ITT Insulin tolerance test

QUICKI Quantitative insulin sensitivity check index MCAD Medium-chain acyl-CoA dehydrogenase TIMP Tissue inhibitor of metalloproteinases

TGF Transforming growth factor

MTP Microsomal triglyceride transfer protein

AMPK AMP-activated protein kinase

Introduction

Obesity-induced liver disease (nonalcoholic fatty liver disease, NAFLD), is now the commonest cause of chronic liver disease in affluent nations. The disease comprises obesity and insulin resistance (IR), with a consequent histopathological spectrum of hepatosteatosis, steatohepatitis (nonalcoholic steatohepatitis, NASH), cirrhosis and possible hepatocellular cancer [1-3]. There are presently no proven treatments for NAFLD or its more severe stage NASH. Bofutsushosan (BTS), a Japanese herbal (Kampo) medicine, long used as an anti-obesity medicine in Japan and other Asian countries [4] has recently been shown in obese Japanese women, to reduce body weight and improve IR [4]. In addition, BTS in experimental animals prevented adipogenesis [5], reduced weight, suppressed visceral and subcutaneous fat accumulation and in parallel decreased plasma glucose, triglycerides (TG), insulin, tumor necrosis factor-alpha [6], and hepatic steatosis induced by high-fat diet feeding [7, 8]. The mechanism of action of BTS however, is not known. Our aim here was to evaluate the ability of BTS to prevent the development of NASH in a recently described murine model involving administration of gold-thioglucose (GTG) and high-fat feeding to induce NASH [9], and to determine the mediators and pathways involved.

Materials and methods

Animal preparation

All procedures conformed to our institutions' guidelines for the care and use of animals in Kochi Medical School. Fourweek-old male C57BL/6 mice were purchased from CLEA Japan Inc. All animals were housed for 12 weeks on a 12 h light/12 h dark cycle, with food and water freely available. Mice were fed high-fat-diet (HF, 640 kcal/100 g, F2HFD2, Oriental Yeast, Tokyo, Japan) or HF admixed with 2 or 5 % BTS (TJ-62, Tsumura & Co., Tokyo, Japan). All groups were fed standard chow (SC) for the first week and

then continued on their respective group diets for remainder of the protocol.

Three experimental groups were studied: (1) intra-peritoneal administered GTG (2 mg/g of body weight, Sigma-Aldrich, St. Louis, MO, USA) and then SC for 1 week followed by HF diet for 11 weeks (GTG + HF) [9]; (2 and 3) intra-peritoneal GTG, SC for 1 week followed by HF diet admixed with either 2 %BTS or 5 % BTS for further 11 weeks (2 %BTS or 5 % BTS). At the end of the treatment period, all animals were fasted overnight, anesthetized with pentobarbital sodium intraperitoneally (25-50 mg/kg of body weight, Nembutal; Abbott Laboratories, Abbott Park, IL, USA). Blood and liver samples were harvested. Livers were fixed in 10 % formalin, or snap frozen in liquid nitrogen and stored at -80 °C, for later analyses.

CT scan analysis for body fat composition

The extent of adiposity in each experimental group was assayed by CT scanning (La Theta, ALOKA, Tokyo, Japan) under isoflurane (2 % v/v) anesthesia as described previously [9]. Animal were scanned at 2-mm intervals from the diaphragm to the pelvis, and visceral fat and subcutaneous fat volumes quantified with La Theta software (version 1.00) [9–11].

Histopathological examination

Five-micrometer sections of formalin-fixed/paraffinembedded livers were processed for haematoxylin and eosin (H&E). Oil Red-O staining of intracellular neutral lipids was performed according to the manufacturer's instructions (Sigma-Aldrich, St. Louis, MO, USA). For estimation of extent of hepatic steatosis, the areas of digital photomicrographs were quantified with a computerized image analysis system (macintosh MacSCOPE version 2.591) as described previously [9, 12]. Degree of oxidative stress was determined by staining and quantification with anti-8-hydroxy-2'-deoxyguanosine (8-OHdG) and anti-4hydroxy-2-nonenal (4-HNE) as previously described [9, 12, 13].

Glucose tolerance test (GTT), insulin tolerance test (ITT) and QUICKI

At 12 weeks, a glucose tolerance test (GTT) (n = 6) and an insulin tolerance test (ITT) (n = 6) were performed. For GTT, mice were fasted for 18 h, and then intra-peritoneally loaded with 20 % glucose at a dose of 1.0 g/kg body weight. For ITT, mice were fasted for 6 h, and then intra-peritoneally challenged with human insulin at 1.0 U/kg body weight [14, 15]. With both GTT and ITT blood



samples from the orbital sinus were taken at times 0, 30, 45, 60 and 120 min and plasma glucose concentrations measured using an automatic blood glucose meter (Glutest; Sanwa Kagaku Kenkyusho Co., Ltd., Nagoya, Japan). Plasma insulin level was measured by Ultrasensitive Mouse Insulin ELISA kit (Mercodia AB, Uppsala, Sweden) according to the manufacture's protocol. The quantitative insulin sensitivity check index (QUICKI), as a measure of IR, was calculated from the fasting insulin and glucose levels.

Measurement of plasma adiponectin

Plasma adiponectin levels were measured by Mouse Adiponectin/Acrp30 (R&D Systems, Minneapolis, MN, USA) according to the manufacture's instructions.

Laboratory evaluation

Asparate aminotransferase (AST), alanine aminotransferase (ALT) and TG were measured by an autoanalyzer (BM6010; JEOL Ltd., Tokyo, Japan).

Real-time RT-PCR for quantitative assessment of mRNA expression

Total RNA was extracted using trizol reagent (Life Technologies, Grand Island, NY, USA) according to the manufacture's protocol. RNA extracts were reverse-transcribed with random hexamers and avian myeloblastosis virus reverse transcriptase using a commercial kit (Takara, Kyoto, Japan). Real time RT-PCR were performed for quantitative assessment of mRNA expression on an ABI Prism 7000 Sequence Detection system (Applied Biosystems, Foster City, CA, USA) according to the manufacturer's protocol. Probes and primers for TNF-α, PPAR-α, PPAR-γ, MTP, DGAT2, Cyp2E1, adiponectin receptor 1/receptor 2 (AdipoR1/R2) were all purchased from Applied Biosystems. Relative expression of target gene mRNA was normalized to the amount of GAPDH mRNA.

Western blot analysis

For in vivo analysis of phosphorylated Akt and total Akt, mice were fasted for 18 h, injected intra-peritoneally with human insulin (10 U/kg) or control, and sacrificed 4 min later. Livers were snap frozen in liquid nitrogen. Liver total protein was analyzed by western blot with a polyclonal antibody to phosphorylated Akt/Akt and phosphorylated AMPK/AMPK (Cell Signaling Technology, Inc., Danvers, MA, USA) as described [16]. For analysis of SREBP-1c, mice were fasted for 18 h, sacrificed, and livers dissected

and homogenized to prepare cell nuclear extracts which were then analyzed by western blotting with anti-SREBP-1c antibody (Santa Cruz Biotechnology, Inc., California, CA, USA) as described [17]. For analysis of 4-HNE, liver total protein was analyzed by western blot with a monoclonal anti-4-HNE antibody (Japan Institute for the Control of Aging, Shizuoka, Japan).

Statistics

Data are shown as mean \pm SD. A univariate analysis was conducted with the Mann–Whitney U test to determine significance between groups. Qualitative data were compared using Fisher's exact test. Statistical significance was accepted at p < 0.05. All analyses were performed using Stat View software (SAS Institute, Cary, NC, USA).

Results

BTS treatment reduces GTG + HF induced obesity and steatohepatitis

Mice administered GTG and then fed a HF diet (GTG + HF), had a comprehensive histological and dysmetabolic phenotype resembling human NASH as reported recently [9]. To then evaluate the effectiveness of BTS in the GTG + HF model, we studied the anthropometric, radiological, biochemical and histological parameters as detailed above in the presence or absence of BTS 2 or 5 % admixed with the HF component of the GTG + HF. Administration of BTS attenuated weight gain in a dose dependent manner (Fig. 1a). Unexpectedly, GTG + HF induced increase in the volume of the visceral and subcutaneous fat were not markedly attenuated by treatment with BTS (Fig. 1b). However, hepatic steatosis on H&E or Oil Red-O staining in 12 weeks was attenuated in a dose dependent manner by BTS treatment (Figs. 2a, 3a). Additionally, BTS treatment also attenuated GTG + HF induced hepatomegaly in a dose-dependent manner (Fig. 2b).

GTG + HF mice livers showed steatohepatitis with marked steatosis and inflammation, hepatocyte ballooning and Mallory–Denk bodies as described previously (Fig. 3a) [9]. BTS treatment attenuated hepatic steatosis and hepatic inflammation (Fig. 3a), and inhibited hepatocyte ballooning and Mallory–Denk bodies. In parallel, oxidative stress makers, 8-OHdG (Fig. 3b) and 4-HNE (supplemental figure), were remarkably reduced by BTS, as was the expression of TNF- α (Fig. 3b). BTS treatment moreover, attenuated GTG + HF induced elevation of transaminases (Table 1).



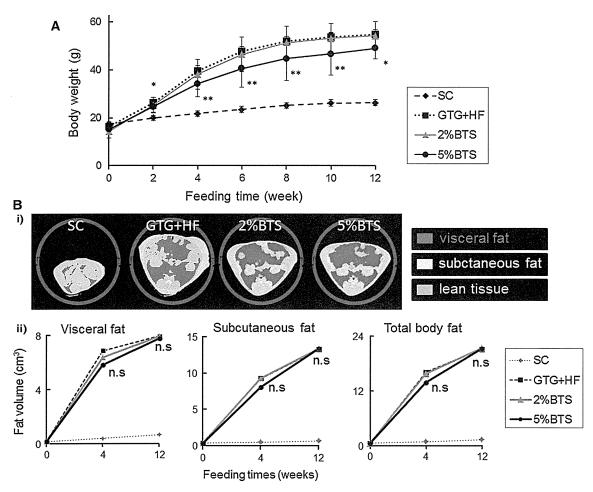


Fig. 1 Effectiveness of BTS for obesity and adiposity. **a** BTS dose-dependently attenuated weight gain in GTG + HF mice. **p < 0.01, *p < 0.05 vs GTG + HF, n = 6. SC standard chow fed mice, GTG + HF GTG treated and HF fed mice, 2 %BTS 2 %BTS + GTG + HF fed mice, 5 %BTS 5 %BTS + GTG + HF fed mice. **b** Anthropometry and evaluation of subcutaneous fat and visceral fat volumes by abdominal CT i representative

(Fig. 4a). In addition, expression of PPAR-γ, an activator

2 %BTS, 5 %BTS vs GTG + HF, p = ns, n = 6

photomicrographs of abdominal CT scan of SC, GTG + HF,

2 %BTS and 5 %BTS mice shown at 12 weeks. Yellow subcutaneous

fat, purple visceral fat, blue lean tissue. ii Time course of increase of

volume of subcutaneous and visceral fat. Neither subcutaneous fat nor visceral fat was significantly attenuated by treatment with BTS;

BTS inhibition of hepatic lipid metabolism occurs through induction of adiponectin signaling

We examined plasma adiponectin levels and liver expression of the adiponectin receptors (AdipoR1 and AdipoR2), an important anti-inflammatory cytokine and receptors [18–20], because both plasma adiponectin levels and liver expression of the adiponectin receptors were decreased in GTG + HF mice as described previously [9]. Plasma adiponectin level and hepatic expression of AdipoR1, R2 were significantly enhanced by BTS (Fig. 4a). We next investigated the pathways of suppression of hepatic lipid metabolism by adiponectin in the presence of BTS. The expression of SREBP-1c was decreased dose dependently by BTS (Fig. 4b). The phosphorylation of AMPK (P-AMPK/AMPK) was here increased by BTS treatment (Fig. 4c) in parallel with activation of AdipoR1 signaling

of AMPK, was increased by BTS treatment (Fig. 4d).

Attenuation of hepatic steatosis with BTS treatment involves activation of fatty acid oxidation

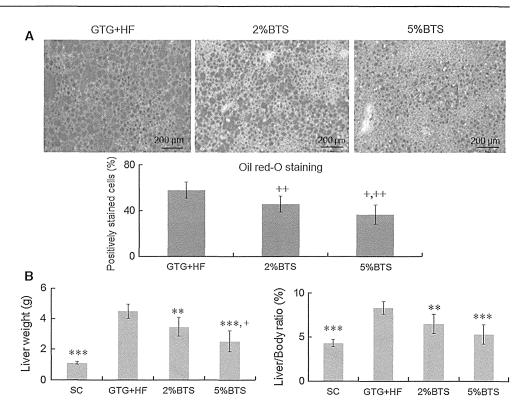
Additionally, the expression of PPAR- α and its target genes, MCAD, involved in mitochondrial β -oxidation and CYP2E-1 involved in microsomal ω -oxidation [21, 22] were increased by treatment with BTS (Fig. 4e–g).

BTS promotes hepatic lipid export

We next determined if enhanced secretion of TG from the liver could contribute to the attenuation of hepatic steatosis by BTS treatment. The expression of microsomal triglyceride transfer protein (MTP) known to play a central role in



Fig. 2 BTS reduces hepatomegaly and hepatic steatosis. a Oil red-O staining and image analysis of livers: Oil red-O staining showed that BTS attenuated hepatic steatosis in GTG + HF fed mice. In addition, image analysis for Oil red-O staining of liver sections confirmed that BTS treatment attenuated hepatic steatosis. Plus symbol p < 0.0001 vs 2 %BTS, double plus symbol p < 0.00001 vs GTG + HF, n = 6. **b** Liver weight and liver/ body weight ratio (liver/body): the increase of both liver weight and liver/body ratio was significantly attenuated by BTS treatment in a dose dependent manner. Triple asterisk p < 0.001, double asterisk p < 0.01 vs GTG + HF, plus symbol p<0.05 vs 2 %BTS,



lipoprotein assembly [23] was increased with BTS treatment (Fig. 4h). Interestingly, expression of DGAT2 reported to be involved in the conversion of free fatty acids into TG in the liver [24] was also increased in the BTS treated groups (Fig. 4i).

BTS reduces glucose intolerance and insulin resistance through induction of Akt

Since IR is regarded as a central pathogenic feature of NAFLD [25], we now investigated the effect of BTS treatment on IR. Fasting plasma glucose and insulin levels were markedly reduced in a dose dependent manner by BTS treatment, and QUICKI as an index of IR was also increased by BTS (Fig. 5a). We next evaluated the attenuation of glucose intolerance and IR using the GTT and the ITT in the presence of BTS. GTT revealed that treatment with BTS remarkably attenuated severe glucose intolerance induced by GTG + HF, and ITT showed that treatment with BTS attenuated IR induced by GTG + HF (Fig. 5b). The reduction of IR by BTS involved its promotion of the phosphorylation of Akt (Fig. 5c) an important factor in glucose metabolism [26].

Discussion

The public health importance of NAFLD [1, 2] and the unavailability of proven and effective therapies drive the search for a greater understanding of its pathophysiology

and novel therapeutic pathways. In this study, we have clarified the mechanisms through which BTS attenuates NASH based on a novel animal model of NASH [9]. BTS attenuated the GTG + HF induced increases in body and liver weight, serum transaminases, hepatic steatosis, degree of oxidative stress and TNF-α expression (Figs. 1, 2, 3; Table 1) without reducing intake volume of diets (data not shown). However, in contrast to previous reports [6], we did not demonstrate statistically remarkable reduction by BTS, in GTG + HF mice, of induced increases in visceral or subcutaneous fat (Fig. 1b), perhaps because the volume of these fats in our mice were much larger than previously reported [6]. Additionally, the reduction of body weight might mainly be through the reduction of fat accumulation in muscles, since it has been reported that the degree of hepatosteatosis is well correlated with the degree of fat accumulation in muscles [27].

To examine the mechanisms though which BTS attenuated hepatic steatosis, we firstly evaluated the effect of BTS on adiponectin, thought to be a central adipokine in the pathogenesis for NASH [18, 19]. The expression of AdipoR1 and AdipoR2 in the livers was increased in a dose dependent manner by BTS (Fig. 4a), indicating that BTS could have PPAR- α agonist like effect, since PPAR- α agonists are known to increase expression of AdipoR1 and AdipoR2 [28]. Interestingly, plasma adiponectin levels were also remarkably induced by BTS treatment (Fig. 4a) even though neither visceral nor subcutaneous fat were decreased. These data indicated that BTS through putative



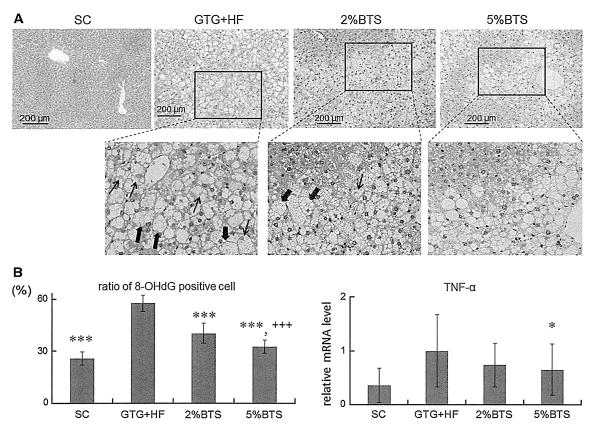


Fig. 3 BTS treatment reduces hepatic inflammation and oxidative stress. a The extent of hepatic inflammation was markedly attenuated with BTS: hepatocyte ballooning and Mallory–Denk bodies in the livers of GTG + HF fed mice were absent in BTS treatment livers. Filled arrow ballooning hepatocyte, arrow Mallory–Denk body. b Oxidative stress and TNF- α expression: BTS reduced hepatic

oxidative stress as shown by reduction of numbers of nuclei stained positive for 8-OHdG in GTG + HF mice. BTS similarly reduced hepatic TNF- α mRNA expression. *Triple asterisk* p < 0.001, asterisk p < 0.01 vs GTG + HF, *Triple plus symbol* p < 0.001 vs 2 %BTS, n = 6

Table 1 Physiological and biochemical analyses in mice treated with BTS

| | SC $(n = 6)$ | GTG + HF (n = 6) | 2 %BTS (n = 6) | 5 %BTS $(n = 6)$ |
|------------|--------------|------------------|----------------|------------------|
| AST (U/L) | 86 ± 11 | 310 ± 114 | 271 ± 27 | 170 ± 46* |
| ALT (U/L) | 31 ± 9 | 514 ± 170 | 433 ± 37 | 299 ± 133* |
| TG (mg/dL) | 36 ± 7 | 37 ± 13 | 52 ± 11* | 49 ± 8 |

Serum ALT and AST levels were significantly reduced in mice treated with ${\rm GTG} + {\rm HF} + 5$ % BTS compared to the control ${\rm GTG} + {\rm HF}$ group, *p < 0.05. There was no change in serum TG level between the ${\rm GTG} + {\rm HF} + 5$ % BTS and ${\rm GTG} + {\rm HF}$ groups although TG was elevated by 2 %BTS compared to ${\rm GTG} + {\rm HF} (*p < 0.05)$

PPAR- γ effects may also function to increase serum adiponectin, since PPAR- γ agonists have been shown to positively regulate serum adiponectin independently of adipose tissue volume regulation [28, 29].

To now study the mechanisms of inhibition of liver lipid metabolism by BTS in the presence of the activated adiponectin signaling pathway, AdipoR1 signaling pathways and their target genes were examined. It is known that activated AdipoR1 signaling decreases expression of SREBP-1c [30], a key regulator of hepatic fatty acid

synthesis [31, 32], through AMPK activation [33]. Here, we showed increased phosphorylation of AMPK by BTS in parallel with activated AdipoR1 signaling (Fig. 4c). Additionally, the phosphorylation of AMPK could also have been induced by activated PPAR-γ, in BTS treated mice, since as above BTS could also have PPAR-γ agonist like actions. Moreover, in the livers of mice treated with BTS, there was a reduced expression of SREBP-1c (Fig. 4b) in parallel with activated adiponectin signaling and phosphorylation of AMPK.



^{*} p < 0.05 vs GTG + HF, n = 6

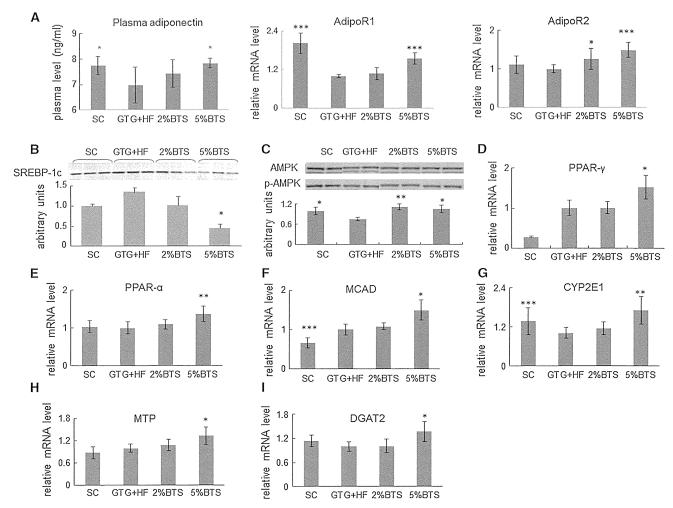


Fig. 4 BTS attenuates hepatosteatosis through activation of adiponectin. **a** Plasma adiponectin and expression of adiponectin receptors 1 (AdipoR1) and 2 (AdipoR2): plasma adiponectin levels and hepatic mRNA expression of AdipoR1 and AdipoR2 were dose dependently increased by BTS. *Asterisk* p < 0.05, *triple asterisk* p < 0.001 vs GTG + HF, n = 6. **b-d** AdioR1 signaling and target genes expression: BTS induced AdipoR1 signaling suppressed nuclear expression of SREBP-1c (n = 4) through phosphorylated AMPK (n = 4). Expression of PPAR-γ mRNA was also increased by BTS treatment

(n=6). Asterisk p < 0.05, double asterisk p < 0.01 vs GTG + HF. e-g Expression of PPAR- α and its target genes: Expression of PPAR- α mRNA was increase by BTS (n=6). Expression of MCAD (n=6) and CYP2E1 (n=6), PPAR- α target genes, was similarly increased by BTS. Asterisk p < 0.05, double asterisk p < 0.01, triple asterisk p < 0.001 vs GTG + HF. h, i BTS promotes hepatic lipid export: MRNA expression of MTP (n=6) and DGAT2 (n=6) was increased by treatment with BTS. Asterisk p < 0.05 vs GTG + HF

TNF- α expression is also known to regulate expression of SREBP-1c through activation of AMPK [31, 34]. Therefore, the suppression of TNF- α expression by BTS, as observed here, could also have contributed to the decreased SREBP-1c expression. Taken together, therefore, the data here suggest that activation of AdipoR1 signaling and consequent suppression of SREBP-1c may be central mechanisms through which BTS attenuates hepatic steatosis.

BTS may also have enhanced hepatic fatty acid oxidation via AdipoR2 signaling, known to increase the expression of PPAR- α [24] and its fatty acid oxidation related target genes. Here, the expression of PPAR- α (Fig. 4e) and its target gene MCAD (Fig. 4f) and CYP2E1 (Fig. 4g), were increased by treatment with BTS. Therefore, activated fatty

acids oxidation could also contribute to the attenuation of hepatic steatosis by BTS. Activation of fatty acid oxidation would be expected to increase production of reactive oxygen species (ROS) in the liver [35, 36]. BTS treatment here reduced ROS levels, probably contributing to the improved hepatic inflammation observed with BTS.

Additionally, MTP expression was confirmed here to be increased by treatment with BTS, possibly contributing to attenuation of steatosis by increasing hepatic export of TG (Fig. 4h). Interestingly, expression of DGAT2 was also increased by BTS treatment (Fig. 4i). Reduced hepatic accumulation of TG in BTS-treated livers may therefore have been due to increased MTP expression and increased expression of DGAT2 to reduce the content of FFA.



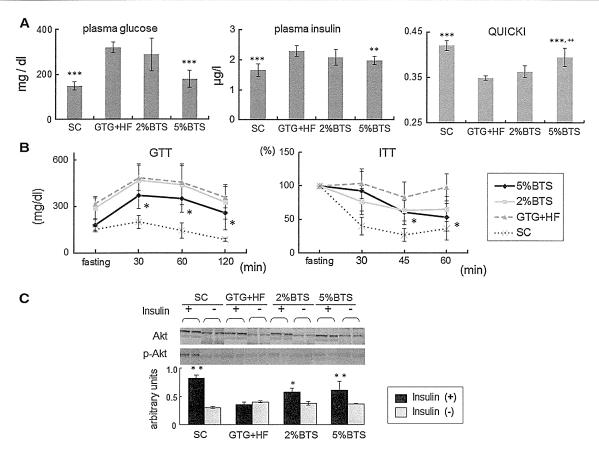


Fig. 5 Glucose intolerance and insulin resistance (IR) were attenuated by BTS. a Fasting plasma glucose, insulin levels and QUICKI: increased level of fasted plasma insulin and glucose were decreased, and QUICKI increased by treatment with 5 %BTS, and ITT confirmed severe IR double asterisk p < 0.01, triple asterisk p < 0.01 vs GTG + HF, double plus symbol p < 0.01 vs 2 %BTS, n = 6. b Glucose tolerance test (GTT) and insulin tolerance test

Furthermore, increased expression of DGAT2 might also have contributed to the reduction of ROS production by reducing hepatic FFA availability.

We next also investigated the effect of BTS on IR. GTG + HF induced elevation of fasting plasma glucose and plasma insulin were markedly reduced by BTS treatment, whilst QUICKI was markedly increased by BTS (Fig. 5a). In addition, ITT also was attenuated by BTS treatment. Furthermore, GTT showed that BTS remarkably attenuated severe glucose intolerance induced by GTG + HF (Fig. 5b). Phosphorylation of hepatic Akt after administration of insulin was reduced by treatment of BTS (Fig. 5c). The mechanisms through which BTS improves insulin sensitivity may involve Akt phosphorylation triggered through as yet uncertain mechanisms, but possibly involving induction of adiponectin signaling by BTS, since adiponectin is known to be involved in the suppression of hepatic gluconeogenesis and insulin secretion by activating AMPK [37].

In conclusion, therefore, BTS—a Japanese anti-obesity herbal (Kampo) medicine—is an effective preventive agent

(ITT): GTT revealed severe glucose intolerance was attenuated, and ITT confirmed IR was attenuated by 5 %BTS. Asterisk p < 0.05 vs GTG + HF, n = 6. c Involvement of Akt in BTS reduction of IR: phosphorylation of Akt was involved in the reduction of IR by BTS treatment as reflected by increased phosphorylation of Akt in the livers of BTS treated mice upon insulin administration. Asterisk p < 0.05, double asterisk p < 0.01 vs GTG + HF, p = 4

against the development of NASH. The challenge is now to identify the active components of BTS to allow its refinement and the rational design of more potent analogues.

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Conflict of interest The authors declare that they have no conflict of interest.

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References

1. Ioannou GN, Boyko EJ, Lee SP. The prevalence and predictors of elevated serum aminotransferase activity in the United States in 1999–2002. Am J Gastroenterol. 2006;101:76–82.

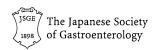


- James O, Day C. Non-alcoholic steatohepatitis: another disease of affluence. Lancet. 1999;353:1634–6.
- Guha IN, Parkes J, Roderick P, Chattopadhyay D, Cross R, Harris S, et al. Noninvasive markers of fibrosis in nonalcoholic fatty liver disease: validating the European liver fibrosis panel and exploring simple markers. Hepatology. 2008;47:455–60.
- Hioki C, Yoshimoto K, Yoshida T. Efficacy of bofu-tsusho-san, an oriental herbal medicine, in obese Japanese women with impaired glucose tolerance. Clin Exp Pharmacol Physiol. 2004; 31:614–9.
- Yamakawa J, Ishigaki Y, Takano F, Takahashi T, Yoshida J, Moriya J, et al. The Kampo medicines Orengedokuto, Bofutsushosan and Boiogito have different activities to regulate gene expressions in differentiated rat white adipocytes: comprehensive analysis of genetic profiles. Biol Pharm Bull. 2008;31:2083–9.
- Shimada T, Kudo T, Akase T, Aburada M. Preventive effects of Bofutsushosan on obesity and various metabolic disorders. Biol Pharm Bull. 2008;31:1362–7.
- Nakayama T, Suzuki S, Kudo H, Sassa S, Nomura M, Sakamoto S. Effects of three Chinese herbal medicines on plasma and liver lipids in mice fed a high-fat diet. J Ethnopharmacol. 2007;109: 236–40.
- Sakamoto S, Takeshita S, Sassa S, Zuki S, Ishikawa Y, Kudo H. Effects of colestimide and/or Bofu-tsusho-san on plasma and liver lipids in mice fed a high-fat diet. In vivo. 2005;19:1029–33.
- Ogasawara M, Hirose A, Ono M, Aritake K, Nozaki Y, Takahashi M, et al. A novel and comprehensive mouse model of human non-alcoholic steatohepatitis with the full range of dysmetabolic and histological abnormalities induced by gold thioglucose and a high-fat diet. Liver Int. 2011;31:542–51.
- Oike Y, Akao M, Yasunaga K, Yamauchi T, Morisada T, Ito Y, et al. Angiopoietin-related growth factor antagonizes obesity and insulin resistance. Nat Med. 2005;11:400–8.
- 11. Fan W, Yanase T, Nomura M, Okabe T, Goto K, Sato T, et al. Androgen receptor null male mice develop late-onset obesity caused by decreased energy expenditure and lipolytic activity but show normal insulin sensitivity with high adiponectin secretion. Diabetes. 2005;54:1000–8.
- Hirose A, Ono M, Saibara T, Nozaki Y, Masuda K, Yoshioka A, et al. Angiotensin II type 1 receptor blocker inhibits fibrosis in rat nonalcoholic steatohepatitis. Hepatology. 2007;45:1375–81.
- Albrecht C, Knaapen AM, Becker A, Höhr D, Haberzettl P, van Schooten FJ, et al. The crucial role of particle surface reactivity in respirable quartz-induced reactive oxygen/nitrogen species formation and APE/Ref-1 induction in rat lung. Respir Res. 2005;6:129.
- Guerre-Millo M, Rouault C, Poulain P, André J, Poitout V, Peters JM, et al. PPAR-alpha-null mice are protected from high-fat dietinduced insulin resistance. Diabetes. 2001;50:2809–14.
- Ma K, Cabrero A, Saha PK, Kojima H, Li L, Chang BH, et al. Increased beta-oxidation but no insulin resistance or glucose intolerance in mice lacking adiponectin. J Biol Chem. 2002;277: 34658–61.
- Taniguchi CM, Ueki K, Kahn R. Complementary roles of IRS-1 and IRS-2 in the hepatic regulation of metabolism. J Clin Invest. 2005;115:718–27.
- DeBose-Boyd RA, Ou J, Goldstein JL, Brown MS. Expression of sterol regulatory element-binding protein 1c (SREBP-1c) mRNA in rat hepatoma cells requires endogenous LXR ligands. Proc Natl Acad Sci USA. 2001;98:1477–82.
- Hui JM, Hodge A, Farrell GC, Kench JG, Kriketos A, George J. Beyond insulin resistance in NASH: TNF-alpha or adiponectin? Hepatology. 2004;40:46–54.
- Kamada Y, Takehara T, Hayashi N. Adipocytokines and liver disease. J Gastroenterol. 2008;43:811–22.

- Tomita K, Oike Y, Teratani T, Taguchi T, Noguchi M, Suzuki T, et al. Hepatic AdipoR2 signaling plays a protective role against progression of nonalcoholic steatohepatitis in mice. Hepatology. 2008;48:458–73.
- Rao MS, Reddy JK. PPARalpha in the pathogenesis of fatty liver disease. Hepatology. 2004;40:783–6.
- Reddy JK, Hashimoto T. Peroxisomal beta-oxidation and peroxisome proliferator-activated receptor alpha: an adaptive metabolic system. Annu Rev Nutr. 2001;21:193–230.
- Namikawa C, Shu-Ping Z, Vyselaar JR, Nozaki Y, Nemoto Y, Ono M, et al. Polymorphisms of microsomal triglyceride transfer protein gene and manganese superoxide dismutase gene in nonalcoholic steatohepatitis. J Hepatol. 2004;40:781–6.
- 24. Yamaguchi K, Yang L, McCall S, Huang J, Yu XX, Pandey SK, et al. Inhibiting triglyceride synthesis improves hepatic steatosis but exacerbates liver damage and fibrosis in obese mice with nonalcoholic steatohepatitis. Hepatology. 2007;45:1366–74.
- Postic C, Girard J. Contribution of de novo fatty acid synthesis to hepatic steatosis and insulin resistance: lessons from genetically engineered mice. J Clin Invest. 2008;118:829–38.
- Whiteman EL, Cho H, Birnbaum MJ. Role of Akt/protein kinase B in metabolism. Trends Endocrinol Metab (TEM). 2002;13:444–51.
- Kitajima Y, Eguchi Y, Ishibashi E, Nakashita S, Aoki S, Toda S, et al. Age-related fat deposition in multifidus muscle could be a marker for nonalcoholic fatty liver disease. J Gastroenterol. 2010; 45:218–24.
- 28. Tsuchida A, Yamauchi T, Takekawa S, Hada Y, Ito Y, Maki T, et al. Peroxisome proliferator-activated receptor (PPAR)alpha activation increases adiponectin receptors and reduces obesity-related inflammation in adipose tissue: comparison of activation of PPARalpha, PPARgamma, and their combination. Diabetes. 2005;54:3358–70.
- Waki H, Yamauchi T, Kamon J, Ito Y, Uchida S, Kita S, et al. Impaired multimerization of human adiponectin mutants associated with diabetes. Molecular structure and multimer formation of adiponectin. J Biol Chem. 2003;278:40352–63.
- Shklyaev S, Aslanidi G, Tennant M, Prima V, Kohlbrenner E, Kroutov V, et al. Sustained peripheral expression of transgene adiponectin offsets the development of diet-induced obesity in rats. Proc Natl Acad Sci USA. 2003;100:14217–22.
- 31. Browning JD, Horton JD. Molecular mediators of hepatic steatosis and liver injury. J Clin Invest. 2004;114:147–52.
- 32. Yamauchi T, Nio Y, Maki T, Kobayashi M, Takazawa T, Iwabu M, et al. Targeted disruption of AdipoR1 and AdipoR2 causes abrogation of adiponectin binding and metabolic actions. Nat Med. 2007;13:332–9.
- 33. Yano W, Kubota N, Itoh S, Kubota T, Awazawa M, Moroi M, et al. Molecular mechanism of moderate insulin resistance in adiponectin-knockout mice. Endocr J. 2008;55:515–22.
- Endo M, Masaki T, Seike M, Yoshimatsu H. TNF-alpha induces hepatic steatosis in mice by enhancing gene expression of sterol regulatory element binding protein-1c (SREBP-1c). Exp Biol Med (Maywood). 2007;232:614–21.
- 35. Evans JL, Goldfine ID, Maddux BA, Grodsky GM. Are oxidative stress-activated signaling pathways mediators of insulin resistance and beta-cell dysfunction? Diabetes. 2003;52:1–8.
- 36. Leclercq IA, Farrell GC, Field J, Bell DR, Gonzalez FJ, Robertson GR. CYP2E1 and CYP4A as microsomal catalysts of lipid peroxides in murine nonalcoholic steatohepatitis. J Clin Invest. 2000;105:1067–75.
- Yamauchi T, Kamon J, Minokoshi Y, Ito Y, Waki H, Uchida S, et al. Adiponectin stimulates glucose utilization and fatty-acid oxidation by activating AMP-activated protein kinase. Nat Med. 2002;8:1288–95.



ORIGINAL ARTICLE—LIVER, PANCREAS, AND BILIARY TRACT



The association of insomnia with gastroesophageal reflux symptoms in biopsy-proven nonalcoholic fatty liver disease

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Abstract

Background It is suggested that nonalcoholic fatty liver disease (NAFLD), including nonalcoholic fatty liver (NAFL) and nonalcoholic steatohepatitis (NASH), can be associated with insomnia and gastro-esophageal reflux disease (GERD). The relationship between GERD and insomnia in subjects with biopsy-proven NAFLD was investigated.

Methods This study enrolled 123 patients with biopsy-proven NAFLD. Insomnia was assessed by the Athens Insomnia Scale (AIS), a self-assessment psychometric instrument designed to quantify sleep difficulty based on ICD-10 criteria; AIS scores ≥ 6 were considered positive for insomnia. GERD symptoms were evaluated using a frequency scale for the symptoms of GERD (FSSG); FSSG scores ≥ 8 were considered positive. Logistic regression models were used to evaluate the association of insomnia with GERD, after adjusting for potential confounders.

Thirteen patients with GERD were treated with the proton pump inhibitor rabeprazole (RPZ; 10 mg/day), for 12 weeks.

Results Of the 123 patients, 76 (62 %) were female and 87 (71 %) were obese, with 34 (28 %) having AIS scores \geq 6 and 31 (25 %) having FSSG scores \geq 8. Liver biopsy revealed that 40 patients (33 %) had NAFL and 83 (67 %) had NASH. FSSG and AIS scores were similar in the two groups. HOMA-IR, FSSG scores and γ GT (GGT) concentrations were significantly higher in insomniacs than in non-insomniacs. Logistic regression analysis demonstrated that FSSG score and GGT concentration were independently associated with insomnia. RPZ treatment resulted in significantly reductions in both AIS and FSSG scores.

Conclusions Nearly 30 % of patients with biopsy-proven NAFLD had insomnia, which was related to GGT and GERD and could be relieved by RPZ treatment.

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Keywords GERD · Insomnia · Nonalcoholic fatty liver disease · Proton-pump inhibitor

Abbreviations

BMI Body mass index

GERD Gastro-esophageal reflux disease NAFLD Nonalcoholic fatty liver disease NASH Nonalcoholic steatohepatitis

Introduction

Nonalcoholic fatty liver disease (NAFLD) [1] is the most common chronic liver disease in many developed countries and results in serious public health problems worldwide. NAFLD includes a wide spectrum of liver diseases, ranging from nonalcoholic fatty liver (NAFL), which is usually benign, to nonalcoholic steatohepatitis (NASH), which may progress to liver cirrhosis (LC), hepatic failure and hepatocellular carcinoma (HCC) in the absence of significant alcohol consumption [2]. A large proportion of NA-FLD patients are asymptomatic, but some occasionally experience fatigue, anxiety, and/or insomnia, resulting in a significant decrement in quality of life (QOL) [3]. In middle-aged Koreans, short sleep duration and poor sleep quality were found to be significantly associated with an increased risk of NAFLD [4]. Similarly, short sleep duration was associated with NAFLD in the general Japanese population [5]. However, the mechanisms underlying the association between insomnia and NAFLD remain unknown. Sleep is important to maintain body homeostasis, with sleep problems associated with all-cause mortality [6].

In addition to being associated with sleep problems, NAFLD was found, in two recent studies from Japan and Italy, to be associated with a high prevalence of the symptoms of gastro-esophageal reflux disease (GERD) [7, 8]. Evidence has emerged suggesting a link between metabolic syndrome, specifically obesity and visceral fat accumulation, and the onset of GERD. Studies throughout the world have shown that GERD is associated with sleep problems [9-14]. For example, a population-based study from Sweden showed positive associations among the presence of insomnia, sleeplessness, problems falling asleep, and risk of GERD [10]. In addition, an analysis of 19864 healthy adults in Japan found that poor sleep quality and irregular dietary habits were strong risk factors for high scores on the frequency scale for the symptoms of GERD (FSSG) [15]. Thus, it can be hypothesized that GERD symptoms may be responsible for insomnia in patients with NAFLD. To our knowledge, no study to date has assessed the prevalence of insomnia or GERD, or their association, in patients with biopsy-proven NAFLD.

Rabeprazole (RPZ), a proton pump inhibitor (PPI), is a potent and irreversible inhibitor of the H(+)/K(+)-ATP-ase gastric pump and is indicated for the treatment of GERD, Zollinger-Ellison syndrome, and duodenal and gastric ulcers. Moreover, the combination of RPZ and antibiotics is indicated for the eradication of Helicobacter pylori. RPZ is therefore expected to be effective in the treatment of GERD patients with sleep disturbances [16, 17]. This study was designed to evaluate the prevalence of insomnia and GERD in patients with biopsy-proven NAFLD; to compare the rates of insomnia and GERD in patients with NASH and NAFL; to determine independent predictors of insomnia, including FSSG score, among these patients; and to evaluate the effect of RPZ on insomnia.

Methods

Study population

The study included a total of 123 patients with wellcharacterized, liver biopsy-confirmed NAFLD who completed the FSSG questionnaire assessing symptoms of GERD and the Athens Insomnia Scale (AIS) questionnaire. All patients underwent biopsies at one of the seven hepatology centers included in the Japan Study Group of NAFLD (JSG-NAFLD): Center for Digestive and Liver Diseases, Nara City Hospital; Division of Gastroenterology, Yokohama City University Graduate School of Medicine; Department of Medicine and Molecular Science, Graduate School of Biomedical Sciences, Hiroshima University; Department of Gastroenterology and Hepatology, Kochi Medical School; Department of Internal Medicine, Saga Medical School, Saga University; Department of Hepatology, Graduate School of Medicine, Osaka City University; and the Department of Gastroenterology and Hepatology, Kyoto Prefectural University of Medicine.

NAFLD was diagnosed based on liver biopsy findings of steatosis in ≥ 5 % of hepatocytes and the exclusion of other liver diseases, including viral hepatitis, autoimmune hepatitis, drug-induced liver disease, primary biliary cirrhosis, biliary obstruction, hemochromatosis, Wilson's disease, and α -1-antitrypsin-deficiency-associated liver disease. Patients consuming more than 20 g of alcohol per day, those with evidence of decompensated LC or HCC, those with psychiatric disorders or psychiatric drug users, and those taking PPIs and/or histamine H2-receptor antagonists were excluded. All patients provided written informed consent at the time of liver biopsy, and the study was conducted in conformance with the Declaration of Helsinki.



Laboratory and clinical parameters

Venous blood samples were taken in the morning after a 12-h overnight fast. Laboratory assays included blood cell counts and measurements of serum concentrations of aspartate aminotransferase (AST), alanine aminotransferase (ALT), gamma glutamyl transpeptidase (GGT), cholinesterase (ChE), total cholesterol, triglycerides, fasting plasma glucose (FPG), immunoreactive insulin (IRI), ferritin, hyaluronic acid, and type IV collagen 7S. These parameters were measured using the standard techniques of clinical chemistry laboratories.

Body mass index (BMI) was calculated as weight in kilograms/(height in meters)², with obesity defined as a BMI > 25 kg/m², according to the criteria of the Japan Society for the Study of Obesity [18]. Patients taking oral hypoglycemic medication, and those with a random glucose concentration > 200 mg/dl or a fasting glucose concentration > 126 mg/dl, were regarded as positive for hyperglycemia [19]. Patients with serum cholesterol concentrations > 220 mg/dl or triglyceride concentrations > 160 mg/dl were diagnosed with dyslipidemia. Patients taking antihypertensive agents and those having a resting recumbent blood pressure \geq 140/90 mmHg on at least two occasions were regarded as having hypertension [20].

GERD score

The FSSG is a questionnaire widely used to diagnose GERD [21–24] and to evaluate the effectiveness of any treatment [21, 25]. The FSSG consisted of 12 questions assessing the frequency of symptoms (never, 0; occasionally, 1; sometimes, 2; often, 3; and always, 4). Patients with FSSG scores \geq 8 were considered positive for GERD; at this cut-off point, the FSSG had a sensitivity of 62 %, a specificity of 59 %, and an accuracy of 60 % in assessing GERD [21].

Insomnia scale

The intensity of sleep difficulty was evaluated using the AIS, a self-administered psychometric tool with high consistency, reliability and external validity (Table 1) [26, 27]. The AIS consists of eight items, five of which are used to assess insomnia, and the three used to assess well-being, functional capacity, and sleepiness during the day. The full eight-item version (AIS-8) was developed for clinical settings, while the five-item version (AIS-5) can be used to assess sleep quantity and quality. These first five questions (AIS-5) are used to assess difficulty with sleep induction, awakenings during the night, early morning awakening, total sleep time and overall quality of sleep. The last three items in the AIS-8 refer to

Table 1 Athens Insomnia Scale (AIS) [26]

| Sleep induction (time | e it takes you to fall asleep after tu | rming-off the lights) | |
|------------------------|--|----------------------------|--|
| 0: No problem | 1: Slightly delayed | 2: Markedly delayed | 3: Very delayed or did not sleep at all |
| Awakening during th | ne night | | |
| 0: No problem | 1: Minor problem | 2: Considerable problem | 3: Serious problem or did not sleep at all |
| Final awakening earl | ier than desired | | |
| 0: Not earlier | 1: A little earlier | 2: Markedly earlier | 3: Much earlier or did not sleep at all |
| Total sleep duration | | | |
| 0: Sufficient | 1: Slightly insufficient | 2: Markedly insufficient | 3: Very insufficient or did not sleep at all |
| Overall quality of sle | eep (no matter how long you slept) | 1 | |
| 0: Satisfactory | 1: Slightly unsatisfactory | 2: Markedly unsatisfactory | 3: Very unsatisfactory or did not sleep at all |
| Sense of well-being | during the day | | |
| 0: Normal | 1: Slightly decreased | 2: Markedly decreased | 3: Very decreased |
| Functioning (physica | l and mental) during the day | | |
| 0: Normal | 1: Slightly decreased | 2: Markedly decreased | 3: Very decreased |
| Sleepiness during the | e day | | |
| 0: None | 1: Mild | 2: Considerable | 3: Intense |

Instructions this scale is intended to record own assessment of any sleep difficulty you might have experienced. Please, check (by circling the appropriate number) the items above to indicate your estimate of any difficulty, provided that it occurred at least three times per week during the last month

The period of the self-assessment may vary, depending on the design of a given study. Whenever the self-assessment pertains to a period other than that of the last month, the second sentence of the instructions should be rephrased accordingly



daytime symptoms that often result from sleep disorders, such as narcolepsy and obstructive sleep apnea, in patients with insomnia. Each item on the AIS was rated from 0 (*no problem at all*) to 3 (*very serious problem*). Total scores can range from 0 to 24, with scores \geq 6 and < 6 representing the presence and absence of insomnia, respectively. This cutoff point had a sensitivity of 93 %, a specificity of 85 % (90 % overall correct case identification), a positive predictive value (PPV) of 41 % and a negative predictive value (NPV) of 99 % [27].

Responders were asked to calculate their scores if they had experienced sleep difficulties at least three times a week during the previous month.

Liver histology

All enrolled patients underwent a percutaneous liver biopsy under ultrasonic guidance or peritoneoscopy. The liver specimens were embedded in paraffin and stained with hematoxylin and eosin, Masson-trichrome, reticulin silver stain, and Perls' Prussian blue. The specimens were evaluated by two hepatic pathologists (S.T. and Y.S.), who were blinded to the clinical findings. An adequate liver biopsy sample was defined as a specimen of length > 1.5 cm and/or having more than 6 portal tracts. NASH was defined as steatosis with lobular inflammation and ballooning degeneration, with or without Mallory-Denk body or fibrosis. Patients with liver biopsy specimens showing simple steatosis or steatosis with nonspecific inflammation were identified as the NAFL [28]. Specimens with steatosis 5-33, > 33-66, and > 66 % were scored as having steatosis grades of 0, 1, 2, and 3, respectively [29]. Histological grade and stage were scored as described [30]. Necroinflammatory grades of 1, 2, and 3, were defined as mild, moderate and severe hepatocellular steatosis,

Table 2 Clinical characteristics of enrolled patients with NAFL and NASH

| Clinical parameter | Total $(n = 123 [100 \%])$ | NAFL $(n = 40 [33 \%])$ | NASH $(n = 83 [67 \%])$ | P value |
|--|----------------------------|-------------------------|-------------------------|----------|
| Age (years) | 59 (14–82) | 56 (20–78) | 62 (14–82) | 0.0025 |
| Gender (female) | 76 (62 %) | 17 (43 %) | 59 (71 %) | 0.0030 |
| BMI (kg/m ²) | 26.6 (16.6–43.4) | 26.6 (18.9–43.4) | 27.3 (16.6–41.0) | 0.1405 |
| Obesity (BMI > 25) | 87 (71 %) | 27 (68 %) | 60 (72 %) | 0.6731 |
| Dyslipidemia (yes [%]) | 46 (37 %) | 12 (30 %) | 34 (41 %) | 0.3202 |
| Hypertension (yes [%]) | 48 (39 %) | 10 (25 %) | 38 (46 %) | 0.0310 |
| Type 2 diabetes (yes [%]) | 55 (45 %) | 13 (33 %) | 42 (51 %) | 0.0811 |
| Hemoglobin (g/dl) | 14.1 (10.5–18.3) | 14.8 (10.6–18.3) | 13.8 (10.5–16.7) | 0.0728 |
| Platelet count ($\times 10^4/\mu l$) | 21.4 (4.6–78.5) | 23.5 (13.0–78.5) | 20.8 (4.6-45.4) | 0.0125 |
| AST (IU/l) | 45 (17–186) | 37 (17–151) | 51 (18–186) | 0.0001 |
| ALT (IU/l) | 69 (12–358) | 61 (15–358) | 71 (12–218) | 0.2425 |
| GGT (IU/l) | 61 (20–391) | 60 (20–319) | 62 (21–391) | 0.6382 |
| Cholinesterase (IU/l) | 371 (167–547) | 378 (266–545) | 370 (167–547) | 0.2873 |
| Total cholesterol (mg/dl) | 209 (87–335) | 218 (127–335) | 203 (87–319) | 0.0183 |
| Triglyceride (mg/dl) | 156 (61–659) | 155 (61–416) | 162 (66–659) | 0.4849 |
| HDL-C (mg/dl) | 50 (23–290) | 49 (31–77) | 52 (23–290) | 0.7727 |
| Ferritin (ng/ml) | 163 (5–1100) | 113 (10–1100) | 210 (5–923) | 0.0160 |
| FPG (mg/dl) | 96 (60–452) | 96 (60–161) | 96 (60–452) | 0.3571 |
| IRI (μU/ml) | 11.4 (1.59–49.5) | 8.4 (1.6–46) | 13.2 (2.8–49.5) | < 0.0001 |
| HOMA-IR | 2.62 (0.38–33.04) | 1.87 (0.38–13.63) | 3.02 (0.65–33.04) | < 0.0001 |
| Hyaluronic acid (ng/ml) | 37 (9–3480) | 22 (9–149) | 49 (9–3480) | 0.0001 |
| Type IV collagen 7S (ng/ml) | 4.5 (2.7–13) | 3.7 (2.8–7.1) | 5.1 (2.7–13.0) | < 0.0001 |
| FSSG | 4 (0–38) | 4 (0–29) | 3 (0–38) | 0.5009 |
| $FSSG \ge 8 \ (n \ [\%])$ | 31 [25 %] | 10 [25 %] | 21 [25 %] | 1.0000 |
| AIS | 3 (0–15) | 3 (0–12) | 4 (0–15) | 0.5591 |
| $AIS \ge 6 \ (n \ [\%])$ | 34 [28 %] | 10 [25 %] | 24 [29 %] | 0.8299 |

Results are presented as numbers with percentages in parenthesis for qualitative data or as mean \pm SD for quantitative data

BMI body mass index, AST aspartate aminotransferase, ALT alanine aminotransferase, GGT gamma glutamyl transpeptidase, FPG fasting plasma glucose, IRI immuno-reactive insulin

P values were calculated by t test or χ^2 analysis



ballooning and inflammation (acinar and portal), respectively. The severity of hepatic fibrosis (stage) was scored as: stage 1, zone 3 perisinusoidal fibrosis; stage 2, zone 3 perisinusoidal fibrosis with portal fibrosis; stage 3, zone 3 perisinusoidal fibrosis and portal fibrosis with bridging fibrosis; and stage 4, cirrhosis.

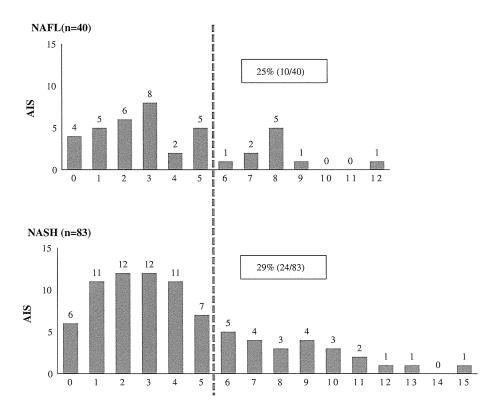
Treatment with RPZ

Thirteen NAFLD patients with GERD symptoms (11 females and 2 males) were administered 10 mg/day RPZ for 12 weeks. These patients completed both the FSSG and AIS before and after RPZ treatment.

Statistical analysis

Quantitative results are presented as medians and ranges, and qualitative results as numbers and percentages. Statistical differences in quantitative data were determined using the Mann–Whitney U test or Wilcoxon rank-sum test, and differences in qualitative data using Fisher's exact probability test or χ^2 analysis (Tables 2, 4, 5; Figs. 1, 2, 3, 4). Correlations were calculated by Spearman rank correlation analysis (Table 3). Multivariate logistic regression analysis was used to identify variables independently associated with the occurrence of insomnia (Table 6). Statistical significance was defined as a P value < 0.05.

Fig. 1 The distribution of Athens Insomnia scale (AIS)



2 Springer

Results

Characteristics of study subjects

Table 2 summarizes the clinical and laboratory data of the patient population. Of the 123 patients with NAFLD, 76 (62 %) were female, and 87 (71 %) were obese (BMI > 25 kg/m²). Histologically, 83 patients (67 %) were diagnosed with NASH, and 40 (33 %) with NAFL. Patients with NASH were significantly older; were more predominantly female; were more likely to have hypertension and type 2 diabetes; had lower platelet counts and total cholesterol concentrations; and had higher levels of AST, ferritin, IRI, HOMA-IR, hyaluronic acid, and type IV collagen 7S. Of the 83 patients with NASH, 41 (49 %), 22 (27 %), 13 (16 %), and 7 (8 %) had stage 0–1, 2, 3, and 4 fibrosis, respectively.

Comparisons between NASH and NAFL

The distribution of AIS scores in patients with NAFL and NASH is shown in Fig. 1. Overall, 34 of the 123 patients (28 %) with NAFLD had AIS scores \geq 6, diagnostic of insomnia, including 10 of 40 (25 %) patients with NAFL and 24 of 83 (29 %) with NASH (P=0.8299). Males and females had similar median AIS scores [3 (range 0–13) vs. 3 (range 0–15), P=0.7954] and a similar prevalence of insomnia [26 % (12/47] vs. 29 % (22/76), P=0.8359].

Fig. 2 Correlation between FSSG and AIS. A significant positive correlation was found between AIS and FSSG

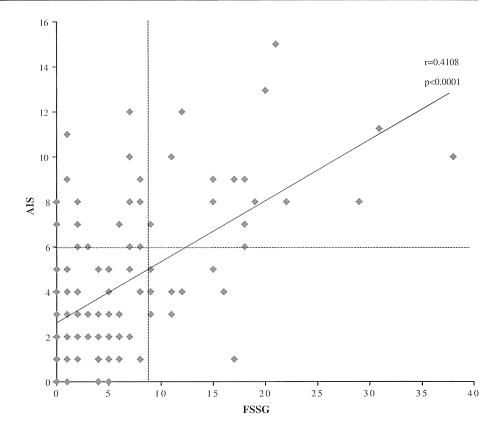
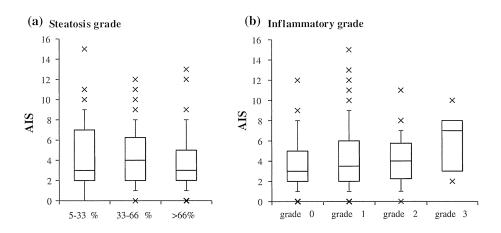


Fig. 3 Relationship between AIS and histological findings. The *box* represents the interquartile ranges (25 and 75 %) from the median (*horizontal line*). The *bars* indicate the 10 and 90 %



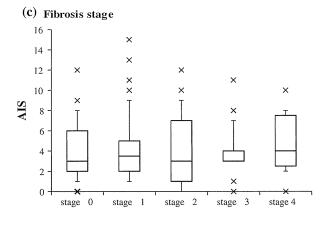




Fig. 4 Effects of rabeprazole (RPZ) on the improvement of GERD symptoms and insomnia. The box represents the interquartile ranges (25 and 75 %) from the median (horizontal line). The bars indicate the 10 and 90 %. a Change in the total FSSG score. RPZ significantly reduced total FSSG scores. *P = 0.0071compared to baseline response before treatment. b Change in the total AIS. RPZ significantly reduced total AIS. *P = 0.0144compared to baseline response before treatment

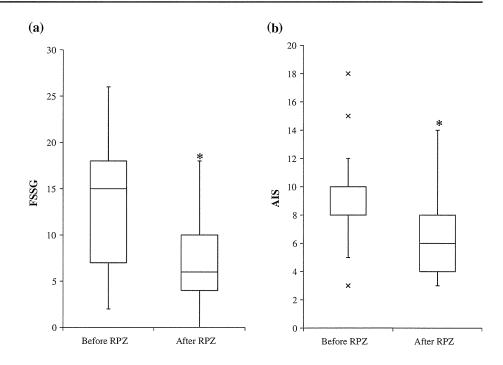


Table 3 Correlation between AIS/FSSG and clinical parameters in 123 patients with biopsy-proven NAFLD

| Variables | AIS | | FSSG | |
|---------------------|-------------------------|----------|-------------------------|---------|
| | Correlation coefficient | P value | Correlation coefficient | P value |
| Age | -0.0431 | 0.6363 | -0.2363 | 0.0085 |
| BMI | $^{-}0.0075$ | 0.9345 | 0.091 | 0.3128 |
| Hemoglobin | 0.0104 | 0.9100 | 0.0328 | 0.7220 |
| Platelet | 0.1079 | 0.2407 | 0.2197 | 0.0159 |
| AST | 0.0588 | 0.5197 | 0.0015 | 0.9868 |
| ALT | 0.0284 | 0.7558 | 0.1148 | 0.2080 |
| AST/ALT ratio | -0.0046 | 0.9597 | -0.1254 | 0.1689 |
| γGT | 0.1545 | 0.0935 | -0.1272 | 0.1681 |
| Cholinesterase | 0.1366 | 0.1646 | 0.1683 | 0.0861 |
| Prothrombin time | 0.0359 | 0.6998 | 0.1403 | 0.1296 |
| Cholesterol | 0.1366 | 0.7233 | 0.0624 | 0.5039 |
| Triglyceride | -0.0343 | 0.7132 | 0.0506 | 0.5879 |
| HDL-C | -0.0128 | 0.8943 | -0.1166 | 0.2230 |
| FPG | 0.0442 | 0.6316 | -0.2010 | 0.0277 |
| IRI | -0.1073 | 0.2412 | -0.1570 | 0.0855 |
| HOMA-IR | -0.0993 | 0.2806 | -0.1948 | 0.0330 |
| Ferritin | 0.0934 | 0.3081 | -0.0490 | 0.5932 |
| Hyaluronic acid | -0.0340 | 0.7088 | -0.2246 | 0.0125 |
| Type IV collagen 7S | -0.0193 | 0.8323 | -0.1277 | 0.1592 |
| FSSG | 0.4108 | < 0.0001 | _ | _ |

P values are based on Spearman's non-parametric correlation analysis



FSSG scored did not differ in NAFL and NASH patients, with the prevalence of GERD being 25 % in each group.

Factors positively correlating with AIS and FSSG scores

AIS score was positively correlated with FSSG score $(r=0.4108,\,P<0.001)$ (Fig. 2), but not with any other parameter (Table 3). FSSG score was positively correlated with platelet count $(r=0.2197,\,P=0.0159)$, and negatively correlated with age $(r=-0.2363,\,P=0.0085)$, FPG concentration $(r=-0.2010,\,P=0.0277)$, HOMA-IR score $(r=-0.1948,\,P=0.0330)$ and hyaluronic acid concentration $(r=-0.2246,\,P=0.0125)$. BMI, transaminase activities, lipid profiles, and iron parameters were not correlated with AIS or FSSG score.

Correlation between histological findings and AIS scores

Assessment of histological findings in the 123 patients with NAFLD showed that 53 (43 %), 40 (33 %), and 30 (24 %) had steatosis grades 1, 2, and 3, respectively; 30 (24 %), 70 (57 %), 18 (15 %), and 5 (4 %) had inflammation grades 0,

1, 2, and 3, respectively; and 44 (36 %), 38 (31 %), 21 (17 %), 13 (11 %), and 7 (6 %) had fibrosis grades 0, 1, 2, 3, and 4, respectively. Evaluation of correlations between AIS scores and histological findings showed that AIS score was not correlated with steatosis, inflammation, or fibrosis grade (Fig. 3).

Clinical findings in patients with and without insomnia

Comparisons of clinical and laboratory findings in patients with and without insomnia showed that γ GT concentrations and FSSG scores were higher, and the prevalence of hypertension and IRI and HOMA-IR scores were lower, in patients with insomnia (Table 4). Moreover, GERD symptoms were significantly more prevalent in patients with than without insomnia (56 vs. 13 %, P < 0.0001).

Drug usage

Drug usage in patients involved in this study was shown in Table 5. Beta-blockers users were more prevalent in patients with GERD compared to those without. The prevalence of other drug users was not different between patients with GERD/insomnia and those without.

Table 4 The comparison between insomniacs and non-insomniacs

| Clinical parameter | Insomniacs ($n = 34 [28 \%]$) | Non-insomniacs ($n = 89 [72 \%]$) | P value |
|--|---------------------------------|-------------------------------------|----------|
| Age (years) | 56 (35–74) | 60 (14–82) | 0.8100 |
| Gender (female) | 22 (65 %) | 54 (61 %) | 0.8359 |
| BMI (kg/m^2) | 26.4 (21.9–38.6) | 26.6 (16.6–43.4) | 0.5028 |
| Obesity (BMI > 25) | 25 (74 %) | 62 (70 %) | 0.8252 |
| Dyslipidemia | 21 (62 %) | 56 (63 %) | 1.0000 |
| Hypertension (yes) | 8 (24 %) | 40 (45 %) | 0.0385 |
| Type 2 diabetes (yes) | 16 (47 %) | 39 (44 %) | 0.8400 |
| Hemoglobin (g/dl) | 14.4 (11.0–18.3) | 14.0 (10.5–17.1) | 0.9072 |
| Platelet count ($\times 10^4/\mu l$) | 22.3 (8.7–33.5) | 21.1 (4.6–78.5) | 0.7137 |
| AST (IU/I) | 47 (20–182) | 44 (17–186) | 0.4682 |
| ALT (IU/l) | 71 (12–358) | 69 (15–218) | 0.6233 |
| GGT (IU/l) | 77 (24–391) | 59 (20–268) | 0.0063 |
| Cholinesterase (IU/l) | 373 (208–547) | 371 (167–545) | 0.7992 |
| Total cholesterol (mg/dl) | 206 (125–314) | 214 (87–335) | 0.3670 |
| Triglyceride (mg/dl) | 146 (68–424) | 164 (61–659) | 0.3864 |
| Ferritin (ng/ml) | 128 (11–1100) | 170 (5–923) | 0.7315 |
| FPG (mg/dl) | 94 (70–452) | 97 (60–171) | 0.7708 |
| IRI (μU/ml) | 9.8 (2.8–11.2) | 12.0 (1.6–49.5) | 0.0326 |
| HOMA-IR | 2.10 (0.65-33.04) | 2.96 (0.38–16.34) | 0.0335 |
| Hyaluronic acid (ng/ml) | 32 (9–392) | 37 (9–3480) | 0.9774 |
| Type IV collagen 7S (ng/ml) | 4.7 (2.7–13.0) | 4.4 (2.8–10.0) | 0.9054 |
| FSSG | 8 (0–38) | 3 (0–17) | < 0.0001 |
| $FSSG \ge 8 \ (n \ [\%])$ | 19 [56 %] | 12 [13 %] | < 0.0001 |
| NASH (n [%]) | 24 [71 %] | 59 [66 %] | 0.8299 |

numbers with percentages in parenthesis for qualitative data or as mean ± SD for quantitative data *BMI* body mass index, *AST* aspartate aminotransferase, *ALT* alanine aminotransferase, *GGT* gamma glutamyl transpeptidase, *FPG* fasting plasma glucose, *IRI* immuno-reactive insulin

Results are presented as

P values were calculated by t test or χ^2 analysis



Table 5 Drug usage

| Drug usage | FSSG < 8 (n = 92) | $FSSG \ge 8$ $(n = 31)$ | P value | Insomniacs $(n = 34)$ | Non-insomniacs $(n = 89)$ | P value |
|-----------------------------|----------------------|-------------------------|---------|-----------------------|---------------------------|---------|
| Use of antihypertensive dru | ugs | | | | | |
| Calcium antagonists | 26 [28 %] | 10 [32 %] | 0.6560 | 7 [21 %] | 29 [33 %] | 0.2680 |
| ARBs | 4 [4 %] | 2 [6 %] | 0.6412 | 2 [6 %] | 4 [4 %] | 0.6679 |
| Beta-blockers | 2 [2 %] | 4 [13 %] | 0.0348 | 2 [6 %] | 4 [4 %] | 0.6679 |
| Use of NSAIDs | 10 [11 %] | 2 [6 %] | 0.7285 | 1 [3 %] | 11 [12 %] | 0.1762 |
| Use of anticoagulants | 2 [2 %] | 0 [0 %] | 1.0000 | 0 [0 %] | 2 [2 %] | 1.0000 |
| Use of digestive drugs | 2 [2 %] | 0 [0 %] | 1.0000 | 0 [0 %] | 2 [2 %] | 1.0000 |

ARBs angiotensin receptor blockers, NSAIDs non-steroidal anti-inflammatory drugs

Table 6 Results of multivariate analysis: independent predictors of insomniacs

| Variables | Adjusted (multivariate) | | | |
|--------------|-------------------------|---------------|----------|--|
| | OR | 95 % CI | P value | |
| FSSG | 1.2315 | 1.1221–1.3516 | < 0.0001 | |
| GGT | 1.0109 | 1.0031-1.0189 | 0.0063 | |
| IRI | 0.9515 | 0.8908-1.0164 | 0.1396 | |
| Hypertension | 0.5503 | 0.1823-1.6610 | 0.2893 | |

OR odds ratio, *CI* confidence interval, *FSSG* frequency scale for the symptoms of GERD, *GGT* gamma glutamyl transpeptidase, *IRI* immuno-reactive insulin

Factors independently predictive of insomnia in patients with NAFLD

Multivariate logistic regression analysis showed that plasma GGT concentration [odds ratio (OR) 1.2315, 95 % confidence interval (CI) 1.1221–1.3516, P < 0.0001] and FSSG score (OR 1.0109, 95 % CI 1.0031–1.0189, P = 0.0063) were significant independent predictors of insomnia (Table 6). In contrast, IRI and HOMA-IR scores and hypertension were not predictive.

Effects of RPZ on insomnia

RPZ treatment of patients with GERD significantly reduced FSSG and AIS scores (Fig. 4). Of the 9 patients with insomnia treated with RPZ, 4 (44 %) showed resolution of insomnia after treatment.

Discussion

This study of Japanese NAFLD patients demonstrated that (1) 28 % had AIS scores \geq 6, indicative of insomnia; (2) 25 % had FSSG scores \geq 8, indicative of GERD; (3) AIS and FSSG scores did not differ significantly in patients

with NASH and NAFL; (4) FSSG score was independently associated with AIS score; and (5) insomnia could be relieved after treatment with RPZ.

The precise prevalence of insomnia in NAFLD patients has been unclear. Using the AIS, we found that 28 % of patients with biopsy-proven NAFLD had insomnia. Although sleep dysfunction has been defined as a Pittsburgh Sleep Quality Index (PSQI) score > 5.5 in other studies [17, 31], the AIS is satisfactorily validated, simple to perform, and well accepted based on ICD-10 criteria. The AIS has been used to assess insomnia in the general population in Japan. For example, a study of approximately 3000 individuals found that 21.4 % had experienced insomnia during the previous month [32]. The prevalence of insomnia in our NAFLD patients does not seem to be markedly different from that in the general population. Though this precise reason is unknown, one plausible explanation is that about 20-30 % of the general population is estimated to have NAFLD. We should obtain data from sex- and age-matched non-NAFLD population to clarify whether the prevalence of insomnia in NAFLD is really higher compared to that in the general population. Assessments of employees of two local governments in Japan found that 1382 of 5951 males (23.2 %) and 465 of 1500 females (31.0 %), aged 34-59 years, had insomnia [33, 34]. Assessments of middle-aged women found that 27.5-43.6 % had AIS scores ≥ 6 [35, 36]. Taken together, these findings indicate that the prevalence of insomnia is higher in women than in men, across countries and cultures. In contrast, we observed no differences in AIS scores between men and women with NAFLD. Since the discrepancy between our results and previous studies can be explained by a small number of patients involved in this study, a larger number of patients should be examined in the future. Since Yoshioka et al. [34] showed that the gender difference disappeared after adjustment for paid work and family responsibilities, detailed characteristics of patients should be considered to clarify the gender differences. In Japanese studies in which insomnia was



diagnosed using AIS, factors associated with insomnia included work at visual display terminals for ≥ 6 h per day [37], job stress [38, 39], and reduced illumination in the workplace [40]. In this study, however, we did not evaluate these work environmental factors.

The mechanisms by which insomnia arises in patients with NAFLD have never been clarified. We found that AIS scores and the incidence of insomnia were similar in patients with NAFL and NASH. AIS scores did not correlate with any histological findings, such as steatosis, inflammation, and fibrosis scores, indicating that histological severity is not important in the pathogenesis of insomnia in patients with NAFLD. In contrast, many studies have explored the associations between life-style related disorders/obesity and sleep disturbance. Changes in secretion of the hormones cortisol, leptin, and ghrelin, and increased insulin resistance due to short sleep duration were found to increase the risks of obesity and diabetes [41–43]. In contrast, insomniacs were more likely to have insulin resistance than non-insomniacs. This study demonstrated that FSSG score was significantly correlated only with AIS score. Multivariate analysis showed that FSSG was an independent risk factor associated with insomnia, suggesting that GERD symptoms are responsible for insomnia in NAFLD patients, findings consistent with previously reported results [9, 17]. For example, sleep disorders, such as inability to sleep, difficult falling asleep, and awakening during the night, were observed in 56.3 % of patients with heartburn [9]. Similarly, we found that 61.3 % (19/31) of NAFLD patients with GERD had insomnia. A study in 134 Japanese patients with GERD found that FSSG score was significantly positively correlated with PSQI score [11]. GERD can affect sleep through two primary mechanisms. First, nighttime reflux, which occurs in 47-79 % of patients with GERD, can cause awakening during the night. Second, GERD can cause short, amnestic arousals (approximately 30 s), resulting in sleep fragmentation. However, recent studies also suggested that the link between GERD and sleep problems may be bidirectional. Sleep stage may influence the esophago-upper esophageal sphincter contractile reflex [44]. Sleep disturbance may reinforce the perception of intraesophageal acid [45]. The association of NAFLD with GERD has been assessed in only two studies, which reported that 37 and 51 % of patients with NAFLD had GERD symptoms [7, 8], percentages higher than observed in the present study. Plausible explanations of a lower prevalence (25 %) of GERD in our NAFLD patients were the difference of ethics, sex/age distribution, and the diagnostic method of GERD or NAFLD between previous studies [7, 8] and ours. Another explanation is the possibility that our NAFLD patients receiving dietary

educations might avoid irregular diet habits, which are known to be the most significant risk factors for GERD symptoms [15]. In the future, sex- and age-matched controlled studies using a larger population is essential to draw conclusions. A recent study of Japanese patients with NAFLD found that GERD symptoms were significantly more severe in the group with higher than lower total cholesterol (T-CHO) and triglyceride (TG) levels [7]. In contrast, we observed no correlation between GERD symptoms and either T-CHO or TG. These conflicting results may be due to our inclusion of only patients with biopsy-proven NAFLD, who are not representative of the general population of patients diagnosed with NAFLD. In contrast, the patients included in the previous study were diagnosed with NAFLD by ultrasound [7]. Moreover, that study did not assess the association of insomnia with GERD symptoms. Thus, to our knowledge, our study is the first to clarify the relationship between insomnia and GERD symptoms in patients with NAFLD.

Previous studies suggested that acid suppression can improve sleep problems in GERD patients. For example, a prospective randomized clinical trial found that a significantly higher percentage of patients treated with esomeprazole than placebo showed resolution of GERD-related sleep disturbances [46]. RPZ treatment also significantly improved subjective indices of sleep quality over placebo [16]. Moreover, an 8-week course of RPZ treatment significantly decreased both FSSG and PSQI scores in Japanese patients [11]. Consistent with these findings, we found that treatment with RPZ significantly decreased both AIS and FSSG scores. These results also indicate that GERD symptoms are at least partly responsible for the occurrence of insomnia in NAFLD patients.

In addition to AIS score, GGT concentration was found to be an independent predictor of insomnia in patients with NAFLD. Serum GGT activity is a marker of oxidative stress [47]. The primary function of GGT is to maintain intracellular concentrations of glutathione, a critical antioxidant molecule. Thus, increased GGT activity can be regarded as a response to oxidative stress, aimed at increasing the intracellular concentration of glutathione. Although little is known about the relationship between insomnia and oxidative stress, a preliminary study showed that anti-oxidant activity was significantly lower and lipid peroxidation levels significantly higher in patients with primary insomnia than in controls [48]. Serum concentrations of GGT and 8-hydroxydeoxyguanosine are correlated in patients with NAFLD, indicating the occurrence of oxidative stress [49]. GGT has also been associated with the occurrence of metabolic syndrome, early atherosclerosis, and cardiovascular events [50]. Moreover, GGT concentrations are higher in patients with obstructive sleep



apnea syndrome (OSAS) than in controls [51] and have been associated with nocturnal arterial oxygen desaturation. Furthermore, continuous positive airway pressure treatment has been shown to decrease GGT, further suggesting that the increase in GGT is directly associated with OSAS. OSAS has also been associated with the presence and severity of NAFLD [52], suggesting that insomnia in patients with NAFLD is at least partly associated with the occurrence of OSAS. Our finding, that elevated GGT is an independent predictor of insomnia, suggests that these patients also have OSAS, although this was not directly evaluated in our patient population.

This study had several limitations. First, its design was cross-sectional, making it difficult to establish a cause–effect relationship, and suggesting the need for prospective studies. Second, the AIS is a self-administered questionnaire and subjective measure of insomnia, suggesting the need for assessment of objective sleep variables such as those obtained during polysomnography. Third, GERD was diagnosed based on FSSG scores alone, without endoscopic examination or 24 h pH monitoring. Because all participants were Japanese, there is a possibility that our results may not be applicable for NAFLD patients of other races or ethnic groups. The current study also did not assess the effects of smoking habits, mental status, dietary habit, and work environments. Additional studies, in larger populations, are needed to assess these variables.

In conclusion, we found that about 30 % of Japanese patients with biopsy-proven NAFLD have insomnia. GERD symptoms may be important in the development of insomnia. PPIs may be clinically useful for treating insomnia in NAFLD patients.

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References

- 1. Angulo P. Nonalcoholic fatty liver disease. N Engl J Med. 2002;346:1221–31.
- Ludwig J, Viggiano TR, McGill DB, Oh BJ. Non-alcoholic steatohepatitis. Mayo Clinic experiences with a hitherto unnamed disease. Mayo Clin Proc. 1980;55:434–8.
- David K, Kowdley KV, Unalp A, Kanwal F, Brunt EM, Schwimmer JB. Quality of life in adults with nonalcoholic fatty liver disease: baseline data from the nonalcoholic steatohepatitis clinical research network. Hepatology. 2009;49:1904–12.
- Kim CW, Yun KE, Jung HS, Chang Y, Choi ES, Kwon MJ, et al. Sleep duration and quality in relation to non-alcoholic fatty liver disease in middle-aged workers and their spouses. J Hepatol. 2013;59(2):351–7.
- Hsieh SD, Muto T, Murase T, Tsuji H, Arase Y. Association of short sleep duration with obesity, diabetes, fatty liver and behavioral factors in Japanese men. Intern Med. 2011;50:2499– 502.

- Dew MA, Hoch CC, Buysse DJ, Monk TH, Begley AE, Houck PR, et al. Healthy older adults' sleep predicts all-cause mortality at 4 to 19 years of follow-up. Psychosom Med. 2003;65:63–73.
- 7. Fujikawa Y, Tominaga K, Fujii H, Machida H, Okazaki H, Yamagami H, et al. High prevalence of gastroesophageal reflux symptoms in patients with non-alcoholic fatty liver disease associated with serum levels of triglyceride and cholesterol but not simple visceral obesity. Digestion. 2012;86:228–37.
- 8. Miele L, Cammarota G, Vero V, Racco S, Cefalo C, Marrone G, et al. Non-alcoholic fatty liver disease is associated with high prevalence of gastro-oesophageal reflux symptoms. Dig Liver Dis. 2012;44:1032–6.
- Kusano M, Kouzu T, Kawano T, Ohara S. Nationwide epidemiological study on gastroesophageal reflux disease and sleep disorders in the Japanese population. J Gastroenterol. 2008;43: 833–41
- Jansson C, Nordenstedt H, Wallander MA, Johansson S, Johnsen R, Hveem K, Lagergren J. A population-based study showing an association between gastroesophageal reflux disease and sleep problems. Clin Gastroenterol Hepatol. 2009;7:960–5.
- 11. Fujiwara Y, Kohata Y, Kaji M, Nebiki H, Yamasaki T, Sasaki E, et al. Sleep dysfunction in Japanese patients with gastroesophageal reflux disease: prevalence, risk factors, and efficacy of rabeprazole. Digestion. 2010;81:135–41.
- Johnson DA. Gastroesophageal reflux disease and sleep disorders: a wake-up call for physicians and their patients. Rev Gastroenterol Disord. 2005;5(Suppl 2):S3–11.
- 13. Mody R, Bolge SC, Kannan H, Fass R. Effects of gastroesophageal reflux disease on sleep and outcomes. Clin Gastroenterol Hepatol. 2009;7:953–9.
- Orr WC. Review article: sleep-related gastro-oesophageal reflux as a distinct clinical entity. Aliment Pharmacol Ther. 2010;31: 47–56.
- Yamamichi N, Mochizuki S, Asada-Hirayama I, Mikami-Matsuda R, Shimamoto T, Konno-Shimizu M, et al. Lifestyle factors affecting gastroesophageal reflux disease symptoms: a crosssectional study of healthy 19864 adults using FSSG scores. BMC Med. 2012;10:45.
- Orr WC, Goodrich S, Robert J. The effect of acid suppression on sleep patterns and sleep-related gastro-oesophageal reflux. Aliment Pharmacol Ther. 2005;21:103–8.
- 17. Fujiwara Y, Kohata Y, Kaji M, Nebiki H, Yamasaki T, Sasaki E, et al. Sleep dysfunction in Japanese patients with gastroesophageal reflux disease: prevalence, risk factors, and efficacy of rabeprazole. Digestion. 2010;81:135–41.
- 18. Examination Committee of Criteria for 'Obesity Disease' in Japan; Japan Society for the Study of Obesity. New criteria for 'obesity disease' in Japan. Circ J. 2002;66:987–92.
- Committee on the Classification and Diagnostic Criteria of Diabetes Mellitus. Report of the Committee on the Classification and Diagnostic Criteria of Diabetes Mellitus. J Jpn diabetes Soc. 2010;53:450–67.
- Ogihara T, Kikuchi K, Matsuoka H, Fujita T, Higaki J, Horiuchi M, et al. The Japanese Society of Hypertension Guidelines for the Management of Hypertension (JSH 2009). Hypertens Res. 2009;32:3–107.
- Kusano M, Shimoyama Y, Sugimoto S, Kawamura O, Maeda M, Minashi K, Kuribayashi S, Higuchi T, Zai H, Ino K, Horikoshi T, Sugiyama T, Toki M, Ohwada T, Mori M. Development and evaluation of FSSG: frequency scale for the symptoms of GERD. J Gastroenterol. 2004;39:888–91.
- Yasaka S, Murakami K, Abe T, Anan J, Mizukami K, Tanahashi J, Okimoto T, Kodama M, Kudo Y, Kawasaki H, Fujioka T. Evaluation of esophageal function in patients with gastroesophageal reflux disease using transnasal endoscopy. J Gastroenterol Hepatol. 2009;24:1677–82.



- 23. Miyamoto M, Haruma K, Takeuchi K, Kuwabara M. Frequency scale for symptoms of gastroesophageal reflux disease predicts the need for addition of prokinetics to proton pump inhibitor therapy. J Gastroenterol Hepatol. 2008;23:746–51.
- 24. Furuta T, Shimatani T, Sugimoto M, Ishihara S, Fujiwara Y, Kusano M, et al. Investigation of pretreatment prediction of proton pump inhibitor (PPI)-resistant patients with gastroesophageal reflux disease and the dose escalation challenge of PPIs-TORNADO study: a multicenter prospective study by the Acid-Related Symptom Research Group in Japan. J Gastroenterol. 2011;46:1273–83.
- Sakamoto Y, Inamori M, Iwasaki T, Lida H, Endo H, Hosono K, et al. Relationship between upper gastrointestinal symptoms and diet therapy: examination using frequency scale for the symptoms of gastroesophageal reflux disease. Hepatogastroenterology. 2010;57:1635–8.
- Soldatos CR, Dikeos DG, Paparrigopoulos TJ. Athens Insomnia Scale: validation of an instrument based on ICD-10 criteria. J Psychosom Res. 2000;48:555–60.
- Soldatos CR, Dikeos DG, Paparrigopoulos TJ. The diagnostic validity of the Athens Insomnia Scale. J Psychosom Res. 2003;55:263–7.
- Matteoni CA, Younossi ZM, Gramlich T, Boparai N, Liu YC, McCullough AJ. Nonalcoholic fatty liver diseases: a spectrum of clinical and pathological severity. Gastroenterology. 1999;116: 1413–9.
- Kleiner DE, Brunt EM, Van Natta M, Behling C, Contos MJ, Cummings OW, Nonalcoholic Steatohepatitis Clinical Research Network, et al. Design and validation of a histological scoring system for nonalcoholic fatty liver disease. Hepatology. 2005;41:1313–21.
- Brunt EM, Janney CG, Di Bisceglie AM, Neuschwander-Tetri BA, Bacon BR. Nonalcoholic steatohepatitis: a proposal for grading and staging the histological lesions. Am J Gastroenterol. 1999;94:2467–74.
- Doi Y, Minowa M, Uchiyama M, Okawa M, Kim K, Shibui K, Kamei Y. Psychometric assessment of subjective sleep quality using the Japanese version of the Pittsburgh Sleep Quality Index (PSQI-J) in psychiatric disordered and control subjects. Psychiatry Res. 2000;97:165–72.
- 32. Kim K, Uchiyama M, Okawa M, Liu X, Ogihara R. An epidemiological study of insomnia among the Japanese general population. Sleep. 2000;23:41–7.
- Yoshioka E, Saijo Y, Kita T, Satoh H, Kawaharada M, Kishi R. Effect of the interaction between employment level and psychosocial work environment on insomnia in male Japanese public service workers. Int J Behav Med. 2012. doi:10.1007/s12529-012-9230-9.
- Yoshioka E, Saijo Y, Kita T, Satoh H, Kawaharada M, Fukui T, Kishi R. Gender differences in insomnia and the role of paid work and family responsibilities. Soc Psychiatry Psychiatr Epidemiol. 2012;47:651–62.
- 35. Monterrosa-Castro A, Marrugo-Flórez M, Romero-Pérez I, Chedraui P, Fernández-Alonso AM, Pérez-López FR. Prevalence of insomnia and related factors in a large mid-aged female Colombian sample. Maturitas. 2013;74:346–51.
- 36. Blümel JE, Cano A, Mezones-Holguín E, Barón G, Bencosme A, Benítez Z, et al. A multinational study of sleep disorders during female mid-life. Maturitas. 2012;72:359–66.
- Yoshioka E, Saijo Y, Fukui T, Kawaharada M, Kishi R. Association between duration of daily visual display terminal work

- and insomnia among local government clerks in Japan. Am J Ind Med. 2008;51:148-56.
- 38. Utsugi M, Saijo Y, Yoshioka E, Horikawa N, Sato T, Gong Y, Kishi R. Relationships of occupational stress to insomnia and short sleep in Japanese workers. Sleep. 2005;28:728–35.
- Nishitani N, Sakakibara H. Job stress factors, stress response, and social support in association with insomnia of Japanese male workers. Ind Health. 2010;48:178–84.
- Kozaki T, Miura N, Takahashi M, Yasukouchi A. Effect of reduced illumination on insomnia in office workers. J Occup Health. 2012;54:331–5.
- 41. Spiegel K, Tasali E, Penev P, Van Cauter E. Brief communication: sleep curtailment in healthy young men is associated with decreased leptin levels, elevated ghrelin levels, and increased hunger and appetite. Ann Intern Med. 2004;141:846–50.
- Spiegel K, Leproult R, L'hermite-Baleriaux M, Copinschi G, Penev PD, Van Cauter E. Leptin levels are dependent on sleep duration: relationships with sympathovagal balance, carbohydrate regulation, cortisol, and thyrotropin. J Clin Endocrinol Metab. 2004;89:5762–71.
- 43. Donga E, Van Dijk M, Van Dijk JG, Biermasz NR, Lammers GJ, van Kralingen KW, et al. A single night of partial sleep deprivation induces insulin resistance in multiple metabolic pathways. J Clin Endocrinol. 2010;95:2963–8.
- 44. Bajaj JS, Bajaj S, Dua KS, Jaradeh S, Rittmann T, Hofmann C, Shaker R. Influence of sleep stages on esophago-upper esophageal sphincter contractile reflex and secondary esophageal peristalsis. Gastroenterology. 2006;130:17–25.
- Schey R, Dickman R, Parthasarathy S, Quan SF, Wendel C, Merchant J, et al. Sleep deprivation is hyperalgesic in patients with gastroesophageal reflux disease. Gastroenterology. 2007;133:1787–95.
- 46. Johnson DA, Orr WC, Crawley JA, Traxler B, McCullough J, Brown KA, Roth T. Effect of esomeprazole on nighttime heartburn and sleep quality in patients with GERD: a randomized, placebo-controlled trial. Am J Gastroenterol. 2005;100:1914–22.
- 47. Lee DH, Jacobs DR Jr. Serum gamma-glutamyltransferase: new insights about an old enzyme. J Epidemiol Community Health. 2009;63:884–6.
- 48. Gulec M, Ozkol H, Selvi Y, Tuluce Y, Aydin A, Besiroglu L, Ozdemir PG. Oxidative stress in patients with primary insomnia. Prog Neuropsychopharmacol Biol Psychiatry. 2012;37:247–51.
- 49. Irie M, Sohda T, Iwata K, Kunimoto H, Fukunaga A, Kuno S, et al. Levels of the oxidative stress marker γ-glutamyltranspeptidase at different stages of nonalcoholic fatty liver disease. J Int Med Res. 2012;40:924–33.
- Kozakova M, Palombo C, Eng MP, Dekker J, Flyvbjerg A, Mitrakou A, et al. Fatty liver index, gamma-glutamyltransferase, and early carotid plaques. Hepatology. 2012;55:1406–15.
- 51. Barceló A, Barbé F, de la Peña M, Vila M, Pérez G, Piérola J, et al. Antioxidant status in patients with sleep apnoea and impact of continuous positive airway pressure treatment. Eur Respir J. 2006;27:756-60.
- 52. Musso G, Cassader M, Olivetti C, Rosina F, Carbone G, Gambino R. Association of obstructive sleep apnoea with the presence and severity of non-alcoholic fatty liver disease. A systematic review and meta-analysis. Obes Rev. 2013;14:417–31.

