

Figure 4 | Host-microbe interactions mediated by butyrate and niacin. Butyrate is mainly produced by clusters IV and XIVa of *Clostridia*. Although butyrate acts as an energy source for normal colonic epithelial cells (trophic effect), it also has the capability to suppress proliferation of cancerous epithelial cells that usually undergo the Warburg effect⁹⁹. Butyrate upregulates histone H3 acetylation at regulatory regions of the *Foxp3* gene and facilitates differentiation of naive CD4+T cells into T_{reg} cells. In contrast, butyrate together with other SCFAs induce TGF-β secretion by epithelial cells through an unknown mechanism⁷¹. Gpr109a was originally described as the receptor for niacin. Butyrate and niacin bind Gpr109a on epithelial cells to trigger production of a cytoprotective cytokine IL-18. These microbial metabolites also stimulate dendritic cells and macrophages to produce IL-10 and retinoic acids, both of which are important for the development of IL-10-producing T_{reg} s in the colon. Therefore, butyrate and niacin contribute to the maintenance of intestinal homeostasis through multiple mechanisms.

hepaticus^{75,76}. Furthermore, sphingolipids of *B. fragilis* reduce cell numbers of invariant NKT (iNKT) cells in the colon. Reduction of iNKT cells in response to colonization with *B. fragilis* results in alleviation of oxazolan-induced experimental colitis^{77,78} because iNKT cells are responsible for the colitis development by producing large amounts of cytokines upon stimulation with glycosphingolipids presented by CD1d⁷⁹. All of these observations indicate that different diet-independent metabolites have distinct roles in the maintenance of gut immune homeostasis.

So far, a variety of diet-dependent metabolites have been linked to the epithelial barrier, immune regulation and inflammation. These products include bile acids, short and long chain fatty acids and vitamins 66,80 . Certain Bifidobacterium spp. actively produce folate (vitamin $B_9)^{81}$, and its derivative, 6-formyl pterin (6-FP), is one of ligands of the MHC class I-like molecule MR1, which presents an antigen to mucosal-associated invariant T (MAIT) cells 82 . Notably, several riboflavin (vitamin B_2)-based metabolites, whose structures are closely related to 6-FP with an extra ribityl moiety, activate Jurkat T-cell lines (expressing invariant MAIT TCR α and β chains) to produce TNF and IFN- γ in an MR1-restricted manner. Given that B-group vitamins including rivoflavin are known to be synthesized de novo by certain commensal bacteria, it is possible that MAIT cells may sense the growth of riboflavin-producing bacteria on the mucosa.

Indole is a quorum-sensing molecule produced from tryptophan by the tryptophanase of a variety of Gram-positive and Gram-negative intestinal bacteria. Indole enhances epithelial barrier functions in vitro and in vivo through upregulation of components of tight junction complexes^{83,84}. Acetate is another metabolite that enhances gut epithelial barrier functions (Fig. 3). GF mice succumb to Escherichia coli O157:H7 lethal infection; however, inoculation of mice with certain Bifidobacteria strains prevents E. coil O157-induced death85. Bifidobacteriaderived acetate inhibits translocation of luminal Shiga toxin from the gut lumen to the blood by improving epithelial defense functions and suppressing colonic inflammation. These findings provide insight into the mode of action through which probiotic bacteria exert a protective effect against pathological infections and open up a new question on how bacterial metabolites regulates epithelial barrier functions.

The anti-inflammatory effect of SCFAs, including acetate, has been well characterized on both epithelial and immune cell levels^{62,86} (Fig. 4). Oral administration of acetate in drinking water suppresses not only DSS-induced experimental colitis but also inflammatory arthritis and ovalbumininduced allergic airway inflammation. The therapeutic effect of acetate is canceled in Gpr43-deficient mice, implying a major role for the acetate-Gpr43 axis in containment of inflammation. Acetate-dependent Gpr43 activation on neutrophils results in upregulation of apoptosis-related gene clusters and thus provokes apoptosis.

SCFAs also have an impact on the terminal differentiation of CD4⁺ helper T cells⁶². Administration of an individual or a cocktail of SCFAs to GF or SPF mice

increased the frequency of colonic Foxp3 $^{\scriptscriptstyle +}$ regulatory $T_{\scriptscriptstyle reg}$ cells 87,88 , which have a central role in the suppression of the inflammatory and allergic immune responses⁸⁹. After oral administration, butyrate per se is less effective on the accumulation of colonic T_{reg}s as compared with that of the other two SCFAs⁸⁷. However, this observation may reflect enhanced migration from lymphoid tissues into the colon rather than an increase in Treg development in the colon, given that a large part of the orally administered SCFAs are absorbed into the portal vein and/or used by epithelial cells (butyrate is a particularly good energy source) in the upper part of the intestine before reaching the colon. The in vitro T_{reg} induction assay indicated that, among SCFAs, butyrate most effectively induces the differentiation of naive T cells into T_{reg} cells^{88,90}. Propionate also shows the T_{reg}-inducing effect to a lesser extent, whereas acetate dose not have any impact on T_{reg} induction. Consistent with in vitro observations, colonic Treg cells are significantly augmented in SPF mice with a diet containing butyrylateresistant starch and slightly augmented with propionylated resistant starch but are hardly induced with acetylated resistant starch90. These observations show that locally produced butyrate is responsible for T_{res} differentiation and that orally administered acetate and propionate may be important for the migration of T_{reg}s into the colon. Indeed, oral administration of propionate upregulates Gpr15 (ref. 91), a T_{reg}-specific gut-homing molecule, in a Gpr43-dependent manner⁸⁷. Butyrate is well known to regulate gene expression epigenetically by inhibiting histone deacetylases (HDACs). Chromatin immunoprecipitation sequencing (ChIP-seq) analysis demonstrated that butyrate upregulates histone H3 acetylation at regulatory regions of the Foxp3 gene locus and therefore facilitates Foxp3 expression (Fig. 4). In addition to such a direct effect on CD4+ T cells, butyrate indirectly induces IL-10-producing T_{reg} cells by imparting anti-inflammatory properties on dendritic cells (DCs) and macrophages in the colonic lamina propria in a Gpr109adependent manner⁵⁶ (Fig. 4). Niacin, a pharmacological agonist for Gpr109a, also exhibits a similar anti-inflammatory activity (Fig. 4). Collectively, these findings demonstrate that microbe-derived SCFAs and niacin contribute to the maintenance of gut immune homeostasis by promoting T_{reg} accumulation in the colon through multiple mechanisms. In support of this idea, butyrate-producing bacteria decreased in the microbial community of IBD patients compared with that of healthy subjects92. It has also been reported that gut microbiota-derived SCFAs, particularly propionate, alleviate an allergic airway response induced by house dust mite extract in mice93. The feeding of a diet rich in fermentable carbohydrates promotes the outgrowth of bacteria that belong to the Bacteroidetes phylum, leading to increased serum levels of acetate and propionate, which in turn increase the hematopoiesis of DC precursors⁹³. These DCs exhibit an impaired capacity to elicit allergy-prone T_H2 responses in the lung. This process requires the receptor Gpr41 but not Gpr43. This study provides key evidence that bacteria-derived metabolites may affect not only the local but the systemic immune system. Collectively, the recent data support the idea that dietdependent metabolites shape host immunity by activating different mechanisms, including Gprs-dependent intracellular signaling activation and epigenetic modifications. Although further investigations will be required to clarify the precise molecular mechanism, the interactions between gut metabolites and host immunocompetent cells are essential for the successful establishment and maintenance of the host immune system.

Gut metabolites and disease development. Disturbance of the microbiota has been implicated in the pathogenesis of several human disorders, including IBD, obesity and cardiovascular disease94. For example, gut microbial metabolism of dietary food-derived phosphatidylcholine and L-carnitine generates trimethylamine (Fig. 3), which is further metabolized to trimethylamine-N-oxide (TMAO), a proatherogenic agent, in mice and humans^{12,13}. A HFD alters gut microbial composition in mice, leading to expansion of deoxycholic acid-producing bacteria, namely cluster XI of the genus Clostridium¹⁴. This secondary bile acid induces a phenotypic change in hepatic stellate cells to secrete proinflammatory cytokines and eventually facilitates hepatocellular carcinoma (Fig. 3).

Accumulating reports have linked gut microbiota to emotional behavior. Alteration of the gut microbial community owing to TLR5 deficiency increases appetite (about 10% increase in food intake) and induces metabolic syndrome (for example, hyperlipidemia, insulin resistance, obesity and hepatic steatosis) in mice without affecting the efficiency of dietary energy harvest⁹⁵. In addition, chronic ingestion of fermented milk with probiotics can modulate the responsiveness of an extensive brain network in healthy subjects%. Although the exact mechanism remains to be elucidated, these observations suggest that metabolites produced by microbial fermentation have a key role in brain activity.

A current study revealed that a mouse model of autism spectrum disorder (ASD) can be triggered by the gut microbial metabolites 4-ethylphenylsulfate (4EPS) and indolepyruvate⁹⁷ (Fig. 3). Maternal immune activation induced by the administration of an immunostimulant polyinosinic-polycytidylic acid (poly I:C) during pregnancy causes ASD-like behavior in offspring, which is associated with gut barrier dysfunction and systemic leakage of luminal metabolites. In line with these experiential observations, several metabolites (for example, p-cresol and indolyl-3-acryloylglycine), which are structurally similar to 4EPS and indole pyruvate, are detected as human autism biomarkers in urine98. On the basis of these observations, the brain-microbiota-metabolite axis can be considered a potential therapeutic target for ASD and other neurodevelopmental illnesses⁹⁷. Future studies will provide mechanistic insights into how these gut microbiota-derived metabolites contribute to the pathophysiology of neurodevelopmental disorders.

Conclusions and perspectives

It is now becoming clear that the gut microbiota and its metabolites have an important role in host physiology. Of note, recent investigations indicate that different environmental factors, such as nutrition and drugs, can profoundly affect the gut bacterial community, thereby changing the gut microbiome activity, which may result in generations of bioactive metabolites (for example, healthpromoting or disease-causing metabolites). These observations will have ramifications on what we eat and as such would influence food, pharmaceutical and medical sciences by providing new concepts. For example, the use of prebiotics and probiotics or folk medicines often rely on traditional beliefs rather than solid scientific evidence and still suffer because of the lack of more precise knowledge concerning their modes of action. Further investigations of the molecular mechanisms by which prebiotics and probiotics exert their effects on animal physiology will provide new opportunities for improving human health. Furthermore, many important immune and metabolic disorders, including diabetes, obesity, behavioral disorders and chronic inflammation, are now known to be in part due to the imbalance of interactions between the host and microbiota or metabolites. Accordingly, we expect that further investigation on the molecular relationship between microbiota and metabolites and host physiology will provide new concepts and strategies for the development of 'gut metabolite concept'-driven therapeutic agents. Considering the very high diversity of gut bacterial species in different metazoans, it is likely that yet-unexpected microbiotametabolite-metazoa relationships and the molecular dynamics of these relationships remain to be explored. The use of powerful invertebrate and vertebrate genetic animal model system together with multi-omics approaches would allow us to discover and decipher the secrets of evolutionarily conserved molecular dialogs among gut microbiota, metabolites and metazoans, which will certainly open new avenues to prevent and/or treat a number of disorders associated with gut microbiota and metabolites.

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References

- Lee, W.J. & Brey, P.T. How microbiomes influence metazoan development: insights from history and *Drosophila* modeling of gut-microbe interactions. Annu. Rev. Cell Dev. Biol. 29, 571-592 (2013).
- Gusarov, I. et al. Bacterial nitric oxide extends the lifespan of C. elegans. Cell 152, 818-830 (2013).
- Cabreiro, F. et al. Metformin retards aging in C. elegans by altering microbial folate and methionine metabolism. Cell 153, 228-239 (2013).
- 4. Buchon, N., Broderick, N.A. & Lemaitre, B. Gut homeostasis in a microbial world: insights from Drosophila melanogaster. Nat. Rev. Microbiol. 11, 615-626
- Lee, K.A. & Lee, W.J. Drosophila as a model for intestinal dysbiosis and chronic
- inflammatory diseases. *Dev. Comp. Immunol.* **42**, 102–110 (2014). Guo, L., Karpac, J., Tran, S.L. & Jasper, H. PGRP-SC2 promotes gut immune homeostasis to limit commensal dysbiosis and extend lifespan. Cell 156, 109-122 (2014).
- Shin, S.C. et al. Drosophila microbiome modulates host developmental and metabolic homeostasis via insulin signaling. Science 334, 670-674 (2011).
- Gems, D. & Partridge, L. Genetics of longevity in model organisms: debates and paradigm shifts. Annu. Rev. Physiol. 75, 621-644 (2013).
- Abada, E.A. et al. C. elegans behavior of preference choice on bacterial food. Mol. Cells 28, 209-213 (2009).

- Lenaerts, I., Walker, G.A., Van Hoorebeke, L., Gems, D. & Vanfleteren, J.R. Dietary restriction of *Caenorhabditis elegans* by axenic culture reflects nutritional requirement for constituents provided by metabolically active microbes. *J. Gerontol. A Biol. Sci. Med. Sci.* 63, 242–252 (2008).
- Saiki, R. et al. Altered bacterial metabolism, not coenzyme Q content, is responsible for the lifespan extension in Caenorhabditis elegans fed an Escherichia coli diet lacking coenzyme Q. Aging Cell 7, 291–304 (2008).
- Tang, W.H. et al. Intestinal microbial metabolism of phosphatidylcholine and cardiovascular risk. N. Engl. J. Med. 368, 1575–1584 (2013).
- Koeth, R.A. et al. Intestinal microbiota metabolism of L-carnitine, a nutrient in red meat, promotes atherosclerosis. Nat. Med. 19, 576–585 (2013).
- Yoshimoto, S. et al. Obesity-induced gut microbial metabolite promotes liver cancer through senescence secretome. Nature 499, 97–101 (2013).
- Johnson, C.H., Patterson, A.D., Idle, J.R. & Gonzalez, F.J. Xenobiotic metabolomics: major impact on the metabolome. *Annu. Rev. Pharmacol. Toxicol.* 52, 37–56 (2012).
- Nicholson, J.K. & Wilson, I.D. Opinion: understanding 'global' systems biology: metabonomics and the continuum of metabolism. *Nat. Rev. Drug Discov.* 2, 668–676 (2003).
- Mahmood, K., Naeem, M. & Rahimnajjad, N.A. Metformin: the hidden chronicles of a magic drug. Eur. J. Intern. Med. 24, 20–26 (2013).
- 18. Pyra, K.A., Saha, D.C. & Reimer, R.A. Prebiotic fiber increases hepatic acetyl CoA carboxylase phosphorylation and suppresses glucose-dependent insulinotropic polypeptide secretion more effectively when used with metformin in obese rats. J. Nutr. 142, 213–220 (2012).
- Shin, N.R. et al. An increase in the Akkermansia spp. population induced by metformin treatment improves glucose homeostasis in diet-induced obese mice. Gut 63, 727–735 (2014).
- Anisimov, V.N. et al. Metformin slows down aging and extends life span of female SHR mice. Cell Cycle 7, 2769–2773 (2008).
- Onken, B. & Driscoll, M. Metformin induces a dietary restriction–like state and the oxidative stress response to extend *C. elegans* healthspan via AMPK, LKB1, and SKN-1. *PLoS ONE* 5, e8758 (2010).
- Grandison, R.C., Piper, M.D. & Partridge, L. Amino-acid imbalance explains extension of lifespan by dietary restriction in *Drosophila*. Nature 462, 1061–1064 (2009).
- Zimmerman, J.A., Malloy, V., Krajcik, R. & Orentreich, N. Nutritional control of aging. Exp. Gerontol. 38, 47–52 (2003).
- Slack, C., Foley, A. & Partridge, L. Activation of AMPK by the putative dietary restriction mimetic metformin is insufficient to extend lifespan in *Drosophila*. PLoS ONE 7, e47699 (2012).
- Ryu, J.H. et al. Innate immune homeostasis by the homeobox gene caudal and commensal-gut mutualism in *Drosophila*. Science 319, 777–782 (2008).
- O'Hara, A.M. & Shanahan, F. The gut flora as a forgotten organ. EMBO Rep. 7, 688–693 (2006).
- Packey, C.D. & Sartor, R.B. Commensal bacteria, traditional and opportunistic pathogens, dysbiosis and bacterial killing in inflammatory bowel diseases. *Curr. Opin. Infect. Dis.* 22, 292–301 (2009).
- 28. Kamada, N., Chen, G.Y., Inohara, N. & Nuñez, G. Control of pathogens and pathobionts by the gut microbiota. *Nat. Immunol.* 14, 685–690 (2013).
- Lemaitre, B. & Hoffmann, J. The host defense of Drosophila melanogaster. Annu. Rev. Immunol. 25, 697–743 (2007).
- Kim, S.H. & Lee, W.J. Role of DUOX in gut inflammation: lessons from Drosophila model of gut-microbiota interactions. Front. Cell. Infect. Microbiol. 3. 116 (2014).
- 31. Lee, K.-A. *et al.* Bacterial-derived uracil as a modulator of mucosal immunity and gut-microbe homeostasis in *Drosophila*. *Cell* **153**, 797–811 (2013).
- Bischoff, V. et al. Downregulation of the Drosophila immune response by peptidoglycan-recognition proteins SC1 and SC2. PLoS Pathog. 2, e14 (2006).
- Paredes, J.C., Welchman, D.P., Poidevin, M. & Lemaitre, B. Negative regulation by amidase PGRPs shapes the *Drosophila* antibacterial response and protects the fly from innocuous infection. *Immunity* 35, 770–779 (2011).
- Lhocine, N. et al. PIMS modulates immune tolerance by negatively regulating Drosophila innate immune signaling. Cell Host Microbe 4, 147–158 (2008).
- Franzenburg, S. et al. Distinct antimicrobial peptide expression determines host species-specific bacterial associations. Proc. Natl. Acad. Sci. USA 110, E3730—E3738 (2013).
- 36. Cabreiro, F. & Gems, D. Worms need microbes too: microbiota, health and aging in *Caenorhabditis elegans*. *EMBO Mol. Med.* **5**, 1300–1310 (2013).
- Bosch, T.C. Cnidarian-microbe interactions and the origin of innate immunity in metazoans. Annu. Rev. Microbiol. 67, 499–518 (2013).
- McFall-Ngai, M. et al. Animals in a bacterial world, a new imperative for the life sciences. Proc. Natl. Acad. Sci. USA 110, 3229–3236 (2013).
- Chu, H. & Mazmanian, S.K. Innate immune recognition of the microbiota promotes host-microbial symbiosis. Nat. Immunol. 14, 668–675 (2013).
- Storelli, G. et al. Lactobacillus plantarum promotes Drosophila systemic growth by modulating hormonal signals through TOR-dependent nutrient sensing. Cell Metab. 14, 403–414 (2011).
- 41. Ley, R.E., Peterson, D.A. & Gordon, J.I. Ecological and evolutionary forces

- shaping microbial diversity in the human intestine. Cell 124, 837-848 (2006).
- Human Microbiome Project Consortium. Structure, function and diversity of the healthy human microbiome. Nature 486, 207–214 (2012).
- Nicholson, J.K. et al. Host-gut microbiota metabolic interactions. Science 336, 1262–1267 (2012).
- Bäckhed, F. et al. The gut microbiota as an environmental factor that regulates fat storage. Proc. Natl. Acad. Sci. USA 101, 15718–15723 (2004).
- Cummings, J.H. Fermentation in the human large intestine: evidence and implications for health. *Lancet* 1, 1206–1209 (1983).
- Cummings, J.H., Pomare, E.W., Branch, W.J., Naylor, C.P. & Macfarlane, G.T. Short chain fatty acids in human large intestine, portal, hepatic and venous blood. Gut 28, 1221–1227 (1987).
- Donohoe, D.R. et al. The microbiome and butyrate regulate energy metabolism and autophagy in the mammalian colon. Cell Metab. 13, 517–526 (2011).
- Kuma, A. et al. The role of autophagy during the early neonatal starvation period. Nature 432, 1032–1036 (2004).
- Asanuma, N., Kawato, M., Ohkawara, S. & Hino, T. Characterization and transcription of the genes encoding enzymes involved in butyrate production in Butyrivibrio fibrisolvens. *Curr. Microbiol.* 47, 203–207 (2003).
- Cho, I. et al. Antibiotics in early life alter the murine colonic microbiome and adiposity. Nature 488, 621–626 (2012).
- Ley, R.E. et al. Obesity alters gut microbial ecology. Proc. Natl. Acad. Sci. USA 102, 11070–11075 (2005).
- Ley, R.E., Turnbaugh, P.J., Klein, S. & Gordon, J.I. Microbial ecology: human gut microbes associated with obesity. *Nature* 444, 1022–1023 (2006).
- 53. Turnbaugh, P.J. et al. An obesity-associated gut microbes with increased
- capacity for energy harvest. *Nature* 444, 1027–1031 (2006).

 54. Thangaraju, M. *et al.* GPR109A is a G protein–coupled receptor for the bacterial fermentation product butyrate and functions as a tumor suppressor in
- colon. Cancer Res. 69, 2826–2832 (2009).
 55. Le Poul, E. et al. Functional characterization of human receptors for short chain fatty acids and their role in polymorphonuclear cell activation. J. Biol. Chem. 278, 25481–25489 (2003).
- Singh, N. et al. Activation of gpr109a, receptor for niacin and the commensal metabolite butyrate, suppresses colonic inflammation and carcinogenesis. *Immunity* 40, 128–139 (2014).
- Tunaru, S. *et al.* PUMA-G and HM74 are receptors for nicotinic acid and mediate its anti-lipolytic effect. *Nat. Med.* 9, 352–355 (2003).
- Hong, Y.-H. et al. Acetate and propionate short chain fatty acids stimulate adipogenesis via GPCR43. Endocrinology 146, 5092–5099 (2005).
- Bjursell, M. et al. Improved glucose control and reduced body fat mass in free fatty acid receptor 2-deficient mice fed a high-fat diet. Am. J. Physiol. Endocrinol. Metab. 300, E211-E220 (2011).
- Kimura, I. et al. The gut microbiota suppresses insulin-mediated fat accumulation via the short-chain fatty acid receptor GPR43. Nat. Commun. 4, 1829 (2013).
- 61. Tolhurst, G. *et al.* Short-chain fatty acids stimulate glucagon-like peptide-1 secretion via the G protein–coupled receptor FFAR2. *Diabetes* **61**, 364–371 (2012).
- Maslowski, K.M. et al. Regulation of inflammatory responses by gut microbiota and chemoattractant receptor GPR43. Nature 461, 1282–1286 (2009).
- Garrett, W.S., Gordon, J.I. & Glimcher, L.H. Homeostasis and inflammation in the intestine. Cell 140, 859–870 (2010).
- Del Rio, D., Borges, G. & Crozier, A. Berry flavonoids and phenolics: bioavailability and evidence of protective effects. *Br. J. Nutr.* 104 Suppl 3, S67–S90 (2010).
- 65. Possemiers, S., Bolca, S., Verstraete, W. & Heyerick, A. The intestinal microbiome: a separate organ inside the body with the metabolic potential to influence the bioactivity of botanicals. *Fitoterapia* 82, 53–66 (2011).
- Brestoff, J.R. & Artis, D. Commensal bacteria at the interface of host metabolism and the immune system. *Nat. Immunol.* 14, 676–684 (2013).
- Hooper, L.V. & Macpherson, A.J. Immune adaptations that maintain homeostasis with the intestinal microbiota. *Nat. Rev. Immunol.* 10, 159–169
- Bouskra, D. et al. Lymphoid tissue genesis induced by commensals through NOD1 regulates intestinal homeostasis. Nature 456, 507–510 (2008).
- Moreau, M.C., Ducluzeau, R., Guy-Grand, D. & Muller, M.C. Increase in the population of duodenal immunoglobulin A plasmocytes in axenic mice associated with different living or dead bacterial strains of intestinal origin. *Infect. Immun.* 21, 532–539 (1978).
- Atarashi, K. et al. Induction of colonic regulatory T cells by indigenous Clostridium species. Science 331, 337–341 (2011).
- 71. Atarashi, K. et al. T_{reg} induction by a rationally selected mixture of Clostridia strains from the human microbiota. Nature 500, 232–236 (2013).
- Mazmanian, S.K., Liu, C.H., Tzianabos, A.O. & Kasper, D.L. An immunomodulatory molecule of symbiotic bacteria directs maturation of the host immune system. *Cell* 122, 107–118 (2005).
- Cahenzli, J., Köller, Y., Wyss, M., Geuking, M.B. & McCoy, K.D. Intestinal microbial diversity during early-life colonization shapes long-term IgE levels. Cell Host Microbe 14, 559–570 (2013).

- Umetsu, D.T., McIntire, J.J., Akbari, O., Macaubas, C. & DeKruyff, R.H. Asthma: an epidemic of dysregulated immunity. *Nat. Immunol.* 3, 715–720 (2002).
- Mazmanian, S.K., Round, J.L. & Kasper, D.L. A microbial symbiosis factor prevents intestinal inflammatory disease. *Nature* 453, 620–625 (2008).
- Round, J.L. et al. The Toll-like receptor 2 pathway establishes colonization by a commensal of the human microbiota. Science 332, 974–977 (2011).
- 77. Olszak, T. *et al.* Microbial exposure during early life has persistent effects on natural killer T cell function. *Science* **336**, 489–493 (2012).
- An, D. et al. Sphingolipids from a symbiotic microbe regulate homeostasis of host intestinal natural killer T cells. Cell 156, 123–133 (2014).
- Kawano, T. et al. CD1d-restricted and TCR-mediated activation of va14 NKT cells by glycosylceramides. Science 278, 1626–1629 (1997).
- Greer, R.L., Morgun, A. & Shulzhenko, N. Bridging immunity and lipid metabolism by gut microbiota. *J. Allergy Clin. Immunol.* 132, 253–262, quiz 263 (2013).
- Rossi, M., Amaretti, A. & Raimondi, S. Folate production by probiotic bacteria. Nutrients 3, 118–134 (2011).
- 82. Kjer-Nielsen, L. et al. MR1 presents microbial vitamin B metabolites to MAIT cells. Nature 491, 717–723 (2012).
- Bansal, T., Alaniz, R.C., Wood, T.K. & Jayaraman, A. The bacterial signal indole increases epithelial-cell tight-junction resistance and attenuates indicators of inflammation. *Proc. Natl. Acad. Sci. USA* 107, 228–233 (2010).
- Shimada, Y. et al. Commensal bacteria-dependent indole production enhances epithelial barrier function in the colon. PLoS ONE 8, e80604 (2013).
- 85. Fukuda, S. *et al.* Bifidobacteria can protect from enteropathogenic infection through production of acetate. *Nature* **469**, 543–547 (2011).
- Inan, M.S. et al. The luminal short-chain fatty acid butyrate modulates NF-κB activity in a human colonic epithelial cell line. Gastroenterology 118, 724–734 (2000).
- Smith, P.M. et al. The microbial metabolites, short-chain fatty acids, regulate colonic T_{reg} cell homeostasis. Science 341, 569–573 (2013).
- Arpaia, N. et al. Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. Nature 504, 451–455 (2013).
- 89. Sakaguchi, S., Yamaguchi, T., Nomura, T. & Ono, M. Regulatory T cells and immune tolerance. *Cell* 133, 775–787 (2008).
- 90. Furusawa, Y. et al. Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. *Nature* **504**, 446–450 (2013).

- 91. Kim, S.V. et al. GPR15-mediated homing controls immune homeostasis in the large intestine mucosa. *Science* **340**, 1456–1459 (2013).
- Frank, D.N. et al. Molecular-phylogenetic characterization of microbial community imbalances in human inflammatory bowel diseases. Proc. Natl. Acad. Sci. USA 104. 13780–13785 (2007).
- 93. Trompette, A. et al. Gut microbiota metabolism of dietary fiber influences allergic airway disease and hematopoiesis. Nat. Med. 20, 159–166 (2014).
- 94. Blumberg, R. & Powrie, F. Microbiota, disease, and back to health: a metastable journey. Sci. Transl. Med. 4, 137rv137 (2012).
- 95. Vijay-Kumar, M. *et al.* Metabolic syndrome and altered gut microbiota in mice lacking Toll-like receptor 5. *Science* **328**, 228–231 (2010).
- Tillisch, K. et al. Consumption of fermented milk product with probiotic modulates brain activity. Gastroenterology 144, 1394–1401 (2013).
- Hsiao, E.Y. et al. Microbiota modulate behavioral and physiological abnormalities associated with neurodevelopmental disorders. Cell 155, 1451–1463 (2013).
- Wang, L., Angley, M.T., Gerber, J.P. & Sorich, M.J. A review of candidate urinary biomarkers for autism spectrum disorder. *Biomarkers* 16, 537–552 (2011)
- Donohoe, D.R. et al. The Warburg effect dictates the mechanism of butyratemediated histone acetylation and cell proliferation. Mol. Cell 48, 612–626 (2012).

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Competing financial interests

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REVIEW

Commensal microbiota regulates T cell fate decision in the gut

Yukihiro Furusawa · Yuuki Obata · Koji Hase

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Abstract Commensal microbiota shapes the intestinal immune system by regulating T helper (T_H) cell lineage differentiation. For example, Bacteroides fragilis colonization not only optimizes the systemic T_H1/T_H2 balance, but also can induce regulatory T (Treg) cell differentiation in the gut. In addition, segmented filamentous bacteria (SFB) facilitate the development of T_H17 cells in the small intestine. The 17 strains within clusters IV, XIVa, and XVIII of Clostridiales found in human feces can also induce the differentiation and expansion of Treg cells in the colon. Thus, the regulation of T_H cell differentiation by commensal bacteria is evident; however, the molecular mechanisms underlying these processes remain uncertain. Recent studies have demonstrated that bacterial components, as well as their metabolites, play a central role in regulating T_H cell development. Furthermore, these metabolites can elicit changes in histone posttranslational modification to modify the expression of critical regulators of T cell fate. In this review, we discuss the mechanisms and biological significance of microbiota-dependent T_H differentiation.

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Y. Furusawa · K. Hase (⋈)
Department of Biochemistry, Graduate School of Pharmaceutical
Science, Keio University, Tokyo 105-8512, Japan
e-mail: hase@ims.u-tokyo.ac.jp

Y. Furusawa Y. Obata K. Hase The Institute of Medical Science, The University of Tokyo, Tokyo 108-8639, Japan

Y. Obata Graduate School of Medicine, Chiba University, Chiba 260-8670, Japan **Keywords** Microbiota \cdot T_H17 cells \cdot Regulatory T cells \cdot Histone modifications \cdot DNA methylation

Introduction

Human beings harbor over 100 trillion bacterial species, comprising more than 1000 strains, in the distal intestine. Commensal bacteria facilitate the breakdown of food necessary for energy metabolism in their hosts [1, 2]. Compelling evidence suggests that colonization by commensal bacteria critically contributes to the development of the mammalian immune system. Given that patients with inflammatory bowel disease (IBD) often present with dysbiosis [3], or an imbalance of commensal bacteria, the interplay between commensals and host cells may play a critical role in the maintenance of gut immune homeostasis. This concept has been partly verified by recent studies using gnotobiotic mice, in which germ-free (GF) mice are colonized by specific strains of commensal bacteria. So far, a select number of these strains have been documented to induce the development of particular immune mediators, such as type 17 helper T cells (TH17) and regulatory T (Treg) cells [4–6]. Although the mechanisms by which gut microbiota regulates host immunity have yet to be elucidated, their derived metabolites can partially mediate this immunomodulatory effect. Such bioactive materials are produced by microbiota through both diet-dependent and dietindependent means [7]. Here, we discuss the molecular interactions between commensal microbiota and TH cell development.

Development of effector T_H cells by commensal bacteria

Systemic immune responses are biased to T_H2 under neonatal and GF conditions [4, 8, 9]. Microbial exposure early in life is