human $\gamma\delta$ T cells with dynamic variation affects the initiation, progression and prognosis of infectious diseases.

Viral infections

Although the mechanism underlying $\gamma\delta$ T cell-mediated immune responses against viruses remains to be delineated, their protective roles have been confirmed in some acute and chronic viral infec-

tions. The activation and cytokine secretions of $\gamma\delta$ T cells are considered as indicators of early viral infection.

Recently, the beneficial effects of human $V_{\gamma}9V\delta2$ T cells against influenza virus infection have been re-ported. Vy9Vδ2 T cells can control infection of several strains of influenza viruses, such as pandemic H1N1, human seasonal H1N1, and the avian H5N1 and H9N2 viruses [144]. A study has suggested that Vγ9Vδ2 T cells expressed both type 1 cytokines and chemokine receptors during influenza virus infection, and IPP-activated cells had a higher capacity to produce IFN-y [145]. It is worth noting that IPP-activated γδ T cells also can inhibit seasonal and pandemic H1N1 viruses in a noncytolytic manner, mainly through IFN-y production [145]. Avian H5N1 and H9N2 viruses can induce higher CCL5 production in Vγ9Vδ2 T cells, which may mediate the migration of Vγ9Vδ2 T cells toward influenza virus-infected cells [145].

In addition, Pam activated human V γ 9V δ 2 T cells could kill influenza virus-infected cells and suppress viral replication *in vitro*. Tu et al. [146] demonstrated that Pam-expanded V γ 9V δ 2 T cells by themselves can control influenza virus infection effectively *in vivo*. Regarding H1N1 viruses, the inhibition of the virus replication by the V γ 9V δ 2 T cells may rely on direct killing of virus-infected cells and secretion of IFN- γ , but, for H5N1 infection, mainly by direct killing.

The antiviral mechanisms elicited by $V\gamma 9V\delta 2$ T cells during different viral infections are diverse. For example, human $V\gamma 9V\delta 2$ T cells can kill Epstein-Barr virus (EBV)-, and herpes simplex virus

(HSV)-infected target cells in an HLA-unrestricted manner in vitro [147]. IPP-activated Vγ9Vδ2 T cells kill human immunodeficiency (HIV)-infected target cells and inhibit viral replication by releasing certain CCR5 ligand chemokines to block the HIV entry co-receptor CCR5. Furthermore, phosphoantigen-activated Vγ9Vδ2 T cells can induce noncytolytic inhibition of hepatitis C virus (HCV) replication by IFN-y secretion [148]. It was revealed that simian $y\delta$ T cells could produce β -chemokines, as macrophage inflammatory protein-1a [MIP1- α], MIP1- β , and RANTES. These factors are known to block virus attachment to the CCR5 co-receptor [149], thus preventing SIV infection.

Furthermore, γδ T cells can induce the maturation of dendritic cells (DCs) to promote the establishment of protective adaptive immunity against West Nile virus [150]. The protective roles of $\gamma\delta$ T cells have also been confirmed in some chronic infectious diseases. During human cytomegalovirus (HCMV) infection [151], V82- T cells, a minor population of peripheral blood γδ T cells, have been found to expand significantly, showing a potent 'virus-specific' cytotoxicity and increased elimination of pathogens [152]. Similarly, yo T cells in HIV-infected patients have been found to exhibit beneficial roles in controlling HIV infection [153] through their cell-lytic functions and cytokine secretions. γδ T cells are potent effectors in antibody-directed cell cytotoxicity [154], which is important for HIV inhibition [155], although the quantity and quality of γδ T cells are generally decreased with the advancement of HIV infection [156]. γδ T cells also help control the infection caused by Epstein-Barr virus [157] and human hepatitis virus C [158].

More recently, Yin's group found negative correlation between the ratios of V δ 2 T cells to Th17 (IL-17-producing CD4+ T) cells and liver damage in HBV-infected immune-activated patients and provided experimental evidence that V δ 2 T cells suppressed Th17 cytokine production through cell contact-dependent and IFN- γ -dependent mechanisms [116].

The applications of $\gamma\delta$ T cells in the antiviral immunity are summarized in Table 2.

Bacterial infections

The discovery that $\gamma\delta$ T cells expanded in the peripheral blood of patients with bacterial infections raised the possibility that the T cell subset can be utilized for the control of bacterial infections. Mounting evidence indicates that $\gamma\delta$ T cells are of importance in human bacterial infections. Human $\gamma\delta$ T cells can recognize HMBPP derived from various bacteria and provoke adaptive immunity in various ways. They

expand during bacterial infections such as tuberculosis (mean, 14%), salmonellosis (mean, 18%), tularemia (mean, 31%), brucellosis (mean, 29%), listeriosis (mean, 12%), and ehrlichiosis (mean, 57%). Activated and expanded $V\gamma 2V\delta 2^+$ T cells might directly participate in antimicrobial immune responses. They recognize HMBPP in a TCR-dependent, MHC-, and CD1-unrestricted manner [6], then kill bacteria-infected cells and bacteria.

Tabel 2. Applications of $\gamma \delta T$ -cell in the antiviral immunity.

Virus Type	The Functions of γδ T Cells	Ref.
Influenza virus	Cytotoxic and noncytolytic antiviral activities	[144]
Simian immunodeficiency virus (SIV)	Produce β-chemokines, block virus attachment to the CCR5 co-receptor	[149]
West Nile virus	Induce the maturation of dendritic cells (DCs)	[150]
Human Cytomegalovirus (HCMV)	Have 'virus-specific' cytotoxicity	[151]
Human immunodeficiency virus (HIV)	Cell-lysis and cytokine secretion	[153]
Hepatitis C virus (HCV)	Mediate non-(MHC)-restricted killing of primary hepatocytes, produce Th1-like cytokine	[158]

Pontiac fever-like disease, which is caused by Legionella micdadei, was found to be related to a significant and long-lasting expansion of Vγ9Vδ2 T cells, implying that the subset may also be pathophysiologically important in a mild and transient form of intracellular bacterial diseases. Surprisingly, patients with the Pontiac fever-like disease showed an early depletion of Vγ9Vδ2 T cells from the circulation, followed by a sharp increase and subsequently, a slow decline over the next 6 months [159]. The ability of the γδ T cells to secrete IFN-γ and TNF-α seemed to be down-regulated after the acute phase of the disease. These results support the assumption that $Vy9V\delta2$ T cells are pathophysiologically important in intracellular bacterial infections, including a mild and transient condition such as Pontiac fever.

Moreover, some intracellular bacterial pathogens, such as *Mycobacterium tuberculosis*, can specifically expand and activate $V\gamma 9V\delta 2$ $\gamma\delta$ T cells by inducing the production of metabolites (e.g., IPP) in infected cells, which strongly demonstrates that $\gamma\delta$ T cells are crucially important in infection control [33]. Consistent with this finding, the suppression of $\gamma\delta$ T cells by chronic tuberculosis infection can contribute to a disastrous outcome.

Other pathogen infections

 $\gamma\delta$ T cells not only play protective roles in viral and bacterial infections, but also control infections by protozoas such as *Leishmania* [160] and *Toxoplasma gondii* [161], whereas the $\gamma\delta$ T cell-mediated inflammation may cause some unwanted destruction of surrounding tissues. Similarly, the protective roles of $\gamma\delta$ T cells during malaria infection have been confirmed in several independent studies [162].

Applications of $\gamma \delta T$ -cell in autoimmune diseases

In recent years, $\gamma\delta$ T cells have been shown roles in the pathogenesis of autoimmune diseases such as rheumatoid arthritis (RA) and systemic lupus erythematosus (SLE). $\gamma\delta$ T cells, by bridging innate and adaptive immunity, may display different functions similar to those of CD4⁺ T-cell subsets such as CTLs, Th1/Th2 cells, Tregs, Th17 cells, and APCs depending on specific microenvironment [163].

Rheumatoid arthritis (RA) is an autoimmune disease that primarily affects the limbs, but the pathogenic mechanism is still unclear. Research has demonstrated that $\gamma\delta$ T cells can function as antigen-presenting cells and are related with rheumatoid arthritis development. During the development of rheumatoid arthritis, $\gamma\delta$ T cells can aggravate immune dysfunction and produce abnormal immune damage by the secretion of cytokines (such as IFN- γ and IL-17) and induction of inflammatory cells to participate in synergistic inflammatory responses [164].

Systemic lupus erythematosus (SLE) is a common autoimmune disease with severe dysregulation of the immune system. Research suggested that the CD27+CD45RA- γδ T cells (a subset constitution of the peripheral blood γδ T cells) were significantly decreased in SLE patients and the numbers of CD27+CD45RA- γδ T cells was negatively correlated with the SLE disease activity. In inactive SLE patients following glucocorticosteroid and cyclophosphamide treatment, Vδ1 cells were significantly increased. This research group also suggested that CD27+CD25high Vδ1 T cells had immunoregulatory activities through cell-to-cell contact, which could express Foxp3 similar to CD4+Foxp3+. These regulatory γδ T cells decreased in the peripheral blood of active SLE patients could be generated in vitro under the stimulation with anti-TCR $y\delta$ in the presence of TGF- β and IL-2 [165]. This finding provides a theoretical basis and feasibility for employing γδ Tregs as a potential therapeutic target in autoimmune disease immunotherapy.

In addition, in an experimental autoimmune encephalomyelitis (EAE) model of the human CNS autoimmune disease multiple sclerosis, $\gamma\delta$ T cells had been shown to regulate CNS inflammation and pro-

mote disease recovery through Fas/FasL-induced apoptosis of encephalitogenic T cells [166].

Applications of γδT-cell in allergic diseases

Some researches have suggested that $\gamma\delta$ T cells may serve as effectors and immunoregulatory cells in allergic disease. Atopic dermatitis (AD), a chronic relapsing inflammatory disease of the skin, is associated with allergic bronchial asthma. Cairo et al. [167] have observed that the circulating V γ 9V δ 2 T cells were significantly increased in AD patients, which is positively correlated between their expansion and the severity of the disease.

Zhang et al. demonstrated that γδ T cells play a proinflammatory role in the development of ovalbumin-induced allergic airway inflammation [168]. Svensson et al. indicated that $\gamma\delta$ T cells promote allergic airway inflammation by enhancing the systemic IgE response and local antibody reactivity without a specific role in the shift of the immune response towards Th2 [169]. IL-17+ γδ T cells, belonging to the Vγ4 subset, have been recently shown to downmodulate central features of allergic reaction in airway inflammation, including Th2 response and lung eosinophilia [143]. When activated, γδ T cells are able to produce a number of cytokines and chemokines, with a unique plasticity to produce Th1, Th2, and Th17 cytokines, contributing to the development and regulation of immune responses [170].

Furthermore, Pawankar et al. have observed an important role for the oligoclonally expanded nasal mucosal gamma delta T cells in the pathogenesis of perennial allergic rhinitis (PAR), with the increase of $\delta 1$ T cells and able to produce mainly interleukin such as IL-4, IL-5 and IL-13 [171, 172].

Summary

In the previous paragraphs, we have elaborated on the mechanism of ligand recognition, activation, cytokine secretion, and applications of $\gamma\delta$ T cells. To be understood better, we summarized the above-mentioned contents in Fig 3.

Obstacles

Clinical applications mentioned above provide enormous opportunities for accelerating the establishment of novel approaches to disease treatment and control, slowing disease progression, reducing comorbidities, and reducing or modifying requirements for antiretroviral therapy. People, however, envisage that some specific obstacles have to be overcome for the development of $\gamma\delta$ T cell immunotherapies.

Firstly, clinical trials have demonstrated that repeated administration of phosphoantigens might

cause anergy, exhaustion or even death of effector γδ T cells. For instance, a macaque study showed that responses occurred after BrHPP/IL-2 injections compared to the first treatment [173]. A similar pattern of declining responses was reported for Zol and IL-2 treatment in prostate cancer patients [66]. Thus, therapies targeting γδ T cells produce short-term responses for a long-term, chronic disease. If we can extend the duration of elevated $y\delta T$ cell levels and functions following treatment, repetitive dosing may not be necessary or the detrimental anergizing effect may not occur. Pauza and his colleagues [22] defined the protocol for yδ T cell activation, including adding immunomodulators such as rapamycin that increase the yield of the cells and potentially modulate the onset of anergy [18].

Secondly, immunostimulatory treatments may elicit significant adverse events (SAEs). For instance, activated $\gamma\delta$ T cells will produce proinflammatory cytokines that may cause SAEs. People, therefore, must minimize or manage the potential consequences of immune reconstitution. These potential obstacles, the risk for SAEs and anergy, have to be addressed by definitively controlled human clinical trials.

Thirdly, although the activation of V γ 2V δ 2 T cells can contribute to the rapid acquisition of APC characteristics ($\gamma\delta$ T-APCs), dominant V γ 2V δ 2 T-cell subset capable of recognizing microbial phosphoantigen exist only in primates. Therefore, current task is

to find an analogue to evaluate the characteristics and clinical potential (including side effects) in murine systems, which may be overcome by the development of a humanized mouse model. This is another current key barrier for the application of $\gamma\delta$ T cells based immunotherapy.

Outlooks

Although great progress has been made in $\gamma\delta$ T cell-based immunothearpies, many aspects need to be improved in future clinical trials.

Regarding tumor cells, it is necessary to explore the adjuvant effect of Toll-like receptor (TLR) stimulation, because in vitro treatment of tumor cells with TLR3 and TLR7 agonists could enhance cytotoxicity of $y\delta$ T cells isolated from cancer patients [174]. As for $y\delta$ T cells, it will be of great importance to evaluate the clinical effects of synthetic TCR agonists such as phosphostim (BrHPP) and picostim (an analog of HMBPP). Another interesting feature is the use of phosphoantigens combined with therapeutic antibodies, as suggested by the improved leukemia and/or lymphoma in vitro killing co-administration of BrHPP and rituximab [175]. In addition, it is important to evaluate the safety of TCR-independent killing strategy by comparing the NKG2D-mediated cytotoxicity against transformed and healthy tissues by NKG2D+ γδ T cells.

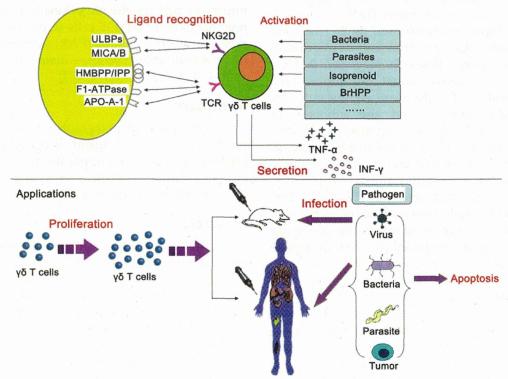


Figure 3. Mechanism underlying $\gamma\delta$ T cell recognition of nonpeptide antigens and clinical applications.

Finally, the identification of biomarkers to predict clinical outcome is crucial for patient selection. A recent study, for example, has identified a panel of 10 genes which encode cell surface proteins that segregated "γδ-susceptible" from "γδ-resistant" hematologic tumors [121]. Equivalent markers could be promptly characterized in multiple cancer types, and their predictive value should be accessed in y8 T cell-based clinical trials. The combination of "susceptible" tumor profiles with improved strategies for γδ T -cell activation *in vivo* may be the way forward for γδ T cell-based cancer immunotherapy.

γδ T cells are attractive targets for cellular immunotherapy, but protocols for their therapeutic use need to be optimized. In addition, it is necessary to explore better antigens which help us stimulate γδ T cell expansion in vitro for the preparation of a large number of cells for adoptive cell transfer. Future studies should focus on the possible advantages of combining v8 T cell-based immunotherapy with conventional chemotherapy or other therapeutic approaches, such as antiangiogenic drugs.

Acknowledgments

We gratefully acknowledge the support by the Program for Zhejiang Leading Team of Science and Technology Innovation (2011R50021), Social Development project of Zhejiang Province (2011C23004) and Zhejiang provincial Natural Science Foundation of China (LY12B02019).

Competing Interests

The authors have declared that no competing interest exists.

References

- Porritt HE, Rumfelt LL, Tabrizifard S, et al. Heterogeneity among DN1 prothymocytes reveals multiple progenitors with different capacities to generate T cell and non-T cell lineages. Immunity. 2004; 20: 735–745. Allman D, Sambandam A, Kim S, et al. Thymopoiesis independent of common
- lymphoid progenitors. Nat Immunol. 2003; 4: 168–174.
- Taghon T, Yui MA, Pant R, et al. Developmental and molecular characterization of emerging beta- and gammadelta-selected pre-T cells in the adult mouse thymus. Immunity. 2006; 24: 53-64.
- Jabeen R, Chang HC, Goswami R, et al. The transcription factor PU.1 regulates γδ T cell homeostasis. PLoS One. 2011; 6: e22189.
- Morita CT, Lee HK, Leslie DS, et al. Recognition of nonpeptide prenyl pyrophosphate antigens by human gammadelta T cells. Microbes Infect. 1999; 1:
- Tanaka Y, Morita CT, Tanaka Y, et al. Natural and synthetic non-peptide antigens recognized by human gamma delta T cells. Nature. 1995; 375:
- Bauer S, Groh V, Wu J, et al. Activation of NK cells and T cells by NKG2D, a receptor for stress-inducible MICA. Science. 1999; 285: 727–729.
- Bukowski JF, Morita CT, Brenner MB. Human gamma delta T cells recognize alkylamines derived from microbes, edible plants, and tea: implications for innate immunity. Immunity. 1999; 11: 57-65
- Hayday AC. γδ T cells: a right time and a right place for a conserved third way of protection. Annu Rev Immunol. 2000; 18: 975-1026.
- Petermann F, Rothhammer V, Claussen MC, et al. $\gamma\delta$ T cells enhance autoimmunity by restraining regulatory T cell responses via an interleukin-23-dependent mechanism. Immunity. 2010; 33: 351–363. Kabelitz D, Glatzel A, Wesch D. Antigen recognition by human $\gamma\delta$ T lym-
- phocytes. International Archives of Allergy and Immunology. 2000; 122: 1-7.

- 12. Groh V, Steinle A, Bauer S, et al. Recognition of stress-induced MHC molecules by intestinal epithelial gammadelta T cells. Science. 1998; 279: 1737–1740.
- Kapp JA, Kapp LM, McKenna KC, et al. gammadelta T-cell clones from intestinal intraepithelial lymphocytes inhibit development of CTL responses ex vivo. Immunology. 2004; 111: 155-164.
- Gould DS, Ploegh HL, Schust DJ. Murine female reproductive tract intraepithelial lymphocytes display selection characteristics distinct from both peripheral and other mucosal T cells. J Reprod Immunol. 2001; 52: 85-99.
- Poles MA, Barsoum S, Yu W, et al. Human immunodeficiency virus type 1 induces persistent changes in mucosal and blood gammadelta T cells despite suppressive therapy. J Virol. 2003; 77: 10456–10467.
- 16. Rakasz E, MacDougall AV, Zayas MT, et al. Gammadelta T cell receptor repertoire in blood and colonic mucosa of rhesus macaques. J Med Primatol.
- Moser B, Eberl M. γδ T-APCs: a novel tool for immunotherapy? Cell Mol Life Sci. 2011; 68(14): 2443-2452.
- Li H, Pauza CD. Rapamycin increases the yield and effector function of human $\gamma\delta$ T cells stimulated in vitro. Cancer Immunol Immunother. 2011; 60: 361–370.
- Mangan BA, Dunne MR, O'Reilly VP, et al. Cutting edge: CD1d restriction and Th1/Th2/Th17 cytokine secretion by human Vδ3 T cells. J Immunol. 2013; 191:
- Kabelitz D, Wesch D, He W. Perspectives of gamma delta T cells in tumor immunology. Cancer Res. 2007; 67: 5-8.
- Cairo C, Armstrong CL, Cummings JS, et al. Impact of age, gender, and race on circulating γδ T cells. Hum Immunol. 2010; 71: 968-975
- Pauza CD, Riedel DJ, Gilliam BL, et al. Targeting $\gamma\delta$ T cells for immunotherapy of HIV disease. Future Virol. 2011; 6: 73-84.
- 23. Hayday AC. Gammadelta T cells and the lymphoid stress-surveillance response. Immunity. 2009; 31: 184-196.
- Caccamo N, Dieli F, Wesch D, et al. Sex-specific phenotypical and functional differences in peripheral human Vgamma9/Vdelta2 T cells. J Leukoc Biol. 2006: 79: 663-666.
- Allison TJ, Winter CC, Fournié JJ, et al. Structure of a human gammadelta T-cell antigen receptor. Nature. 2001; 411: 820-824.
- Morita CT, Beckman EM, Bukowski JF, et al. Direct presentation of nonpeptide prenyl pyrophosphate antigens to human gamma delta T cells. Immunity.
- 27. Scotet E, Martinez LO, Grant E, et al. Tumor recognition following Vgamma9Vdelta2 T cell receptor interactions with a surface F1-ATPase-related structure and apolipoprotein A-1. Immunity. 2005; 22: 71-80.
- Harly C, Guillaume Y, Nedellec S, et al. Key implication of CD277/butyrophilin-3 (BTN3A) in cellular stress sensing by a major human γδ T-cell subset. Blood. 2012; 120: 2269-2279.
- Jr Janeway CA, Jones B, Hayday A. Specificity and function of T cells bearing γδ receptors. Immunol Today. 1988; 9: 73–76.
- Bluestone JA, Khattri R, Sciammas R, et al. TCR gamma delta cells: a specialized T cell subset in the immune system. Annu Rev Cell Dev Biol. 1995; 11:
- Kazen AR, Adams EJ. Evolution of the V, D, and J gene segments used in the primate gammadelta T-cell receptor reveals a dichotomy of conservation and diversity. Proc Natl Acad Sci USA. 2011; 108: E332-340.
- Beagley KW, Husband AJ. Intraepithelial lymphocytes: origins, distribution, and function. Crit Rev Immunol. 1998; 18(3): 237-254.
- Zheng J, Liu Y, Lau YL, et al. γ 6-T cells: an unpolished sword in human anti-infection immunity. Cell Mol Immunol. 2013; 10(1): 50-57.
- 34. Pfeffer K, Schoel B, Gulle H, et al. Primary responses of human T cells to mycobacteria: a frequent set of gamma/delta T cells are stimulated by protease-resistant ligands. Eur J Immunol. 1990; 20: 1175-1179.
- 35. Pfeffer K, Schoel B, Plesnila N, et al. A lectin-binding, protease-resistant mycobacterial ligand specifically activates Vgamma9+ human gamma delta T cells. J lmmunol. 1992; 148: 575-583.
- Porcelli SA, Morita CT, Modlin RL. T-cell recognition of non-peptide antigens. Curr Opin Immunol. 1996; 8: 510-516.
- Burk MR, Mori L, De Libero G. Human V gamma 9-Vdelta 2 cells are stimulated in a cross-reactive fashion by a variety of phosphorylated metabolites. Eur J fmmunol, 1995; 25: 2052-2058.
- Tanaka Y, Morita CT, Nieves E, et al. Natural and synthetic nonpeptide antigens recognized by human gamma/delta T cells. Nature. 1995; 375: 55-58. Wang H, Fang Z, Morita CT. Vgamma2Vdelta2 T Cell Receptor recognition of
- prenyl pyrophosphates is dependent on all CDRs. J Immunol. 2010; 184(11):
- Vavassori S, Kumar A, Wan GS, et al. Butyrophilin 3A1 binds phosphorylated antigens and stimulates human $\gamma\delta$ T cells. Nat Immunol. 2013; 14: 908–916.
- Messal N, Mamessier E, Sylvain A, et al. Differential role for CD277 as a co-regulator of the immune signal in T and NK cells. Eur J Immunol. 2011; 41:
- Wang H, Henry O, Distefano MD, et al. Butyrophilin 3A1 plays an essential role in prenyl pyrophosphate stimulation of human $V\gamma 2V\delta 2$ T cells. J Immunol. 2013; 191: 1029-1042.
- Agea E, Russano A, Bistoni O, et al. Human CD1-restricted T cell recognition of lipids from pollens. J Exp Med. 2005; 202(2): 295-308.
- Dieudé M, Striegl H, Tyznik AJ, et al. Cardiolipin binds to CD1d and stimulates CD1d-restricted $\gamma\delta$ T cells in the normal murine repertoire. J Immunol. 2011; 186(8): 4771-4781.

- 45. Born WK, Kemal Aydintug M, O'Brien RL. Diversity of $\gamma\delta$ T-cell antigens. Cell Mol Immunol. 2013; 10(1): 13–20.
- Willcox CR, Pitard V, Netzer S, et al. Cytomegalovirus and tumor stress surveillance by binding of a human γδ T cell antigen receptor to endothelial protein C receptor. Nat Immunol. 2012; 13: 872–879.
- Rust CJ, Verreck F, Vietor H, et al. Specific recognition of staphylococcal enterotoxin A by human T cells bearing receptors with the V gamma 9 region. Nature, 1990; 346(6284): 572–574.
- Guo Y, Ziegler HK, Safley SA, et al. Human T-cell recognition of Listeria monocytogenes: recognition of listeriolysin O by TcR alpha beta* and TcR gamma delta* T cells. Infect Immun. 1995; 63(6): 2288–2294.
- Champagne E. γδ T cell receptor ligands and modes of antigen recognition. Arch Immunol Ther Exp (Warsz). 2011; 59(2):117–137.
- Zhao Y, Yokota K, Ayada K, et al. Helicobacter pylori heat-shock protein 60 induces interleukin-8 via a Toll-like receptor (TLR)2 and mitogen-activated protein (MAP) kinase pathway in human monocytes. J Med Microbiol. 2007; 56(Pt 2): 154–164.
- Tsan MF, Gao B. Heat shock proteins and immune system. J Leukoc Biol. 2009; 85(6): 905–910.
- Bai L, Picard D, Anderson B, et al. The majority of CD1d-sulfatide-specific T cells in human blood use a semiinvariant V81 TCR. Eur J Immunol. 2012; 42(9):2505–2510.
- Lanca T, Correia DV, Moita CF, et al. The MHC class Ib protein ULBP1 is a nonredundant determinant of leukemia/lymphoma susceptibility to gammadelta T-cell cytotoxicity. Blood. 2010; 115: 2407–2411.
- Kong Y, Cao W, Xi X, et al. The NKG2D ligand ULBP4 binds to TCR gamma9/delta2 and induces cytotoxicity to tumor cells through both TCR gammadelta and NKG2D. Blood. 2009; 114: 310–317.
- Chien YH, Jores R, Crowley MP. Recognition by gamma/delta T cells. Annu Rev Immunol. 1996; 14: 511–532.
- Asarnow DM, Kuziel WA, Bonyhadi M, Tigelaar RE, Tucker PW, Allison JP. Limited diversity of gamma delta antigen receptor genes of Thy-1* dendritic epidermal cells. Cell. 1988; 55: 837–847.
- epidermal cells. Cell. 1988; 55: 837–847.

 57. Adams EJ, Chien YH, Garcia KC. Structure of a gammadelta T cell receptor in complex with the nonclassical MHC T22. Science. 2005; 308: 227–231.
- Pechhold K, Wesch D, Schondelmaier S, et al. Primary activation of V gamma 9-expressing gamma delta T cells by Mycobacterium tuberculosis. Requirement for Th1-type CD4 T cell help and inhibition by IL-10. J Immunol. 1994; 152(10): 4984-4992.
- Behr C, Poupot R, Peyrat MA, et al. Plasmodium falciparum stimuli for human gammadelta T cells are related to phosphorylated antigens of mycobacteria. Infect Immun. 1996; 64(8): 2892–2896.
- Arigoni D, Sagner S, Latzel C, et al. Terpenoid biosynthesis from 1-deoxy-D-xylulose in higher plants by intramolecular skeletal rearrangement. Proc Natl Acad Sci USA. 1997; 94: 10600–10605.
- Espinosa E, Belmant C, Pont F, et al. Chemical synthesis and biological activity
 of bromohydrin pyrophosphate, a potent stimulator of human gamma delta T
 cells. J Biol Chem. 2001; 276: 18337–18344.
- Rincon-Orozco B, Kunzmann V, Wrobel P, et al. Activation of Vgamma9Vdelta2 T cells by NKG2D. J Immunol. 2005; 175: 2144–2151.
- 63. Hebbeler AM, Cairo C, Cummings JS, et al. Individual Vgamma2-Jgamma1.2* T cells respond to both isopentenyl pyrophosphate and Daudi cell stimulation: generating tumor effectors with low molecular weight phosphoantigens. Cancer Immunol Immunother. 2007; 56: 819–829.
- Zgani I, Menut C, Seman M, et al. Synthesis of prenyl pyrophosphonates as new potent phosphoantigens inducing selective activation of human Vgamma9Vdelta2 T lymphocytes. J Med Chem. 2004; 47: 4600–4612.
- Bonneville M, Scotet E. Human Vgamma9Vdelta2 T cells: promising new leads for immunotherapy of infections and tumors. Curr Opin Immunol. 2006; 18: 539–546.
- Gober HJ, Kistowska M, Angman L, et al. De Libero G. Human T cell receptor gammadelta T cells recognize endogenous mevalonate metabolites in tumor cells. J Exp Med. 2003; 197: 163–168.
- Dieli F, Vermijlen D, Fulfaro F, et al. Targeting human gammadelta T cells with zoledronate and interleukin-2 for immunotherapy of hormone-refractory prostate cancer. Cancer Res. 2007; 67: 7450–7457.
- Kim S, Jizuka K, Aguila HL, et al. In vivo natural killer cell activities revealed by natural killer cell-deficient mice. Proc Natl Acad Sci USA. 2000; 97: 2731–2736.
- Nishio N, Fujita M, Tanaka Y, et al. Zoledronate sensitizes neuroblastoma-derived tumor-initiating cells to cytolysis mediated by human γδ T cells. J Immunother. 2012; 35: 598–606.
- Idrees ASM, Sugie T, Inoue C, et al. Comparison of γδ T cell responses and farnesyl diphosphate synthase inhibition in tumor cells pretreated with zoledronic acid. Cancer Sci. 2013; 104: 536–542.
- Miyagawa F, Tanaka Y, Yamashita S, et al. Essential requirement of antigen presentation by monocyte lineage cells for the activation of primary human gamma delta T cells by aminobisphosphonate antigen. J Immunol. 2001; 166: 5508-5514.
- Brandes M, Willimann K, Moser B. Professional antigen-presentation function by human gamma delta T cells. Science. 2005; 309: 264–268.
- Shin S, El-Diwany R, Schaffert S, et al. Antigen recognition determinants of gamma delta T cell receptors. Science. 2005; 308: 252–255.
- Sarikonda G, Wang H, Puan KJ, et al. Photoaffinity antigens for human gammadelta T cells. J Immunol. 2008; 181: 7738–7750.

- Wei H, Huang D, Lai X, et al. Definition of APC presentation of phosphoantigen (E)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate to Vgamma2Vdelta2 TCR. I Immunol. 2008: 181: 4798–4806.
- Kabelitz D. CD277 takes the lead in human T-cell activation. Blood. 2012; 120: 2159–2161.
- Sutton CE, Lalor SJ, Sweeney CM, et al. Interleukin-1 and IL-23 induce innate IL-17 production from gammadelta T cells, amplifying Th17 responses and autoimmunity. Immunity. 2009; 31: 331–341.
- Eberl M, Engel R, Beck E, Jomaa H. Differentiation of human gamma-delta T cells towards distinct memory phenotypes. Cell Imminol. 2002; 218(1-2): 1-6.
- Ribot JC, deBarros A, Pang DJ, et al. CD27 is a thymic determinant of the balance between interferon-gamma- and interleukin 17-producing gammadelta T cell subsets. Nat Immunol. 2009; 10: 427–436.
- Gioia C, Agrati C, Casetti R, et al. Lack of CD27-CD45RA-Vgamma 9V delta2* T cell effectors in immunocompromised hosts and during active pulmonary tuberculosis. J Immunol. 2002; 168: 1484–1489.
- Cho JS, Pietras EM, Garcia NC, et al. IL-17 is essential for host defense against cutaneous Staphylococcus aureus infection in mice. J Clin Invest. 2010; 120: 1762–1773.
- Ribot JC, Debarros A, Silva-Santos B. Searching for "signal 2": costimulation requirements of gammadelta T cells. Cell Mol Life Sci. 2011; 68: 2345–2355.
- Sarikonda G, Wang H, Puan KJ, et al. Photoaffinity antigens for human cdT Cells. J Immunol. 2008; 181: 7738–7750.
- Budd RC, Russell JQ, van Houten N, et al. CD2 expression correlates with proliferative capacity of alpha beta1 or gamma delta1 CD42 CD82 T cells in lpr mice. J Immunol. 1992; 148: 1055–1064.
- Lafont V, Liautard J, Gross A, et al. Tumor necrosis factor-alpha production is differently regulated in gamma delta and alpha beta human T lymphocytes. J Biol Chem. 2000; 275: 19282–19287.
- Mohan JF, Levisetti MG, Calderon B, et al. Unique autoreactive T cells recognize insulin peptides generated within the islets of Langerhans in autoimmune diabetes. Nat Immunol. 2010; 11: 350–354.
- Kozbor D, Trinchieri G, Monos DS, et al. Human TCR-gamma1/delta1, CD81
 T lymphocytes recognize tetanus toxoid in an MHC-restricted fashion. J Exp Med. 1989; 169: 1847–1851.
- Silva-Santos B, Pennington DJ, Hayday AC. Lymphotoxin-mediated regulation of gammadelta cell differentiation by alphabeta T cell progenitors. Science. 2005; 307: 925–928.
- 89. Vantourout P, Hayday A. Six-of-the-best: unique contributions of $\gamma\delta$ T cells to immunology. Nat Rev Immunol. 2013; 13(2):88–100.
- Caccamo N, Battistini L, Bonneville M, et al. CXCR5 identifies a subset of Vgamma9Vdelta2 T cells which secrete IL-4 and IL-10 and help B cells for antibody production. Immunol. 2006; 177(8): 5290-5295.
- Devilder MC, Maillet S, Bouyge-Moreau I, et al. Potentiation of antigen-stimulated Vgamma9Vdelta2 T cell cytokine production by immature dendritic cells (DC) and reciprocal effect on DC maturation. J Immunol. 2006; 176: 1386–1393.
- Ismaili J, Olislagers V, Poupot M, et al. Human gamma delta T cells induce dendritic cell maturation. Clin Immunol. 2002; 103: 296–302.
- 93. Leslie DS, Vincent MS, Spada FM, et al. CD1-mediated gamma/delta T cell maturation of dendritic cells. J Exp Med. 2002; 196: 1575–1584.
- Collins C, Wolfe J, Roessner K, et al. Lyme arthritis synovial gammadelta T cells instruct dendritic cells via fas ligand. The Journal of Immunology. 2005; 175: 5656–5665.
- Dieli F, Caccamo N, Meraviglia S, et al. Reciprocal stimulation of gammadelta T cells and dendritic cells during the anti-mycobacterial immune response. Eur J Immunol. 2004; 34: 3227–3235.
- Martino A, Casetti R, D'Alessandri A, et al. Complementary function of gamma delta T-lymphocytes and dendritic cells in the response to isopentenyl-pyrophosphate and lipopolysaccharide antigens. J Clin Immunol. 2005; 25: 230–237.
- DiTirro J, Rhoades ER, Roberts AD, et al. Disruption of the cellular inflammatory response to Listeria monocytogenes infection in mice with disruptions in targeted genes. Infect Immun. 1998; 66: 2284–2289.
- Sharp LL, Jameson JM, Cauvi G, et al. Dendritic epidermal T cells regulate skin homeostasis through local production of insulin-like growth factor 1. Nat Immunol. 2005; 6: 73–79.
- Jameson JM, Cauvi G, Sharp LL, et al. Gammadelta T cell-induced hyaluronan production by epithelial cells regulates inflammation. J Exp Med. 2005; 201: 1269–1279.
- 100. Skeen MJ, Freeman MM, Ziegler HK. Changes in peritoneal myeloid populations and their proinflammatory cytokine expression during infection with Listeria monocytogenes are altered in the absence of gamma/delta T cells. J Leukocyte Biol. 2004; 76: 104–115.
- 101. Schilbach KE, Geiselhart A, Wessels J T, et al. Human gammadelta T lymphocytes exert natural and IL-2-induced cytotoxicity to neuroblastoma cells. J Immunother. 2000; 23: 536–548.
- 102. Todaro M, D'Asarob M, Caccamo N, et al. Efficient killing of human colon cancer atem cells by gammadelta T lymphocytes. J Immunol. 2009; 182: 7287–7296.
- 103. D'Asaro M., La Mendola C, Di Liberto D, et al. V gamma 9V delta 2 T lymphocytes efficiently recognize and kill zoledronate-sensitized, imatinib-sensitive, and imatinib-resistant chronic myelogenous leukemia cells. J Immunol. 2010; 184: 3260-3268.

- 104. Chargui J, Combaret V, Scaglione V, et al. Bromohydrin pyrophosphate-stimulated Vgamma9delta2 T cells expanded ex vivo from patients with poor-prognosis neuroblastoma lyse autologous primary tumor cells. J Immunother. 2010; 33: 591–598.
- 105. Kato Y, Tanaka Y, Tanaka H, et al. Requirement of species-specific interactions for the activation of human gamma delta T cells by pamidronate. J Immunol. 2003; 170: 3608–3613.
- 106. Kato Y, Tanaka Y, Hayashi M, et al. Involvement of CD166 in the activation of human gamma delta T cells by tumor cells sensitized with nonpeptide antigens. J Immunol. 2006; 177: 877–884.
- 107. Li W, Kubo S, Okuda A, et al. Effect of IL-18 on expansion of gammadelta T cells stimulated by zoledronate and IL-2. J Immunother. 2010; 33: 287–296.
- 108. Laggner U, Di Meglio P, Perera GK, et al. Identification of a novel proinflammatory human skin-homing $V\gamma9V\delta2$ T cell subset with a potential role in psoriasis. J Immunol. 2011; 187(5): 2783–2793.
- Brandes M, Willimann K, Moser B. Professional antigen-presentation function by human gammadelta T Cells. Science. 2005; 309(5732): 264–268.
- Li H, Luo K, Pauza CD. TNF-alpha is a positive regulatory factor for human Vgamma2Vdelta2 T cells. J Immunol. 2008; 181: 7131–7137.
- 111. van den Broek MF, Muller U, et al. Immune defence in mice lacking type I and/or type II interferon receptors. Immunological Reviews. 1995; 148: 5–18.
- 112. Beetz S, Wesch D, Marischen L, et al. Innate immune functions of human gammadelta T cells. Immunobiology. 2008; 213:173–182.
- 113. Spinozzi F, Agea E, Bistoni O, et al. Local expansion of allergen-specific CD30+Th2-type gamma delta T cells in bronchial asthma. Mol Med. 1995; 1: 821–826.
- 114. Rhodes KA, Andrew EM, Newton DJ, et al. A subset of IL-10-producing gammadelta T cells protect the liver from Listeria-elicited, CD8 (+) T cell-mediated injury. Eur J Immunol. 2008; 8: 2274–2283.
- 115. Jensen KD, Su X, Shin S, et al. Thymic selection determines gammadelta T cell effector fate: antigen-naive cells make interleukin-17 and antigen-experienced cells make interferon gamma. Immunity. 2008; 29: 90-100.
- 116. Wu XL, Zhang JY, Huang A, et al. Decreased Vδ2γδ T cells associate with liver damage through regulating Th17 response in chronic hepatitis B patients. J Infect Dis. 2013; doi:10.1093/infdis/jit312.
- Monforte A, Abrams D, Pradier C, et al. HIV-induced immunodeficiency and mortality from AIDS-defining and non-AIDS-defining malignancies. AIDS. 2008; 22: 2143–2153.
- 118. Salavoura K, Kolialexi A, Tsangaris G, et al. Development of cancer in patients with primary immunodeficiencies. Anticancer Research. 2008; 28: 1263–1269.
- Waldmann TA. Immunotherapy: past, present and future. Nature Medicine. 2003; 9: 269–277.
- 120. Cordova A, Toia F, La Mendola C, et al. Characterization of human $\gamma\delta$ T lymphocytes infiltrating primary malignant melanomas. PLoS One. 2012; 7(11): e49878.
- 121. Gomes AQ, Correia DV, Grosso AR, et al. Identification of a panel of ten cell surface protein antigens associated with immunotargeting of leukemias and lymphomas by peripheral blood gamma delta T cells. Haematologica. 2010; 95: 1397–1404.
- 122. Liu Z, Guo BL, Gehrs BC, et al. Ex vivo expanded human Vgamma9Vdelta2 gammadelta-T cells mediate innate antitumor activity against human prostate cancer cells in vitro. J Urol. 2005; 173:1552–1556.
- 123. Saito A, Narita M, Yokoyama A, et al. Enhancement of anti-tumor cytotoxicity of expanded gammadelta T cells by stimulation with monocyte-derived dendritic cells. J Clin Exp Hematop. 2007; 47: 61–72.
- 124. Ferrarini M, Pupa SM, Zocchi MR, et al. Distinct pattern of HSP72 and monomeric laminin receptor expression in human lung cancers infiltrated by gamma/delta T lymphocytes. Int J Cancer. 1994; 57: 486–490.
- 125. Chou chary A, Davodeau F, Moreau A, et al. Selective lysis of autologous tumor cells by recurrent gamma delta tumor-infiltrating lymphocytes from renal carcinoma. J Immunol. 1995; 154: 3932–3940.
- 126. Dhar S, Chiplunkar SV. Lysis of aminobisphosphonate-sensitized MCF-7 breast tumor cells by V γ 9V δ 2 cells. Cancer Immun. 2010; 10: 10.
- 127. Lamb LS Jr, Henslee-Downey PJ, Parrish RS, et al. Increased frequency of TCR γδ* T cells in disease-free survivors following T cell-depleted, partially mismatched, related donor bone marrow transplantation for leukemia. J Hematother . 1996; 5: 503–509.
- 128. Viey E, Fromont G, Escudier B, et al. Phosphostim-activated gamma delta T cells kill autologous metastatic renal cell carcinoma. J Immunol. 2005; 174: 1338–7347
- 129. Sugie T, Murata-Hirai K, Iwasaki M, et al. Zoledronic acid-induced expansion of γδ T cells from early-stage breast cancer patients: effect of IL-18 on helper NK cells. Cancer Immunol Immunother. 2013; 62: 677–687.
- Liotta LA, Kohn EC. The microenvironment of the tumour-host interface. Nature. 2001; 411: 375–379.
- 131. Brand es M, Willimann K, Bioley G, et al. Cross-presenting human gammade Ita T cells induce robust CD8+ alpha beta T cell responses. Proc Natl Acad Sci USA. 2009; 106: 2307–2312.
- Kabel-itz D, Wesch D. Features and functions of gammadelta T lymphocytes: focus on chemokines and their receptors. Crit Rev Immunol. 2003; 23: 339–370.
- 133. Bennouma J, Bompas E, Neidhardt EM, et al. Phase-I study of Innacell gammade Ita, an autologous cell-therapy product highly enriched in gamma9d elta2 T lymphocytes, in combination with IL-2, in patients with metastatic renal cell carcinoma. Cancer Immunol Immunother. 2008; 57: 1599-1609.

- 134. Dokouhaki P, Han M, Joe B, et al. Adoptive immunotherapy of cancer using ex vivo expanded human $\gamma\delta$ T cells: A new approach. Cancer Lett. 2010; 297: 126–136.
- 135. Kikuchi E, Yamazaki K, Torigoe T, et al. HLA class I antigen expression is associated with a favorable prognosis in early stage non-small cell lung cancer. Cancer Sci. 2007; 98: 1424–1430.
- 136. Meraviglia S, Eberl M, Vermijlen D, et al. In vivo manipulation of $V\gamma9V\delta2$ T cells with zoledronate and low-dose interleukin-2 for immunotherapy of advanced breast cancer patients. Clin Exp Immunol. 2010; 161: 290–297.
- 137. Kobayashi H, Tanaka Y, Shimmura H, et al. Complete remission of lung metastasis following adoptive immunotherapy using activated autologous gammadelta T-cells in a patient with renal cell carcinoma. Anticancer Res. 2010; 30: 575–579.
- Wilhelm M, Kunzmann V, Eckstein S, et al. γδ T cells for immune therapy of patients with lymphoid malignancies. Blood. 2003; 102: 200–206.
- 139. Corvaisier M, Moreau-Aubry A, Diez E, et al. V gamma 9 V delta 2 T cell response to colon carcinoma cells. J Immunol. 2005; 175: 5481–5488.
- 140. Marten A, Lilienfeld-Toal M, Buchler MW, et al. Zoledronic acid has direct antiproliferative and antimetastatic effect on pancreatic carcinoma cells and acts as an antigen for delta2 gamma/delta T cells. J Immunother. 2007; 30: 370-377.
- 141. Lalor SJ, Dungan LS, Sutton CE, et al. Caspase-1-processed cytokines IL-1b and IL-18 promote IL-17 production by γδ and CD4 T cells that mediate autoimmunity. J Immunol. 2011; 186: 5738–5748.
- 142. Fink DR, Holm D, Schlosser A, et al. Elevated numbers of SCART11 gammadelta T cells in skin inflammation and inflammatory bowel disease. Mol Immunol. 2010; 47: 1710–1718.
- 143. Murdoch JR, Lloyd CM. Resolution of allergic airway inflammation and airway hyperreactivity is mediated by IL-17 producing γδ T cells. Am J Respir Crit Care Med. 2010; 182: 464–476.
- 144. Qin G, Mao H, Zheng J, et al. Phosphoantigen-expanded human gammadelta T cells display potent cytotoxicity against monocyte-derived macrophages infected with human and avian influenza viruses. J Infect Dis. 2009; 200: 858–865.
- 145. Qin G, Liu Y, Zheng J, et al. Type 1 responses of human Vγ9Vδ2 T cells to influenza A viruses. J Virol. 2011; 85(19): 10109–10116.
- 146. Tu W, Zheng J, Liu Y, et al. The aminobisphosphonate pamidronate controls influenza pathogenesis by expanding a gammadelta T cell population in humanized mice. J Exp Med. 2011; 208(7): 1511–1522.
- Fujishima N, Hirokawa M, Fujishima M, et al. Skewed T cell receptor repertoire of Vö1* T lymphocytes after human allogeneic haematopoietic stem cell transplantation and the potential role for Epstein-Barr virus-infected B cells in clonal restriction. Clin Exp Immunol. 2007; 149: 70-79.
 Agrati C, Alonzi T, De Santis R, et al. Activation of Vy9Vô2 T cells by
- 148. Agrati C, Alonzi T, De Santis R, et al. Activation of Vγ9Vδ2 T cells by non-peptidic antigens induces the inhibition of subgenomic HCV replication. Int Immunol. 2006; 18: 11–18.
- 149. Tikhonov I, Deetz CO, Paca R, et al. Human Vgamma2Vdelta2 T cells contain cytoplasmic RANTES. Int Immunol. 2006; 18: 1243–1251.
- Fang H, Welte T, Zheng X, et al. Gamma delta T cells promote the maturation of dendritic cells during West Nile virus infection. FEMS Immunol Med Microbiol. 2010; 59: 71–80.
- 151. Knight A, Madrigal AJ, Grace S, et al. The role of Vdelta2-negative gamma-delta T cells during cytomegalovirus reactivation in recipients of allogeneic stem cell transplants. Blood. 2010; 116: 2164–2172.
- 152. Devaud C, Bilhere E, Loizon S, et al. Antitumor activity of T cells reactive against cytomegalovirus-infected cells in a mouse xenograft tumor model. Cancer Res. 2009; 69: 3971–3978.
- Sciammas R, Bluestone JA. TCR gamma delta cells and viruses. Microbes Infect. 1999; 1: 203–212.
- 154. Chen Z, Freedman MS. CD16⁺ gamma delta T cells mediate antibody dependent cellular cytotoxicity: potential mechanism in the pathogenesis of multiple sclerosis. Clin Immunol. 2008; 128: 219–227.
- 155. Riedel DJ, Sajadi MM, Armstrong CL, et al. Natural viral suppressors of HIV-1 have a unique capacity to maintain gammadelta T cells. AIDS. 2009; 23: 1955–1964.
- Poccia F, Gioia C, Martini F, et al. Zoledronic acid and interleukin-2 treatment improves immunocompetence in HIV-infected persons by activating Vgamma9Vdelta2 T cells. AIDS. 2009; 23: 555–565.
- 157. de Paoli P, Gennari D, Martelli P, et al. Gammadelta T cell receptor-bearing lymphocytes during Epstein-Barr virus infection. J Infect Dis. 1990; 161: 1013–1016.
- 158. Tseng CT, Miskovsky E, Houghton M, et al. Characterization of liver T-cell receptor gammadelta T cells obtained from individuals chronically infected with hepatitis C virus (HCV): evidence for these T cells playing a role in the liver pathology associated with HCV infections. Hepatology. 2001; 33: 1312–1320.
- 159. Kroca M, Johansson A, Sjöstedt A, et al. V gamma 9V delta 2 T cells in human legionellosis. Clin Diagn Lab Immunol. 2001; 8(5): 949–954.
- 160. Xu S, Han Y, Xu X, et al. IL-17A-producing $\gamma\delta$ T cells promote CTL responses against Listeria monocyto genes infection by enhancing dendritic cell cross-presentation. J Immunol. 2010; 185: 5879–5887.
- 161. Egan CE, Dalton JE, Andrew EM, et al. A requirement for the Vgamma1⁺ subset of peripheral gammadelta T cells in the control of the systemic growth of Toxoplasma gondii and infection-induced pathology. J Immunol. 2005; 175: 8191–8199.

- 162. Costa G, Loizon S, Guenot M, et al. Control of Plasmodium falciparum erythrocytic cycle: $\gamma\delta$ T cells target the red blood cell-invasive merozoites. Blood. 2011; 118(26): 6952–6962.
- 163. Su D, Shen M, Li X, et al. Roles of $\gamma\delta$ T cells in the pathogenesis of autoimmune diseases. Clin Dev Immunol. 2013; 2013: 985753.
- 164. Hu C, Qian L, Miao Y, et al. Antigen-presenting effects of effector memory Vγ9Vδ2 T cells in rheumatoid arthritis. Cell Mol Immunol. 2012; 9(3): 245–254.
- 165. Li X, Kang N, Zhang X, et al. Generation of human regulatory gammadelta T cells by TCR gammadelta stimulation in the presence of TGF-beta and their involvement in the pathogenesis of systemic lupus erythematosus. J Immunol. 2011; 186(12): 6693–700.
- 166. Ponomarev ED, Dittel BN. Gamma delta T cells regulate the extent and duration of inflammation in the central nervous system by a Fas ligand-dependent mechanism. J Immunol. 2005; 174(8): 4678–4687.
- 167. Cairo C, Arabito E, Landi F, et al. Analysis of circulating gammadelta T cells in children affected by IgE-associated and non-IgE-associated allergic atopic eczema/dermatitis syndrome. Clin Exp Immunol. 2005; 141(1): 116–121.
- 168. Zhang L, Liu J, Wang E, et al. Respiratory syncytial virus protects against the subsequent development of ovalbumin -induced allergic responses by inhibiting Th2-type $\gamma\delta$ T cells. J Med Virol. 2013; 85 (1): 149–156.
- 169. Svensson L, Lilliehook B, Larsson R, et al. Gamma/delta T cells contribute to the systemic immunoglobulin E response and local B cell reactivity in allergic eosinophilic airway inflammation. Immunology. 2003; 108:98–108.
- eosinophilic airway inflammation. Immunology. 2003; 108:98–108. 170. de Oliveira Henriques MD, Penido C. γδ T Lymphocytes Coordinate Eosinophil Influx during Allergic Responses. Front Pharmacol. 2012; 3: 200.
- 171. Pawankar R. γδ T cells in allergic airway diseases. Clin Exp Allergy. 2000; 30:
- 172. Pawankar R, Okuda M, Suzuki K, et al. Phenotypic and molecular characteristics of nasal mucosal γδ T cells in allergic and infectious rhinitis. Am J Respir Crit Care Med. 1996; 153: 1655–1665.
- 173. Sicard H, Ingoure S, Luciani B, et al. In vivo immunomanipulation of vgam-ma9vdelta2 T cells with a synthetic phosphoantigen in a preclinical nonhuman primate model. J Immunol. 2005; 175: 5471–5480.
- 174. Shojaei H, Oberg H. H, Juricke M, et al. Toll-like receptors 3 and 7 agonists enhance tumor cell lysis by human gammadelta T cells. Cancer Research. 2009; 69: 8710–8717.
- 175. Gertner-Dardenne J, Bonnafous C, Bezombes C, et al. Bromohydrin pyrophosphate enhances antibody-dependent cell-mediated cytotoxicity induced by therapeutic antibodies. Blood. 2009; 113: 4875–4884.

Author biography



Dr. Yan-Ling Wu is a professor in Molecular Immunology and now heads the Cellular and Molecular Immunology Research Group. She received Master and Doctoral degrees in Applied Life Science in 2003 and in Medicine Science in 2006, respectively, from Tohoku

University, Japan. After that, she entered to Professor Minato's group of School of Medicine, Kyoto University, Japan, as a senior researcher working in the field of molecular immunology. Her current research focuses on understanding the molecular mechanisms of gene regulation related to diseases by immune inhibitory receptors. Dr. Wu has published better papers as the first/corresponding author in excellent Journals including *Chem & Biol, ChemBioChem, Int J Biol Sci* etc. and given oral presentations in international conferences.



Yan-Ping Ding received her bachelor's degree in biological sciences from Zhejiang University of Chinese Traditional Medicine, China. Since graduation, she has been working in Zhejiang Center of Disease Control and Prevention, China. Her research mainly focuses

on the field of Molecular Immunology under the guidance of Prof. Yanling Wu.



Yoshimasa Tanaka received his Ph. D in Hokkaido University Graduate School of Agriculture with a specialization in Enzymology and Biochemistry. After graduation, he continued his research in the field of Immunobiology. Since 2008, he is an associate professor and works

in the Center for Innovation in Immunoregulative Immunology and Therapeutics that belongs to the Kyoto University Graduate School of Medicine.



Li-Wen Shen is a postgraduate majoring in pharmacy. He obtained the Bachelor's degree in Biological Engineering in 2012 from Shanxi Datong University, China. After that, he entered Lab of Molecular Immunology, Zhejiang Provincial Center for Disease Control and

Prevention, China, working with small molecules to regulate disease-related gene to explore gene-targeted drugs under the direction of Profs Y.-L. Wu and W. Zhang.



Chuan-He Wei received his Bachelor's degree in 2011 from Jilin Agricultural Science and Technology College, China. Then, he entered Prof. Zhang's group as a postgraduate of College of Pharmaceutical Science, Zhejiang University of Technology, China, his

current research mainly focusing on the field of Molecular Immunology under the direction of Profs W. Zhang and Y.-L. Wu.



Dr. Nagahiro Minato is a full professor of Department of Immunology and Cell Biology, Graduate School of Medicine, Kyoto University, Japan, since 1992, and since 1998, also a professor of Department of Bio-Regulation, Graduate School of

Biostudies, Kyoto University (Joint appointment), Kyoto, Japan. He received his MD & PhD from Kyoto University in 1975. At present, his research focuses on Immunobiology. Prof Minato has published more than 150 papers in excellent Journals, including Nature Medicine, Nature Cell Biol, Nature Immunol, Nature Med, Proc Natl Acad Sci USA, J Immunol, Blood, J Exp Med etc.