Pulmonary atresia (PA)	***	-	- 0		- +	H -		-	-	-	-	-		NA -	***	**	-	-		***	-	-	-			-		-	-	-		-	-	100	-	-	-		-	-			(600)		2%	% (1/49)
Positional anomaly of valsalva sinus	-	-		-		-	-	-	-	-	-	-	- 1	NA -	-	_	_	-	~			-	-	-	-	-		-	-	-	-	-	-	-	-	-	-	+	-			-	_	-	27	/ <sub>*</sub> (1/49)
Functional abnormalities																																														
Pulmonary hypertension (PH)	***	-	-	-	~ *		200	***	-	-	six	-		NA -		***	946	+	-			-	-		-	-	-	+	-	100		90.	-	+	-	-	-	100	-	100		-	-		8%	% (4/49)
Left ventricular noncompaction (LVNC)	-	-		-	ener i	-: -	-	-	-	-					+	-		-	-	-	+	-	100	+	-	-	-			-	-	-	+	-	-	-	-	900	-	- 4	+ -		-	_	10	7% (5/49)
Dilated cardiomyopathy	***	-	-	-		-	***	+	-	+	***	-	- 1	NA -	-	$\rightarrow$	-	100	$\sim$	-		-	***	-	-	-		-	-	-	-	-	-		-	-	-	-	-	-			-	- 100	49	% (2/49)
5. Sensory organs																																														
Ears																																														
Hearing problems	-	+		-		+	-	-	-	+	-	_	- ,		-	+	+	-	-	+		_	+	+	-	-	+ -	_	NA	+	- 4	+	+	-	-	+	+	+	+	+ .			+	-	39	9% (19/49)
Eyes																																														
Strabismus			+	- 1		- +	N/	A -	-	+	-	+	-		+	+	-		+	-	+	-	-	-	-	+	NA -	4-	NA	+		+	_	-	+	+	-	NA	_	. 9	+ -		-	+	33	3% (15/46)
Ametropia	-		+ -	-			N/	١ -	-	+			+		+	+	+		+		٠.	+	-	-		-	NA -	-	NA			-	-	-	+	-	-	NA	-		+ -	+				5% (12/46)
Nystagmus	-	-				-	N/	+	-	-	-	+			-	_	_	_	-	+	-	_	-	_	+	-	NA -	_	NA	+	-	_	_	-	+	-		NA	+		- +				-	0% (9/46)
Oculomotor disturbance	-	-		_		-	N/	A -	-	-	-	-	-	-	-		-	-	-	~	-	-	-	-	_	_	NA -	_	NA	_			-	_	_	-	_	NA	_		+ -		_			% (1/46)
Colohoma	_	-					N/	٠ -	-	-	-	_	_		-	_	_	-	-			-	_	_	-			_	NA						_	_			_	_						% (1/46)
Congenital microphthalmia	-	-					N/	١ -	-	-		-			-	-		-	-	-		-	-	~	_		NA -		NA			_						NA					+			% (1/46)
Retinitis pigmentosa		-	-			-	N/	١ -	-	-		-			-	-	-		-				+	-	_		NA -	-	NA		_								***							% (1/46)
6. Urogenital abnormalities																																						1.74								0 (1) 40]
Renal abnormalities																																														
Renal hypoplasia	_	-	NA -	_			IA N		_			_	_	NA -			_	_		_			-		NA			NA	_	NA															-	
Hydronephrosis	_		NA -				IA N		-	_	-			NA -	_	-	-	-	-	_		-	+	_		_			-		+ .	-	-	-	-	-	-	_	_	-		-	-	_		% (1/43)
Pyelectasis	_				_		IA N	-	-	-				NA -			_		-			-	_	-						NA	+ .	-	-	-			-	-	-	-	-			_		% (2/43)
Nephronophthisis	_		NA .				IA N		( = )					NA -		-	-		20				+	_	NA			NA NA		NA NA	-	-	-	-			-			-	-	-	-	-		% (1/43)
Extragenital abnormalities			INA			- 1	175									-	-	- 1	-			-	Τ.		NA	7	-	NA	-	NA	-	-	-		-	-	-		-	-	-		-	94	25	% (1/43)
Ectopic kidney	_		NA				A N					_	_	NA -		-	_	_	-	-		_	+	-	***			-	_	NA	_															
Ambiguous external genitalia							A N		-	-				NA -			_	-				_		_	NA					NA NA		-	-	-	-	-	-	-	+		-			-		74 (1/44)
Cryptorchidism	NA	_	NA I						NIA	NIA																	NA NA								-	-				+			IA -			% (3/43)
Scrotal hypoplasia	NA		NA I																															A +	+	NA			NA		NA N			NA N		4% (9/14)
7. Endocrinology and nutrition	1374		INA I	IVA	INA I	NA P	NA 141	I INM	INA	NA	INA	MA	-	NA -	N	A N	INA	-	NA	NA	- 18	A N		NA	NA	NA	NA NA		-	NA	+	N	A N	A -	-	NA	NA	-	NA	+	NA P	NA P	IA N	AN	A 14	4% (2/14)
Hypothyroidism	-		NA -				NA -																					_	-							/										
Obesity	_	_	NA .	-	NIA .			-	+	+	-	+	-		-	-	-	NA	-	-	-	-	-	_	-	_	+ -	-	-	NA	+ -	-	+	+	NA NA		-	-	+	-		- 1	A -	-		6% (7/45)
	-	-	INA	-	INA .		- +	-	T	т		т .		-		-	-	NA	+	-				-	-	-	-			-	-	-	-	-	NA	-	-	7.7	-				-	-	11	1% (5/46)
8. Other complications																																														
Respiratory tract related																																														
Stenosis of nasal cavity		NA					NA N																					-						A NA										NA N		
Laryngomalacia and/or tracheomalacia		NA																									NA -																			
Adenoidal hypertrophy and/or enlarged tonsil		+					NA +																					-						A NA												
Obstructive sleep apnea	NA	NA	NA	-	NA I	NA N	A +	NA	NA	NA	NA	+	NA	NA N	IA N	A N	A NA	NA	NA	NA	NA N	A N	A NA	NA	NA	NA	NA -	80.	NA	NA	NA 1	NA N	A N	A NA	NA	NA	NA	NA	NA	NA	NA 1	NA N	IA N	IA N	A	
Allergy related																																														
Atopic dermatitis		NA					h N																				NA -							A NA												
Food allergy		NA					NA N																					-						A NA												
Bronchial asthma	NA	+	NA ·	-	NA I	NA N	NA N	A NA	NA	NA	NA	NA	NA	NA N	IA N	A N	A NA	NA	NA	NA	NA N	A N	A NA	NA	NA	NA	NA -	-	NA	NA	NA I	NA N	A N	A NA	NA	NA	NA	NA	NA	NA	NA 1	NA N	NA N	A N	A	
Related to secondary																																														
Precocious puherty		NA																									NA -	-						A NA										NA N		
Breast enlargement	NA	NA	NA	***	NA I	NA N	NA N	A NA	NA	NA	NA	NA	NA	+ 1	NA N	A N	A NA	NA	NA	NA	NA N	A N	A NA	NA	NA	NA	NA -	100	NA	NA	NA I	NA N	A N	A NA	NA	NA	NA	NA	NA	NA	NA I	NA N	NA N	IA N	A	
Others																																														
Atresia of external acoustic foramen		NA																									NA -	-						A NA										+ N		
Dental abnormalities		NA					NA N																											A NA							NA 1			NA N		
Non alcoholic steatohepatitis (NASII)		NA																									NA -							A NA												
Constipation	NA	NA	NA	-	NA I	NA N	NA N.	A NA	NA	NA	NA	NA	NA	NA N	NA N	A N	A NA	NA	NA	NA	NA N	A +	NA	NA	NA	NA	NA -	+	NA	NA	NA I	NA N	A N	A NA	NA	NA	NA	NA	+	NA :	NA I	NA N	NA N	IA N	A	
NA not available: S severe: I	3 bor	derl	ine:	М	m	ode	rate																																							

NA, not available; S, severe; B, borderline; M, moderate.



Fig. 1. Facial features of the patients with variably sized 1p36 deletions. Pt 1 (a; at 14 years of age) shows edematous eyelids rather than deep-set eyes. Pt 3 (b; 6 years), 6 (c; 5 years), and 14 (d; 15 years) share characteristic features, including deep-set eyes, hypotelorism, and pointed chins. Pt 47 (e; 4 years) and 48 (f; 8 years) do not exhibit such characteristic features, with round faces rather than hypotelorism and pointed chins. Pt 50 (g; 3 years) exhibits distinctive features with arched eyebrows and hypertelorism. Written informed consent to publish patient photos was obtained from all the patient families.

involved in chromatin remodeling and gene transcription, regulating the expression of neuronal genes [29]. Thus, *CHD5* also may be a modifier gene for severe ID.

It has been suggested that two genes, gamma-aminobutyric acid (GABA) A receptor delta (GABRD; chr1: 1,950,768–1,962,192), and KCNAB2 (chr1: 6,105, 981–6,161,253), are associated with the manifestations of epilepsy [27]. This is also been suggested by our present study, as there was no history of epilepsy in a patient (Pt 2) with a 1.8 Mb terminal deletion and a patient (Pt 50) with a 10.0 Mb interstitial deletion; both of the deletions includes neither GABRD nor KCNAB2 (Fig. 2). The incidence of epilepsy was higher in the patients with severe ID (30/38; 79%) than in the patients with moderate ID (4/8; 50%). Thus, the severity of ID was associated with the incidence of epilepsy and the same gene/set of genes may be involved in both of these neurological manifestations.

Several case reports have suggested an association between periventricular nodular heterotopia (PVNH) and 1p36 deletion [16,30–32], and the candidate region for polymicrogyria has been mapped to the distal 4.8 Mb region [33]. As the smallest deletion among the patients with abnormal neuronal migration was 3.0 Mb (Pt 8), the gene(s) responsible for this phenotype may be narrowed down to the distal 3.0 Mb region (Fig. 2; region D). Chiari malformation type II was identified only in Pt 34, who showed an unbalanced translocation with chromosome 4. Thus, this rare feature may be attributable to the partial trisomy of chromosome 4.

#### 4.4. Cardiac abnormality

Previously, the genetic region responsible for left ventricular noncompaction (LVNC) was assigned to the 1.9-3.4 Mb region [34-36]. On the other hand, there are many reports which show an association between Ebstein anomaly and 1p36 deletion [7,37–40]. The genomic region responsible for Ebstein anomaly was assigned to the 2.9-3.8 Mb region [39,40]. In 2005, Sinkovec et al. reported two patients with LVNC associated with Ebstein anomaly [41]. In this study, we identified a patient (Pt 24) who showed both LVNC and Ebstein anomalies. Given this perspective, it might be reasonable to conclude that the critical regions involved in LVNC and Ebstein anomaly are relatively close. As mentioned above PRDM16 located on chr1: 2,985,742-3,355,185 was reported as a gene responsible for cardiomyopathy and LVNC [28]. This is in agreement with our study, as the smallest deletion identified in a patient (Pt 9) with DCM was 3.1 Mb in size. It is possible that PRDM16 may also be related not only to LVNC but also to the Ebstein anomaly.

Although double-outlet right ventricle (DORV) has never been reported in individuals with 1p36 deletions, we found DORV in two patients. We found a relatively small deletion (2.5 Mb) in a patient (Pt 6) with DORV (Fig. 2; region D). There is a possibility that the protein kinase C zeta gene (*PRKCZ*; chr1 chr1: 1,981,909–2,116,834) is related to cardiac abnormalities, because this gene had been implicated in a variety of process including cardiac muscle function [42,43]. The positional

S. Shimada et al. | Brain & Development xxx (2014) xxx-xxx

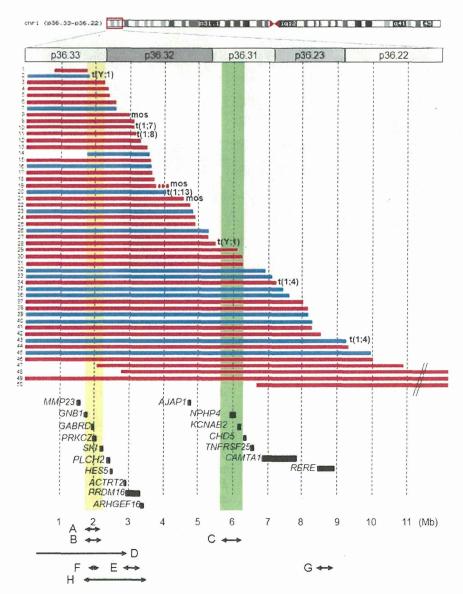


Fig. 2. Result of chromosomal microarray testing depicted in a genome map of the 1p36 region. The scheme of chromosome 1 (top) is downloaded from the UCSC genome browser. Red and blue bars indicate the deletion regions identified in female and male patients, respectively. Black bars indicate the locations of the genes, discussed in the text. The numbers depicted on the left side of each bar indicate patients' numbering. "t" and "mos" indicate unbalanced translocations and mosaicism, respectively. Yellow and green translucent vertical lines emphasize the proposed responsible regions for ID. Proposed responsible regions for each phenotype; A, distinctive craniofacial findings; B, ID; C, modifier effect for ID; D, LVNC and Ebstein anomaly; E, DORV; F, cardiac anomalies; G, cryptochidisms. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

effects for *PRDM16* may be another possibility in this case.

The arginine-glutamic acid dipeptide (RE) repeats gene (RERE; chr1: 8,412,464–8,877,699) has been reported to play a critical role in early cardiovascular development [44]. In this study, all patients with deletions larger than 8.4 Mb, which involve RERE, showed cardiac anomalies. Thus, RERE may be involved in the pathogenesis of congenital heart defects (Fig. 2; region G).

Only Pt 20, with an unbalanced translocation between 13q32.3, showed hypoplasia of the left ventricle (HLHS) in this study. HLHS accounts for 2–3% of all congenital heart defects, and a minority of HLHS cases have been associated with congenital anomaly syndromes, e.g., the Jacobsen, Turner, and Potocki–Lupski syndromes, respectively [45–47]. As 13q duplication has been reported to be associated with this manifestation, the findings of HLHS found in Pt 20 may be due to a partial trisomy of 13q [48].

Please cite this article in press as: Shimada S et al. Microarray analysis of 50 patients reveals the critical chromosomal regions responsible for 1p36 deletion syndrome-related complications. Brain Dev (2014), http://dx.doi.org/10.1016/j.braindev.2014.08.002

# 4.5. Other complications

In patients with 1p36 monosomy, a Prader-Willi syndrome (PWS)-like phenotype has been described [6,13,49]. The clinical features that overlap between the 1p36 deletion syndrome and PWS are ID, neonatal hypotonia, obesity, craniofacial anomalies, hyperphagia, short stature, and behavior problems. D'Angelo et al. described a patient with a 2.5 Mb deletion within the chromosome region 1p36.33-1p36.32 [13]. Tsuyusaki et al. hypothesized that the critical region for the PWS-like phenotype was within 4 Mb from 1pter [49]. Rosenfeld et al. suggested a critical region for the PWS-like phenotype in the 1.7-2.3 Mb region [12]. In this study, all five patients with obesity (Pt 8, 10, 11, 13, and 21) were female, and acquired ambulatory ability within the ages of 2–8 years. Two of the patients (Pt 8 and 21) showed mosaic deletions [17]. From these perspectives, we speculate that female patients who showed 1p36 deletions involving the critical region and who acquired ambulatory ability are likely to be at risk for obesity.

#### 5. Conclusion

In this study, we successfully accumulated the genotype-phenotype data of 50 patients with the deletions of 1p36 regions. As hypotelorism was commonly observed in patients, it may be characteristic of Asian patients. The genotype-phenotype correlation analysis narrowed down the regions responsible for distinctive craniofacial features and ID to the 1.8-2.1 and 1.8-2.2 Mb regions, respectively. Patients with deletions larger than 6.2 Mb showed no ambulation, indicating that severe neurodevelopmental prognosis may be modified by haploinsufficiencies of KCNAB2 and/or CHD5, located 6.2 Mb away from the telomere. Although the genotype-phenotype correlation for the cardiac abnormalities is unclear, PRDM16, PRKCZ, and RERE may be related to this complication. One more finding revealed by this study for the first time, is that female patients who acquired ambulatory ability are likely to be at a risk for obesity.

# Acknowledgments

We would like to express our gratitude to the patients and their families for their cooperation. We also thank the Japanese Society of Child Neurology and the Japan Society of Pediatric Genetics for their supports. This work was supported by a Grant-in-Aid for Scientific Research from Health Labor Sciences Research Grants from the Ministry of Health, Labor, and Welfare, Japan.

#### References

- Shapira SK, McCaskill C, Northrup H. Spikes AS. Elder FFB, Sutton VR, et al. Chromosome 1p36 deletions: the clinical phenotype and molecular characterization of a common newly delineated syndrome. Am J Hum Genet 1997;61:642–50.
- [2] Gajecka M, Yu W. Ballif BC, Glotzbach CD, Bailey KA, Shaw CA, et al. Delineation of mechanisms and regions of dosage imbalance in complex rearrangements of 1p36 leads to a putative gene for regulation of cranial suture closure. Eur J Hum Genet 2005;13:139–49.
- [3] Heilstedt HA, Ballif BC, Howard LA, Lewis RA, Stal S, Kashork CD, et al. Physical map of 1p36, placement of breakpoints in monosomy 1p36, and clinical characterization of the syndrome. Am J Hum Genet 2003;72:1200–12.
- [4] Shao L, Shaw CA, Lu XY, Sahoo T, Bacino CA, Lalani SR, et al. Identification of chromosome abnormalities in subtelomeric regions by microarray analysis: a study of 5,380 cases. Am J Med Genet A 2008;146A:2242–51.
- [5] Heilstedt HA, Ballif BC, Howard LA, Kashork CD, Shaffer LG. Population data suggest that deletions of 1p36 are a relatively common chromosome abnormality. Clin Genet 2003:64:310–6.
- [6] Slavotinek A, Shaffer LG, Shapira SK, Monosomy 1p36, J Med Genet 1999;36:657-63.
- [7] Battaglia A, Hoyme HE, Dallapiccola B, Zackai E, Hudgins L, McDonald-McGinn D, et al. Further delineation of deletion 1p36 syndrome in 60 patients: a recognizable phenotype and common cause of developmental delay and mental retardation. Pediatrics 2008;121:404–10.
- [8] Battaglia A. Del 1p36 syndrome: a newly emerging clinical entity. Brain Dev 2005;27:358-61.
- [9] Zenker M. Rittinger O. Grosse KP, Speicher MR, Kraus J, Rauch A, et al. Monosomy 1p36-a recently delineated, clinically recognizable syndrome. Clin Dysmorphol 2002;11:43–8.
- [10] Kurosawa K, Kawame H, Okamoto N, Ochiai Y, Akatsuka A, Kobayashi M, et al. Epilepsy and neurological findings in 11 individuals with 1p36 deletion syndrome. Brain Dev 2005;27:378–82.
- [11] Buck A, du Souich C, Boerkoel CF. Minimal genotype-phenotype correlation for small deletions within distal 1p36. Am J Med Genet A 2011;155A:3164-9.
- [12] Rosenfeld JA, Crolla JA, Tomkins S, Bader P, Morrow B. Gorski J, et al. Refinement of causative genes in monosomy 1p36 through clinical and molecular cytogenetic characterization of small interstitial deletions. Am J Med Genet A 2010;152A:1951–9.
- [13] D'Angelo CS, Da Paz JA, Kim CA, Bertola DR, Castro CIE, Varela MC, et al. Prader-Willi-like phenotype: investigation of 1p36 deletion in 41 patients with delayed psychomotor development, hypotonia, obesity and/or hyperphagia, learning disabilities and behavioral problems. Eur J Med Genet 2006;49:451-60.
- [14] Okamoto N, Toribe Y, Nakajima T, Okinaga T, Kurosawa K, Nonaka I, et al. A girl with 1p36 deletion syndrome and congenital fiber type disproportion myopathy. J Hum Genet 2002;47:556-9.
- [15] Hiraki Y, Fujita H, Yamamori S, Ohashi H, Eguchi M, Harada N, et al. Mild craniosynostosis with 1p36.3 trisomy and 1p36.3 deletion syndrome caused by familial translocation t(Y:1). Am J Med Genet A 2006;140A:1773-7.
- [16] Saito Y. Kubota M. Kurosawa K, Ichihashi I, Kaneko Y, Hattori A, et al. Polymicrogyria and infantile spasms in a patient with 1p36 deletion syndrome. Brain Dev 2011;33:437–41.
- [17] Shimada S. Maegaki Y. Osawa M. Yamamoto T. Mild developmental delay and obesity in two patients with mosaic 1p36 deletion syndrome. Am J Med Genet A 2014;164A:415-20.
- [18] Shimada S, Okamoto N, Hirasawa K, Yoshii K, Tani Y, Sugawara M, et al, Clinical manifestations of Xq28 functional

- disomy involving MECP2 in one female and two male patients. Am J Med Genet A 2013;161A:1779–85.
- [19] Shimada S, Okamoto N, Ito M, Arai Y, Momosaki K, Togawa M, et al. MECP2 duplication syndrome in both genders. Brain Dev 2013;35:411-9.
- [20] Kang SH, Scheffer A, Ou Z, Li J, Scaglia F, Belmont J, et al. Identification of proximal 1p36 deletions using array-CGH: a possible new syndrome. Clin Genet 2007;72:329–38.
- [21] Shimojima K, Paez MT, Kurosawa K, Yamamoto T. Proximal interstitial 1p36 deletion syndrome: the most proximal 3.5-Mb microdeletion identified on a dysmorphic and mentally retarded patient with inv(3)(p14.1q26.2). Brain Dev 2009;31:629–33.
- [22] Benzing T. Schermer B. Clinical spectrum and pathogenesis of nephronophthisis. Curr Opin Nephrol Hypertens 2012;21:272–8.
- [23] Otto EA, Ramaswami G, Janssen S, Chaki M, Allen SJ. Zhou W, et al. Mutation analysis of 18 nephronophthisis associated ciliopathy disease genes using a DNA pooling and next generation sequencing strategy. J Med Genet 2011;48:105–16.
- [24] Gajecka M. Mackay K.L. Shaffer L.G. Monosomy 1p36 deletion syndrome. Am J Med Genet C 2007;145C:346–56.
- [25] Wu YQ, Heilstedt HA, Bêdell JA, May KM, Starkey DE, McPherson JD, et al. Molecular refinement of the 1p36 deletion syndrome reveals size diversity and a preponderance of maternally derived deletions. Hum Mol Genet 1999;8:313–21.
- [26] Colmenares C, Heilstedt HA, Shaffer LG, Schwartz S, Berk M, Murray JC, et al. Loss of the SKI proto-oncogene in individuals affected with 1p36 deletion syndrome is predicted by straindependent defects in Ski mice. Nat Genet 2002;30:106–9.
- [27] Heilstedt HA, Burgess DL, Anderson AE, Chedrawi A, Tharp B, Lee O, et al. Loss of the potassium channel beta-subunit gene. KCNAB2, is associated with epilepsy in patients with 1p36 deletion syndrome. Epilepsia 2001;42:1103-11.
- [28] Arndt AK, Schafer S, Drenckhahn JD, Sabeh MK, Plovie ER, Caliebe A, et al. Fine mapping of the 1p36 deletion syndrome identifies mutation of *PRDM16* as a cause of cardiomyopathy. Am J Hum Genet 2013;93:67–77.
- [29] Potts RC, Zhang P, Wurster AL, Precht P, Mughal MR, Wood III WH, et al. CHD5, a brain-specific paralog of Mi2 chromatin remodeling enzymes, regulates expression of neuronal genes. PLoS One 2011;6:e24515.
- [30] Neal J. Apse K, Sahin M, Walsh CA, Sheen VL. Deletion of chromosome 1p36 is associated with periventricular nodular heterotopia. Am J Med Genet A 2006;140:1692–5.
- [31] Saito S. Kawamura R, Kosho T, Shimizu T, Aoyama K, Koike K, et al. Bilateral perisylvian polymicrogyria, periventricular nodular heterotopia, and left ventricular noncompaction in a girl with 10.5–11.1 Mb terminal deletion of 1p36. Am J Med Genet A 2008;146A;2891–7.
- [32] Descartes M, Mikhail FM, Franklin JC, McGrath TM, Bebin M. Monosomy1p36.3 and trisomy 19p13.3 in a child with periventricular nodular heterotopia. Pediatr Neurol 2011;45:274–8.
- [33] Dobyns WB, Mirzaa G, Christian SL, Petras K, Roseberry J, Clark GD, et al. Consistent chromosome abnormalities identify novel polymicrogyria loci in 1p36.3. 2p16.1-p23.1, 4q21.21-q22.1, 6q26-q27, and 21q2. Am J Med Genet A 2008;146A:1637-54.

- [34] Thienpont B, Mertens L, Buyse G, Vermeesch JR, Devriendt K, Left-ventricular non-compaction in a patient with monosomy 1p36. Eur J Med Genet 2007;50:233–6.
- [35] Cremer K, Ludecke HJ, Ruhr F, Wieczorek D. Left-yentricular non-compaction (LVNC): a clinical feature more often observed in terminal deletion 1p36 than previously expected. Eur J Med Genet 2008;51:685–8.
- [36] Gajecka M. Saitta SC, Gentles AJ, Campbell L, Ciprero K, Geiger E, et al. Recurrent interstitial 1p36 deletions: evidence for germline mosaicism and complex rearrangement breakpoints. Am J Med Genet A 2010;152A;3074–83.
- [37] Faivre L, Morichon-Delvallez N, Viot G, Martinovic J, Pinson MP, Aubry JP, et al. Prenatal detection of a 1p36 deletion in a fetus with multiple malformations and a review of the literature. Prenat Diagn 1999;19:49–53.
- [38] Riegel M, Castellan C, Balmer D, Brecevic L, Schinzel A. Terminal deletion, del(1)(p36.3), detected through screening for terminal deletions in patients with unclassified malformation syndromes. Am J Med Genet 1999;82:249–53.
- [39] Redon R, Rio M, Gregory SG, Cooper RA, Fiegler H, Sanlaville D, et al. Tiling path resolution mapping of constitutional 1p36 deletions by array-CGH: contiguous gene deletion or "deletion with positional effect" syndrome? J Med Genet 2005;42:166-71.
- [40] Digilio MC, Bernardini L, Lepri F, Giuffrida MG, Guida V. Baban A, et al. Ebstein anomaly: genetic heterogeneity and association with microdeletions 1p36 and 8p23.1. Am J Med Genet A 2011;155A:2196–202.
- [41] Sinkovec M, Kozelj M, Podnar T. Familial biventricular myocardial noncompaction associated with Ebstein's malformation. Int J Cardiol 2005;102:297–302.
- [42] Sentex E, Wang X, Liu X, Lukas A, Dhalla NS. Expression of protein kinase C isoforms in cardiac hypertrophy and heart failure due to volume overload. Can J Physiol Pharmacol 2006;84:227–38.
- [43] Wu SC, Solaro RJ. Protein kinase C zeta. A novel regulator of both phosphorylation and de-phosphorylation of cardiac sarcomeric proteins. J Biol Chem 2007;282:30691–8.
- [44] Kim BJ, Zaveri HP, Shchelochkov OA, Yu Z, Hernandez-Garcia A, Seymour ML, et al. An allelic series of mice reveals a role for RERE in the development of multiple organs affected in chromosome 1p36 deletions. PLoS One 2013;8:e57460.
- [45] Barron DJ, Kilby MD, Davies B, Wright JG, Jones TJ, Brawn WJ. Hypoplastic left heart syndrome. Lancet 2009;374:551–64.
- [46] Grossfeld PD. The genetics of hypoplastic left heart syndrome. Cardiol Young 1999;9:627–32.
- [47] Sanchez-Valle A, Pierpont ME, Potocki L. The severe end of the spectrum: hypoplastic left heart in Potocki-Lupski syndrome. Am J Med Genet A 2011;155A:363-6.
- [48] Chen CP, Chern SR, Hsu CY, Lee CC, Lee MS, Wang W. Prenatal diagnosis of de novo partial trisomy 13q (13q22 qter) and partial monosomy 8p (8p23,3 pter) associated with holoprosencephaly, premaxillary agenesis, hexadactyly, and a hypoplastic left heart. Prenat Diagn 2005;25:334-6.
- [49] Tsuyusaki Y, Yoshihashi H, Furuya N, Adachi M, Osaka H, Yamamoto K, et al. 1p36 deletion syndrome associated with Prader-Willi-like phenotype. Pediatr Int 2010;52:547–50.



Brain & Development xxx (2014) xxx-xxx



www.elsevier.com/locate/braindev

# Original article

# Clinical and genetic features of acute encephalopathy in children taking theophylline

Makiko Saitoh ", ", Mayu Shinohara ", Atsushi Ishii b, Yukiko Ihara b, Shinichi Hirose b, Masashi Shiomi b, Hisashi Kawawaki d, Masaya Kubota b, Takanori Yamagata f, Akie Miyamoto g, Gaku Yamanaka b, Kaoru Amemiya f, Kenjiro Kikuchi f, Atsushi Kamei k, Manami Akasaka k, Yuki Anzai f, Masashi Mizuguchi b

a Department of Developmental Medical Sciences, Graduate School of Medicine, The University of Tokyo, Japan
b Department of Pediatrics, Fukuoka University, Japan
c Department of Pediatrics, Child Medical Center, Osaka City General Hospital, Japan
d Department of Pediatric Neurology, Child Medical Center, Osaka City General Hospital, Japan
c Department of Neurology, National Center for Child Health and Development, Japan
f Department of Pediatrics, Jichi Medical University, Japan
g Department of Pediatrics, Asahikawa Habilitation Center for Disabled Children, Japan
h Department of Pediatrics, Tokyo Medical University, Japan
f Department of Neurology, Tokyo Metropolitan Hachioji Children's Hospital, Japan
f Division of Neurology, Saitama Children's Medical Center, Japan
k Department of Pediatrics, Iwate Medical University, Japan
Department of Pediatrics, Saiseikai Yokohamashi Tobu Hospital, Japan

Received 21 May 2014; received in revised form 30 July 2014; accepted 30 July 2014

#### Abstract

Background: Theophylline has recently been suspected as a risk factor of acute encephalopathy with biphasic seizures and late reduced diffusion (AESD), although there has been no systematic study on the relationship between acute encephalopathy in children taking theophylline (AET) and AESD.

Methods: We recruited 16 Japanese patients (11 male and 5 female, median age of 2 years and 7 months) with AET from 2008 to 2013. We evaluated their clinical features, such as the duration of first seizure, biphasic clinical course and cranial CT/MRI imaging and compared them with those of AESD. We analyzed the polymorphisms or mutations of genes which are associated with AESD.

Results: Clinically, 12 patients had neurological and/or radiological features of AESD. Only one patient died, whereas all 15 surviving patients were left with motor and/or intellectual deficits. Genetically, 14 patients had at least one of the following polymorphisms or mutations associated with AESD: thermolabile variation of the carnitine palmitoyltransferase 2 (CPT2) gene, polymorphism causing high expression of the adenosine receptor A2A (ADORA2A) gene, and heterozygous missense mutation of the voltage gated sodium channel 1A (SCN1A) and 2A (SCN2A) gene.

http://dx.doi.org/10.1016/j.braindev.2014.07.010

0387-7604/© 2014 The Japanese Society of Child Neurology. Published by Elsevier B.V. All rights reserved.

Please cite this article in press as: Saitoh M et al. Clinical and genetic features of acute encephalopathy in children taking theophylline. Brain Dev (2014), http://dx.doi.org/10.1016/j.braindev.2014.07.010

<sup>\*</sup> Corresponding author. Address: Department of Developmental Medical Sciences, Graduate School of Medicine, The University of Tokyo, 7-3-1 Hongo, Bunkyo, Tokyo 113-0033, Japan. Tel.: +81 3 5841 3615; fax: +81 3 5841 3628.

E-mail address: makisaito-tky@umin.ac.jp (M. Saitoh).

Conclusions: Our results demonstrate that AET overlaps with AESD, and that AET is a multifactorial disorder sharing a genetic background with AESD.

© 2014 The Japanese Society of Child Neurology. Published by Elsevier B.V. All rights reserved.

Keywords: Theophylline; Adenosine receptors; Acute encephalopathy; Status epilepticus

#### 1. Introduction

Theophylline is a methylxanthine that exerts multiple pharmacologic effects by inhibiting phosphodiesterases. Until recently, it has been commonly used in clinical practice for the treatment of bronchial asthma and acute bronchitis, especially in Japan. However, theophylline may trigger seizures in patients with or without epilepsy, even when the concentration is within the therapeutic range [1,2]. The pro-convulsive effects of theophylline are explained by its activity as a non-selective, competitive antagonist of adenosine. In the central nervous system (CNS), adenosine plays a role as an endogenous anticonvulsant [3,4], since the effects of anti-excitatory A1 receptor (ADORA1) predominate over those of pro-excitatory A2A receptor (ADORA2A). Theophylline-associated seizures (TASs) are most prevalent among children under 6 years of age and usually occur during a febrile infectious disease [5]. TASs often persist and resist first-line anticonvulsants, leading to refractory status epilepticus and a poor neurologic outcome [6,7].

When a post-ictal coma lasts for more than 24 h, the condition should be regarded as acute encephalopathy rather than a mere seizure [8]. Acute encephalopathy with inflammation-mediated status epilepticus includes multiple syndromes [9], such as fever-induced refractory epileptic encephalopathy in school-aged children (FIRES) (or its eponym, acute encephalitis with refractory, repetitive partial seizures (AERRPS)), and acute encephalopathy with biphasic seizures and late reduced diffusion (AESD) [10] (or its eponym, acute encephalopathv with febrile convulsive status epilepticus (AEFCSE)) [11]. In a case series in a referral hospital in Japan, many children taking theophylline reportedly had clinical and radiological features of AESD or AEFCSE [12]. Thus, theophylline has recently been suspected as a risk factor of AESD [8], although there has been no systematic study on the relationship between acute encephalopathy in children taking theophylline (AET) and AESD.

In this paper, we recruited Japanese patients with AET by means of a nationwide, multi-institutional study supported by the Japanese Society of Child Neurology. We reviewed their clinical data and examined whether the findings meet the diagnostic criteria of AESD. We also conducted genetic analysis of these patients, focusing on genes that were shown to be

associated with AESD in our previous studies: carnitine palmitoyltransferase 2 (*CPT2*), *ADORA2A*, and voltage-gated sodium channel subunit 1A (*SCN1A*) and 2A (*SCN2A*) [12–15]. The aim of this study was to elucidate the relationship between AET and AESD from both clinical and genetic viewpoints.

#### 2. Methods

#### 2.1. Patients

We defined acute encephalopathy based on the following criteria [16,17]: (1) acute onset of severe and sustained impairment of consciousness after a preceding infection, and (2) exclusion of CNS inflammation. We defined AET as acute encephalopathy with the onset with status epilepticus within several hours after administration of oral theophylline or intravenous aminophylline, and recruited patients with AET from hospitals in Japan during 2008–2012 in a retrospective manner. Sixteen Japanese patients (11 male and 5 female) aged from 6 months to 4 years and 4 months (median, 2 years and 7 months), participated in this study. One case (Case 2) had been reported previously [14]. Their clinical characteristics including the family and past history, preceding infection, serum concentration of theophylline, duration of status epilepticus, presence or absence of biphasic seizures, cranial CT and/or MRI findings, therapy and outcome, were evaluated. The diagnosis of AESD was based on the criteria described previously [16]. It was regarded as 'definite' when both the characteristic clinical course (biphasic seizures) and CT/MRI findings (delayed appearance of cerebral cortical edema, distribution of lesions showing lobar or hemispheric involvement and peri-Rolandic sparing, and restricted diffusion of the subcortical white matter (so-called bright tree appearance) were present [8,10], 'probable' when either clinical or CT/MRI features were present, and 'possible' when prolonged febrile seizures were followed by non-specific CT/MRI findings (diffuse cortical damage) and other diagnostic possibilities were unlikely. In some patients whose CT/MRI findings in the acute/subacute period were either unavailable or insufficient, distribution of lesions was inferred on the basis of those in the convalescence. Other conditions that occasionally show bright tree appearance, such as hemorrhagic shock and encephalopathy syndrome, head

injury and hypoxic-ischemic encephalopathy, were excluded based on the clinical history and laboratory data.

# 2.2. Standard protocol approvals, registrations, and patient consent

The procedures in this study were approved by the University of Tokyo Ethics Committee. Written informed consent was obtained from all guardians of patients participating in the study.

#### 2.3. Procedures

Peripheral blood samples were collected from all 16 patients and from 100 control subjects, namely healthy Japanese volunteers. Genomic DNA was extracted from the blood using standard protocols and was used for the analysis of *CPT2*, *ADORA1*, *ADORA2A*, *SCN1A* and *SCN2A* genes.

# 2.3.1. CPT2

We analyzed exon 4 and 5 of the *CPT2* gene by direct sequencing or real-time polymerase chain reaction (PCR) using the TaqMan Probe and Faststart Universal Probe Master ROX (Roche, Basel, Switzerland), as described previously [12]. In this study, we focused on the F352C genotype. We had previously found that at least one allele C in F352C is associated with AESD and other syndromes of acute encephalopathy [12].

# 2.3.2. ADORA1 and ADORA2A

All coding regions and intron-exon splicing sites of the ADORA1 and ADORA2A genes were PCR amplified with flanking intronic primers under standard PCR conditions. PCR products of ADORA1 and ADORA2A were sequenced on a 310 Genetic Analyzer, 3100 Genetic Analyzer or 3130xl Genetic Analyzer (Life Technologies, Carlsbad, CA, USA). To identify rs5751876 and rs2298383 SNPs of ADORA2A, the PCR-restriction fragment length polymorphism (PCR-RFLP) method was adopted. Based on the combination of four SNPs showing complete linkage disequilibrium Japanese (human HapMap project, http:// Apr2011.archive.ensemble.org), we determined whether the subjects had either haplotype A (C at rs2298383, T at rs5751876, deletion at rs35320474 and C at rs4822492) or haplotype B (T at rs2298383, C at rs5751876, T at rs35320474 and G at rs4822492). We had previously demonstrated that haplotype A is a risk factor for AESD [14].

#### 2.3.3. SCN1A and SCN2A

The entire coding regions of the *SCN1A* and *SCN2A* genes were sequenced on a 310 Genetic Analyzer (Life Technologies) [14,15].

#### 3. Results

# 3.1. Clinical findings

Clinical data were similar among the 16 patients studied (Table 1). Family history and past history were unremarkable, except for the presence of febrile seizures in two cases each. In all the cases, theophylline or aminophylline was administered temporarily for the treatment of acute asthma attacks (2 cases) and acute bronchitis (14 cases). Blood concentration of theophylline was within the therapeutic range (3.9–11.8 μg/ml) in all 5 cases examined. All patients had fever due to acute respiratory infection. The first convulsion, mostly status epilepticus, occurred within 24 h from the onset of fever. Of the 14 patients who had seizures lasting longer than 15 min, seven patients required continuous intravenous infusion of barbiturates for 2-11 days. Two underwent hypothermia. Eleven showed biphasic seizures typical for AESD. Cranial CT or MRI findings during the acute/subacute period were available in 15 cases. Ten had one of the features characteristic of AESD: delayed cerebral edema, lobar or hemispheric involvement, and bright tree appearance (Fig. 1). One of the remaining five showed, during convalescence, cerebral cortical sparing of the peri-Rolandic regions, another feature typical of AESD. Cranial CT/MRI during convalescence showed diffuse atrophy in 11 patients.

# 3.2. Genetic findings

## 3.2.1. CPT2

Eight out of the 16 patients had at least one allele C in F352C (Table 2). The frequency was higher in the patients (8/16, 50%) than in controls (26/100, 26%), although the difference did not reach statistical significance (p = 0.07).

## 3.2.2. ADORA1 and ADORA2A

First, we confirmed the absence of mutations in the entire coding region of *ADORA1* and *ADORA2A* in all the patients. Next, we analyzed genetic variations of *ADORA2A*. The number of homozygous/heterozygous haplotype A (AA/AB diplotype) in patients was 3 and 11, respectively. Only 2 patients had homozygous haplotype B (BB diplotype) (Table 2). The frequency of BB diplotype (2/16, 12.5%) was lower in the TAE patients than in controls (56/184, 30.4%) [13], although the difference did not reach statistical significance.

# 3.2.3. SCN1A

We found in one case (Case 2) a missense mutation, V982L, which was not found in the 100 control subjects. The valine 982 residue is located on transmembrane segment 6, domain II of SCN1A (Na $_{\rm v}$ 1.1) protein, and is highly conserved among vertebrates and among other

Please cite this article in press as: Saitoh M et al. Clinical and genetic features of acute encephalopathy in children taking theophylline. Brain Dev (2014), http://dx.doi.org/10.1016/j.braindev.2014.07.010

Table 1 Clinical characteristics of patients with acute encephalopathy in children taking theophylline (AET).

	Age, sex	History seizure	y of febrile s	Blood concentration of	Initial seizure (duration) / intravenous barbiturate /	Cranial CT/MRI		Diagnosis of AESD	Outcome	
		Past	Family	theophylline	biphasic seizures	Subacute period	Convalescence		Intellectual disabilities	Motor disabilities
1	2ylm, M	+	_	NR	<15 min/-/+	Delayed cerebral . edema	Diffuse cerebral atrophy	Definite	Severe	Severe
2	2y3m, F	-	+	NR	>15 min/+/+	NR (Normal on day 2)	Diffuse cerebral atrophy, CS	Definite	Severe	Severe
3	4y0m, F	-	-	Therapeutic range	>30 min/-/+	Not available	Diffuse cerebral atrophy	Probable	Severe	Severe
4	2y7m, M	-	-	NR	>15 min/+/-	Mild cerebral edema	Diffuse cerebral atrophy	Possible	Severe	Severe
5	2y2m, M	-	-	13.4 μg/ml	>30 min/+/+	Delayed cerebral edema	Diffuse cerebral atrophy	Definite	Severe	Severe
6	1y0m, M	NR	-	NR	<15 min/-/+	Delayed cerebral edema, BTA, CS	Bilateral frontal atrophy	Definite	Moderate	Full recovery
7	3y5m, M	+	_	NR	>30 min/- "/-	Left temporal subcortical edema	Diffuse cerebral atrophy	Probable	Severe	Severe
8	2y4m, F	+	-	NR	>30 min /+/+	Delayed cerebral edema, right parietal dominant	Diffuse cerebral atrophy	Definite	Severe	Mild
9	3y3m, M	-	, -	NR	>15 min/+/-	Delayed cerebral edema, bilateral parietal dominant	Diffuse cerebral atrophy	Probable	Severe	Mild
10	4y0m, F	-	-	5.8 μg/ml	>30 min/-/-	NR (Mild cortical edema on day 2)	Diffuse cerebral	Possible	Severe	Mild
11	lyllm,M	-	+	NR	>15 min/+/+	BTA, left temporal dominant	Left temporal atrophy	Definite	Mild	Full recovery
12	2y7m, F	_	_	3.9 µg/ml	>15 min/+/-	Early cerebral edema	-	Unlikely	Death	ĺ
13	2y6m, F	-	_	NR	>15 min/-/+	Delayed cerebral edema, bilateral frontal dominant	Diffuse cerebral atrophy, bilateral frontal dominant	Definite	Severe	Mild
14	0y6m, M	-	_	NR	>15 min/+/+	Normal	Bilateral hippocampal sclerosis	Possible	Moderate	Full
15	2y10m, M	-	-	5.6 μg/ml	>15 min/-/+	Hemispheric cortical edema	Hemispheric atrophy	Definite	Mild	Mild
16	4y4m, M	NR	-	NR	>15 min/-/+	Delayed cerebral edema	Diffuse cerebral atrophy bilateral frontal dominant	Definite	Severe	Full recovery

NR, not recorded; BTA, bright tree appearance; CS, central sparing.

# Continuous intravenous midazolam administration.

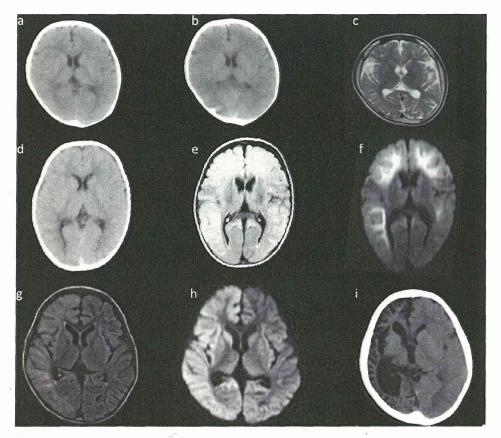


Fig. 1. Cranial CT/MRI findings in acute encephalopathy in children taking theophylline (AET). In Case 1, cranial CT on day 3 showed slight narrowing of the cerebrospinal fluid space, but no clear evidence of cerebral edema (a). On day 7, however, CT showed mild narrowing of the cerebrospinal fluid space and hypodensity of the white matter, indicating delayed cerebral edema (b). Seven years later, MRI (T2-weighted imaging) demonstrated diffuse cerebral atrophy with bilateral subdural effusion (c). In Case 6, CT on day 1 was normal (d). MRI on day 7 showed narrowing of the cerebrospinal fluid space and hyperintensity of the bilateral frontal and temporal cortex on fluid-attenuated inversion recovery (FLAIR) imaging, indicating delayed cerebral edema (e). Diffusion-weighted imaging visualized restricted diffusion in the subcortical white matter (bright tree appearance), with sparing of bilateral peri-Rolandic regions (f). In Case 15, MRI in the subacute period (day 28) showed T1/T2 prolongation of the right cerebral cortex ((g) T1-weighted imaging, (h) FLAIR imaging). Two months later, CT showed atrophy of the right hemisphere (i).

Table 2
Genetic Background of ATE.

Genetic background of ATE.												
Patient No.	CPT2 diplotype	ADORA2A diplotype <sup>b</sup>	SCN1A mutation	SCN2A mutation								
1	FC	AB	No	No								
2	FF	AB	V982L	No								
3	FF	BB	No	No								
4	CC	AB	No	No								
5	FF	AA	No	No								
6	FC	AB	No	No								
7	FF	AA	No	No								
8	FC	AB	No	No								
9	FC	AA	No	No								
10	FC	AB	No	No								
11	FC	AB	No	No								
12	FC	AB	No	No								
13	FF	AB	No	No								
14	FF	AB	No	F328V								
15	FF	BB	No	No								
16	FF	AB	No	No								

<sup>&</sup>lt;sup>a</sup> F352C polymorphism. Allele C is thermolabile variation.

types of sodium channels. This mutation was previously reported in a patient with Dravet syndrome without myoclonic seizures and ataxia [18]. Case 2 with V982L of *SCN1A* had typical AESD ("definite" AESD in this study). The clinical course of this case was reported previously [14].

# 3.2.4. SCN2A

We found in one case (Case 14) a missense mutation, F328V (Fig. 2). The phenylalanine 328 residue is located on the loop between the transmembrane segments 5 and 6, domain I of SCN2A (Na<sub>v</sub>1.2) protein (Fig. 2). The F328V mutation had previously been reported in a patient with Dravet syndrome [19]. Case 14 with F328V of SCN2A was born to a family with no history of epilepsy and seizure disorders. He had no seizures during the neonatal period. At 6 months old, he had acute bronchiolitis and took theophylline for 4 days. He then developed prolonged generalized tonic convulsions with the eyes deviated to the right. Status

<sup>&</sup>lt;sup>b</sup> Combination of four SNPs. Haplotype A is associated with high expression of ADORA2A.