

with no major adverse effects, and that PPV treatment resulted in longer survival (MST of 10.1 or 15.2 months) [50, 71]. A clinical study in 10 advanced small cell lung cancer (SCLC) also showed the safety and feasibility of PPV [72].

### 2.3.6. Urothelial Cancer

A phase I clinical trial of PPV was conducted in 10 HLA-A2<sup>+</sup> or HLA-A24<sup>+</sup> refractory urothelial cancer patients [73]. In this study, some patients treated by PPV showed clear clinical responses as evaluated by the Response Evaluation Criteria in Solid Tumors (RECIST) criteria with boosted immune responses: CR in 1, PR in 1, and SD in 2 patients. These 4 responders showed better progression-free survival (MST, 21 months) and overall survival (MST, 24 months), suggesting the potential clinical efficacy of PPV for advanced urothelial cancer.

### 2.3.7. Other Cancers

We also conducted phase I clinical trials for other advanced cancers, including metastatic renal cell carcinoma (RCC) [74], gynecologic cancers [49], and malignant melanoma [51]. All of these studies demonstrated that PPV was safe and well tolerated with no major adverse effects, and that good immune responses to vaccine antigens were induced in many of the patients after PPV. Further clinical trials would be required to clearly prove the clinical benefits of PPV in these cancers.

## 2.4. Biomarkers for PPV (Table 3)

Recent clinical trials of cancer immunotherapies, including peptide-based cancer vaccines, have demonstrated that only a subset of patients show clinical benefits. Furthermore, unexpectedly, some large clinical trials in the past several years have demonstrated that cancer vaccines might sometimes show worse clinical outcomes [75, 76]. It would thus be important to identify predictive biomarkers that could accurately assess anti-tumor immune responses and predict patient prognosis following the administration of cancer vaccines. In some clinical trials, several post-vaccination biomarkers, including CTL responses, Th1 responses, delayed-type hypersensitivity (DTH), and autoimmunity, have been reported to be associated with clinical responses [77-80]. However, there are currently no validated biomarkers for cancer vaccines in widespread use.

To identify biomarkers for PPV, we statistically reviewed 500 advanced cancer patients undergoing PPV from October 2000 to October 2008 [36]. Both lymphocyte counts before vaccination ( $P = 0.0095$ ) and increased IgG response ( $P = 0.0116$ ) to the vaccine peptides after vaccination, along with performance status ( $P < 0.0001$ ), were well correlated with overall survival. In CRPC patients treated with PPV ( $n = 40$ ), a comprehensive study of soluble factors assessed by multiplexed bead array in plasma and gene expression profiles by DNA microarray in PBMC demonstrated that higher IL-6 level and granulocytic myeloid-derived suppressor cells (MDSC) in the peripheral blood before vaccination were closely related to poorer prognosis in the vaccinated patients [81]. By multivariate Cox regression analyses in patients with refractory NSCLC ( $n = 41$ ), higher C-reactive protein (CRP) level before vaccination was a significant predictor of

unfavorable overall survival (HR = 10.115, 95% CI = 2.447 - 41.806,  $P = 0.001$ ) [71]. In addition, in refractory biliary tract cancer patients ( $n = 25$ ), multivariate Cox regression analyses showed that higher IL-6 and lower albumin levels before vaccination were significantly unfavorable factors for overall survival [HR = 1.123, 95% CI = 1.008 - 1.252,  $P = 0.035$ ; HR = 0.158, 95% CI = 0.029 - 0.860,  $P = 0.033$ ; respectively] [66].

Collectively, these findings suggested that less inflammation may contribute to better responses to PPV, indicating that the evaluation of inflammatory factors before vaccination could be useful for selecting cancer patients who are appropriate for PPV (Table 3). An early phase clinical trial is under way to reveal whether or not the blockage of IL-6-mediated inflammatory signaling with a humanized anti-IL-6 receptor monoclonal antibody, tocilizumab, would be beneficial for enhancing the immune and/or clinical responses after PPV in advanced cancer patients who show higher plasma IL-6 levels [82, 83].

## 3. OTHER NEW TYPES OF PEPTIDE VACCINES

Recent early phase clinical trials have also demonstrated significant advances in other types of therapeutic peptide-based vaccines [19, 20]. Several new types of peptide-based vaccines are reviewed in this section (Fig. 1).

### 3.1. Multi-Peptide Vaccine Consisting of CTL and Helper T-Cell Epitopes

Numerous helper T-cell epitopes have been identified from TAA. Since helper T cells are known to play crucial roles in the efficient induction of CTL responses, cancer vaccines, which consist of both HLA class II-restricted helper epitopes recognized by CD4 T cells and class I-restricted CTL epitopes recognized by CD8 T cells, have been developed and clinically tested [84-89]. For example, Kuball *et al.* conducted a phase I study of a multi-peptide vaccine consisting of multiple CTL epitopes from Wilms tumor gene-1 (WT-1), proteinase 3 (Pr3) and mucin 1 (MUC1), and MUC1-helper epitope or pan HLA-DR epitope (PADRE) [84]. Each peptide was formulated separately and injected at a different site. In this study, an increase in PADRE-specific CD4 T cells, which appeared unable to produce IL2, was observed after vaccination, and regulatory T cells were increased, suggesting that helper epitope peptides have the potential to induce not only helper T cells but also regulatory T cells. Krug *et al.* tested the safety and immunogenicity of a WT1 vaccine comprised of four class I and class II-restricted peptides in patients with malignant pleural mesothelioma or NSCLC expressing WT1 [85]. They showed that this multivalent WT1 peptide vaccine induced both CD4 and CD8 T-cell responses in a high proportion of patients with minimal toxicity.

### 3.2. Multi-Peptide Cocktail Vaccine

If each of multiple peptides are formulated separately and injected at a separate site, the number of peptides employed for vaccination might be limited. One strategy for overcoming this limitation is to generate multi-peptide cocktail vaccines, since one preparation could contain more than 10 different peptides. Although the issue of competition between

individual peptides to bind to HLA molecules on the APCs still remains [46], different types of multi-peptide cocktail vaccines have been developed; vaccines consisting of CTL epitope peptides alone [90, 91] or those of both CTL epitope and helper epitope peptides [86-89].

Barve *et al.* conducted a phase I/II study of a multi-peptide cocktail vaccine, IDM-2101, consisting of nine CTL epitope peptides and the PADRE helper epitope peptide with Montanide ISA51 in patients with metastatic non-small cell lung cancer [86]. No significant adverse events were noted except for low-grade erythema and pain at the injection site. One-year survival in the treated patients was 60%, with a median overall survival of 17.3 months. One complete response (CR) patient was observed in the total of 63 patients. Slingsluff *et al.* conducted a multicenter randomized trial to examine the immunogenicity of a multi-peptide cocktail vaccine containing 12 melanoma-associated HLA class I-restricted peptides (12MP) for CD8<sup>+</sup> T cells and tetanus peptide or a mixture of six melanoma-associated helper peptides (6MHP) for CD4<sup>+</sup> T cells in the presence or absence of cyclophosphamide pretreatment in 167 patients with resected stage IIB to IV melanoma [87]. However, the combination of 6MHP with 12MP paradoxically reduced the circulating CD8<sup>+</sup> T-cell response, and cyclophosphamide pretreatment had no measurable effect on CD8<sup>+</sup> or CD4<sup>+</sup> responses. Clinical outcome was not improved by adding melanoma-associated helper peptides or by adding cyclophosphamide.

Rammensee and his colleagues also reported a phase I/II trial of a multi-peptide cocktail vaccine, which consisted of 13 synthetic peptides (11 HLA-A\*0201-restricted CTL epitopes and 2 helper epitopes derived from prostate tumor antigens) for 19 HLA-A2<sup>+</sup> hormone-sensitive prostate cancer patients with biochemical recurrence after primary surgical treatment [88]. The vaccine was well tolerated, and stabilized or slowed down PSA progress in 4 of the 19 patients. The same group also developed another cocktail vaccine, IMA901, which consisted of nine HLA-A\*0201-restricted CTL epitopes and one helper epitope from renal cell cancer antigens with hepatitis B virus epitope as a marker peptide, for advanced renal cell cancer [89]. In a randomized phase II trial with a single dose of cyclophosphamide, the number of regulatory T cells was reduced, and immune responses to the vaccine peptides were associated with longer overall survival. A randomized phase III study to determine the clinical benefit of IMA901 is ongoing.

### 3.3. Hybrid Peptide Vaccine

Peptides used in most clinical trials for peptide-based vaccines possess native amino acid sequences with or without slight modification in anchor amino acids to increase their binding capability to HLA molecules. However, hybrid-type peptide vaccines, which use a new artificial peptide fusing two or more peptides, have been devised. For example, the Ii-Key/HER-2/neu hybrid peptide vaccine, a fusion peptide made up of the Ii-Key 4-mer peptide and HER-2/neu (776-790) helper epitope peptide, has been reported [92, 93]. The Ii/Key 4-mer peptide is the shortest active sequence of the Ii protein, which catalyzes direct charging of MHC class II epitopes to the peptide-binding groove, circumventing the need for intracellular epitope processing [94]. Phase I studies

of the Ii-Key/HER-2/neu hybrid peptide vaccine in patients with prostate cancer showed that this vaccine is safe and can induce HER-2/neu-specific cellular immune responses in vaccinated patients [93]. In addition, significant decreases in circulating regulatory T-cell frequencies, plasma HER2/neu, and serum TGF-beta levels were observed.

Nishimura *et al.* reported an artificially synthesized helper/killer-hybrid epitope long peptide (H/K-HELP) of MAGE-A4 cancer antigen [95]. In the first case report, a patient with pulmonary metastasis of colon cancer was vaccinated with MAGE-A4-H/K-HELP in combination with OK432 and Montanide ISA51. There were no severe side effects except for a skin reaction at the injection site. Vaccination with MAGE-A4-H/K-HELP induced MAGE-A4-specific Th1 and Tc1 immune responses and the production of MAGE-A4-specific complement-fixing IgG antibodies. Tumor growth and tumor markers were significantly decreased in this patient.

### 3.4. Long Peptide Vaccine

The classical types of peptide vaccines have consisted of short epitope peptides with minimal optimal lengths, which are recognized by CTLs or helper T cells in an HLA class I- or class II-restricted manner, respectively. However, direct binding of short peptides to nonspecific cells without a costimulatory capacity has been reported to bear the potential to induce tolerance to antigen-specific T cells rather than to induce their activation in some mouse models [39-41]. Therefore, a novel approach using synthetic long peptides, which need to be taken up by professional APCs and processed for presentation by HLA class I and/or class II molecules, has been developed for cancer vaccination, although the efficiency and mechanisms of presentation of exogenous long peptides in human HLA class I remain to be fully elucidated [96]. Synthetic long peptides may contain not only HLA class I-restricted but also HLA class II-restricted epitopes, which can activate helper T cells important for the efficient induction of antigen-specific CTL responses.

Several clinical studies using a pool of multiple synthetic long peptides have been reported, since a mixture of multiple synthetic long peptides is likely to contain multiple HLA class I-restricted and class II-restricted T-cell epitopes, which could be applicable to any patients irrespective of their HLA types [42-45, 97-100]. Melief and his colleagues showed that a vaccine composed of a synthetic long peptide pool derived from high-risk-type human papillomavirus (HPV)-16 E6/E7 oncoproteins successfully induced HPV-specific immune responses [42, 43]. They conducted a phase I study of HPV16 E6 and E7 overlapping long peptides in end-stage cervical cancer patients [42]. Cocktails of nine E6 peptides and/or four E7 peptides covering the entire sequences of E6 and E7 proteins showed a strong and broad T-cell response dominated by immunity against E6 after four subcutaneous administrations with Montanide ISA51 at 3-week intervals. Subsequently, they conducted a phase II study of the same vaccine in patients with HPV-positive grade 3 vulvar intraepithelial neoplasia, which is a chronic disorder caused by HPV [43]. At 3 months after the last vaccination, 12 of 20 patients (60%) had clinical responses and reported relief of symptoms. Five women had complete regression of the lesions. At 12 months of follow-up, 15 of 19

patients (79%) had clinical responses, with a complete response in 9 of 19 patients (47%).

The same group also reported a synthetic long peptide vaccine targeted for p53. This p53 synthetic long peptide vaccine (p53-SLP) consisted of 10 synthetic 25-mer to 30-mer long overlapping peptides, spanning amino acids 70–248 of the wild-type p53 protein. In a phase I/II trial of the p53-SLP vaccine in 10 patients with metastatic colorectal cancer, p53-specific T-cell responses were induced in 9 of 10 patients as measured by IFN- $\gamma$  enzyme-linked immunospot (ELISPOT), proliferation, and cytokine bead arrays [97]. Subsequently, a phase II study of the same vaccine in 20 ovarian cancer patients with recurrent elevation of CA-125 showed that SD, as determined by CA-125 levels and CT scans, was observed in 2 out of 20 patients (10%) as the best clinical response, but no relationship was found between the clinical response and vaccine-induced immunity [44]. IFN- $\gamma$ -producing p53-specific responses were induced in CD4 T cells, but not in CD8 T cells, in all patients who received four immunizations. The absence of p53-specific CD8 T-cell responses might be attributable to the dominant production of Th2 cytokines by CD4 T cells, which have inhibitory effects on CTL induction. Nevertheless, the combined use of p53-SLP vaccine and a low dose of cyclophosphamide or IFN- $\alpha$  has recently been reported to efficiently induce more IFN- $\gamma$ -producing p53-specific T cells, suggesting that these combinations may potentiate the immunogenicity of the p53-SLP vaccine [98, 99].

Kakimi *et al.* also conducted a phase I trial of an NY-ESO-1 synthetic long peptide vaccine. A 20-mer peptide spanning from amino acid 91 to 110 of NY-ESO-1, called NY-ESO-1f, which includes multiple epitopes recognized by antibodies and CD4 and CD8 T cells, was administered along with OK-432 and Montanide ISA51 to patients with advanced cancers [100]. Both antigen-specific CD4 and CD8 T-cell responses, as well as antibody responses, were increased in 9 of 10 patients.

### 3.5. Novel Approach for Targeting Peptides to Professional APCs

The goal of cancer immunotherapy is to induce and amplify functional antigen-specific immune responses in order to develop long-lasting immunological memory specific to tumor cells [101, 102]. However, one hurdle to the use of peptide-based vaccines is that the uptake and/or presentation of vaccine peptides by nonspecific cells, but not by professional APCs, leads to CTL anergy through insufficient stimulation [103]. For efficient priming and activation of antigen-specific CTL through vaccination, sufficient amounts of antigens should be presented to T cells by functionally activated, professional APCs for sufficient periods of time [104–107]. In this respect, a novel delivery system for peptide vaccines remains to be developed.

For example, nanotechnology-based antigen delivery has been developing as a vaccine strategy due to its dose-sparing and prolonged antigen presentation features [108, 109]. In particular, polymeric nanoparticles (NP) have attracted increasing attention as carriers of therapeutic immunogens [110]. Antigen peptides encapsulated in polymeric NP are shown to be directly and specifically delivered to profes-

sional APCs via phagocytosis without proteolytic degradation, and efficiently cross-presented to induce strong T-cell immunity, whereas those in solution that are internalized by APCs via macropinocytosis are reported to be poorly presented as peptides in complex with MHC class I molecules on cell surfaces [111, 112]. Indeed, we have demonstrated the feasibility of NP consisting of a biodegradable, biocompatible copolymer, poly(D,L-lactide-co-glycolide) (PLGA) carrying antigenic peptides and a toll-like receptor 4 agonist, monophosphoryl lipid A, to efficiently induce CTL responses against TAA in murine tumor models [113]. To increase the efficacy of peptide-based vaccines, such a novel antigen delivery system remains to be developed and clinically examined.

### CONCLUSIONS

In the field of cancer immunology and immunotherapy, excitement and enthusiasm have risen around the latest approvals of immunotherapy-based treatments in various cancer types. However, several issues remain to be addressed in order to achieve further development of cancer vaccines. In particular, in view of the complexity and diversity of tumor cell characteristics and host immune cell repertoires, the selection of vaccine peptides appropriate for individual patients based on the pre-existing host immunity before vaccination could be critical for the efficient induction of beneficial antitumor responses in cancer patients. In a series of clinical trials, we have demonstrated promising results of PPV as a new treatment modality for patients with various types of advanced cancer. Further randomized phase III clinical trials are essential to validate the clinical benefits of PPV. Moreover, novel biomarkers for selecting patients who would benefit most from PPV remain to be addressed.

### CONFLICT OF INTEREST

Akira Yamada is an Executive Officer for Green Peptide Company, Ltd. Kyogo Itoh received a research grant from the Green Peptide Company, Ltd. and owns stock in the Green Peptide Company, Ltd.

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### REFERENCES

- [1] Finn, O.J. Cancer immunology. *N. Engl. J. Med.*, **2008**, *358*(25), 2704–2715.
- [2] Rosenberg, S.A. Cell transfer immunotherapy for metastatic solid cancer—what clinicians need to know. *Nat. Rev. Clin. Oncol.*, **2011**, *8*(10), 577–585.
- [3] Mellman, I.; Coukos, G.; Dranoff, G. Cancer immunotherapy comes of age. *Nature*, **2011**, *480*(7378), 480–489.
- [4] Schlom, J. Therapeutic cancer vaccines: current status and moving forward. *J. Natl. Cancer Inst.*, **2012**, *104*(8), 599–613.
- [5] Itoh, K.; Yamada, A.; Mine, T.; Noguchi, M. Recent advances in cancer vaccines: an overview. *Jpn. J. Clin. Oncol.*, **2009**, *39*(2), 73–80.
- [6] Kantoff, P.W.; Higano, C.S.; Shore, N.D.; Berger, E.R.; Small, E.J.; Penson, D.F.; Redfern, C.H.; Ferrari, A.C.; Dreicer, R.; Sims, R.B.; Xu, Y.; Frohlich, M.W.; Schellhammer, P.F. IMPACT Study

- Investigators. Sipuleucel-T immunotherapy for castration-resistant prostate cancer. *N. Engl. J. Med.*, **2010**, *363*(5), 411-422.
- [7] Hodi, F.S.; O'Day, S.J.; McDermott, D.F.; Weber, R.W.; Sosman, J.A.; Haanen, J.B.; Gonzalez, R.; Robert, C.; Schadendorf, D.; Hassel, J.C.; Akerley, W.; van den Eertwegh, A.J.; Lutzky, J.; Lorigan, P.; Vaubel, J.M.; Linette, G.P.; Hogg, D.; Ottensmeier, C.H.; Lebbé, C.; Peschel, C.; Quirt, I.; Clark, J.I.; Wolchok, J.D.; Weber, J.S.; Tian, J.; Yellin, M.J.; Nichol, G.M.; Hoos, A.; Urban, W.J. Improved survival with ipilimumab in patients with metastatic melanoma. *N. Engl. J. Med.*, **2010**, *363*(8), 711-723.
- [8] Topalian, S.L.; Hodi, F.S.; Brahmer, J.R.; Gettinger, S.N.; Smith, D.C.; McDermott, D.F.; Powderly, J.D.; Carvajal, R.D.; Sosman, J.A.; Atkins, M.B.; Leming, P.D.; Spigel, D.R.; Antonia, S.J.; Horn, L.; Drake, C.G.; Pardoll, D.M.; Chen, L.; Sharfman, W.H.; Anders, R.A.; Taube, J.M.; McMiller, T.L.; Xu, H.; Korman, A.J.; Jure-Kunkel, M.; Agrawal, S.; McDonald, D.; Kollia, G.D.; Gupta, A.; Wigginton, J.M.; Sznol, M. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N. Engl. J. Med.*, **2012**, *366*(26), 2443-2454.
- [9] Brahmer, J.R.; Tykodi, S.S.; Chow, L.Q.; Hwu, W.J.; Topalian, S.L.; Hwu, P.; Drake, C.G.; Camacho, L.H.; Kauh, J.; Odunsi, K.; Pitot, H.C.; Hamid, O.; Bhatia, S.; Martins, R.; Eaton, K.; Chen, S.; Salay, T.M.; Alaparthi, S.; Grosso, J.F.; Korman, A.J.; Parker, S.M.; Agrawal, S.; Goldberg, S.M.; Pardoll, D.M.; Gupta, A.; Wigginton, J.M. Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. *N. Engl. J. Med.*, **2012**, *366*(26), 2455-2465.
- [10] van der Bruggen, P.; Traversari, C.; Chomez, P.; Lurquin, C.; De Plaen, E.; Van den Eynde, B.; Knuth, A.; Boon, T. A gene encoding an antigen recognized by cytolytic T lymphocytes on a human melanoma. *Science*, **1991**, *254*(5038), 1643-1647.
- [11] Türeci, O.; Sahin, U.; Schobert, I.; Koslowski, M.; Schmitt, H.; Schild, H.J.; Stenner, F.; Seitz, G.; Rammensee, H.G.; Pfreundschuh, M. The SSX-2 gene, which is involved in the t(X;18) translocation of synovial sarcomas, codes for the human tumor antigen HOM-MEL-40. *Cancer Res.*, **1996**, *56*(20), 4766-4772.
- [12] Cheever, M.A.; Allison, J.P.; Ferris, A.S.; Finn, O.J.; Hastings, B.M.; Hecht, T.T.; Mellman, I.; Prindiville, S.A.; Viner, J.L.; Weiner, L.M.; Matrisian, L.M. The prioritization of cancer antigens: a national cancer institute pilot project for the acceleration of translational research. *Clin. Cancer Res.*, **2009**, *15*(17), 5323-5337.
- [13] Singh-Jasuja, H.; Emmerich, N.P.; Rammensee, H.G. The Tübingen approach: identification, selection, and validation of tumor-associated HLA peptides for cancer therapy. *Cancer Immunol. Immunother.*, **2004**, *53*(3), 187-195.
- [14] Kessler, J.H.; Melief, C.J. Identification of T-cell epitopes for cancer immunotherapy. *Leukemia*, **2007**, *21*(9), 1859-1874.
- [15] Hu, X.; Chakraborty, N.G.; Sporn, J.R.; Kurtzman, S.H.; Ergin, M.T.; Mukherji, B. Enhancement of cytolytic T lymphocyte precursor frequency in melanoma patients following immunization with the MAGE-1 peptide loaded antigen presenting cell-based vaccine. *Cancer Res.*, **1996**, *56*(11), 2479-2483.
- [16] Rosenberg, S.A.; Yang, J.C.; Restifo, N.P. Cancer immunotherapy: moving beyond current vaccines. *Nat. Med.*, **2004**, *10*(9), 909-915.
- [17] Purcell, A.W.; McCluskey, J.; Rossjohn, J. More than one reason to rethink the use of peptides in vaccine design. *Nat. Rev. Drug Discov.*, **2007**, *6*(5), 404-414.
- [18] Schwartzentruber, D.J.; Lawson, D.H.; Richards, J.M.; Conry, R.M.; Miller, D.M.; Treisman, J.; Gailani, F.; Riley, L.; Conlon, K.; Pockaj, B.; Kendra, K.L.; White, R.L.; Gonzalez, R.; Kuzel, T.M.; Curti, B.; Leming, P.D.; Whitman, E.D.; Balkissoon, J.; Reintgen, D.S.; Kaufman, H.; Marincola, F.M.; Merino, M.J.; Rosenberg, S.A.; Choyke, P.; Vena, D.; Hwu, P. gp100 peptide vaccine and interleukin-2 in patients with advanced melanoma. *N. Engl. J. Med.*, **2011**, *364*(22), 2119-2127.
- [19] Yamada, A.; Sasada, T.; Noguchi, M.; Itoh, K. Next-generation peptide vaccines for advanced cancer. *Cancer Sci.*, **2013**, *104*(1), 15-21.
- [20] Perez, S.A.; von Hofe, E.; Kallinteris, N.L.; Gritzapis, A.D.; Peoples, G.E.; Papanichail, M.; Baxevasis, C.N. A new era in anticancer peptide vaccines. *Cancer*, **2010**, *116*(9), 2071-2080.
- [21] Schreiber, R.D.; Old, L.J.; Smyth, M.J. Cancer immunoeediting: integrating immunity's roles in cancer suppression and promotion. *Science*, **2011**, *331*(6024), 1565-1570.
- [22] Bei, R.; Scardino, A. TAA polyepitope DNA-based vaccines: a potential tool for cancer therapy. *J. Biomed. Biotechnol.*, **2010**, *2010*, 102758.
- [23] Yewdell, J.W.; Bennink, J.R. Immunodominance in major histocompatibility complex class I-restricted T lymphocyte responses. *Annu. Rev. Immunol.*, **1999**, *17*, 51-88.
- [24] Chen, W.; McCluskey, J. Immunodominance and immunodomination: critical factors in developing effective CD8+ T-cell-based cancer vaccines. *Adv. Cancer Res.*, **2006**, *95*, 203-247.
- [25] Itoh, K.; Yamada, A. Personalized peptide vaccines: a new therapeutic modality for cancer. *Cancer Sci.*, **2006**, *97*(10), 970-976.
- [26] Mochizuki, K.; Sato, Y.; Tsuda, N.; Shomura, H.; Sakamoto, M.; Matsuura, K.; Ushijima, K.; Macda, Y.; Katagiri, K.; Yamada, A.; Todo, S.; Kamura, T.; Harada, M.; Itoh, K. Immunological evaluation of vaccination with pre-designated peptides frequently selected as vaccine candidates in an individualized peptide vaccination regimen. *Int. J. Oncol.*, **2004**, *25*(1), 121-131.
- [27] Pilla, L.; Rivoltini, L.; Patuzzo, R.; Marrari, A.; Valdagni, R.; Parmiani, G. Multi-peptide vaccination in cancer patients. *Expert Opin. Biol. Ther.*, **2009**, *9*(8), 1043-1055.
- [28] Nakao, M.; Shichijo, S.; Imaizumi, T.; Inoue, Y.; Matsunaga, K.; Yamada, A.; Kikuchi, M.; Tsuda, N.; Ohta, K.; Takamori, S.; Yamana, H.; Fujita, H.; Itoh, K. Identification of a gene coding for a new squamous cell carcinoma antigen recognized by the CTL. *J. Immunol.*, **2000**, *164*(5), 2565-2574.
- [29] Shichijo, S.; Nakao, M.; Imai, Y.; Takasu, H.; Kawamoto, M.; Niiya, F.; Yang, D.; Toh, Y.; Yamana, H.; Itoh, K. A gene encoding antigenic peptides of human squamous cell carcinoma recognized by cytotoxic T lymphocytes. *J. Exp. Med.*, **1998**, *187*(3), 277-288.
- [30] Harashima, N.; Tanaka, K.; Sasatomi, T.; Shimizu, K.; Miyagi, Y.; Yamada, A.; Tamura, M.; Yamana, H.; Itoh, K.; Shichijo, S. Recognition of the Lck tyrosine kinase as a tumor antigen by cytotoxic T lymphocytes of cancer patients with distant metastases. *Eur. J. Immunol.*, **2001**, *31*(2), 323-332.
- [31] Yamada, A.; Kawano, K.; Koga, M.; Matsumoto, T.; Itoh, K. Multidrug resistance-associated protein 3 is a tumor rejection antigen recognized by HLA-A2402-restricted cytotoxic T lymphocytes. *Cancer Res.*, **2001**, *61*(17), 6459-6466.
- [32] Harada, M.; Kobayashi, K.; Matsueda, S.; Nakagawa, M.; Noguchi, M.; Itoh, K. Prostate-specific antigen-derived epitopes capable of inducing cellular and humoral responses in HLA-A24+ prostate cancer patients. *Prostate*, **2003**, *57*(2), 152-159.
- [33] Inoue, Y.; Takae, Y.; Takei, M.; Kato, K.; Kanai, S.; Harada, Y.; Tobisu, K.; Noguchi, M.; Kakizoe, T.; Itoh, K.; Wakasugi, H. Induction of tumor specific cytotoxic T lymphocytes in prostate cancer using prostatic acid phosphatase derived HLA-A2402 binding peptide. *J. Urol.*, **2001**, *166*(4), 1508-1513.
- [34] Kobayashi, K.; Noguchi, M.; Itoh, K.; Harada, M. Identification of a prostate-specific membrane antigen-derived peptide capable of eliciting both cellular and humoral immune responses in HLA-A24+ prostate cancer patients. *Cancer Sci.*, **2003**, *94*(7), 622-627.
- [35] Yoshida, K.; Noguchi, M.; Mine, T.; Komatsu, N.; Yutani, S.; Ueno, T.; Yanagimoto, H.; Kawano, K.; Itoh, K.; Yamada, A. Characteristics of severe adverse events after peptide vaccination for advanced cancer patients: Analysis of 500 cases. *Oncol. Rep.*, **2011**, *25*(1), 57-62.
- [36] Noguchi, M.; Mine, T.; Komatsu, N.; Suekane, S.; Moriya, F.; Matsuoka, K.; Yutani, S.; Shichijo, S.; Yamada, A.; Toh, U.; Kawano, K.; Azuma, K.; Uemura, H.; Okuno, K.; Matsumoto, K.; Yanagimoto, H.; Yamanaka, R.; Oka, M.; Todo, S.; Sasada, T.; Itoh, K. Assessment of immunological biomarkers in patients with advanced cancer treated by personalized peptide vaccination. *Cancer Biol. Ther.*, **2011**, *10*(12), 1266-1279.
- [37] Avogadri, F.; Merghoub, T.; Maughan, M.F.; Hirschhorn-Cymerman, D.; Morris, J.; Ritter, E.; Olmsted, R.; Houghton, A.N.; Wolchok, J.D. Alphavirus replicon particles expressing TRP-2 provide potent therapeutic effect on melanoma through activation of humoral and cellular immunity. *PLoS One*, **2010**, *5*(9), e12670.
- [38] Hong, S.; Qian, J.; Li, H.; Yang, J.; Lu, Y.; Zheng, Y.; Yi, Q. CpG or IFN- $\alpha$  are more potent adjuvants than GM-CSF to promote anti-tumor immunity following idiotype vaccine in multiple myeloma. *Cancer Immunol. Immunother.*, **2012**, *61*(4), 561-571.

- [39] Melief, C.J.; van der Burg, S.H. Immunotherapy of established (pre)malignant disease by synthetic long peptide vaccines. *Nat. Rev. Cancer*, **2008**, *8*(5), 351-360.
- [40] Bijker, M.S.; van den Eeden, S.J.; Franken, K.L.; Melief, C.J.; Offringa, R.; van der Burg, S.H. CD8+ CTL priming by exact peptide epitopes in incomplete Freund's adjuvant induces a vanishing CTL response, whereas long peptides induce sustained CTL reactivity. *J. Immunol.*, **2007**, *179*(8), 5033-5040.
- [41] Toes, R.E.; Offringa, R.; Blom, R.J.; Melief, C.J.; Kast, W.M. Peptide vaccination can lead to enhanced tumor growth through specific T-cell tolerance induction. *Proc. Natl. Acad. Sci. USA*, **1996**, *93*(15), 7855-7860.
- [42] Kenter, G.G.; Welters, M.J.; Valentijn, A.R.; Lowik, M.J.; Berends-van der Meer, D.M.; Vloon, A.P.; Drijfhout, J.W.; Wafelman, A.R.; Oostendorp, J.; Fleuren, G.J.; Offringa, R.; van der Burg, S.H.; Melief, C.J. Phase I immunotherapeutic trial with long peptides spanning the E6 and E7 sequences of high-risk human papillomavirus 16 in end-stage cervical cancer patients shows low toxicity and robust immunogenicity. *Clin. Cancer Res.*, **2008**, *14*(1), 169-177.
- [43] Kenter, G.G.; Welters, M.J.; Valentijn, A.R.; Lowik, M.J.; Berends-van der Meer, D.M.; Vloon, A.P.; Essahsah, F.; Fathers, L.M.; Offringa, R.; Drijfhout, J.W.; Wafelman, A.R.; Oostendorp, J.; Fleuren, G.J.; van der Burg, S.H.; Melief, C.J. Vaccination against HPV-16 oncoproteins for vulvar intraepithelial neoplasia. *N. Engl. J. Med.*, **2009**, *361*(19), 1838-1847.
- [44] Leffers, N.; Lambeck, A.J.; Gooden, M.J.; Hoogeboom, B.N.; Wolf, R.; Hamming, I.E.; Hepkema, B.G.; Willemse, P.H.; Molmans, B.H.; Hollema, H.; Drijfhout, J.W.; Shuiter, W.J.; Valentijn, A.R.; Fathers, L.M.; Oostendorp, J.; van der Zee, A.G.; Melief, C.J.; van der Burg, S.H.; Daemen, T.; Nijman, H.W. Immunization with a P53 synthetic long peptide vaccine induces P53-specific immune responses in ovarian cancer patients, a phase II trial. *Int. J. Cancer*, **2009**, *125*(9), 2104-2113.
- [45] Welters, M.J.; Kenter, G.G.; Piersma, S.J.; Vloon, A.P.; Löwik, M.J.; Berends-van der Meer, D.M.; Drijfhout, J.W.; Valentijn, A.R.; Wafelman, A.R.; Oostendorp, J.; Fleuren, G.J.; Offringa, R.; Melief, C.J.; van der Burg, S.H. Induction of tumor-specific CD4+ and CD8+ T-cell immunity in cervical cancer patients by a human papillomavirus type 16 E6 and E7 long peptides vaccine. *Clin. Cancer Res.*, **2008**, *14*(1), 178-187.
- [46] Rosenberg, S.A.; Sherry, R.M.; Morton, K.E.; Yang, J.C.; Topalian, S.L.; Royal, R.E.; Kammula, U.S.; Restifo, N.P.; Hughes, M.S.; Schwarz, S.L.; Ngo, L.T.; Mavroukakis, S.A.; White, D.E. Altered CD8(+) T-cell responses when immunizing with multiepitope peptide vaccines. *J. Immunother.*, **2006**, *29*(2), 224-231.
- [47] Yajima, N.; Yamanaka, R.; Mine, T.; Tsuchiya, N.; Homma, J.; Sano, M.; Kuramoto, T.; Obata, Y.; Komatsu, N.; Arima, Y.; Yamada, A.; Shigemori, M.; Itoh, K.; Tanaka, R. Immunologic evaluation of personalized peptide vaccination for patients with advanced malignant glioma. *Clin. Cancer Res.*, **2005**, *11*(16), 5900-5911.
- [48] Hida, N.; Maeda, Y.; Katagiri, K.; Takasu, H.; Harada, M.; Itoh, K. A simple culture protocol to detect peptide-specific cytotoxic T lymphocyte precursors in the circulation. *Cancer Immunol. Immunother.*, **2002**, *51*(4), 219-228.
- [49] Tsuda, N.; Mochizuki, K.; Harada, M.; Sukehiro, A.; Kawano, K.; Yamada, A.; Ushijima, K.; Sugiyama, T.; Nishida, T.; Yamana, H.; Itoh, K.; Kamura, T. Vaccination with predesignated or evidence-based peptides for patients with recurrent gynecologic cancers. *J. Immunother.*, **2004**, *27*(1), 60-72.
- [50] Mine, T.; Gouhara, R.; Hida, N.; Imai, N.; Azuma, K.; Rikimaru, T.; Katagiri, K.; Nishikori, M.; Sukehiro, A.; Nakagawa, M.; Yamada, A.; Aizawa, H.; Shirouzu, K.; Itoh, K.; Yamana, H. Immunological evaluation of CTL precursor-oriented vaccines for advanced lung cancer patients. *Cancer Sci.*, **2003**, *94*(6), 548-556.
- [51] Tanaka, S.; Harada, M.; Mine, T.; Noguchi, M.; Gohara, R.; Azuma, K.; Tamura, M.; Yamada, A.; Morinaga, A.; Nishikori, M.; Katagiri, K.; Itoh, K.; Yamana, H.; Hashimoto, T. Peptide vaccination for patients with melanoma and other types of cancer based on pre-existing peptide-specific cytotoxic T-lymphocyte precursors in the periphery. *J. Immunother.*, **2003**, *26*(4), 357-366.
- [52] Sharma, P.; Wagner, K.; Wolchok, J.D.; Allison, J.P. Novel cancer immunotherapy agents with survival benefit: recent successes and next steps. *Nat. Rev. Cancer*, **2011**, *11*(11), 805-812.
- [53] Whiteside, T.L. Immune monitoring of clinical trials with biotherapies. *Adv. Clin. Chem.*, **2008**, *45*, 75-97.
- [54] Komatsu, N.; Shichijo, S.; Nakagawa, M.; Itoh, K. New multiplexed flow cytometric assay to measure anti-peptide antibody: a novel tool for monitoring immune responses to peptides used for immunization. *Scand. J. Clin. Lab. Invest.*, **2004**, *64*(6), 535-545.
- [55] Mine, T.; Sato, Y.; Noguchi, M.; Sasatomi, T.; Gouhara, R.; Tsuda, N.; Tanaka, S.; Shomura, H.; Katagiri, K.; Rikimaru, T.; Shichijo, S.; Kamura, T.; Hashimoto, T.; Shirouzu, K.; Yamada, A.; Todo, S.; Itoh, K.; Yamana, H. Humoral responses to peptides correlate with overall survival in advanced cancer patients vaccinated with peptides based on pre-existing, peptide-specific cellular responses. *Clin. Cancer Res.*, **2004**, *10*(3), 929-937.
- [56] Noguchi, M.; Uemura, H.; Naito, S.; Akaza, H.; Yamada, A.; Itoh, K. A phase I study of personalized peptide vaccination using 14 kinds of vaccine in combination with low-dose estramustine in HLA-A24-positive patients with castration-resistant prostate cancer. *Prostate*, **2011**, *71*(5), 470-479.
- [57] Terasaki, M.; Shibui, S.; Narita, Y.; Fujimaki, T.; Aoki, T.; Kajiwara, K.; Sawamura, Y.; Kurisu, K.; Mineta, T.; Yamada, A.; Itoh, K. Phase I trial of a personalized peptide vaccine for patients positive for human leukocyte antigen-A24 with recurrent or progressive glioblastoma multiforme. *J. Clin. Oncol.*, **2011**, *29*(3), 337-344.
- [58] Noguchi, M.; Kobayashi, K.; Suetsugu, N.; Tomiyasu, K.; Suekane, S.; Yamada, A.; Itoh, K.; Noda, S. Induction of cellular and humoral immune responses to tumor cells and peptides in HLA-A24 positive hormone-refractory prostate cancer patients by peptide vaccination. *Prostate*, **2003**, *57*(1), 80-92.
- [59] Noguchi, M.; Itoh, K.; Suekane, S.; Yao, A.; Suetsugu, N.; Katagiri, K.; Yamada, A.; Yamana, H.; Noda, S. Phase I trial of patient-oriented vaccination in HLA-A2-positive patients with metastatic hormone-refractory prostate cancer. *Cancer Sci.*, **2004**, *95*(1), 77-84.
- [60] Noguchi, M.; Mine, T.; Yamada, A.; Obata, Y.; Yoshida, K.; Mizoguchi, J.; Harada, M.; Suekane, S.; Itoh, K.; Matsuoka, K. Combination therapy of personalized peptide vaccination and low-dose estramustine phosphate for metastatic hormone refractory prostate cancer patients: an analysis of prognostic factors in the treatment. *Oncol. Res.*, **2007**, *16*(7), 341-349.
- [61] Noguchi, M.; Kakuma, T.; Uemura, H.; Nasu, Y.; Kumon, H.; Hirao, Y.; Moriya, F.; Suekane, S.; Matsuoka, K.; Komatsu, N.; Shichijo, S.; Yamada, A.; Itoh, K. A randomized phase II trial of personalized peptide vaccine plus low dose estramustine phosphate (EMP) versus standard dose EMP in patients with castration resistant prostate cancer. *Cancer Immunol. Immunother.*, **2010**, *59*(7), 1001-1009.
- [62] Noguchi, M.; Moriya, F.; Suekane, S.; Matsuoka, K.; Arai, G.; Matsueda, S.; Sasada, T.; Yamada, A.; Itoh, K. Phase II study of personalized peptide vaccination for castration-resistant prostate cancer patients who failed in docetaxel-based chemotherapy. *Prostate*, **2012**, *72*(8), 834-845.
- [63] Yanagimoto, H.; Mine, T.; Yamamoto, K.; Sato, S.; Terakawa, N.; Takahashi, K.; Nakahara, K.; Honma, S.; Tanaka, M.; Mizoguchi, J.; Yamada, A.; Oka, M.; Kamiyama, Y.; Itoh, K.; Takai, S. Immunological evaluation of personalized peptide vaccination with gemcitabine for pancreatic cancer. *Cancer Sci.*, **2007**, *98*(4), 605-611.
- [64] Yanagimoto, H.; Shiomi, H.; Sato, S.; Mine, T.; Toyokawa, H.; Yamamoto, T.; Tani, T.; Yamada, A.; Kwon, A.H.; Komatsu, N.; Itoh, K.; Noguchi, M. A phase II study of personalized peptide vaccination combined with gemcitabine for non-resectable pancreatic cancer patients. *Oncol. Rep.*, **2010**, *24*(3), 795-801.
- [65] Burris, H.A.; Moore, M.J.; Andersen, J.; Green, M.R.; Rothenberg, M.L.; Modiano, M.R.; Cripps, M.C.; Portenoy, R.K.; Storniolo, A.M.; Tarassoff, P.; Nelson, R.; Dorr, F.A.; Stephens, C.D.; Von Hoff, D.D. Improvements in survival and clinical benefit with gemcitabine as first-line therapy for patients with advanced pancreas cancer: a randomized trial. *J. Clin. Oncol.*, **1997**, *15*(6), 2403-2413.
- [66] Yoshitomi, M.; Yutani, S.; Matsueda, S.; Ioji, T.; Komatsu, N.; Shichijo, S.; Yamada, A.; Itoh, K.; Sasada, T.; Kinoshita, H. Personalized peptide vaccination for advanced biliary tract cancer: IL-6, nutritional status and pre-existing antigen-specific immunity

- as possible biomarkers for patient prognosis. *Exp. Ther. Med.*, **2012**, *3*(3), 463-469.
- [67] Sato, Y.; Shomura, H.; Maeda, Y.; Mine, T.; Une, Y.; Akasaka, Y.; Kondo, M.; Takahashi, S.; Shinohara, T.; Katagiri, K.; Sato, M.; Okada, S.; Matsui, K.; Yamada, A.; Yamana, H.; Itoh, K.; Todo, S. Immunological evaluation of peptide vaccination for patients with gastric cancer based on pre-existing cellular response to peptide. *Cancer Sci.*, **2003**, *94*(9), 802-808.
- [68] Sato, Y.; Maeda, Y.; Shomura, H.; Sasatomi, T.; Takahashi, M.; Une, Y.; Kondo, M.; Shinohara, T.; Hida, N.; Katagiri, K.; Sato, K.; Sato, M.; Yamada, A.; Yamana, H.; Harada, M.; Itoh, K.; Todo, S. A phase I trial of cytotoxic T-lymphocyte precursor-oriented peptide vaccines for colorectal carcinoma patients. *Br. J. Cancer*, **2004**, *90*(7), 1334-1342.
- [69] Sato, Y.; Fujiwara, T.; Mine, T.; Shomura, H.; Homma, S.; Maeda, Y.; Tokunaga, N.; Ikeda, Y.; Ishihara, Y.; Yamada, A.; Tanaka, N.; Itoh, K.; Harada, M.; Todo, S. Immunological evaluation of personalized peptide vaccination in combination with a 5-fluorouracil derivative (TS-1) for advanced gastric or colorectal carcinoma patients. *Cancer Sci.*, **2007**, *98*(7), 1113-1119.
- [70] Hattori, T.; Mine, T.; Komatsu, N.; Yamada, A.; Itoh, K.; Shiozaki, H.; Okuno, K. Immunological evaluation of personalized peptide vaccination in combination with UFT and UZEL for metastatic colorectal carcinoma patients. *Cancer Immunol. Immunother.*, **2009**, *58*(11), 1843-1852.
- [71] Yoshiyama, K.; Terazaki, Y.; Matsueda, S.; Shichijo, S.; Noguchi, M.; Yamada, A.; Mine, T.; Ioji, T.; Itoh, K.; Shirouzu, K.; Sasada, T.; Takamori, S. Personalized peptide vaccination in patients with refractory non-small cell lung cancer. *Int. J. Oncol.*, **2012**, *40*(5), 1492-1500.
- [72] Terazaki, Y.; Yoshiyama, K.; Matsueda, S.; Watanabe, N.; Kawahara, A.; Naito, Y.; Suekane, S.; Komatsu, N.; Ioji, T.; Yamada, A.; Mine, T.; Terasaki, M.; Itoh, K.; Takamori, S.; Sasada, T. Immunological evaluation of personalized peptide vaccination in refractory small cell lung cancer. *Cancer Sci.*, **2012**, *103*(4), 638-644.
- [73] Matsumoto, K.; Noguchi, M.; Satoh, T.; Tabata, K.; Fujita, T.; Iwamura, M.; Yamada, A.; Komatsu, N.; Baba, S.; Itoh, K. A phase I study of personalized peptide vaccination for advanced urothelial carcinoma patients who failed treatment with methotrexate, vinblastine, adriamycin and cisplatin. *BJU Int.*, **2011**, *108*(6), 831-838.
- [74] Suekane, S.; Nishitani, M.; Noguchi, M.; Komohara, Y.; Kokubu, T.; Naitoh, M.; Honma, S.; Yamada, A.; Itoh, K.; Matsuoka, K.; Kanayama, H. Phase I trial of personalized peptide vaccination for cytokine-refractory metastatic renal cell carcinoma patients. *Cancer Sci.*, **2007**, *98*(12), 1965-1968.
- [75] Sasada, T.; Komatsu, N.; Suekane, S.; Yamada, A.; Noguchi, M.; Itoh, K. Overcoming the hurdles of randomised clinical trials of therapeutic cancer vaccines. *Eur. J. Cancer*, **2010**, *46*(9), 1514-1519.
- [76] Eggermont, A.M. Therapeutic vaccines in solid tumours: can they be harmful? *Eur. J. Cancer*, **2009**, *45*(12), 2087-2090.
- [77] Disis, M.L. Immunologic biomarkers as correlates of clinical response to cancer immunotherapy. *Cancer Immunol. Immunother.*, **2011**, *60*(3), 433-442.
- [78] Hoos, A.; Eggermont, A.M.; Janetzki, S.; Hodi, F.S.; Ibrahim, R.; Anderson, A.; Humphrey, R.; Blumenstein, B.; Old, L.; Wolchok, J. Improved endpoints for cancer immunotherapy trials. *J. Natl. Cancer Inst.*, **2010**, *102*(18), 1388-1397.
- [79] Amos, S.M.; Duong, C.P.; Westwood, J.A.; Ritchie, D.S.; Junghans, R.P.; Darcy, P.K.; Kershaw, M.H. Autoimmunity associated with immunotherapy of cancer. *Blood*, **2011**, *118*(3), 499-509.
- [80] López, M.N.; Pereda, C.; Segal, G.; Muñoz, L.; Aguilera, R.; González, F.E.; Escobar, A.; Ginesta, A.; Reyes, D.; González, R.; Mendoza-Naranjo, A.; Larrondo, M.; Compán, A.; Ferrada, C.; Salazar-Onfray, F. Prolonged survival of dendritic cell-vaccinated melanoma patients correlates with tumor-specific delayed type IV hypersensitivity response and reduction of tumor growth factor beta-expressing T cells. *J. Clin. Oncol.*, **2009**, *27*(6), 945-952.
- [81] Komatsu, N.; Matsueda, S.; Tashiro, K.; Ioji, T.; Shichijo, S.; Noguchi, M.; Yamada, A.; Doi, A.; Suekane, S.; Moriyama, F.; Matsuoka, K.; Kuhara, S.; Itoh, K.; Sasada, T. Gene expression profiles in peripheral blood as a biomarker in cancer patients receiving peptide vaccination. *Cancer*, **2012**, *118*(12), 3208-3221.
- [82] Sansone, P.; Bromberg, J. Targeting the interleukin-6/Jak/stat pathway in human malignancies. *J. Clin. Oncol.*, **2012**, *30*(9), 1005-1014.
- [83] Nishimoto, N.; Kishimoto, T. Interleukin 6: from bench to bedside. *Nat. Clin. Pract. Rheumatol.*, **2006**, *2*(11), 619-626.
- [84] Kuball, J.; de Boer, K.; Wagner, E.; Wattad, M.; Antunes, E.; Weeratna, R.D.; Vicari, A.P.; Lotz, C.; van Dorp, S.; Hol, S.; Greenberg, P.D.; Heit, W.; Davis, H.L.; Theobald, M. Pitfalls of vaccinations with WT1-, Proteinase3- and MUC1-derived peptides in combination with MontanideISA51 and CpG7909. *Cancer Immunol. Immunother.*, **2011**, *60*(2), 161-171.
- [85] Krug, L.M.; Dao, T.; Brown, A.B.; Maslak, P.; Travis, W.; Bekele, S.; Korontsvit, T.; Zakhaleva, V.; Wolchok, J.; Yuan, J.; Li, H.; Tyson, L.; Scheinberg, D.A. WT1 peptide vaccinations induce CD4 and CD8 T cell immune responses in patients with mesothelioma and non-small cell lung cancer. *Cancer Immunol. Immunother.*, **2010**, *59*(10), 1467-1479.
- [86] Barve, M.; Bender, J.; Senzer, N.; Cunningham, C.; Greco, F.A.; McCune, D.; Steis, R.; Khong, H.; Richards, D.; Stephenson, J.; Ganesa, P.; Nemunaitis, J.; Ishioka, G.; Pappen, B.; Nemunaitis, M.; Morse, M.; Mills, B.; Maples, P.B.; Sherman, J.; Nemunaitis, J.J. Induction of immune responses and clinical efficacy in a phase II trial of IDM-2101, a 10-epitope cytotoxic T-lymphocyte vaccine, in metastatic non-small-cell lung cancer. *J. Clin. Oncol.*, **2008**, *26*(27), 4418-4425.
- [87] Slingluff, C.L.; Petroni, G.R.; Chianese-Bullock, K.A.; Smolkin, M.E.; Ross, M.L.; Haas, N.B.; von Mehren, M.; Grosh, W.W. Randomized multicenter trial of the effects of melanoma-associated helper peptides and cyclophosphamide on the immunogenicity of a multipptide melanoma vaccine. *J. Clin. Oncol.*, **2011**, *29*(21), 2924-2932.
- [88] Feyerabend, S.; Stevanovic, S.; Gouttefangeas, C.; Wernet, D.; Hennenlotter, J.; Bedke, J.; Dietz, K.; Pascolo, S.; Kuczyk, M.; Rammensee, H.G.; Stenzl, A. Novel multi-peptide vaccination in Hla-A2+ hormone sensitive patients with biochemical relapse of prostate cancer. *Prostate*, **2009**, *69*(9), 917-927.
- [89] Walter, S.; Weinschenk, T.; Stenzl, A.; Zdrojowy, R.; Pluzanska, A.; Szczylik, C.; Stachler, M.; Brugger, W.; Dietrich, P.Y.; Mendrzyk, R.; Hilf, N.; Schoor, O.; Fritsche, J.; Mahr, A.; Maurer, D.; Vass, V.; Trautwein, C.; Lewandrowski, P.; Flohr, C.; Pohla, H.; Stanczak, J.J.; Bronte, V.; Mandruzzato, S.; Biedermann, T.; Pawelec, G.; Derhovanessian, E.; Yamagishi, H.; Miki, T.; Hongo, F.; Takaha, N.; Hirakawa, K.; Tanaka, H.; Stevanovic, S.; Frisch, J.; Mayer-Mokler, A.; Kirner, A.; Rammensee, H.G.; Reinhardt, C.; Singh-Jasuja, H. Multipptide immune response to cancer vaccine IMA901 after single-dose cyclophosphamide associates with longer patient survival. *Nat. Med.*, **2012**, *18*(8), 1254-1261.
- [90] Meyer, R.G.; Korn, S.; Micke, P.; Becker, K.; Huber, C.; Wölfel, T.; Buhl, R. An open-label, prospective phase I/II study evaluating the immunogenicity and safety of a ras peptide vaccine plus GM-CSF in patients with non-small cell lung cancer. *Lung Cancer*, **2007**, *58*(1), 88-94.
- [91] Morse, M.A.; Secord, A.A.; Blackwell, K.; Hobeika, A.C.; Sinnathamby, G.; Osada, T.; Hafner, J.; Philip, M.; Clay, T.M.; Lyerly, H.K.; Philip, R. MHC class I-presented tumor antigens identified in ovarian cancer by immunoproteomic analysis are targets for T-cell responses against breast and ovarian cancer. *Clin. Cancer Res.*, **2011**, *17*(10), 3408-3419.
- [92] Holmes, J.P.; Benavides, L.C.; Gates, J.D.; Carmichael, M.G.; Hueman, M.T.; Mittendorf, E.A.; Murray, J.L.; Amin, A.; Craig, D.; von Hofe, E.; Ponniah, S.; Peoples, G.E. Results of the first phase I clinical trial of the novel II-key hybrid preventive HER-2/neu peptide (AE37) vaccine. *J. Clin. Oncol.*, **2008**, *26*(20), 3426-3433.
- [93] Perez, S.A.; Kallinteris, N.L.; Bisias, S.; Tzonis, P.K.; Georgakopoulou, K.; Varla-Leftherioti, M.; Papamichail, M.; Thanos, A.; von Hofe, E.; Baxevanis, C.N. Results from a phase I clinical study of the novel II-Key/HER-2/neu(776-790) hybrid peptide vaccine in patients with prostate cancer. *Clin. Cancer Res.*, **2010**, *16*(13), 3495-3506.
- [94] Kallinteris, N.L.; Lu, X.; Blackwell, C.E.; von Hofe, E.; Humphreys, R.E.; Xu, M. II-Key/MHC class II epitope hybrids: a strategy that enhances MHC class II epitope loading to create more potent peptide vaccines. *Expert Opin. Biol. Ther.*, **2006**, *6*(12), 1311-1321.
- [95] Takahashi, N.; Ohkuri, T.; Homma, S.; Ohtake, J.; Wakita, D.; Togashi, Y.; Kitamura, H.; Todo, S.; Nishimura, T. First clinical

- trial of cancer vaccine therapy with artificially synthesized helper/ killer-hybrid epitope long peptide of MAGE-A4 cancer antigen. *Cancer Sci.*, **2012**, *103*(1), 150-153.
- [96] Zandvliet, M.L.; Kester, M.G.; van Liempt, E.; de Ru, A.H.; van Veelen, P.A.; Griffioen, M.; Guchelaar, H.J.; Falkenburg, J.H.; Meij, P. Efficiency and mechanism of antigen-specific CD8+ T-cell activation using synthetic long peptides. *J. Immunother.*, **2012**, *35*(2), 142-153.
- [97] Speetjens, F.M.; Kuppen, P.J.; Welters, M.J.; Essahsah, F.; Voet van den Brink, A.M.; Lantrua, M.G.; Valentijn, A.R.; Oostendorp, J.; Fathers, L.M.; Nijman, H.W.; Drijfhout, J.W.; van de Velde, C.J.; Melief, C.J.; van der Burg, S.H. Induction of p53-specific immunity by a p53 synthetic long peptide vaccine in patients treated for metastatic colorectal cancer. *Clin. Cancer Res.*, **2009**, *15*(3), 1086-1095.
- [98] Vermeij, R.; Leffers, N.; Hoogeboom, B.N.; Hamming, I.L.; Wolf, R.; Reyners, A.K.; Molmans, B.H.; Hollema, H.; Bart, J.; Drijfhout, J.W.; Oostendorp, J.; van der Zee, A.G.; Melief, C.J.; van der Burg, S.H.; Daemen, T.; Nijman, H.W. Potentiation of a p53-SLP vaccine by cyclophosphamide in ovarian cancer: a single-arm phase II study. *Int. J. Cancer*, **2012**, *131*(5), E670-680.
- [99] Zeestraten, E.C.; Speetjens, F.M.; Welters, M.J.; Saadatmand, S.; Stynenbosch, L.F.; Jongen, R.; Kapiteijn, E.; Gelderblom, H.; Nijman, H.W.; Valentijn, A.R.; Oostendorp, J.; Fathers, L.M.; Drijfhout, J.W.; van de Velde, C.J.; Kuppen, P.J.; van der Burg, S.H.; Melief, C.J. Addition of interferon- $\alpha$  to the p53-SLP vaccine results in increased production of interferon- $\gamma$  in vaccinated colorectal cancer patients: A phase I/II clinical trial. *Int. J. Cancer*, **2013**, *132*(7), 1581-1591.
- [100] Kakimi, K.; Isobe, M.; Uenaka, A.; Wada, H.; Sato, E.; Doki, Y.; Nakajima, J.; Seto, Y.; Yamatsuji, T.; Naomoto, Y.; Shiraishi, K.; Takigawa, N.; Kiura, K.; Tsuji, K.; Iwatsuki, K.; Oka, M.; Pan, L.; Hoffman, E.W.; Old, L.J.; Nakayama, E. A phase I study of vaccination with NY-ESO-1 peptide mixed with Picibanil OK-432 and Montanide ISA-51 in patients with cancers expressing the NY-ESO-1 antigen. *Int. J. Cancer*, **2011**, *129*(12), 2836-2846.
- [101] Berzofsky, J.A.; Terabe, M.; Oh, S.; Belyakov, I.M.; Ahlers, J.D.; Janik, J.E.; Morris, J.C. Progress on new vaccine strategies for the immunotherapy and prevention of cancer. *J. Clin. Invest.*, **2004**, *113*(11), 1515-1525.
- [102] Pulendran, B.; Ahmed, R. Translating innate immunity into immunological memory: implications for vaccine development. *Cell*, **2006**, *124*(4), 849-863.
- [103] Steinman, R.M.; Hawiger, D.; Nussenzweig, M.C. Tolerogenic dendritic cells. *Annu. Rev. Immunol.*, **2003**, *21*, 685-711.
- [104] Banchereau, J.; Palucka, A.K. Dendritic cells as therapeutic vaccines against cancer. *Nat. Rev. Immunol.*, **2005**, *5*(4), 296-306.
- [105] Gilboa, E. DC-based cancer vaccines. *J. Clin. Invest.*, **2007**, *117*(5), 1195-1203.
- [106] Reddy, S.T.; Swartz, M.A.; Hubbell, J.A. Targeting dendritic cells with biomaterials: developing the next generation of vaccines. *Trends Immunol.*, **2006**, *27*(12), 573-579.
- [107] Tacken, P.J.; de Vries, I.J.; Torensma, R.; Figdor, C.G. Dendritic-cell immunotherapy: from ex vivo loading to in vivo targeting. *Nat. Rev. Immunol.*, **2007**, *7*(10), 790-802.
- [108] Ferrari, M.; Downing, G. Medical nanotechnology: shortening clinical trials and regulatory pathways? *BioDrugs*, **2005**, *19*(4), 203-210.
- [109] Almeida, A.J.; Souto, E. Solid lipid nanoparticles as a drug delivery system for peptides and proteins. *Adv. Drug Deliv. Rev.*, **2007**, *59*(6), 478-490.
- [110] Vasir, J.K.; Labhsetwar, V. Biodegradable nanoparticles for cytosolic delivery of therapeutics. *Adv. Drug Deliv. Rev.*, **2007**, *59*(8), 718-728.
- [111] Langer, R.; Cleland, J.L.; Hanes, J. New advances in microsphere-based single-dose vaccines. *Adv. Drug Deliv. Rev.*, **1997**, *28*(1), 97-119.
- [112] Shen, H.; Ackerman, A.L.; Cody, V.; Giodini, A.; Hinson, E.R.; Cresswell, P.; Edelson, R.L.; Saltzman, W.M.; Hanlon, D.J. Enhanced and prolonged cross-presentation following endosomal escape of exogenous antigens encapsulated in biodegradable nanoparticles. *Immunology*, **2006**, *117*(1), 78-88.
- [113] Zhang, Z.; Tongchusak, S.; Mizukami, Y.; Kang, Y.J.; Ioji, T.; Tourma, M.; Reinhold, B.; Keskin, D.B.; Reinherz, E.L.; Sasada, T. Induction of anti-tumor cytotoxic T cell responses through PLGA-nanoparticle mediated antigen delivery. *Biomaterials*, **2011**, *32*(14), 3666-3678.
- [114] Noguchi, M.; Itoh, K.; Suekane, S.; Morinaga, A.; Sukehiro, A.; Suetsugu, N.; Katagiri, K.; Yamada, A.; Noda, S. Immunological monitoring during combination of patient-oriented peptide vaccination and estramustine phosphate in patients with metastatic hormone refractory prostate cancer. *Prostate*, **2004**, *60*(1), 32-45.
- [115] Noguchi, M.; Itoh, K.; Yao, A.; Mine, T.; Yamada, A.; Obata, Y.; Furuta, M.; Harada, M.; Suekane, S.; Matsuoka, K. Immunological evaluation of individualized peptide vaccination with a low dose of estramustine for HLA-A24+ HRPC patients. *Prostate*, **2005**, *63*(1), 1-12.
- [116] Noguchi, M.; Yao, A.; Harada, M.; Nakashima, O.; Komohara, Y.; Yamada, S.; Itoh, K.; Matsuoka, K. Immunological evaluation of neoadjuvant peptide vaccination before radical prostatectomy for patients with localized prostate cancer. *Prostate*, **2007**, *67*(9), 933-942.

RESEARCH ARTICLE

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# Feasibility study of personalized peptide vaccination for metastatic recurrent triple-negative breast cancer patients

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## Abstract

**Introduction:** Since treatment modalities for metastatic recurrent triple-negative breast cancer (mTNBC) are limited, a novel treatment approach including immunotherapy is required. We have developed a novel regimen of personalized peptide vaccination (PPV), in which vaccine antigens are individually selected from a pool of different peptide candidates based on the pre-existing host immunity. Herein we conducted a phase II study of PPV for metastatic recurrent breast cancer patients to investigate the feasibility of PPV for mTNBC.

**Methods:** Seventy-nine patients with metastatic recurrent breast cancer who had metastases and had failed standard chemotherapy and/or hormonal therapy were enrolled. They were subgrouped as the mTNBC group (n = 18), the luminal/human epidermal growth factor receptor 2 (HER2)-negative group (n = 41) and the HER2-positive group (n = 18), while the remaining two patients had not been investigated. A maximum of four human leukocyte antigen (HLA)-matched peptides showing higher peptide-specific immunoglobulin G (IgG) responses in pre-vaccination plasma were selected from 31 pooled peptide candidates applicable for the four HLA-IA phenotypes (HLA-A2, -A24, or -A26 types, or HLA-A3 supertypes), and were subcutaneously administered weekly for 6 weeks and bi-weekly thereafter. Measurement of peptide-specific cytotoxic T lymphocyte (CTL) and IgG responses along with other laboratory analyses were conducted before and after vaccination.

**Results:** No severe adverse events associated with PPV were observed in any of the enrolled patients. Boosting of CTL and/or IgG responses was observed in most of the patients after vaccination, irrespective of the breast cancer subtypes. There were three complete response cases (1 mTNBC and 2 luminal/HER2-negative types) and six partial response cases (1 mTNBC and 5 luminal/HER2-negative types). The median progression-free survival time and median overall survival time of mTNBC patients were 7.5 and 11.1 months, while those of luminal/HER2-negative patients were 12.2 and 26.5 months, and those of HER2-positive patients were 4.5 and 14.9 months, respectively.

**Conclusions:** PPV could be feasible for mTNBC patients because of the safety, immune responses, and possible clinical benefits.

**Clinical Trial Registration Number:** UMIN000001844 (Registration Date: April 5, 2009)

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## Introduction

Recent advances in chemotherapies, hormonal therapies and anti-human epidermal growth factor receptor 2 (HER2) therapies have significantly improved the prognosis in metastatic recurrent breast cancer patients. For example, new chemotherapies using agents such as nanoparticle albumin-bound paclitaxel (nab-PTX) [1,2], eribulin mesylate [3,4] and bevacizumab [5-7], new hormonal therapies such as flvestrant injection [8,9] or new anti-HER2 therapies such as those using pertuzumab [10,11] and trastuzumab emtansine (T-DM1) [12] have shown significant clinical benefits in metastatic recurrent breast cancer patients. Despite these novel therapeutic advances, the treatment modalities for chemotherapy-resistant triple-negative breast cancer (TNBC) remain limited, and thus a novel treatment approach including immunotherapy is required. Nevertheless, no randomized controlled trials of cancer vaccine have shown promise of clinical benefit for metastatic recurrent breast cancer patients, particularly in metastatic recurrent TNBC (mrTNBC).

We have developed a novel regimen of personalized peptide vaccination (PPV), in which vaccine antigens are selected from a pool of 31 different peptide candidates based on the pre-existing immunoglobulin G (IgG) responses specific to each peptide before vaccination [13-17]. Most of the peptides employed for PPV, except for those derived from prostate-related antigens, are known to be commonly expressed in various types of advanced cancers. Our previous clinical trials of PPV for patients with advanced cancers demonstrated the safety and feasibility of this new approach [13-17]. Here we conducted a phase II study of PPV for metastatic recurrent breast cancer to investigate the feasibility of PPV for mrTNBC.

## Methods

### Patients and methods

Women with a histological diagnosis of metastatic recurrent breast cancer were eligible for inclusion in the present study. All patients were required to have evaluable recurrent and/or metastatic tumors at the time of entry. Patients were divided into three different intrinsic subtypes as follows: luminal (estrogen-receptor-positive)/HER2-negative type, HER2-positive type (immunohistochemical score 3+ or HER2 gene/chromosome 17 ratio >2.2 in fluorescence *in situ* hybridization) and TNBC (hormone-receptor-negative and HER2-negative). Most patients had failed standard chemotherapy, but a few patients who had failed hormonal therapy alone were also eligible for this study. All patients were required to show positive IgG responses to at least 2 of the 31 different vaccine candidate peptides, as reported previously [13-17]. Other inclusion criteria were as follows: age between 20 and 80 years; an Eastern Cooperative Oncology

Group (ECOG) performance status of 0 or 1; positive status for human leukocyte antigen (HLA)-A2, -A24 or -A26 types, or HLA-A3 supertypes (A3, A11, A31 or A33); life expectancy of at least 12 weeks; and adequate hematologic, hepatic and renal function. Exclusion criteria included pulmonary, cardiac or other systemic diseases; an acute infection; a history of severe allergic reactions; pregnancy or nursing; and other inappropriate conditions for enrollment as judged by clinicians. Patients with a lymphocyte count of <1,000/ $\mu$ L were excluded from the study, since we previously reported that pre-vaccination lymphocytopenia (<1,000 cells/ $\mu$ L) is an unfavorable factor for overall survival (OS) in cancer patients receiving PPV [17]. The protocol was approved by the Kurume University Ethical Committee and registered in the UMIN Clinical Trials Registry (Registration, number UMIN000001844; Registration date, 5 April 2009). All patients were given a full explanation of the protocol and provided their informed consent before enrollment.

### Clinical protocol

This was a phase II study to evaluate the safety and immunological responses in metastatic recurrent breast cancer patients under PPV. Thirty-one peptides, whose safety and immunological effects for other types of cancer were confirmed in previously conducted clinical studies [14-17], were employed for vaccination (12 peptides for HLA-A2, 16 peptides for HLA-A24, 9 peptides for HLA-A3 supertypes (-A3, -A11, -A31, and -A33) and 4 peptides for HLA-A26) (Additional file 1: Table S1). These peptides were prepared under the conditions of Good Manufacturing Practice (GMP) by the PolyPeptide Laboratories (San Diego, CA, USA) and American Peptide Company (Vista, CA, USA). Peptides for vaccination of individual patients were selected in consideration of the pre-existing host immunity before vaccination, as assessed by the titers of IgG specific to each of the 31 different vaccine candidates.

A maximum of four peptides (3 mg/each peptide), which were selected based on the results of HLA typing and peptide-specific IgG titers, were subcutaneously administered with incomplete Freund's adjuvant (Montanide ISA51; Seppic, Paris, France) once a week for six consecutive weeks. After the first cycle of six vaccinations, up to four antigen peptides, which were re-selected according to the titers of peptide-specific IgG at the sixth vaccination, were administered every two weeks. After the second cycle of six vaccinations, up to four antigen peptides, which were also re-selected, were administered every four to eight weeks according to the immune responses after PPV. These protocols were continued until remarkable disease progression or disease in remission was shown, according to the will of the individual patient. During the PPV, patients were allowed to receive combination therapies

such as chemotherapy, hormonal therapy, anti-HER2 therapy and radiotherapy. Adverse events were monitored according to the National Cancer Institute Common Terminology Criteria for Adverse Events version 3.0 (NCI-CTC Ver.-3.0). Complete blood counts and serum biochemistry tests were performed after every six vaccinations. The clinical responses were determined by the Response Evaluation Criteria in Solid Tumors (RECIST) in the vaccinated patients. The RECIST-based clinical responses were evaluated after nearly 12 vaccinations by radiological findings of computed tomography (CT) scan and/or magnetic resonance imaging (MRI), and the best overall responses during PPV treatment were shown. For the patients who did not complete the second cycle of 12 vaccinations, the newest radiological findings

were evaluated, except in the case of patients who had died before the RECIST-based radiological evaluation.

#### Measurement of humoral and cellular immune responses

Humoral immune responses specific to each of the 31 peptide candidates were determined by peptide-specific IgG levels using the Luminex system (Luminex, Austin, TX, USA), as previously reported [18]. If the titers of peptide-specific IgG to at least one of the vaccinated peptides in the post-vaccination plasma were more than two-fold higher than those in the pre-vaccination plasma, the changes were considered to be significant, as previously reported [14-17]. Cellular immune responses specific to the vaccinated peptides were evaluated by interferon (INF)- $\gamma$  ELISPOT using peripheral blood mononuclear cells (PBMCs) as previously

**Table 1 Comparison of patient characteristics for overall and breast cancer subtypes**

Character	Overall (number = 79)	mrTNBC (number = 18)	Luminal/HER2-negative (number = 41)	HER2-positive (number = 18)	P-value <sup>a</sup>
Median age (range)	57 (30 to 77)	55 (30 to 65)	55 (39 to 76)	62 (51 to 70)	0.019
Performance status					0.046
0/1	58/21	14/4	32/9	11/7	
Median time to the first PPV from recurrence, months (range)	35 (1.2 to 165)	8 (1.2 to 99)	40 (2 to 123)	55 (19 to 165)	0.101
Histopathology					0.079
Ductal carcinoma	71	16	37	17	
Lobular carcinoma	4	1	2	1	
Others	4	1	2		
Positive status of HLA-A24	55	10	29	14	0.039
Positive status of HLA-A2	22	6	8	8	0.015
Visceral metastasis					0.009
Yes/No	60/19	8/6	29/11	16/2	
Brain metastasis					0.002
Yes/No	11/68	3/11	2/38	6/12	
Median duration of previous chemotherapies, months (range)	12 (2 to 148)	9 (4 to 43)	12 (2 to 9)	37 (10 to 148)	<0.0001
Usage of previous standard chemotherapy					
Anthracycline	50	16	22	12	0.003
Taxane	58	16	24	17	0.0006
Trastuzumab	18		3	15	<0.0001
Regimen number of previous chemotherapies					0.008
<4/ $\geq$ 4	36/36	8/9	22/14	6/12	
Combined therapies					
Oral chemotherapy	19	5	8	6	0.060
Infusion chemotherapy	32	10	15	7	0.038
Anti-HER2 therapy	11			11	<0.0001
Hormonal therapy	23	2	17	4	0.006
Median times of peptide vaccination	14 (2 to 39)	12 (2 to 30)	15 (4 to 39)	12 (6 to 22)	0.021

<sup>a</sup>The Mann-Whitney U test and Fisher-Freeman-Halton exact test were performed to examine P-values for continuous values and categorical values. HER2, human epidermal growth factor 2; HLA, human leukocyte antigens; mrTNBC, metastatic recurrent triple-negative breast cancer; PPV, personalized peptide vaccination.

**Table 2 Adverse events during the PPV**

Adverse event	Grade 1	Grade 2	Grade 3	Grade 4
Injection site reaction	42	37		
Constitutional symptom				
Fever	8	1		
Malaise	7	3		
Edema limbs	2	3		
Pain	5			
Tumor pain	4	9		
Gastrointestinal				
Nausea	4			
Mucositis oral		1		
Abdominal pain			1	
Constipation	1	1		
Diarrhea	2			
Respiratory				
Dyspnea	5	1	1	
Cough	5			
Hoarseness	2			
Pneumonitis		1		
Pleural effusion	1			
Hypoxia		1	2	
Neurological				
Headache	1	1		
Dysgeusia	1			
Dizziness	2	1		
Peripheral sensory neuropathy	8	2		
Peripheral motor neuropathy	1			
Endocrine disorder				
Hypothyroidism		1		
Skin and subcutaneous				
Pruritis	33	3		
Urticaria	4	1		
Reproductive system				
Vaginal hemorrhage	2			
Vascular disorders				
Hot flashes	1			
Lymphedema		1		
Hypertension		1		
Blood/Bone marrow				
Anemia	18	9	2	2
Hemoglobin increased	1			
Leukocytopenia	22	11	4	
Neutropenia	2	6	5	1

**Table 2 Adverse events during the PPV (Continued)**

Lymphocytopenia	29	7	12
Thrombocytopenia	6		1
Metabolism and nutrition			
Anorexia	2		
Hyponatremia	1		
Hyperkalemia	3		
Hypocalcemia	2		
Hyperglycemia	2		
Laboratory			
AST increased	14	9	2
ALT increased	19	4	2
γ-GTP increased	7	3	2
ALP increased	6	1	1
Hyperbilirubinemia	3	1	1
Creatinine increased	10	3	1
Cholesterol high	4		
Hypoalbuminemia	46	7	
INR increased	1	1	1
APTT increased	1		

ALP, alkaline phosphatase; ALT, alanine aminotransferase; AST, aspartate aminotransferase; APTT, activated partial thromboplastin time; γ-GTP, gamma-glutamyl transpeptidase; INR, international normalized ratio; PPV, personalized peptide vaccination.

reported [14-17]. As a control, cellular immune responses specific to CEF peptides (MABTECH, Cincinnati, OH, USA), a mixture of virus-derived cytotoxic T lymphocyte (CTL) epitopes, were also examined.

### Statistical analyses

The Mann-Whitney U test and Fisher-Freeman-Halton exact test were used to examine statistical differences for continuous values and categorical values, respectively. *P*-values less than 0.05 were considered to be statistically significant. Progression-free survival (PFS) or OS was calculated from the date of the first vaccination until the date of disease progression or death, respectively, or the last date when the patient was known to be alive. The survival analysis was performed using the Kaplan-Meier method, and a comparison of the survival curves was performed with the log-rank test. Statistical tests were performed using JMP version 10 (SAS Institute Inc., Cary, NC, USA) and StatXact version 8 (Cytel Inc., Cambridge, MA, USA).

### Results

#### Patient characteristics

Between January 2009 and April 2013, 79 metastatic recurrent breast cancer patients were enrolled in this study. The patient characteristics are shown in Table 1

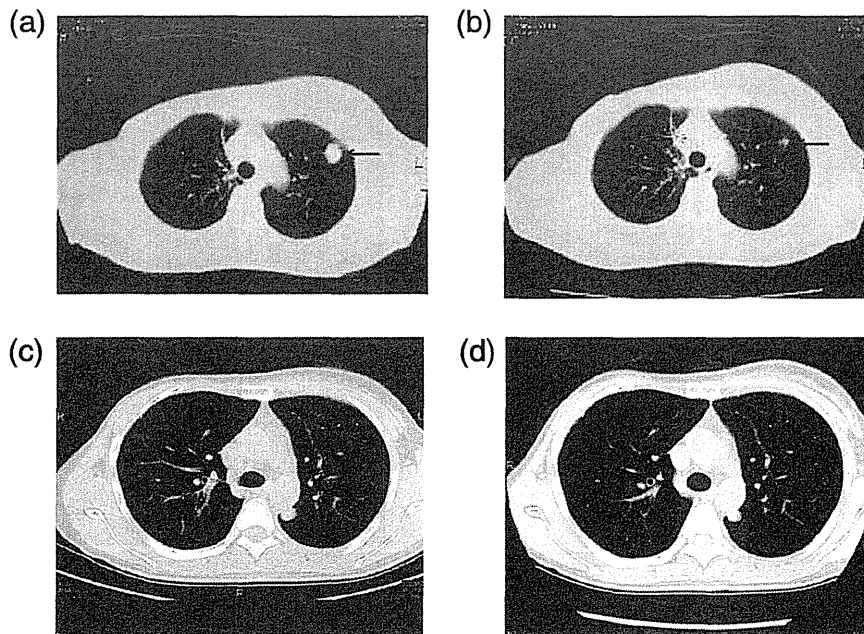
for the overall patient group and each of the three subtypes. Among the 79 patients, 77 patients had been investigated to determine their intrinsic subtype before vaccination, while the remaining 2 patients had not. The HER2-positive group was associated with older median age ( $P = 0.019$ ), restricted performance status ( $P = 0.046$ ), higher positivity of HLA-A24 or -A2 ( $P = 0.039$  or  $P = 0.015$ ), higher frequency of visceral or brain metastasis ( $P = 0.009$  or  $P = 0.002$ ) and longer duration of previous chemotherapies ( $P < 0.0001$ ). Although the mrTNBC group had a shorter duration of previous chemotherapies ( $P < 0.0001$ ), most of the mrTNBC patients had received previous standard chemotherapy (anthracycline,  $P = 0.003$ ; taxane,  $P = 0.0006$ ).

#### Combined therapies and adverse events

The median number of peptide vaccinations was 14, with a range from 2 to 39 vaccinations (Table 1). Table 2 shows all adverse events during the PPV. As the vaccination-related adverse events, all patients showed grade 1 or 2 dermatological reactions to PPV at the injection sites, but no patients showed severe adverse events (grade 3 or more). Forty-one patients (52.0%) showed grade 3 or 4 adverse events strongly associated

with combined chemotherapies and disease progression (Table 2).

During the PPV, 51 patients (64.6%; 32 cases with infusion chemotherapy and 19 cases with oral chemotherapy) received combined chemotherapies, while 23 patients (29.1%) and 11 patients (13.9%) received hormonal therapies and anti-HER2 therapies, respectively (Table 1). The mrTNBC patients received combined infusion chemotherapy more frequently than other breast cancer subtypes ( $P = 0.038$ ). The most commonly used chemotherapy drug was capecitabine (fifteen cases; 19.0%), followed by gemcitabine (eight cases; 10.1%), eribulin mesylate (six cases; 7.6%), FEC (5-fluorouracil, epirubicin and cyclophosphamide), nab-PTX or vinorelbine (four cases each; 5.1%), paclitaxel (three cases; 3.8%), bevacizumab, irinotecan or S-1 (two cases each; 2.5%), and docetaxel, oral cyclophosphamide or tegafur (one case each; 1.3%). Eleven patients (13.9%) received combined anti-HER2 therapies including trastuzumab (six cases; 7.6%) and lapatinib (five cases; 6.3%); combined anti-HER2 therapy was the most used treatment for the HER2-positive group ( $P < 0.0001$ ). In addition, 23 patients (29.1%) received combined hormonal therapies using agents, such as aromatase inhibitor (16 cases; 20.3%), high-dose toremifene (5 cases; 6.3%) and



**Figure 1** Clinical responses to PPV. **a, b**) Computed tomography findings of a PR case with mrTNBC (case 2 in Table S2) before and after the 12th vaccination. A 63-year-old woman with a recurrent lung mass underwent 12 vaccinations in combination with gemcitabine (1,000 mg/m<sup>2</sup>/week for three weeks followed by one week intermission). At four months after the first vaccination, the lung mass was remarkably reduced in size (arrow). **c, d**) Computed tomography findings of a SD case with mrTNBC (case 18 in Table S2) before and after the eighth vaccination. A 34-year-old woman with a recurrent lung mass underwent eight vaccinations in combination with two cycles of eribulin mesylate (1.4 mg/m<sup>2</sup>/week for two weeks followed by one week intermission). At three months after the first vaccination, the lung mass was slightly decreased in size (arrow). mrTNBC, metastatic recurrent triple negative breast cancer; PPV, personalized peptide vaccination; PR, partial response; SD, stable disease.

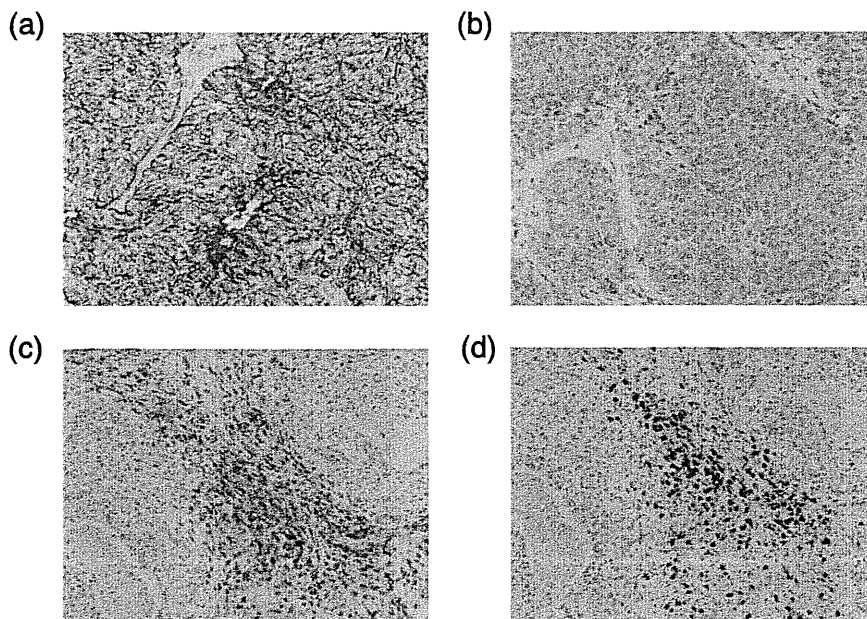
fluvestrant (2 cases; 2.5%); combined hormonal therapy was the most used treatment for the luminal/HER2-negative group ( $P = 0.046$ ).

#### Immune responses to the vaccinated peptides

Both humoral and cellular immune responses specific to the vaccinated peptides were analyzed in blood samples before and after vaccination. Plasma samples were collected from 79, 75, or 53 patients before vaccination, at the 6th vaccination, or at the 12th vaccination, respectively. For the monitoring of humoral immune responses, peptide-specific IgGs reactive to each of the 31 different peptides, including both vaccinated and non-vaccinated peptides, were measured by bead-based multiplex assay. The numbers of peptides employed for the first cycle of vaccinations were 2, 3, or 4 in 8, 6, or 63 patients, respectively (Additional file 2: Table S2, Additional file 3: Table S3, and Additional file 4: Table S4). Augmentation of IgG responses specific to at least one of the vaccinated peptides after 6 or 12 vaccinations was observed in 53/75 (70.7%) patients or 50/53 (94.3%) patients, respectively. Peptide-specific IgG responses after 6 or 12 vaccinations were augmented in 7/15 (46.7%) patients or 9/10 (90%) patients with mrTNBC (Additional file 2: Table S2). Such augmentation was seen in 28/40 (70.0%) patients or 29/31 (93.5%)

patients in the luminal/HER2-negative group (Additional file 3: Table S3) and in 16/18 (88.9%) patients or 11/11 (100%) patients of the HER2-positive group (Additional file 4: Table S4), respectively.

Cellular immune responses to vaccinated peptides were assessed by IFN- $\gamma$  ELISPOT assay. Antigen-specific CTL responses were detectable in 17/66 (25.8%) patients before vaccination (Additional file 2: Table S2, Additional file 3: Table S3 and Additional file 4: Table S4). In contrast, augmentation of the CTL responses specific to at least one of the vaccinated peptides after six vaccinations was observed in 34/63 patients (54.0%). Peptide-specific CTL responses after six vaccinations were augmented in 7/14 (50.0%) patients with mrTNBC (Additional file 2: Table S2), while such augmentation was seen in 18/31 (58.1%) patients and 7/16 (43.8%) patients in the luminal/HER2-negative group (Additional file 3: Table S3) and HER2-positive group (Additional file 4: Table S4), respectively. We also tested CTL responses to CEF peptides, a mixture of virus-derived CTL epitopes, as a control. CTL responses to CEF peptides were observed in 27/62 (43.5%) patients before vaccination and 15/58 (25.9%) patients after six vaccinations (Additional file 2: Table S2, Additional file 3: Table S3 and Additional file 4: Table S4).



**Figure 2 Expressions of TAAs and pathological responses to PPV.** After completion of eight vaccinations in combination with two cycles of eribulin mesylate (1.4 mg/m<sup>2</sup>/week for two weeks followed by one week intermission), the lung metastasis of a SD case with mrTNBC (case 18 in Table S2) was resected at three months after the first vaccination. The TAA expression and T cell infiltration in the resected lung tissue were examined by immunohistochemistry. **a, b** Among the four TAAs, that is, SART2, PSA, EGF-R and LCK, two TAAs were expressed in the lung tumor. **a**) EGF-R (X200); **b**) SART2 (X200). **c, d** Peritumoral infiltration of T lymphocytes was confirmed in the lung tumor. **c**) CD4<sup>+</sup> T lymphocytes (X200); **d**) CD8<sup>+</sup> T lymphocytes (X200). EGF-R, epidermal growth factor receptor; LCK, lymphocyte specific protein tyrosine kinase; mrTNBC, metastatic recurrent triple negative breast cancer; PPV, personalized peptide vaccination; PSA, prostate specific antigen; SART2, squamous cell carcinoma antigen recognized by T-cells 2; SD, stable disease; TAA, tumor associated antigens.

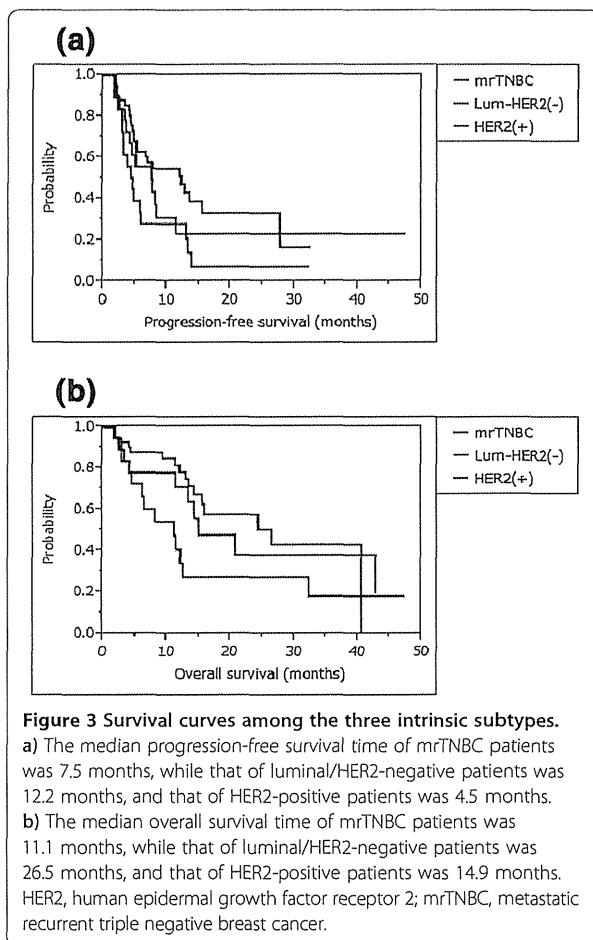
Collectively, 30/63 (47.6%) patients showed both increased CTL and IgG responses to the vaccinated peptides, 23/63 (36.5%) patients showed either increased CTL or IgG responses, and the remaining 10 (15.9%) patients showed neither CTL nor IgG boosting. In patients treated with PPV alone ( $n = 27$ ), IgG responses were more frequently increased than those in patients treated with combined chemotherapies ( $n = 47$ ) ( $P = 0.002$ ), although there was no significant difference in the increase in CTL responses ( $P = 1.000$ ).

#### Clinical responses to PPV

The RECIST-based clinical responses were evaluated in 64 patients by radiological findings. There were 3 complete response (CR), 6 partial response (PR), 27 stable disease (SD) and 28 progressive disease (PD). The overall response rate of PPV was 14%, including three CR and six PR cases. Among the responsive patients, combined chemotherapy was used in eight cases and hormonal therapy in one case. The intrinsic subtypes showed one mrTNBC and two luminal/HER2-negative types in the CR cases and one mrTNBC and five luminal/HER2-negative types in the PR cases. Computed tomography findings of each of the mrTNBC cases showing PR or SD are shown in Figure 1. The PR case (case 2 in Additional file 2: Table S2) was a 63-year-old woman with a recurrent lung mass treated with a combination of gemcitabine and PPV. At four months after the first vaccination, the lung mass was remarkably reduced in size (Figure 1a and Figure 1b). She survived 32 months after the first vaccination and died due to disease progression. The SD case (case 18 in Additional file 2: Table S2) was a 34-year-old woman with a recurrent lung mass treated with a combination of eribulin mesylate and PPV. At three months after the first vaccination, the lung mass was slightly decreased in size (Figure 1c and Figure 1d). She was subsequently treated by radical resection of the lung mass and pathological evaluation. The lung mass was metastatic TNBC with a high Ki-67 labeling index (42.0%). It expressed epidermal growth factor receptor (EGF-R) and squamous cell carcinoma antigen recognized by T-cells 2 (SART2) antigens which were vaccinated antigens (Figure 2a and Figure 2b), and peritumoral infiltration of T lymphocytes was confirmed (Figure 2c and Figure 2d). She is still alive at 13 months following the first vaccination.

#### Survival analyses by intrinsic subtypes

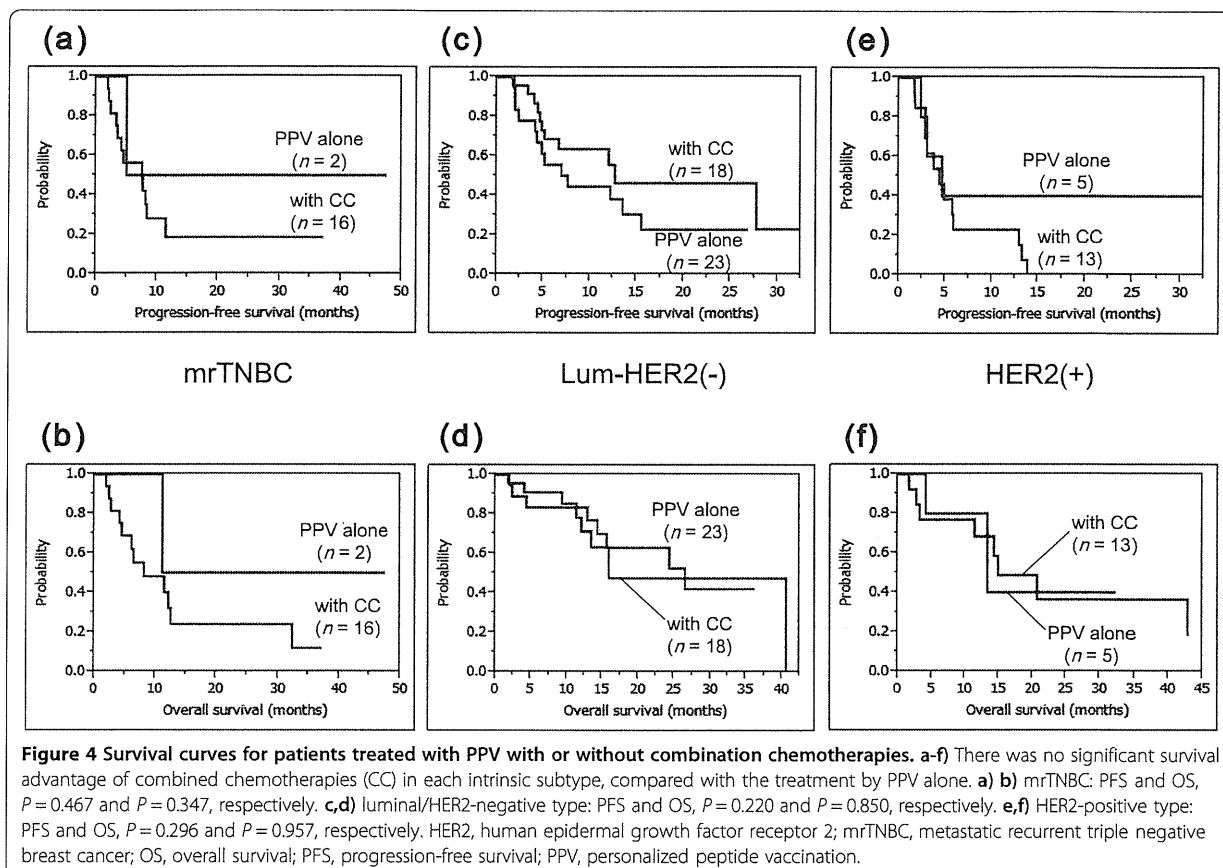
Figure 3 shows survival curves for the three intrinsic subtypes. The median progression-free survival time (MPFS) and median overall survival time (MST) of mrTNBC patients were 7.5 and 11.1 months, while those of luminal/HER2-negative patients were 12.2 and 26.5 months, and those of HER2-positive patients were 4.5 and 14.9 months, respectively. For each intrinsic subtype, the survival curves



**Figure 3 Survival curves among the three intrinsic subtypes.**  
**a)** The median progression-free survival time of mrTNBC patients was 7.5 months, while that of luminal/HER2-negative patients was 12.2 months, and that of HER2-positive patients was 4.5 months.  
**b)** The median overall survival time of mrTNBC patients was 11.1 months, while that of luminal/HER2-negative patients was 26.5 months, and that of HER2-positive patients was 14.9 months.  
 HER2, human epidermal growth factor receptor 2; mrTNBC, metastatic recurrent triple negative breast cancer.

were compared for patients treated with PPV plus concurrent chemotherapies and those treated with PPV alone (Figure 4). There was no significant advantage of concurrent chemotherapies in each intrinsic subtype, compared with treatment with PPV alone (mrTNBC: PFS and OS,  $P = 0.467$  and  $P = 0.347$ , respectively; luminal/HER2-negative type: PFS and OS,  $P = 0.220$  and  $P = 0.850$ , respectively; and HER2-positive type: PFS and OS,  $P = 0.296$  and  $P = 0.957$ , respectively).

Survival analyses along with analyses of the immune responses to PPV were also conducted in the three subtypes. Figure 5 shows the survival curves for patients with or without increased IgG responses after PPV in each intrinsic subtype. IgG boosting was a significant prognostic factor for OS and PFS in HER2-positive patients, whereas there was no significant difference between increased IgG responses and these prognoses in mrTNBC and luminal/HER2-negative patients (mrTNBC: PFS and OS,  $P = 0.274$  and  $P = 0.152$ , respectively; luminal/HER2-negative type: PFS and OS,  $P = 0.732$  and  $P = 0.571$ , respectively; HER2-positive type: PFS and OS,  $P = 0.0001$  and  $P = 0.001$ , respectively). Figure 6 shows the survival curves for patients



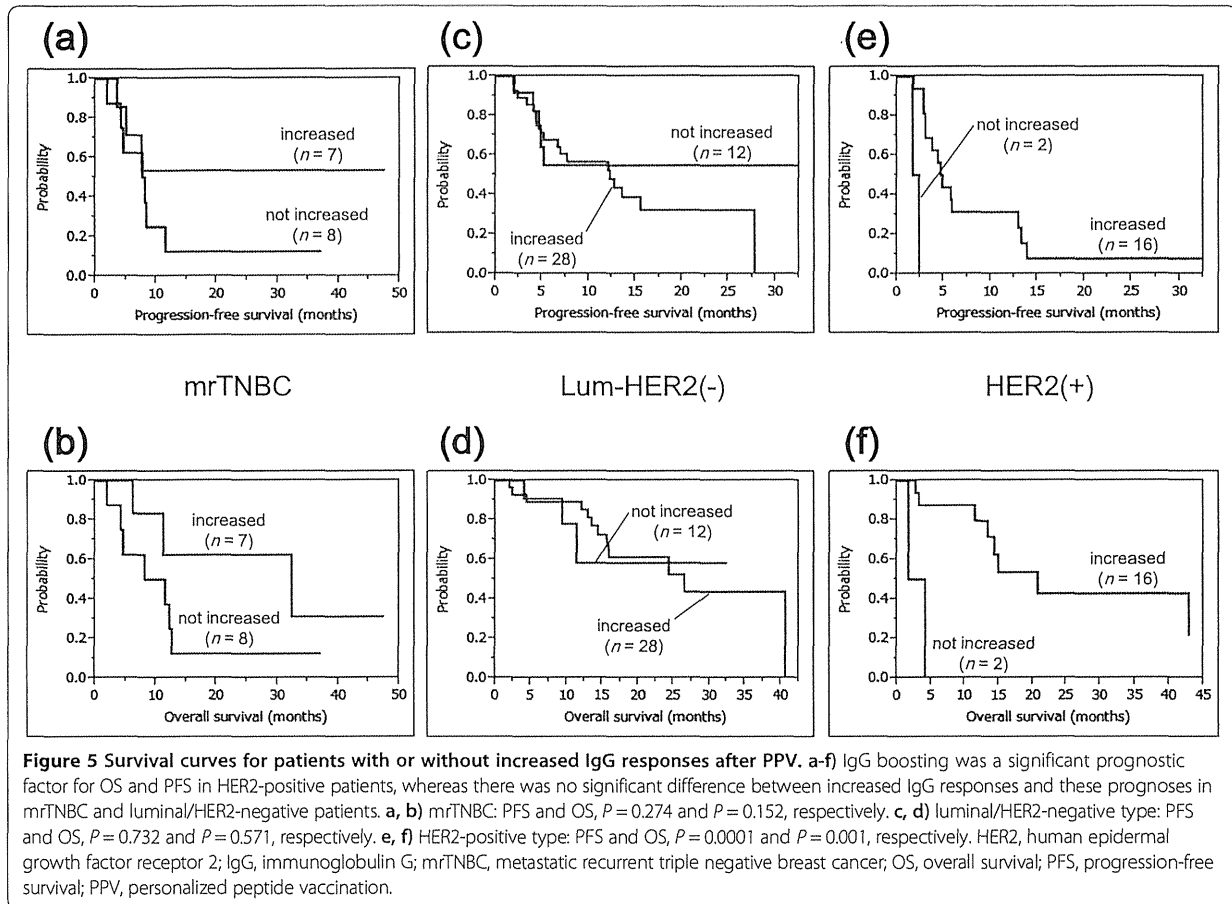
with or without increased CTL responses after PPV in each intrinsic subtype. CTL boosting was suggested to be a potential prognostic factor for OS but not for PFS in mrTNBC patients, whereas there was no significant difference between CTL boosting and these prognoses in luminal/HER2-negative and HER2-positive patients (mrTNBC: PFS and OS,  $P = 0.345$  and  $P = 0.053$ , respectively; luminal/HER2-negative type: PFS and OS,  $P = 0.272$  and  $P = 0.740$ , respectively; HER2-positive type: PFS and OS,  $P = 0.714$  and  $P = 0.758$ , respectively).

## Discussion

Since treatment outcomes in mrTNBC patients remain poor [19-21], a novel treatment modality including immunotherapy is required. Several tumor associated antigens (TAAs), such as cancer testis antigens, EGF-R, aldehyde dehydrogenase 1 (ALDH1) and enhancer of zeste homolog 2 (EZH2), are frequently expressed in TNBC, particularly in basal-like subtypes [22-24]. Despite these potential molecular targets for immunotherapy in TNBC, no randomized controlled trials of cancer vaccine have shown promise of clinical benefit to date. We have developed a novel regimen of PPV, in which vaccine antigens

are selected and administered from a pool of 31 different peptide candidates based on the pre-existing IgG responses specific to peptides before vaccination [13-17]. In previous studies, PPV was feasible for the vast majority of cancer patients with different HLA-types [13-17]. Based on these results in cancer patients, we conducted a phase II study of PPV for metastatic recurrent breast cancer patients to investigate the feasibility of PPV for mrTNBC. There were no severe adverse events associated with PPV, and most of the mrTNBC patients showed augmented immune responses to PPV.

The current study suggested the feasibility of PPV for mrTNBC patients who had failed standard chemotherapy, since the MPFST and MST of mrTNBC patients were 7.5 and 11.1 months from the first vaccination, respectively. In previously reported studies, the MPFST of mrTNBC patients treated by various chemotherapy and/or targeted therapy regimens was between 2.5 and 6.5 months [7,25-28]. Therefore, the MPFST of 7.5 months in mrTNBC patients treated by PPV in the current study seemed to be promising. Regarding OS in TNBC patients, Dent *et al.* demonstrated that the MST from recurrence to death was nine months, although the details



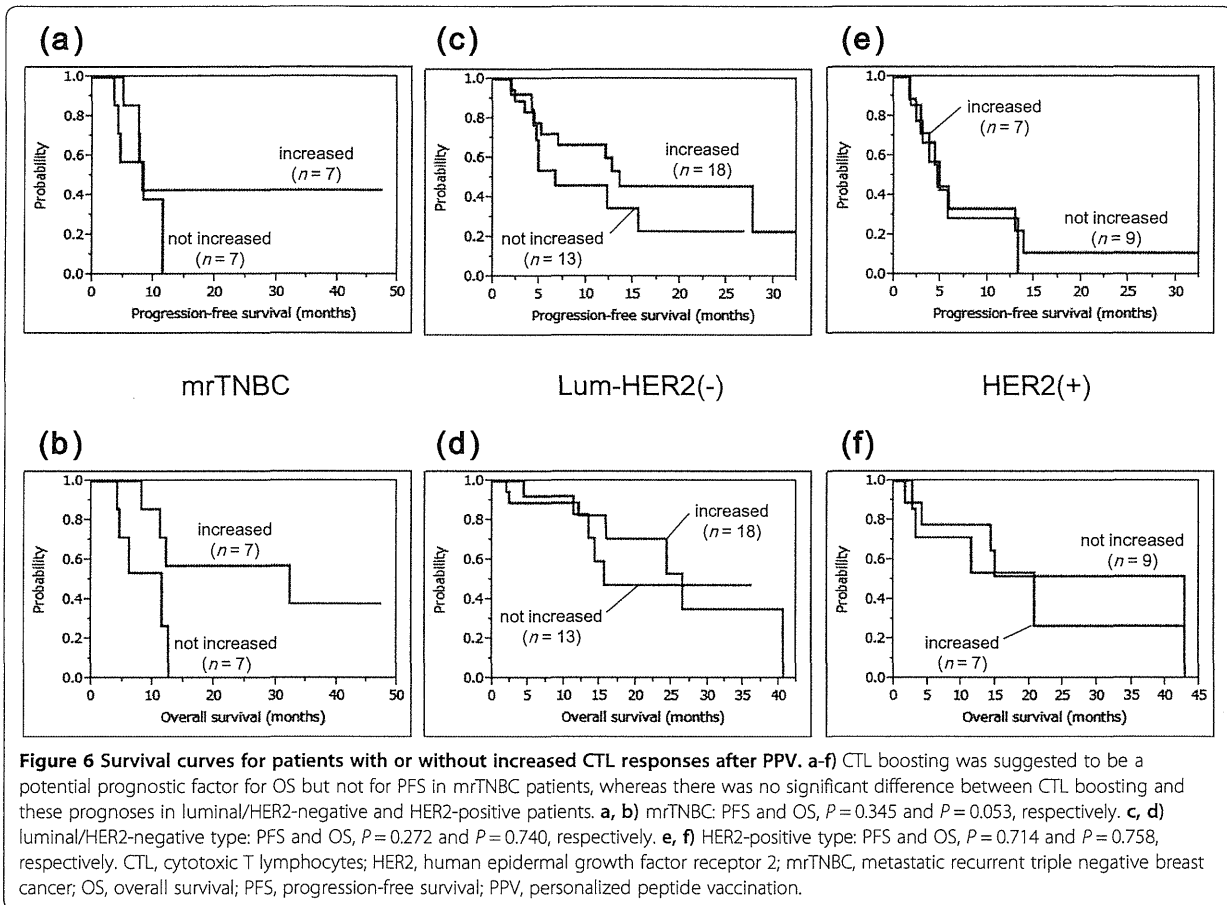
of chemotherapy regimens were not described [19]. More recently conducted studies showed that the MST of mrTNBC patients treated by various chemotherapy and/or targeted therapy regimens was between 7.7 and 17.9 months [26-28]. Although almost all patients in these previous studies were enrolled as a first-line and/or second-line treatment [26-28], most patients in the current study were enrolled as a third or more line treatment. In addition, considering that the median duration of previous chemotherapies in the mrTNBC patients in the current study was 9 months, the MST of 11.1 months in mrTNBC patients treated by PPV seems to be encouraging. As a next step, to clarify the clinical benefit of PPV in mrTNBC, we need to conduct a randomized controlled study, in which patients are treated with standard of care (SOC) alone or with PPV plus SOC.

Although the results of immune responses were not significantly different by intrinsic subtypes, a high population of HER2-positive patients showed IgG responses at the sixth vaccination. Since all of the HER2-positive patients had been treated with trastuzumab, antigen-dependent cellular cytotoxicity might have affected their

humoral immunities [29]. Notably, IgG boosting was a significant prognostic factor for OS and PFS in HER2-positive patients, although the number of patients was too small to confirm this. Combined chemotherapies also might affect the status of IgG responses, but no survival advantages of combined chemotherapies were shown in our metastatic recurrent breast cancer patients.

The clinical response rate in the present series was 14.0%, including three CR and six PR cases. Among these responsive patients, combined chemotherapy was used in eight cases and hormonal therapy in one case. The intrinsic subtype of these patients was luminal/HER2-negative type in seven cases and mrTNBC in two cases. Notably, the number of previous chemotherapy regimens was one regimen in two patients and two regimens in seven patients. In our study, more than four regimens of previous chemotherapy were significantly correlated with poor prognosis (data not shown). From these results, we would recommend PPV within three regimens of previous chemotherapy for metastatic recurrent breast cancer patients. A greater number of previous chemotherapy regimens could not sufficiently enhance





clinical responses to PPV, particularly in HER2-positive patients. Because of the significant clinical benefit and conventional usage of trastuzumab, the duration of previous chemotherapy was significantly prolonged in HER2-positive patients. The status of tumor molecular biology could change and be complicated by this long-term chemotherapy, eventually leading to a poor prognosis. For HER2-positive patients, the induction of PPV should be earlier than that in our HER2-positive patients. Since combined chemotherapies increase the number of severe adverse events in metastatic breast cancer patients, treatment with PPV alone should be performed to maintain their quality of life.

We had an opportunity to confirm the peritumoral infiltration of lymphocytes in the lung metastasis of mrTNBC. Two of the four vaccinated peptide antigens (EGF-R and squamous cell carcinoma antigen recognized by T-cells 2 (SART2)) were expressed in the lung tumor, and CTL responses to the SART2-93 antigen were significantly increased in this case (case 18 in Additional file 2: Table S2). Although IgG responses to the two peptide antigens were not significantly increased, PPV could enhance the anti-tumor immunity and efficacy of combined

chemotherapy in this case. We have investigated the expressions of 15 TAAs in primary and recurrent breast cancer tissues by immunohistochemistry (RT, unpublished data). We found that 10 of 15 TAAs were expressed in both primary and recurrent breast cancer tissues, except for lymphocyte specific protein tyrosine kinase (LCK), prostate specific antigen (PSA), prostate specific membrane antigen (PSMA), prostatic acid phosphatase (PAP) and multidrug resistance-associated protein 3 (MRP3). However, four of these five TAAs, including LCK, PSA, PAP and MRP3, were reported to be expressed in breast cancer tissues, although the frequency of expression was lower than that of other TAAs [30-34]. Therefore, 14 of the 15 TAAs could be potential molecular targets for immunotherapy in breast cancer patients.

### Conclusions

In conclusion, PPV could be feasible for mrTNBC patients because of the safety, immune responses and possible clinical benefits. For mrTNBC patients, we are planning a randomized controlled study, in which patients are treated with SOC alone or with PPV plus SOC to further clarify the clinical benefit of PPV.

## Additional files

**Additional file 1: Table S1.** Information on the peptide candidates used for PPV.

**Additional file 2: Table S2.** Immune responses to vaccinated peptides in mTNBC patients.

**Additional file 3: Table S3.** Immune responses to vaccinated peptides in luminal/HER2-negative patients.

**Additional file 4: Table S4.** Immune responses to vaccinated peptides in HER2-positive patients.

## Abbreviations

ALDH1: aldehyde dehydrogenase 1; CR: complete response; CT: computed tomography; CTL: cytotoxic T lymphocyte; ECOG: Eastern Cooperative Oncology Group; EGF-R: epidermal growth factor receptor; EZH2: enhancer of zeste homolog 2; FEC: 5- fluorouracil/epirubicin/cyclophosphamide; HER2: human epidermal growth factor 2; INF: interferon; LCK: lymphocyte specific protein tyrosine kinase; MPFST: median progression-free survival time; MRI: magnetic resonance imaging; MRP3: multidrug resistance-associated protein 3; mTNBC: metastatic recurrent triple-negative breast cancer; MST: median overall survival time; nab-PTX: nanoparticle albumin-bound paclitaxel; NCI-CTC Ver.-3.0: National Cancer Institute common terminology criteria for adverse events version 3.0; OS: overall survival; PAP: prostatic acid phosphatase; PBMCs: peripheral blood mononuclear cells; PD: progressive disease; PFS: progression-free survival; PPV: personalized peptide vaccination; PR: partial response; PSA: prostate specific antigen; PSMA: prostate specific membrane antigen; RECIST: Response Evaluation Criteria in Solid Tumors; SART2: squamous cell carcinoma antigen recognized by T-cells 2; SD: stable disease; SOC: standard of care; TAAs: tumor associated antigens; T-DM1: trastuzumab emtansine; TNBC: triple-negative breast cancer.

## Competing interests

Akira Yamada, is a Board member of the Green Peptide Company, Ltd. Kyogo Itoh and Akira Yamada have stock of the Green Peptide Company, Ltd. Kyogo Itoh received research fund from Taiho Pharmaceutical Company. The other authors declare that they have no competing interests.

## Authors' contributions

RT, UT and KI are responsible for the conception and design of the study, the acquisition, analysis and interpretation of data, and drafting the work. TS is responsible for the interpretation of data and drafting the work. NI, MT, HO, MF, TF, NS, YA and AY are responsible for the interpretation of data and revising the work critically. AK, MK and SM are responsible for the acquisition and analysis of data and revising the work critically. All authors read and approved the final manuscript.

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## References

1. Gradishar WJ, Krasnojon D, Cheporov S, Makhson AN, Manikhas GM, Clawson A, Bhar P: Significantly longer progression-free survival with nab-paclitaxel compared with docetaxel as first-line therapy for metastatic breast cancer. *J Clin Oncol* 2009, **27**:3611–3619.
2. Robinson DM, Keating GM: Albumin-bound Paclitaxel: in metastatic breast cancer. *Drugs* 2006, **66**:941–948.
3. Cortes J, O'Shaughnessy J, Loesch D, Blum JL, Vahdat LT, Petrakova K, Chollet P, Manikas A, Diéras V, Delozier T, Vladimirov V, Cardoso F, Koh H, Bougnoux P, Dutcus CE, Seegobin S, Mir D, Meneses N, Wanders J, Twelves C: Eribulin monotherapy versus treatment of physician's choice in patients with metastatic breast cancer (EMBRACE): a phase 3 open-label randomised study. *Lancet* 2011, **377**:914–923.
4. Twelves C, Cortes J, Vahdat LT, Wanders J, Akerele C, Kaufman PA: Phase III trials of eribulin mesylate (E7389) in extensively pretreated patients with locally recurrent or metastatic breast cancer. *Clin Breast Cancer* 2010, **10**:160–163.
5. Miller K, Wang M, Gralow J, Dickler M, Cobleigh M, Perez EA, Shenker T, Cella D, Davidson NE: Paclitaxel plus bevacizumab versus paclitaxel alone for metastatic breast cancer. *N Engl J Med* 2007, **357**:2666–2676.
6. Miles DW, Chan A, Dirix LY, Cortés J, Pivot X, Tomczak P, Delozier T, Sohn JH, Provencher L, Puglisi F, Harbeck N, Steger GG, Schnee Weiss A, Wardley AM, Chlistalla A, Romieu G: Phase III study of bevacizumab plus docetaxel compared with placebo plus docetaxel for the first-line treatment of human epidermal growth factor receptor 2-negative metastatic breast cancer. *J Clin Oncol* 2010, **28**:3239–3247.
7. Robert NJ, Diéras V, Glaspy J, Brufsky AM, Bondarenko I, Lipatov ON, Perez EA, Yardley DA, Chan SY, Zhou X, Phan SC, O'Shaughnessy J: RIBBON-1: randomized, double-blind, placebo-controlled, phase III trial of chemotherapy with or without bevacizumab for first-line treatment of human epidermal growth factor 2-negative, locally recurrent or metastatic breast cancer. *J Clin Oncol* 2011, **29**:1252–1260.
8. Di Leo A, Jerusalem G, Petruzella L, Torres R, Bondarenko IN, Khasanov R, Verhoeven D, Pedrini JL, Smirnova I, Lichinitser MR, Pendergrass K, Garnett S, Lindemann JP, Sapunar F, Martin M: Results of the CONFIRM phase III trial comparing fulvestrant 250 mg with fulvestrant 500 mg in postmenopausal women with estrogen receptor-positive advanced breast cancer. *J Clin Oncol* 2010, **28**:4594–4600.
9. Mehta RS, Barlow WE, Albain KS, Vandenberg TA, Dakhil SR, Tirumali NR, Lew DL, Hayes DF, Gralow JR, Livingston RB, Hortobagyi GN: Combination anastrozole and fulvestrant in metastatic breast cancer. *N Engl J Med* 2012, **367**:435–444.
10. Baselga J, Swain SM: CLEOPATRA: a phase III evaluation of pertuzumab and trastuzumab for HER2-positive metastatic breast cancer. *Clin Breast Cancer* 2010, **10**:489–491.
11. Baselga J, Cortés J, Kim SB, Im SA, Hegg R, Im YH, Roman L, Pedrini JL, Pienkowski T, Knott A, Clark E, Benyunes MC, Ross G, Swain SM, CLEOPATRA Study Group: Pertuzumab plus trastuzumab plus docetaxel for metastatic breast cancer. *N Engl J Med* 2012, **366**:109–119.
12. Verma S, Miles D, Gianni L, Krop IE, Welslau M, Baselga J, Pegram M, Oh DY, Diéras V, Guardino E, Fang L, Lu MW, Olsen S, Blackwell K, EMILIA Study Group: Trastuzumab emtansine for HER2-positive advanced breast cancer. *N Engl J Med* 2012, **367**:1783–1791.
13. Sasada T, Komatsu N, Suekane S, Yamada A, Noguchi M, Itoh K: Overcoming the hurdles of randomized clinical trials of therapeutic cancer vaccines. *Euro J Cancer* 2010, **46**:1514–1519.
14. Terasaki M, Shibui S, Narita Y, Fujimaki T, Aoki T, Kajiwara K, Sawamura Y, Kurisu K, Mineta T, Yamada A, Itoh K: Phase I trial of a personalized peptide vaccine for patients positive for human leukocyte antigen-A24 with recurrent or progressive glioblastoma multiforme. *J Clin Oncol* 2011, **29**:337–344.
15. Yanagimoto H, Shiomi H, Satou S, Mine T, Toyokawa H, Yamamoto T, Tani T, Yamada A, Kwon AH, Komatsu N, Itoh K, Noguchi M: A phase II study of personalized peptide vaccination combined with gemcitabine for non-resectable pancreatic cancer patients. *Oncol Rep* 2010, **24**:795–801.
16. Hattori T, Mine T, Komatsu N, Yamada A, Itoh K, Shiozaki H, Okuno K: Immunological evaluation of personalized peptide vaccination in combination with UFT and UZEL for metastatic colorectal carcinoma patients. *Cancer Immunol Immunother* 2009, **58**:1843–1852.
17. Noguchi M, Mine T, Komatsu N, Suekane S, Moriya F, Matsuoka K, Yutani S, Shichijo S, Yamada A, Toh U, Kawano K, Azuma K, Uemura H, Okuno K,

- Matsumoto K, Yanagimoto H, Yamanaka R, Oka M, Todo S, Sasada T, Itoh K: Assessment of immunological biomarkers in patients with advanced cancer treated by personalized peptide vaccination. *Cancer Biol Ther* 2011, **10**:1266–1279.
18. Komatsu N, Shichijo S, Nakagawa M, Itoh K: New multiplexed flow cytometric assay to measure anti-peptide antibody: a novel tool for monitoring immune responses to peptides used for immunization. *Scand J Clin Lab Invest* 2004, **64**:535–545.
19. Dent R, Trudeau M, Pritchard KI, Hanna WM, Kahn HK, Sawka CA, Lickley LA, Rawlinson E, Sun P, Narod SA: Triple-negative breast cancer: clinical features and patterns of recurrence. *Clin Cancer Res* 2007, **13**:4429–4434.
20. Kassam F, Enright K, Dent R, Dranitsaris G, Myers J, Flynn C, Fralick M, Kumar R, Clemons M: Survival outcomes for patients with metastatic triple-negative breast cancer; implications for clinical practice and trial design. *Clin Breast Cancer* 2009, **9**:29–33.
21. Foulkes WD, Smith IE, Reis-Filho JS: Triple-negative breast cancer. *N Engl J Med* 2010, **363**:1938–1948.
22. Curigliano G, Viale G, Ghioni M, Jungbluth AA, Bagnardi V, Spagnoli GC, Neville AM, Nolè F, Rotmensz N, Goldhirsch A: Cancer-testis antigen expression in triple-negative breast cancer. *Ann Oncol* 2011, **22**:98–103.
23. Ueno NT, Zhang D: Targeting EGFR in triple negative breast cancer. *J Cancer* 2011, **2**:324–328.
24. De Brot M, Rocha RM, Soares FA, Gobbi H: Prognostic impact of the cancer stem cell related markers ALDH1 and EZH2 in triple negative and basal-like breast cancers. *Pathology* 2012, **44**:303–312.
25. Finn RS, Press MF, Dering J, Arbushites M, Koehler M, Oliva C, Williams LS, Di Leo A: Estrogen receptor, progesterone receptor, human epidermal growth factor receptor 2 (HER2), and epidermal growth factor receptor expression and benefit from lapatinib in a randomized trial of paclitaxel with lapatinib or placebo as first-line treatment in HER2-negative or unknown metastatic breast cancer. *J Clin Oncol* 2009, **27**:3908–3915.
26. O'Shaughnessy J, Osborne C, Pippen JE, Yoffe M, Patt D, Rocha C, Koo IC, Sherman BM, Bradley C: Iniparib plus chemotherapy in metastatic triple-negative breast cancer. *N Engl J Med* 2011, **364**:205–214.
27. Brufsky A, Valero V, Tiangco B, Dakhil S, Brize A, Rugo HS, Rivera R, Duenne A, Bousfoul N, Yardley DA: Second-line bevacizumab-containing therapy in patients with triple-negative breast cancer: subgroup analysis of the RIBBON-2 trial. *Breast Cancer Res Treat* 2012, **133**:1067–1075.
28. Baselga J, Segalla JG, Roché H, Del Giglio A, Pinczowski H, Ciruelos EM, Filho SC, Gómez P, Van Eyll B, Bermejo B, Llombart A, Garicochea B, Durán MÁ, Hoff PM, Espià M, de Moraes AA, Ribeiro RA, Mathias C, Gil Gil M, Ojeda B, Morales J, Kwon Ro S, Li S, Costa F: Sorafenib in combination with capecitabine: an oral regimen for patients with HER2-negative locally advanced or metastatic breast cancer. *J Clin Oncol* 2012, **30**:1484–1491.
29. Musolino A, Naldi N, Bortesi B, Pezzuolo D, Capelletti M, Missale G, Laccabue D, Zerbini A, Camisa R, Bisagni G, Neri TM, Ardizzoni A: Immunoglobulin G fragment C receptor polymorphisms and clinical efficacy of trastuzumab-based therapy in patients with HER-2/neu-positive metastatic breast cancer. *J Clin Oncol* 2008, **26**:1789–1796.
30. Elsberger B, Fullerton R, Zino S, Jordan F, Mitchell TJ, Brunton VG, Mallon EA, Shiels PG, Edwards J: Breast cancer patients' clinical outcome measures are associated with Src kinase family member expression. *Br J Cancer* 2010, **103**:899–909.
31. Narita D, Raica M, Suciuc C, Cimpean A, Anghel A: Immunohistochemical expression of androgen receptor and prostate-specific antigen in breast cancer. *Folia Histochem Cytobiol* 2006, **44**:165–172.
32. Wang Y, Harada M, Yano H, Ogasawara S, Takedatsu H, Arima Y, Matsueda S, Yamada A, Itoh K: Prostatic acid phosphatase as a target molecule in specific immunotherapy for patients with nonprostate adenocarcinoma. *J Immunother* 2005, **28**:535–541.
33. Partanen L, Staaf J, Tanner M, Tuominen VJ, Borg Å, Isola J: Amplification and overexpression of the ABCC3 (MRP3) gene in primary breast cancer. *Genes Chromosomes Cancer* 2012, **51**:832–840.
34. Chang SS, Reuter VE, Heston WD, Bander NH, Grauer LS, Gaudin PB: Five different anti-prostate-specific membrane antigen (PSMA) antibodies confirm PSMA expression in tumor-associated neovasculature. *Cancer Res* 1999, **59**:3192–3198.

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RESEARCH ARTICLE

## Feasibility study of personalized peptide vaccination for recurrent ovarian cancer patients

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### Abstract

**Context:** To develop a personalized peptide vaccine (PPV) for recurrent ovarian cancer patients and evaluate its efficacy from the point of view of overall survival (OS), Phase II study of PPV was performed.

**Patients and methods:** Forty-two patients, 17 with platinum-sensitive and 25 with platinum-resistant recurrent ovarian cancer, were enrolled in this study and received a maximum of four peptides based on HLA-A types and IgG responses to the peptides in pre-vaccination plasma. **Results:** Expression of 13 of the 15 parental tumor-associated antigens encoding the vaccine peptides, with the two prostate-related antigens being the exceptions, was confirmed in the ovarian cancer tissues. No vaccine-related systemic severe adverse events were observed in any patients. Boosting of cytotoxic T lymphocytes or IgG responses specific for the peptides used for vaccination was observed in 18 or 13 of 42 cases at 6th vaccination, and 19 or 29 of 30 cases at 12th vaccination, respectively. The median survival time (MST) values of the platinum-sensitive- and platinum-resistant recurrent cases were 39.3 and 16.2 months, respectively. The MST of PPV monotherapy or PPV in combination with any chemotherapy during the 1st to 12th vaccination of platinum-sensitive cases was 39.3 or 32.2 months, and that of platinum-resistant cases was 16.8 or 16.1 months, respectively. Importantly, lymphocyte frequency and epitope spreading were significantly prognostic of OS.

**Discussion and conclusion:** Because of the safety and possible prolongation of OS, a clinical trial of PPV without chemotherapy during the 1st to 12th vaccination in recurrent ovarian cancer patients is merited.

### Keywords

Cytotoxic T-lymphocytes, epitopes, ovarian cancer, peptide vaccine, personalized medicine

### History

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### Introduction

Ovarian cancer is the leading cause of mortality among patients with gynecologic malignancies<sup>1</sup>. Although the majority of patients respond to a first-line chemotherapy with platinum and taxane agents, most patients experience relapse and develop resistance to platinum and subsequent chemotherapeutic agents<sup>2,3</sup>. Thus, it is important to develop new therapeutic approaches including cancer vaccines and molecular targeting therapy.

We and other groups previously reported the existence of tumor-reactive cytotoxic T lymphocytes (CTLs) among the tumor-infiltrating lymphocytes (TILs) in ovarian cancers<sup>4–6</sup>. In addition, a correlation between TILs and clinical outcome was reported in several studies<sup>7–9</sup>. These findings and several

clinical trials of immunotherapy in patients with ovarian cancer indicated that ovarian cancers are responsive to immunotherapy<sup>10–15</sup>. We developed and clinically tested a novel regimen of personalized peptide vaccine (PPV) in which the vaccine antigens are selected and administered based on the pre-existing host immunity before vaccination<sup>16–19</sup>. The results suggested that PPV could prolong overall survival (OS) but not progression-free survival in advanced cancer patients who fail to respond to standard chemotherapy. Moreover, a randomized clinical trial of PPV in advanced prostate cancer patients showed a favorable clinical outcome in the vaccinated group<sup>20</sup>. In this study, we examined whether PPV would be feasible as a cancer vaccine for the treatment of recurrent ovarian cancer from the viewpoint of OS.

### Materials and methods

#### Immunohistochemical analysis

Tissue specimens were collected from 22 ovarian cancer patients, including three patients who were enrolled in a

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