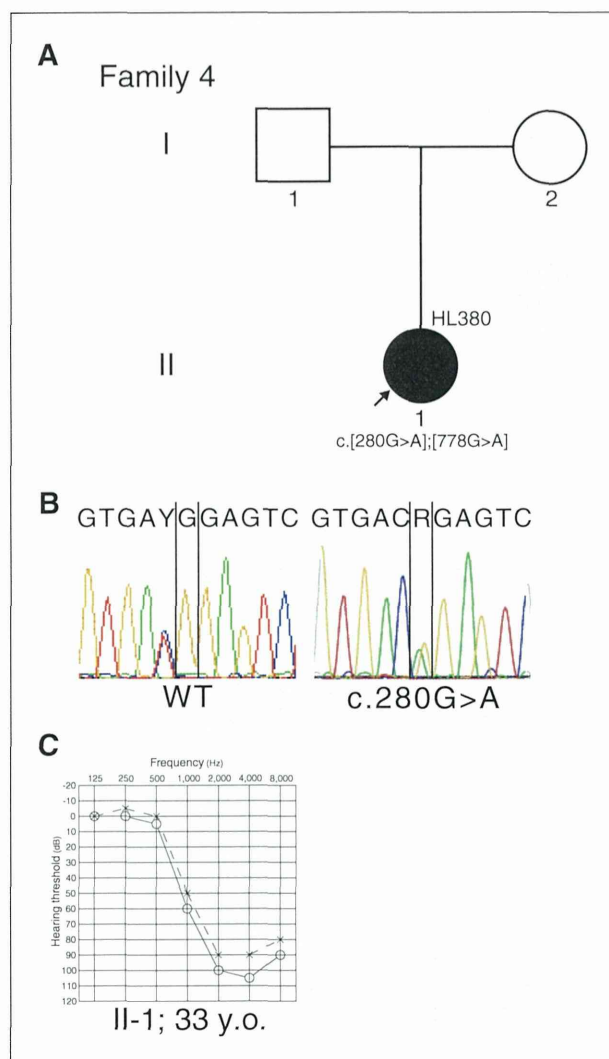


**Figure 3.** (A) The patient (SNS5134) shows compound heterozygous *TMPRSS3* mutations, c.[212T>C];[617-4\_-3dupAT] (p.[F71S];[T205fs]). (B) The results of Sanger sequencing. (C) Pre- and postoperative audiograms indicating good hearing preservation after EAS. (D) Japanese monosyllable test (65 dB SPL in quiet) after bilateral EAS showing a dramatic improvement. EAS, electric acoustic stimulation.



**Figure 4.** (A) The patient (HL0380) shows compound heterozygous *TMPRSS3* mutations, c.[280G>A];[778G>A] (p.[G94R];[A260T]). (B) The results of Sanger sequencing. (C) Audiogram at 33 years old.

sensorineural hearing loss with tinnitus. There were large variations in age of onset, although the hearing loss in patient 4541 and her brother (4540) developed in early childhood.

**Family 1 (Figure 1: SNS5355).** Patient SNS5355 (48-year-old male) had compound heterozygous mutation, c.[390C>G];[647G>T](p.[H130R];[R216L]). The patient noticed he could not hear his electric alarm at 33 years old due to high-frequency progressive hearing loss, and he started to use a hearing aid at 39 years old. Due to the inconvenience associated with using a hearing aid, he received EAS (MEDEL PULSAR FLEX<sup>24</sup>) at 45 years old. His residual

hearing in low frequencies was completely preserved, and his discrimination score was improved after EAS.

**Family 2 (Figure 2: 4540, 4541).** Patient 4541 (41-year-old female) was identified with a compound heterozygous mutation, c.[226C>T];[778G>A](p.[Q76X];[A260T]). The mutation and brief clinical features have been reported previously.<sup>10,31</sup> Her hearing loss was first detected by mass screening in primary school. It appeared to slowly progress, and by age 25, she suffered some inconvenience in hearing and communication. Progressive, ski slope type hearing loss was noted (Figure 2C). The threshold level for 1000 Hz was preserved at 17 years old but thereafter decreased rapidly until 36 years old. The average rate of progression for 1000 Hz was 4.5 dB/year. EAS (MEDEL PULSAR FLEX<sup>24</sup>) was applied at the ages of 38 and 39 bilaterally. Residual hearing for acoustic amplification was preserved, and the hearing level with bilateral EAS was around 30 dB. The patients showed a dramatic improvement in scores for the Japanese monosyllable test (65 dB SPL in quiet) after bilateral EAS, improving from 18% to 90% 1 year after receiving the second EAS.

The same compound heterozygous mutation, c.[226C>T];[778G>A](p.[Q76X];[A260T]), was identified in her brother (patient 4540), who had experienced postlingual hearing loss from 10 years old. His hearing loss was progressive, and he experienced profound hearing loss at 46 years old.

**Family 3 (Figure 3: SNS5134).** Patient SNS5134 (54-year-old female) had a compound heterozygous mutation, c.[212T>C];[617-3\_-4dupAT](p.[F71S];[T205fs]). The patient's age at onset was 30; however, due to rapid progression of the hearing loss, she experienced some inconvenience in hearing and communication by 44 years old. She did not suffer any associated vertigo but did complain of tinnitus. She showed a ski slope type audiogram and received EAS (MEDEL PULSAR FLEX<sup>24</sup>) when she was 51 years old. Her word discrimination score on the Japanese monosyllable test improved after receiving EAS.

**Family 4 (Figure 4: HL0380).** Patient HL0380 was found to have a compound heterozygous mutation, c.[280G>A];[778G>A](p.[G94R];[A260T]). She had noticed the onset of hearing loss, particularly involving high frequencies, when she was 15 years old. She showed ski slope type audiograms at age 33 when she visited an ENT clinic. As she cannot obtain sufficient amplification by use of a hearing aid, she is planning to have genetic counseling, including recommended intervention such as EAS.

Three patients with *TMPRSS3* mutations (SNS5355, 4541, SNS5134) showed normal vestibular function, as evaluated by caloric test and cervical vestibular evoked myogenic potentials (cVEMP) (Figure 5). In addition, no symptoms except hearing loss were confirmed.

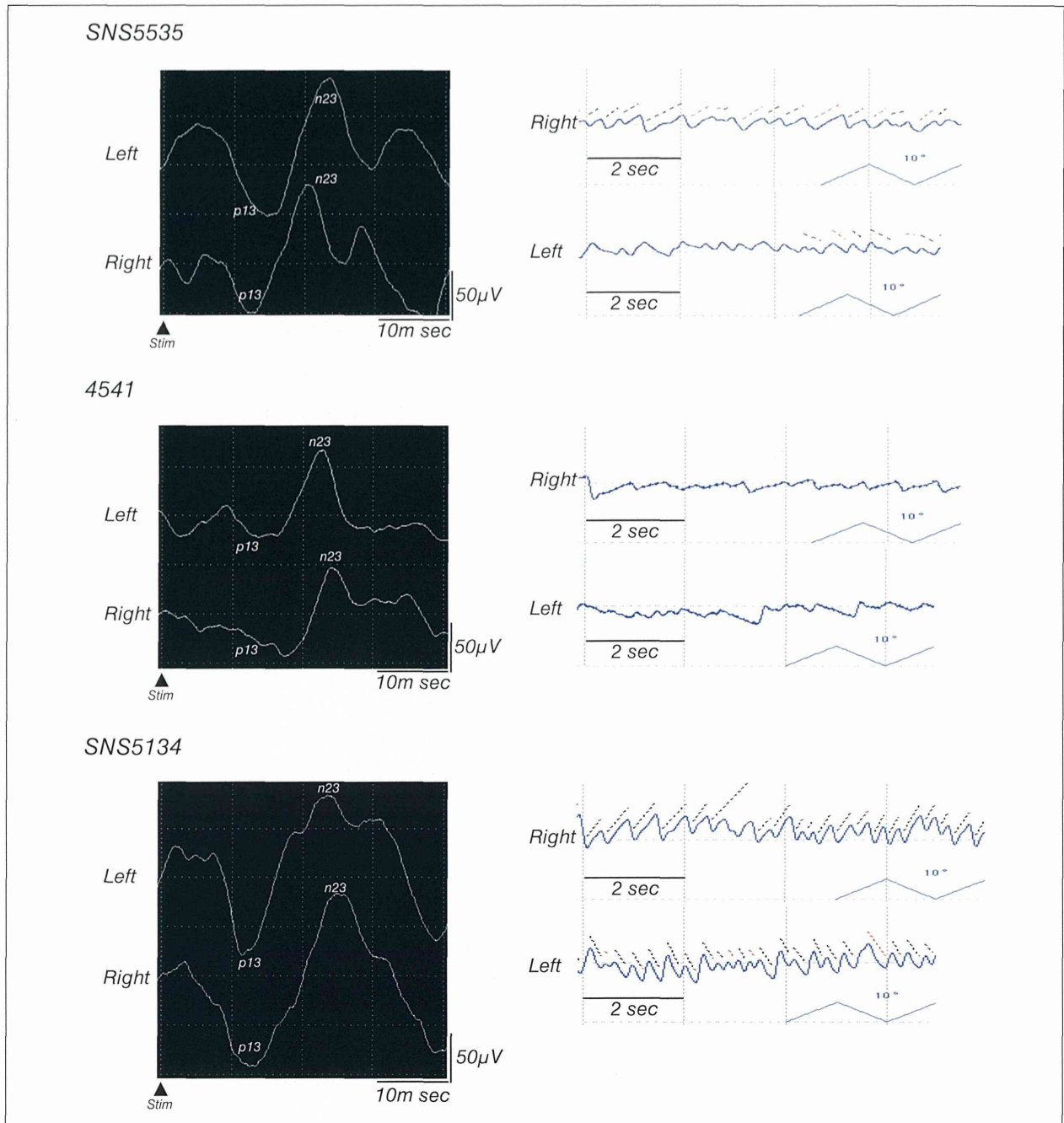
**Table 2.** Clinical Features of 5 Patients With Hearing Loss Caused by *TMPRSS3* Mutations.

Family No.	Patient ID	Nucleotide Change	Amino Acid Change	Age	Onset Age	Intervention	Age at Time of Surgery	Hearing Level (dB) <sup>a</sup>	Hearing Level at Low Frequencies (dB) <sup>b</sup>	Progressiveness	Tinnitus	Vertigo	Caloric Test	cVEMP
1	SNS5355	c.[390C>G];[647G>T]	p.[H130R];[R216L]	48	33	EAS	45	93.8	33.3	+	+	-	Normal	Normal
2	4541	c.[226C>T];[778G>A]	p.[Q76X];[A260T]	41	6	EAS	38/39	106.3	81.7	+	+	-	Normal	Normal
2	4540	c.[226C>T];[778G>A]	p.[Q76X];[A260T]	46	10	None		105	86.7	+	+	-	N/A	N/A
3	SNS5134	c.[212T>C];[617-3_4dupAT]	p.[F71S];[T205fs]	54	30	EAS	51	78.8	38.3	+	+	-	Normal	Normal
4	HL0380	c.[280G>A];[778G>A]	p.[G94R];[A260T]	33	15	None		57.5	-1.7	+	+	-	N/A	N/A

Abbreviations: EAS, electric acoustic stimulation; cVEMP, cervical vestibular evoked myogenic potential; +, existing symptoms; -, without symptoms.

<sup>a</sup>Average of 500, 1000, 2000, and 4000 Hz.

<sup>b</sup>Average of 125, 250, and 500 Hz.

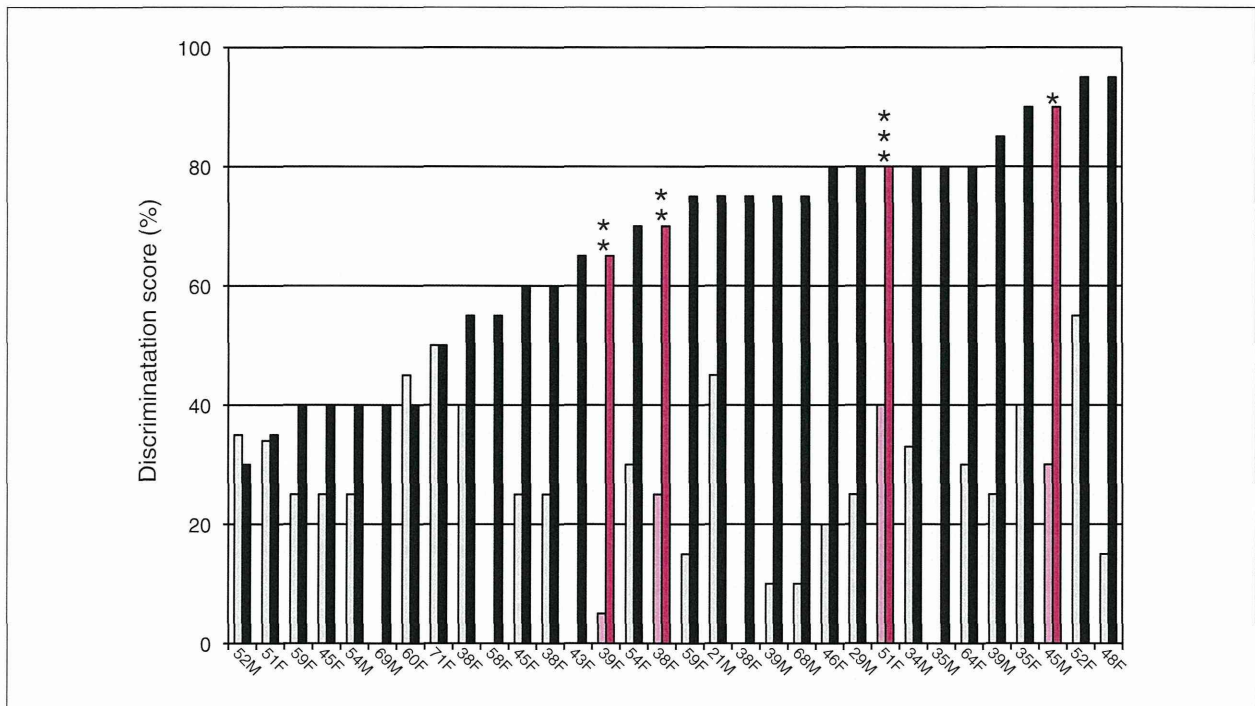


**Figure 5.** Three patients with *TMPRSS3* mutations (SNS5355, 4541, SNS5134) showed normal vestibular function on (left) caloric tests and (right) vestibular evoked myogenic potentials (VEMP).

### Outcome of EAS

Three patients with *TMPRSS3* compound heterozygous mutations (SNS5355, 4541, SNS5134) received EAS. We evaluated the improvement in speech discrimination and perception scores (using the 67S Japanese monosyllable test)

preoperatively and at 12 months after the initial EAS stimulation between 3 patients with *TMPRSS3* mutations who underwent EAS and the other 27 patients (Figure 6). Hearing preservation was achieved in all 30 patients (32 ears), with good speech perception observed for all patients. The *TMPRSS3* patients, in particular, showed good outcomes.



**Figure 6.** Speech discrimination scores (using the 67S Japanese monosyllable test, 70 dB SPL) preoperatively (grey) and at 12 months after the initial EAS (black). The 3 patients with *TMPRSS3* mutations (\*SNS5355, \*\*4541, \*\*\*SNS5134) showed significant improvement.

## Discussion

In previous studies, the frequency of *TMPRSS3* mutations in hearing loss patients was found to be <0.5% in Europe, 3% in Pakistan, and 5.9% in Korea.<sup>7,12,13</sup> In this study, MPS technology successfully identified *TMPRSS3* mutations and the frequency of *TMPRSS3* mutations in a Japanese population. The rate of patients with *TMPRSS3* mutations was 0.36% (4/1120) among Japanese and 0.7% (4/600) in autosomal recessive sensorineural hearing loss (ARSNHL) patients, which are similar to the figures for Europe.

Five patients were detected with compound heterozygous mutations in this study, with no specific mutation found to occur at a significantly higher frequency. The existence of certain frequent mutations, such as c.916G>A (p.A306T), has been reported.<sup>8,13,14</sup> The mutation spectrum found in Japanese is quite different from those reported in other populations. Based on the present results, the carrier rate of *TMPRSS3* mutations is extremely low in Japan in comparison with *GJB2* or *SLC26A4*.

For such rare causative mutations/genes, targeted exon sequencing of selected genes using MPS technology was extremely useful. In fact, in this study we successfully identified 7 *TMPRSS3* mutations among 4 families. All of the patients with *TMPRSS3* mutations showed typical ski slope

hearing loss with postlingual onset. However, the clinical characteristics varied among the patients.

Weegerink et al<sup>8</sup> hypothesized that *TMPRSS3* mutations are associated with 2 types of hearing impairment phenotypes, (1) DFNB10: a severe congenital or early childhood onset type with prelingual hearing impairment caused by the presence of 2 severe mutations and (2) DFNB8: a later onset progressive but initially milder type with postlingual hearing impairment caused by the presence of 1 mild and 1 severe mutation. In this study, patient ID 4541 had early onset and relatively rapid progressive hearing loss, resembling the DFNB10 phenotype. Her brother (patient ID 4540) also had a more severe phenotype that demonstrated earlier onset and deteriorated to profound hearing loss from young adulthood. Therefore, the c.226C>T (p.Q76X) and c.778G>A (p.A260T) mutations identified in this family are thought to be associated with early onset hearing loss. Conversely, patients SNS5355 and SNS5134 showed late-onset hearing loss. Those mutations might be associated with milder mutations, leading to a DFNB8 phenotype.

With regard to the function of *TMPRSS3* in the inner ear, it was hypothesized that *Tmprss3* participates in the regulation of sodium homeostasis through its ability to activate the inner ear-expressed sodium channel (ENaC) in vitro.<sup>5</sup> In the *Tmprss3*-related deafness mouse model, degeneration

of hair cells in the organ of Corti starts at the basal turn at the onset of hearing loss (postnatal day 12) and progresses toward the apex within 2 days.<sup>32</sup> This phenomenon observed in the mouse model is in line with the human phenotype that presents as high-frequency hearing loss.

Concerning vestibular function, the patients showed normal vestibular function, as evaluated by caloric test and cVEMP (Figure 5), and none of the patients showed vestibular symptoms. Similarly, none of the patients reported previously were found to have vestibular symptoms, but the vestibular function in some patients was reported to be affected. Weegerink et al<sup>8</sup> reported that 5 out of 9 patients showed mild hyperreflexia/hyporeflexia on rotatory and caloric tests. It is not surprising to find this type of associated vestibular hypofunction as *Tmprss3* is expressed both in the cochlear and vestibular hair cells.<sup>32</sup> A similar discrepancy between gene expression, vestibular testing, and vestibular symptoms has also been reported for *GJB2*-associated deafness.<sup>33</sup> Vestibular compensation may be related to these complications, and this possibility should be further examined in future studies.

Three of the 5 patients with mutations detected in this study received EAS. A good outcome for EAS in a *TMPRSS3* patient (patient ID: 4541) was previously reported.<sup>10</sup> In this study, 2 additional patients with *TMPRSS3* mutations also showed good outcomes, further confirming that patients with *TMPRSS3* mutations are good candidates for EAS.

The outcomes for CI for *TMPRSS3* patients remain controversial.<sup>8,10,14,34</sup> A majority of cases (13/15) were reported to show good outcomes for CI, while 2 cases reported by Eppsteiner et al<sup>34</sup> showed a poorer performance. *Tmprss3* is reported to be expressed not only in the organ of Corti but also in the spiral ganglion, and the loss of ganglion cells has been reported.<sup>32,34</sup> Therefore, it is possible that neuronal cell loss may negatively affect CI performance. However, the majority of cases, including our 3 EAS cases, showed good performance, indicating that CI and/or EAS is a potential therapeutic option. If the progression of hearing loss results in the patients losing the benefits of EAS acoustic stimulation, it is possible to cover all frequencies by electric stimulation as is common in CI. A recent study on the human temporal bones indicated that the behavior of human ganglion cells is different from those in animals, being more resistant to degeneration.<sup>35</sup> Although no study of the temporal bone in relation to patients with *TMPRSS3* mutations is available, such behavioral differences may provide an explanation of why CI/EAS is effective in patients with *TMPRSS3* mutations.

Clinicians should keep in mind that hearing loss caused by *TMPRSS3* mutations may be progressive and should consider proper intervention for these patients.

In conclusion, the present study provided additional evidence that the patients associated with *TMPRSS3* mutations

are good candidates for EAS. Genetic testing based on next-generation sequencing will facilitate candidate selection and personalized intervention.

### Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

### Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This study was supported by a Health and Labour Sciences Research Grant for Research on Rare and Intractable Diseases (H24-Nanchiou(Nan)-Ippan-032) and Comprehensive Research on Disability Health and Welfare (H25-Kankaku-Ippan-002) from the Ministry of Health, Labour and Welfare of Japan (S.U.) and by a Grant-in-Aid for Scientific Research (A) (22249057) from the Ministry of Education, Science and Culture of Japan (S.U.).

### References

1. Smith RJ, Bale JF Jr, White KR. Sensorineural hearing loss in children. *Lancet*. 2005;365(9462):879-890.
2. Wilson BS. The future of cochlear implants. *Br J Audiol*. 1997;31(4):205-225.
3. von Ilberg CA, Baumann U, Kiefer J, Tillein J, Adunka OF. Electric-acoustic stimulation of the auditory system: a review of the first decade. *Audiol Neurootol*. 2011;16(suppl 12):1-30.
4. Scott HS, Kudoh J, Wattenhofer M, et al. Insertion of beta-satellite repeats identifies a transmembrane protease causing both congenital and childhood onset autosomal recessive deafness. *Nat Genet*. 2001;27(1):59-63.
5. Guipponi M, Vuagniaux G, Wattenhofer M, et al. The transmembrane serine protease (*TMPRSS3*) mutated in deafness DFNB8/10 activates the epithelial sodium channel (ENaC) in vitro. *Hum Mol Genet*. 2002;11(23):2829-2836.
6. Guipponi M, Toh MY, Tan J, et al. An integrated genetic and functional analysis of the role of type II transmembrane serine proteases (*TMPRSSs*) in hearing loss. *Hum Mutat*. 2008;29(1):130-141.
7. Wattenhofer M, Di Iorio MV, Rabionet R, et al. Mutations in the *TMPRSS3* gene are a rare cause of childhood nonsyndromic deafness in Caucasian patients. *J Mol Med (Berl)*. 2002;80(2):124-131.
8. Weegerink NJ, Schraders M, Oostrik J, et al. Genotype-phenotype correlation in DFNB8/10 families with *TMPRSS3* mutations. *J Assoc Res Otolaryngol*. 2011;12(6):753-766.
9. Lee K, Khan S, Islam A, et al. Novel *TMPRSS3* variants in Pakistani families with autosomal recessive non-syndromic hearing impairment. *Clin Genet*. 2012;82(1):56-63.
10. Miyagawa M, Nishio SY, Ikeda T, Fukushima K, Usami S. Massively parallel DNA sequencing successfully identifies new causative mutations in deafness genes in patients with cochlear implantation and EAS. *PLoS One*. 2013;8(10):e75793.
11. Charif M, Abidi O, Boulouiz R, et al. Molecular analysis of the *TMPRSS3* gene in Moroccan families with

- non-syndromic hearing loss. *Biochem Biophys Res Commun.* 2012;419(4):643-647.
12. Ben-Yosef T, Wattenhofer M, Riazuddin S, et al. Novel mutations of *TMPRSS3* in four DFNB8/B10 families segregating congenital autosomal recessive deafness. *J Med Genet.* 2001;38(6):396-400.
  13. Chung J, Park SM, Chang SO, et al. A novel mutation of *TMPRSS3* related to milder auditory phenotype in Korean postlingual deafness: a possible future implication for a personalized auditory rehabilitation. *J Mol Med (Berl).* 2014;92(6):651-663.
  14. Elbracht M, Senderek J, Eggermann T, et al. Autosomal recessive postlingual hearing loss (DFNB8): compound heterozygosity for two novel *TMPRSS3* mutations in German siblings. *J Med Genet.* 2007;44(6):e81.
  15. Wattenhofer M, Sahin-Calapoglu N, Andreassen D, et al. A novel *TMPRSS3* missense mutation in a DFNB8/10 family prevents proteolytic activation of the protein. *Hum Genet.* 2005;117(6):528-535.
  16. Masmoudi S, Antonarakis SE, Schwede T, et al. Novel missense mutations of *TMPRSS3* in two consanguineous Tunisian families with non-syndromic autosomal recessive deafness. *Hum Mutat.* 2001;18(2):101-108.
  17. Ahmed ZM, Li XC, Powell SD, et al. Characterization of a new full length *TMPRSS3* isoform and identification of mutant alleles responsible for nonsyndromic recessive deafness in Newfoundland and Pakistan. *BMC Med Genet.* 2004;5:24.
  18. Fasquelle L, Scott HS, Lenoir M, et al. *Tmprss3*, a transmembrane serine protease deficient in human DFNB8/10 deafness, is critical for cochlear hair cell survival at the onset of hearing. *J Biol Chem.* 2011;286(19):17383-17397.
  19. Wang K, Li M, Hakonarson H. ANNOVAR: functional annotation of genetic variants from high-throughput sequencing data. *Nucleic Acids Res.* 2010;38(16):e164.
  20. Chang X, Wang K. wANNOVAR: annotating genetic variants for personal genomes via the web. *J Med Genet.* 2012;49(7):433-436.
  21. Abecasis GR, Auton A, Brooks LD, et al. An integrated map of genetic variation from 1,092 human genomes. *Nature.* 2012;491(7422):56-65.
  22. NHLBI Exome Sequencing Project (ESP) Exome Variant Server. <http://evs.gs.washington.edu/EVS/>. Accessed February 10, 2015.
  23. Narahara M, Higasa K, Nakamura S, et al. Large-scale East-Asian eQTL mapping reveals novel candidate genes for LD mapping and the genomic landscape of transcriptional effects of sequence variants. *PLoS One.* 2014;9(6):e100924.
  24. Pollard KS, Hubisz MJ, Rosenbloom KR, et al. Detection of nonneutral substitution rates on mammalian phylogenies. *Genome Res.* 2010;20(1):110-121.
  25. Kumar P, Henikoff S, Ng PC. Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. *Nat Protoc.* 2009;4(7):1073-1081.
  26. Adzhubei IA, Schmidt S, Peshkin L, et al. A method and server for predicting damaging missense mutations. *Nat Methods.* 2010;7(4):248-249.
  27. Chun S, Fay JC. Identification of deleterious mutations within three human genomes. *Genome Res.* 2009;19(9):1553-1561.
  28. Schwarz JM, Rodelsperger C, Schuelke M, et al. MutationTaster evaluates disease-causing potential of sequence alterations. *Nat Methods.* 2010;7(8):575-576.
  29. Cooper GM, Stone EA, Asimenos G, et al. Distribution and intensity of constraint in mammalian genomic sequence. *Genome Res.* 2005;15(7):901-913.
  30. Usami S, Moteki H, Tsukada K, et al. Hearing preservation and clinical outcome of 32 consecutive electric acoustic stimulation (EAS) surgeries. *Acta Otolaryngol.* 2014;134(7):717-727.
  31. Miyagawa M, Naito T, Nishio SY, Kamatani N, Usami S. Targeted exon sequencing successfully discovers rare causative genes and clarifies the molecular epidemiology of Japanese deafness patients. *PLoS One.* 2013;8(8):e71381.
  32. Fasquelle L, Scott HS, Lenoir M, et al. *Tmprss3*, a transmembrane serine protease deficient in human DFNB8/10 deafness, is critical for cochlear hair cell survival at the onset of hearing. *J Biol Chem.* 2011;286(19):17383-17397.
  33. Tsukada K, Fukuoka H, Usami S. Vestibular functions of hereditary hearing loss patients with *GJB2* mutations. *Audiol Neurootol.* In press.
  34. Eppsteiner RW, Shearer AE, Hildebrand MS, et al. Prediction of cochlear implant performance by genetic mutation: the spiral ganglion hypothesis. *Hear Res.* 2012;292(1-2):51-58.
  35. Liu W, Edin F, Atturo F, et al. The pre- and post-somatic segments of the human type I spiral ganglion neurons—structural and functional considerations related to cochlear implantation. *Neuroscience.* 2015;284:470-482.

# Detailed Hearing and Vestibular Profiles in the Patients with *COCH* Mutations

Annals of Otolaryngology & Laryngology  
1–11

© The Author(s) 2015  
Reprints and permissions:  
sagepub.com/journalsPermissions.nav  
DOI: 10.1177/0003489415573074  
aor.sagepub.com



Keita Tsukada, MD, PhD<sup>1</sup>, Aya Ichinose, MD<sup>1</sup>, Maiko Miyagawa, MD, PhD<sup>1,2</sup>,  
Kentaro Mori, MD<sup>1</sup>, Mitsuru Hattori, PhD<sup>1</sup>, Shin-ya Nishio, PhD<sup>1,2</sup>,  
Yasushi Naito, MD, PhD<sup>3</sup>, Shin-ichiro Kitajiri, MD, PhD<sup>4</sup>,  
and Shin-ichi Usami, MD, PhD<sup>1,2</sup>

## Abstract

**Objectives:** To evaluate the clinical features of Japanese DFNA9 families with mutations of the *COCH* gene.

**Methods:** Mutation screening was performed using targeted next-generation sequencing (NGS) for 63 previously reported deafness genes. The progression of hearing loss and vestibular dysfunction were evaluated by pure-tone audiometry, caloric testing, cVEMP, and computed dynamic posturography.

**Results:** We detected 1 reported mutation of p.G88E and 2 novel mutations of p.I372T and p.C542R. The patients with the novel mutations of p.I372T and p.C542R within the vWFA2 domain showed early onset progressive hearing loss, and the patients with the p.G88E mutation showed late onset hearing loss and acute hearing deterioration over a short period. Vestibular symptoms were reported in the patients with p.G88E and p.C542R. Vestibular testing was performed for the family with the p.G88E mutation. Severe vestibular dysfunction was observed in the proband, and the proband's son showed unilateral semicircular canal dysfunction with mild hearing loss.

**Conclusions:** Targeted exon resequencing of selected genes using NGS successfully identified mutations in the relatively rare deafness gene, *COCH*, in the Japanese population. The phenotype is compatible with that described in previous reports. Additional supporting evidence concerning progressive hearing loss and deterioration of vestibular function was obtained from our study.

## Keywords

*COCH*, DFNA9, progressive hearing loss, vestibular dysfunction, Massively Parallel Sequencing, next-generation sequencing

## Introduction

The majority of genetic hearing loss is autosomally inherited, and autosomal dominant nonsyndromic hearing loss (ADNSHL) is estimated to be responsible for 20% of those with a genetic cause,<sup>1</sup> with 30 causative genes identified to date (Hereditary Hearing Loss Homepage, <http://hereditary-hearingloss.org/>). Mutations in the *COCH* gene are well known to cause ADNSHL with vestibular dysfunction (DFNA9).<sup>2</sup>

*COCH* encodes a 550 amino acid extracellular protein “cochlin” that includes 1 late gestation lung protein Lgl1 (LCCL) domain and 2 von Willebrand factor A (vWFA) domains.<sup>3,4</sup> Although the precise function of cochlin remains unclear, it is well known to be predominantly expressed in the inner ear<sup>3,5</sup> and constitutes 70% of all inner ear proteins,<sup>6</sup> suggesting that this protein plays an important role in the inner ear.

Our previous report showed a Japanese family with autosomal dominant progressive cochleo-vestibular dysfunction caused by a p.A119T mutation in the *COCH* gene.<sup>7</sup> However, mutations of the *COCH* gene are thought to be rare in the Japanese hearing loss patients. In this study, we identified 4 families with causative *COCH* mutations in a large cohort of hearing loss patients by the use of

targeted next-generation sequencing (NGS) of deafness genes, and we herein summarize the clinical features observed for Japanese families with *COCH* mutations.

## Subjects and Methods

### Subjects

A total of 1120 Japanese hearing loss (HL) patients (ADSNHL, 266; ARSNHL, 600; unknown, 254) from 53 ENT departments nationwide participated in this study.

<sup>1</sup>Department of Otorhinolaryngology, Shinshu University School of Medicine, Matsumoto, Japan

<sup>2</sup>Department of Hearing Implant Sciences, Shinshu University School of Medicine, Matsumoto, Japan

<sup>3</sup>Department of Otorhinolaryngology, Kobe City Medical Center General Hospital, Hyogo, Japan

<sup>4</sup>Department of Otolaryngology, Head and Neck Surgery, Kyoto University Graduate School of Medicine, Kyoto, Japan

### Corresponding Author:

Shin-ichi Usami, MD, PhD, Department of Otorhinolaryngology, Shinshu University School of Medicine, 3-1-1 Asahi, Matsumoto 390-8621, Japan.  
Email: [usami@shinshu-u.ac.jp](mailto:usami@shinshu-u.ac.jp)



Written informed consent was obtained from all subjects (or from their next of kin, caretaker, or guardian in the case of minors/children) prior to enrollment in the project. This study was approved by the Shinshu University Ethical Committee as well as the respective ethical committees of the other participating institutions.

### Amplicon Library Preparation

Amplicon libraries were prepared using an Ion AmpliSeq Custom Panel (Applied Biosystems, Life Technologies, Carlsbad, California, USA), according to the manufacturer's instructions, for 63 genes reported to cause nonsyndromic hearing loss. The detailed protocol was described elsewhere.<sup>8</sup> After preparation, the amplicon libraries were diluted to 20 pM, and equal amounts of 6 libraries for 6 patients were pooled for 1 sequence reaction.

### Emulsion Polymerase Chain Reaction and Sequencing

Emulsion polymerase chain reaction (PCR) and sequencing were performed according to the manufacturer's instructions. The detailed protocol was described elsewhere.<sup>8</sup> Massively Parallel Sequencing was performed with an Ion Torrent Personal Genome Machine (PGM) system using an Ion PGM 200 Sequencing Kit and an Ion 318 Chip (Life Technologies).

### Base Call and Data Analysis

The sequence data were mapped against the human genome sequence (build GRCh37/hg19) with a Torrent Mapping Alignment Program. After sequence mapping, the DNA variant regions were piled up with Torrent Variant Caller plug-in software. After variant detection, their effects were analyzed using ANNOVAR software.<sup>9,10</sup> The missense, nonsense, insertion/deletion, and splicing variants were selected from among the identified variants. Variants were further selected as less than 1% of (1) the 1000 genome database<sup>11</sup>, (2) the 6500 exome variants, (3) the Human Genetic Variation Database (data set for 1208 Japanese exome variants)<sup>12</sup>, and (4) the 269 in-house Japanese normal hearing controls.

The pathogenicity of the missense variants was predicted using the following functional prediction software: PhyloP<sup>13</sup>, Sorting Intolerant from Tolerant (SIFT)<sup>14</sup>, Polymorphism Phenotyping<sup>15</sup>, LRT<sup>16</sup>, MutationTaster<sup>17</sup>, and GERP++.<sup>18</sup>

Candidate mutations were confirmed by Sanger sequencing, and the responsible mutations were identified by segregation analysis using samples from among the patients' family members.

### Audiologic Evaluations

Audiometric evaluation from 125 to 8000 Hz was performed by pure-tone audiometry. To evaluate speech perception outcomes, speech discrimination scores (using the 67S Japanese monosyllable test) and speech perception scores (using the Japanese CI2004 word and sentence test) were used. Subjects sat 1 m away from and facing the loudspeaker (azimuth = 0°) and recorded monosyllable words presented at 70 dB SPL in quiet condition.

### Vestibular Evaluations

**Caloric Testing.** Caloric testing involved the measurement of the maximum slow phase velocity (SPV) by cold water irrigation (20°C, 5 ml, 20 s). We defined a maximum SPV value below 10 deg/s as representing areflexia and a value between 10 to 20 deg/s as representing hyporeflexia.

**cVEMP.** For cVEMP testing, electromyography (EMG) was performed using a pair of surface electrodes mounted on the upper half and sternal head of the sternocleidomastoid (SCM) muscle, respectively. The electrographic signal was recorded using a Neuropack evoked potential recorder (Nihon Kohden Co Ltd, Tokyo, Japan). Clicks lasting for 0.1 ms at 105 dB nHL were presented through a headphone. The stimulation rate was 5 Hz, the bandpass filter intensity was 20 to 2000 Hz, and the analysis time was 50 ms. The responses to 100 stimuli were averaged twice.

**Computed Dynamic Posturography (Equitest) by Sensory Organization Testing.** Sensory Organization Testing (SOT), which was used to assess overall balance and compensation of balance, was performed using the Balance Manager system (NeuroCom, Clackamas, Oregon, USA) under 6 sensory conditions, each of which was performed in triplicate, as follows: condition 1: eyes open, fixed platform surface and background; condition 2: eyes closed, fixed platform surface and background; condition 3: eyes open, moving background (directly proportional to patient antero-posterior sway); condition 4: eyes open, moving platform surface (directly proportional to patient antero-posterior sway); condition 5: eyes closed, moving platform surface; and condition 6: eyes open, moving platform and background.

Equilibrium scores (ES), which indicate the amplitude of the sway angle based on the maximum displacement of the center of gravity, were calculated from the ratio of the estimated antero-posterior sway to the theoretical range of normal antero-posterior sway (12.5°), and the mean ES from 3 trials was used. When a trial ended with a fall or the mean ES was beyond the 95% confidence interval of age-specific normative data, it was considered a failure. Sensory analysis (SA) was also used to assess total balance and

compensation. Based on average balance scores in specific pairs of trials, SA classifies sensory dysfunction and the abnormal sensory preference including (1) somatosensory (SOM; the ratio between conditions 2 and 1), (2) visual (VIS; conditions 4:1), (3) vestibular (VEST; conditions 5:1), and (4) visual preference (PREF; conditions 3+6:2+5).

## Results

We identified 4 families among the study cohort that had causative mutations of the *COCH* gene.

### Mutation Analysis

We found 1 previously reported causative mutation, p.G88E, in 1 family (Figure 1) as well as 2 novel *COCH* missense mutations. One of the novel mutations detected in 2 of the Japanese families, FAM 475 and FAM 535, is a heterozygous c.T1115C (NM\_004086) (Figure 4A) mutation in exon 11 that substitutes a threonine residue for an isoleucine (p.I372T) in the vWFA2 domain. We performed Sanger sequencing as part of a family segregation study and confirmed the co-segregation of p.I372T and the disease phenotype in FAM 475. In addition, p.I372T was predicted to be deleterious by both PolyPhen2 (score of 1.0) and SIFT (score of 0.996) programs.

The other heterozygous c.T1624C (NM\_004086) mutation (Figure 5A) in exon 12, leading to a p.C542R substitution in the vWFA2 domain, was found in a member of family FAM 986. Both PolyPhen2 and SIFT predicted that the substitution caused “probably damaging” with a highest possible score of 1.0 and 0.998, respectively.

To exclude common polymorphisms, the 1000 genome database, the 6500 exome variants, and the Human Genetic Variation Database (data set for 1208 Japanese exome variants and 269 in-house Japanese normal hearing controls) were analyzed. These mutations were not found in any of the databases. Therefore, we concluded that the p.I372T and p.C542R mutations were causative mutations associated with hearing loss.

### Details of a Case of p.G88E

The pedigree and audiogram of the family of the subject with a p.G88E mutation are shown in Figure 1A and 1B, respectively.

### Audiologic Profile

The proband was a 70-year-old man (III-2), and changes in his hearing are shown in Figure 2A. He first noticed hearing loss without tinnitus or vertigo in his early 50s, and his first visit to the Shinshu University Hospital was at the age

of 57. Pure-tone audiograms showed hearing loss in the higher frequencies (an average threshold of 45 dB, bilaterally), and his speech discrimination scores were 80% in the right ear and 90% in the left ear. The patient experienced a sudden deterioration in hearing in his left ear with tinnitus, resulting in profound hearing loss in all frequencies at age 64. Moreover, acute progression of hearing loss in his right ear occurred soon after that event, and he developed bilateral deafness with a speech discrimination score of 16% even with hearing aid at age 65. Cochlear implantation (CI) was therefore performed for the right ear at age 68. He received a MED-EL PULSAR with FLEXSOFT electrode (MED-EL, Innsbruck, Austria) through a round window approach. The speech perception scores using the Japanese CI2004 word and sentence test in quiet thereafter improved from 43% and 63% preoperatively using a hearing aid to 76% and 75% with CI, respectively (Figure 2B).

The proband's son (IV-2), aged 42, showed mild to moderate hearing loss at the higher frequencies (2 kHz, 4 kHz, and 8 kHz) with tinnitus in the left ear without awareness of hearing loss (Figure 1B). Otoacoustic emissions (OAEs) showed an abnormal response at the higher frequencies.

### Vestibular Profiles

The detailed results of vestibular testing are shown in Figure 3.

The proband (III-2) experienced an onset of dizziness at the age of 64, and caloric testing, cVEMP, and SOT were subsequently performed on the proband and his son. The proband showed no response bilaterally in the caloric testing and cVEMP. In the SOT, he fell into a safety harness under conditions 2, 3, and 4, and could not continue under conditions 5 and 6.

Although the proband's son (IV-2) had no symptoms of vestibular dysfunction and a normal bilateral response in cVEMP, caloric testing revealed areflexia in the left ear (MSPV: right, 22.2 deg/s; left, 7.3 deg/s) and a fall during 1 of 3 trials under condition 5 occurred in the SOT.

### Novel p.I372T Mutation

The pedigrees and audiograms of patients with p.I372T are shown in Figure 4.

The proband (III-2) of FAM 475, a 38-year-old female, first noticed hearing loss at the age of 33, after which it progressed gradually. Hearing loss was more severe in the higher frequencies, with only mild impairment in the lower frequencies. His father (II-4), who started to lose his hearing at age 42, had a downward sloping audiogram that was worse than that of the proband. Neither subject complained of any vestibular symptoms at the time of testing.