

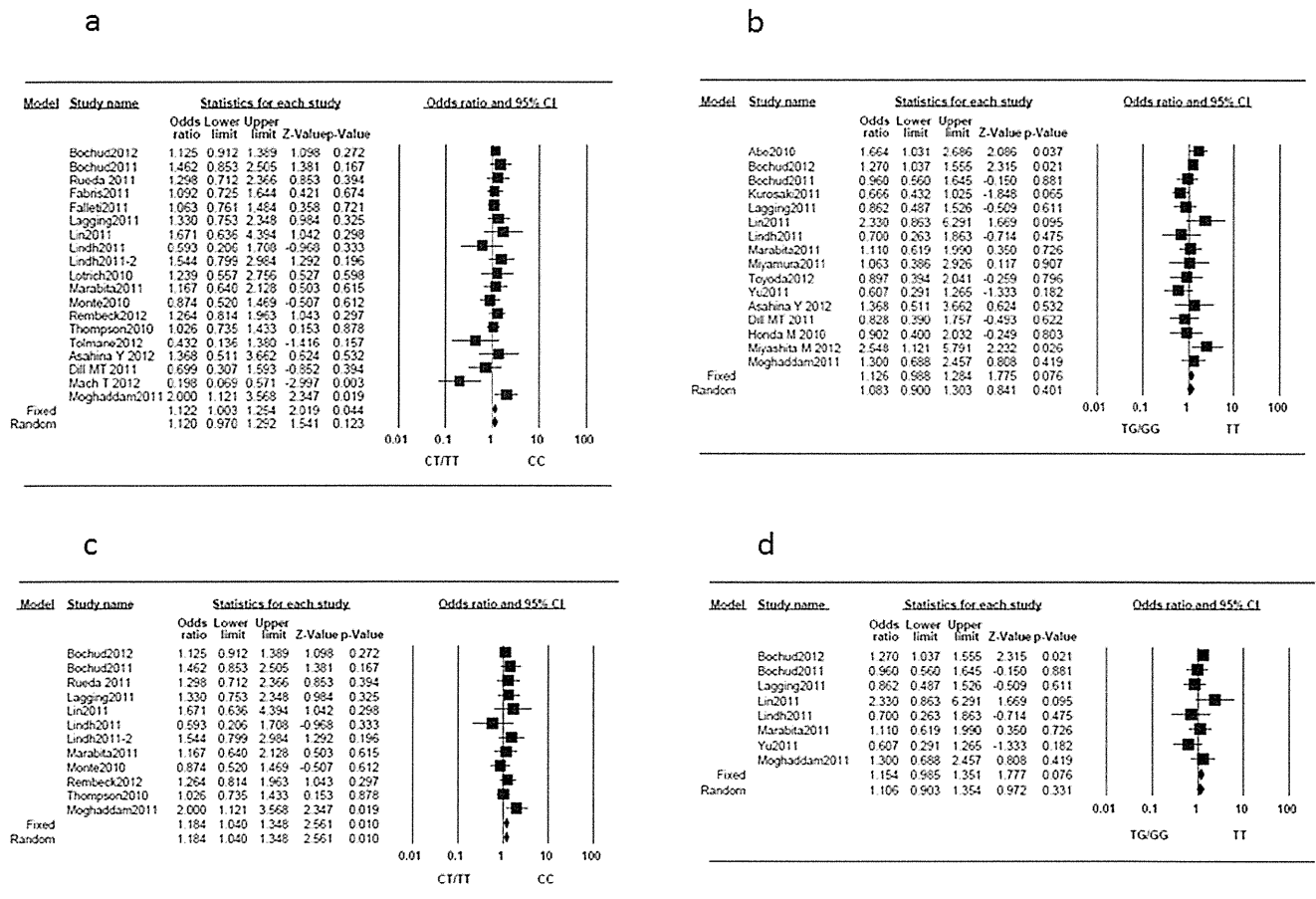
**Table 1.** Main characteristics of all studies included in the meta-analysis.

First author (year)	Ref.	Population ethnicity, region	IL-28B SNP rsID, Allele	Outcome measure F(Fibrosis) A(Activity) S(Steatosis)	Patients*			HCV genotype	Genotype for patients rs12979860		Genotype for patients rs8099917	
					Male	Female	Total		CC	CT/TT	TT	TG/GG
Abe (2010)	[48]	Asian, Japan	rs8099917 T/G	F, A: Inuyama	212	152	364	1/2			265	99
Honda (2010)	[49]	Asian, Japan	rs8099917 T/G	F, A: Inuyama	58	33	91	1			60	31
Lotrich (2010)	[50]	Mixed (African-American/Caucasian), USA	rs12979860 C/T	F: Ishak	101	32	133	1/2	57	76		
Monte (2010)	[51]	Caucasian, Spain	rs12979860 C/T	F: Scheuer	166	117	283	1–4	129	154		
Thompson (2010)	[52]	Mixed (African-American/Caucasian/Asian/Hispanic), USA	rs12979860 C/T	F: METAVIR	986	642	1628	1	538	1090		
Bochud (2011)	[53]	Caucasian, Switzerland	rs12979860 C/T rs8099917 T/G	F: Ishak, A: ALT S: 163 Histological finding		79	242	1–3	90	150	150	92
Dill MT (2011)	[54]	Caucasian, Switzerland	rs12979860 C/T rs8099917 T/G	F, A: METAVIR	30	79	109	1–4	33	96	52	57
Fabris (2011)	[44]	Caucasian, Italy	rs12979860 C/T	F: Ishak	N.A	N.A	434	1–4	133	301		
Falleti (2011)	[55]	Caucasian, Italy	rs12979860 C/T	F: Ishak	357	272	629	1–4	205	424		
Kurosaki (2011)	[56]	Asian, Japan	rs8099917 T/G	F: METAVIR S: Histological finding	250	246	496	1			269	106
Lagging (2011)	[57]	Caucasian, Sweden	rs12979860 C/T rs8099917 T/G	F: Ishak S: Histological finding	169	83	252	1–4	93	159	153	99
Lin (2011)	[58]	Asian, Taiwan	rs12979860 C/T rs8099917 T/G	F: METAVIR	123	68	191	1	171	20	170	21
Lindh (2011)-1	[59]	Mixed (Caucasian/Asian), Sweden	rs12979860 C/T rs8099917 T/G	F: Batts Ludwig	67	43	110	1	38	72	66	44
Lindh (2011)-2	[60]	Caucasian, Sweden	rs12979860 C/T	F: Ishak	204	137	341	2/3	150	191		
Marabita (2011)	[61]	Caucasian, Italy	rs12979860 C/T rs8099917 T/G	F: Ishak	129	118	247	1–4	88	159	131	116
Miyamura (2011)	[62]	Asian, Japan	rs8099917 T/G	F, A: Inuyama	37	42	79	1			53	26
Moghaddam(2011)	[63]	Caucasian, Norway	rs12979860 C/T rs8099917 T/G	F: APRI score	166	115	281	3	129	152	201	80
Rueda (2011)	[64]	Caucasian, Spain	rs12979860 C/T	F, A: Scheuer	246	177	423	1–4	83	184		
Tillman (2011)	[35]	Mixed (African-American/Caucasian/Asian), USA	rs12979860 C/T rs8099917 T/G	S: Histological finding	215	110	325	1	88	237	97	67
Yu (2011)	[65]	Asian, Taiwan	rs8099917 T/G	F: Knodell and Scheuer	264	218	482	2			315	34
Asahina (2011)	[66]	Asian, Japan	rs12979860 C/T rs8099917 T/G	F: Inuyama	28	60	88	1	54	34	54	34

**Table 1.** Cont.

First author (year)	Ref.	Population ethnicity, region	IL-28B SNP rsID, Allele	Outcome measure F(Fibrosis) A(Activity) S(Steatosis)	Patients*			HCV genotype	Genotype for patients rs12979860		Genotype for patients rs8099917	
					Male	Female	Total		CC	CT/TT	TT	TG/GG
Bochud (2012)	[47]	Caucasian, Switzerland	rs12979860 C/T rs8099917 T/G	F, A: METAVIR	870	657	1527	1–4	534	993	855	672
Mach (2012)	[67]	Slav: Poland	rs12979860 C/T	F: Batts Ludwig	82	60	142	1	38	104		
Miyashita (2012)	[68]	Asian, Japan	rs8099917 T/G	F, A: Desmet	88	132	220	1/2			155	63
Ohnishi (2012)	[69]	Asian, Japan	rs8099917 T/G	S: Histological finding	83	70	153	1			116	37
Rembeck (2012)	[70]	Caucasian, Sweden	rs12979860 C/T	F: Ishak	199	140	339	2/3	144	179		
Tolmane (2012)	[71]	Caucasian, Latvia	rs12979860 C/T	F: Knodell histology activity index S: Histological finding	84	58	142	1–3	41	80		
Toyoda (2012)	[72]	Asian, Japan	rs8099917 T/G	F, A: METAVIR	139	133	272	1			187	59

\*Patients included in the original study. Thus, patients without information regarding IL28B polymorphism were also included. APRI, aminotransferase platelet ratio index. doi:10.1371/journal.pone.0091822.t001



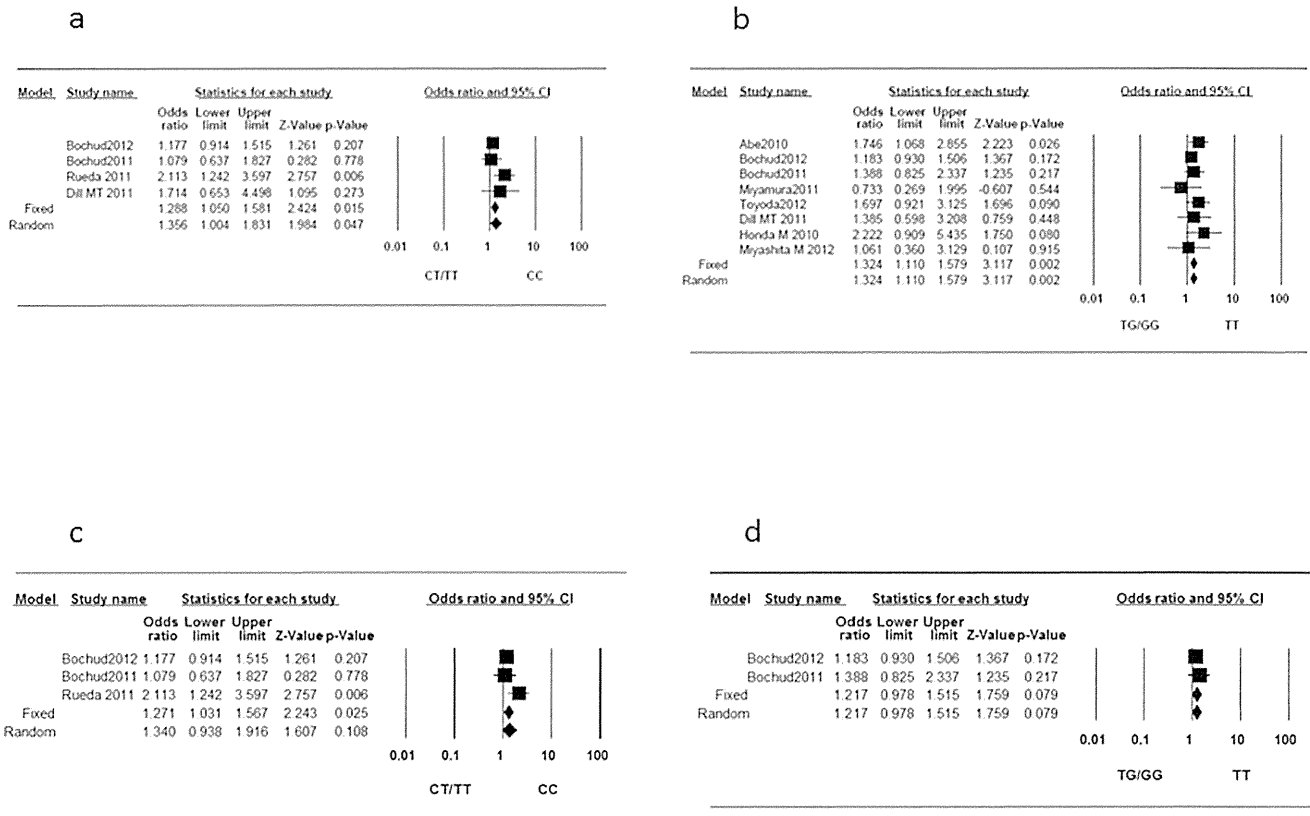
**Figure 2. Forest plot of the IL28B genotypes and the risk of severe fibrosis.** (a) rs12979860 in all patients, (b) rs8099917 in all patients, (c) rs12979860 in treatment-naïve patients, and (d) rs8099917 in treatment-naïve patients. doi:10.1371/journal.pone.0091822.g002

strong predictors of a sustained viral response [17–20] or spontaneous clearance of HCV [21]. The level of IL28B gene transcripts is reportedly higher in patients homozygous for the IFN responsive allele [18,19]. Therefore, in patients with the rs12979860 CC and rs8099917 TT genotype, IL28B production, which induces expression of interferon-stimulated genes, including some inflammatory cytokines, was thought to be increased. This may be the underlying cause of the higher inflammation activity and progressed fibrosis in patients with the IFN responsive allele. In analysis with the studies involving only patients without a history of IFN-based treatment, rs12979860 CC and rs8099917 TT genotypes were associated with higher possibility of having severe inflammation activity; however, the differences did not reach to the significant level. Only three studies according to rs12979860 polymorphism and two studies according to rs8099917 polymorphism were included when restricted to studies with only treatment-naïve patients, and may be underpowered to detect the effects of IL28B polymorphisms on inflammation activity. The further analyses with larger sample are needed to confirm this association. Additionally, meta-regression analysis showed that the effect of the rs12979860 polymorphism was influenced by viral genotype distribution. This result may imply a different influence of rs12979860 polymorphism on immune response according to viral genotype in treatment-naïve patients.

IL28B polymorphisms were also shown to be associated with lipid metabolism [25]. In the present study, the rs8099917 TT

genotype was significantly associated with a lower possibility of severe steatosis. This association still remained statistically significant after we restricted to studies in which only treatment-naïve patients were included. The lower hepatic steatosis in patients with the IFN responsive allele could be explained by a more efficient export of lipids from hepatocytes. Higher interferon expression was shown to lead to suppression of lipoprotein lipase, which would result in decreased conversion of VLDL to LDL and subsequent higher steatosis [30–33]. The difference in IL28B expression might cause an aberration of lipid metabolism in patients with CHC. We found no significant association of rs12979860 with steatosis. And when we restricted to treatment-naïve patients, rs12979860 CC genotype was significantly associated with a higher possibility of severe steatosis. Previous studies have shown that racial differences or viral genotypes make a difference in the effects of rs12979860 and rs8099917 polymorphisms [34,35]. This may explain the discrepancy between the effect of rs12979860 and rs8099917 on hepatic steatosis. However, only four studies (962 patients) were included in the analysis of rs12979860; or when it comes to the studies with only treatment-naïve patients, only two studies (495 patients) were extracted. Thus, we should not make any definite conclusion on this matter right now. Further studies with larger sample sizes are needed to identify their exact correlation.

According to the meta-regression analysis, the effect of rs8099917 polymorphisms on steatosis became smaller with the



**Figure 3. Forest plot of the IL28B genotypes and the risk of severe inflammation activity.** (a) rs12979860 and (b) rs8099917. (c) rs12979860 in treatment-naïve patients, and (d) rs8099917 in treatment-naïve patients. doi:10.1371/journal.pone.0091822.g003

increase in the male proportion (Fig. 5), suggesting that a sexual dimorphism might be involved in the effect of rs8099917 polymorphisms on the liver fat content. Although the present study cannot explain the interaction between the polymorphism and sex, immune systems responding to IFN are reportedly controlled by estrogenic sex hormones [36,37]. Differences in IL28B expression mediated by sex hormones could be a possible

mechanism for the sexual dimorphism in the effect of rs8099917 polymorphisms on liver steatosis.

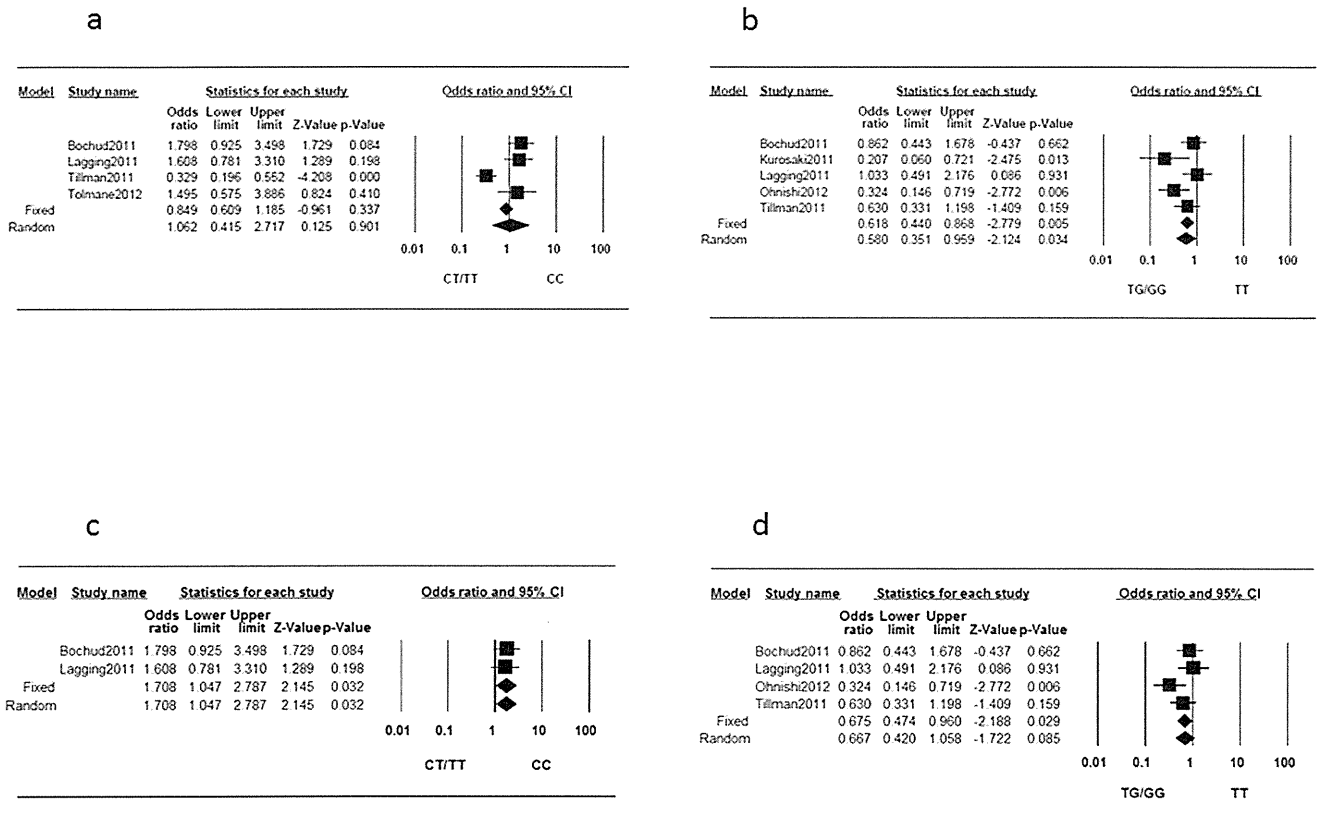
The rs738409 genotype within the patatin-like phospholipase domain containing 3 locus was also reported to be associated with hepatic steatosis in patients with CHC [38–40]. Notably, previous meta-analysis evaluating the effect of patatin-like phospholipase domain containing 3 polymorphisms on steatosis also reported a

**Table 2. Meta-regression analysis between each continuous variable among the studies (only treatment-naïve patients were included) and the effect (log odds ratio) of IL28B polymorphisms on inflammation activity.**

Variables	Slope*	Standard error	P-value
<b>Proportion of patients with genotype 1 or 4 virus, per 1% increase</b>			
rs12979860	2.992	1.497	0.046
<b>Proportion of male patients, per 1% increase</b>			
rs12979860	-2.963	5.802	0.610
<b>Proportion of Caucasian patients, per 1% increase</b>			
rs12979860†	—	—	—
<b>Proportion of African-American patients, per 1% increase</b>			
rs12979860†	—	—	—
<b>Proportion of Asian patients, per 1% increase</b>			
rs12979860†	—	—	—

\*Positive (negative) slope values indicate that the proportions of patients with the rs12979860 CC genotype with severe inflammation activity are increasing (decreasing) as the values of each contentions variable (proportions of genotype 1 or 4 virus, male, or each race) is increasing.

†We could not perform meta-regression analyses for these outcomes because only caucasian patients were included in all 3 studies included in this analysis. doi:10.1371/journal.pone.0091822.t002



**Figure 4. Forest plot of the IL28B genotypes and the risk of hepatic steatosis.** (a) rs12979860 and (b) rs8099917. (c) rs12979860 in treatment-naïve patients, and (d) rs8099917 in treatment-naïve patients. doi:10.1371/journal.pone.0091822.g004

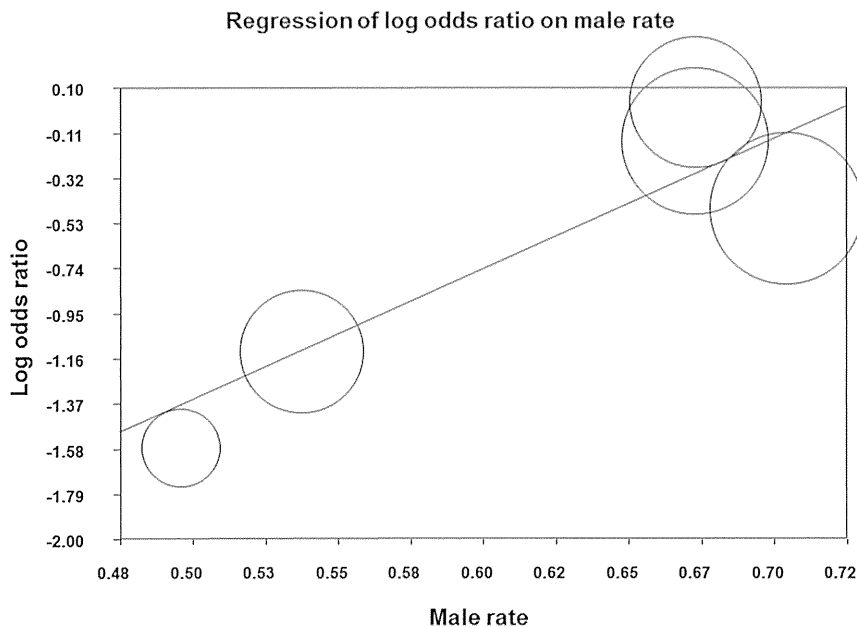
**Table 3. Meta-regression analysis between each continuous variable among the studies and the effect (log odds ratio) of IL28B polymorphisms on steatosis.**

Variables	Slope*	Standard error	P-value
Proportion of patients with genotype 1 or 4 virus, per 1% increase			
rs12979860	-4.947	1.086	<0.001
rs8099917	-2.704	1.277	0.034
Proportion of male patients, per 1% increase			
rs12979860	-2.899	16.577	0.861
rs8099917	6.225	2.530	0.014
Proportion of Caucasian patients, per 1% increase			
rs12979860	7.361	1.569	<0.001
rs8099917	1.168	0.422	0.006
Proportion of African-American patients, per 1% increase			
rs12979860	-8.996	1.918	<0.001
rs8099917	0.142	2.147	0.947
Proportion of Asian patients, per 1% increase			
rs12979860†	-	-	-
rs8099917	-1.049	0.398	0.008

\*Positive (negative) slope values indicate that the proportions of patients with the rs12979860 CC or rs8099917 TT genotypes with severe steatosis are increasing (decreasing) as the values of each contentions variable (proportions of genotype 1 or 4 virus, male, or each race) is increasing.

†We could not perform a meta-regression analysis for this outcome because only one patient was included in the corresponding studies.

doi:10.1371/journal.pone.0091822.t003



**Figure 5. Meta-regression plot for log odds ratios in rates of patients with severe hepatic steatosis by proportion of males (%) in rs8099917.**

doi:10.1371/journal.pone.0091822.g005

negative correlation between the male proportion and the effect of rs738409 on the liver fat content in nonalcoholic fatty liver disease [41]. Interestingly, the meta-regression analysis in the present study showed that the effect of the IL28B (rs12979860 and rs8099917) polymorphisms on steatosis was also influenced by racial and viral genotype distributions.

In the present study, we included studies that did not report the associations between IL28B genotypes and background liver diseases as study outcomes, but provided raw data that allowed us to calculate the OR for each outcome, which may have minimized potential publication bias. In fact, no publication bias was observed in the present study. The Human Genome Epidemiology Network highlighted the necessity of meta-analysis before evidence for a particular association can be regarded as strong [42]. The impact of IL28B genotypes on the disease progression found in the present meta-analysis may provide clinically important information in the follow-up of patients with CHC. The effect of IL28B polymorphisms on hepatocarcinogenesis, which is also crucial information in the HCC screening of patients with CHC, remains controversial [43–47]. Further analysis with larger sample sizes may be needed to elucidate the exact effect of IL28B polymorphisms on hepatocarcinogenesis.

A potential limitation of this study is inter-study variability in the outcome measure and the definition of “severe” among studies, where some discrepancies among studies exist. The studies without a pathological diagnosis, using laboratory data as

surrogates, were also included. These studies may have diminished the accuracy of our research results concerning liver disease severity.

In conclusion, the present study highlighted the impact of IL28B polymorphisms on liver fibrosis, inflammation activity, and steatosis in patients with CHC. Disease progression appeared to be promoted in patients with rs12979860 GC or rs8099917 TT genotypes. The current findings may provide clinically important information in the follow-up of patients with CHC.

## Supporting Information

**Checklist S1 PRISMA 2009 Checklist.**  
(DOC)

## Acknowledgments

The English in this document has been checked by at least two professional editors, both native speakers of English. For a certificate, please see: <http://www.textcheck.com/certificate/IWcYpT>.

## Author Contributions

Conceived and designed the experiments: MS RT NK. Performed the experiments: MS MK RT. Analyzed the data: MS RT. Contributed reagents/materials/analysis tools: MS. Wrote the paper: MS RT HY. Critical revision of manuscript: NF MT KK.

## References

- Barrera JM, Bruguera M, Ercilla MG, Gil C, Celis R, et al. (1995) Persistent hepatitis C viremia after acute self-limiting posttransfusion hepatitis C. *Hepatology* 21: 639–644.
- Poynard T, Bedossa P, Opolon P (1997) Natural history of liver fibrosis progression in patients with chronic hepatitis C. The OBSVIRC, METAVIR, CLINIVIR, and DOSVIRC groups. *Lancet* 349: 825–832.
- Hourigan LF, Macdonald GA, Purdie D, Whitehall VH, Shorthouse C, et al. (1999) Fibrosis in chronic hepatitis C correlates significantly with body mass index and steatosis. *Hepatology* 29: 1215–1219.
- Powell EE, Edwards-Smith CJ, Hay JL, Clouston AD, Crawford DH, et al. (2000) Host genetic factors influence disease progression in chronic hepatitis C. *Hepatology* 31: 828–833.
- Massard J, Ratziu V, Thabut D, Moussalli J, Lebray P, et al. (2006) Natural history and predictors of disease severity in chronic hepatitis C. *J Hepatol* 44: S19–24.
- Bochud PY, Cai T, Overbeck K, Bochud M, Dufour JF, et al. (2009) Genotype 3 is associated with accelerated fibrosis progression in chronic hepatitis C. *J Hepatol* 51: 655–666.

7. De Nicola S, Aghemo A, Rumi MG, Colombo M (2009) HCV genotype 3: an independent predictor of fibrosis progression in chronic hepatitis C. *J Hepatol* 51: 964–966.
8. Thursz M, Yallop R, Goldin R, Trepo C, Thomas HC (1999) Influence of MHC class II genotype on outcome of infection with hepatitis C virus. The HENCORE group. *Hepatitis C European Network for Cooperative Research. Lancet* 354: 2119–2124.
9. Pradat P, Tillmann HL, Sauleda S, Braconier JH, Saracco G, et al. (2007) Long-term follow-up of the hepatitis C HENCORE cohort: response to therapy and occurrence of liver-related complications. *J Viral Hepat* 14: 556–563.
10. Kato N, Ji G, Wang Y, Baba M, Hoshida Y, et al. (2005) Large-scale search of single nucleotide polymorphisms for hepatocellular carcinoma susceptibility genes in patients with hepatitis C. *Hepatology* 42: 846–853.
11. Urabe Y, Ochi H, Kato N, Kumar V, Takahashi A, et al. (2013) A genome-wide association study of HCV-induced liver cirrhosis in the Japanese population identifies novel susceptibility loci at the MHC region. *J Hepatol*; 58 (5): 875–82
12. Patin E, Kutalik Z, Guernon J, Bibert S, Nalpas B, et al. (2012) Genome-Wide Association Study Identifies Variants Associated with Progression of Liver Fibrosis from HCV Infection. *Gastroenterology*; 143 (5): 1244–52
13. Hadziyannis SJ, Sette H Jr, Morgan TR, Balan V, Diago M, et al. (2004) Peginterferon-alpha2a and ribavirin combination therapy in chronic hepatitis C: a randomized study of treatment duration and ribavirin dose. *Ann Intern Med* 140: 346–355.
14. Manns MP, McHutchison JG, Gordon SC, Rustgi VK, Shiffman M, et al. (2001) Peginterferon alfa-2b plus ribavirin compared with interferon alfa-2b plus ribavirin for initial treatment of chronic hepatitis C: a randomised trial. *Lancet* 358: 958–965.
15. McHutchison JG, Everson GT, Gordon SC, Jacobson IM, Sulkowski M, et al. (2009) Telaprevir with peginterferon and ribavirin for chronic HCV genotype 1 infection. *N Engl J Med* 360: 1827–1838.
16. Poordad F, McCone J Jr, Bacon BR, Bruno S, Manns MP, et al. (2011) Boceprevir for untreated chronic HCV genotype 1 infection. *N Engl J Med* 364: 1195–1206.
17. Ge D, Fellay J, Thompson AJ, Simon JS, Shianna KV, et al. (2009) Genetic variation in IL28B predicts hepatitis C treatment-induced viral clearance. *Nature* 461: 399–401.
18. Tanaka Y, Nishida N, Sugiyama M, Kurosaki M, Matsuura K, et al. (2009) Genome-wide association of IL28B with response to pegylated interferon-alpha and ribavirin therapy for chronic hepatitis C. *Nat Genet* 41: 1105–1109.
19. Suppiah V, Moldovan M, Ahlenstiel G, Berg T, Weltman M, et al. (2009) IL28B is associated with response to chronic hepatitis C interferon-alpha and ribavirin therapy. *Nat Genet* 41: 1100–1104.
20. Rauch A, Kutalik Z, Descombes P, Cai T, Di Iulio J, et al. (2010) Genetic variation in IL28B is associated with chronic hepatitis C and treatment failure: a genome-wide association study. *Gastroenterology* 138: 1338–1345, 1345 e1331–1337.
21. Thomas DL, Thio CL, Martin MP, Qi Y, Ge D, et al. (2009) Genetic variation in IL28B and spontaneous clearance of hepatitis C virus. *Nature* 461: 798–801.
22. Yoneyama M, Kikuchi M, Natsukawa T, Shinobu N, Imaizumi T, et al. (2004) The RNA helicase RIG-I has an essential function in double-stranded RNA-induced innate antiviral responses. *Nat Immunol* 5: 730–737.
23. Moriyama M, Kato N, Otsuka M, Shao RX, Taniguchi H, et al. (2007) Interferon-beta is activated by hepatitis C virus NS5B and inhibited by NS4A, NS4B, and NS5A. *Hepatology Int* 1: 302–310.
24. Li CZ, Kato N, Chang JH, Muroyama R, Shao RX, et al. (2009) Polymorphism of OAS-1 determines liver fibrosis progression in hepatitis C by reduced ability to inhibit viral replication. *Liver Int* 29: 1413–1421.
25. Li JH, Lao XQ, Tillmann HL, Rowell J, Patel K, et al. (2010) Interferon-lambda genotype and low serum low-density lipoprotein cholesterol levels in patients with chronic hepatitis C infection. *Hepatology* 51: 1904–1911.
26. Borenstein M, Hedges LV, Higgins JP, Rothstein HR (2009) *Introduction to Meta-analysis*. West Sussex: John Wiley & Sons Ltd.
27. Baker WL, White CM, Cappelleri JC, Kluger J, Coleman CI (2009) Understanding heterogeneity in meta-analysis: the role of meta-regression. *Int J Clin Pract* 63: 1426–1434.
28. Thompson SG, Sharp SJ (1999) Explaining heterogeneity in meta-analysis: a comparison of methods. *Stat Med* 18: 2693–2708.
29. Egger M, Davey Smith G, Schneider M, Minder C (1997) Bias in meta-analysis detected by a simple, graphical test. *BMJ* 315: 629–634.
30. Schectman G, Kaul S, Mueller RA, Borden EC, Kissebah AH (1992) The effect of interferon on the metabolism of LDLs. *Arterioscler Thromb* 12: 1053–1062.
31. Ehnholm C, Aho K, Huttunen JK, Kostiaainen E, Mattila K, et al. (1982) Effect of interferon on plasma lipoproteins and on the activity of postheparin plasma lipases. *Arteriosclerosis* 2: 68–73.
32. Shinohara E, Yamashita S, Kihara S, Hirano K, Ishigami M, et al. (1997) Interferon alpha induces disorder of lipid metabolism by lowering postheparin lipases and cholesteryl ester transfer protein activities in patients with chronic hepatitis C. *Hepatology* 25: 1502–1506.
33. Andrade RJ, Garcia-Escano MD, Valdivielso P, Alcantara R, Sanchez-Chaparro MA, et al. (2000) Effects of interferon-beta on plasma lipid and lipoprotein composition and post-heparin lipase activities in patients with chronic hepatitis C. *Aliment Pharmacol Ther* 14: 929–935.
34. Sarrazin C, Susser S, Doehring A, Lange CM, Muller T, et al. (2011) Importance of IL28B gene polymorphisms in hepatitis C virus genotype 2 and 3 infected patients. *J Hepatol* 54: 415–421.
35. Tillmann HL, Patel K, Muir AJ, Guy CD, Li JH, et al. (2011) Beneficial IL28B genotype associated with lower frequency of hepatic steatosis in patients with chronic hepatitis C. *J Hepatol*; 55 (6): 1195–200
36. Nakaya M, Tachibana H, Yamada K (2006) Effect of estrogens on the interferon-gamma producing cell population of mouse splenocytes. *Biosci Biotechnol Biochem* 70: 47–53.
37. Siracusa MC, Overstreet MG, Housseau F, Scott AL, Klein SL (2008) 17beta-estradiol alters the activity of conventional and IFN-producing killer dendritic cells. *J Immunol* 180: 1423–1431.
38. Cai T, Dufour JF, Muellhaupt B, Gerlach T, Heim M, et al. (2011) Viral Genotype-Specific Role of PNPLA3, PPARG, MTTP and IL28B in Hepatitis C Virus-Associated Steatosis. *J Hepatol*.
39. Trepo E, Pradat P, Potthoff A, Momozawa Y, Quertinmont E, et al. (2011) Impact of patatin-like phospholipase-3 (rs738409 C>G) polymorphism on fibrosis progression and steatosis in chronic hepatitis C. *Hepatology* 54: 60–69.
40. Valenti L, Rumi M, Galmozzi E, Aghemo A, Del Menico B, et al. (2011) Patatin-like phospholipase domain-containing 3 I148M polymorphism, steatosis, and liver damage in chronic hepatitis C. *Hepatology* 53: 791–799.
41. Sookoian S, Pirola CJ (2011) Meta-analysis of the influence of I148M variant of the patatin-like phospholipase domain containing 3 gene (PNPLA3) on the susceptibility and histological severity of nonalcoholic fatty liver disease. *Hepatology* 53: 1883–1894.
42. Ioannidis JP, Boffetta P, Little J, O'Brien TR, Uitterlinden AG, et al. (2008) Assessment of cumulative evidence on genetic associations: interim guidelines. *Int J Epidemiol* 37: 120–132.
43. Asahina Y, Tanaka K, Suzuki Y, Tamaki N, Hoshioka T, et al. (2011) Association between IL28B gene variation and development of hepatocellular carcinoma after interferon therapy in patients with chronic hepatitis c. *Journal of Hepatology* 54: S37.
44. Fabris C, Falletti E, Cussigh A, Bitetto D, Fontanini E, et al. (2011) IL-28B rs12979860 C/T allele distribution in patients with liver cirrhosis: role in the course of chronic viral hepatitis and the development of HCC. *J Hepatol* 54: 716–722.
45. Joshita S, Umemura T, Katsuyama Y, Ichikawa Y, Kimura T, et al. (2011) Association of IL28B gene polymorphism with development of hepatocellular carcinoma in Japanese patients with chronic hepatitis C virus infection. *Hum Immunol*; 73 (3): 298–300
46. Miura M, Maekawa S, Kadokura M, Sueki R, Komase K, et al. (2011) Analysis of viral amino acids sequences and the IL28B SNP influencing the development of hepatocellular carcinoma in chronic hepatitis C. *Hepatology Int*; Aug 17 [Epub ahead of print].
47. Bochud PY, Bibert S, Kutalik Z, Patin E, Guernon J, et al. (2011) IL28B alleles associated with poor hepatitis C virus (HCV) clearance protect against inflammation and fibrosis in patients infected with non-1 HCV genotypes. *Hepatology*; 55 (2): 384–94
48. Abe H, Ochi H, Maekawa T, Hayes CN, Tsuge M, et al. (2010) Common variation of IL28 affects gamma-GTP levels and inflammation of the liver in chronically infected hepatitis C virus patients. *J Hepatol* 53: 439–443.
49. Honda M, Sakai A, Yamashita T, Nakamoto Y, Mizukoshi E, et al. (2010) Hepatic ISG expression is associated with genetic variation in interleukin 28B and the outcome of IFN therapy for chronic hepatitis C. *Gastroenterology* 139: 499–509.
50. Lotrich FE, Loftis JM, Ferrell RE, Rabinovitz M, Hauser P (2010) IL28B Polymorphism Is Associated with Both Side Effects and Clearance of Hepatitis C During Interferon-Alpha Therapy. *J Interferon Cytokine Res*; Dec 6 [Epub ahead of print].
51. Montes-Cano MA, Garcia-Lozano JR, Abad-Molina C, Romero-Gomez M, Barroso N, et al. (2010) Interleukin-28B genetic variants and hepatitis virus infection by different viral genotypes. *Hepatology* 52: 33–37.
52. Thompson AJ, Muir AJ, Sulkowski MS, Ge D, Fellay J, et al. (2010) Interleukin-28B polymorphism improves viral kinetics and is the strongest pretreatment predictor of sustained virologic response in genotype 1 hepatitis C virus. *Gastroenterology* 139: 120–129 e118.
53. Bochud PY, Bibert S, Negro F, Haagmans B, Soulier A, et al. (2011) IL28B polymorphisms predict reduction of HCV RNA from the first day of therapy in chronic hepatitis C. *J Hepatol*; 55 (5): 980–8.
54. Dill MT, Duong FH, Vogt JE, Bibert S, Bochud PY, et al. (2011) Interferon-induced gene expression is a stronger predictor of treatment response than IL28B genotype in patients with hepatitis C. *Gastroenterology* 140: 1021–1031.
55. Falletti E, Bitetto D, Fabris C, Cussigh A, Fornasiero E, et al. (2011) Role of Interleukin 28B rs12979860 C/T Polymorphism on the Histological Outcome of Chronic Hepatitis C: Relationship with Gender and Viral Genotype. *J Clin Immunol*; 31 (5): 891–9.
56. Kurosaki M, Tanaka Y, Nishida N, Sakamoto N, Enomoto N, et al. (2011) Pre-treatment prediction of response to pegylated-interferon plus ribavirin for chronic hepatitis C using genetic polymorphism in IL28B and viral factors. *J Hepatol* 54: 439–448.
57. Lagging M, Askari G, Negro F, Bibert S, Soderholm J, et al. (2011) Response prediction in chronic hepatitis C by assessment of IP-10 and IL28B-related single nucleotide polymorphisms. *PLoS One* 6: e17232.

58. Lin CY, Chen JY, Lin TN, Jeng WJ, Huang CH, et al. (2011) IL28B SNP rs12979860 is a critical predictor for on-treatment and sustained virologic response in patients with hepatitis C virus genotype-1 infection. *PLoS One* 6: e18322.
59. Lindh M, Lagging M, Arnholm B, Eilard A, Nilsson S, et al. (2011) IL28B polymorphisms determine early viral kinetics and treatment outcome in patients receiving peginterferon/ribavirin for chronic hepatitis C genotype 1. *J Viral Hepat* 18: e325–331.
60. Lindh M, Lagging M, Farkkila M, Langeland N, Morch K, et al. (2011) Interleukin 28B gene variation at rs12979860 determines early viral kinetics during treatment in patients carrying genotypes 2 or 3 of hepatitis C virus. *J Infect Dis* 203: 1748–1752.
61. Marabita F, Aghemo A, De Nicola S, Rumi MG, Cheroni C, et al. (2011) Genetic variation in the interleukin-28B gene is not associated with fibrosis progression in patients with chronic hepatitis C and known date of infection. *Hepatology* 54: 1127–1134.
62. Miyamura T, Kanda T, Nakamoto S, Wu S, Fujiwara K, et al. (2011) Hepatic STAT1-nuclear translocation and interleukin 28B polymorphisms predict treatment outcomes in hepatitis C virus genotype 1-infected patients. *PLoS ONE*; 6 (12): e28617.
63. Moghaddam A, Melum E, Reinton N, Ring-Larsen H, Verbaan H, et al. (2011) IL28B genetic variation and treatment response in patients with hepatitis C virus genotype 3 infection. *Hepatology* 53: 746–754.
64. de Rueda PM, Lopez-Nevot MA, Saenz-Lopez P, Casado J, Martin-Casares A, et al. (2011) Importance of Host Genetic Factors HLA and IL28B as Predictors of Response to Pegylated Interferon and Ribavirin. *Am J Gastroenterol*.
65. Yu ML, Huang CF, Huang JF, Chang NC, Yang JF, et al. (2011) Role of interleukin-28B polymorphisms in the treatment of hepatitis C virus genotype 2 infection in Asian patients. *Hepatology* 53: 7–13.
66. Asahina Y, Tsuchiya K, Muraoka M, Tanaka K, Suzuki Y, et al. (2011) Association of gene expression involving innate immunity and genetic variation in IL28B with antiviral response. *Hepatology*.
67. Mach T, Ciesla A, Sanak M, Golwacki M, Warunek W, et al. (2012) The importance of IL28B polymorphism in response to pegylated interferon (alpha) and ribavirin in chronic hepatitis caused by HCV genotype 1b. *Przegląd Gastroenterologiczny* 7: 38–42.
68. Miyashita M, Ito T, Sakaki M, Kajiwara A, Nozawa H, et al. (2012) Genetic polymorphism in cyclooxygenase-2 promoter affects hepatic inflammation and fibrosis in patients with chronic hepatitis C. *J Viral Hepat* 19: 608–614.
69. Ohnishi M, Tsuge M, Kohno T, Zhang Y, Abe H, et al. (2012) IL28B polymorphism is associated with fatty change in the liver of chronic hepatitis C patients. *J Gastroenterol* 47: 834–844.
70. Rembeck K, Alsio A, Christensen PB, Farkkila M, Langeland N, et al. (2012) Impact of IL28B-related single nucleotide polymorphisms on liver histopathology in chronic hepatitis C genotype 2 and 3. *PLoS One* 7: e29370.
71. Tolmane I, Rozentale B, Keiss J, Ivancenko L, Subnikova N, et al. (2012) Interleukin 28B Gene Polymorphism and Association with Chronic Hepatitis C Therapy Results in Latvia. *Hepat Res Treat*: 324090.
72. Toyoda H, Kumada T, Tada T, Hayashi K, Honda T, et al. (2012) Predictive value of early viral dynamics during peginterferon and ribavirin combination therapy based on genetic polymorphisms near the IL28B gene in patients infected with HCV genotype 1b. *J Med Virol* 84: 61–70.



**Original Article**

# Slight elevation of high-sensitivity C-reactive protein to predict recurrence and survival in patients with early stage hepatitis C-related hepatocellular carcinoma

Naoto Fujiwara,<sup>1</sup> Ryosuke Tateishi,<sup>1</sup> Hayato Nakagawa,<sup>1</sup> Ryo Nakagomi,<sup>1</sup> Mayuko Kondo,<sup>1</sup> Tatsuya Minami,<sup>1</sup> Masaya Sato,<sup>1</sup> Koji Uchino,<sup>1</sup> Kenichiro Enooku,<sup>1</sup> Yuji Kondo,<sup>1</sup> Yoshinari Asaoka,<sup>1</sup> Shuichiro Shiina,<sup>2</sup> Haruhiko Yoshida<sup>1</sup> and Kazuhiko Koike<sup>1</sup>

<sup>1</sup>Department of Gastroenterology, Graduate School of Medicine, The University of Tokyo, and <sup>2</sup>Department of Gastroenterology, Juntendo University, Tokyo, Japan

**Aim:** Hepatocellular carcinoma (HCC) is associated with chronic inflammation derived from various origins. We investigated whether high-sensitivity C-reactive protein (hsCRP) could predict recurrence and survival after curative treatment for early stage hepatitis C virus-related HCC (C-HCC).

**Methods:** We enrolled 387 patients with three or fewer C-HCC nodules, none of which exceeded 3 cm, and of Child–Pugh class A or B who underwent radiofrequency ablation. We divided the patients into high and low hsCRP groups based on the optimal cut-off value for recurrence using a split-sample method and maximally selected rank statistics. Differences in recurrence and survival rates were evaluated by the Kaplan–Meier method and the log–rank test. Hazard ratios of hsCRP were adjusted with confounding factors using a multiple Cox regression model. We also assessed the correlations between hsCRP levels and clinical parameters.

**Results:** The optimal hsCRP cut-off value was 0.08 mg/dL. The cumulative recurrence rates after 5 years in the high and low hsCRP groups were 90.0% and 82.2%, respectively ( $P = 0.028$ ), and the corresponding survival rates were 50.9% and 71.8%, respectively ( $P < 0.001$ ). Higher hsCRP was an independent predictor for recurrence (adjusted hazard ratio [aHR], 1.32; 95% confidence interval [CI], 1.03–1.67;  $P = 0.026$ ) and survival (aHR, 1.59; 95% CI, 1.14–2.22;  $P = 0.007$ ). hsCRP was correlated with central obesity as well as tumor burden and liver dysfunction.

**Conclusion:** Slight elevation of the hsCRP level, even within the normal range, can predict recurrence and survival after curative treatment among patients with early stage C-HCC.

**Key words:** curative treatment, hepatocellular carcinoma, high-sensitivity C-reactive protein, prognosis, recurrence

## INTRODUCTION

HEPATOCELLULAR CARCINOMA (HCC) is the fifth most frequently diagnosed cancer and the third most frequent cause of cancer-related death.<sup>1</sup>

*Correspondence:* Dr Ryosuke Tateishi, Department of Gastroenterology, Graduate School of Medicine, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8655, Japan. Email: [tateishi-tky@umin.ac.jp](mailto:tateishi-tky@umin.ac.jp)

*Conflict of interest:* The authors have nothing to disclose.

*Funding source:* This work was supported by Health Sciences Research Grants of The Ministry of Health, Labor and Welfare of Japan (Research on Hepatitis). No additional external funding was received for this study. The funders had no role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Received 15 February 2014; revision 13 July 2014; accepted 22 July 2014.

Chronic hepatitis C virus (HCV) infection is the predominant cause of HCC in Japan and other countries. These patients are often under surveillance for HCC because of frequent hepatocarcinogenesis; as a result, HCC is usually detected when the tumors are small and treatment is more likely to succeed.<sup>2</sup> Nevertheless, recurrence eventually occurs in most patients, which can lead to a long-term poor prognosis.<sup>3</sup> Several methods have been used to predict recurrence or survival among patients with HCC, including biomarkers,<sup>4,5</sup> but these may become less useful as HCC is more frequently diagnosed at earlier stages.

C-Reactive protein (CRP) is one of the major acute-phase proteins and is a marker of systemic inflammation. CRP has been reported to be associated with a wide range of diseases, including atherosclerosis, diabetes mellitus and various cancers.<sup>6–8</sup> Notably, high-sensitivity

CRP (hsCRP) can be used to detect low-grade inflammation, and slight elevation in the hsCRP level is therefore a useful indicator of future cardiovascular disease development even within the normal range of the conventional CRP tests.<sup>8</sup> The use of hsCRP as a prognostic predictor also has been evaluated in patients with a variety of malignancies.<sup>9,10</sup> Indeed, there was a report on the relationship between elevated hsCRP and survival of patients with HCC. However, that study included patients with advanced stage HCC, such as those with tumors of more than 5 cm in diameter, multiple nodules and portal venous tumor invasion, who were treated by various methods, including resection, ablation and transarterial chemoembolization. To clarify what represents elevated hsCRP and whether hsCRP may predict survival and recurrence in HCC patients, it would be ideal to enroll patients with early HCC treated with single-modality therapies.

The aim of this study was to assess whether the hsCRP level at the time of diagnosis of HCC could predict recurrence and long-term outcomes in patients with HCV-related HCC, classified as stage 0 or A according to the Barcelona Clinic Liver Cancer staging system (BCLC).<sup>11</sup>

## METHODS

**T**HIS RETROSPECTIVE STUDY was conducted according to the ethical guidelines for epidemiological research of the Japanese Ministry of Education, Culture, Sports, Science and Technology and the Ministry of Health, Labor and Welfare. The study design was included in a comprehensive protocol from the Department of Gastroenterology, the University of Tokyo Hospital, and approved by The University of Tokyo Medical Research Center Ethics Committee (approval no. 2058).

### Patients

Of the 750 patients undergoing percutaneous radiofrequency ablation (RFA) for naïve HCC in our hospital between January 2004 and December 2009, there were 413 patients with BCLC stage 0 or A HCC. To clarify recurrence patterns, we excluded two patients in whom RFA was only intended to reduce tumor burden because it had been judged that curative treatment, either before or after RFA, was not feasible. Serum hsCRP levels and/or other variables were unavailable for 24 patients. Thus, we reviewed the remaining 387 (94.2%) patients retrospectively.

### Laboratory data

All laboratory values recorded in this study were evaluated within 3 days prior to RFA. Because we postponed the treatment when a patient had clinical evidence of an infection at the time of admission, such as pneumonia or spontaneous bacterial peritonitis, and when the hsCRP value had substantially increased compared to the value at the last outpatient clinic prior to the admission, we could exclude patients with elevated hsCRP related to overt infection. Slightly increased hsCRP levels may reflect inapparent infection related to cirrhosis. However, such elevations of hsCRP may be useful in terms of long-term mortality prediction. Therefore, we included these patients in this study.

### Anthropometric parameters

A previous report showed that obesity influenced serum inflammatory markers in chronic hepatitis C.<sup>12</sup> Thus, we assessed the following anthropometric parameters in patients with HCC: body weight, body mass index (BMI; the weight in kilograms divided by the square of the height in meters), waist circumference and fat tissue area. The fat tissue area and waist circumference were measured in each patient by analyzing a computed tomography (CT) image at the level of the umbilicus using Slim Vision software (KGT, Tokyo, Japan).<sup>13</sup> The subcutaneous fat area (SFA) was defined as the sum of the extraperitoneal fat areas between the skin and muscle on the CT image, which showed attenuation ranging from -150 to -50 Hounsfield units. The visceral fat area (VFA) was defined as the sum of the intraperitoneal fat areas showing the same attenuation.

### Diagnosis of HCC

Hepatocellular carcinoma was diagnosed using dynamic CT, considering hyperattenuation in the arterial phase with washout in the late phase as a definite sign of the disease.<sup>14</sup> The diagnosis of HCC was based on typical findings on CT, including hyperattenuation in the arterial phase and hypoattenuation in the equilibrium phase.<sup>15,16</sup> Most nodules (88.4%) were also confirmed histopathologically with ultrasound-guided biopsies. The pathological grade was determined based on the Edmondson–Steiner criteria.<sup>17</sup>

### Biopsy of non-tumorous liver

Ultrasound-guided biopsy of the non-tumorous liver was performed in 315 patients (81.4%), excluding patients with a risk of hemorrhage such as those with

severe thrombocytopenia. The background liver was pathologically graded using the METAVIR system.<sup>18</sup>

### Treatment and follow up

The inclusion criteria for percutaneous ablation were as follows: total bilirubin concentration of less than 3.0 mg/dL, platelet count of  $50 \times 10^3/\text{mm}^3$  or more, and prothrombin activity of 50% or more. Patients with portal vein tumor thrombosis, massive refractory ascites or extrahepatic metastasis were excluded. Generally, we performed percutaneous ablation in patients with three or fewer lesions, all of which were 3.0 cm or less in diameter. However, we also performed ablation in patients with more than three lesions or lesions of more than 3.0 cm if the procedure was considered to be beneficial clinically. This procedure has been described elsewhere.<sup>19</sup> After several sessions of percutaneous ablation, dynamic CT was performed with a section thickness of 0.5 cm to evaluate treatment efficacy. Complete ablation was defined as hypoattenuation of the whole lesion along with the surrounding liver parenchyma. We usually attempt to ablate an area larger than the size of the tumors with consideration of possible underestimation of the tumor boundary by imaging modalities. Patients received additional sessions of ablation until complete ablation was confirmed for each HCC nodule. The follow up comprised monthly blood tests and monitoring of tumor markers at the outpatient clinic, with ultrasonography and dynamic CT scans performed every 4 months. Tumor recurrence was diagnosed according to the same criteria as the initial HCC. Chest CT or bone scintigraphy was performed if extrahepatic recurrence was suspected. RFA was used for the treatment of recurrence if the patient still met the indication criteria. If multiple recurrences were not treatable by RFA, we typically performed transcatheter arterial chemoembolization.

### Analysis of recurrence and survival

Recurrence time was defined as the interval between the first ablation and the detection of HCC recurrence or the last examination before 31 December 2012, whichever came first. Survival analysis was performed on a patient basis. Survival time was defined as the interval between the first treatment and death or the last visit to the outpatient clinic prior to 31 December 2012.

### Statistical analyses

Quantitative variables are expressed as medians and interquartile ranges (IQR) unless otherwise indicated.

Numbers and percentages are used for qualitative variables.

To identify the optimal hsCRP cut-off value, we randomly split the data into two sets: a training set (193 patients) and the validation set (194 patients). In the training set, we estimated the optimal cut-off value of hsCRP for predicting recurrence after curative treatment using maximally selected rank statistics, as described by Lausen and Schumacher.<sup>20</sup> This cut-off value is optimal for separation into high and low levels of prognosis. Statistically significant differences in prognosis using this cut-off value were examined using an adjustment for *P*-values, taking into account the arising multiple test situation. We then validated the cut-off value by the log-rank test in the validation set. We classified patients into high and low hsCRP groups according to the cut-off value and compared their baseline characteristics using Student's *t*-test or the Mann-Whitney *U*-test for continuous data and the  $\chi^2$ -test for categorical data.

Correlations between hsCRP and the following clinical parameters were estimated using Spearman's rank correlation coefficient: age, body height, body weight, BMI, waist circumference, SFA, VFA, platelet count, aspartate aminotransferase (AST) level, alanine aminotransferase (ALT) level, total bilirubin, albumin, tumor factors (including the size and number of nodules), and HCC-specific biomarkers  $\alpha$ -fetoprotein (AFP) and des- $\gamma$ -carboxyprothrombin.

Cumulative recurrence and survival curves were plotted using the Kaplan-Meier method, and differences were assessed using the log-rank test. We investigated predictors of recurrence and survival using the Cox proportional hazard regression model. Stepwise variable selection with the Akaike information criterion (AIC) was used to find the best model in the multivariate analysis. The Cox proportional hazards assumption was checked using the smoothed plots of Schoenfeld residuals. Moreover, the hazard ratio (HR) of hsCRP for survival was estimated as a continuous number using a restricted cubic spline with three knots after adjusting for other significant predictors.<sup>21</sup>

Regarding long-term survival, subgroup analyses using the Cox proportional hazards model were used to estimate the HR of higher versus lower hsCRP, with two-tailed *P*-values, for explanatory variables. The explanatory variables used for HR estimation were as follows: age, sex, serum albumin concentration, BMI, ALT level, platelet counts, clinical cirrhosis, AFP concentration and tumor stage.

Statistical analyses were performed using the "R" software (ver. 2.13.0; www.r-project.org), with the

“survival”, “maxstat” and “rms” packages. All tests were two-sided and  $P < 0.05$  was considered to indicate statistical significance.

## RESULTS

### Optimal cut-off value of hsCRP

THE DISTRIBUTION OF hsCRP values is shown in Figure 1(a). The median hsCRP was 0.05 mg/dL (IQR, 0.02–0.11). In the training set, maximally selected rank statistics showed that the optimal cut-off value of hsCRP to predict HCC recurrence was 0.08 mg/dL (Fig. 1b, Fig. S1A,B). This cut-off value was confirmed in the validation set (Fig. S1C,D). Thus, we classified patients with a hsCRP of 0.08 mg/dL or less as the low hsCRP group and the remaining patients as the high hsCRP group.

### Patients

Baseline characteristics of the patients are shown in Table 1. Of the 387 patients, 139 (35.9%) were classified into the high hsCRP group, and they were more likely to be heavier in weight, especially with central obesity, and to have worse liver function (Table 1). Diabetes was also more prevalent among patients in the high hsCRP group. Tumor size in the high hsCRP group tended to be larger.

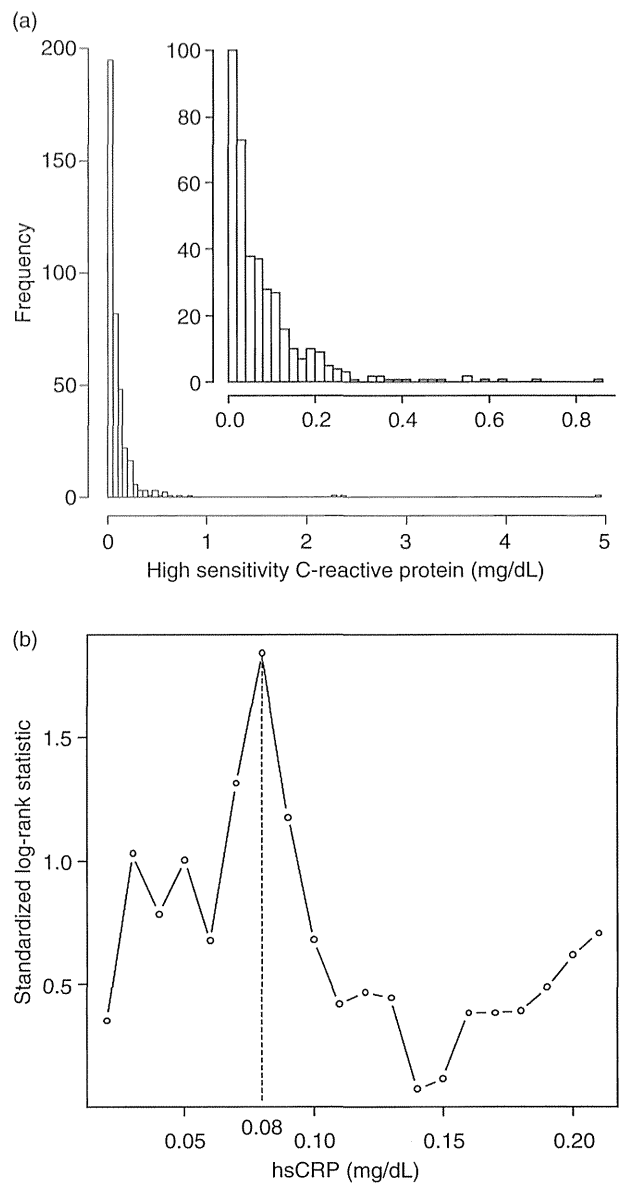
### Correlation between hsCRP and clinical parameters

Correlations between hsCRP and clinical parameters are shown in Table 2. Serum hsCRP was correlated not only with liver dysfunction and tumor size but also with high weight in HCC patients. Serum hsCRP had a positive correlation with VFA but not with SFA.

### Recurrence rate and long-term survival

The median observation period was 4.4 years (IQR, 3.1–6.0). In the low hsCRP group, the cumulative recurrence rates at 1, 3 and 5 years were 27.1%, 68.8% and 82.2%, respectively, while in the high hsCRP group, the rates were 39.1%, 79.3% and 90.0%, respectively (Fig. 2a). The recurrence rate curves for the two groups differed significantly (log-rank test,  $P = 0.0028$ ).

The overall survival rates at 1, 3 and 5 years were 98.4%, 87.9% and 71.8% in the low hsCRP group, respectively, and 94.2%, 73.1% and 50.9% in the high hsCRP group, respectively (Fig. 2b). The survival curves for the two groups differed significantly (log-rank test,  $P < 0.001$ ).



**Figure 1** Distribution and the optimal cut-off value of serum high-sensitivity C-reactive protein (hsCRP) concentrations. (a) The distribution of serum hsCRP concentrations is shown. The median serum hsCRP concentration was 0.05 mg/dL (interquartile range, 0.02–0.11). The upper right histogram shows the frequency of patients whose hsCRP levels ranged within 1.0 mg/dL. (b) Maximally selected log-rank statistics performed for hsCRP to determine an optimal cut-off value for separation of the two groups with different recurrence distribution. In the training set, the estimated cut-off point was 0.08 mg/dL with an M statistic of 1.85 and a corresponding corrected  $P$ -value of less than 0.001.

**Table 1** Baseline characteristics

Characteristics	Overall (n = 387)	Low hsCRP (n = 248)	High hsCRP (n = 139)	P value
Sex, male, n (%)	210 (54.3)	126 (50.8)	84 (60.4)	0.086
Age†	70.1 ± 8.0	70.7 ± 7.8	69.1 ± 8.4	0.062
Body weight (kg)	57.0 (50.1–64.0)	56.0 (49.1–62.8)	58.4 (52.0–66.3)	0.009
BMI (kg/m <sup>2</sup> )	22.8 (20.8–24.8)	22.4 (20.6–24.4)	23.6 (21.1–26.0)	0.005
Waist circumference (cm)	81.0 (75.5–88.1)	80.3 (75.1–86.8)	83.8 (77.0–89.0)	0.003
Visceral fat area (cm <sup>2</sup> )	65.7 (36.4–96.0)	61.1 (35.5–90.9)	73.8 (38.9–106.4)	0.028
Subcutaneous fat area (cm <sup>2</sup> )	111.1 (68.4–154.9)	110.1 (66.6–150.2)	117.6 (73.1–163.8)	0.31
Diabetes				0.014
Yes, n (%)	81 (20.9)	42 (16.9)	39 (28.1)	
Atherosclerosing disease				0.35
Yes, n (%)	189 (48.8)	126 (50.8)	63 (45.3)	
Alcohol consumption, n (%)				0.68
>80 g per day	41 (10.6)	28 (11.3)	13 (9.4)	
Clinical cirrhosis, n (%)	321 (82.9)	197 (79.4)	124 (89.2)	0.021
Fibrotic stage, n (%)‡				0.38
F0–2	58 (18.4)	42 (20.0)	16 (15.2)	
F3–4	257 (81.6)	168 (80.0)	89 (84.8)	
Inflammatory activity in non-cancerous tissue‡				0.38
0	0 (0)	0 (0)	0 (0)	
1	195 (61.9)	132 (62.9)	63 (60.0)	
2	117 (37.1)	75 (35.7)	42 (40.0)	
3	3 (1.0)	3 (1.4)	0 (0)	
hsCRP (mg/dL)	0.05 (0.02–0.11)	0.03 (0.02–0.05)	0.14 (0.11–0.22)	
Platelet count (×10 <sup>3</sup> /mm <sup>3</sup> )	101 (76–136)	101 (78–137)	100 (76–132)	0.82
AST (IU/L)	58 (45–78)	56 (42–76)	61 (50–80)	0.026
ALT (IU/L)	51 (34–77)	50 (33–77)	54 (36–75)	0.43
Total bilirubin (mg/dL)	0.8 (0.6–1.1)	0.8 (0.6–1.0)	0.9 (0.7–1.3)	0.004
Albumin (g/dL)	3.6 (3.3–3.9)	3.7 (3.4–4.0)	3.4 (3.1–3.8)	<0.001
Tumor size (cm)	2.0 (1.7–2.5)	2.0 (1.6–2.4)	2.1 (1.7–2.5)	0.06
No. of nodules				0.95
Solitary, n (%)	250 (64.6)	161 (64.9)	89 (64.0)	
2 or 3 nodules, n (%)	137 (35.4)	87 (35.1)	50 (36.0)	
Tumor stage, n (%)				0.11
Solitary nodule <2 cm	115 (29.7)	81 (32.7)	34 (24.5)	
Others	272 (70.3)	167 (67.3)	105 (75.5)	
Serum AFP (ng/mL)	21.6 (8.2–74.9)	24.5 (8.1–81.9)	17.1 (9.0–53.7)	0.33
Serum DCP (mAU/mL)§	21.0 (15.0–42.0)	21.0 (15.0–40.5)	21.0 (15.0–47.0)	0.49

†Expressed as mean ± standard deviation.

‡Biopsies were available in 315 patients (81.4%). Background liver was pathologically graded based on the METAVIR system.

§Serum DCP level could not be measured in two patients because they were taking warfarin.

AFP,  $\alpha$ -fetoprotein; ALT, alanine transaminase; AST, aspartate aminotransferase; BMI, body mass index; DCP, des- $\gamma$ -carboxyprothrombin; hsCRP, high-sensitivity C-reactive protein.

### Risk factors for survival and recurrence

Univariate analysis showed that a high hsCRP level was a significant predictor of overall survival (HR, 1.95; 95% confidence interval [CI], 1.43–2.66;  $P < 0.001$ ). The risk of recurrence after RFA increased in accordance with high hsCRP (HR, 1.43; 95% CI, 1.13–1.82;  $P = 0.003$ ). Other risk factors for recurrence and survival are shown in Table 3.

Stepwise multivariate analysis using the AIC established the best model for survival and recurrence (Table 3). Patients with higher hsCRP levels were at a significantly higher risk for both survival (HR, 1.59; 95% CI, 1.14–2.22;  $P = 0.007$ ) and recurrence (HR, 1.32; 95% CI, 1.03–1.67;  $P = 0.026$ ). After adjusting for other risk factors, we found that the estimated log-transformed HR of survival in relation to hsCRP peaked

**Table 2** Spearman's rank correlation coefficients and *P*-values between hsCRP and clinical parameters

Characteristics	Spearman's rho	<i>P</i>
Age	-0.055	0.28
Body weight (kg)	0.199	<0.001
BMI (kg/m <sup>2</sup> )	0.181	<0.001
Waist circumference (cm)	0.200	<0.001
Visceral fat area (cm <sup>2</sup> )	0.176	<0.001
Subcutaneous fat area (cm <sup>2</sup> )	0.064	0.21
Platelet count (×10 <sup>3</sup> /mm <sup>3</sup> )	0.044	0.39
AST (IU/L)	0.091	0.074
ALT (IU/L)	-0.002	0.96
Total bilirubin (mg/dL)	0.120	0.019
Albumin (g/dL)	-0.285	<0.001
Tumor size (cm)	0.124	0.015
No. of tumors	0.091	0.072
Serum AFP (ng/mL)	-0.086	0.091
Serum DCP (mAU/mL)†	0.050	0.32

†Serum DCP level could not be measured in two patients because they were taking warfarin.

AFP,  $\alpha$ -fetoprotein; ALT, alanine transaminase; AST, aspartate aminotransferase; BMI, body mass index; DCP, des- $\gamma$ -carboxyprothrombin; hsCRP, high-sensitivity C-reactive protein.

at approximately 0.16 mg/dL hsCRP and then plateaued (Fig. 3). This estimated HR was significant after adjustment ( $P = 0.0019$ ).

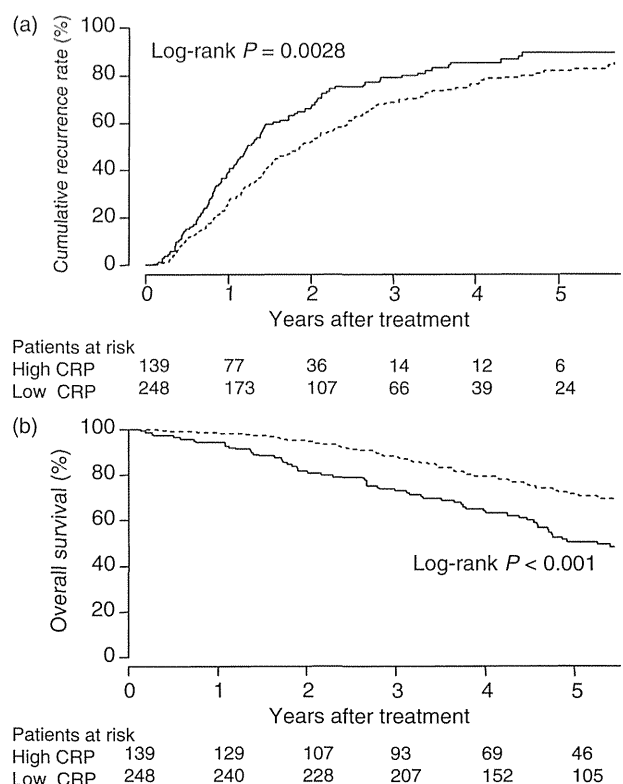
The effects of high hsCRP on the risk of HCC survival were also evaluated in subgroup analyses to assess whether high hsCRP was a significant risk factor over strata (Fig. 4). Indeed, a higher hsCRP level was found to be a significant risk factor for survival excluding the subgroups of patients with non-liver cirrhosis and serum albumin levels of more than 4.0 g/dL. A higher hsCRP level was an especially strong predictor for survival in patients with HCC with a single nodule of less than 2 cm (HR, 2.88; 95% CI, 1.46–5.66;  $P < 0.001$ ).

## DISCUSSION

**I**N THIS STUDY, we identified an optimal hsCRP cut-off value of 0.08 mg/dL to predict recurrence after curative treatment for early stage HCC. This value was much lower than the conventional cut-off value of 0.3 mg/dL and was below the range detected by conventional CRP assays. Using the hsCRP assay enabled us to predict early recurrence and poor prognosis after curative treatment for early/very early stage HCC.

What pathological mechanism(s) may underlie this association between hsCRP and prognosis? We specu-

lated that visceral fat was the dominant factor associated with elevations in hsCRP level compared with inflammation of non-tumorous liver tissue, considering that neither AST, ALT nor the grade of liver inflammation were associated with higher hsCRP levels. As shown in Table 2, there were weak correlations between hsCRP and anthropometric parameters related to central obesity, including waist circumference and VFA. Accordingly, visceral adipose accumulation may link higher hsCRP with a poor prognosis, because obesity-induced dysregulation of adipokines can directly accelerate HCC



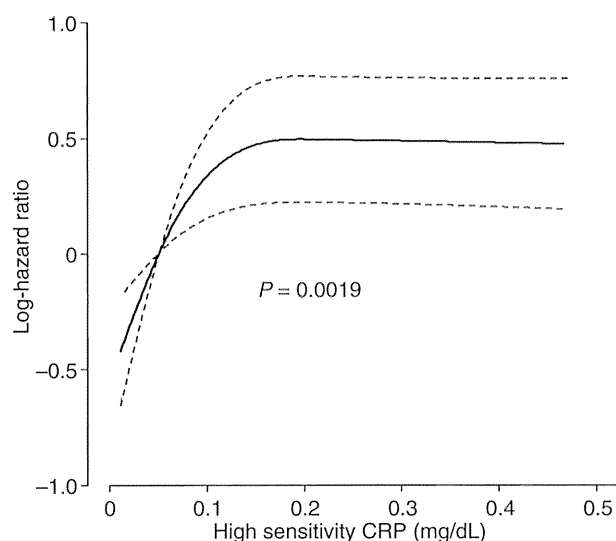
**Figure 2** Cumulative recurrence rate and overall survival stratified by high-sensitivity C-reactive protein (hsCRP) level. (a) The cumulative recurrence rate is shown. In the low hsCRP group, the cumulative recurrence rates at 1, 3 and 5 years were 27.1%, 68.8% and 82.2%, respectively, while in the high hsCRP group, the rates were 39.1%, 79.3% and 90.0%, respectively. There was a significant difference between the two cumulative rates (log-rank test,  $P = 0.0028$ ). (b) The overall survival is shown. In the low hsCRP group, the overall survival rates at 1, 3 and 5 years were 98.4%, 87.9% and 71.8%, respectively, while in the high hsCRP group, the rates were 94.2%, 73.1% and 50.9%, respectively. The survival curves of the two groups were significantly different (log-rank test,  $P < 0.001$ ). —, High hsCRP; ---, low hsCRP.

Table 3 Risk factors for recurrence and survival

Variables	Survival						Recurrence					
	Univariate			Multivariate			Univariate			Multivariate		
	HR	95% CI	P	HR	95% CI	P	HR	95% CI	P	HR	95% CI	P
High hsCRP	1.95	1.43–2.66	<0.001	1.59	1.14–2.22	0.007	1.43	1.13–1.82	0.003	1.32	1.03–1.67	0.026
Sex, female	1.05	0.77–1.43	0.78				0.86	0.68–1.08	0.20			
Age per 1 year	1.03	1.01–1.05	0.005	1.04	1.02–1.06	<0.001	1.01	0.99–1.02	0.49			
BMI per 1 kg/m <sup>2</sup>	0.97	0.92–1.01	0.14				0.98	0.95–1.02	0.28			
VFA per 10 cm <sup>2</sup>	0.98	0.94–1.01	0.22				1.00	0.97–1.02	0.71			
Presence of diabetes	1.07	0.73–1.57	0.72				1.03	0.78–1.37	0.83			
Presence of atherosclerosis	1.03	0.76–1.40	0.86				1.09	0.87–1.37	0.44			
Alcohol consumption >80 g/day	1.08	0.65–1.78	0.78				0.98	0.66–1.45	0.92			
Serum albumin, per 1 g/dL	0.33	0.23–0.48	<0.001	0.47	0.32–0.70	<0.001	0.61	0.48–0.78	<0.001	0.61	0.47–0.78	<0.001
Serum total bilirubin, per 1 mg/dL	2.28	1.64–3.17	<0.001	1.76	1.22–2.56	0.003	1.30	0.99–1.72	0.063			
Platelet count, per 10 × 10 <sup>3</sup> /mm <sup>3</sup>	0.96	0.93–1.00	0.033				0.98	0.96–1.00	0.11			
Prothrombin time, per 1%	0.99	0.98–1.01	0.26				1.00	0.99–1.01	0.79			
Tumor size per 1 cm	1.61	1.19–2.20	0.002	1.56	1.14–2.14	0.006	1.45	1.16–1.82	0.001	1.44	1.15–1.81	0.002
No. of nodules per 1 nodule	1.12	0.91–1.38	0.29				1.31	1.11–1.53	0.001	1.34	1.14–1.58	<0.001
Serum AFP ≥100 ng/dL	1.41	0.98–2.01	0.061	1.50	1.04–2.19	0.033	1.32	0.99–1.75	0.055	1.39	1.04–1.86	0.024
Serum DCP ≥100 mAU/mL†	1.61	1.00–2.61	0.051				1.44	0.97–2.13	0.067	1.59	1.07–2.37	0.021

†Serum DCP level could not be measured in two patients because they were taking warfarin.

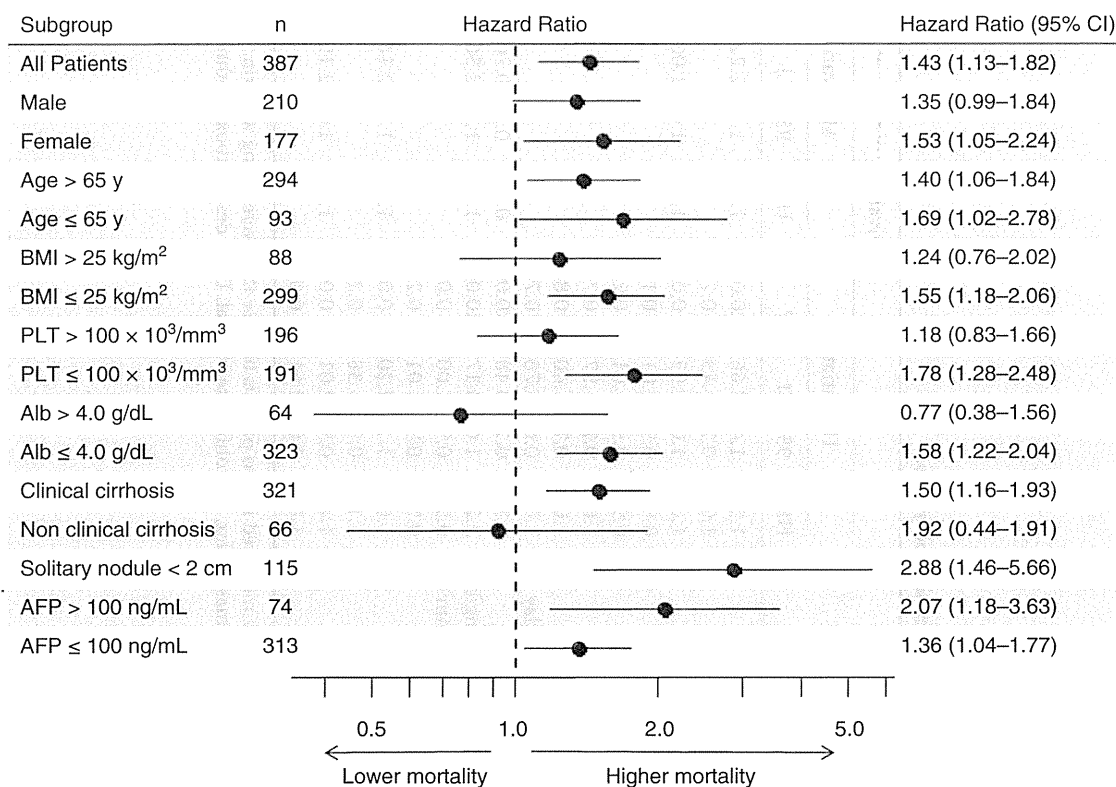
AFP,  $\alpha$ -fetoprotein; BMI, body mass index; CI, confidence interval; DCP, des- $\gamma$ -carboxyprothrombin; HR, hazard ratio; hsCRP, high-sensitivity C-reactive protein; VFA, visceral fat area.



**Figure 3** Log-hazard ratio of survival related to serum high-sensitivity C-reactive protein (hsCRP) concentration. Serum hsCRP concentration was fixed at 0.05 mg/dL. The estimated log-transformed hazard ratio of survival in relation to hsCRP peaked at approximately 0.16 mg/dL and then plateaued.

development by enhancing cancer cell proliferation, inhibiting apoptosis and inducing migration which are liver steatosis- and inflammation-independent mechanisms.<sup>22,23</sup> However, this present study could not address the causality. Further study is required to elucidate the association between hsCRP, adipokines and the outcomes of HCC after curative treatment.

Based on previous reports,<sup>24-26</sup> interleukin (IL)-6 could partially explain the relationship of elevated hsCRP with higher recurrence rate and poorer survival in HCC patients. IL-6, the principal regulator of CRP production, is a multifunctional cytokine largely responsible for the hepatic response to infections or systemic inflammation. In fact, we previously reported that a higher serum IL-6 level correlated with future HCC development in patients with chronic hepatitis C.<sup>27</sup> IL-6 is also a key molecule linking obesity with hepatocarcinogenesis.<sup>28</sup> Moreover, promising molecular innovations support these results. Hoshida *et al.* reported a transcriptomic signature in liver tissue



**Figure 4** Subgroup analyses stratified by risk factor. A higher high-sensitivity C-reactive protein (hsCRP) concentration was a significant risk factor for mortality over nearly all strata. A higher hsCRP level was also an independent risk factor for survival in patients with a single nodule of less than 2 cm. AFP,  $\alpha$ -fetoprotein; Alb, albumin; BMI, body mass index; CI, confidence interval; PLT, platelets.



adjacent to HCC that could predict survival and late recurrence after HCC resection.<sup>29</sup> In this study, the poor prognosis signature involved genes associated with inflammation, including those related to interferon signaling and activation of nuclear factor- $\kappa$ B. Intriguingly, the downstream targets of IL-6 were strongly associated with the poor-prognosis signature in non-tumorous liver tissue. Thus, higher CRP may independently indicate carcinogenic potential in the underlying liver. Additionally, the transcription factor signal transducer and activator of transcription-3, which mediates the effects of IL-6, was found to be activated in most HCC and seems to be associated with more aggressive tumors. This may indicate that higher hsCRP levels reflect the HCC malignancy grade.<sup>30</sup>

Exploration of serum biomarkers to predict survival, progression and treatment efficacy among HCC patients in various stages is an ever-improving field.<sup>5,31,32</sup> Several reports have shown associations of conventional CRP with recurrence rate and long-term survival in HCC patients who were treated by surgical resection,<sup>33</sup> non-surgical procedures<sup>34</sup> and liver transplantation.<sup>35</sup> In these studies, 1.0 mg/dL was used as the cut-off value. However, only three patients with early HCC (0.8%) showed a hsCRP of more than 1.0 mg/dL in this study (Fig. 1a). Thus, there is a need for an optimal cut-off value in early/very early stage HCC patients.<sup>36,37</sup> On the basis of a statistically robust method, this study set a useful hsCRP cut-off value to identify patients at risk for rapid progression to death despite curative treatment among patients with early HCC, who have intrinsically better prognosis.<sup>38</sup> Additionally, measuring hsCRP is practical in real-world clinical settings because it is readily determinable, simple, widely available and inexpensive.

This study has several limitations. First, due to its retrospective nature, hsCRP was not measured in some patients, which might have caused selection bias. However, the proportion of these patients (5.9%) was small. Second, elevation of hsCRP may reflect non-specific or undetectable infection, especially that related to cirrhosis. Thus, hsCRP may be a surrogate marker for poor liver function. However, hsCRP remained significant in terms of recurrence and prognosis prediction after adjusting for indicators of liver functional reserve. Third, we did not validate the determined optimal cut-off value of hsCRP in an external cohort. However, we could validate the cut-off value using a split-sample method.

Although we analyzed only patients with C-HCC in order to restrict the background conditions of patients,

whether hsCRP is an independent predictor of recurrence and survival in HCC patients with hepatitis B virus or non-alcoholic steatohepatitis is an interesting issue. Further study is required to clarify the predictability of slightly elevated hsCRP in patients with HCC of other etiologies.

In conclusion, slightly elevated hsCRP, even though below the range detected by conventional CRP assays, can identify patients with early stage C-HCC at risk for recurrence and death.

## REFERENCES

- Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. *CA Cancer J Clin* 2011; **61**: 69–90.
- Llovet JM, Bruix J. Novel advancements in the management of hepatocellular carcinoma in 2008. *J Hepatol* 2008; **48** (Suppl 1): S20–37.
- Sherman M. Recurrence of hepatocellular carcinoma. *N Engl J Med* 2008; **359**: 2045–7.
- Tateishi R, Shiina S, Yoshida H *et al*. Prediction of recurrence of hepatocellular carcinoma after curative ablation using three tumor markers. *Hepatology* 2006; **44**: 1518–27.
- Nault JC, Guyot E, Laguillier C *et al*. Serum proteoglycans as prognostic biomarkers of hepatocellular carcinoma in patients with alcoholic cirrhosis. *Cancer Epidemiol Biomarkers Prev* 2013; **22**: 1343–52.
- Danesh J, Wheeler JG, Hirschfield GM *et al*. C-reactive protein and other circulating markers of inflammation in the prediction of coronary heart disease. *N Engl J Med* 2004; **350**: 1387–97.
- Siemes C, Visser LE, Coebergh JW *et al*. C-reactive protein levels, variation in the C-reactive protein gene, and cancer risk: the Rotterdam Study. *J Clin Oncol* 2006; **24**: 5216–22.
- Ridker PM. High-sensitivity C-reactive protein: potential adjunct for global risk assessment in the primary prevention of cardiovascular disease. *Circulation* 2001; **103**: 1813–8.
- Villasenor A, Flatt SW, Marinac C, Natarajan L, Pierce JP, Patterson RE. Postdiagnosis C-reactive protein and Breast Cancer Survivorship: findings from the WHEL Study. *Cancer Epidemiol Biomarkers Prev* 2014; **23**: 189–99.
- Proctor MJ, Horgan PG, Talwar D, Fletcher CD, Morrison DS, McMillan DC. Optimization of the systemic inflammation-based Glasgow prognostic score: a Glasgow Inflammation Outcome Study. *Cancer* 2013; **119**: 2325–32.
- Bruix J, Sherman M; American Association for the Study of Liver Diseases. Management of hepatocellular carcinoma: an update. *Hepatology* 2011; **53**: 1020–2.
- Jonsson JR, Barrie HD, O'Rourke P, Clouston AD, Powell EE. Obesity and steatosis influence serum and hepatic inflammatory markers in chronic hepatitis C. *Hepatology* 2008; **48**: 80–7.

- 13 Zhao B, Colville J, Kalaigian J *et al.* Automated quantification of body fat distribution on volumetric computed tomography. *J Comput Assist Tomogr* 2006; 30: 777–83.
- 14 Torzilli G, Minagawa M, Takayama T *et al.* Accurate preoperative evaluation of liver mass lesions without fine-needle biopsy. *Hepatology* 1999; 30: 889–93.
- 15 Teratani T, Yoshida H, Shiina S *et al.* A novel display of reconstruction computed tomography for the detection of small hepatocellular carcinoma. *Liver Int* 2004; 24: 619–24.
- 16 Fujishima T, Yoshida H, Obi S *et al.* Analysis of factors influencing hepatocellular carcinoma detection: efficient use of computed tomography during arterial portography and during hepatic arteriography. *J Gastroenterol* 2005; 40: 266–73.
- 17 Edmondson HA, Steiner PE. Primary carcinoma of the liver: a study of 100 cases among 48,900 necropsies. *Cancer* 1954; 7: 462–503.
- 18 Bedossa P, Poynard T. An algorithm for the grading of activity in chronic hepatitis C. the METAVIR Cooperative Study Group. *Hepatology* 1996; 24: 289–93.
- 19 Omata M, Tateishi R, Yoshida H, Shiina S. Treatment of hepatocellular carcinoma by percutaneous tumor ablation methods: ethanol injection therapy and radiofrequency ablation. *Gastroenterology* 2004; 127: S159–166.
- 20 Lausen B, Schumacher M. Maximally selected rank statistics. *Biometrics* 1992; 48: 73–85.
- 21 Durrleman S, Simon R. Flexible regression models with cubic splines. *Stat Med* 1989; 8: 551–61.
- 22 Sharma D, Wang J, Fu PP *et al.* Adiponectin antagonizes the oncogenic actions of leptin in hepatocellular carcinogenesis. *Hepatology* 2010; 52: 1713–22.
- 23 Saxena NK, Fu PP, Nagalingam A *et al.* Adiponectin modulates C-jun N-terminal kinase and mammalian target of rapamycin and inhibits hepatocellular carcinoma. *Gastroenterology* 2010; 139: 1762–73, 1773 e1761–5.
- 24 Jang JW, Oh BS, Kwon JH *et al.* Serum interleukin-6 and C-reactive protein as a prognostic indicator in hepatocellular carcinoma. *Cytokine* 2012; 60: 686–93.
- 25 Rhodes B, Furnrohr BG, Vyse TJ. C-reactive protein in rheumatology: biology and genetics. *Nat Rev Rheumatol* 2011; 7: 282–9.
- 26 Ganter U, Arcone R, Toniatti C, Morrone G, Ciliberto G. Dual control of C-reactive protein gene expression by interleukin-1 and interleukin-6. *EMBO J* 1989; 8: 3773–9.
- 27 Nakagawa H, Maeda S, Yoshida H *et al.* Serum IL-6 levels and the risk for hepatocarcinogenesis in chronic hepatitis C patients: an analysis based on gender differences. *Int J Cancer* 2009; 125: 2264–9.
- 28 Park EJ, Lee JH, Yu GY *et al.* Dietary and genetic obesity promote liver inflammation and tumorigenesis by enhancing IL-6 and TNF expression. *Cell* 2010; 140: 197–208.
- 29 Hoshida Y, Villanueva A, Kobayashi M *et al.* Gene expression in fixed tissues and outcome in hepatocellular carcinoma. *N Engl J Med* 2008; 359: 1995–2004.
- 30 Calvisi DF, Ladu S, Gorden A *et al.* Ubiquitous activation of Ras and Jak/Stat pathways in human HCC. *Gastroenterology* 2006; 130: 1117–28.
- 31 Tsuchiya K, Asahina Y, Matsuda S *et al.* Changes in plasma vascular endothelial growth factor at 8 weeks after sorafenib administration as predictors of survival for advanced hepatocellular carcinoma. *Cancer* 2014; 120: 229–37.
- 32 Raoul JL, Bruix J, Greten TF *et al.* Relationship between baseline hepatic status and outcome, and effect of sorafenib on liver function: SHARP trial subanalyses. *J Hepatol* 2012; 56: 1080–8.
- 33 Hashimoto K, Ikeda Y, Korenaga D *et al.* The impact of preoperative serum C-reactive protein on the prognosis of patients with hepatocellular carcinoma. *Cancer* 2005; 103: 1856–64.
- 34 Sieghart W, Pinter M, Huckle F *et al.* Single determination of C-reactive protein at the time of diagnosis predicts long-term outcome of patients with hepatocellular carcinoma. *Hepatology* 2013; 57: 2224–34.
- 35 An HJ, Jang JW, Bae SH *et al.* Serum C-reactive protein is a useful biomarker for predicting outcomes after liver transplantation in patients with hepatocellular carcinoma. *Liver Transpl* 2012; 18: 1406–14.
- 36 Dufour JF. C-reactive protein, a prognostic marker in hepatocellular carcinoma. *Hepatology* 2013; 57: 2103–5.
- 37 Pang RW, Poon RT. Diagnosis: novel prognostic biomarkers in hepatocellular carcinoma. *Nat Rev Gastroenterol Hepatol* 2012; 9: 691–2.
- 38 Livraghi T, Meloni F, Di Stasi M *et al.* Sustained complete response and complications rates after radiofrequency ablation of very early hepatocellular carcinoma in cirrhosis: is resection still the treatment of choice? *Hepatology* 2008; 47: 82–9.

## SUPPORTING INFORMATION

ADDITIONAL SUPPORTING INFORMATION may be found in the online version of this article at the publisher's website:

**Figure S1** Cumulative recurrence rate and survival stratified by high-sensitivity C-reactive protein (hsCRP) in the training set and the validation set, respectively. (A) Cumulative recurrence rate in the training set. In the low hsCRP group, the cumulative recurrence rates at 1, 3 and 5 years were 27.7%, 68.0% and 83.0%, respectively; while in the high hsCRP group, the rates were 38.3%, 81.4% and 88.0%, respectively. There was a significant difference between the two cumulative rates (log-rank test,  $P = 0.049$ ). (B) Overall survival in the training set. In the low hsCRP group, the overall survival rates at 1, 3 and 5 years were 97.5%, 84.6% and 68.2%, respectively.

