

図3 樹状細胞反応とTh1・Th2系細胞反応

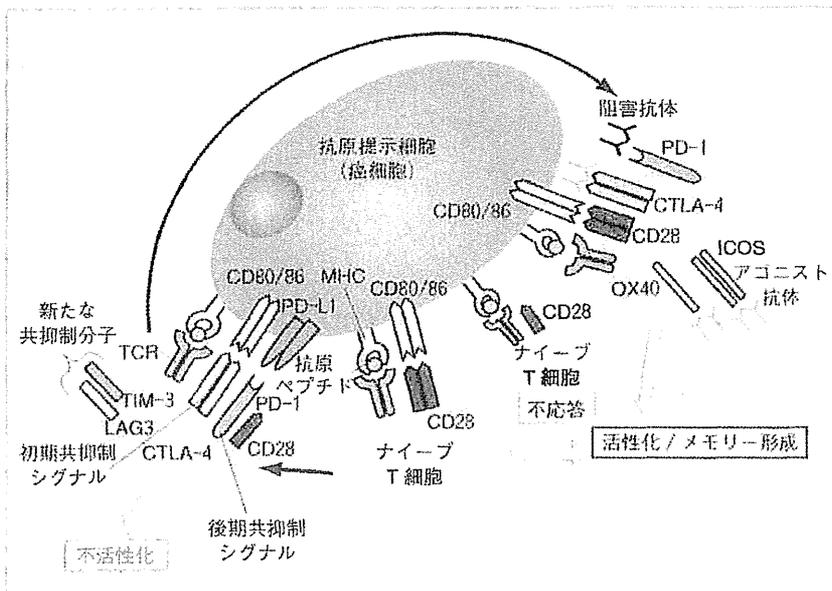


図4 T細胞の活性化と抑制の調節分子とその作用

体を認識し活性化する。MHC-IIの発現は樹状細胞、マクロファージ、B細胞などの抗原提示細胞に限定される⁶⁾。今日注目されている腫瘍免疫機構において、腫瘍細胞を特異的に認識して細胞障害性作用を示すのはCD8⁺T細胞である。その理由としては、ほとんどの腫瘍細胞がMHC-IIを発現しないことに起因している。なお、重要な機序としてクロスプレゼンテーション機構の存在がある。それは樹状細胞にみられる現象で、腫瘍細胞由来のペプチドを取り込み、MHC-Iを介してCD8⁺T細胞の活性化が生じることが知られている⁷⁾。また、免疫監視機構における初期防御や転換抑制に重要な役割を担う細胞として、natural killer (NK)細胞およびNKT細胞が知られている。前感作なしに癌細胞を傷害しうるリンパ球として知られるNKT細胞がNK細胞の表面マーカーを恒常的に発現していることが、上記のような機能を発揮する機序として明らかになっている。

2 獲得免疫における需要分子

1) MHC-IとTCR

CD8⁺T細胞のTCRとの結合には、MHC-Iとペプチドのみで可能なことから、co-stimulatory moleculeの存在を問われていない。ただし、樹状細胞にあっては、CD80/86とCD8⁺細胞のCD28の結合がMHC-Iでも重要となることが明らかになっている(図4)。

2) MHC-IIとTCR

MHC-IIと抗原分子をCD4⁺細胞が認識するにあたり、DCの「CD40」

が CD4⁺ 細胞の CD40L と、「CD80/86」が CD4⁺ 細胞の CD28 と結合することが必須条件となる (図 4)。これは、副刺激分子として重要な存在で相互の結合なくして補助シグナルが細胞内へ伝搬はありえないことから必須の分子となっている。

3) 抗原提示細胞, 腫瘍細胞あるいは感染細胞とナイーブ T 細胞

ナイーブ T 細胞の活性化には, MHC-II と CD80/86 に対しての TCR と CD28 分子の関与となるのは原則である (図 4)。一方, ナイーブ T 細胞の不活化には, 共抑制分子の存在が知られ, 初期共抑制シグナルとして CD80/86 に対応する CTLA-4 (cytotoxic T-lymphocyte antigen 4) が後期共抑制シグナルとして PD-L1 と PD-1, そしてナイーブ T 細胞の結合部位以外の細胞膜に TIM-3, LAG3 の発現が左右することも知られるに至っている。また, 同様の部位に OX40, ICOS 分子が発現し, それらに対する抗体の使用により CD28 の副刺激のもと活性化およびメモリー形成が成立されることも知られている。

4) Treg 関連分子

Treg は「アポトーシスの誘導」, 「IL-2 の消費」抗原提示細胞に対する

「抑制性サイトカインの産生」, 「アデノシン代謝を介しての IL-2, IFN- γ 産生の抑制」, 「APC 成熟化の抑制」などにより免疫反応を抑制する。アポトーシス誘導には Treg の FasLigand (FasL) の CD39/73, APC 成熟化の抑制には Treg の LAG3 が MHC-II に, CTLA-4 が CD80/86 へ結合することが知られている (図 4)。

上記に示した以外にも多くの分子の存在, 機能分担細胞の種別化に関する研究が進んでいるが, 序論として以上の紹介にとどめたい。

おわりに

今回の特集においての免疫に関する知見の紹介としては, 自然免疫研究に重点を置いての企画となっている。そのため序論として, 免疫反応全体を生体防御として捉えたうえでの自然免疫の位置付けの詳細を学ぶにあたって, 本稿ではそのための前知識としての内容を記述した。生体は, 「optimal immunity-disease protection」であることが望ましく, 「compromized immunity-disease-At risk」に外科医は悩まされている。日頃のエクササイズが, 自然免疫能に好影響を与えるとの見解が得られたとの報告はあるが, ヒト臨床上のエビデンスとしては, 不

十分な状況にある。T 細胞, B 細胞にあっては, エクササイズ終了後 24 時間を経ると元の木阿弥と化すとも報告されている。今後どのような研究展開によって, ヒトへの応用が有効となるのであろうか。研究展開に期待したいところである。

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Trials of vaccines for pancreatic ductal adenocarcinoma: Is there any hope of an improved prognosis?

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Abstract Pancreatic tumors are chemoresistant and malignant, and there are very few therapeutic options for pancreatic cancer, as the disease is normally diagnosed at an advanced stage. Although attempts have been made to develop vaccine therapies for pancreatic cancer for a couple of decades, none of the resultant protocols or regimens have succeeded in improving the clinical outcomes of patients. We herein review vaccines tested within the past few years, including peptide, biological and multiple vaccines, and describe the three sets of criteria used to evaluate the therapeutic activity of vaccines in solid tumors.

Keywords Pancreatic cancer · Vaccine · Immunomodulation

Introduction

Pancreatic cancer is the fourth leading cause of cancer-related death in the United States [1–3] and the fifth most common cause of such deaths in Japan [4]. Although surgical resection is considered to be the only curative therapy for pancreatic cancer, only 20 % of patients have resectable disease at the time of diagnosis [5, 6]. In addition, advanced pancreatic cancer patients exhibit a median survival time (MST) of approximately six months and a 5-year overall survival rate of less than 5 %, despite efforts to manage the tumors with chemotherapy, radiotherapy and other treatments [3, 5–8].

In 1997, Burris et al. reported that gemcitabine monotherapy is superior to fluorouracil (5-FU) monotherapy for

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Table 1 Chemotherapy for advanced pancreatic cancer

	Median survival time (months)	Overall response rate (%)	Trial name	References
Gemcitabine	5.65	5.4		<i>J Clin Oncol</i> 1997;15: 2403–13.
Gemcitabine + erlotinib	6.24	8.6	NCIC CTG PA.3	<i>J Clin Oncol</i> 2007;25: 1960–6.
FOLFIRINOX	11.1	31.6	ACCORD 11	<i>N Engl J Med</i> 2011;364: 1817–25.
Nab-paclitaxel + gemcitabine	8.7	29.2	MPACT trial NCT00844649	<i>N Engl J Med</i> 2013;369: 1691–703.
Gemcitabine +TS-1	10.1	29.3	GEST trial	<i>J Clin Oncol</i> 2013; 31:640–8.

treating pancreatic ductal adenocarcinoma (PDAC) [9]. Gemcitabine monotherapy has subsequently become the standard chemotherapy for PDAC, resulting in an MST of 5.65 months (Table 1). Currently, three protocols have proven to be superior to gemcitabine monotherapy. Combining gemcitabine with erlotinib improved the MST of PDAC to 6.24 months in the NCIC CTG PA3 trial [10], while combining gemcitabine with nab-paclitaxel improved the MST to 8.7 months in the MPACT trial [11]. FOLFIRINOX achieved the longest MST for PDAC (11.1 months) in the ACCORD11 trial [12], and the GEST study obtained similar clinical outcomes. S-1 is an oral fluoropyrimidine derivative that has been shown to be effective against various cancers, and a previous study found that it is at least as effective as gemcitabine against PDAC [13]. In addition, treatment with a combination of gemcitabine + S-1 has been demonstrated to result in an MST of 10.1 months [14]. Although these chemotherapies extend the survival period among PDAC patients, they also result in serious adverse events. Therefore, the optimal chemotherapy regimen for PDAC depends on the patient's performance status.

There have been numerous attempts to develop vaccine therapies for cancer over the past century [2, 3]. Although clinical trials of such vaccines have obtained promising results in specific patients, none of the tested vaccines has exhibited significant improvements in efficacy compared with established therapies. In addition, several issues must be resolved before vaccine therapies can be used in the clinical setting. Tumor-associated antigens (TAA) have been demonstrated to recognize specific human leukocyte antigens (HLA) [15]. Theoretically, the tumor lysate contains all of the antigens expressed by the tumor, and cytotoxic T lymphocytes (CTL) are capable of recognizing some of these antigens [16]. All vaccines for pancreatic cancer are based on the fact that CTL recognize TAA expressed on tumor cells and subsequently attack these cells. The question is how strongly and specifically each TAA stimulates CTL in vivo in the clinical setting. Immune tolerance can develop via various mechanisms, including the downregulation of the major histocompatibility complex (MHC) molecule expression, induction of

T cell anergy, reductions in the number of immune effectors and increases in the number of regulatory T cells [17, 18], which may explain why no cancer vaccine therapy has been established as a standard treatment for advanced PDAC. Therefore, in this study, we comprehensively reviewed the clinical outcomes of vaccine therapy against advanced PDAC.

Peptide-based vaccines developed within the past few years

MUC1

Mucin 1, cell surface associated, (MUC1) is a type I transmembrane protein containing multiple tandem repeats of a 20-amino acid sequence. Several MUC1 peptides have been tested as vaccines in the clinical setting; however, most of them have failed to activate CTL [19–21]. Ramanathan et al. [22]; Yamamoto et al. [23] injected pancreatic patients with a vaccine containing a 100-mer extracellular tandem repeat domain of MUC1 and Montanide ISA-51, and both studies obtained similar clinical responses; i.e., the authors detected cytokines (interferon (IFN)- γ or interleukin (IL)-4) and anti-MUC1 antibodies in the patients' sera but did not observe any significant clinical effects. Another recent study involving a vaccine based on a different MUC1 epitope showed similar clinical outcomes, i.e., all seven patients had progressive disease (PD), although some of the patients exhibited immunological responses, such as IFN- γ and granzyme B secretion [24].

K-RAS mutants

K-RAS mutations are frequently found in patients with PDAC. Vaccines targeting mutations in codon 12 of the K-RAS gene have been tested as treatments for advanced [25] or postoperative [26] PDAC in the clinical setting. Gjertsen et al. [[21]] investigated the utility of a K-RAS peptide vaccine containing granulocyte-macrophage colony-stimulating factor (GM-CSF) in 10 patients who had undergone potentially curative

resection (CTN RAS 95002) and 38 patients with advanced disease (CTN RAS 97004). In that study, one patient achieved a partial response (PR), which lasted for 28 months, and the MST of the immunological responders was 4.9 months, compared to 2.0 months for the non-responders.

Human telomerase reverse transcriptase (hTERT)

Human telomerase reverse transcriptase (hTERT) is frequently expressed in cancer cells [27]. hTERT maintains functional telomeres at the end of chromosomes, which protect against cell senescence [28]. A vaccine against pancreatic cancer containing the telomerase peptide GV1001: hTERT (611-626) and GM-CSF was examined by Bernhardt et al. [29], who found the MST of the immunological responders and non-responders to be 7.2 and 2.9 months, respectively.

Vascular endothelial growth factor receptor 2 (VEGFR2)

Vascular endothelial growth factor (VEGF) plays an important role in the progression of PDAC. The type 2 VEGF receptor (VEGFR2) is expressed in PDAC and associated with tumor neovascularization. Miyazawa et al. [30] investigated the efficacy of combined treatment consisting of PDAC with a VEGFR2-169 peptide-based vaccine and gemcitabine chemotherapy and reported that one patient achieved a PR, while the disease control rate was 67%. In addition, the MST was 7.7 months, although 15/18 patients were chemotherapy naive.

G17DT (gastrimmune)

Gastrin is expressed in PDAC and plays a role in regulating the autocrine, paracrine and endocrine systems [31]. The administration of the anti-gastrin immunogen G17DT results in increased serum antibody levels and reduced tumor growth in patients with gastrointestinal malignancies [32]. A randomized, double-blind, placebo-controlled multicenter trial of G17DT was also recently performed [33]. Although, among the intention to treat (ITT) population, no significant differences in MST were detected between the PDAC patients treated with G17DT and those given the placebo, the MST of the two groups differed significantly after excluding major protocol violators and censoring for chemotherapy.

Heat shock protein (HSP)

Heat shock protein (HSP) itself is not an immunogen; however, it acts as a chaperone or carrier of antigenic peptides and possesses a repertoire of cellular peptides for

pancreatic cancer [34]. Furthermore, HSPPC-96 (Onco-phage) has been tested as a vaccine in the adjuvant setting after complete resection of PDAC [35]. In the latter study, the MST of PDAC was reported to be 2.9 months after surgery; however, this did not result in further clinical studies because only two of 10 patients exhibited increased enzyme-linked immunospot (ELISPOT) reactivity.

Biological vaccines

Fowlpox viral vaccine

Carcinoembryonic antigen (CEA) and MUC1 are highly expressed in PDAC [36]. Viral vectors carrying CEA, MUC1 and TRICOM [a triad of costimulatory molecules: B7.1, intercellular adhesion molecule 1 (ICAM-1) and lymphocyte function-associated antigen 3 (LFA-3)] have been investigated as vaccines against advanced PDAC [37]. In one study, a vaccinia viral vector was used for the initial T cell priming, and a fowlpox viral vector was used for immune boosting. Although this treatment resulted in an MST of 6.3 months (1.5–21.1 months), the five patients who showed T cell responses achieved a longer survival period than the five patients who did not (15.1 and 3.9 months, respectively; $P = 0.002$) [38]. It should be noted that GM-CSF was used as a vaccine adjuvant in the latter trial (Table 2).

Live-attenuated, double-deleted (LADD) *Listeria monocytogene* vaccine

ANZ-100 is a live-attenuated double-deleted *Listeria monocytogene* strain (LADD; Lm Δ actA/ Δ inlB) found to induce a local proinflammatory response, resulting in the activation of innate and adaptive effector cells [39]. Mesothelin is expressed in PDAC and plays an important role in tumor progression [40]. CRS-207 is a LADD Lm strain that delivers mesothelin antigens into class I and II antigen-processing pathways [41]. In a study examining the utility of CRS-207 as a treatment for advanced cancer, three of the seven subjects with PDAC were long-term survivors, although the detection of a mesothelin-specific T cell response was not correlated with survival [41].

Recent vaccine therapies

WT1

Kobayashi et al. reported a retrospective analysis of 255 advanced PDAC patients who were treated with dendritic

Table 2 Peptide-based vaccines and biological vaccines for advanced pancreatic cancer

Author	Journal	Antigen peptide	Sequences	Combination	Patients	Outcome/MST
Yamamoto	Anticancer Res. 2005;25:3575–9	MUC1	10-mer extracellular tandem repeat domain: (GVTSAPDTRPAPGSTAPPAH) ₅	Montanide ISA-51	6	1/6 SD
Rong	Clin Exp Med. 2012;12:173–80	MUC1	PDTRPAPGSTAPPAHGVTSA	DC cells	7	All PD
Gjertsen	Int J Cancer. 2001;92:441–50	K-ras	KLVVVGAGGVGKSALTI Asp: D Arg: R Val: V Cys: C	GM-CSF	38	1 PR 10 SD (10.2 M; 3-23 M) 27 PD 4.9 M responders 2.0 M non-responders
Abou-Alfa	Am J Clin Oncol. 2011;34:321–5	ras12R ras12 V ras12D Wild-type ras	TEYKLVWGARGVVGKSALTIQ TEYKLVWGAVGVGKSALTIQ TEYKLVWGADGVGKSALTIQ TEYKLVWGAGGVGKSALTIQ	hGM-CSF	24	Postoperative adjuvant treatment
Bernhardt	Br J Cancer. 2006;95:1474–82	Telomerase hTERT (611–626)	GV1001; EARPALLTSRLRFIPK	GM-CSF	38	7.2 M (24 responders) 2.9 M (14 non-responders)
Miyazawa	Cancer Sci. 2010;101:433–9	VEGFR2-169	RFVDPGNRI	Gemcitabine	18	7.7 M
Gilliam	Pancreas. 2012;41:374–9	Anti-gastrin G17DT Gastrimmune	EGPWLEEEEEAYGWMDf-DT (diphtheria toxoid)	G17DT vs. placebo	152	5.0 M vs 2.8 M
Maki	Dig Dis Sci. 2007;52:1964–72	HSP HSPPC-96 (gp96, Oncophage)			10	Postoperative adjuvant treatment 2.7 Y
Kaufman	J Transl Med. 2007;5:60	MUC1 and CEA	CEA agonist peptide CAP1-6D (YLSGADLNL) MUC-1 agonist peptide P-93L (ALWGQDVTSV)	B7.1, ICAM-1, LFA-3 (TRICOM) Vaccinia virus: PANVAC-V Fowlpox virus: PANVAC-F GM- CSF	10	6.3 M
Le	Clin Cancer Res. 2012;18:858–68	Listeria vaccine ANZ- 100, CRS-207			9 vs. 17	NA

Table 3 Recently developed peptide-based vaccines and multiple vaccines for advanced pancreatic cancer

Author	Journal	Antigen peptide	Sequences	Restricted HLA	Combination	Patients	Outcome/MST
Kobayashi	Cancer Immunol Immunother. 2014;63:797–806	WT1 MUC1	CYTWNQMNL RMFPNAPYL TRPAPGSTAPPAHG-VTSAP DTRPAPGSTAP	A24:02 A02:01/02:06 Any A	DC cells OK432	255	9.9 M 10.4 M (erythema)
Nishida	J Immunother. 2014;37:105–14	WT1	CYTWNQMNL	A24:02	Weekly 1000 mg/m ² GEM	31	8.1 M 10.9 M (DTH)
Asahara	J Translation Res. 2013;11:291	KIF20A-66	KVYLRVRPLL	A2402	Montanide ISA51VG	31	4.7 M 6.1 M (reaction)
Suzuki	J Immunother. 2014;37:36–42	KIF20A-10-66	KVYLRVRPLL	A2402	Montanide ISA51VG	9	5.8 M
Geynisman	J ImmunoThera Cancer. 2013;1:8	CEA CAP1-6D	YLSGADLNL	A2	Montanide/GM-CSF	19	11.1 M
Kameshima	Cancer Sci. 2013;104:124–9	SVN2B	AYACNTSTL	A2402	Montanide/IFN- α c	6	(9.6 M)
Yutani	Oncology Reports. 2013;30:1094–100	31 vaccine peptides		A2, A24, A3, A26	Mono: 8 Chemo: 33	41	7.9 M 9.6 M (chemo)
Kimura	Pancreas. 2012;41:195–205	WT1, Her2, CEA, MUC1, CA125, autologous tumor lysate			DC cells plus LAK plus GEM and S1 OK432	49	S: 8.0 M G: 12.0 M GS + LAK: 16.9 M
Le	J Clin Oncol. 2014;32(suppl 3):Abstract 177	GVAX pancreas and CRS-207 vs. GVAX pancreas alone	Irradiated GM-CSF-secreting allogeneic pancreatic tumor vaccine (GVAX pancreas)		Cyclophosphamide	90	6.1 M vs. 3.9 M 9.7 M (3 or more rounds of vaccine therapy)

cell (DC) vaccines containing Wilms tumor 1 (WT1) and MUC1 after being recruited from seven institutions that followed a unified standard operating procedure. The MST of these patients was 9.9 months [42]. Nishida et al. also examined the utility of chemo-vaccine therapy in which a WT1-based vaccine was used in combination with the administration of 1,000 mg/m² of gemcitabine weekly. The latter regimen resulted in an MST of 8.1 months among 31 advanced PDAC patients [43]. In addition, the MST of the immunological responders in these two studies was very similar (10.4 and 10.9 months, respectively) (Table 3).

KIF20A

Kinesin family member 20A (KIF20A) plays an important role in the trafficking of molecules and organelles [44] and is one of the molecules targeted by vaccines against PDAC. A KIF20A vaccine was recently tested using different regimens, including vaccine monotherapy [45] and chemo-vaccine therapy involving gemcitabine [46], and similar MST values were reported in both studies (4.7 and 5.8 months, respectively).

Carcinoembryonic antigen (CEA)

CEA is a 180-kDa immunoglobulin-like molecule expressed on the surface of 90 % of PDAC tumor cells [47]. CAP1-6D, a modified CEA peptide, was combined with Montanide/GM-CSF to produce a vaccine against pancreatic cancer that was subsequently tested in advanced PDAC patients [48]. The MST of the 19 patients was 11.1 months, and one patient, randomized into the 0.01 mg arm, achieved a complete response (CR).

Survivin2B

Survivin is a member of the inhibitors of apoptosis (IAP) family of proteins that protect apoptotic signals by inhibiting the caspase activity [49, 50]. Hence, survivin-expressing cancer cells escape from apoptosis and do not die. Using a peptide-binding assay, we found that the survivin2B 80–88 peptide induces a strong CTL response [51]. We also examined the effects of a survivin2B 80–88 peptide-based vaccine on various cancers in the clinical setting and obtained promising outcomes. In particular, the anti-tumor effect of the survivin2B 80–88 peptide was enhanced by combining it with incomplete Freud's adjuvant and IFN- α injection. Our preliminary clinical study demonstrated a 66.6 % disease control rate in advanced PDAC patients (four of six patients) [52]. Moreover, the PDAC patients in our recent clinical phase I study exhibited an MST of 9.6 months.

Table 4 Evaluation of therapeutic activity in solid tumors

Method	WHO	RECIST	IIRC
	Sum of the products of the two longest perpendicular dimensions (bidimensional)	Sum of the longest dimensions (unidimensional)	Sum of the products of the two longest perpendicular dimensions (SPD) of all index lesions. (bidimensional)
No. of measured lesions	All lesions	Five per organ, 10 in total	Five per organ, 10 in total, and five cutaneous index lesions
CR	Disappearance of all known disease, confirmed at 4 weeks	Disappearance of all known disease, confirmed at 4 weeks	Disappearance of all known disease, confirmed at 4 weeks apart
PR	>50 % decrease in total tumor size, confirmed at 4 weeks	>30 % decrease in total tumor size, confirmed at 4 weeks	>50 % decrease in tumor burden compared with baseline in two observations at least 4 weeks apart
SD	CR, PR, and PD criteria not met	CR, PR, and PD criteria not met	CR, PR, and PD criteria not met
PD	>25 % increase in total tumor size; no CR, PR, or SD documented before increase in tumor size; new lesion (s); > 25 % increase in size of one lesion	>20 % increase in total tumor size; no CR, PR, or SD documented before increase in tumor burden; new lesion (s)	>25 % increase in tumor burden compared with nadir (at any single time point) in two consecutive observations at least 4 weeks apart

Tumor burden = SPD_{index lesions} + SPD new, measurable lesions

Multiple vaccines

Personalized peptides

In a previous study, a set of 31 peptides was used to create personalized vaccines for advanced PDAC [53]. A maximum of four peptides were selected from among the 31-peptide set based on the results of HLA typing and the patients' peptide-specific IgG titers. Eight patients received vaccine monotherapy, and 31 patients received chemo-vaccine therapy. In the chemo-vaccine therapy group, gemcitabine was administered in eight patients, S-1 was administered in six patients and gemcitabine + S-1 was given in eight patients. The overall MST was 9.6 months, although that of the patients who underwent monotherapy was 7.9 months. Yanagimoto et al. reported similar clinical outcomes for chemo-vaccine therapy involving personalized vaccines and gemcitabine based on the same regimen [54]. The MST of the patients in the latter study was 9.0 months, although that of the immunological responders was 15.5 months. None of the patients in either study achieved CR (Table 3).

Autologous tumor lysate combined with lymphokine-activated killer cell therapy

Kimura et al. treated 49 PDAC patients with vaccines based on five different peptides and autologous tumor lysate, although the vaccine preparation regimens and anti-tumor therapies varied in each case [16]. Two patients achieved CR after treatment with a combination of DC cell and lymphokine-activated killer cell (LAK) therapy. The MST of the patients treated with LAK + gemcitabine and S-1 was 16.9 months, whereas that of all patients was 12.0 months. It should be noted that the survival time was calculated from the day after the first vaccination, which may have resulted in a shorter survival time (by a couple of months) than would have been obtained using the methods employed in other studies. It is very difficult to evaluate the clinical results of this study due to the effects of the different therapeutic strategies used in each case. However, the fact that multiple patients achieved CR will encourage researchers to pursue this approach further.

GVAX pancreas with CRS-207

GVAX is a series of irradiated GM-CSF-secreting allogeneic pancreatic cell lines that elicit broad antigenic responses. CRS-207 is a LADD Lm strain (Lm Δ actA/ Δ inlB) that expresses mesothelin and stimulates the innate and adaptive immune systems. A phase II randomized control trial of GVAX pancreas combined with CRS-207 versus GVAX pancreas alone was presented at the 2014

American Society of Clinical Oncology (ASCO) Gastrointestinal Cancers Symposium [55]. Interestingly, the clinical results demonstrated that both treatments had dose-dependent survival benefits. The MST of the patients who received three or more rounds of vaccine therapy was 9.7 months, and the MST of the GVAX with CRS-207 arm was longer than that of the GVAX-alone arm (6.1 vs. 3.9 months; $P = 0.01$) [56].

Evaluation of therapeutic activity in solid tumors

The response of solid tumors is evaluated using either the WHO [57] or RECIST criteria [58]. The RECIST criteria were developed because the WHO criteria are quite complex and measuring all visible lesions in two dimensions is both time consuming and subject to measuring bias [59]. However, the use of immunotherapeutic agents in cancer patients is associated with the following problems: (a) The measurable anti-tumor activity can take longer to appear during immunotherapy than during cytotoxic therapy; (b) Responses to immunotherapy can occur after the standard criteria for progressive disease (PD) have been met; (c) Discontinuing immunotherapy may not be appropriate in some cases, unless PD is confirmed; (d) Allowing for "clinically insignificant" PD (e.g., small new lesions developing in the presence of other responsive lesions) is recommended; and (e) Durable stable disease (SD) may represent the anti-tumor activity [60]. Therefore, the immune-related response criteria (irRC) were developed to evaluate the immunotherapeutic activity in solid tumors [61]. The most important aspects of the irRC criteria are that (a) new lesions are not classified as PD and (b) two consecutive observations obtained at least four weeks apart are required to diagnose PD. However, the clinical utility of the irRC remains unclear and these criteria may require further optimization [61] (Table 4).

Future research topics

Initial time point for survival assessments

The initial time point for survival assessments should be unified to allow clinical outcomes to be compared between studies. Most PDAC patients already have advanced disease at the time of diagnosis [6]. In addition, the adverse effects of chemotherapies differ markedly among the various regimens [8]. Therefore, the status of PDAC patients at the time point at which they are registered can differ both within and between clinical studies. Kobayashi et al. reported that the MST from the date of diagnosis and the MST from the first vaccination are very different (16.5 vs.

9.9 months) [42]. Therefore, MST data must be interpreted carefully.

Vaccine therapy and chemotherapy

The goal of vaccine therapy for cancer is to increase the native immunity of cancer patients. However, chemotherapy causes irreversible damage to proliferating cancer cells as well as immune cells, including T and B cells. Therefore, there is a conflict between the fundamental principles of these two treatments. Chemotherapy is currently the gold standard treatment for advanced PDAC. Although the biological mechanisms of vaccine therapy and chemotherapy conflict with each other, the anti-cancer activity of vaccine monotherapy or chemo-vaccine combination therapy should be greater than that of chemotherapy alone.

Slow clinical response to vaccine therapy

It is very hard to achieve a complete response (CR) with vaccine therapy alone. We reviewed 19 studies involving a total of 860 patients and found that CR responses were obtained in only three cases. Although none of these studies involved a large number of patients, the poor reported response rates are a concern. One of the patients who achieved a CR was administered CEA CAPI-6D + Montanide/GM-CSF therapy, while the other two were treated with WT1, Her2, CEA, MUC1, cancer antigen 125 and autologous tumor lysate vaccines combined with DC cell-based LAK therapy and chemotherapy. Immunological responses require a long time to control tumor growth and achieve remission. The primary goal of vaccine therapy is to achieve long-term SD [62]. Most previous clinical studies of PDAC involved patients with advanced disease for whom no other therapies were available. Therefore, vaccine therapy may be suitable for patients in other clinical stages or possibly a useful postoperative adjuvant therapy. The main advantage of vaccine therapy is that it has few adverse effects, although it has also demonstrated minimal clinical effects in previous trials. We are currently conducting a phase II study of the survivin2B 80–88 peptide + Montanide + IFN- β as a treatment for PDAC (SUCCESS, Study of Unresectable CanCER with Survivin-2B peptide vaccine in Sapporo: UMIN000012146), in which half of the required patients have been recruited. The clinical results of the SUCCESS phase II study will be reported by the end of next year.

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Phase I Clinical Study of Survivin-Derived Peptide Vaccine for Patients with Advanced Gastrointestinal Cancers

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Abstract

Survivin is a member of the Inhibitor of Apoptosis Protein (IAP) family. It is expressed in fetal tissues but not in normal adult tissues. Since Survivin is over expressed in various types of tumor tissues as well as tumor cell lines, it is considered to be suitable as a target antigen for cancer vaccine therapy. We identified an HLA-A24-restricted antigenic peptide, SVN-2B (AYACNTSTL), derived from a splicing variant of Survivin-2B. In the present study, we carried out a phase I clinical study assessing the safety and efficacy of vaccination with the peptide in patients having advanced gastrointestinal cancer. Vaccinations with 0.1mg, 1.0mg, or 3.0mg doses of the SVN-2B peptide were given subcutaneously four times at 14-day intervals. In 20 patients who received at least one vaccination, grade 1 and grade 2 treatment-related adverse events were observed, including injection site extravasation (grade 2), injection site reaction (grade 1), skin induration (grade 1) and fever (grade 1). No severe adverse event was observed in any patient. Based on tumor size evaluated by computed tomography, eight of the 15 patients who completed the vaccination schedule were considered to have stable disease as assessed by the RECIST criteria. Analysis of peripheral blood lymphocytes using HLA-A24/peptide tetramers revealed the highest increase of SVN-2B-specific cytotoxic T lymphocyte frequency in the 1.0mg dose group. The present clinical study indicates that SVN-2B peptide vaccination is safe and can be considered a potent immunotherapy for HLA-A24-positive gastrointestinal cancer patients.

Keywords

Survivin, Cancer vaccine, Gastrointestinal cancer, Tetramer, Phase I trial

Abbreviations

IAP: Inhibitor of Apoptosis Protein, CTLs: Cytotoxic T lymphocytes, HLA: Human Leukocyte Antigen, CT: Computed Tomography, PBLs: Peripheral Blood Lymphocytes, AEs: Adverse Events, HIV: Human Immunodeficiency Virus, PD: Progressive Disease, SD: Stable Disease, IFN: Interferon

Introduction

Cytotoxic T lymphocytes (CTLs) can recognize MHC class I-bound peptides derived from tumor antigens in cancer cells. Following the first report of the identification of a human tumor antigen, melanoma antigen-1 (MAGE-1), in 1991 [1] a large number of antigenic peptides from various human cancers have been identified [2-7]. They have been employed in immunotherapy for cancer and clinical trials of peptide-based vaccine therapies have taken place [8-11].

We have identified a human leukocyte antigen (HLA)-A24-restricted antigenic peptide, SVN-2B (AYACNTSTL), which was derived from the exon 2B-encoded region of Survivin-2B, a splicing variant of Survivin [12]. Survivin is a member of the inhibitor of apoptosis protein (IAP) family with a single baculovirus IAP repeat domain [13]. It is expressed during fetal development but undetectable in terminally differentiated normal adult tissues. In contrast to normal tissues, Survivin and Survivin-2B are expressed in transformed cell lines and in most common cancers, including gastrointestinal cancer and pancreatic cancer [13,14]. We reported previously that SVN-2B peptide-specific CTLs were increased by

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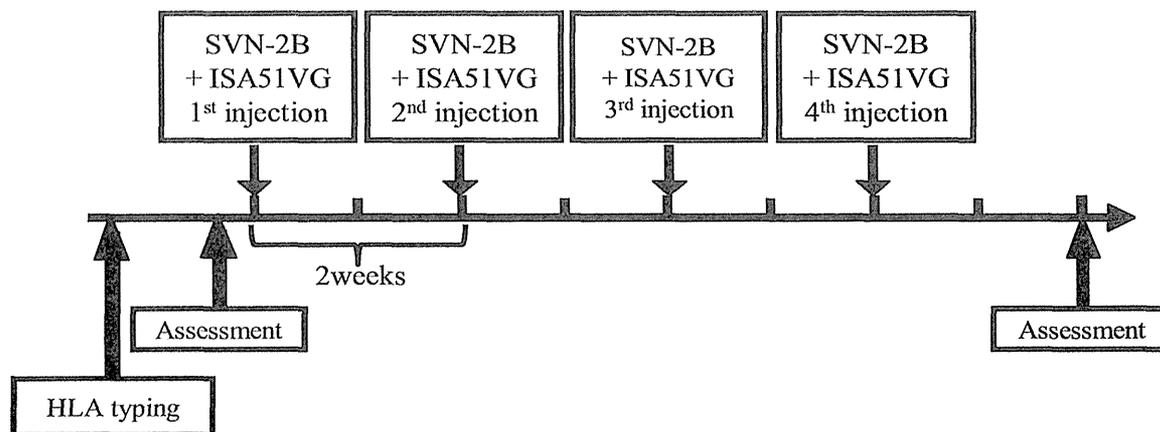


Figure 1: Protocols of the clinical study

The SVN-2B peptide at a dose of 0.1mg/1mL, 1mg/1mL, or 3mg/1mL was emulsified with Montanide ISA51VG at a volume of 0.8mL immediately before vaccination. The patients were then vaccinated subcutaneously (s.c.) four times at 14-day intervals. Tumor size and the immunological response were evaluated before treatment and at two weeks after the 4th vaccination.

stimulating peripheral blood lymphocytes (PBLs) of cancer patients with the peptide *in vitro* [15]. The induced CTLs showed specific cytotoxicity against HLA-A24-positive cancer cells [15-17]. We have carried out clinical trials of SVN-2B vaccination. The SVN-2B peptide was given subcutaneously to patients six times or more at biweekly intervals for colon, breast, oral cavity, and urinary bladder cancer patients [18-24]. There were no severe adverse effects and, clinically, certain patients showed reductions in tumor markers and tumor size as assessed by Computed Tomography (CT). In the present clinical study, we reevaluated the safety and efficacy of SVN-2B vaccination in accordance with good clinical practice guidelines and evaluated the optimal dose of the peptide.

Methods

Patient selection

The study protocol was approved by the Institutional Review Board of Sapporo Medical University. All patients gave informed consent before being enrolled. This study was conducted in accordance with the International Conference on Harmonisation E6 requirements for Good Clinical Practice and with the ethical principles outlined in the Declaration of Helsinki.

Patients enrolled in this study were required to conform to the following criteria: (1) to have histologically confirmed gastrointestinal, bile duct, or pancreatic cancer, (2) to be HLA-A*2402 positive, (3) to have Survivin-positive cancer tissue confirmed by immunohistochemical staining, (4) to be between 20 and 85 years old, (5) to have lesions measurable by CT at the time of registration, (6) to have a history of standard chemotherapy, (7) to have grade 0 or 1 in Eastern Cooperative Oncology Group (ECOG) performance status, and (8) to have no serious organ failure within 30 days at the time of registration.

Exclusion criteria included: (1) prior cancer therapy such as chemotherapy, radiation therapy or other immunotherapy within the previous 4 weeks, (2) presence of other cancers that might influence the prognosis, (3) administration of immunosuppressive drugs such as systemic steroid therapy, (4) severe cardiac insufficiency, acute infection, or hematopoietic failure, (5) uncontrollable diabetes or hypertension, (6) pregnancy or ongoing breast-feeding, and (7) unsuitability for the trial based on clinical judgment. In addition, patients with a high frequency of the peptide-specific CTLs at the time of registration were excluded since such patients were poor responders to the vaccine in our previous studies [23,24]. The number of the HLA-A24/SVN-2B peptide tetramer-positive CTLs per 10,000 CD8-positive T cells (CTLpre) was analyzed at the time of registration

and patients who had a value of log₁₀ (1+CTLpre) higher than 1.6 were excluded.

Peptide preparation

The peptide SVN-2B with the sequence AYACNTSTL was prepared under good manufacturing practice conditions by PolyPeptide Laboratories San Diego (San Diego, CA, USA). The identity of the peptide was confirmed by mass spectral analysis, and the purity was shown to be more than 98% as assessed by high pressure liquid chromatography analysis. The peptide was supplied as a freeze-dried, sterile white powder. It was dissolved in 1.0 ml of physiological saline (Ohtsuka Pharmaceutical Co., Ltd., Tokyo, Japan) and stored at -80°C until just before use.

Patient treatment

This study was carried out as an open-label, randomized parallel group study at the Department of Surgery, Surgical Oncology and Science of Sapporo Medical University Hospital to evaluate the safety and efficacy of the SVN-2B peptide vaccine for patients who had advanced or recurrent gastrointestinal or pancreatic cancer (UMIN000008611). The patients were randomly assigned into the following three dosage groups: group 1 patients received 0.1mg, group 2 received 1.0mg and group 3 received 3mg. Each group included five patients. SVN-2B at a dose of 0.1mg/1mL, 1mg/1mL, or 3mg/1mL was emulsified with Montanide ISA51VG (Seppic, Paris, France) at a volume of 0.8mL immediately before vaccination. The patients were then vaccinated subcutaneously (s.c.) four times at 14-day intervals (Figure 1).

Toxicity evaluation

Patients were examined closely for signs of toxicity during and after vaccination. Adverse events (AEs) were recorded using CTCAE (version 4.03) criteria and graded for severity.

Clinical response evaluation

Physical and hematological examinations were conducted before and after each vaccination. Changes in tumor marker levels (CEA and CA19-9) were evaluated by comparison of the serum levels before the first vaccination and those after the fourth vaccination. Tumor size was evaluated by CT scans before treatment and at two weeks after the fourth vaccination (Figure 1). The antitumor response was evaluated according to the Response Evaluation Criteria in Solid Tumors (RECIST: version 1.1) guideline [25]. Briefly, a complete response (CR) was defined as complete disappearance of all measurable and evaluable disease. A partial response (PR) was defined as a $\geq 30\%$

Table 1: Profiles of patients in the full analysis set for safety assessment (N=20)

Clinical variables		0.3 mg (n=7)	1.0 mg (n=7)	3.0 mg (n=6)	Total (n=20)
Gender	Men: Women	2:5	5:2	3:3	10:10
Age	Median (min-max)	69.5 (53-80)	63.0 (51-84)	64.4 (41-86)	65.1(41-84)
Type of cancer	Pancreatic cancer	5	2	5	12
	Colon cancer	2	3	1	6
	Gastric cancer	0	1	0	1
	Bile duct cancer	0	1	0	1
Metastasis	(positive: negative)	7:0	5:2	5:1	17:3
Prior surgery	(positive: negative)	4:3	5:2	3:3	12:8
Prior radiation therapy	(positive: negative)	2:5	2:5	3:3	7:13
Prior chemotherapy	(positive: negative)	6:1	6:1	6:0	18:2
ECOG PS	(0:1)	1:6	1:6	2:4	4:16
Treatment-related AEs					
Fever	Grade 1	1			1
Injection site extravasation	Grade 2			1	1
Injection site reaction	Grade 1	1		1	2
Skin induration	Grade 1	1	1		2

decrease from baseline in the size of all measurable lesions (sum of maximal diameters). Progressive disease (PD) was defined as an increase in the sum of maximal diameters by at least 20% or the appearance of new lesions. Stable disease (SD) was defined as the absence of criteria matching those for CR, PR or PD. Patients who received fewer than four vaccinations were excluded from clinical response evaluations in this study.

In vitro stimulation of PBLs

PBLs were isolated by Ficoll-Conray density gradient centrifugation using Lymphoprep (AXIS-SHIELD, Oslo, Norway). They were then frozen and stored at -80°C. The frozen PBLs were thawed and incubated in the presence of 40µg/mL SVN-2B in AIM-V medium (Life Technologies, Carlsbad, CA, USA) containing 10% human serum at room temperature. Next, interleukin-2 was added at a final concentration of 50 U/mL 1 hour, 2 days and 4 days after addition of the peptide. On day 7 of culture, the PBLs were analyzed by tetramer staining assay and ELISPOT assay.

Tetramer staining

FITC-labeled HLA-A*2402/human immunodeficiency virus (HIV)-derived peptide (RYLRDQQLL) and PE-labeled HLA-A*2402/SVN-2B peptide tetramers were purchased from MBL, Inc. (Nagoya, Japan). For flow cytometric analysis, PBLs, which were stimulated *in vitro* as above, were stained with the FITC-labeled tetramer and PE-labeled tetramer at 37°C for 20 min, followed by staining with a PC5-labeled anti-CD8 monoclonal antibody (Beckton Dickinson Biosciences, San Jose, CA, USA) at 4°C for 30 min. The cells were washed twice with PBS before fixation in 1% formaldehyde. Flow cytometric analysis was performed using FACSCalibur and CellQuest software (Beckton Dickinson Biosciences). The frequency of CTL precursors was calculated as the number of HLA-A24/SVN-2B tetramer-positive cells per 10,000 CD8-positive cells.

ELISPOT assay

ELISPOT plates were coated sterily overnight with an IFN-γ capture antibody (Beckton Dickinson Biosciences) at 4°C. The plates were then washed once and blocked with AIM-V medium containing 10% human serum for 2 h at room temperature. CD8-positive T cells separated from patients' PBLs (5x10³ cells/well), which were stimulated *in vitro* as above, were then added to each well along with HLA-A24-transfected CIR cells (CIR-A24) (5x10⁴ cells/well) preincubated with SVN-2B (10ng/mL, 100ng/mL, 10µg/mL) or the HIV peptide (RYLRDQQLL) as a negative control. After incubation in a 5% CO₂ humidified chamber at 37°C for 24 h, the wells were washed

vigorously five times with PBS and incubated with a biotinylated anti-human IFN-γ detection antibody (Beckton Dickinson Biosciences) and horseradish peroxidase-conjugated avidin. Spots were visualized and analyzed using KS ELISPOT (Carl Zeiss, Jena, Germany).

Immunohistochemistry

Immunohistochemical study of the HLA class I expression in the patients' primary cancer tissues was done with anti-HLA class I heavy chain monoclonal antibody EMR8-5 according to the standard methods described previously [26].

Statistical analysis

All statistical analyses were done using SAS Version 9.3 and JMP Version 11.0 (SAS Institute, Inc.). For the tetramer assay, statistical analysis was performed using a one-sided t-test. Statistical analysis of ELISPOT assay was performed using the student t-test.

Results

Patient profiles

From August 2012 to May 2013, 38 patients were assessed for eligibility and 21 patients were initially enrolled in this trial (Figure 2). However, one patient was withdrawn before the first vaccination due to deterioration of the systemic condition. Twenty patients who received at least one vaccination were evaluated for safety as a full analysis set (FAS). Five patients discontinued halfway through the protocol due to progression of the disease. None of the interruptions was due to treatment-related AEs. Fifteen patients received the complete regimen including four vaccinations and were evaluated for efficacy of the vaccine (Figure 2). The patient profiles are shown in Table 1. The primary malignant tumors of the 20 patients were 12 pancreatic cancers, 6 colon cancers (including 2 appendix cancers), 1 gastric cancer and 1 bile duct cancer.

Safety

Peptide vaccination was well tolerated in all patients. The treatment-related AEs are listed in Table 1. They included injection site extravasation (grade 2), injection site reaction (grade 1), skin induration (grade 1) and fever (grade 1). No serious toxicity-associated adverse event was observed during or after the vaccination.

Clinical responses

Table 2 summarizes the clinical outcomes of the 15 patients who received the complete regimen. CT evaluation of tumor size showed that 8 patients had SD and 7 patients PD, although none had PR or

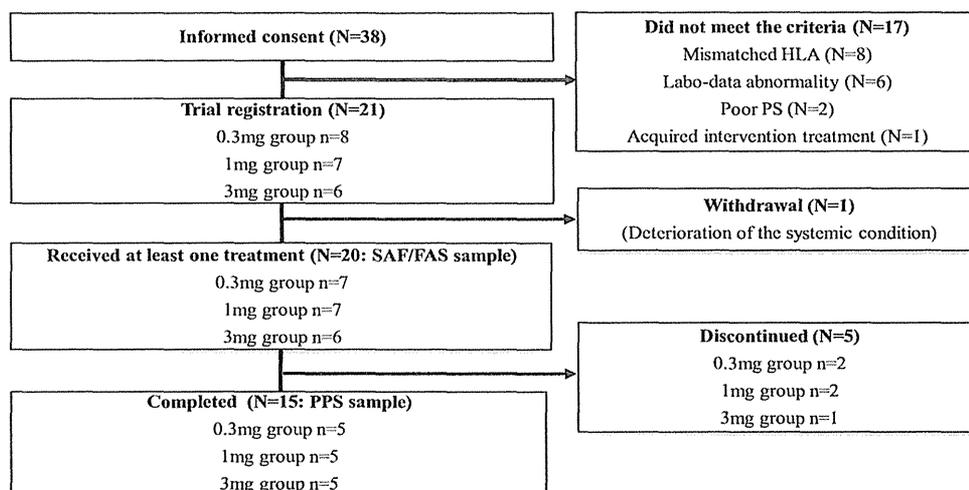


Figure 2: Enrollment of patients

Thirty-eight patients were assessed for eligibility and 21 were initially enrolled in this trial. One patient was withdrawn before the first vaccination due to deterioration of the systemic condition. Twenty patients who received at least one vaccination were evaluated for safety as the full analysis set. Five patients discontinued halfway through the protocol due to progression of the disease. Fifteen patients received the complete regimen and were evaluated for efficacy of the vaccine as the per protocol set. SAF: Safety Analysis Set, FAS: Full Analysis Set, PPS: Per Protocol Set, HLA: Human Leukocyte Antigen, PS: Performance Status.

Table 2: Profiles and clinical outcomes of patients who completed the regimen

Clinical Background						Immunological Response		Antitumor Response		
Dose	Age	Gender	Origin	Status	HLA class I	Tetramer increase	ELISPOT increase	RECIST	CEA	CA19-9
0.3 mg	63	Woman	Pancreas	Inoperable	+	35	-17	SD	Decreased	Decreased
0.3 mg	69	Woman	Pancreas	Inoperable	+	5	-31	SD	WNL	Increased
0.3 mg	53	Woman	Pancreas	Post-op	-	7	6	PD	Increased	Increased
0.3 mg	68	Man	Pancreas	Post-op	+	8	17	PD	Increased	Increased
0.3 mg	78	Man	Colon	Post-op	+	-4	2	PD	Increased	Increased
1.0 mg	61	Man	Pancreas	Inoperable	+	21	-1	SD	Increased	Increased
1.0 mg	84	Woman	Colon	Post-op	+	28	14	SD	Increased	Increased
1.0 mg	69	Man	Stomach	Post-op	+	7	26	SD	Increased	Increased
1.0 mg	59	Man	Colon	Post-op	+	29	16	PD	Increased	Increased
1.0 mg	62	Man	Colon	Post-op	+	15	2	PD	Increased	WNL
3.0 mg	41	Woman	Pancreas	Post-op	+	12	158	SD	WNL	Stable
3.0 mg	66	Man	Pancreas	Inoperable	+	9	19	SD	Decreased	Increased
3.0 mg	64	Man	Pancreas	Post-op	+	2	-16	SD	WNL	Decreased
3.0 mg	50	Man	Pancreas	Post-op	+	9	21	PD	WNL	Increased
3.0 mg	64	Woman	Pancreas	Inoperable	+	0	10	PD	Increased	Increased

Post-op: Post-Operative, SD: Stable Disease, PD: Progressive Disease, WNL: Within the Normal Limit

CR. The disease control rate was 53.3%. Among the 8 patients who were defined as having SD, the CEA levels and the CA19-9 levels were decreased or at least stable during vaccination in 2 patients and 3 patients, respectively. The CEA levels stayed within the normal range (0~5.9ng/ml) throughout the study in 4 patients, and the CA19-9 level stayed within the normal range (0~37 U/ml) in one patient. It was noted that all three patients who had undergone immunotherapy before the registration had PD. Moreover, the result for one patient who had HLA class I-negative cancer tissue was also PD.

Tetramer assay and ELISPOT assay

We investigated whether the SVN-2B peptide vaccination could

actually induce specific immune responses in the enrolled patients. The peptide-specific CTL frequencies in PBLs before the first vaccination (CTLpre) and after the fourth vaccination (CTLpost) were assessed using the HLA-A24/SVN-2B tetramer, and the tetramer increase (CTLpost-CTLpre) was calculated (Table 2). The HLA-A24/HIV peptide (RYLRDQQL) tetramer was used as a negative control. SVN-2B-specific CTL frequencies were increased after the vaccination in all patients except two who had undergone immunotherapy before the registration. We compared the tetramer increases between the PD group (non-responders) and SD group (responders). The mean tetramer increase of the SD group was higher than that of the PD group (Figure 3A), although there was no statistical significance

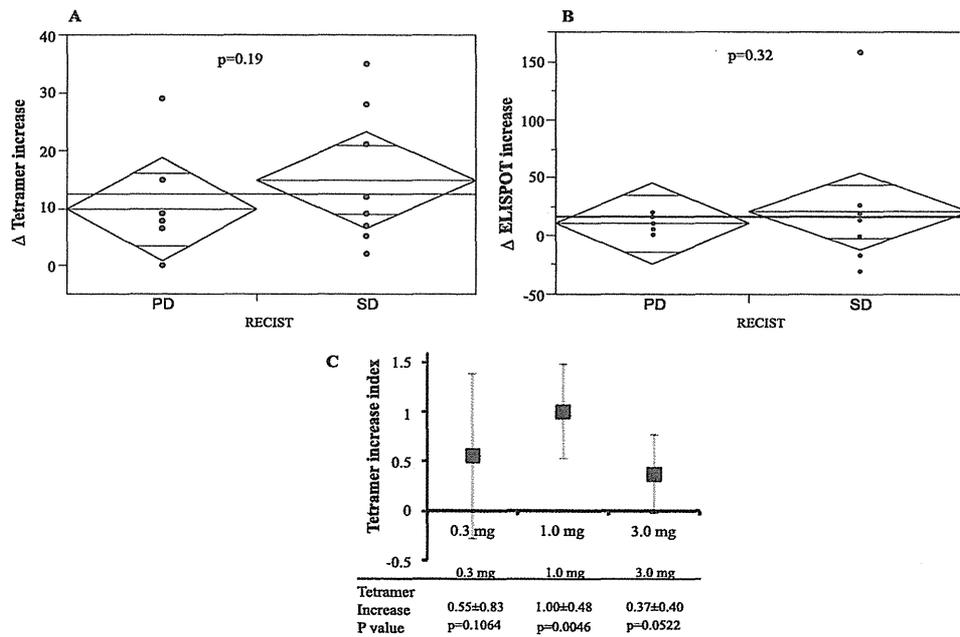


Figure 3: Tetramer assay and ELISPOT assay

(A) The tetramer increase (CTLpost-CTLpre) was calculated from the peptide-specific CTL frequency in PBLs before the first vaccination (CTLpre) and after the fourth vaccination (CTLpost) using the HLA-A24/SVN-2B tetramer. The mean tetramer increases of the PD (non-responders) and SD groups (responders) were compared. (B) The ELISPOT increase was calculated from the numbers of the peptide-specific IFN- γ spots before the first vaccination and after the fourth vaccination. The mean ELISPOT increases of the PD and SD groups were compared. (C) The mean tetramer increase index was calculated according to the following formula: Tetramer increase index= $\text{Log}_{10}(1+\text{CTLpost})-\text{Log}_{10}(1+\text{CTLpre})$. The mean tetramer indices of the three groups (0.1mg dose, 1.0mg dose, and 3.0mg dose) were compared.

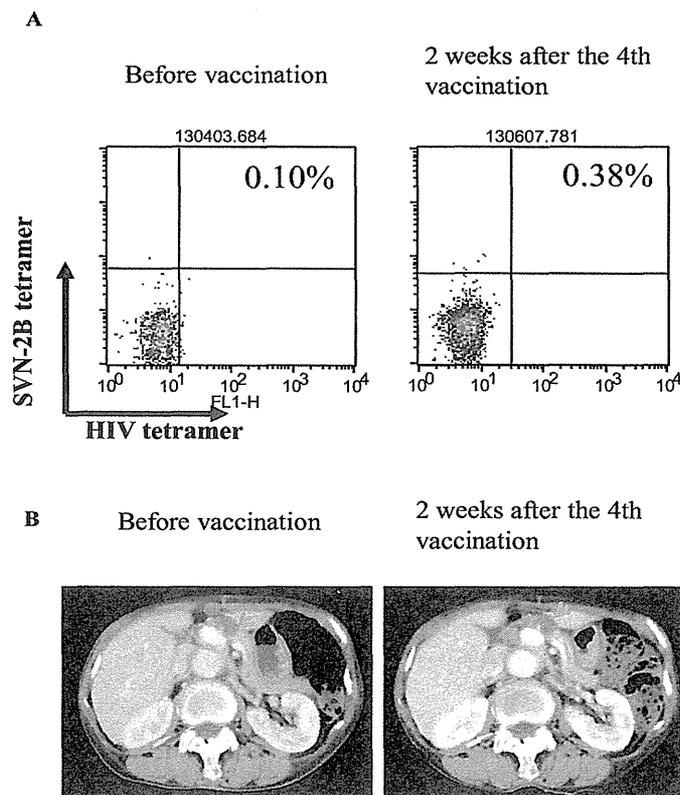


Figure 4: Tetramer assay and CT scan images of the patient with a metastatic pancreatic tumor

An 84-year-old woman with primary colon cancer and metastatic pancreatic tumor. (A) Tetramer assay before vaccination (left panel) and 2 weeks after the 4th vaccination (right panel). (B) CT scan images before vaccination (left panel) and 2 weeks after the 4th vaccination (right panel). The arrowhead indicates the metastatic pancreatic tumor. The tumor grew slightly from 16 mm to 17mm during the 8 weeks of the study.

($p=0.19$). To determine the optimal dose of the peptide to induce specific CTLs in patients, the mean tetramer increase indices of the three groups (0.1mg dose, 1.0mg dose, and 3.0mg dose) were compared (Figure 3C). It was found that 1.0mg was the most effective dose for the induction of peptide-specific T cells after the fourth vaccination ($p=0.0046$).

We also analyzed the peptide-specific IFN- γ responses of CD8-positive T cells by ELISPOT assay. The HIV peptide (RYLRDQQLL) was used as a negative control. The numbers of peptide-specific IFN- γ spots before the first vaccination and after the fourth vaccination were counted, and the ELISPOT increase was calculated (Table 2). There was no significant difference in the mean ELISPOT increase between the SD group and PD group (Figure 3B).

Overall, this study suggests that the immunological response of the vaccine is well represented by tetramer assay rather than ELISPOT assay and that the immunological responses, at least in some patients, appropriately reflect the antitumor responses.

A Case Study

An 84-year-old woman who had primary colon cancer and metastatic liver and pancreatic tumors received the 1.0 mg dose of the SVN-2B vaccine. CT images and tetramer staining data are shown in Figure 4. In this case, the clinical response was defined as SD, and the peptide-specific CTL frequency was increased after the vaccination (Figure 4A). The metastatic pancreatic tumor barely changed from 16 mm to 17 mm during the 8 weeks of the study (Figure 4B). She continued the vaccination after the study. After 6 months, the pancreatic tumor size had increased by 31%, and a new lesion appeared in the caudate lobe of the liver. The time to progression was 267 days. There was no treatment-related AE and she could maintain high quality of daily life for almost one and a half years.

Discussion

The present study demonstrated the safety and clinical efficacy of the survivin-2B peptide vaccine for patients with advanced gastrointestinal cancer. However, the efficacy of vaccination with the SVN-2B peptide plus oil adjuvant Montanide ISA51VG was limited and not sufficient to elicit overt clinical responses. It is obvious that superior clinical and immunological responses are necessary for cancer immunotherapy. It should be considered that vaccination in combination with immunostimulatory adjuvants or cytokines may lead to greater immune and clinical responses. We have reported that type I interferon (IFN) can enhance the antitumor and immunological responses of the peptide vaccine [19,20]. On the basis of the results in this phase I study, we have started a phase II study of the SVN-2B peptide vaccine in combination with IFN- γ .

Immunomonitoring revealed that the tetramer increases were well correlated with antitumor responses as compared with ELISPOT analysis. Therefore, we used the tetramer increase as an index of vaccine-specific immune responses and determined the optimal peptide dose. A significantly higher frequency of tetramer-positive CTLs was induced in the 1mg dose group. However, the optimal dose may vary depending on conditions such as the vaccination interval and combination with distinct adjuvants and/or cytokines, and may have to be reevaluated in combination with IFN. It is enigmatic why the 3mg dose vaccination caused less induction of the peptide-specific CTLs. It was reported previously that persistent vaccine depots could induce sequestration, dysfunction and depletion of antigen-specific CTLs [27]. That may explain, at least in part, the mechanism of the bell-shaped dose effect of the antigenic peptide.

Three patients with a history of immunotherapy such as a dendritic cell vaccine and certain peptide vaccine failed to respond to the SVN-2B peptide vaccine clinically and immunologically. It is possible that their cancers may have had immunoescape phenotypes, thereby maintaining resistance to the vaccine as well as the prior immunotherapy. Alternatively, prior immunotherapy might have affected the immune system, thereby inducing tolerance against the vaccination. In any case, a history of immunotherapy was considered

to be a predictive factor for a worse response, and was therefore added to the exclusion criteria in the ongoing phase II clinical study.

In conclusion, we demonstrated the safety and clinical efficacy of the SVN-2B peptide vaccine for patients with advanced gastrointestinal cancer, although clinical interpretation of the results was limited due to this being a phase I study with a small number of patients. At present, a phase II study (UMIN000012146) of the SVN-2B peptide vaccine for advanced pancreatic cancer is ongoing in combination with IFN- γ .

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