

TABLE 3: Correlation analysis of CSF biomarkers in HHV-6 encephalopathy.

Biomarker		8-OHdG	HEL	Tau	IL-6	IL-10	TNF- α
8-OHdG	Spearman <i>r</i>	1.000	-0.292	-0.239	-0.484	-0.046	-0.315
	<i>P</i>	<0.0001	0.312	0.373	0.068	0.871	0.253
HEL	Spearman <i>r</i>	-0.292	1.000	0.277	0.476	-0.286	0.497
	<i>P</i>	0.312	<0.0001	0.338	0.086	0.322	0.070
Tau	Spearman <i>r</i>	-0.239	0.277	1.000	-0.036	-0.224	0.091
	<i>P</i>	0.373	0.338	<0.0001	0.899	0.422	0.748
IL-6	Spearman <i>r</i>	-0.484	0.476	-0.036	1.000	0.226	0.783
	<i>P</i>	0.068	0.086	0.899	<0.0001	0.418	0.0006
IL-10	Spearman <i>r</i>	-0.046	-0.286	-0.224	0.226	1.000	0.166
	<i>P</i>	0.871	0.322	0.422	0.418	<0.0001	0.555
TNF- α	Spearman <i>r</i>	-0.315	0.497	0.091	0.783	0.166	1.000
	<i>P</i>	0.253	0.070	0.748	0.0006	0.555	<0.0001

CSF: cerebrospinal fluid; HHV: human herpesvirus; 8-OHdG: 8-hydroxy-2'-deoxyguanosine; HEL: hexanoyl-lysine adduct; IL: interleukin; TNF: tumor necrosis factor.

TABLE 4: Clinical profile of patients receiving edaravone treatment.

Patient	Clinical diagnosis	Age/sex	Initiation and dosage of edaravone treatment	Other treatments	Outcomes
1	HHV-6 encephalopathy	14 m/M	Day 5 0.5 mg/kg \times 2/day \times 7 days	Mannitol, Dexamethasone, Ganciclovir, MDL, PHT	Intellectual disability
2	HHV-6 encephalopathy (AESD)	12 m/F	Day 5 0.5 mg/kg \times 2/day \times 8 days	Mannitol, Dexamethasone, Ganciclovir, DZP, MDL	Without sequelae
3	HHV-6 encephalopathy	14 m/M	Day 4 0.5 mg/kg \times 2/day \times 10 days	Mannitol, Dexamethasone, Ganciclovir/acyclovir, DZP, MDL	Hemophagocytic syndrome Died of fulminant hepatitis
4	HHV-6 encephalopathy (AESD)	20 m/F	Day 7 0.5 mg/kg \times 2/day \times 7 days	Mannitol, Dexamethasone, Aciclovir, MDL	Lt hemiparesis
5	HHV-6 encephalopathy (AESD)	12 m/M	Day 3 15 mg/day \times 10 days	DZP, MDL, steroid pulse therapy, mild therapeutic hypothermia	Moderate psychomotor retardation
6	HHV-6 febrile seizures	10 m/F	Day 1 0.5 mg/kg \times 2/day \times 12 days	Mannitol, Dexamethasone, Ganciclovir/acyclovir, DZP, MDL, PHT	Without sequelae

m: months; M: male; F: female; DZP: diazepam; MDL: midazolam; PHT: phenytoin; Lt: left; HHV: human herpesvirus; AESD: acute encephalopathy with biphasic seizures and late reduced diffusion.

in healthy children and they peaked at the second seizures [18]. They speculated that 8-OHdG was produced by ROS from cytokines associated with inflammation and apoptosis following brain edema because changes in urinary 8-OHdG levels reflected the degree of brain edema. However, we found that increased levels of 8-OHdG were observed not only in HHV-6 encephalopathy but also in complex FS associated with HHV-6 infection. We also showed that CSF IL-6 and TNF- α levels were elevated only in the HHV-6 encephalopathy group, but not in the HHV-6 complex FS group. In

addition, we analyzed correlations among biomarkers and observed no significant correlations between increased 8-OHdG levels and cytokine production or increased tau levels. These results suggest that oxidative DNA damage in the brain caused by HHV-6 infection may be independent of inflammatory reactions and subsequent axonal damage.

In contrast with the increased levels of 8-OHdG, there was no significant increase of CSF-HEL levels in HHV-6 encephalopathy compared with HHV-6 complex FS and controls. We previously demonstrated that oxidative stress

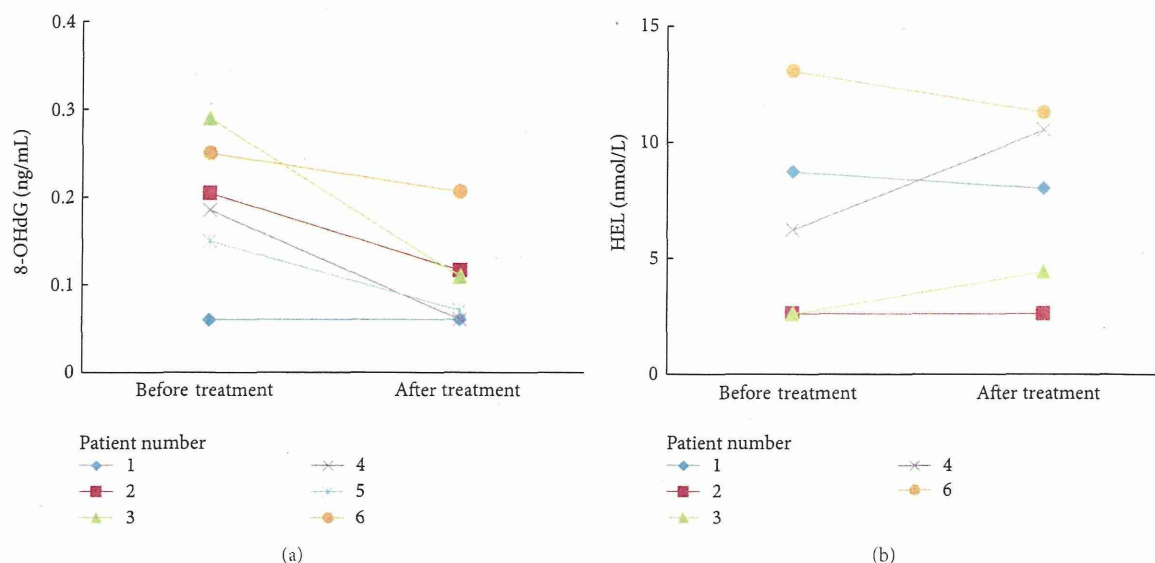


FIGURE 4: Changes in cerebrospinal fluid (CSF)-8-OHdG (a) and CSF-HEL (b) levels before and after edaravone treatment in HHV-6-associated acute encephalopathy and complex febrile seizure (FS) patients. CSF-8-OHdG levels are decreased after edaravone treatment ($P = 0.0202$, paired t -test).

of DNA contributes to early neuronal damage, whereas lipid peroxidation is related to subsequent neurodegeneration in subacute sclerosing panencephalitis [8]. In the present study, we only examined levels of CSF biomarkers in the acute phase of the diseases. Further investigation is required to clarify whether lipid peroxidation may be involved in the chronic phase.

Most patients with HHV-6 encephalopathy in this study (81.3%) were AESD, which has a high incidence of neurological sequelae. We previously indicated that levels of CSF tau protein were elevated in patients with AESD [19]. The current study demonstrates that CSF levels of tau protein were significantly increased at days 3–8 in HHV-6 encephalopathy compared with those of HHV-6 encephalopathy at days 1–2, HHV-6 complex FS, and controls. As CSF tau protein is considered a useful biomarker of axonal damage [20], our results raise the possibility that the high incidence of neurological sequelae in AESD is attributable to axonal injury. However, there was no correlation between the increased levels of tau protein and the presence or absence of neurological sequelae. These findings suggest that tau protein is a sensitive biomarker that might help diagnose HHV-6 encephalopathy, but it is difficult to make an early diagnosis for acute encephalopathy using this biomarker. In terms of the prognostic prediction for HHV-6 encephalopathy, another biomarker will be required because increased levels of tau protein do not always reflect a poor prognosis.

Edaravone is a free radical scavenger that interacts biochemically with a wide range of free radicals [21]. In experimental models, edaravone protects against apoptotic neuronal cell death and improves cerebral function after traumatic brain injury (TBI) [21]. In addition, Ohta et al. reported that administration of edaravone to mice immediately after TBI suppressed traumatic axonal injury and oxidative stress,

which protected against trauma-induced memory deficits [22]. Edaravone is used clinically in Japan for the treatment of acute ischemic stroke. Although childhood ischemic stroke is different than in adults, the use of edaravone was recently approved for the treatment of stroke in children. In the present study, we reported 5 cases of edaravone treatment for HHV-6-associated acute encephalopathy and one case of HHV-6 complex FS. Our study is very preliminary and it is likely that the efficacy of edaravone treatment in combination with other therapies at this time was poor.

There were several limitations in this study. First, the initiation of edaravone treatment was delayed in HHV-6 encephalopathy because it is difficult to distinguish HHV-6 encephalopathy from HHV-6 complex FS during the initial seizures. Early diagnosis of HHV-6 encephalopathy, especially AESD, will be required to overcome this problem. Second, a clinical trial of edaravone for the treatment of acute encephalopathy might be difficult ethically, as placebo control cannot be used because of the severe nature of this disease. We confirmed that CSF-8-OHdG levels decreased after edaravone treatment, although there were no significant differences of mean values of other biomarkers between before and after the treatment. These results suggest edaravone treatment was partially effective for HHV-6 encephalopathy. Although these findings are encouraging, the therapeutic implications of ROS and RNS scavengers are complex, owing to their potential to exert toxic as well as protective effects [23].

5. Conclusion

In summary, we found oxidative DNA damage is involved in acute encephalopathy/febrile seizures associated with HHV-6 infection and may be independent of inflammatory reactions and subsequent axonal damage.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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References

- [1] J. Takanashi, H. Oba, A. J. Barkovich et al., "Diffusion MRI abnormalities after prolonged febrile seizures with encephalopathy," *Neurology*, vol. 66, no. 9, pp. 1304–1309, 2006.
- [2] J.-I. Takanashi, "Two newly proposed infectious encephalitis/encephalopathy syndromes," *Brain and Development*, vol. 31, no. 7, pp. 521–528, 2009.
- [3] A. Hoshino, M. Saitoh, A. Oka et al., "Epidemiology of acute encephalopathy in Japan, with emphasis on the association of viruses and syndromes," *Brain and Development*, vol. 34, no. 5, pp. 337–343, 2012.
- [4] M. Mizuguchi, "Overview of acute encephalitis and encephalopathy," *Nippon Rinsho. Japanese Journal of Clinical Medicine*, vol. 69, no. 3, pp. 391–398, 2011.
- [5] M. Hayashi, "Oxidative stress in developmental brain disorders," *Neuropathology*, vol. 29, no. 1, pp. 1–8, 2009.
- [6] J. Emerit, M. Edeas, and F. Bricaire, "Neurodegenerative diseases and oxidative stress," *Biomedicine and Pharmacotherapy*, vol. 58, no. 1, pp. 39–46, 2004.
- [7] K. Dasuri, L. Zhang, and J. N. Keller, "Oxidative stress, neurodegeneration, and the balance of protein degradation and protein synthesis," *Free Radical Biology and Medicine*, vol. 62, pp. 170–185, 2013.
- [8] M. Hayashi, N. Arai, J. Satoh et al., "Neurodegenerative mechanisms in subacute sclerosing panencephalitis," *Journal of Child Neurology*, vol. 17, no. 10, pp. 725–730, 2002.
- [9] M. Hayashi, M. Itoh, S. Araki et al., "Oxidative stress and disturbed glutamate transport in hereditary nucleotide repair disorders," *Journal of Neuropathology & Experimental Neurology*, vol. 60, no. 4, pp. 350–356, 2001.
- [10] M. Hayashi, S. Araki, J. Kohyama, K. Shioda, and R. Fukatsu, "Oxidative nucleotide damage and superoxide dismutase expression in the brains of xeroderma pigmentosum group A and Cockayne syndrome," *Brain and Development*, vol. 27, no. 1, pp. 34–38, 2005.
- [11] M. Hayashi, S. Araki, N. Arai et al., "Oxidative stress and disturbed glutamate transport in spinal muscular atrophy," *Brain and Development*, vol. 24, no. 8, pp. 770–775, 2002.
- [12] E. Otomo, H. Tohgi, K. Kogure et al., "Effect of a novel free radical scavenger, edaravone (MCI-186), on acute brain infarction: Randomized, placebo-controlled, double-blind study at multicenters," *Cerebrovascular Diseases*, vol. 15, no. 3, pp. 222–229, 2003.
- [13] B. J. Lee, Y. Egi, K. van Leyen, E. H. Lo, and K. Arai, "Edaravone, a free radical scavenger, protects components of the neurovascular unit against oxidative stress in vitro," *Brain Research*, vol. 1307, pp. 22–27, 2010.
- [14] Y. Kitagawa, "Edaravone in acute ischemic stroke," *Internal Medicine*, vol. 45, no. 5, pp. 225–226, 2006.
- [15] Y. Higashi, "Edaravone for the treatment of acute cerebral infarction: role of endothelium-derived nitric oxide and oxidative stress," *Expert Opinion on Pharmacotherapy*, vol. 10, no. 2, pp. 323–331, 2009.
- [16] T. Valyi-Nagy and T. S. Dermody, "Role of oxidative damage in the pathogenesis of viral infections of the nervous system," *Histology and Histopathology*, vol. 20, no. 3, pp. 957–967, 2005.
- [17] T. Akaike, "Role of free radicals in viral pathogenesis and mutation," *Reviews in Medical Virology*, vol. 11, no. 2, pp. 87–101, 2001.
- [18] M. Fukuda, H. Yamauchi, H. Yamamoto et al., "The evaluation of oxidative DNA damage in children with brain damage using 8-hydroxydeoxyguanosine levels," *Brain & Development*, vol. 30, no. 2, pp. 131–136, 2008.
- [19] N. Tanuma, R. Miyata, S. Kumada et al., "The axonal damage marker tau protein in the cerebrospinal fluid is increased in patients with acute encephalopathy with biphasic seizures and late reduced diffusion," *Brain and Development*, vol. 32, no. 6, pp. 435–439, 2010.
- [20] M. Öst, K. Nylén, L. Csajbok et al., "Initial CSF total tau correlates with 1-year outcome in patients with traumatic brain injury," *Neurology*, vol. 67, no. 9, pp. 1600–1604, 2006.
- [21] T. Itoh, T. Satou, S. Nishida et al., "Edaravone protects against apoptotic neuronal cell death and improves cerebral function after traumatic brain injury in rats," *Neurochemical Research*, vol. 35, no. 2, pp. 348–355, 2010.
- [22] M. Ohta, Y. Higashi, T. Yawata et al., "Attenuation of axonal injury and oxidative stress by edaravone protects against cognitive impairments after traumatic brain injury," *Brain Research*, vol. 1490, pp. 184–192, 2013.
- [23] T. Satoh and S. A. Lipton, "Redox regulation of neuronal survival mediated by electrophilic compounds," *Trends in Neurosciences*, vol. 30, no. 1, pp. 37–45, 2007.

Original article

Microarray analysis of 50 patients reveals the critical chromosomal regions responsible for 1p36 deletion syndrome-related complications

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Abstract

Objective: Monosomy 1p36 syndrome is the most commonly observed subtelomeric deletion syndrome. Patients with this syndrome typically have common clinical features, such as intellectual disability, epilepsy, and characteristic craniofacial features.

Method: In cooperation with academic societies, we analyzed the genomic copy number aberrations using chromosomal microarray testing. Finally, the genotype–phenotype correlation among them was examined.

Results: We obtained clinical information of 86 patients who had been diagnosed with chromosomal deletions in the 1p36 region. Among them, blood samples were obtained from 50 patients (15 males and 35 females). The precise deletion regions were successfully genotyped. There were variable deletion patterns: pure terminal deletions in 38 patients (76%), including three cases of mosaicism; unbalanced translocations in seven (14%); and interstitial deletions in five (10%). Craniofacial/skeletal features, neurodevelopmental impairments, and cardiac anomalies were commonly observed in patients, with correlation to deletion sizes.

Conclusion: The genotype–phenotype correlation analysis narrowed the region responsible for distinctive craniofacial features and intellectual disability into 1.8–2.1 and 1.8–2.2 Mb region, respectively. **Patients with deletions larger than 6.2 Mb showed no ambulation, indicating that severe neurodevelopmental prognosis may be modified by haploinsufficiencies of *KCNAB2* and *CHD5*, located at 6.2 Mb away from the telomere. Although the genotype–phenotype correlation for the cardiac abnormalities is unclear, *PRDM16*, *PRKCZ*, and *RERE* may be related to this complication.** Our study also revealed that female patients who acquired ambulatory ability were likely to be at risk for obesity.

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Keywords: 1p36 deletion syndrome; Chromosomal deletion; Genotype–phenotype correlation; Intellectual disability; Ambulation; Epilepsy; Distinctive features

1. Introduction

Monosomy 1p36 syndrome is a congenital malformation syndrome caused by the subtelomeric deletion of the short arm of chromosome 1 [1–3]. This syndrome is the most commonly observed subtelomere deletion syndrome, with an estimated incidence of 1:5000–1:10,000 [4,5]. Patients with this syndrome exhibit common clinical features, including intellectual disability (ID) and characteristic craniofacial features; such as straight eyebrows, deep-set eyes, epicanthus, and a pointed chin [6–9]. Although the levels of ID vary among patients, craniofacial features are commonly seen [10]. The patients with the 1p36 deletion syndrome also show many other complications, including hypotonia, seizures, hearing loss, structural heart defects, cardiomyopathy, ophthalmological abnormalities, and behavior abnormalities [7]. Recent advances in microarray-based chromosomal testing have helped us to identify small chromosomal rearrangements that are invisible by conventional G-banded chromosomal tests/karyotyping [11,12]. Using this method, the precise locations of the aberrations can be revealed at the molecular level. These advances have also allowed the study of more in-depth genotype–phenotype correlations for this syndrome, as

well as the identification of some of the regions responsible for individual complications [12,13].

In this study, we performed a nation-wide survey for the 1p36 deletion syndrome in Japan. The aim of this study was to identify the chromosomal regions responsible for individual complications in patients with 1p36 deletions. We analyzed the affected genomic regions in 50 patients with 1p36 deletions, and performed correlational analyses of the genotype data with the clinical information.

2. Materials and methods

2.1. Patients and samples

We performed a nation-wide survey for the 1p36 deletion syndrome with the cooperation of two academic societies; the Japanese Society of Child Neurology and the Japan Society of Pediatric Genetics. The study subjects were Japanese patients who had already been diagnosed using various diagnostic methods, including conventional karyotyping, subtelomere fluorescence *in situ* hybridization (FISH) analysis, multiplex ligation-dependent probe amplification (MLPA), and chromosomal microarray testing. Five patients

(patient [Pt] 8, 9, 21, 28, and 43), whose clinical features have been previously reported [14–17], were also included in this study. With the questionnaire survey for attending physicians, we accumulated the patients' clinical information, including craniofacial/skeletal features, neurodevelopmental features, brain structural abnormalities, cardiac abnormalities, sensory-organs abnormalities, urogenital abnormalities, endocrinological and nutritional findings among others. This study was approved by the ethics committee in Tokyo Women's Medical University.

On receipt of written informed consents from the families of the patients, we obtained the patients' blood samples to determine genomic copy number losses in the patients. Genomic DNA was extracted from the blood samples using the QIA quick DNA Extraction Kit (QIAGEN, Hamburg, Germany). Metaphase spreads were also prepared from blood samples and used for FISH analyses. In cases, if we could obtain written informed consent, parental samples were also analyzed.

2.2. Molecular and cytogenetic analyses

Chromosomal microarray testing was performed using any of the Agilent Oligo Microarray Kits 44, 60, 105, 180, and 244 K (Agilent Technologies, Santa Clara, CA), as described previously [18,19]. Genomic copy number aberrations were visualized using Agilent Genomic Workbench version 6.5 (Agilent Technologies). For cases in which variations of unknown significance were identified or suspected, parental samples were also analyzed. In cases of complex chromosomal rearrangements or mosaicism, metaphase spreads prepared using the patients' samples were used for FISH analyses for confirmation. The bacterial artificial clones were selected from the UCSC genome browser (<http://genome.ucsc.edu/>) for use as probes. For the target probes, RP11-425E15 (1p36.33: 949,400–1,132,489), RP11-82D16 (1p36.33: 2,046,751–2,208,312), RP11-70N12 (1p36.32: 2,740,703–2,922,551), CTD-3209F18 (1p36.32: 3,530,092–3,769,006), and RP11-933B18 (1p36.31: 5,988,719–6,177,261) were selected, while CTB-167K11 (1q44: 249,250,621–249,250,621) was used as a marker of chromosome 1. All of the genomic regions are described according to the February 2009 human reference sequence (GRCh37/hg19) in this study.

3. Results

3.1. Molecular-cytogenetic findings

We obtained clinical information from 86 patients with chromosomal deletions involving 1p36 regions. Among them, 50 patients (15 males and 35 females) were successfully genotyped. All of the genotypes were summarized in Tables 1 and 2, and 1p36 deletions identified

in the patients were depicted in the genome map (Fig. 2). The minimum and maximum deletion sizes was 0.9 and 12.9 Mb, respectively. Pure terminal deletions were identified in 38 patients (76%). Among them, three patients (Pt 8, 19, and 21) exhibited mosaicism. Pt 8 was first diagnosed with mosaic 1p36 deletion by chromosomal microarray testing, and Pt 21 had been diagnosed with 1p36 deletion using subtelomere FISH analysis; however, mosaicism was not reported at that time [17]. Although the mosaic deletion of 1p36 in Pt 19 had been firstly confirmed by FISH, we could not detect the breakpoint by chromosomal microarray testing due to low frequency (28% mosaic ratio). As the breakpoint was determined to be between two FISH probes (CTD-3209F18 and RP11-933B18), the proximal end of CTD-3209F18 was used as the minimum deletion region in this patient.

Additional aberrations with the sizes over 0.5 Mb were identified in eight patients (Pt 2, 10, 11, 15, 20, 28, 34, and 43) involving chromosomes 4, 7, 8, 13, and Y (Table 2), including a possible benign copy number aberration in Pt 15, which was also observed in the healthy mother. The other seven patients were confirmed to have unbalanced translocations by cytogenetic evaluation (14%), using either G-banding or FISH analysis. Two translocations were diagnosed as de novo, and the others were designated as unknown because of the lack of availability of parental information.

Five patients (Pt 1, 14, 47, 48, and 50) had interstitial deletions (10%) with a deletion size between 0.9 and 10.3 Mb.

3.2. Clinical findings

Clinical information of the 50 patients successfully genotyped is summarized in Table 3. Estimated frequencies of each complication are also included in Table 3. Pt 26 and 49 suddenly died at 24 and 10 months old of age, respectively. Pt 49 probably died due to heart failure but Pt 26 died of an unknown cause (detailed information unavailable).

3.2.1. Craniofacial features

Most of the patients showed craniofacial features, including straight eyebrows (84%), deep-set eyes (93%), broad nasal bridge (97%), low set ears (88%), and a pointed chin (89%). Constellations of these findings make distinctive facial impressions for 1p36 deletion syndrome, observed in Pt 3, 6, and 14 (Fig. 1b–d). This observation suggests that hypotelorism is rather characteristic among these patients. On the other hand, Pt 1 did not show deep-set eyes (Fig. 1a). The craniofacial features of three patients (Pt 47, 48, and 50) did not exhibit hypotelorism (Fig. 2e–g). From the genotypic point of view, these three patients (Pt 47, 48, and 50) would be diagnosed as having the proximal 1p36 deletion syndrome [20,21].

Table 1
The ranges of 1p36 deletions analyzed by chromosomal microarray testing.

Patient number	Age (year)	Gender	Platform (k)	Start	End	Additional aberration	FISH probe	Mosaic ratio ^a (%)	References
1	14	F	180	834,101	1,770,669	Interstitial	RP11-425E15		
2	9	M	44	1	1,820,584	der(1)t(Y;1), idic(Y)			
3	6	F	180	1	2,186,829				
4	1	F	60	1	2,239,497				
5	3	F	44	1	2,281,699				
6	5	F	60	1	2,553,982				
7	2	M	60	1	2,553,982				
8	5	F	44	1	3,044,953	Mosaicism	RP11-82D16	70	Shimada et al. [17] Okamoto et al. [14]
9	13	F	44	1	3,102,718				
10	18	F	44	1	3,102,718	der(1)t(1;7)			
11	17	F	60	1	3,138,565	der(1)t(1;8)			
12	8	F	60	1	3,265,702				
13	11	F	244	1	3,408,152				
14	5	M	60	1,786,789	3,472,907	Interstitial			
15	2	F	180	1	3,564,328				
16	13	M	60	1	3,582,084				
17	4	F	44	1	3,607,275				
18	2	F	60	1	3,660,110				
19	3	F	60	1	3,769,006	Mosaicism	CTD-3209F18	28	
20	3	M	44	1	4,070,842	der(1)t(1;13)			
21	17	F	44	1	4,481,324	Mosaicism	RP11-82D16	77	Shimada et al. [17]
22	2	F	180	1	4,703,581				
23	6	M	60	1	4,779,157				
24	3	F	60	1	4,843,370				
25	6	F	44	1	4,843,718				
26	2	M	60	1	5,252,985				
27	0	F	44	1	5,252,985				
28	25	F	44	1	5,411,803	der(1)t(Y;1)			Hiraki et al. [15]
29	3	F	44	1	6,128,223				
30	3	F	60	1	6,282,562				
31	1	F	60	1	6,282,562				
32	3	M	60	1	6,882,431				
33	7	M	60	1	7,035,075				
34	1	F	60	1	7,187,535	der(1)t(1;4)			
35	10	M	60	1	7,392,688				
36	8	M	60	1	7,581,058				
37	3	F	44	1	8,077,959				
38	2	F	60	1	8,104,671				
39	3	M	44	1	8,104,671				
40	4	M	44	1	8,181,042				
41	5	F	44	1	8,181,042				
42	1	F	60	1	8,427,633				
43	3	M	60	1	9,180,975	der(1)t(1;4)			Saito et al. [16]
44	5	F	60	1	9,251,936				
45	4	M	60	1	9,953,030				

46	2	F	44	1	10,001,011	Interstitial
47	4	F	44	2,080,309	10,869,155	Interstitial
48	8	F	60	2,785,042	12,743,178	Interstitial
49	0	F	44	1	12,917,483	Interstitial
50	22	F	180	6,614,950	16,890,814	Interstitial

^a The genomic position referring build19.

^b The mosaic ratio was confirmed by FISH; F, female; M, male.

3.2.2. Neurological features

Almost all patients showed ID (98%) but a patient (Pt 2) having a deletion in the far distal region of 1p36 showed borderline ID, with an intelligence quotient (IQ) of 80. Therefore, this region could be eliminated from the responsible region for ID (Fig. 2). The smallest deletion, an interstitial deletion between genomic positions 0.8 and 1.8 Mb, was identified in Pt 1 (Fig. 2). In spite of having this smallest deletion, Pt 1 had severe ID, i.e., she was locomotive but aphasic and required support for all activities in her daily life. This was probably a consequence of intractable epilepsy associated with tonic seizures, caused by factors other than the interstitial deletion of this region. The proximal and distal ends of the breakpoints in Pt 3 and 14 defined the shortest region of overlap for ID, spanning the 1.8–2.2 Mb region (Fig. 2; region B). Axial hypotonia (92%) and poor sucking (70%) were also commonly observed. Epilepsy, one of the major complications in 1p36 deletion syndrome, was observed in 70% of the patients. Infantile spasms were observed in 16% of the patients.

In this study, many types of structural brain abnormalities were identified; not only in the cerebral cortex but also in the white matter (Table 3), indicating that there is no major pattern. The most frequently observed abnormality was a nonspecific finding with enlargement of lateral ventricles.

3.2.3. Cardiac abnormality

Cardiac abnormality is one of the most frequently observed complications in patients with 1p36 deletions. In this study, congenital heart defects and functional abnormalities were observed in 69% (34/49) and 22% (11/49) of the patients, respectively. The most frequently observed patterns were patent ductus arteriosus (PDA; 37% [18/49]) and ventricular septal defects (VSD; 37% [18/49]).

3.2.4. Other complications

Many kinds of complications were observed in many organs. Cryptorchidism was the most frequently observed complication in male patients (64% [9/14]). As Pt 14, with a small interstitial deletion spanning from 1.8 to 3.5 Mb, had cryptorchidism, the deleted region was likely involved in abnormalities of the external genitalia (Fig. 2; region H). Hearing problems (39% [19/49]) and strabismus (33% [15/46]) were relatively common among the patients. Obesity was observed in 5 patients (11% [5/46]).

Renal abnormalities were rare and identified only in three patients. Among them, Pt 26, who had a 5.3 Mb deletion, was diagnosed with the autosomal recessive cystic kidney disease of nephronophthisis (this patient died at 2 years of age) [22]. One of the genes responsible for this condition, the nephronophthisis 4 gene

Table 2
Additional aberrations identified in the patients.

Patient number	Chr	Start ^a	End ^a	Remark	Attribute	Origin
2	Y		59,373,566	der(1)t(Y;1)(p36.3;q12), idic(Y)(q12)	dup	NA
10	7		6,870,943	der(1)t(1;7)(p36.32;p22.1)	dup	NA
11	8		3,909,039	der(1)t(1;8)(p36.22;p23.2)	dup	NA
15	1	146,324,068	149,192,104	del(1)(q21.1;q21.2)	del	Common with mother
20	13	100,462,233	115,169,878	der(1)t(1;13)(p26.32;q32.3)	dup	De novo
28	Y	26,435,039	59,373,566	der(1)t(Y;1)(q12;p36.32) [#]	dup	NA
34	4		13,396,747	der(1)t(1;4)(p36.31;p15.33)	dup	De novo
43	4	189,012,426	191,154,276	der(1)t(1;4)(p36.31;q35.2)	dup	NA

^a The genomic position referring build19; dup, duplication; del, deletion; NA, not available.

[#] This case was previously reported by Hiraki et al. [15].

(*NPHP4*), is located on 1p36 (chr1: 5,946,555–5,965,543) [23], proximal to the deletion region of three patients with renal abnormalities (Pt 26, 33, and 35). It is unclear whether there is a correlation between *NPHP4* and the renal abnormalities observed in this study.

4. Discussion

4.1. Previous genetic studies on the 1p36 deletion syndrome

Many cohort studies have been performed to delineate the phenotypic features of patients with 1p36 deletion syndrome and to evaluate the frequency of complications [1,6,7]. It has been reported that there is no correlation between the deletion size and the number of observed clinical features [24], while the critical region responsible for core phenotypic features, including clefting, hypothyroidism, cardiomyopathy, hearing loss, large fontanel, and hypotonia, has been narrowed down to a region 2.2 Mb from the telomere [3]. Compared to such core phenotypic features, other complications tend to vary with the size of the deletion, and study subjects with larger deletions tend to have more phenotypic features [25], suggesting that the various phenotypic features are dependent on genes involved in the deletion regions. Thus, precise knowledge of the genotype–phenotype correlations could potentially lead to more personalized treatments for individuals with 1p36 deletions and might identify mutations for single gene disorders [3]. The potassium voltage-gated channel, shaker-related subfamily, beta member 2 gene (*KCNAB2*) and the v-ski sarcoma viral oncogene homolog gene (*SKI*) were identified as candidate genes for the epilepsy phenotype and clefting abnormalities, respectively [26,27]. More recently, the PR domain containing 16 gene (*PRDM16*) was identified as a possible candidate gene for cardiomyopathy, as *PRDM16* was included in a minimal deletion among patients with 1p36 deletions associated with cardiomyopathy, while in patients with pure cardiomyopathy, single nucleotide variants

of *PRDM16* were identified as the cause of cardiomyopathy [28]. This was one of the most successful studies of genotype–phenotype correlation in patients with 1p36 deletions [28].

4.2. Craniofacial features

As mentioned above, a region 2.2 Mb from the telomere has been reported to be responsible for core phenotypic features of 1p36 deletion syndrome [3]. Compared to this, we observed atypical facial features in four patients (Pt 1, 47, 48, and 50) whose deletions did not include the 1.8–2.1 Mb region, in this study. Thus, the region responsible for typical facial features is narrowed into this region (Fig. 2; region A). Because hypotelorism has never been listed in the clinical delineations of 1p36 deletion syndrome reported from Western countries, we did not include this finding in the questionnaire survey and the frequency of this finding in Japanese patients could not be calculated. However, it is commonly observed in Japanese patients with typical 1p36 deletion syndrome. Therefore, hypotelorism may be a characteristic finding among Asian patients.

4.3. Neurological features

Although more severe ID was reported to be associated with larger 1p36 deletions [10], the genomic region responsible for severe ID has never been identified. In this study, a patient (Pt 28) having a 5.4 Mb deletion acquired independent gait, while patients with >6.1 Mb deletions had not yet acquired independent gait, and exhibited severe ID. Thus, the region between 5.4 and 6.1 Mb would appear to be the borderline for independent gait (Fig. 2; region C), and the modifier genes for prognosis of development may be located in the region proximal to this borderline. *KCNAB2*, mentioned above, may be one of the modifier genes responsible for severe ID. Chromodomain helicase DNA-binding protein 5 (*CHD5*; chr1: 6,161,847–6,240,194), which encodes a neuron-specific protein, is

Table 3
Summary of clinical features of the patients with 1p36 deletions.

Patient number	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44	45	46	47	48	49	50	Frequencies	
1. Craniofacial and skeletal features																																																				
Characteristic craniofacial features																																																				
Microcephaly	NA	+	NA	+	+	NA	NA	-	+	NA	NA	-	+	NA	NA	+	+	NA	NA	+	+	NA	NA	NA	NA	NA	NA	NA	+	+	NA	-	-	+	NA	NA	NA	+	NA	+	NA	-	NA	NA	+	+	+	+	83% (20/24)			
Brachycephaly	+	NA	-	+	+	NA	NA	-	+	NA	+	NA	NA	-	NA	NA	NA	+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	-	-	+	NA	NA	NA	NA	NA	NA	NA	+	+	+	+	+	+	65% (11/17)				
Straight eyebrow	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	84% (32/38)				
Deep-set eyes	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	93% (37/40)				
Epicanthus	+	NA	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	86% (30/35)					
Broad nasal root/bridge	+	NA	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	97% (32/33)					
Long philtrum	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	63% (22/35)					
Low set ears	-	-	-	NA	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	88% (29/33)					
Pointed chin	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	89% (34/38)					
Late closure of the anterior fontanel	-	NA	+	-	-	-	NA	NA	+	NA	NA	+	NA	+	NA	+	NA	+	NA	+	NA	+	NA	+	NA	+	NA	+	NA	+	NA	+	NA	+	NA	+	NA	+	NA	+	NA	+	NA	+	NA	+	NA	56% (18/32)				
Cleft palate problem																																																				
High palate	-	-	-	-	-	-	+	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	29% (14/49)				
Cleft palate	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	22% (11/49)					
Cleft lip	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	16% (8/49)					
Cleft jaw	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	2% (1/50)					
Limb abnormalities																																																				
Finger abnormalities	-	-	-	NA	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	15% (7/48)				
Limb deformity	-	-	-	NA	-	-	-	-	-	-	-	NA	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	4% (2/48)					
Other skeletal abnormalities																																																				
Rib abnormalities such as 11 ribs	-	-	-	NA	-	-	-	-	-	-	-	NA	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	4% (12/48)				
Scoliosis	-	-	-	NA	-	-	-	-	-	-	-	NA	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	25% (12/48)					
Congenital dislocated hip	-	-	-	NA	-	-	-	-	-	-	-	NA	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	4% (2/48)					
Developmental dysplasia of the hip	-	-	-	NA	-	-	-	-	-	-	-	NA	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	2% (1/48)					
2. Neurological features (clinical)																																																				
Axial hypotonia																																																				
Poor sucking	+	+	NA	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	92% (44/48)		
Difficulty of swallowing	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	70% (32/46)			
Developmental delay	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	56% (27/48)					
Developmental delay																																																				
Intellectual disability	S	B	S	S	M	S	S	M	S	S	S	S	S	M	S	S	M	S	S	M	S	M	S	S	M	S	S	M	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	98% (49/50)			
Acquire independent gait	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	32% (16/50)				
Expressive language																																																				
Sentence	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	6% (3/50)				
Only words	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	16% (8/50)				
Dysarthria	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	8% (4/50)				
Gestures	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	10% (5/50)				
Behavior disorders																																																				
Self-injury	-	+	+	-	NA	+	-	NA	+	+	NA	+	NA	-	NA	-	NA	+	-	-	NA	-	NA	-	NA	-	NA	-	NA	+	+	-	-	NA	-	-	NA	-	-	-	-	-	-	-	-	-	-	-	30% (11/37)			
Temper tantrum	+	-	-	-	NA	+	+	NA	+	+	NA	+	NA	-	NA	-	NA	+	-	-	NA	-	NA	-	NA	-	NA	-	NA	+	-	-	NA	-	-	NA	-	-	-	-	-	-	-	-	-	-	-	-	30% (11/37)			
Poor social interaction	+	-	+	-	NA	+	+	NA	+	+	NA	+	NA	-	NA	-	NA	+	-	-	NA	-	NA	-	NA	-	NA	-	NA	+	-	-	NA	-	-	NA	-	-	-	-	-	-	-	-	-	-	-	-	27% (10/37)			
Epilepsy																																																				
History of epilepsy	+	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	70% (35/50)				
Infantile spasms	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	16% (8/50)					
3. Neurological features (radiological)																																																				
Cerebral cortex																																																				
Periventricular nodular heterotopia (PVNI)	-	-	NA	-	-	-	+	-	NA	-	-	+	-	NA	NA	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	10% (4/42)				
Polymicrogyria	-	-	NA	-	-	-	-	-	NA	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	7% (3/42)				
Cortical dysplasia	-	-	NA																																																	