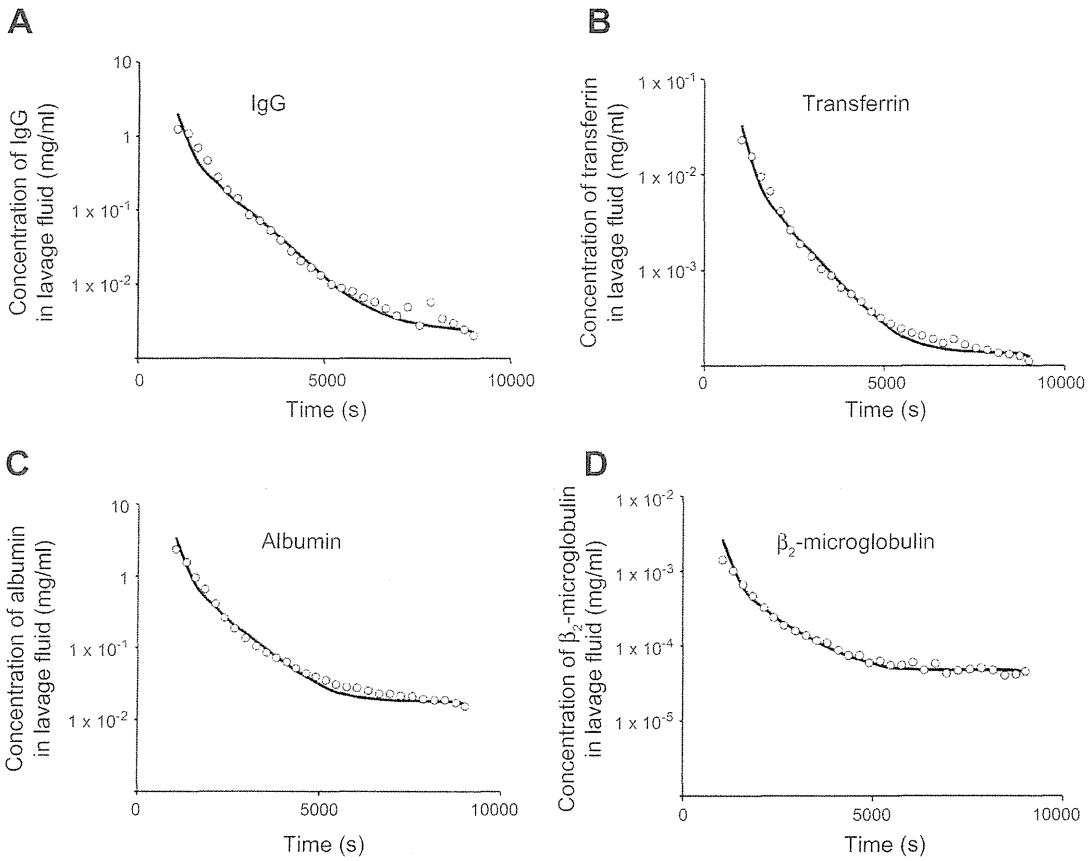

Fig. 2. Theoretical concentrations (lines) and measured concentration (plots) of IgG (*A*), transferrin (*B*), albumin (*C*), and β_2-microglobulin (*D*) in the drained aliquot of lavage fluid for each cycle. The vertical axis is the concentration of the protein on a log scale, and the horizontal axis indicates the time after the beginning of WLL.

when the cumulative eliminated albumin in case 1 was estimated in silico with fixed lavage cycle time at 240 s, the eliminated albumin appeared to increase as the instilled volume increased during 0 to ~3,200 s. After 3,200 s, the eliminated albumin gradually increased, but the volume effect seemed to be diminished.

Exceptional Substances That Fail to Follow the Mathematical Model

Although we applied our mathematical model to the transfer of various substances during WLL, we found that the following substances did not follow the model.

Gastrin and urea. Measured levels of gastrin and urea did not exhibit an exponential decreasing phase but instead reached a plateau in the early stage of WLL (Fig. 5, *A* and *B*). Thus calculation of K_s was difficult. Permeation of gastrin and urea from the blood to the lavage fluid occurred so quickly that the theoretical curves were hardly matched with the actual measurements, which themselves fluctuated markedly during the plateau phase.

SP-D. The SP-D concentration in the drained lavage fluid decreased consistently to a minor extent in the four lungs in the absence of an exponential phase and quickly reached a plateau in the early phase (Fig. 5*C*). As alveolar type II cells and nonciliated Clara cells abundantly release SP-D into the lower respiratory tract, this early plateau phase reflects its active release in situ.

GM-CSF autoantibody. Although the quantified GM-CSF autoantibody belongs to an IgG isotype, theoretical curves of the concentration in the drained lavage fluid did not fit with the measured autoantibody concentration even upon substitution of various sets of coefficients with K_s and K_b in all 17 lungs (Fig. 5*D*).

DISCUSSION

By using a mathematical model based on measured concentrations of proteins, this study investigated the transfer of proteins from the surfactant and blood into the lavage fluid during WLL. We confirmed that the transfer followed a time-dependent differential equation, which assumes that the rate of transfer is proportional to the transmission coefficient, the effective surface area, and the protein gradient between the body compartment and lavage fluid (44).

By using various methods (e.g., comparisons of the protein concentrations between the plasma, sputum, and BALF) and by proving that the IgG1/IgG2 ratio between the BALF and serum are comparable, previous studies demonstrated the transfer of circulating proteins into the alveolar spaces (2, 14, 18, 28, 39).

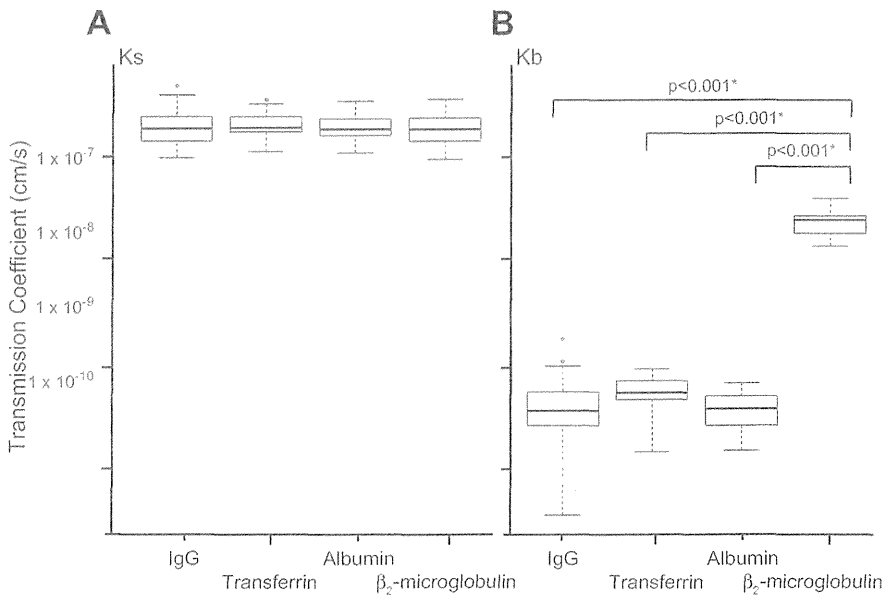

Fig. 3. Coefficients of transfer of IgG, transferrin, albumin, and β_2-microglobulin from surfactant (K_s) (*A*) and blood (K_b) (*B*) to the lavage fluid. The vertical axis indicates the transmission coefficients (cm/s) on a log scale. Statistical significance of coefficients between 2 proteins are shown in the figure.

More recently, intravenously injected GM-CSF autoantibodies were detected in the BALF of nonhuman primates and were observed to reproduce PAP (35). These results indicate that the antibody can cross the air-blood barrier (35). The kinetics of transfer from the blood to the air space and vice versa was studied both in vitro and in vivo (3, 23, 26, 27, 34). In one study, the transmission coefficient (10^{-7}-10^{-5} cm/s) of various proteins across a monolayer of A549 cells was shown to indicate bidirectional transfer. These coefficients appear to be inversely correlated with the molecular weight of proteins (22). In another study, the transmission coefficient for proteins in a monolayer of rat alveolar epithelial cells in vitro was within 10^{-9}-10^{-7} cm/s, whereas that for albumin in sheep lung in vivo was 5×10^{-10} cm/s (11, 17). Thus mass transfer from the blood to the air spaces may be continuously taking place even at steady state.

In previous studies by Ikegami et al. (15), surface tension maintained by surfactant materials covering the alveolar surface was found to have a probable role in interfering with massive transfer and subsequent accumulation of circulating proteins in the air spaces. Interference with the transfer is known to be disrupted by the elimination or deficiency of SP-B (15, 16). Lung lavage may remove surface-active materials in the alveoli and thus temporally disrupt the mechanisms that interfere with the influx of circulating proteins. It is for this reason that we focused on WLL to clarify the mechanism of protein transfer from the blood or surfactant to the lavage fluid. We found that the protein transfer followed a time-dependent mathematical model that was made analogous to the heat transmission model. To our knowledge, this is the first study that has clarified the mechanism of protein transfer in the lung during WLL.

To postulate a mathematical model, we assumed that the transfer of proteins from each body compartment to the lavage fluid consists of two pathways, namely transfer from the accumulated surfactant to the lavage fluid and transfer from the blood to the lavage fluid. The latter may be further divided into two pathways, namely transfer from the blood through the surfactant and direct transfer to the lavage fluid. However, we did not distinguish between these two latter pathways in this study because the transfer of a protein across the air-blood barrier seemed to be rate limiting. We found that protein transfer from the surfactant to the lavage fluid appeared to have K_s values independent of the molecular weight and other properties. It is notable that the K_s values did not differ among patients, indicating the reproducibility of the model. However, mass transfer from the blood to the lavage fluid with variable K_b values did appear to be affected by the molecular weight of the protein because the protein was transferred through a semipermeable membrane consisting of endothelial cells, basement membrane, and type I pneumocytes. Transcytosis was proposed as the primary mechanism of protein transfer for large molecules and of partial paracellular diffusion of small molecules (7, 23). However, the true mechanism remains controversial. As indicated in this study, transfer of β_2-microglobulin (molecular weight of 11 kDa) from the blood to the lavage fluid had K_b values that were two orders of magnitude higher than those of albumin, transferrin, and IgG, which had molecular weights of 66, 80, and 150 kDa, respectively. This difference suggests that β_2-microglobulin diffusion possesses a mechanism that is different from that of other proteins, i.e., it is supposed to be mainly transcytosis for albumin, transferrin, and IgG but mainly paracellular diffusion for β_2-microglobulin. Further analyses will be required to clarify the mechanisms by measuring the permeability of various substances with molecular weight of 10–60 kDa to confirm a "gap" in permeability coefficient K_b among substances with molecular weights in this range.

It is notable that the decrease in concentrations of low-molecular-weight substances in the lavage fluid, namely urea (molecular weight of 60 kDa) and gastrin (molecular weight of 2.1 kDa), was inconsistent with our mathematical model. The measured concentrations appeared to fluctuate and appeared to be independent of time. Moreover, the phase of exponential

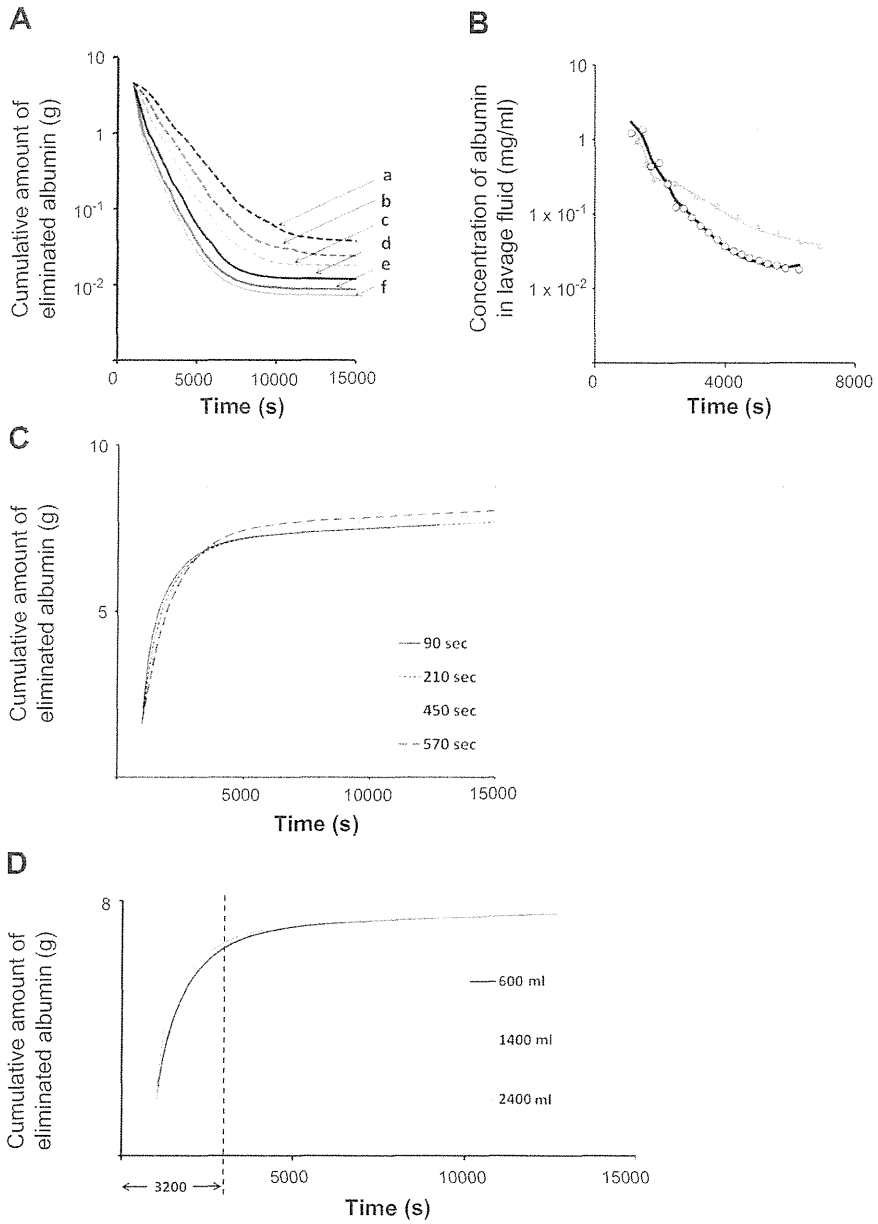

Fig. 4. *A*: durable effect of the retaining stage in each lavage cycle on the theoretical decreasing curve of albumin concentration in the drained lavage fluid. The time assumed for the retaining stage was variable: *a*, 540 s; b, 360 s; *c*, 240 s; *d*, 120 s; *e*, 60 s; and *f*, 30 s. The vertical axis indicates the albumin concentration in the lavage fluid (mg/ml). The horizontal axis indicates the time after the beginning of WLL. *B*: theoretical (lines; black, left; gray, right) and measured (plots; ○, left, ▲, right) concentrations of albumin in the drained lavage fluid in each cycle. The vertical axis indicates the albumin concentration in the lavage fluid (mg/ml). The horizontal axis indicates the time after the beginning of WLL. *C*: simulation curves of cumulative amount of albumin drained in the drained lavage fluid when the retaining time varied with 90 (solid line), 210(small dashed line), 450(dotted line), or 570(large dashed line) s. *D*: cumulative amount of eliminated albumin in the drained lavage fluid. An in silico evaluation by changing instilled saline volume varied with 600 (black solid line), 1,400 (dotted line), or 2,400 (gray solid line) ml.

decrease was hardly defined in six out of ten lungs examined; when there was any decrease, the phase lasted within 1,000 s after the start of WLL (data not shown). This characteristic was likely due to the high permeability of the air-blood barrier to the molecules. Similarly, Rennard et al. (32) reported that urea was more able than glucose and albumin to permeate into the lavage fluid, as observed in normal volunteers with saline instilled into their lung segments.

SP-D is produced by alveolar type II cells and nonciliated Clara cells in the lower respiratory tracts and is secreted into the air space (43). Although SP-D is detectable in the sera of patients with aPAP, its levels are much lower than those of BAL (12). Thus SP-D transfer from the blood to the air space is negligible. The high concentration of SP-D in the lavage fluid was likely due to its continuous production in the lung. The rate of its production was estimated to be 6–13 mg/h on the basis of evaluation of four lungs (data not shown).

The lung is the organ that most abundantly produces GM-CSF, a factor that is critical for terminal differentiation of alveolar macrophages, as it promotes the expression of the transcription factor, PU.1 (38). It is suggested that IgG-type GM-CSF autoantibody is pathogenic and is known to be transferred from the lung capillaries into the air spaces immediately formed by GM-CSF autoantibody complex to become undetectable by our GM-CSF autoantibody ELISA system (30).

Furthermore, we had better to reconsider the adequacy of the present mathematical model when it was applied to substances

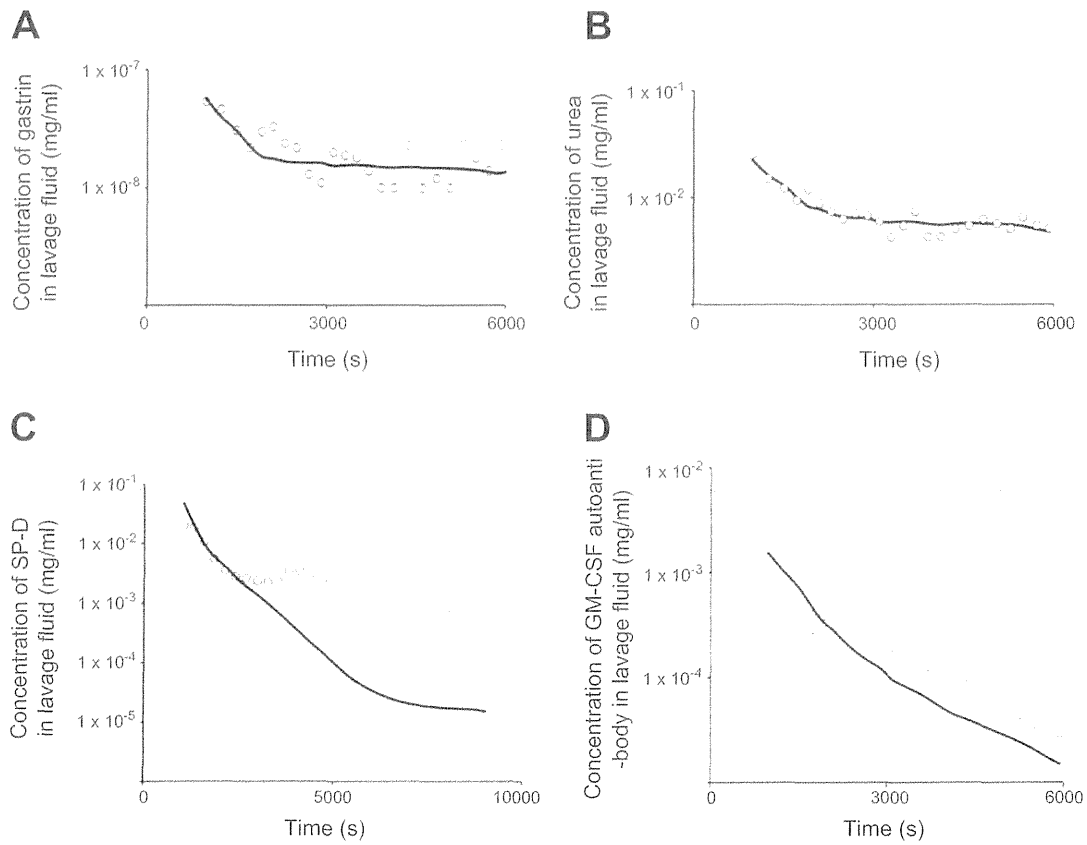

Fig. 5. *A* and *B*: actual measurements (plots) of gastrin or urea concentration in the drained lavage fluid did not exhibit the exponential decreasing phase but reached a plateau fluctuating in the early term. These seemed to migrate immediately from the blood to the lavage fluid. Thus the theoretical curves (lines) were hardly fitted with the actual measured concentration. *C*: concentration of surfactant protein D (SP-D) in the drained lavage fluid revealed slight decrease without exponential phase and soon reached a plateau phase in the early term. As SP-D is abundantly released from alveolar type II cells into the lower respiratory tracts, this early plateau phase probably reflected the active release in situ. *D*: actually measured granulocyte/macrophage colony-stimulating factor (GM-CSF) autoantibody concentrations were consistently under the theoretical curve especially in the early stage.

with lower molecular weights by assuming two permeation coefficients, such as K_{b1} (coefficients from the blood to the lavage fluid through surfactant) and K_{b2} (from the blood directly to the lavage fluid).

In the present study, the recovery rate in the first draining lavage fluid was lower than those after the second lavage. Although the first instilled saline remained in the lower respiratory tracts, we did not mind the remaining volume at the first draining because we thought that the remaining lavage fluid could be recovered after the second draining. Therefore, we did not intentionally extend the first draining time longer than those of other cycles. Although we usually perform percussion or vibration on the patient's chest, the recovery rate at the first draining was not improved by these procedures. It is likely that the low recovery rate and its variability of the first lavage shown in Table 4 were due to the early cessation of the first draining.

To date, methods of WLL for the treatment of PAP have not been standardized (25). Michaud et al. (29) recommended instilling 1 l of saline into the lavage lung and then to clamp the draining tube for 4–5 min (29). Bonella et al. (4) and Paschen et al. (31) determined the number of lavage cycles by measuring the optical density of each lavage fluid. They applied statistical evaluation to data from a number of WLLs to find the relationship between instilled saline volume and eliminated proteins. Although their approach is fundamentally different from ours, their finding that instilling volume is an important element for determining the amount of eliminated protein was confirmed in this study (Fig. 4*D*). The protocol for WLL used in this study were variable among participating hospitals, and thus time of each cycle varied between 213–630 s, including 120–540 s for the retaining time. As for our mathematical model, the number of cycles and the retaining times did not influence the efficiency of WLL. Based on *Eq. 1*, the amount of proteins eliminated by WLL was dependent on time after the beginning. According to the volume effect demonstrated by in silico simulation in this study (Fig. 4*D*), larger instilled volume appeared to improve the efficiency of lavage. However, the simulation also suggested that the effect is limited within some range of time. Previous studies, however, demonstrated the volume effect (4). In this regard, total eliminated albumin concentration significantly correlated with instilling saline volume in actually measured values in 17 WLLs of the present study with Rho value at 0.69. However, we have to consider the possibility that it also prolonged the duration of instilling and draining time, and thus longer time for each lavage cycle increases the elimi-

nated protein(s). Thus our mathematical model may be useful to predict the amount of eliminated proteins at a certain time point after the beginning of WLL.

In conclusion, we demonstrated that protein transfer in the lung during WLL followed a relatively simple, mathematical model based on diffusion and that this model could be expressed in terms of a number of differential equations. As an exception of the present mathematical model, substances with low molecular weight do not follow the theory. Our study, not only contributes to the design of an efficient regimen for WLL, but also reveals the mechanism of delivery of specific large drug molecules across the air-blood barrier, such as antibody drugs.

APPENDIX

The Effective Alveolar Surface Area

The effective alveolar surface area was calculated from the data for the alveolar volume, V_A according to the following equations: $A_s = 6.4 \cdot 10^3 \cdot V_A^{2/3}$. For a person with 74 kg body wt, both A_s and V_A were reported to be 143 m^2 and 3,338 ml, respectively (10). The effective surface area of the pulmonary capillaries, A_b, was estimated from the following formula (10): $A_b = 0.89 \cdot A_s$. The relationship between alveolar surface area, S_A, and alveolar volume, V_A, depends on the number of alveoli. S_A increases as the number of alveoli increases at a fixed value of V_A. According to Ref. 10, the average lung volume is 4,300 ml, and the average alveolar surface is $(143 \pm 12) \times 10^4$ cm^2 in normal subjects with an average body weight of 74 kg at 19–40 yr of age. Under these conditions, air-space volume density is 0.865 ± 0.013 cm^2/cm^3, and alveolar surface density is 370.6 ± 28.9 cm^2/cm^3.

We set

$$\beta = \frac{S_A^{\frac{1}{2}}}{V_A^{\frac{1}{3}}} \qquad (A1)$$

where, the right side of the equation is an expression for the constant shape parameter, β.

According to the report described above (V_A and S_A in the space V)

$$\frac{S_A}{V} = 370.6 \text{ cm}^2/\text{cm}^3 \qquad (A2)$$

$$S_A = 143 \times 10^4 \text{ cm}^2 \qquad (A3)$$

$$\frac{V_A}{V} = 0.865 \text{ cm}^3/\text{cm}^3 \qquad (A4)$$

From *Eqs. A2* and *A3*,

$$V = 3859 \text{ ml} \qquad (A5)$$

and from *Eqs. A4* and *A5*

$$V_A = 3338 \text{ ml} \qquad (A6)$$

where the anatomical dead space is 4,300 − 3,338 = 962 ml. Introducing *Eqs. A3* and *A6* into *Eq. A1*,

$$\beta = \frac{\sqrt{143 \times 10^4}}{\sqrt[3]{3338}} = 80.02 \qquad (A7)$$

On the basis of *Eq. A1* (note that S_A is in m^2 and V_A is in ml),

$$S_A = \beta^2 \cdot V_A^{\frac{2}{3}} = 6.403 \times 10^3 \cdot V_A^{\frac{2}{3}} \qquad (A8)$$

The value of β may be considered as constant even with a change in V_A in the same subject, as the number of alveoli and the shape do not change, particularly in the supine position.

Method for Optimizing the Transmission Coefficients

A program was written in Visual Basic Application using various coefficients to calculate the theoretical substance concentrations in the lavage aliquots. For explanation, we show an example of simulation used to obtain the best fitting curve shown in Fig. 2*C*. As shown in Appendix Fig. A1*A*, the value for K_s could be determined to be 1.8×10^{-7} cm/s by the least-square method until 3,000 s when K_b was assumed to be 0 cm/s. Next, K_b value was determined to be 5.2×10^{-10} cm/s again by the least-square method by 9,018 s. As shown in Appendix Fig. A1*B*, the theoretical curve appeared closer to the dotted actual measurements. Then K_b was changed to 6.1×10^{-10} cm/s manually, as shown in Appendix Fig. A1*C*: the theoretical curve

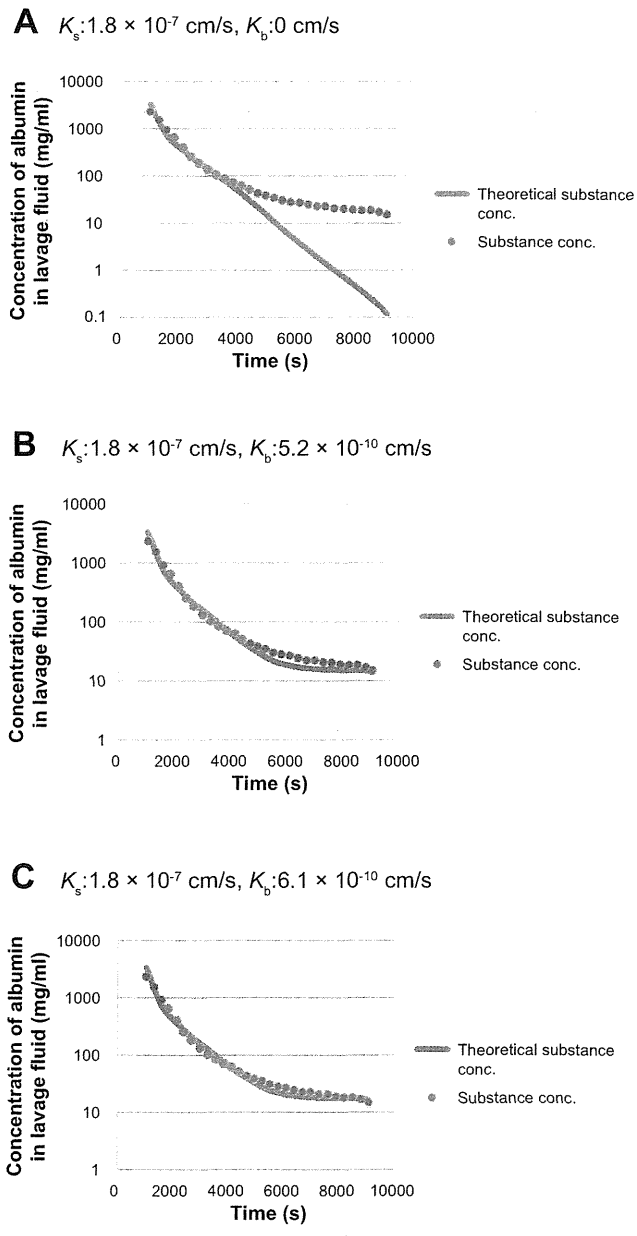

Fig. A1. Example of simulation used to obtain the best fitting curve shown in Fig. 2*C*.

completely coincides with the dotted actual measurements. Therefore, K_b was determined to be 6.1×10^{-10} cm/s.

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DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

AUTHOR CONTRIBUTIONS

K.A., T.M., K.U., R.T., T.I., Y. Inoue, and K.N. conception and design of research; K.A., A.H., Y. Ito, H.W., T.W., Takero Arai, H.M., S.O., R.T., T. Takada, E.Y., T.I., M. Hirose, and Toru Arai performed experiments; K.A., T. Tanaka, T.M., N.K., M. Hayashi, M. Hirose, and K.N. analyzed data; K.A., T. Tanaka, T.M., N.K., R.T., E.Y., H.K., and K.N. interpreted results of experiments; K.A., T. Tanaka, R.T., and K.N. prepared figures; K.A., T. Tanaka, T.M., K.U., T.I., and K.N. drafted manuscript; K.A., T.M., K.U., R.T., Toru Arai, H.K., and K.N. edited and revised manuscript; K.A., T. Tanaka, T.M., N.K., A.H., H.W., Takero Arai, M. Hayashi, H.M., K.U., S.O., R.T., T. Takada, E.Y., T.I., M. Hirose, Toru Arai, Y. Inoue, H.K., and K.N. approved final version of manuscript.

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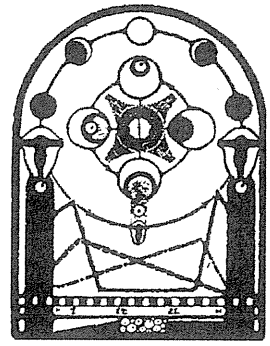

NOTE *Immunology*

Up-Regulation of Cluster of Differentiation (CD) 11b Expression on the Surface of Canine Granulocytes with Human Granulocyte-Macrophage Colony-Stimulating Factor (GM-CSF)

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ABSTRACT. Granulocyte-macrophage colony-stimulating factor (GM-CSF) is a pleiotropic cytokine, sharing a common beta subunit (CDw131) with interleukins 3 and 5. GM-CSF is important for its direct and indirect involvement in host defense. In veterinary medicine, human (h) GM-CSF has been used as a substitute for canine GM-CSF to stimulate canine granulocytes and macrophages. In this study, we compared the effects of three distinct hGM-CSFs produced by bacteria, yeasts and Chinese hamster ovary (CHO) cells with those of *Escherichia* (E) *coli*-produced canine GM-CSF on the cluster of differentiation 11b (CD11b) expression in canine granulocytes. The median effective dose (ED_{50}) of hGM-CSFs from bacteria, yeasts and CHO cells was 3.09, 4.09 and 4.27 *ng/ml*, respectively, with no significant difference among three. In contrast, a significant difference was observed between ED_{50} of canine GM-CSF (0.56 *ng/ml*) and three hGM-CSFs according to the paired *t*-test ($P<0.05$). We conclude that hGM-CSF can activate canine granulocytes, but the average activity of the three rhGM-CSFs was approximately 15% of that of canine GM-CSF.

KEY WORDS: canine, CD11b, flow cytometry, granulocyte-macrophage colony-stimulating factor, median fluorescence intensity, xenostimulation

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Human granulocyte-macrophage colony-stimulating factor (hGM-CSF) is a protein of 144 amino acids (AA), including the signal peptide of 18 AA, and is produced by various types of cells. The protein is monomeric, but its active form basically takes a noncovalent homodimmer in nature. Although GM-CSF is a major cytokine for hemopoiesis like granulocyte colony-stimulating factor, macrophage colony-stimulating factor and erythropoietin, the cytokine has been known to be involved in the enhancement of eosinophil chemotaxis [7], maturation of macrophages and dendritic cells [17], granulocyte activation [1], adjuvant effect [3] and inhibition of apoptosis [4].

Cluster of differentiation molecule 11b (CD11b), known as its integrin α M subunit, consists of macrophage-1 antigen (Mac-1) with CD18. The molecule is expressed in many types of cells, and the CD11b expression on the surfaces of granulocytes and macrophages is increased by their activation, playing an important role in host defense. Mac-1 has been reported to support neutrophil immobilization and migration [6] and is also known as complement receptor 3 (CR3) that binds to iC3b, eliminating pathogens and immune complexes by neutrophils, macrophages and the reticuloendothelial system. CD11b is rapidly elevated by the activation of neutrophils and macrophages, and the amount of CD11b in neutrophils correlates with their activation and inflammation [11].

Clinical trials of the adjuvant therapy and the prevention form leukocytopenia with GM-CSF in veterinary cancer medicine have been started, but the preparation of canine GM-CSF for clinical use is still unavailable. Thus, we just have to choose that of human GM-CSF (hGM-CSF) at the present time. Because hGM-CSF is active in canine cells, it has been empirically employed as a substitute for canine GM-CSF [18, 22]; however, its quantitative activity in canine cells has not been elucidated. Here, we compared the effects of hGM-CSF to those of canine GM-CSF in canine granulocytes and also measured the median effective doses (ED_{50}) of three different rhGM-CSFs in canine granulocytes.

Anti-CD11b (M1/70) conjugated with allophycocyanin-Cy7, Gr-1 with allophycocyanin and anti-human CD14 with phycoerythrin were purchased from BioLegend Co., Ltd. (San Diego, CA, U.S.A.; provided by Tomy Digital, Tokyo, Japan). Molgramostim; *Escherichia (E.) coli*-produced recombinant human GM-CSF (rhGM-CSF), sargramostim produced by yeasts and canine recombinant GM-CSF were obtained from Amoytop Biotech (Xiamen, Fujian, People's Republic of China), Genzyme corporation (Cambridge, MA, U.S.A.) and R&D systems (Minneapolis, MN, U.S.A.), respectively. JCR Pharmaceuticals Co., Ltd. (Akashi, Japan) donated rhGM-CSF produced by Chinese hamster ovary (CHO) cells.

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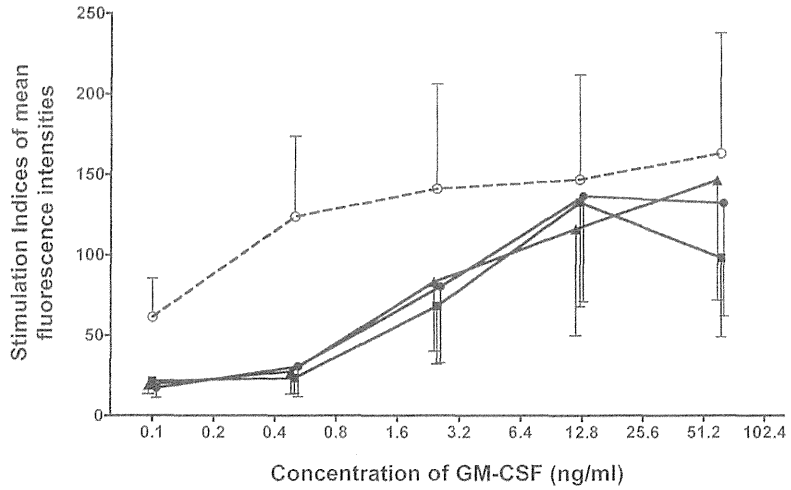

Fig. 1. Dose-response curves of CD11b expression with various granulocyte-macrophage colony-stimulating factors in canine granulocytes. Sigmoid curves represent dose-responses of molgramostim (solid circle), sargramostim (solid square), CHO-produced hGM-CSF (solid triangle) and canine GM-CSF (open circle and broken line). The points and bars show the average values and standard deviations from 4 animals, respectively. X and Y axes show the concentrations of GM-CSF (*ng*/m*l*) and stimulation indices, respectively.

Heparinized canine blood was obtained from 2 male and 2 female beagles for practical trainings of students at Nippon Veterinary and Life Science University. These beagles were individually housed, fed dog chows once a day and drank water *ad libitum*. These bloods were transported to our laboratory and processed at room temperature within 1 hr. Briefly, 100-μl aliquots of the blood were aseptically placed in 2.0-m*l* sterile microtubes, to which various amounts of canine or hGM-CSF were added at final concentrations of 0.02–62.5 *ng*/m*l* or macrophage-serum free medium (macrophage-SFM; Invitrogen Corporation, Carlsbad, CA, U.S.A.) alone. Subsequently, all the samples were incubated for 15 min at a 37°C in a 5% CO_2 incubator without shaking.

After stimulation, antibody cocktail was added to each tube, which was then incubated for 30 min at 4°C. The blood was hemolyzed with 0.15 M ammonium chloride containing 1 mM $KHCO_3$ and 0.1 mM EDTA 4Na (pH 7.3), washed twice with flow cytometer buffer (PBS containing 2% BSA and 0.1% sodium azide) and then fixed in FluoroFix™ buffer (BioLegend), as per the manufacturer's instructions. The cells were re-suspended in 100 μl flow cytometer buffer (PBS containing 2% BSA and 0.05% sodium azide). Data were acquired using FACSArray (BD Bioscience, San Jose, CA, U.S.A.), gating the granulocyte area on a forward vs. side scatter. The median fluorescence intensities (MFIs) of $CD11b^+$ population were obtained under the gate of granulocytes at SSC vs. FSC scatter and $CD14^-$. The indices of MFIs were determined by dividing MFIs from GM-stimulated cultures by MFI from PBS-cultured granulocytes. ED_{50}, determined from MFI values using the probit method, was statistically analyzed using paired *t*-tests at every GM-CSF dose.

Three hGM-CSFs revealed increased CD11b expression on canine granulocytes in a dose-dependent manner (Fig. 1). ED_{50} of molgramostim, sargramostim and hGM-CSF from CHO cells was 3.09, 4.09 and 4.27 *ng*/m*l*, respectively; moreover, no significant difference was observed among these rhGM-CSFs (Table 1). In contrast, ED_{50} of canine rGM-CSF was 0.56 *ng*/m*l*, which was significantly different from the three rhGM-CSFs according to the paired *t*-test results ($P<0.05$). Further, ED_{50} of molgramostim, sargramostim and rhGM-CSF from the CHO cells was 18.1%, 13.7% and 13.1%, respectively, compared with the canine rGM-CSF for canine granulocytes.

GM-CSF is not only an important hemopoietic cytokine, but also involved in the upregulation of the immune system and host-defense [5, 19, 21], because immune cells express its receptor [10, 14]. In experiments using dogs, rhGM-CSF has been employed as a substitute for the canine reagent [2, 16].

GM-CSF activity is usually measured by the proliferation of cells that are GM-CSF-dependent; e.g. TF-1 for hGM-CSF [9]. The detection of augmented CD11b with GM-CSF is rapid and easy. CD11b expression on the surface of neutrophils has been reported to elevate by GM-CSF stimulation [12, 15]. Uchida *et al.* have reported that the quick elevation of CD11b expression on human neutrophils by GM-CSF stimulation was caused by its endogenous molecules but not de novo synthesis [20]. According to a modified Uchida method [20], we detected the activities of rhGM-CSFs in canine neutrophils in a dose-dependent manner. We conclude it may not be a problem to employ rhGM-CSF to canine experiment. This technique doesn't require any GM-CSF-dependent cell line and is applicable to every animal species.

Table 1. Median effective doses of various granulocyte-macrophage colony-stimulating factors in expression of CD11b in canine granulocytes

GM-CSF	ED_{50} (ng/ml)	Specific activities (units/μg)	Relative activities (%) to canine GM-CSF in ED_{50}
Molgramostim	3.09 ± 1.18 [a]	323.6	18.1
Sargramostim	4.09 ± 1.56 [a]	244.5	13.7
CHO hGM-CSF	4.27 ± 1.51 [a]	234.2	13.1
Canine GM-CSF	0.56 ± 0.46 [a]	1,785.7	–

a) Median effective dose (ED_{50}) of canine granulocyte-macrophage colony-stimulating factor (GM-CSF) significantly differed from those of hGM-CSFs according to the paired *t*-test results ($P<0.05$).

Furthermore, it has been reported that some mouse cells are not stimulated by hGM-CSF. However, McClure *et al.* proved that rhGM-CSF activated the BaF-B03 mouse cell line transfected with human GM-CSF receptor α subunit gene [13]. The intracytoplasmic region of the subunit did not participate in the signal transduction [23], which suggests that the α subunit plays an important role in binding species-specifically to GM-CSF. Therefore, the α subunit of canine GM-CSF may have an effective affinity to rhGM-CSF, although rhGM-CSFs had a weaker impact on canine granulocytes compared with canine rGM-CSF in this study. Therefore, to obtain an effect equivalent to an expected activity in dogs with hGM-CSF, we must employ an approximately septuplet dose of rhGM-CSF (Table 1). Nevertheless, this indicates that rhGM-CSF can be a valuable tool for a canine study.

In addition, we also compared GM-CSFs from three different sources: *E. coli*, yeasts and CHO cells; although no significant difference was determined in ED_{50} for the three sources, *E. coli*-produced rhGM-CSF (molgramostim) revealed the highest activity. Moreover, Kelleher *et al.* determined that *E. coli*-produced hGM-CSF had higher efficacy with regard to the proliferation of TF-1 cells compared with that of CHO protein [8]. Although we are not able to explain why molgramostim exhibited the highest activity in our study, Kelleher *et al.* suggested that the difference was the result of the higher affinity of *E. coli* protein [8]. Molgramostim is not much different from the other two types investigated without their glycosylation, which may be involved in their 3-D conformation and homodimmer formation and/or interfere with their interactions with GM-CSFR, affecting GM-CSF activity. Thus, the differences in glycosylation may be responsible for their varied activities.

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Low concentrations of recombinant granulocyte macrophage-colony stimulating factor derived from Chinese hamster ovary cells augments long-term bioactivity with delayed clearance *in vitro*

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ABSTRACT

To date, the biological activity of granulocyte macrophage-colony stimulating factor (GM-CSF) has been investigated by using mostly *Escherichia coli*- or yeast cell-derived recombinant human GM-CSF (erhGM-CSF and yrhGM-CSF, respectively). However, Chinese hamster ovary cell-derived recombinant human GM-CSF (crhGM-CSF), as well as natural human GM-CSF, is a distinct molecule that includes modifications by complicated oligosaccharide moieties. In the present study, we reevaluated the bioactivity of crhGM-CSF by comparing it with those of erhGM-CSF and yrhGM-CSF. The effect of short-term stimulation (0.5 h) on the activation of neutrophils/monocytes or peripheral blood mononuclear cells (PBMCs) by crhGM-CSF was lower than those with erhGM-CSF or yrhGM-CSF at low concentrations (under 60 pM). Intermediate-term stimulation (24 h) among the different rhGM-CSFs with respect to its effect on the activation of TF-1 cells, a GM-CSF-dependent cell line, or PBMCs was not significantly different. In contrast, the proliferation/survival of TF-1 cells or PBMCs after long-term stimulation (72–168 h) was higher at low concentrations of crhGM-CSF (15–30 pM) than that of cells treated with other GM-CSFs. The proportion of apoptotic TF-1 cells after incubation with crhGM-CSF for 72 h was lower than that of cells incubated with other rhGM-CSFs. These effects were attenuated by desialylation of crhGM-CSF. Clearance of crhGM-CSF but not desialylated-crhGM-CSF by both TF-1 cells and PBMCs was delayed compared with that of erhGM-CSF or yrhGM-CSF. These results suggest that sialylation of oligosaccharide moieties delayed the clearance of GM-CSF, thus eliciting increased long-term bioactivity *in vitro*.

1. Introduction

Granulocyte macrophage-colony stimulating factor (GM-CSF) is a hematopoietic growth factor that regulates the growth, differentiation, and maturation of myeloid precursor cells and promotes the function of mature neutrophils, eosinophils, and monocytes [1–4]. It elicits these diverse effects through interaction with a unique dodecameric receptor complex on cells, which consists of α and common β chains [5–7]. GM-CSF signaling induces phosphorylation of Janus kinase 2 (JAK2) and the common β chains, followed by activation of signal transducers and activators of transcription (STATs) [5,7,8]. Upon immune stimulation, it is produced by a variety of cell types, including T cells, macrophages, endothelial cells, and fibroblasts. Although GM-CSF is produced locally [3], it can

Abbreviations: ACN, acetonitrile; ANOVA, analysis of variance; CHO, Chinese hamster ovary; crhGM-CSF, CHO-derived recombinant human GM-CSF; erhGM-CSF, *Escherichia coli*-derived recombinant human GM-CSF; FCS, fetal calf serum; FITC, fluorescein isothiocyanate; GM-CSF, granulocyte macrophage-colony stimulating factor; JAK2, Janus kinase 2; MIP-1α, macrophage inflammatory protein; NaN_3, sodium azide; PBMCs, peripheral blood mononuclear cells; SDS–PAGE, sodium dodecyl sulfate–polyacrylamidegel electrophoresis; STAT5, signal transduction and activator of transcription; TFA, trifluoroacetic acid; TOF mass spectrometer, time-of-flight mass spectrometer; yrhGM-CSF, yeast cell-derived recombinant human GM-CSF.

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act in a paracrine fashion to recruit circulating neutrophils, monocytes, and lymphocytes to enhance their function in host defense [9,10]. GM-CSF is used clinically to prevent neutropenia and associated infections by promoting the proliferation of hematopoietic progenitor cells after chemotherapy, by promoting the differentiation of myeloid cells, and by enhancing the antibacterial activities of neutrophils and macrophages [10–14].

Natural human GM-CSF (hGM-CSF) has been purified from several sources, including medium conditioned with placenta cells or activated blood lymphocytes [15–19]. It is a glycoprotein that consists of 127 amino acid residues, with four cysteines involved in two disulfide bonds, forming a compact globular structure that comprises four α-helices joined by loops. It is found extracellularly as a homodimer [6,7] with two N-glycosylation sites at Asn27 and Asn37 and three O-glycosylation sites at Ser7, Ser9, and Thr10 [15]. The most heavily glycosylated hGM-CSF, with a molecular weight of 28–32 kDa, has two N-linked carbohydrate moieties, whereas the partially glycosylated hGM-CSF, with a molecular weight of 23–25 kDa, contains one N-linked carbohydrate moiety. A minimally glycosylated hGM-CSF with molecular weight of 16–18 kDa consists of only one O-linked carbohydrate [15,20].

Cells from various species can produce recombinant hGM-CSF (rhGM-CSF) [21,22]. However, only commercial preparations produced from *Escherichia coli* and *Saccharomyces cerevisiae* are available for clinical use. Commercial *E. coli*-derived recombinant hGM-CSF (erhGM-CSF), Molgramostim, is non-glycosylated, consists of 127 amino acid residues, has a molecular weight of 14.5 kDa, and is methylated at the N-terminal end [23]. Commercial *Saccharomyces*-derived recombinant hGM-CSF (yrhGM-CSF), Sargramostim, is a glycoprotein of 127 amino acids composed of three primary molecular species having molecular weights of 19.5, 16.8, and 15.5 kDa [23]. Its amino acid sequence differs from hGM-CSF by a substitution of leucine at position 23 [23]. On the other hand, rhGM-CSF derived from Chinese hamster ovary (CHO) cells (crhGM-CSF) has a molecular weight of 15–32 kDa with the same N-glycosylation and O-glycosylation sites as those of hGM-CSF, although the carbohydrate moieties added are probably different. Forno et al. demonstrated that the N-glycan terminal contains mono- and disialic acid residues, but has predominantly tri- or tetrasialic acid residues with and without N-acetyllactosamine repeat units. N-glycans contain more than 90% α-1,6-linked fucose at the proximal end [20].

The pattern of glycosylation on GM-CSF is known to affect its biological activity. Proliferation of a human monocytic leukemia cell line incubated with the heavily glycosylated hGM-CSF (28–32 kDa) was reduced six fold compared with proliferation after treatment with non-glycosylated erhGM-CSF, while neutrophil superoxide anion production was reduced by up to 10-fold. Partially glycosylated hGM-CSF (23–25 kDa) and minimally glycosylated hGM-CSF (16–18 kDa) have biological activity similar to that of erhGM-CSF. The binding capacity of these hGM-CSFs for the rhGM-CSF receptor on neutrophils decreases with increasing molecular weight [15]. Similarly, most studies on mammalian cell-derived, glycosylated GM-CSF (including crhGM-CSF) demonstrate that glycosylation of GM-CSF prolongs the *in vivo* half life by stabilizing the protein, but reduces its binding avidity to the GM-CSF receptor and decreases its biological activities such as colony-forming activity of bone marrow cells and neutrophil superoxide anion production [15,24].

In contrast to previous studies [15,24], we showed in the present study that glycosylated rhGM-CSF produced by CHO cells exhibited increased proliferation/survival of TF-1 cells, PBMCs and monocytes at low GM-CSF concentrations compared with that of erhGM-CSF and yrhGM-CSF *in vitro*. Desialylation of crhGM-CSF attenuated this effect, indicating that the sialyl residue is crucial for augmenting the long-term activity of GM-CSF. Moreover, we examined the mechanism of this effect by measuring the clearance of rhGM-CSF by cells.

2. Materials and methods

2.1. Material

2.1.1. Cells

TF-1, a GM-CSF-dependent cell line, was kindly provided by Kitamura et al. [22].

Peripheral blood mononuclear cells (PBMCs) and monocytes were isolated from the peripheral blood of healthy donors as described previously [8]. Written informed consent was obtained under protocols approved by the institutional review boards of the Niigata University Medical Dental Hospital.

2.1.2. rhGM-CSF

Molgramostim and Sargramostim were purchased from Amoytop Biotech Co., Ltd. (Xiamen, Fujian, PRC) and Genzyme Corporation (Cambridge, MA, USA), respectively. crhGM-CSF was kindly provided by JCR Pharmaceuticals Co., Ltd. (Ashiya, Hyogo, Japan).

2.1.3. Desialylation of crhGM-CSF

crhGM-CSF (1 mg/ml) was incubated with neuraminidase agarose from *Clostridium perfringens* (0.05 U/ml, Sigma–Aldrich, MO, USA) in 100 mM sodium acetate buffer with $CaCl_2$ (pH 5.0) for 60 min at 37 °C. After the agarose was removed, the solution was dialyzed against PBS overnight at 4 °C.

2.2. Mass spectrometry

Protein (10 μl) was mixed with 90 μl of 0.1% trifluoroacetic acid (TFA) and 0.5 μl of MB-HIC8 magnetic C8 beads (Bruker Daltonics, Hercules, MA, USA) in a PCR tube and then incubated for 5 min at room temperature. The tube was subsequently placed in a magnetic beads separator and the supernatant was removed by using a pipette. The magnetic beads were then washed three times with 100 μl of 0.1% TFA. The bound proteins were eluted from the magnetic beads by using 4.5 μl of 60% acetonitrile (ACN) in 0.1% TFA. Two microliters of the eluate was mixed with 1 μl of matrix solution (10 g/l sinapinic acid in 70% ACN, 0.1% TFA) and was spotted on a polished steel plate. The mass spectra were obtained on an Ultraflex TOF/TOF mass spectrometer (Bruker Daltonics, Hercules, MA, USA) operated in positive-ion linear mode.

2.3. Phosphorylated STAT5 detection assay

Heparinized fresh whole blood was incubated with 15, 30, 60, or 500 pM rhGM-CSF, for 30 min at 37 °C and fixed, and then red blood cells were lysed in Fix/Lyse buffer (BD Biosciences, Franklin Lakes, New Jersey, USA) for 20 min at 37 °C. White blood cells were collected by centrifugation and fixed in ice-cold methanol at −20 °C for 1 h. After centrifugation, the cells were resuspended in 3% FCS/0.01% NaN_3/PBS solution and incubated with Alexa Fluor 647-labeled anti-pSTAT5 (BD Biosciences, San Jose, CA, New Jersey, USA). Cells with phosphorylated STAT5 in granulocytes/monocytes detected by flow cytometry (Cell Analyzer, Sony, Tokyo, Japan).

2.4. Neutrophil CD11b stimulation index assay

The neutrophil CD11b assay was performed as described previously [25]. Aliquots of heparinized fresh whole blood were incubated with rhGM-CSF, and cell-surface CD11b levels were quantified by flow cytometry (Sony, Tokyo, Japan). The CD11b

stimulation index was calculated as the mean fluorescent intensity of stimulated neutrophils minus the mean fluorescent intensity of unstimulated neutrophils divided by the mean fluorescent intensity of unstimulated neutrophils and multiplied by 100.

2.5. Measurement of GM-CSF-induced MIP-1α in PBMCs

To evaluate GM-CSF-induced MIP-1α production in normal PBMCs, 1×10^6 cells were incubated with or without GM-CSF in macrophage-serum-free medium (GIBCO BRL, Palo Alto, CA, USA). MIP-1α levels in the supernatant were measured by ELISA (Quantikine, R&D Systems, Mincapolis, MN, USA) according to the manufacturer's instructions [26].

2.6. Cell proliferation/survival assay

TF-1 cells, PBMCs and monocytes (2×10^4 cells/well) were incubated with various concentrations of GM-CSF in macrophage serum free medium (GIBCO BRL, Palo Alto, CA, USA) for 3 and 7 days, respectively [27]. At the end of the incubation, 10 μl of 100 μl (5-[2,4-bis(sodiooxysulfonyl)phenyl-3-(2-methoxy-4-nitrophenyl)-2-(4-nitrophenyl)-2*H*-tetrazole-3-ium]) CCK-8, Doujindo, Kumamoto, Japan) was added to each well. Cells were further incubated at 37 °C under 5% CO_2 for 4 h, and formazan formation was measured as absorbance at 450 nm by using a microplate reader (Bio-Rad, CA, USA).

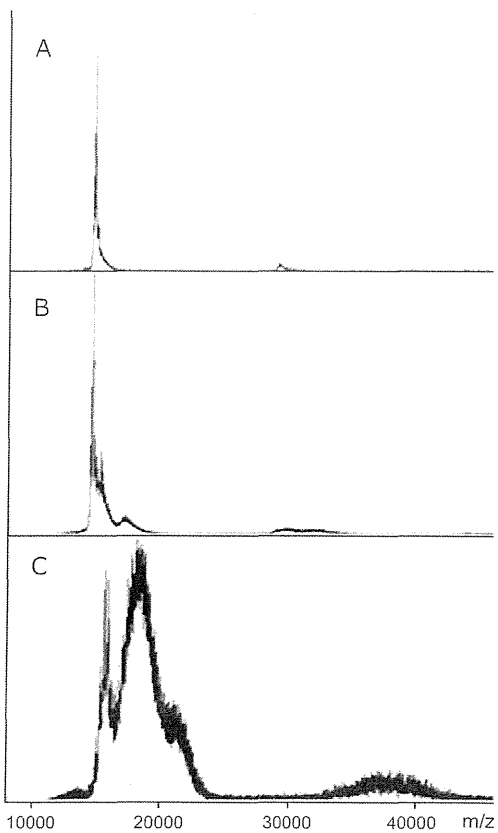

Fig. 1. Mass spectra of rhGM-CSFs. (A) *E. coli*-derived recombinant human GM-CSF. (B) Yeast-derived recombinant human GM-CSF. (C) CHO cell-derived recombinant human GM-CSF. The horizontal axis is the molecular weight (Da) and the vertical axis is the intensity.

2.7. Inhibition of TF-1 cell growth by antibodies

A cell proliferation/survival assay was performed in the presence or absence of 500 ng/ml goat anti-GM-CSF antibody (R&D Systems, Mincapolis, MN, USA), which was purified from the serum of a goat immunized with erhGM-CSF.

2.8. Morphology and cell-survival assay

TF-1 cells (1×10^5 cells) incubated with rhGM-CSF were cytocentrifuged at 200 rpm for 2 min by using a Cytospin (Thermo Scientific, Waltham, MA, USA) and were then stained with Diff-Quick (Sysmex, Hyogo, Japan). The sizes of five hundred cells were measured under a high magnification field by using a micrometer (MeCan Imaging, Saitama, Japan). The percentage of living cells was determined by flow cytometry (Sony, Tokyo, Japan) using staining with propidium iodide solution (Annexin-V-FLUOS Staining Kit, Roche, Basel, Switzerland) according to the manufacturer's instructions.

2.9. SDS–PAGE

rhGM-CSFs (6.5 ng) were subjected to SDS–PAGE under reducing conditions. The gel was stained by using gel stain solution (ORIOLE Fluorescent Gel Stain, Bio-Rad, CA, USA), and the banding pattern was visualized under an image analyzer (MiniLumi, Berthold Technologies, Bad Wildbad, Germany).

2.10. Detection of apoptosis

2.10.1. FITC-Annexin V preparation

TF-1 cells (1×10^6 cells) were stained with FITC-labeled anti-Annexin-V antibody (Annexin-V-FLUOS Staining Kit, Roche,

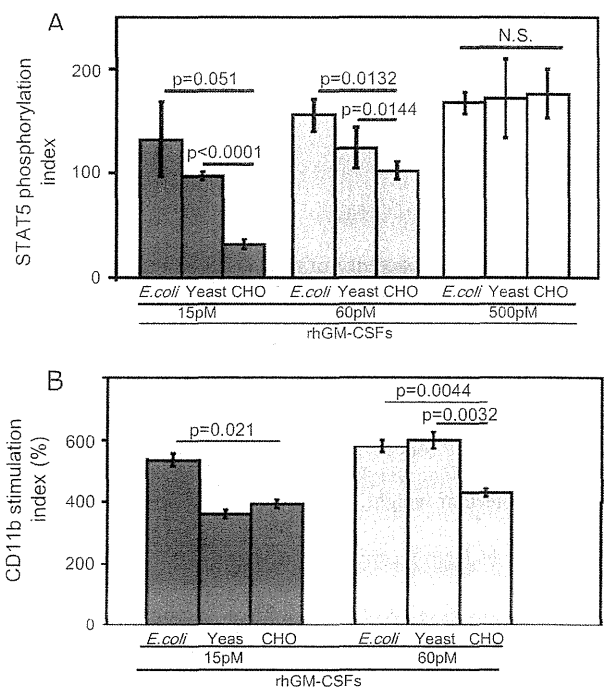

Fig. 2. Effect of short-term stimulation (0.5 h) by *E. coli*-, yeast-, and CHO cell-derived rhGM-CSF. The phosphorylation of STAT5 (A) and CD11b expression of neutrophils and monocytes (B). Whole blood cells were incubated for 0.5 h with 15, 60, or 500 pM of each rhGM-CSF for in (A) and 15 or 60 pM in (B). The vertical axis is STAT5 phosphorylation index (A) and CD11b stimulation index (B) is defined as described in Section 2.

Basel, Switzerland) for 15 min at 4 °C, and the stained cells were detected by flow cytometry (Sony, Tokyo, Japan). FITC-labeled mouse IgG isotype was used as the control.

2.10.2. DNA fragmentation assay

TF-1 cells (2×10^5 cells/ml) were incubated with 15 pM of rhGM-CSF for 3 days [27].

At the end of the incubation, DNA was extracted from TF-1 cells by using a QIAamp DNA Mini Kit (QIAGEN, Valencia, CA, USA). DNA (3.5 μg) was loaded on 1% agarose gel and electrophoresed for 25 min at 100 V (constant voltage). After the gel was stained with ethidium bromide solution (10 mg/ml, Nippon Gene, Tokyo, Japan), the banding pattern was visualized under an image analyzer (Mini-Lumi, Berthold Technologies, Bad Wildbad, Germany).

2.11. GM-CSF clearance assay

GM-CSF clearance assay was performed as described previously [8]. To assess receptor-mediated binding and uptake of exogenous GM-CSF, 1×10^6 PBMCs or 4×10^5 TF-1 cells per well in a 24-well culture plate were incubated in RPMI 1640 (GIBCO BRL, Palo Alto, CA, USA) containing 10% FCS (Nichirei, Bioscience Inc, Tokyo, Japan) 100 mg/ml streptomycin and 100 U/ml penicillin under 5% CO_2 at 37 °C. rhGM-CSF was added at concentrations of 5 and 15 pM to PBMCs and TF-1 cells, respectively. The concentration of rhGM-CSF in the supernatant of each well was then measured at 5, 10, 24, and 48 h by ELISA.

2.12. Statistical analysis

Numerical data were evaluated for normal distribution by using Shapiro–Wilk tests. Parametric data are presented as means (±SE). Parametric data were analyzed by using one-way factorial ANOVA measurements. Multiple comparisons were performed through a Bonferroni-adjusted *t*-test, with non-significance set at $p > 0.05$. All tests were two-sided and *p* values <0.05 were considered statistically significant. Data were analyzed by using JMP (10.0.0) software (SAS, Cary, NC, USA).

3. Results

3.1. Molecular weight of rhGM-CSF

In this study, the bioactivity of rhGM-CSF derived from *E. coli*, yeast, and CHO cells was evaluated and compared. The mass spectrum of each GM-CSF shows distinct characteristic peaks: a single peak at 14.5 kDa for erhGM-CSF; peaks at 14.2, 14.4, and 15.0 kDa for yrhGM-CSF corresponding to a mean molecular weight of 14.7 kDa; and a number of peaks ranging from 16–28 kDa for crhGM-CSF corresponding to mean molecular weight of 19.0 kDa (Fig. 1A). The molar concentration of each rhGM-CSF was calculated from the original weight and volume, and then dividing by each mean molecular weight.

3.2. Short-term biological activity of rhGM-CSF

To compare the short-term bioactivity of the three rhGM-CSFs, we first evaluated the phosphorylation of STAT5 in monocytes and neutrophils stimulated for 0.5 h with the rhGM-CSFs. At both 15 and 60 pM rhGM-CSF, the percentage of pSTAT5-positive cells was significantly lower in crhGM-CSF-treated cells than in erhGM-CSF- or yrhGM-CSF-treated cells; whereas at 500 pM, this percentage was similar among the three rhGM-CSFs (Fig. 2A). Maximal values of CD11b stimulation indices at 60 pM of rhGM-CSF were 425 ± 15%, 576 ± 27%, and 625 ± 33%, for crhGM-CSF, erhGM-CSF, and yrhGM-CSF, respectively (Fig. 2B). These results indicate that the short-term effect of stimulation with crhGM-CSF was smaller than that with erhGM-CSF and yrhGM-CSF.

3.3. Intermediate-term biological activity of rhGM-CSF

When TF-1 cells were incubated with 30–120 pM rhGM-CSF for 24 h, the proliferation/survival was similar after treatment with crhGM-CSF, erhGM-CSF, and yrhGM-CSF (Fig. 3A). Likewise, MIP-1α production in PBMCs was not different among the three rhGM-CSFs at both 15 and 500 pM (Fig. 3B).

3.4. Long-term biological activity of rhGM-CSF

We then investigated the long-term biological effect of GM-CSF on TF-1 cells, monocytes, and PBMCs incubated for 72, 168, and 168 h, respectively. The effect on the proliferation/survival rate of TF-1 cells was significantly greater in cells incubated with 15 pM crhGM-CSF than that on cells incubated with the same concentration of erhGM-CSF or yrhGM-CSF. However, the effects were equivalent among the three rhGM-CSFs at 60 pM. The ED_{50} of each rhGM-CSF was 21 and 24 pM for erhGM-CSF and yrhGM-CSF, respectively, whereas it was 3.9 pM for crhGM-CSF (Fig. 4A). When monocytes were incubated with the GM-CSFs, the proliferation/survival rate was higher at 4 pM crhGM-CSF than that of cells incubated with the same concentration of other GM-CSFs. The ED_{50} was 10.7, 4.9, and 1.8 pM for erhGM-CSF, yrhGM-CSF, and crhGM-CSF, respectively (Fig. 4B). Similarly, the proliferation/survival rate of PBMCs was higher with 2–4 pM crhGM-CSF compared with that with other GM-CSFs (Fig. 4C). Proliferation/survival in the presence of goat anti-GM-CSF antibody was comparable, whereas the

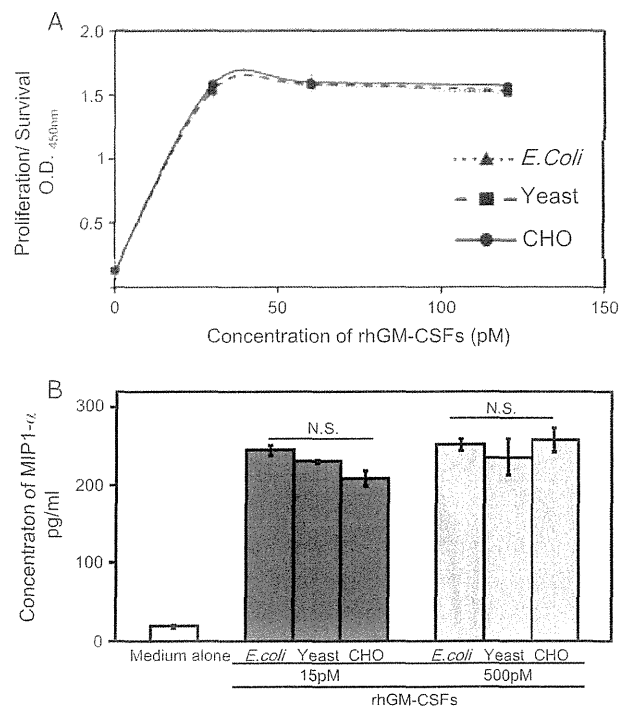

Fig. 3. Effect of intermediate-term stimulation (24 h) by rhGM-CSFs. (A) The proliferation/survival of TF-1 cells incubated for 24 h with various concentrations of rhGM-CSF derived from *E. coli* (▲), yeast (■), and CHO (●) was measured by MTT assay, as described in Section 2. The vertical axis indicates formazan formation expressed as the OD at 450 nm. (B) MIP-1α production of PBMCs incubated for 24 h with 0, 15, or 500 pM of *E. coli*, yeast-, and CHO-derived rhGM-CSF was measured by ELISA as described in Section 2.

inhibitory effect of the antibody was highest in crhGM-CSF. These data suggested that the effect of crhGM-CSF on the proliferation/survival of TF-1 cells was not due to oligosaccharide moieties but rather due to the GM-CSF peptide (Fig. 4D). After 3-day incubation with 30 pM erhGM-CSF, yrhGM-CSF, or crhGM-CSF, the number of viable TF-1 cells increased by multiples of 1.95 ± 0.5, 2.0 ± 0.7, and 6.45 ± 0.25, respectively, compared with the number of viable cells before incubation (Fig. 4E). The size histogram of TF-1 cells incubated with crhGM-CSF displays a bimodal pattern with a mean value of 24.09 μm, which is larger than that of erhGM-CSF-treated cells (22.09 μm) and yrhGM-CSF-treated cells (22.0 μm) (Fig. 4F). The viability of crhGM-CSF-stimulated TF-1 cells was significantly higher than that of TF-1 cells stimulated with other rhGM-CSFs. These results demonstrate that low concentrations of crhGM-CSF not only promote proliferation/survival but also stimulate the growth of these cells more efficiently than do erhGM-CSF and yrhGM-CSF, and that the long-term effect of rhGM-CSF differs from the short- and intermediate-term outcomes. The long-term effects of erhGM-CSF and yrhGM-CSF for each condition were similar.

3.5. Modified bioactivity of crhGM-CSF after treatment with sialidase

To investigate the effect of sialyl residues located at the distal end of the oligosaccharide moieties [20] on cell proliferation/survival, we studied sialidase-treated crhGM-CSF. After treatment, mass spectrometry revealed a drastic reduction in the intensity of peaks corresponding to mono-, di-, tri-, and tetra-sialyl carbohydrates (Fig. 5A). This is also consistent with the banding pattern

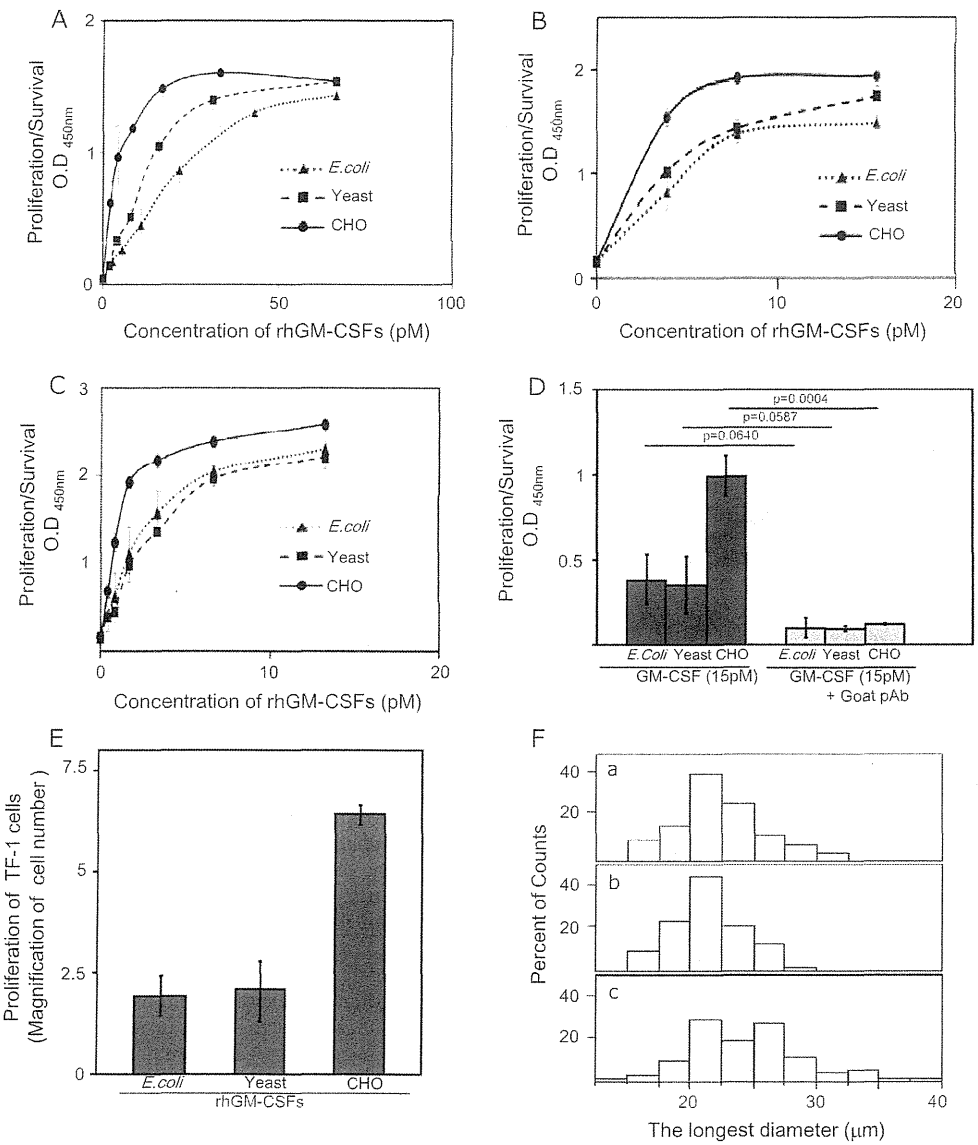

Fig. 4. Effect of long-term stimulation with various concentrations of rhGM-CSFs on the proliferation/survival of TF-1 cells, monocytes, and PBMCs. (A) Proliferation/survival of TF-1 cells incubated for 72 h with various concentrations (0–60 pM) of rhGM-CSF derived from *E. coli* (▲), yeast (■), and CHO (●) was measured as described in Section 2. (B) Proliferation/survival of monocytes incubated for 168 h with various concentrations (0–15 pM) of rhGM-CSF derived from *E. coli* (▲), yeast (■), and CHO (●) was measured as described in Section 2. (C) Proliferation/survival of PBMCs incubated for 168 h with various concentrations (0–15 pM) of rhGM-CSF derived from *E. coli* (▲), yeast (■), and CHO (●) was measured as described in Section 2. (D) Effect of neutralizing goat anti-*E. coli*-derived GM-CSF antibody on the proliferation/survival of TF-1 cells incubated with 15 pM rhGM-CSFs. The vertical axis is the proliferation/survival of TF-1 cells (OD at 450 nm). (E) Magnification of proliferation was measured by enumerating viable TF-1 cells under a phase contrast microscopy before and after 72 h incubation with 30 pM rhGM-CSFs. (F) Size distribution of TF-1 cells incubated with 15 pM rhGM-CSFs for 72 h. The horizontal axis is the largest diameter of cells and the vertical axis is the number of cells.

obtained by SDS–PAGE, in which several bands characteristic for crhGM-CSF are absent or weaker (Fig. 5B). Desialylation of crhGM-CSF markedly reduced the proliferation/survival rates to levels observed with erhGM-CSF- or yrhGM-CSF-treated cells (Fig. 5C). These results suggest that the up-regulated proliferation/survival induced by crhGM-CSF is likely due to its sialyl residues.

3.6. The effect of GM-CSF on apoptosis of TF-1 cells

The effect of GM-CSF on the apoptosis of TF-1 cells was evaluated by Annexin V expression with flow cytometry. When TF-1 cells were incubated with 30 pM crhGM-CSF for 3 days, 8.8% of the cells were apoptotic. In contrast, 17.0%, 21.4%, and 15.9% of cells were apoptotic upon incubation with erhGM-CSF, yrhGM-CSF, and sialidase-treated crhGM-CSF, respectively (Fig. 6A). TF-1 cells incubated with crhGM-CSF had fewer vacuolated nuclei and coagulated chromatin than those of cells incubated with other GM-CSFs (Fig. 6B). TF-1 cell apoptosis was also confirmed by DNA ladder formation via agarose gel electrophoresis (Fig. 6C). These results suggested that apoptotic TF-1 cells were less frequently observed in the presence of low concentration of crhGM-CSF than erhGM-CSF, yrhGM-CSF and sialidase-treated crh GM-CSF as TF-1 cells are GM-CSF dependent cell line. It is plausible that GM-CSF bioactivity is likely to remain in culture supernatant of the cells incubated with crhGM-CSF compared with other rhGM-CSFs.

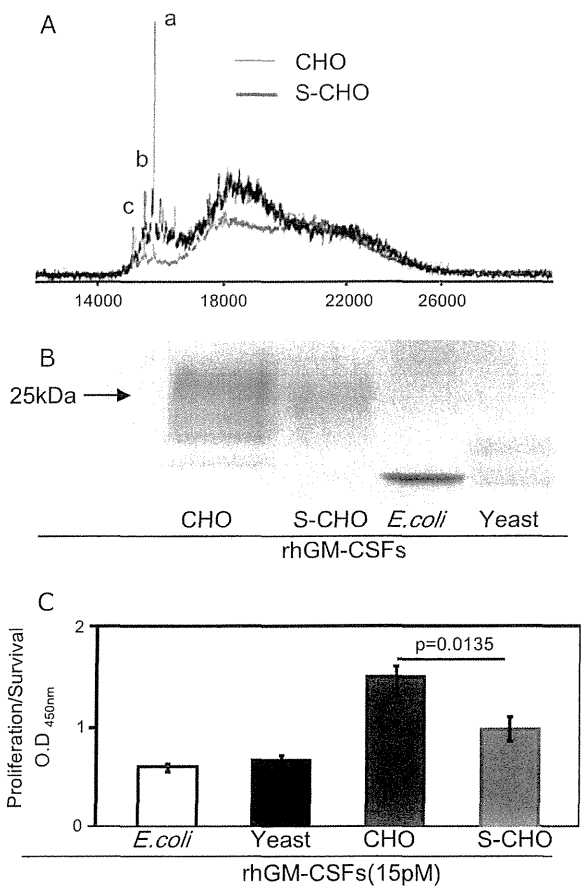

Fig. 5. Sialidase treatment of CHO-cell-derived rhGM-CSF and its biological activity. (A) Mass spectra of CHO cell-derived rhGM-CSF before (blue line) and after (red line) treatment with sialidase. (B) SDS–PAGE appearance of CHO cell-derived GM-CSF, CHO cell-derived GM-CSF after sialidase treatment, *E. coli*-derived GM-CSF, and yeast-derived GM-CSF. (C) The effect of sialidase treatment on the proliferation/survival of TF-1 cells after 72 h incubation with *E. coli*-, yeast-, CHO cells-derived rhGM-CSF or CHO cells-derived rhGM-CSF after sialidase treatment.

3.7. Clearance of rhGM-CSF by TF-1 cells and PBMCs

The clearance of crhGM-CSF by TF-1 cells and PBMCs was delayed compared with that of other GM-CSFs. After 24 and 48 h clearance assays, 13% and 9.5% of the initial crhGM-CSF concentration remained in the culture supernatant of PBMCs, whereas only 4.5% and 1.1% of erhGM-CSF, and 3.1%, 1% of yrhGM-CSF and 5.6% and 2.7% of sialidase-treated crhGM-CSF remained, respectively (Fig. 7A). On the other hand, after 24 and 48 h clearance assays, 7.5% and 3% of the initial crhGM-CSF concentration remained in the culture supernatant of TF-1 cells, whereas only 1.3% and 1.1% of erhGM-CSF, 1.1% and 1.0% of yrhGM-CSF and 2.9% and 2.7% of sialidase-treated crhGM-CSF remained, respectively (Fig. 7B). After 48 h incubation with erhGM-CSF, yrhGM-CSF, and sialidase treated crhGM-CSF, 15 pM of the same rhGM-CSF was except for crhGM-CSF added into each well. As shown in Fig. 7C, addition of each rhGM-CSF improved the proliferation/survival of TF-1 cells in the next 24 h reaching a similar level of those incubated with original 15 pM of crhGM-CSF for three days. Taken together with the data of proliferation/survival assay, it is likely that delayed clearance crhGM-CSF might prolong its biological activity *in vitro* (Fig. 7C).

4. Discussion

A number of studies have reported the expression of human GM-CSF by using natural or recombinant cells. These studies revealed that mammalian cells secrete GM-CSF proteins with variable molecular masses [20]. It has also been shown that its properties such as pharmacokinetics, binding affinity to the GM-CSF receptor, bioactivity, and immunogenicity are affected by glycosylation. In the present study, we demonstrated that compared with erhGM-CSF or yrhGM-CSF, crhGM-CSF promoted more efficiently the proliferation/survival of TF-1 cells, especially at low concentrations. In contrast to the results of the present study, natural hGM-CSF is thought to have lower biological activity with increasing glycosylation [15,24]. The pattern of glycosylation on GM-CSF has been found to affect its specific biological activity. Non-human expression systems such as yeast-, CHO cell-, or COS cell-derived rhGM-CSFs have distinct carbohydrate moieties and show different biological activities [18,28]. The half-life of hGM-CSF injected into rats decreases upon deglycosylation, indicating that the carbohydrate moieties influence the clearance, increase the stability, or alter the distribution of hGM-CSF. The carbohydrate structure of hematopoietic growth factors may therefore be important in determining their effective half-life *in vivo*. In this regard, we confirmed that *in vitro* GM-CSF clearance was also affected largely by the carbohydrate moieties of GM-CSF, especially its sialyl residues at the distal end of the oligosaccharide moieties.

The significance of the glycosylation of hematopoietic growth factors has been investigated previously. First, it is important for the secretion of glycoproteins. Erythropoietin secretion is prevented by site-directed mutagenesis of the N- or O-linked glycosylation sites [29–31]. As tunicamycin does not interfere with secretion of hGM-CSF, the N-linked carbohydrate is not crucial for this process [32]. Second, the N-linked carbohydrate influences the biological activity and receptor binding of other glycoprotein hormones and cytokines [29,33]. The *in vitro* activity of erythropoietin requires oligosaccharide moieties, but N-linked carbohydrates markedly reduce the *in vitro* activity of calcitonin. Glycosylation of luteinizing hormone is required for signal transduction, although

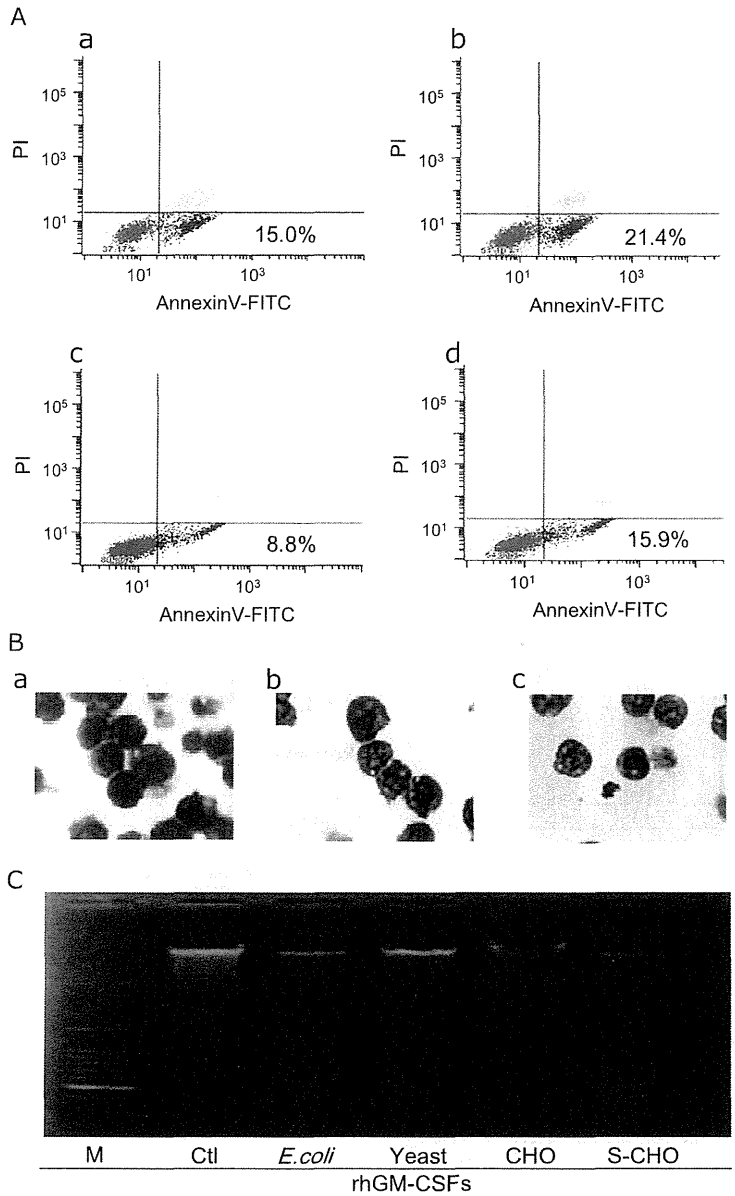

Fig. 6. Apoptosis of TF-1 cells incubated with 30 pM rhGM-CSFs for 72 h. (A) Flow cytometry results for apoptotic TF-1 cells incubated with rhGM-CSF derived from *E. coli* (a), yeast (b), CHO cells (c), or sialidase-treated CHO cells (d). The horizontal axis is the fluorescence intensity of Annexin V-FITC and the vertical axis is the fluorescence intensity of propidium iodide. (B) Morphology of TF-1 cells incubated with GM-CSF derived from CHO cells (a), yeast (b), and *E. coli* (c) at high magnification (1000×). Cells were cytocentrifuged and stained with Diff-Quick stain. (C) Agarose gel electropherogram of DNA extracted from TF-1 cells incubated with *E. coli*-, yeast-, CHO cells-derived rhGM-CSF or CHO cells-derived rhGM-CSF after sialidase treatment.

deglycosylated luteinizing hormone has higher receptor binding affinity [33]. Similarly, deglycosylation of hGM-CSF increases the receptor binding affinity [15]. However, in contrast to hGM-CSF, the most active forms are heavily glycosylated in luteinizing hormone [32].

Sialyl residues on carbohydrates in crhGM-CSF are considered crucial to the upregulation of the proliferation/survival of TF-1 cells because desialylation remarkably reduces this effect. Various sialylated forms of GM-CSF are produced in various tissues of mice and confer different physicochemical characteristics to murine GM-CSF [34]. Molecular weights of GM-CSF purified from various organs range from 37 to 200 kDa [32]; thus, it is possible that the bioactivity of GM-CSF produced in different tissues is regulated by the degree of sialylation. Since sialyl residues at the distal end of oligosaccharides can affect the specific activity of hGM-CSF as well as its isoelectric points and affinities to the GM-CSF receptor, sialylation may alter the activity of hGM-CSF in a tissue-specific manner. The aforementioned studies are clinically important because therapy using hGM-CSF has been associated with side effects, which may relate to its activities as a mediator of inflammation rather than to its function as a growth factor [15]. If different glycosylation patterns allow hGM-CSF activity to be regulated, manipulation of the carbohydrate moieties may enable reduction of the inflammatory mediator effects of hGM-CSF without affecting the stimulation of myeloid cell production.

GM-CSF exerts its biological activities by binding to specific high-affinity cell-surface receptors. After binding, the ligand/receptor complex is rapidly internalized in most hematopoietic cells [35,36]. It is not fully known whether the turnover time of this internalization differs between different rhGM-CSFs. It is possible

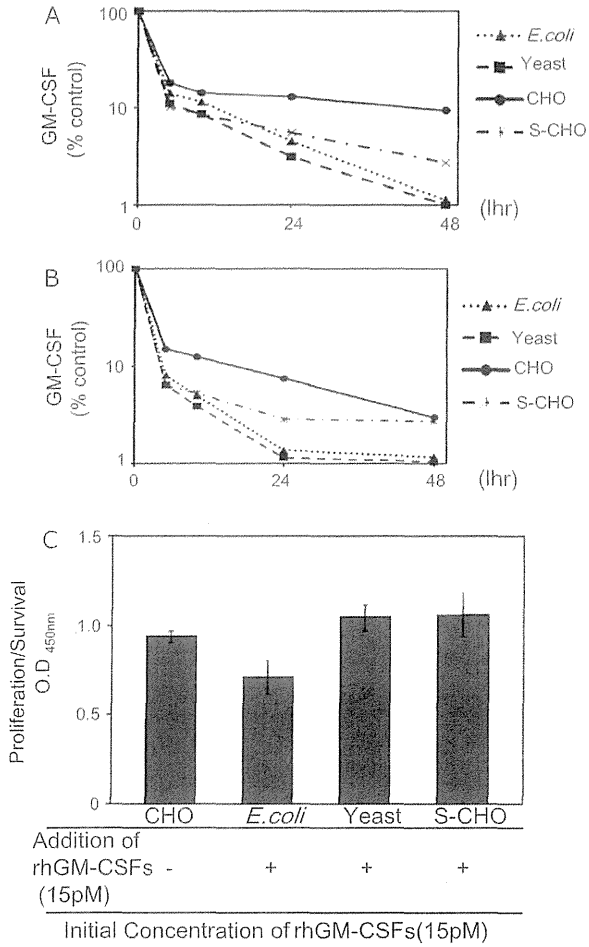

Fig. 7. GM-CSF clearance assay of TF-1 cells and peripheral blood mononuclear cells. (A) Peripheral blood mononuclear cells were incubated for 0–48 h with each 15 pM of *E. coli*-, yeast-, CHO cells-derived rhGM-CSF or CHO cells-derived rhGM-CSF after sialidase treatment. (B) TF-1 cells were incubated for 0–48 h with each 15 pM of *E. coli*-, yeast-, CHO cells-derived rhGM-CSF or CHO cells-derived rhGM-CSF after sialidase treatment. The horizontal axis is the time after the start of incubation. The vertical access is percent each rhGM-CSF concentration per initial concentration at each time point in the culture supernatant. (C) After 48 h incubation with *E. coli*-, yeast-, CHO cells-derived rhGM-CSF or CHO cells-derived rhGM-CSF after sialidase treatment, 15 pM of the same rhGM-CSF was added into each well. The vertical axis is the proliferation/survival of TF-1 cells (OD at 450 nm).

that the oligosaccharide sialyl residue of crhGM-CSF can attenuate its binding to the low-affinity rhGM-CSF receptor α and/or associate with the rhGM-CSF β chain, resulting in downregulation of signal transduction and delayed clearance of the molecule [15]. The present study revealed that stimulation with low concentrations of crhGM-CSF augmented STAT5 phosphorylation less effectively than did low concentrations of erhGM-CSF and yrhGM-CSF. The sialyl residue may prolong the turnover cycle (known to be 40 s for erhGM-CSF) and thus maintain rhGM-CSF bioactivity for a longer period [35]. In the future, it is necessary to determine whether the sialyl residues of GM-CSF attenuate its binding to low-affinity receptors on hematopoietic cells or delay the process of its internalization into cells.

5. Conclusion

We have demonstrated for the first time that sialylated oligosaccharide moieties prolong the proliferation/survival of rhGM-CSF *in vitro*. Further studies are warranted to determine the correlation of the oligosaccharide structure of crhGM-CSF with both signal transduction and internalization.

Authorship

A. Hashimoto and K. Nakata wrote the manuscript and designed the project.

A. Hashimoto performed experiments. Y. Ito and A. Yamagata assisted technical issues. T. Tanaka and N. Kitamurta contributed to the statistical analysis of data. R. Tazawa participated in preparation of materials. K. Nakagaki provided variable information for methods. All authors read and approved the final manuscript.

Disclosure

The authors declare that they have no competing interests.

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