Chhibber et al.

each individual, and used to estimate pathway-specific heritability in GCTA. Empirical p-values were generated by calculating the probability of obtaining a heritability estimate greater than that estimated from observed data.

### **PLINK Set Test**

Briefly, the PLINK Set test as implemented in this study calculates the mean of all significant (p < 0.05) per-SNP p-values after filtering for SNPs in linkage disequilibrium ( $r^2 \ge 0.5$ ). An empirical p-value is applied to each set test by permuting phenotype labels across individuals. SNP p-values for severity and onset of neuropathy were calculated in PLINK by linear regression of residuals from regression of grade of neuropathy on number of minor alleles, with log cumulative dose of paclitaxel and principal components as covariates in both initial regression and PLINK set test.

### Results

The variance explained by common (MAF > 1%) SNPs for paclitaxel-induced neuropathy was estimated in a cohort of 845 unrelated Caucasian breast cancer patients treated with single agent paclitaxel. Two outcomes were of interest − severity of neuropathy (measured on a grade of 0 to 5) and cumulative dose administered at onset of neuropathy (≥ grade 2), both treated as continuous quantitative variables. The variance explained by all genotyped SNPs across the genome was estimated as 41% for severity of neuropathy and 55% for onset of neuropathy, but with high standard errors (44% and 47%, respectively) due to the small sample size. To narrow in on the causative SNPs driving heritability and reduce noise from non-causative SNPs, two methods were applied: (1) a genomic position based SNP selection, extracting SNPs in genic regions, and (2) a biological pathway based selection that extracted SNPs that fall in biological pathways that are associated with putative mechanisms for susceptibility to paclitaxel-induced neuropathy.

When partitioning the genome in SNP sets by genomic location (Figure 2), a trend toward higher heritability was found in genic regions for severity ( $h^2 = 49\% \pm 37\%$ , p = 0.07) and onset of peripheral neuropathy ( $h^2 = 48\% \pm 35\%$ , p = 0.08). For severity of peripheral neuropathy, pathway specific results show highest heritability estimates for the Axonogenesis gene set ( $h^2 = 21\% \pm 12\%$ , h = 0.040; Table 1). A complementary pathway analysis approach, the PLINK set test, was used to further extend our pathway based heritability results. Consistent with the GCTA analysis, only the Axonogenesis set is significant (h = 0.012) for severity of neuropathy using the set test (Supplemental Table 1). For onset of peripheral neuropathy, no significant signal of heritability was detected in any of the pathways tested (Supplemental Table 2).

"Children" of the GO Axonogenesis term, defined as terms with a "is\_a" or "part\_of" relationship with the Axonogenesis term, were subsequently tested for the severity of neuropathy phenotype (Table 2). Of the ten terms tested, GO Regulation of Axonogenesis (GO: 0050770), GO Axon Extension (GO: 0048675), and GO CNS Neuron Axonogenesis (GO: 0021955) showed strong heritability signals ( $h^2 = 13\% \pm 6\%$  (p = 0.009),  $10\% \pm 5\%$  (p = 0.020) and  $5\% \pm 3\%$  (p = 0.020), respectively). To determine whether the signal from these three terms comes from independent genes in each set or overlapping genes in the

Pharmacogenomics J. Author manuscript; available in PMC 2015 February 01.

three sets, heritability estimates were calculated using the pair-wise and three-way union or intersection of the GO Regulation of Axonogenesis, GO Axon Extension, and GO CNS Neuron Axonogenesis sets. The union or intersection of the GO CNS Neuron Axonogenesis set with GO Axon Extension or GO Regulation of Axonogenesis sets resulted in lower heritability estimates than either independent set with high standard error (data not shown). For the GO Axon Extension and GO Regulation of Axonogenesis sets, the heritability signal from each independent set and the union and intersection sets are very similar (Figure 3), suggesting that a large portion of the SNPs driving the heritability in the Regulation and Extension sets come from the 44 genes found in both gene sets.

Heritability estimates were also calculated using imputed data; as with the genotyped SNPs, whole genome estimates of heritability with imputed SNPs had very high standard errors. For genomic position and pathway analyses, results from imputed data were similar to those described above for genotyped data, with a trend to higher heritability estimates in genic versus intergenic regions for the severity of peripheral neuropathy (Supplemental Table 3) and in the GO Axonogenesis set for severity of peripheral neuropathy (Supplemental Tables 4-6).

## Discussion

These results suggest that a portion of variation in severity and onset of paclitaxel-induced sensory peripheral neuropathy is captured by additive effects of common SNPs in this clinical trial population. Previous studies have indicated that heritability is driven primarily by SNPs in genic regions<sup>40</sup>, and a similar trend is found in our study. Within genic regions, we also noted a higher proportion of variance in severity and onset of peripheral neuropathy captured by SNPs in intronic regions (data not shown), but it is unclear whether this is due to a bias in the design of the genotyping chip or true bias in the genomic location of SNPs associated with paclitaxel induced neuropathy. If real, the enrichment of heritability signal in introns suggests that the majority of causal SNPs have subtle biological effects – for example, small changes in expression or stability that may be regulated by intronic SNPs, rather than overt changes in protein structure or function caused by variation in exons. This is consistent with a polygenic model in which many small, additive effects together contribute to the phenotype.

Further, a set of genes was identified that drive a substantial portion of the heritability of severity of paclitaxel-induced peripheral neuropathy, implicating axonogenesis, and more specifically the regulation of axon outgrowth, in the pathophysiology of this adverse event. These results are supported by evidence from human biopsies, electrophysiological studies, and animal and cell-based models that paclitaxel causes a distal axonopathy, in which the degeneration of axons occurs first at axon ends. This pattern of neuronal damage is consistent with a length-dependent neuropathy, targeting the long axons that extend into the hands and feet first, as typically occurs with paclitaxel induced neuropathy<sup>41–44</sup>. Further, there is evidence that demyelination and ganglionopathy, if they do occur, are secondary to axon damage<sup>41, 44, 45</sup>. The current results suggest that susceptibility to paclitaxel-induced neuropathy is caused in part by heightened sensitivity to or reduced capacity to repair this distal axon damage.

Of the 44 genes in the GO Axon Extension and GO Regulation of Axonogenesis overlap set (Supplemental Table 7), a number have been implicated in neuropathy, including hereditary neuropathy genes (MAP1B<sup>46</sup>, NGF<sup>47</sup>,FXN<sup>48</sup>), genes with variants or expression signatures associated with diabetic or HIV-induced peripheral neuropathy (APOE<sup>49, 50</sup>, MAPT<sup>51</sup>, CDH4<sup>51</sup>), genes involved in neurological pain pathways (MT3<sup>52</sup>, TRPV2<sup>53</sup>, CCR5<sup>54</sup>, CXCL12<sup>55</sup>), and genes involved in response to or repair/prevention of peripheral nerve damage (RYK<sup>56</sup>, SLIT1<sup>57</sup>, NTRK3<sup>58</sup>, NGF<sup>59, 60</sup>, TRPV2<sup>53</sup>, NTN1<sup>61</sup>, NDEL1<sup>62</sup>). The majority (38) of these 44 genes fall in the GO term Regulation of Axon Extension (GO 0030516), which is a subset of both GO Regulation of Axonogenesis and GO Axon Extension.

The pathway results are also consistent with gene expression analyses in mouse and human studies of diabetic neuropathy. In a study examining the pathophysiology of diabetes-induced neuropathy the GO Axonogenesis term was identified as an overrepresented pathway in a differential expression analysis in the *db/db* vs *db/+* mouse sciatic nerve<sup>51</sup>. Similarly, the GO Regulation of Axonogenesis term was identified as an overrepresented set in genes up-regulated in sural nerve biopsies from patients with advanced progression of diabetic neuropathy<sup>63</sup>. Although neuron damage is caused by different mechanisms in diabetes and following paclitaxel treatment, these results suggest that susceptibility to sensory peripheral neuropathy is driven by the same sets of genes.

Despite success in estimating heritability for paclitaxel-induced neuropathy and identifying a subset of the genome driving this heritability, some limitations in available methods and data are noted. One of the primary limitations of any pathway or gene set based analysis is the gene set definitions available. All available set definitions are limited by current knowledge about the pathway in question, and well curated sets are restricted to those pathways of interest to researchers. Further, the number of SNPs captured per gene varies, either because of true differences between number of variants or haplotype structure between genes, or because of differences in coverage between genes on the genotyping platform that was used. Such variability in local coverage is known to be a limitation in all commercial genotyping platforms<sup>64</sup>. While imputation of missing SNPs did increase SNP density in each set, heritability estimates with imputed data were close to those with just genotyped data; because of the high imputation quality threshold used ( $r^2 > 0.9$ ), it is likely that additional SNPs are in high LD with genotyped SNPs, adding little additional information. For onset of peripheral neuropathy, no significant signal of heritability was detected in any of the pathways tested, either because genes driving heritability of onset of neuropathy are in a pathway we did not select, or because the use of deviance residuals from the Cox proportional hazards regression rather than a direct proportional hazards regression did not adequately model the data. It is also possible that one or more of the selected pathways is incompletely annotated. Gene Ontology terms are annotated using a combination of experimental evidence and computational analyses, and can be both manually and electronically annotated<sup>32, 65</sup>. The extensive set of sources for term annotation makes Gene Ontology the most comprehensive source of annotated terms available, but also contributes to significant noise (incorrectly assigned genes) being built into the terms. Unfortunately, highly accurate manually annotated gene sets are currently limited, and those that exist reflect the current body of knowledge regarding a given pathway. The Gene

Ontology was the only database that included gene sets for each of the peripheral neuropathy mechanisms of interest. For the GO set Axonogenesis, more restrictive set definitions were investigated, including limiting pathway genes to those annotated to Axonogenesis by experimental evidence and those that were direct associations. The GO Axonogenesis experimental set gave an estimate of heritability significantly lower than that derived from the complete gene set (8% vs 22% for the complete set), suggesting that using a more conservative gene annotation would result in loss of power (Supplemental Table 8).

The standard errors for the whole-genome heritability analyses are high due to the limited sample size. Large sample sizes are difficult to obtain in genomic studies of drug toxicities, since recruitment into these studies is often limited to existing clinical trials. However, by narrowing in on the "causative" SNPs, signals of heritability were obtained even with relatively small sample sizes. In this study, constraints were also imposed by the linear mixed modeling method applied, which requires a continuous or dichotomous phenotype. Although severity of neuropathy is best modeled as an ordinal variable, it is treated as a continuous quantitative variable for the purpose of this study. Likewise, onset of neuropathy is best fit in a survival model but deviance residuals from a survival model were used as a continuous trait in the current analysis. Despite these limitations, the results from the modified phenotype definitions are likely close to those that would be estimated from the application of non-linear phenotype definitions. For example, effect estimates for SNPs in biological pathways from severity of neuropathy modeled as a linear or ordinal variable (Supplemental Figure 3) or onset of neuropathy modeled as a linear phenotype or time-toevent analysis (Supplemental Figure 4) are highly correlated ( $r^2 = 0.91$  and 0.97, respectively). However it is important to note that, because of the constraints on the phenotype definition, we treat heritability estimates obtained from our analyses simply as an indication of association between a certain sets of SNPs and our phenotypes of interest, rather than absolute measures of percent of variance explained by a particular SNP set. Finally, a gene boundary cutoff of 10 kb was selected to ensure that the SNPs are associated with the genes in our pathway (as opposed to a neighboring gene), though at the cost of losing potential causative SNPs in upstream and downstream regulatory regions of a gene. Because most genetic variability appears to be explained by SNPs in or near genes<sup>40</sup> our approach likely captures a significant fraction of the variability explained by the genes in a given set.

In summary, these results suggest that there is a heritable component to the severity and dose to onset of paclitaxel-induced sensory peripheral neuropathy. Further, genes involved in axon outgrowth may modulate the severity of paclitaxel-induced neuropathy. Understanding the mechanisms and pathways involved in susceptibility to paclitaxel-induced sensory peripheral neuropathy will help identify therapies that can mitigate the toxicity and guide future drug development.

# **Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

## **Acknowledgments**

The research for CALGB 60202 and 40101 was supported, in part, by grants from the National Cancer Institute (CA31946) to the Alliance for Clinical Trials in Oncology (Monica M. Bertagnolli, M.D., Chair) and to the Alliance Statistics and Data Center (Daniel J. Sargent, Ph.D., CA33601). The content of this manuscript is solely the responsibility of the authors and does not necessarily represent the official views of the National Cancer Institute.

This work was also supported in part by NIH grants U01 GM61390, U01 GM61393 and U01 HL065962 and the Biobank Japan Project funded by the Japanese Ministry of Education, Culture, Sports, Science and Technology. The genotyping used for this work was generated as part of the NIH Pharmacogenomics Research Network-RIKEN Center for Genomic Medicine Global Alliance. Aparna Chhibber and Megan Li were supported in part by NIH Training Grant T32 GM007175.

#### References

- Rowinsky EK, Donehower RC. Paclitaxel (taxol). N Engl J Med. 1995; 332:1004–1014. [PubMed: 7885406]
- Lee JJ, Swain SM. Peripheral neuropathy induced by microtubule-stabilizing agents. J Clin Oncol. 2006; 24:1633–1642. [PubMed: 16575015]
- 3. Donehower RC, Rowinsky EK, Grochow LB, Longnecker SM, Ettinger DS. Phase I trial of taxol in patients with advanced cancer. Cancer Treat Rep. 1987; 71:1171–1177. [PubMed: 2891441]
- 4. Postma TJ, Vermorken JB, Liefting AJ, Pinedo HM, Heimans JJ. Paclitaxel-induced neuropathy. Ann Oncol. 1995; 6:489–494. [PubMed: 7669713]
- Rowinsky EK, Chaudhry V, Cornblath DR, Donehower RC. Neurotoxicity of taxol. J Natl Cancer Inst Monogr. 1993:107–115. [PubMed: 7912516]
- 6. Rowinsky EK, Eisenhauer EA, Chaudhry V, Arbuck SG, Donehower RC. Clinical toxicities encountered with paclitaxel (Taxol). Semin Oncol. 1993; 20:1–15. [PubMed: 8102012]
- 7. Lipton RB, Apfel SC, Dutcher JP, Rosenberg R, Kaplan J, Berger A, et al. Taxol produces a predominantly sensory neuropathy. Neurology. 1989; 39:368–373. [PubMed: 2564647]
- Flatters SJ, Bennett GJ. Studies of peripheral sensory nerves in paclitaxel-induced painful peripheral neuropathy: evidence for mitochondrial dysfunction. Pain. 2006; 122:245–257. [PubMed: 16530964]
- Leandro-Garcia LJ, Leskela S, Jara C, Green H, Avall-Lundqvist E, Wheeler HE, et al. Regulatory
  polymorphisms in β-tubulin IIa are associated with paclitaxel-induced peripheral neuropathy. Clin
  Cancer Res. 2012; 18:4441–4448. [PubMed: 22718863]
- Leskela S, Jara C, Leandro-Garcia LJ, Martinez A, Garcia-Donas J, Hernando S, et al. Polymorphisms in cytochromes P450 2C8 and 3A5 are associated with paclitaxel neurotoxicity. Pharmacogenomics J. 2011; 11:121–129. [PubMed: 20212519]
- Sissung TM, Mross K, Steinberg SM, Behringer D, Figg WD, Sparreboom A, et al. Association of *ABCB1* genotypes with paclitaxel-mediated peripheral neuropathy and neutropenia. Eur J Cancer. 2006; 42:2893–2896. [PubMed: 16950614]
- 12. Bergmann TK, Green H, Brasch-Andersen C, Mirza MR, Herrstedt J, Holund B, et al. Retrospective study of the impact of pharmacogenetic variants on paclitaxel toxicity and survival in patients with ovarian cancer. Eur J Clin Pharmacol. 2011; 67:693–700. [PubMed: 21327421]
- Ofverholm A, Einbeigi Z, Manouchehrpour S, Albertsson P, Skrtic S, Enerback C. The ABCB1 3435 T allele does not increase the risk of paclitaxel-induced neurotoxicity. Oncol Lett. 2010; 1:151–154. [PubMed: 22966274]
- 14. Baldwin RM, Owzar K, Zembutsu H, Chhibber A, Kubo M, Jiang C, et al. A genome-wide association study identifies novel loci for paclitaxel-induced sensory peripheral neuropathy in CALGB 40101. Clin Cancer Res. 2012; 18:5099–5109. [PubMed: 22843789]
- Schneider BP, Li L, Miller K, Flockhart DA, Radovich M, Hancock BA, et al. Genetic associations with taxane-induced neuropathy by a genome-wide association study in E5103. J Clin Oncol. 2011; 29(Suppl):1000.

16. Bergmann TK, Vach W, Feddersen S, Eckhoff L, Green H, Herrstedt J, et al. GWAS-based association between RWDD3 and TECTA variants and paclitaxel induced neuropathy could not be confirmed in Scandinavian ovarian cancer patients. Acta Oncol. 2013; 52:871–874. [PubMed: 22877241]

- 17. Motsinger-Reif AA, Jorgenson E, Relling MV, Kroetz DL, Weinshilboum R, Cox NJ, et al. Genome-wide association studies in pharmacogenomics: successes and lessons. Pharmacogenet Genomics. 2010; 23:389–394.
- 18. Koren G, Cairns J, Chitayat D, Gaedigk A, Leeder SJ. Pharmacogenetics of morphine poisoning in a breastfed neonate of a codeine-prescribed mother. Lancet. 2006; 368:704. [PubMed: 16920476]
- Lee SH, Wray NR, Goddard ME, Visscher PM. Estimating missing heritability for disease from genome-wide association studies. Am J Hum Genet. 2011; 88:294

  –305. [PubMed: 21376301]
- Yang J, Benyamin B, McEvoy BP, Gordon S, Henders AK, Nyholt DR, et al. Common SNPs explain a large proportion of the heritability for human height. Nat Genet. 2010; 42:565–569. [PubMed: 20562875]
- Li Y, Willer C, Sanna S, Abecasis G. Genotype imputation. Annu Rev Genomics Hum Genet. 2009; 10:387–406. [PubMed: 19715440]
- Li Y, Willer CJ, Ding J, Scheet P, Abecasis GR. MaCH: using sequence and genotype data to estimate haplotypes and unobserved genotypes. Genet Epidemiol. 2010; 34:816–834. [PubMed: 21058334]
- Abecasis GR, Altshuler D, Auton A, Brooks LD, Durbin RM, Gibbs RA, et al. A map of human genome variation from population-scale sequencing. Nature. 2010; 467:1061–1073. [PubMed: 209810921
- Shulman LN, Cirrincione CT, Berry DA, Becker HP, Perez EA, O'Regan RO, et al. Six cycles of doxorubicin and cyclophosphamide or paclitaxel are not superior to 4 cycles as adjuvant chemotherapy for breast cancer in women with 0-3 positive axillary nodes: CALGB 40101. J Clin Oncol. 2012; 30:4071–4076. [PubMed: 22826271]
- Muggia FM, Braly PS, Brady MF, Sutton G, Niemann TH, Lentz SL, et al. Phase III randomized study of cisplatin versus paclitaxel versus cisplatin and paclitaxel in patients with suboptimal stage III or IV ovarian cancer: a gynecologic oncology group study. J Clin Oncol. 2000; 18:106–115. [PubMed: 10623700]
- Perez EA, Vogel CL, Irwin DH, Kirshner JJ, Patel R. Multicenter phase II trial of weekly paclitaxel in women with metastatic breast cancer. J Clin Oncol. 2001; 19:4216–4223. [PubMed: 11709565]
- Bouzigon E, Ulgen A, Dizier MH, Siroux V, Lathrop M, Kauffmann F, et al. Evidence for a
  pleiotropic QTL on chromosome 5q13 influencing both time to asthma onset and asthma score in
  French EGEA families, Hum Genet. 2007; 121:711–719. [PubMed: 17473937]
- 28. Dickson MR, Li J, Wiener HW, Perry RT, Blacker D, Bassett SS, et al. A genomic scan for age at onset of Alzheimer's disease in 437 families from the NIMH Genetic Initiative. Am J Med Genet B Neuropsychiatr Genet. 2008; 147B:784–792. [PubMed: 18189239]
- Hunt KJ, Lehman DM, Arya R, Fowler S, Leach RJ, Goring HH, et al. Genome-wide linkage analyses of type 2 diabetes in Mexican Americans: the San Antonio Family Diabetes/Gallbladder Study. Diabetes. 2005; 54:2655–2662. [PubMed: 16123354]
- 30. Therneau TM. A package for survival analysis in S. 2012
- 31. Therneau, TM.; Grambsch, PM. Modeling Survival Data: Extending the Cox Model. Springer-Verlag: New York; 2000.
- 32. Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, et al. Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. Nat Genet. 2000; 25:25–29. [PubMed: 10802651]
- 33. Timmerman, V. Inherited peripheral neuropathies Mutation database. 2011. http://www.molgen.ua.ac.be/cmtmutations/Home/Default.cfm
- 34. Oshiro C, Marsh S, McLeod H, Carrillo M, Klein T, Altman R. Taxane Pathway. Pharmacogenet Genomics. 2009; 19:979–983. [PubMed: 21151855]
- 35. Smigielski EM, Sirotkin K, Ward M, Sherry ST. dbSNP: a database of single nucleotide polymorphisms. Nucleic Acids Res. 2000; 28:352–355. [PubMed: 10592272]

 Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. Am J Hum Genet. 2007; 81:559–575. [PubMed: 17701901]

- Pruitt KD, Harrow J, Harte RA, Wallin C, Diekhans M, Maglott DR, et al. The consensus coding sequence (CCDS) project: Identifying a common protein-coding gene set for the human and mouse genomes. Genome Res. 2009; 19:1316–1323. [PubMed: 19498102]
- 38. Bush WS, Dudek SM, Ritchie MD. Biofilter: a knowledge-integration system for the multi-locus analysis of genome-wide association studies. Pac Symp Biocomput. 2009:368–379. [PubMed: 19209715]
- 39. Yang J, Lee SH, Goddard ME, Visscher PM. GCTA: a tool for genome-wide complex trait analysis. Am J Hum Genet. 2011; 88:76–82. [PubMed: 21167468]
- Yang J, Manolio TA, Pasquale LR, Boerwinkle E, Caporaso N, Cunningham JM, et al. Genome partitioning of genetic variation for complex traits using common SNPs. Nat Genet. 2011; 43:519– 525. [PubMed: 21552263]
- Chaudhry V, Rowinsky EK, Sartorius SE, Donehower RC, Cornblath DR. Peripheral neuropathy from taxol and cisplatin combination chemotherapy: clinical and electrophysiological studies. Ann Neurol. 1994; 35:304–311. [PubMed: 7907208]
- 42. Letourneau PC, Ressler AH. Inhibition of neurite initiation and growth by taxol. J Cell Biol. 1984; 98:1355–1362. [PubMed: 6143759]
- 43. Melli G, Jack C, Lambrinos GL, Ringkamp M, Hoke A. Erythropoietin protects sensory axons against paclitaxel-induced distal degeneration. Neurobiol Dis. 2006; 24:525–530. [PubMed: 17010629]
- 44. Sahenk Z, Barohn R, New P, Mendell JR. Taxol neuropathy. Electrodiagnostic and sural nerve biopsy findings. Arch Neurol. 1994; 51:726–729. [PubMed: 7912506]
- 45. Roytta M, Raine CS. Taxol-induced neuropathy: chronic effects of local injection. J Neurocytol. 1986; 15:483–496. [PubMed: 2427662]
- Allen E, Ding J, Wang W, Pramanik S, Chou J, Yau V, et al. Gigaxonin-controlled degradation of MAP1B light chain is critical to neuronal survival. Nature. 2005; 438:224–228. [PubMed: 16227972]
- 47. Einarsdottir E, Carlsson A, Minde J, Toolanen G, Svensson O, Solders G, et al. A mutation in the nerve growth factor beta gene (*NGFB*) causes loss of pain perception. Hum Mol Genet. 2004; 13:799–805. [PubMed: 14976160]
- 48. Becker E, Richardson DR. Frataxin: its role in iron metabolism and the pathogenesis of Friedreich's ataxia. Int J Biochem Cell Biol. 2001; 33:1–10. [PubMed: 11167127]
- Bedlack RS, Edelman D, Gibbs JW 3rd, Kelling D, Strittmatter W, Saunders AM, et al. APOE genotype is a risk factor for neuropathy severity in diabetic patients. Neurology. 2003; 60:1022– 1024. [PubMed: 12654974]
- Corder EH, Robertson K, Lannfelt L, Bogdanovic N, Eggertsen G, Wilkins J, et al. HIV-infected subjects with the E4 allele for APOE have excess dementia and peripheral neuropathy. Nat Med. 1998; 4:1182–1184. [PubMed: 9771753]
- 51. Pande M, Hur J, Hong Y, Backus C, Hayes JM, Oh SS, et al. Transcriptional profiling of diabetic neuropathy in the BKS *db/db* mouse: a model of type 2 diabetes. Diabetes. 2011; 60:1981–1989. [PubMed: 21617178]
- 52. Zurowski D, Nowak L, Machowska A, Wordliczek J, Thor PJ. Exogenous melatonin abolishes mechanical allodynia but not thermal hyperalgesia in neuropathic pain. The role of the opioid system and benzodiazepine-gabaergic mechanism. J Physiol Pharmacol. 2012; 63:641–647. [PubMed: 23388480]
- Hori K, Ozaki N, Suzuki S, Sugiura Y. Upregulations of P2X(3) and ASIC3 involve in hyperalgesia induced by cisplatin administration in rats. Pain. 2010; 149:393

  –405. [PubMed: 20378247]
- 54. Padi SS, Shi XQ, Zhao YQ, Ruff MR, Baichoo N, Pert CB, et al. Attenuation of rodent neuropathic pain by an orally active peptide, RAP-103, which potently blocks CCR2- and CCR5-mediated monocyte chemotaxis and inflammation. Pain. 2012; 153:95–106. [PubMed: 22033364]

 Benamar K, Geller EB, Adler MW. First in vivo evidence for a functional interaction between chemokine and cannabinoid systems in the brain. J Pharmacol Exp Ther. 2008; 325:641–645.
 [PubMed: 18281594]

- 56. Li X, Li YH, Yu S, Liu Y. Upregulation of Ryk expression in rat dorsal root ganglia after peripheral nerve injury. Brain Res Bull. 2008; 77:178–184. [PubMed: 18773946]
- 57. Yi XN, Zheng LF, Zhang JW, Zhang LZ, Xu YZ, Luo G, et al. Dynamic changes in Robo2 and Slit1 expression in adult rat dorsal root ganglion and sciatic nerve after peripheral and central axonal injury. Neurosci Res. 2006; 56:314–321. [PubMed: 16979769]
- Gao WQ, Dybdal N, Shinsky N, Murnane A, Schmelzer C, Siegel M, et al. Neurotrophin-3 reverses experimental cisplatin-induced peripheral sensory neuropathy. Ann Neurol. 1995; 38:30– 37. [PubMed: 7611721]
- Apfel SC, Lipton RB, Arezzo JC, Kessler JA. Nerve growth factor prevents toxic neuropathy in mice. Ann Neurol. 1991; 29:87–90. [PubMed: 1705109]
- Peterson ER, Crain SM. Nerve growth factor attenuates neurotoxic effects of taxol on spinal cordganglion explants from fetal mice. Science. 1982; 217:377–379. [PubMed: 6124041]
- 61. Wilson BD, Ii M, Park KW, Suli A, Sorensen LK, Larrieu-Lahargue F, et al. Netrins promote developmental and therapeutic angiogenesis. Science. 2006; 313:640–644. [PubMed: 16809490]
- 62. Toth C, Shim SY, Wang J, Jiang Y, Neumayer G, Belzil C, et al. Ndel1 promotes axon regeneration via intermediate filaments. PLoS One. 2008; 3:e2014. [PubMed: 18431495]
- 63. Hur J, Sullivan KA, Pande M, Hong Y, Sima AA, Jagadish HV, et al. The identification of gene expression profiles associated with progression of human diabetic neuropathy. Brain. 2011; 134:3222–3235. [PubMed: 21926103]
- 64. Lindquist KJ, Jorgenson E, Hoffmann TJ, Witte JS. The impact of improved microarray coverage and larger sample sizes on future genome-wide association studies. Genet Epidemiol. 2013; 37:383–392. [PubMed: 23529720]
- Harris MA, Clark J, Ireland A, Lomax J, Ashburner M, Foulger R, et al. The Gene Ontology (GO) database and informatics resource. Nucleic Acids Res. 2004; 32:D258–D261. [PubMed: 14681407]

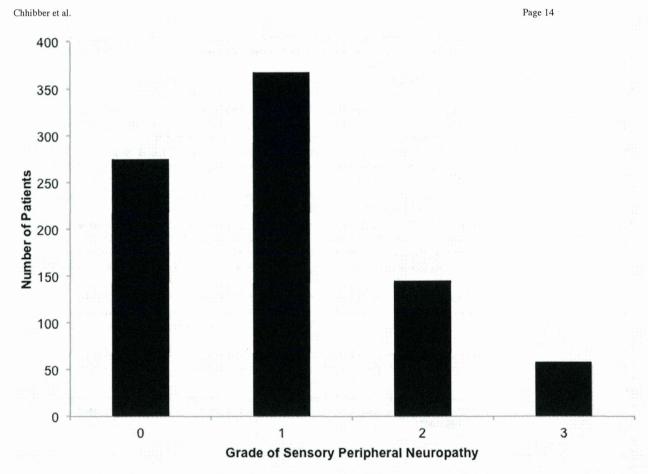


Figure 1. Distribution of sensory peripheral neuropathy in the study population
The distribution of the highest reported grade of sensory peripheral neuropathy is shown for 849 unrelated genetic Europeans from the paclitaxel arm of CALGB 40101. Toxicity is measured using the NCI-CTCAE Scale v2.

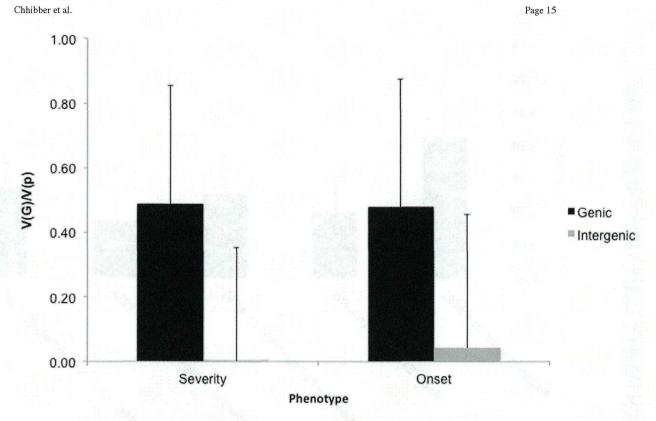


Figure 2. Heritability estimates for severity and onset of paclitaxel-induced sensory peripheral neuropathy for SNPs in genic and intergenic regions

Total genomic variance for both severity and onset of neuropathy was partitioned onto sen

Total genomic variance for both severity and onset of neuropathy was partitioned onto genic and intergenic regions. The error bars denote the SE for the heritability estimates.