

manifestations of a blockade of liver blood outflow, but the elevation in inferior vena cava pressure caused by a sharp rise in intrapleural pressure using MI-E might have been a potential cause of peliosis hepatis.

In conclusion, peliosis hepatis is a rare but important fatal complication that may occur more often once genetic or other therapies for XLMTM become available with a resulting increase in life expectancy. To avoid fatal hepatic hemorrhage from peliosis hepatis, routine liver function tests and abdominal imaging studies are recommended for all XLMTM patients. In addition, it might be necessary to carefully check liver imaging tests, especially at the time of using mechanical ventilation.

## References

- [1] Spiro AJ, Shy GM, Gonatas NK. Myotubular myopathy. Persistence of fetal muscle in an adolescent boy. *Arch Neurol* 1966;14:1–14.
- [2] Romero NR. Centronuclear myopathies: a widening concept. *Neuromuscul Disord* 2010;20:223–8.
- [3] Laporte J, Biancalana V, Tanner SM, et al. MTM1 mutation in X-linked myotubular myopathy. *Hum Mutat* 2000;15:393–409.
- [4] Herman GE, Finegold M, Zhao W, Gouyon B, Metznerberg A. Medical complications in long-term survivors with X-linked myotubular myopathy. *J Pediatr* 1999;134:206–14.
- [5] Wang SY, Ruggles S, Vade A, Newman BM, Borge MA. Hepatic rupture by peliosis hepatis. *J Pediatr Surg* 2001;36:1456–9.
- [6] Karger B, Varchmin-Schultheib K, Fechner G. Fatal hepatic haemorrhage in a child-peliosis hepatis versus maltreatment. *Int J Legal Med* 2005;119:44–6.
- [7] Yano T, Toyono M, Watanabe Y, Sawaishi Y. A 5-year old boy of X-linked myotubular myopathy died of hepatic rapture from peliosis hepatis (in Japanese). *No To Hattatsu* 2010;42:S348.
- [8] Tsai TC, Horinouchi H, Noguchi S, et al. Characterization of *MTM1* mutations in 31 Japanese families with myotubular myopathy, including a patient carrying 240 kb deletion in Xq28 without male hypogenitalism. *Neuromuscul Disord* 2005;15:245–52.
- [9] Zak FG. Peliosis hepatis. *Am J Pathol* 1950;26:1–15.
- [10] Simon DM, Krause R, Galambos JT. Peliosis hepatis in a patient with marasmus. *Gastroenterology* 1988;95:805–9.
- [11] Perkocha LA, Geaghan SM, Yen B, et al. Clinical and pathological features of bacillary peliosis hepatis in association with human immunodeficiency virus infection. *N Engl J Med* 1990;323:1581–6.
- [12] Scoazec J-Y, Marche C, Girard P-M, et al. Peliosis hepatis and sinusoidal dilatation during infection by human immunodeficiency virus (HIV). *Am J Pathol* 1988;131:38–47.
- [13] Testa G, Panaro F, Sankary H, et al. Peliosis hepatis in a living related liver transplantation donor candidate. *J Gastroenterol Hepatol* 2006;21:1075–7.
- [14] Bagheri SA, Boyer JL. Peliosis hepatis associated with androgenic-anabolic steroid therapy. A severe form of hepatic injury. *Ann Intern Med* 1974;81:610–8.
- [15] Koehler JE, Sanchez MA, Garrido CS, et al. Molecular epidemiology of bartonella infections in patients with bacillary angiomatosis–peliosis. *N Engl J Med* 1997;337:1876–83.
- [16] Hyodo M, Mogensen AM, Larsen PN, et al. Idiopathic extensive peliosis hepatis treated with liver transplantation. *Hepatobiliary Pancreat Surg* 2004;11:371–4.
- [17] Tsigiotis P, Sella T, Shapira MY, et al. Peliosis hepatis following treatment with androgen-steroids in patients with bone marrow failure syndromes. *Haematologica* 2007;92:e106–10.
- [18] Battal B, Kocaoglu M, Atay AA, Bulakbasi N. Multifocal peliosis hepatis: MR and diffusion-weighted MR-imaging findings of an atypical case. *Ups J Med Sci* 2010;115:153–6.
- [19] Jacquemin E, Pariente D, Fabre M, Huault G, Valayer J, Bernard O. Peliosis hepatis with initial presentation as acute hepatic failure and intraperitoneal hemorrhage in children. *J Hepatol* 1999;30:1146–50.
- [20] Hayward SR, Lucas CE, Ledgerwood AM. Recurrent spontaneous intrahepatic hemorrhage from peliosis hepatis. *Arch Surg* 1991;126:782–3.
- [21] Iannaccone R, Federle MP, Brancatelli G, et al. Peliosis hepatis: spectrum of imaging findings. *Am J Roentgenol* 2006;187:W43–52.
- [22] Miske LJ, Hickey EM, Kolb SM, Weiner DJ, Panitch HB. Use of the mechanical in-exsufflator in pediatric patients with neuromuscular disease and impaired cough. *Chest* 2004;125:1406–12.
- [23] Degott C, Rueff B, Kreis H, Duboust A, Potet F, Benhamou JP. Peliosis hepatis in recipients of renal transplants. *Gut* 1978;19:748–53.
- [24] Zafrani ES, Cazier A, Baudelot AM, Feldmann G. Ultrastructural lesions of the liver in human peliosis. A report of 12 cases. *Am J Pathol* 1984;114:349–59.



## Limb-girdle muscular dystrophy type 2I is not rare in Taiwan

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### Abstract

Alpha-dystroglycanopathy is caused by the glycosylation defects of  $\alpha$ -dystroglycan ( $\alpha$ -DG). The clinical spectrum ranges from severe congenital muscular dystrophy (CMD) to later-onset limb girdle muscular dystrophy (LGMD). Among all  $\alpha$ -dystroglycanopathies, LGMD type 2I caused by *FKRP* mutations is most commonly seen in Europe but appears to be rare in Asia. We screened uncategorized 40 LGMD and 10 CMD patients by immunohistochemistry for  $\alpha$ -DG and found 7 with reduced  $\alpha$ -DG immunostaining. Immunoblotting with laminin overlay assay confirmed the impaired glycosylation of  $\alpha$ -DG. Among them, five LGMD patients harbored *FKRP* mutations leading to the diagnosis of LGMD2I. One common mutation, c.948delC, was identified and cardiomyopathy was found to be very common in our cohort. Muscle images showed severe involvement of gluteal muscles and posterior compartment at both thigh and calf levels, which is helpful for the differential diagnosis. Due to the higher frequency of LGMD2I with cardiomyopathy in our series, the early introduction of mutation analysis of *FKRP* in undiagnosed Taiwanese LGMD patients is highly recommended.

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### 1. Introduction

Alpha-dystroglycanopathy is a group of muscular dystrophies caused by altered glycosylation of  $\alpha$ -dystroglycan ( $\alpha$ -DG), which is one of the components of dystrophin–glycoprotein complex [1,2]. The clinical

phenotypes form a broad spectrum, ranging from severe congenital muscular dystrophy (CMD) with or without ocular and central nervous system involvement to later-onset limb girdle muscular dystrophy (LGMD) [3–5].

A number of genes have been reported to cause  $\alpha$ -dystroglycanopathy, including *POMT1*, *POMT2*, *POMGnT1*, *FKTN*, *FKRP*, and *LARGE* that are known to be involved in glycosylation of  $\alpha$ -DG, and *DAG1*, which encodes DG itself [6–11]. Recently, the number of genes associated with  $\alpha$ -dystroglycanopathy has been increasing to include *ISPD*, *TMEM5*, *GTDC2*, *B3GNT1*, *DOLK*, *DPM2* and *DPM3* [12–19]. Patients with all

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kinds of  $\alpha$ -dystroglycanopathy are inherited with autosomal recessive trait.

Among those causative genes for  $\alpha$ -dystroglycanopathy, *FKRP* mutations are the most frequently seen in the Caucasian population, causing LGMD2I and congenital muscular dystrophy type 1C (MDC1C). In the Asian population, on the other hand, the most common  $\alpha$ -dystroglycanopathy is Fukuyama congenital muscular dystrophy and LGMD2M caused by the mutations in *FKTN* [20–24]. This phenomenon may be caused by the founder effect of c.826C>A substitution in *FKRP* and the ancestral insertion of a SINE-VNTR-Alu (SVA) retrotransposon in *FKTN* in different geographic areas [21,25]. Recently, an increasing number of patients having *FKTN* mutations were identified outside Asia but so far few Asian patients with LGMD2I caused by *FKRP* mutations have been reported [26–29].

In this study, we found that LGMD2I is common in the Taiwanese patients with  $\alpha$ -dystroglycanopathy due to a common mutation, c.948delC (p.Cys317Alafs\*111), which may cause more severe phenotype and cardiomyopathy.

## 2. Materials and methods

### 2.1. Patients

Forty patients clinically and pathologically diagnosed as LGMD and 10 patients with CMD who received muscle biopsy in Kaohsiung Medical University Hospital from January, 2008 to December, 2011 were enrolled. LGMD was defined as progressive proximal-dominant muscle weakness with characteristic dystrophic changes in muscle pathology. CMD was recognized as infantile floppiness with dystrophic muscle. Patients with deficiencies of dystrophin, sarcoglycans, dysferlin, merosin or collagen VI were excluded by immunohistochemistry beforehand. All merosin deficiency patients were confirmed to have *LAMA2* mutations [30]. This study was approved by the institutional review board of the Kaohsiung Medical University Hospital.

### 2.2. Histochemistry

Biopsied muscle specimens were frozen in isopentane cooled in liquid nitrogen. A serial frozen section was stained by a battery of histochemical methods including hematoxylin and eosin (H&E), modified Gomori-trichrome (mGt) and NADH-tetrazolium reductase (NADH-TR).

### 2.3. Immunohistochemistry

Frozen sections of 6  $\mu$ m thickness were used for immunohistochemistry according to the standard protocols with Vantana Benchmark automated stainer. Primary antibodies used in this study were monoclonal anti- $\alpha$ -DG (VIA4-1; Upstate Biotechnology, Lake Placid,

NY, USA) and anti- $\beta$ -DG (43DAG1/8D5; Novocastra Laboratories, Newcastle upon Tyne, UK) antibodies.

### 2.4. Immunoblotting and laminin overlay assay

The detailed techniques of immunoblotting, and laminin overlay assay have been described previously [31]. The following antibodies were used for immunoblotting analysis: monoclonal anti- $\alpha$ -DG (VIA4-1) and polyclonal anti- $\alpha$ -DG (GT20ADG, kindly provided by Prof. K. Campbell, Iowa Univ.), polyclonal anti-laminin-1 (Sigma, St. Louis, MO, USA), and monoclonal anti- $\beta$ -DG (43DAG1/8D5).

### 2.5. Mutation analyses of $\alpha$ -DGP associated genes

Genomic DNA was extracted from leukocytes in peripheral blood lymphocytes according to standard protocols. All exons and their flanking intronic regions of *FKRP* (NM\_024301.4), *FKTN* (NM\_001079802.1), *POMGnT1* (NM\_001243766.1), *POMT1* (NM\_007171.3), *POMT2* (NM\_013382.5), and *LARGE* (NM\_004737.4) were amplified and sequenced using an automated 3100 DNA sequencer (Applied Biosystems, Foster, CA, USA). Primer sequences are available upon request. DNA samples from 100 Taiwanese individuals without apparent neuromuscular disorders were analyzed as controls.

## 3. Results

### 3.1. Patients with $\alpha$ -DGP caused by *FKRP* mutations

Seven of 50 patients with unclassified LGMD and CMD had a reduced  $\alpha$ -DG immunoreaction using VIA4-1 antibody, which recognizes glycosylated forms of  $\alpha$ -DG on muscles, and they were thus considered to have  $\alpha$ -dystroglycanopathy (Fig. 1). Among these seven patients, six had LGMD phenotype and one was CMD. Mutation screening revealed that five LGMD patients from four families harbored *FKRP* mutations (Fig. 2). No mutation in *FKTN*, *POMGnT1*, *POMT1*, *POMT2* and *LARGE* was identified in these seven patients. The clinical, pathological and biochemical information of all five patients with *FKRP* mutations are summarized in Table 1 together with that of a previously reported Taiwanese LGMD2I patient who was the first reported case in East-Asia (Patient 6) [26]. The c.948delC (p.Cys317Alafs\*111) mutation was found heterozygously in four newly diagnosed patients (Patients 2, 3, 4 and 5) as well as in Patient 6. Patients 2, 3, and 4 carried a c.545A>G (p.TyrY182Cys) mutation, which was previously reported in two Brazilian patients, and a c.823C>T (p.Arg275Cys) mutation was identified in Patients 5 and 6. The compound heterozygous mutations for Patients 2, 5, and 6 were also found to lie on different parental alleles. Patient 1 bears two different novel

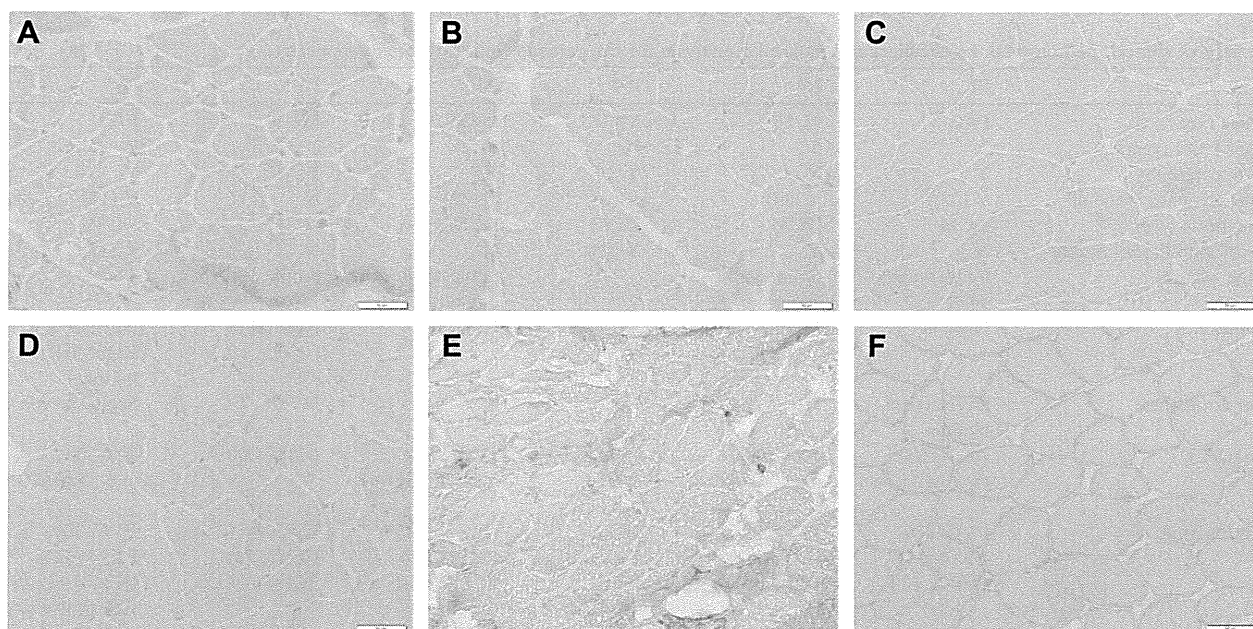


Fig. 1. Immunohistochemistry for  $\alpha$ -DG (VIA4-I) in Patients 1, 2, 4, 5, and 6 (A–E). All patients' muscle samples showed markedly reduced staining, as compared with controls (F). Bar: 50  $\mu$ m.

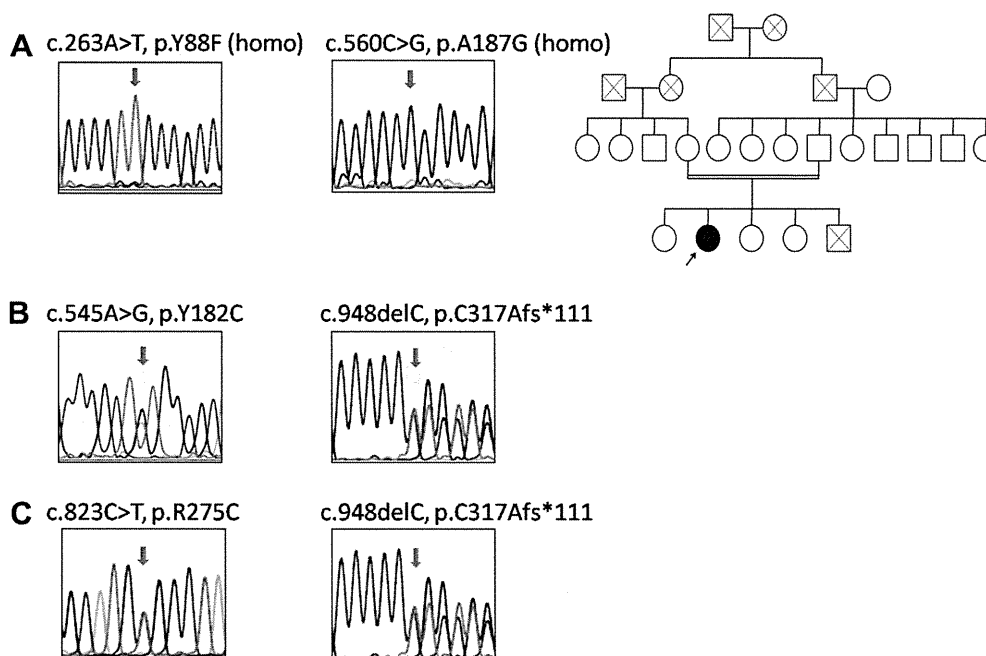


Fig. 2. Sequence analysis of *FKRP* revealed homozygous c.263A>T and c.560C>G mutations in Patient 1 (A), compound heterozygous c.545A>G and c.948delC mutations in Patients 2, 3, and 4 (B), and compound heterozygous c.823C>T and c.948delC mutations in Patients 5 and 6 (C). The pedigree of Patient 1 is also shown; the youngest brother of Patient 1 died of unknown causes at 7 months of age (A).

homozygous mutations, c.263A>T (p.Try88Phe) and c.560C>G (p.Ala187Gly), neither of which were identified in the human genome mutation database (HGMD) and 100 healthy individuals. The consanguineous healthy parents of Patient 1 carried these two missense mutations, heterozygously.

### 3.2. Reduced glycosylation of $\alpha$ -DG in *LGMD2I* patients

We further confirmed the altered glycosylation of  $\alpha$ -DG in our *LGMD2I* patients (P2, 4, 5 and 6) using immunoblotting analysis and laminin overlay assay. On immunoblotting analysis using VIA4-1 antibody, skeletal

Table 1

Summary of clinical, pathological, biochemical and molecular analyses for the patients with  $\alpha$ -dystroglycanopathy.

	P1	P2	P3 <sup>a</sup>	P4 <sup>a</sup>	P5	P6 <sup>b</sup>
Sex/age (years)	F/35	M/16	M/31	M/30	M/10	F/23
Age of onset (years)	2	5	10	17	2	2
Calf hypertrophy	Y	Y	Y	Y	Y	Y
Cardiomyopathy (age of diagnosis, years)	Y (28) DCM	Y (14) DCM	Y (31) DCM	Y (30) DCM	N	Y (17) DCM
Loss of ambulatory ability (age, years)	N 6-min walk: 76 m	Y 6-min walk: 343 m	Y (29)	N 6-min walk: not done	N 6-min walk: 210 m	Y (12)
Cognition	Normal	Normal	Normal	Normal	Normal	Normal Brain MRI: negative finding
Other anomalies	Over-active bladder	N	N	N	N	Scoliosis with op
CK (IU/L)	1000–1500 max: unknown	4000–8000 max: >10,000	1500–2000 max: unknown	1500–2000 max: unknown	6000–9000	200–500 max: >10,000
Lung function	FVC: 32% FEV1: 33% PCF: 2.12 L/s	FVC: 45% FEV1: 53% PCF: 6.91 L/s	FVC: 43% FEV1: 36% PCF: 3.46 L/s	FVC: 62% FEV1: 73% PCF: 3.7 L/s	FVC: 64% FEV1: 76% PCF: 4.64 L/s	FVC: 10% FEV1: 12% PCF: 0.42 L/s BiPAP use at night
FKRP mutations	c.263A>T homo (F & M, hetero) c.560C>G homo (F & M, hetero)	c.545A>G (F, hetero) c.948delC (M, hetero)	c.545A>G c.948delC	c.545A>G c.948delC	c.823C>T (M, hetero) c.948delC (F, hetero)	c.823C>T (F, hetero) c.948delC (M, hetero)

Y: yes; N: nil; DCM: dilated cardiomyopathy; min: minute; m: meter; op: operation; max: maximum; F: Father; M: Mother.

<sup>a</sup> Siblings.<sup>b</sup> Previously reported (Reference [21]).

muscles from all four patients showed fainter and smaller sized bands than the control (Fig. 3A). With GT20ADG antibody for the core region of  $\alpha$ -DG, all skeletal muscles from these patients showed fainter broadbands with smaller molecular mass than that detected in the control (Fig. 3B). Laminin overlay assay displayed greatly reduced binding ability of  $\alpha$ -DG to laminin in all patients (Fig. 3C).

### 3.3. Clinical findings of LGMD2I patients (Table 1)

The mean age of all 6 LGMD2I patients at examination was  $24.2 \pm 9.7$  years, and the mean disease duration was  $17.8 \pm 9.1$  years. The disease onset was variable, ranging from early childhood to late teens ( $2-17$  years;  $6.3 \pm 6.1$ ). All patients had calf hypertrophy and proximal dominant muscle weakness, starting from lower extremities and

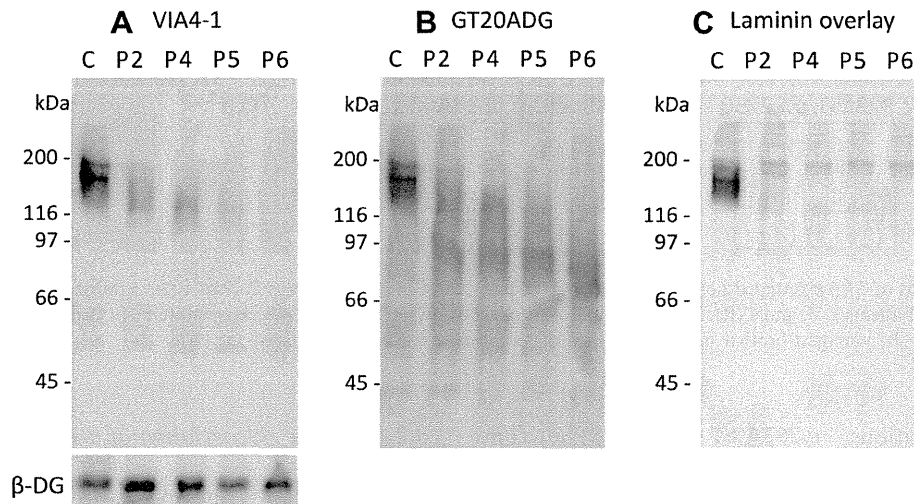


Fig. 3. Immunoblotting analysis. All 4 patients (P2, P4, P5 and P6) examined showed fainter and smaller sized bands than controls using  $\alpha$ -DG (VIA4-1) (A). With the antibody of GT20ADG, fainter broadbands with smaller molecular weights were detected (B). Laminin overlay assay displayed greatly reduced binding ability of  $\alpha$ -DG to laminin in all patients (C).

then extending to shoulder girdle and arms. Patient 2 became wheelchair-bound at the age of 29 years while Patient 6 lost her ambulatory ability at 14 years of age. Dilated cardiomyopathy (DCM) was seen in five of six patients (83.3%) and they are currently under medication. DCM was diagnosed with echocardiogram in Patients 2, 3, and 4 at their first visit to our hospital, so that the exact onset age of cardiac involvement was unclear. All patients had impaired pulmonary function with different degrees of severity but only Patient 6 required ventilator assistance (1 in 6; 16.7%). All patients had normal cognitive functions and the brain MRI of Patient 6 showed no notable abnormal changes. As for other abnormalities, only Patient 6 received an operation for scoliosis at 13 years of age. Serum creatine kinase levels were usually up to 10,000 IU/L at disease onset and then declined to hundreds at a later stage.

#### 3.4. Muscle CT of LGMD2I patients

On muscle CT, all assessed patients (Patients 1–5) showed similar patterns of muscle involvement (Fig. 4) Lower extremities were more severely affected than upper extremities. Gluteus maximus was the most affected muscle (Fig. 4A), followed by posterior compartment of thigh muscles, among which biceps femoris and then adductors showed marked hypodensity (Fig. 4B). In the anterior compartment of thigh, vastus muscles and rectus femoris were equally involved. At the calf level, posterior compartment muscles, especially gastrocnemius and

soleus, were also more affected than anterior part (Fig. 4C). As for upper extremities, involvement of shoulder girdle muscles including subscapularis, infraspinatus and supraspinatus was more prominent than trapezium and deltoid muscles (Fig. 4D).

#### 4. Discussion

Wide variability in clinical picture has been reported in LGMD2I, of which the clinical features can be Duchenne muscular dystrophy-like, late-onset LGMD phenotypic and even asymptomatic [32,33]. In European countries, homozygosity of the most common missense mutation of c.826C>A (p.Leu276Ile) has been reported to confer a relatively milder phenotype than patients with compound heterozygous mutations [34]. A homozygous mutation of c.545A>G identified in the Brazilian patients has previously been reported to cause mild clinical phenotypes and disease course [32]. In our series, Patients 2–4 harbor the same compound heterozygous mutations of c.545A>G and c.948delC while Patients 5 and 6 both carry the same c.823C>T and c.948delC mutations. The patients carrying c.823C>T and c.948delC seem to show more severe clinical features than the patients having c.545A>G and c.948delC in terms of the age at onset, disease course, motor deterioration and complications. Because only a limited number of patients were included, however, additional patients with each mutation are required to clarify the phenotype and genotype correlation more clearly.

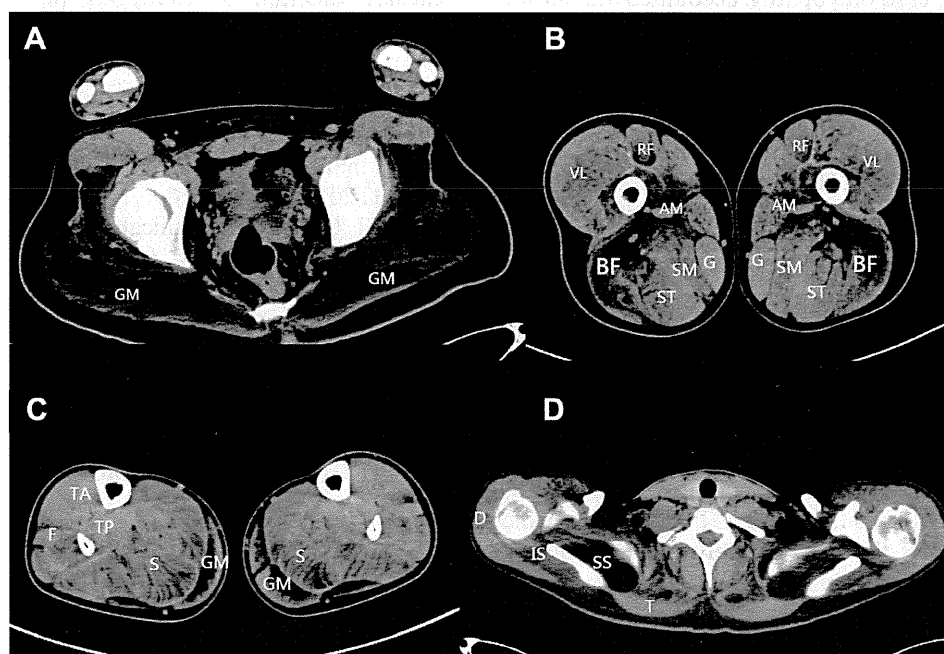


Fig. 4. Muscle CT on Patient 2. Gluteus maximus muscles were severely affected (A), followed by biceps femoris and adductors (B). At the calf level, gastrocnemius and soleus muscles were severely involved (C). In the upper extremities, involvement of subscapularis, infraspinatus and supraspinatus were more severe than trapezium and deltoid (D). (D: deltoid; IS: infraspinatus; SC: subscapularis; BF: biceps femoris; ST: semitendinosus; SM: semimembranosus; S: soleus; F: fibularis; G: gastrocnemius; GM: gluteus maximus; RF: rectus femoris; VL: vastus lateralis; AM: adductor magus; G: gracilis).

Noteworthy, c.948delC in *FKRP* is a common mutation in Taiwanese LGMD2I patients. The mutation could cause frame shift and premature termination in translation (p.Cys317Alafs\*111). We further screened 300 controls without neuromuscular diseases to determine the carrier frequency of c.948delC but none carried this mutation. This result suggests that the prevalence of the homozygosity of c.948delC is at least lower than 1 in 360,000, which may be too low to identify a homozygous patient. On the other hand, this result may also indicate that the homozygosity of this frame shift mutation is too severe to survive, since none of the homozygous null mutations in *FKRP* has been reported to date and *FKRP* knockout mice also showed embryonic lethality [35].

Interestingly, two different homozygous mutations, c.263A>T (p.Tyr88Phe) and c.560C>G (p.Ala187Gly), were found in Patient 1, but not in 100 controls. Her parents were consanguineous (cousins) and both harbored these two mutations heterozygously. Compared the amino acid sequences of the FKRP protein among different species, p.Tyr88 is highly conserved in mammals while p.Ala187 is preserved among primates and some mammals, but not in rodents. Furthermore, predictions of functional effects of these two variants using software showed that p.Tyr88Phe change is probably damaging but p.Ala187Gly is benign in terms of functional impact (<http://genetics.bwh.harvard.edu/pph2/index.shtml>). Accordingly, c.263A>T (p.Tyr88Phe) is more likely to be pathogenic in Patient 1 although further functional studies are still necessary.

In our cohort, cardiomyopathy accounted for 83% of our patients, whereas about 10–55% of European LGMD2I patients were reported to have cardiac problems [36]. As for respiratory function, only one of our patients (Patient 6) was ventilator-dependent at night although the other five developed variable degrees of respiratory impairment. However, the proportion of respiratory aid requirement was slightly lower than other reports [20, 36–38], probably because the assessment age and disease duration of our patients were also lower. Similar to previously reported LGMD2I patients, none of our patients had overt mental retardation.

So far few papers have focused specifically on the muscle imaging of LGMD2I patients [37,39]. Based on previous related literature, gluteal muscles and posterior compartment of thigh muscles were more affected than anterior compartment in LGMD2I. In our report, similar muscle involvement was seen on CT images in which gluteal maximus was the most severely affected, followed by adductors and biceps femoris. Some of these changes may overlap with those seen in other common LGMD, especially LGMD2A [39], such as the early involvement of gluteal muscles and predominant involvement of posterior compartment. However, selective involvement of medial gastrocnemius and soleus and relative sparing of vastus lateralis are characteristic for LGMD2A [12,21,40], which suggests that muscle images are still

helpful for a differential diagnosis. In addition, different clinical phenotypes including commonly-seen calf hypertrophy and cardiac involvement in LGMD2I and the presence of characteristic lobulated fibers on muscle pathology of LGMD2A are also important to make the differentiation. In our series, all patients showed calf hypertrophy and 83% had cardiac problems; lobulated fibers were not observed in skeletal muscle from any patient and molecular analysis of *CAPN3* revealed no mutation.

LGMD2I is one of the most prevalent LGMD in Europe but is very rare in Asia. Only one from Taiwan (P6), two from China and another Asian patient from North America have been reported on thus far [26–28]. Also in Japan, only one LGMD2I patient was identified by the National Center of Neurology and Psychiatry, which has the largest muscle repository in Japan. Therefore, our report discloses that LGMD2I is not rare at least in Taiwan. Considering that the glycosylation defect may be too mild to be detected by immunohistochemical screening, there must be more LGMD2I patients who are as yet undiagnosed in Taiwan. Larger scale mutation analysis for uncategorized LGMD patients may be necessary for an early diagnosis of LGMD2I to be made. One common mutation, c.948delC, in the Taiwanese population may be associated with higher frequency and early development of cardiomyopathy although a larger number of patients is required to make this conclusive. However, it is still suggested that clinicians should closely monitor the cardiac function of LGMD2I patients harboring this mutation from late childhood or their early teens.

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#### References

- [1] Muntoni F, Brockington M, Blake DJ, Torelli S, Brown SC. Defective glycosylation in muscular dystrophy. *Lancet* 2002;360:1419–21.
- [2] Toda T, Kobayashi K, Takeda S, et al. Fukuyama-type congenital muscular dystrophy (FCMD) and alpha-dystroglycanopathy. *Congenit Anom (Kyoto)* 2003;43:97–104.
- [3] Muntoni F, Torelli S, Brockington M. Muscular dystrophies due to glycosylation defects. *Neurotherapeutics* 2008;5:627–32.

- [4] Mercuri E, Messina S, Bruno C, et al. Congenital muscular dystrophies with defective glycosylation of dystroglycan: a population study. *Neurology* 2009;72:1802–9.
- [5] Hara Y, Balci-Hayta B, Yoshida-Moriguchi T, et al. A dystroglycan mutation associated with limb-girdle muscular dystrophy. *N Engl J Med* 2011;364:939–46.
- [6] Beltran-Valero de Bernabe D, Currier S, Steinbrecher A, et al. Mutations in the *O*-mannosyltransferase gene *POMT1* give rise to the severe neuronal migration disorder Walker–Warburg syndrome. *Am J Hum Genet* 2002;71:1033–43.
- [7] van Reeuwijk J, Janssen M, van den Elzen C, et al. *POMT2* mutations cause alpha-dystroglycan hypoglycosylation and Walker–Warburg syndrome. *J Med Genet* 2005;42:907–12.
- [8] Yoshida A, Kobayashi K, Manya H, et al. Muscular dystrophy and neuronal migration disorder caused by mutations in a glycosyltransferase, *POMGnT1*. *Dev Cell* 2001;1:717–24.
- [9] Kobayashi K, Nakahori Y, Miyake M, et al. An ancient retrotransposal insertion causes Fukuyama-type congenital muscular dystrophy. *Nature* 1998;394:388–92.
- [10] Brockington M, Blake DJ, Prandini P, et al. Mutations in the fukutin-related protein gene (*FKRP*) cause a form of congenital muscular dystrophy with secondary laminin alpha2 deficiency and abnormal glycosylation of alpha-dystroglycan. *Am J Hum Genet* 2001;69:1198–209.
- [11] Longman C, Brockington M, Torelli S, et al. Mutations in the human *LARGE* gene cause *MDC1D*, a novel form of congenital muscular dystrophy with severe mental retardation and abnormal glycosylation of alpha-dystroglycan. *Hum Mol Genet* 2003;12:2853–61.
- [12] Roscioli T, Kamsteeg EJ, Buysse K, et al. Mutations in *ISPD* cause Walker–Warburg syndrome and defective glycosylation of alpha-dystroglycan. *Nat Genet* 2012;44:581–5.
- [13] Willer T, Lee H, Lommel M, et al. *ISPD* loss-of-function mutations disrupt dystroglycan *O*-mannosylation and cause Walker–Warburg syndrome. *Nat Genet* 2012;44:575–80.
- [14] Vuillaumier-Barrot S, Bouchet-Séraphin C, Chelbi M, et al. Identification of mutations in *TMEM5* and *ISPD* as a cause of severe cobblestone lissencephaly. *Am J Hum Genet* 2012;91:1135–43.
- [15] Manzini MC, Tambunan DE, Hill RS, et al. Exome sequencing and functional validation in zebrafish identify *GTDC2* mutations as a cause of Walker–Warburg syndrome. *Am J Hum Genet* 2012;91:541–7.
- [16] Buysse K, Riemersma M, Powell G, et al. Missense mutations in  $\beta$ -1,3-N-acetylglucosaminyltransferase I (*B3GNT1*) cause Walker–Warburg syndrome. *Hum Mol Genet* 2013;22:1746–54.
- [17] Lefeber DJ, de Brouwer AP, Morava E, et al. Autosomal recessive dilated cardiomyopathy due to *DOLK* mutations results from abnormal dystroglycan *O*-mannosylation. *PLoS Genet* 2011;7:e1002427.
- [18] Barone R, Aiello C, Race V, et al. *DPM2-DCG*: a muscular dystrophy-dystroglycanopathy syndrome with severe epilepsy. *Ann Neurol* 2012;72:550–8.
- [19] Lefeber DJ, Schönberger J, Morava E, et al. Deficiency of Dol-P-Man synthase subunit *DPM3* bridges the congenital disorders of glycosylation with the dystroglycanopathies. *Am J Hum Genet* 2009;85:76–86.
- [20] Sveen ML, Schwartz M, Vissing J. High prevalence and phenotype-genotype correlations of limb girdle muscular dystrophy type 2I in Denmark. *Ann Neurol* 2006;59:808–15.
- [21] Watanabe M, Kobayashi K, Jin F, et al. Founder *SVA* retrotransposal insertion in Fukuyama-type congenital muscular dystrophy and its origin in Japanese and Northeast Asian populations. *Am J Med Genet A* 2005;138:344–8.
- [22] Murakami T, Hayashi YK, Noguchi S, et al. Fukutin gene mutations cause dilated cardiomyopathy with minimal muscle weakness. *Ann Neurol* 2006;60:597–602.
- [23] Brockington M, Yuva Y, Prandini P, et al. Mutations in the fukutin-related protein gene (*FKRP*) identify limb girdle muscular dystrophy 2I as a milder allelic variant of congenital muscular dystrophy *MDC1C*. *Hum Mol Genet* 2001;10:2851–9.
- [24] Lim BC, Ki CS, Kim JW, et al. Fukutin mutations in congenital muscular dystrophies with defective glycosylation of dystroglycan in Korea. *Neuromuscul Disord* 2010;20:524–30.
- [25] Frosk P, Greenberg CR, Tennese AA, et al. The most common mutation in *FKRP* causing limb girdle muscular dystrophy type 2I (*LGMD2I*) may have occurred only once and is present in Hutterites and other populations. *Hum Mutat* 2005;25:38–44.
- [26] Lin YC, Murakami T, Hayashi YK, et al. A novel *FKRP* gene mutation in a Taiwanese patient with limb-girdle muscular dystrophy 2I. *Brain Dev* 2007;29:234–8.
- [27] Hong D, Zhang W, Wang W, Wang Z, Yuan Y. Asian patients with limb girdle muscular dystrophy 2I (*LGMD2I*). *J Clin Neurosci* 2011;18:494–9.
- [28] Kang PB, Feener CA, Estrella E, et al. *LGMD2I* in a North American population. *BMC Musculoskelet Disord* 2007;8:115.
- [29] Yis U, Uyanik G, Heck PB, et al. Fukutin mutations in non-Japanese patients with congenital muscular dystrophy: less severe mutations predominate in patients with a non-Walker–Warburg phenotype. *Neuromuscul Disord* 2011;21:20–30.
- [30] Topaloglu H, Brockington M, Yuva Y, et al. *FKRP* gene mutations cause congenital muscular dystrophy, mental retardation, and cerebellar cysts. *Neurology* 2003;60:988–92.
- [31] Matsumoto H, Hayashi YK, Kim DS, et al. Congenital muscular dystrophy with glycosylation defects of alpha-dystroglycan in Japan. *Neuromuscul Disord* 2005;15:342–8.
- [32] de Paula F, Vieira N, Starling A, et al. Asymptomatic carriers for homozygous novel mutations in the *FKRP* gene: the other end of the spectrum. *Eur J Hum Genet* 2003;11:923–30.
- [33] Poppe M, Cree L, Bourke J, et al. The phenotype of limb-girdle muscular dystrophy type 2I. *Neurology* 2003;60:1246–51.
- [34] Mercuri E, Brockington M, Straub V, et al. Phenotypic spectrum associated with mutations in the fukutin-related protein gene. *Ann Neurol* 2003;53:537–42.
- [35] Chan YM, Keramaris-Vrantsis E, Lidov HG, et al. Fukutin-related protein is essential for mouse muscle, brain and eye development and mutation recapitulates the wide clinical spectrums of dystroglycanopathies. *Hum Mol Genet* 2010;19:3995–4006.
- [36] Poppe M, Bourke J, Eagle M, et al. Cardiac and respiratory failure in limb-girdle muscular dystrophy 2I. *Ann Neurol* 2004;56:738–41.
- [37] Bourteel H, Vermersch P, Cuisset JM, et al. Clinical and mutational spectrum of limb-girdle muscular dystrophy type 2I in 11 French patients. *J Neurol Neurosurg Psychiatry* 2009;80:1405–8.
- [38] Stensland E, Lindal S, Jonsrud C, et al. Prevalence, mutation spectrum and phenotypic variability in Norwegian patients with limb girdle muscular dystrophy 2I. *Neuromuscul Disord* 2011;21:41–6.
- [39] Fischer D, Walter MC, Kesper K, et al. Diagnostic value of muscle **MRI** in differentiating *LGMD2I* from other *LGMDs*. *J Neurol* 2005;252:538–47.
- [40] Wattjes MP, Kley RA, Fischer D. Neuromuscular imaging in inherited muscle diseases. *Eur Radiol* 2010;20:2447–60.



## RESEARCH PAPER

# Rapidly progressive scoliosis and respiratory deterioration in Ullrich congenital muscular dystrophy

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## ABSTRACT

**Objective** To characterise the natural history of Ullrich congenital muscular dystrophy (UCMD).

**Patients and methods** Questionnaire-based nationwide survey to all 5442 certified paediatric and adult neurologists in Japan was conducted from October 2010 to February 2011. We enrolled the 33 patients (age at assessment, 11±6.6 years) who were reported to have collagen VI deficiency on immunohistochemistry in muscle biopsies. We analysed the development, clinical manifestations, Cobb angle and %vital capacity (%VC) in spirogram.

**Results** Cobb angle over 30° was noted at age 9.9±5.3 years (n=17). The maximum progression rate was 16.2±10°/year (n=13). %VC was decreased exponentially with age, resulting in severe respiratory dysfunction before pubescence. Scoliosis surgery was performed in 3 patients at ages 5 years, 9 years and 10 years. Postoperative %VC was relatively well maintained in the youngest patient. Non-invasive ventilation was initiated at age 11.2±3.6 years (n=13). Twenty-five (81%) of 31 patients walked independently by age 1.7±0.5 years but lost this ability by age 8.8±2.9 years (n=11). Six patients never walked independently.

**Conclusions** The natural history of scoliosis, respiratory function and walking ability in UCMD patients were characterised. Although the age of onset varied, scoliosis, as well as restrictive respiratory dysfunction, progressed rapidly within years, once they appeared.

## INTRODUCTION

Ullrich congenital muscular dystrophy (UCMD) is, after Fukuyama CMD, the second most common CMD in Japan.<sup>1</sup> UCMD is characterised by proximal joint contractures, distal joint hyperlaxity, proximal muscle weakness, scoliosis and respiratory failure.<sup>1-4</sup> The prevalence of UCMD is reported to be 1.3 per million in northern England.<sup>5</sup> Mutations in either *COL6A1*, *COL6A2* or *COL6A3* gene, each encoding a subunit of collagen VI (COL6), are known to cause UCMD. We have previously shown that there are two modes of COL6 deficiency: complete COL6 deficiency (CD) and sarcolemma specific COL6 deficiency (SSCD),<sup>6,7</sup> which are associated with recessive and de novo dominant mutations in *COL6* genes, respectively.<sup>1</sup>

To date, there is no cure for UCMD, and patients rely on supportive treatment of symptoms such as spinal deformity and respiratory failure. However,

pathological hypotheses leading to myofibre degeneration in COL6-deficient skeletal muscle have been proposed and therapeutic targets have been suggested.<sup>8</sup> There is currently a clinical trial for UCMD patients based upon the theory of impaired autophagy.<sup>9,10</sup> Furthermore, a gene-based therapy to inhibit mutant transcripts by antisense has also been proposed because an abnormal mutated subunit can be assembled into growing supra-molecular structures and sequester normal subunits into non-functional complexes, thus exerting dominant-negative effect.<sup>11,12</sup> Advances in such therapeutic research make knowledge of the natural course of the disease and appropriate outcome measures necessary. However, only limited information is available, especially regarding the rate of disease progression.<sup>13</sup> We have therefore attempted to determine the natural history of UCMD.

## PATIENTS AND METHODS

This clinical study was performed in conformity with the Declaration of Helsinki for investigation involving human subjects and was approved by the ethics committee of the National Center of Neurology and Psychiatry.

A questionnaire-based nationwide survey was conducted from October 2010 to February 2011. The questionnaire was mailed to all 5442 certified paediatric and adult neurologists by the Japanese Society of Child Neurology and the Japanese Society of Neurology, and we received 1881 (34.6%) responses. This survey consisted of questions about perinatal and developmental aspects, age and clinical manifestations at diagnosis, age at assessment, age at loss of ambulation, age at scoliosis surgery, age at initiation of non-invasive ventilation (NIV), data of Cobb angle on x-ray and % vital capacity (VC) in spirogram, in addition to pathological findings and COL6 immunohistochemistry in biopsied muscle (table 1).

Among 40 patients reported to have UCMD, we enrolled 33 patients (15 men and 18 women) with COL6 deficiency: 5 with CD and 28 with SSCD, on immunohistochemistry in skeletal muscle. Sequence analysis of *COL6* genes was performed using genomic DNA from 32 patients, 19 (59.4%) of whom carried identifiable mutations (table 2). All the 14 patients, who were not genetically confirmed, manifested clinical features compatible with UCMD in addition to COL6 deficiency (tables 2 and 3).<sup>14</sup> Age at muscle biopsy was 3.2±2 years

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Table 1 Summary of questionnaire items in this study

Items	Items	Items	Items
Patient's visit	Yes/No	Age at muscle biopsy	Age
Age at assessment	Age	Clinical manifestation at diagnosis	
Sex	Male/Female	Distal joint hyperlaxity	Yes/No
Muscle biopsy	Yes/No	Protruding calcaneus	Yes/No
COL6 deficiency on IHC	Yes/No	High arched palate	Yes/No
Mode of COL6 deficiency	CD/SSCD	Proximal joint contractures	Yes/No
Perinatal history		Scoliosis	Yes/No
Decreased fetal movement	Yes/No	Spinal rigidity	Yes/No
Poor sucking	Yes/No	Ankle joint contracture	Yes/No
Floppiness	Yes/No	Facial weakness	Yes/No
CHD	Yes/No	Skin lesions	Yes/No
Torticollis	Yes/No		
AMC	Yes/No	Age at loss of ambulation	Age
Developmental history		NIV	Yes/No
Head control	Age	Age at initiation of NIV	Age
Sitting	Age	Data of %VC	
Walking	Age	Age at scoliosis surgery	Age
Phrases	Age	Data of Cobb angle	

AMC, arthrogryposis multiplex congenita; CD, complete collagen VI deficiency; CHD, congenital hip dislocation; COL6, collagen VI; IHC, immunohistochemistry; NIV, non-invasive ventilation; SSCD, sarcolemma-specific collagen VI deficiency; VC, vital capacity.

(mean±SD). Age at assessment was 11±6.6 years. We analysed the information on perinatal abnormalities, development, clinical features, deterioration of ability to walk, progression of scoliosis and respiratory dysfunction.

## RESULTS

Perinatal abnormalities and clinical manifestations are shown in tables 2 and 3. Congenital hip dislocation, torticollis and arthrogryposis multiplex were noted in 36.4%, 27.3% and 20% of patients, respectively. More than 50% of patients had distal joint hyperlaxity, protruding calcaneus, high arched palate, proximal joint contractures and scoliosis. Creatine kinase level at muscle biopsy was 315±110 IU/L (n=31). Among patients with CD, homozygous or compound heterozygous mutations were identified in three but a heterozygous mutation was identified in one patient. All the 15 patients with SSCD carried a heterozygous mutation (table 2).

Twenty-five (81%) of 31 patients were able to sit and walk independently. Six patients (19%) never walked, three of whom had CD by muscle immunohistochemistry (tables 2 and 3). Head control, sitting and independent ambulation were completed at median ages of 4 months, 9 months and 18 months, respectively (figure 1A). In contrast, achievement of speaking phrases was not delayed, ranging from ages 12 months to 25 months (figure 1A). Most patients became able to walk independently by age 2 years but this ability deteriorated with age (figure 1B). Loss of ambulation occurred at age 8.8±2.9 years (n=11). Patient 18 was reported to walk with knee-ankle-foot orthoses at age 11 years. Patients 20 and 30, respectively, required a wheelchair at ages 13 years and 6 years. Half of the patients became wheelchair-bound by age 11 years (figure 1B). Six patients became wheelchair-bound by age 7 years, two of whom had CD and loss of ambulation at ages 5.5 years and 6 years, respectively. Two of four patients with SSCD who carried a heterozygous c.850G>A (p.Gly284Arg) mutation in *COL6A1* did not acquire independent ambulation (tables 2 and 3).

The severity and progression of scoliosis were assessed by Cobb angle (n=23). Maximum Cobb angles are shown in table 3. Patient 28 was reported to suffer from marked scoliosis albeit no data of Cobb angle was available. Cobb angle over 30° was noted at age 9.9±5.3 years in 17 patients (table 4). Among them 13 patients had Cobb angle data available for 3 or more years and show a maximum progression rate of 16.2±10°/year. Overall, although the onset of scoliosis varied, it progressed rapidly within years once scoliosis was noted (figure 1C). Surgical intervention for scoliosis was performed in Patients 8, 9 and 21 at ages 10 years, 9 years and 5 years, respectively. Presurgical and postsurgical Cobb angle data were available in the last two patients, both of whom showed improvement from 107° to 80° and 90° to 40°, respectively.

Percent VCs at survey are shown in table 3. Patients 8 and 27 had 32% and 70.4% of predicted VC at ages 8 years and 6 years, respectively. Respiratory function, measured by %VC (n=20), decreased exponentially with age accompanied by a sharp decline below age 10 years (figure 1D). Importantly, post-operative VC was relatively well maintained in the patient who underwent surgery at age 5 years (figure 1D). The percentage of patients requiring NIV increased with age (figure 1E). Half of the patients required NIV by age 12 years. Age at initiation of NIV was 11.2±3.6 years (n=13) (table 3). On the other hand, %VC at mean age of NIV initiation was estimated at around 36%.

## DISCUSSION

This is the first nationwide survey of the natural history of UCMD in Japan. This study confirmed the previously reported clinical features of UCMD: delayed motor milestone, absence of mental retardation, distal joint hyperlaxity, proximal joint contractures, scoliosis and respiratory involvement.<sup>1-4</sup> Furthermore, we characterised the natural history of scoliosis, respiratory function and ambulation in this relatively large UCMD series.

UCMD is on a disease spectrum of COL6 related myopathy. Intermediate phenotypes, named mild UCMD or severe Bethlehem myopathy, have been known, and currently there is no

Table 2 Clinical, pathological and genetical findings in the 33 patients

Pt	Age at biopsy (years)	Clinical manifestations and CK level at muscle biopsy										Collagen VI	
		Distal joint hyperlaxity	Protruding calcaneus	High arched palate	Proximal joint contracture	Scoliosis	Spinal rigidity	Ankle joint contracture	Facial weakness	Skin lesion	CK	Deficiency on IHC	Gene mutation
1	3	(+)	ND	(+)	(-)	ND	ND	(-)	(+)	(-)	440	CD	COL6A2 c.1771-3G>C COL6A2 c.1270-1G>C
2	2	(+)	(+)	(-)	(+)	(+)	(-)	(-)	(-)	(+)	413	CD	COL6A3 c.5692delG p.Val1898fs COL6A3 c.8737delG p.Ala2913fs
3	2	(+)	(+)	(-)	(-)	(-)	(-)	(-)	(-)	(+)	474	CD	NF
4	1	ND	(+)	(-)	(-)	(-)	(-)	(-)	(+)	(-)	339	CD	COL6A2 c.2678_2700del23 p.Pro893fs homozygous
5	1	ND	(+)	(-)	(+)	ND	(+)	(+)	(+)	(-)	ND	CD	COL6A3 c.4184G>A p.Arg1395Gln
6	2	(+)	(+)	(+)	(-)	ND	ND	(-)	(-)	(-)	290	SSCD	COL6A1 c.850G>A p.Gly284Arg
7	1	(+)	ND	(+)	(+)	(+)	(+)	(+)	(-)	ND	195	SSCD	COL6A1 c.850G>A p.Gly284Arg
8	1	(+)	(+)	ND	(+)	ND	(+)	(+)	ND	(-)	319	SSCD	NF
9	3	(+)	(-)	ND	(+)	(+)	ND	(-)	ND	(-)	ND	SSCD	NF
10	3	(+)	(+)	(+)	(+)	(+)	(-)	(-)	(-)	(+)	428	SSCD	COL6A2 c.950_954+8del
11	5	(+)	(+)	(+)	(+)	(+)	(+)	(-)	(-)	(-)	151	SSCD	NA
12	4	(+)	(+)	(-)	(+)	(+)	ND	(-)	(-)	ND	207	SSCD	COL6A2 c.901G>T p.Gly301Cys
13	6	ND	(-)	(+)	(-)	(-)	(-)	(+)	(-)	(-)	564	SSCD	NF
14	7	(-)	(+)	(+)	(-)	(-)	(-)	(-)	(-)	(-)	329	SSCD	COL6A2 c.812G>A p.Gly271Asp
15	5	(+)	(-)	(+)	(-)	(+)	(-)	(-)	(+)	(-)	289	SSCD	COL6A1 c.958_966del9 p.Gly320_Lys322del
16	3	(-)	ND	(+)	(-)	(-)	(-)	(-)	(-)	(-)	425	SSCD	COL6A3 c.6210+2T>A
17	3	ND	ND	(+)	ND	ND	ND	ND	(-)	ND	242	SSCD	COL6A1 c.850G>A p.Gly284Arg
18	2	(+)	(+)	(+)	(+)	(-)	(+)	(+)	(+)	(-)	299	SSCD	NF
19	3	(-)	(+)	(+)	(+)	(-)	(-)	(+)	(-)	(-)	464	SSCD	NF
20	1	ND	ND	(+)	(-)	(-)	(-)	(-)	(-)	ND	254	SSCD	COL6A1 c.958_966del9 p.Gly320_Lys322del
21	5	(+)	ND	(-)	(-)	(+)	(+)	(-)	(-)	(-)	138	SSCD	COL6A1 c.868G>A p.Gly290Arg
22	6	(+)	ND	(-)	(+)	(-)	(-)	(+)	(-)	(-)	495	SSCD	NF
23	1	(+)	(+)	(+)	(+)	(-)	(-)	(-)	(-)	(+)	300	SSCD	COL6A1 c.850G>A p.Gly284Arg
24	2	(+)	(+)	(+)	(+)	(-)	(-)	(-)	(+)	(-)	388	SSCD	NF
25	4	(+)	(+)	(-)	(-)	(+)	(+)	(-)	(-)	(-)	277	SSCD	COL6A1 c.860G>A p.Gly287Glu
26	6	(+)	(+)	(+)	(-)	(+)	(+)	(+)	(-)	(-)	103	SSCD	NF
27	5	ND	ND	ND	ND	ND	ND	ND	ND	ND	262	SSCD	COL6A1 c.1056+1G>A
28	8	ND	(-)	(+)	(+)	ND	(+)	(-)	(+)	(+)	254	SSCD	NF
29	2	ND	ND	(-)	(-)	(+)	(-)	(-)	(-)	(-)	361	SSCD	COL6A3 c.6210+1G>A
30	3	(-)	(+)	ND	(-)	(-)	(-)	(-)	ND	(-)	278	SSCD	COL6A3 c.6310-2A>G
31	1	(+)	ND	(-)	(+)	(+)	(-)	(-)	(-)	(-)	261	SSCD	NF
32	4	(+)	(+)	(+)	(+)	(+)	(+)	(+)	(-)	(-)	323	SSCD	NF
33	0	(+)	(-)	(+)	(+)	(+)	(+)	(-)	(-)	(-)	214	SSCD	NF
		21/25 (84%)	18/23 (78.3%)	19/29 (65.5%)	17/31 (54.3%)	14/26 (53.8%)	11/27 (40.7%)	9/31 (29%)	7/29 (24.1%)	5/28 (17.9%)			

CD, complete deficiency; CK, creatine kinase; IHC, immunohistochemistry; NA, not available; ND, no data; NF, not found; Pt, patient; SSCD, sarcolemma specific collagen VI deficiency; (+), present; (-), absent.

Table 3 Perinatal, developmental history and disease progression in our series

Pt	Age at survey (years)	Perinatal history					Developmental milestone (months)					Motor function at survey		Respiratory function		Maximum Cobb angle (°) (yrs)	
		Floppiness	Poor sucking	CHD	Decreased fetal movement	Torticollis	AMC	Head control	Sit	Walk	Phrases	Walk independently	Age at loss of ambulation (yrs)	%VC at survey (%)	NIV		Age at initiation of NIV (yrs)
1	14	(+)	(+)	(+)	(+)	(+)	(+)	6	30	(-)	ND	(-)		29	(+)	8	45 (11)
2	10	(-)	(+)	(+)	(-)	(-)	ND	5	12	(-)	ND	(-)		22.3	(+)	8	12 (10)
3	7	(+)	(-)	(+)	(-)	(+)	(-)	ND	9	24	24	(-)	5.5	55	(-)		ND
4	3	(-)	(+)	(-)	(-)	(+)	(-)	7	12	(-)	25	(-)		ND	(-)		0 (3)
5	23	(+)	(+)	(+)	(-)	(-)	(-)	ND	ND	18	ND	(-)	6	14.9	(+)	15	50 (23)
6	23	(+)	(+)	(+)	(+)	(+)	(-)	5	18	(-)	18	(-)		10.4	(+)	15	54 (23)
7	14	(+)	(+)	(+)	(-)	(+)	(-)	3	11	(-)	23	(-)		ND	(+)	5	99 (9)
8	21	(-)	(-)	(-)	(-)	(-)	ND	5	12	(-)	ND	(-)		ND	(+)	9	53 (10)
9	20	ND	ND	(-)	ND	(+)	(-)	4	7	18	16	(-)	6	12.2	(+)	10	107 (9)
10	6	(-)	(-)	(+)	(-)	(-)	(-)	3	12	24	18	(-)	6	ND	(-)		20 (6)
11	10	(-)	(-)	(-)	(+)	(-)	(-)	3	7	18	19	(-)	7	46	(-)		10 (10)
12	15	(+)	(-)	(-)	(-)	(-)	(-)	4	8	22	24	(-)	10	37.9	(-)		31 (3)
13	15	(-)	(-)	(+)	(-)	(-)	ND	5	6	18	18	(-)	10	27.6	(+)	12	60 (15)
14	15	(+)	(-)	(+)	(+)	(-)	(-)	5	12	16	12	(-)	11	15.7	(+)	9.5	110 (15)
15	13	(+)	(+)	(-)	(+)	(-)	(-)	4	7	14	24	(-)	11	21.9	(+)	11	94 (14)
16	11	(-)	(+)	(-)	(-)	(-)	(+)	4	6	16	16	(-)	11	30.7	(-)		70 (11)
17	23	(-)	(+)	(-)	(-)	(-)	(-)	4	8	16	24	(-)	14	19.8	(+)	16	41 (23)
18	10	(+)	(+)	(-)	(+)	(-)	(-)	7	9	27	24	(+)		45	(-)		0(9)
19	3	(-)	(-)	(-)	(-)	(-)	(-)	8	11	24	24	(+)		ND	(-)		ND
20	16	(+)	(-)	(-)	(-)	(-)	(+)	4	8	20	12	(+)		ND	(+)	10	90(10)
21	12	(-)	(-)	(+)	(-)	(-)	(-)	6	15	30	14	(+)		44.3	(-)		90(5)
22	10	(-)	(-)	(-)	(+)	(-)	(-)	4	8	14	24	(+)		58.9	(-)		0(10)
23	5	(+)	(-)	(-)	(-)	(-)	(-)	2	7	14	21	(+)		ND	(-)		2(4)
24	5	(-)	(-)	(+)	ND	(-)	(-)	3	8	22	18	(+)		ND	(-)		5(5)
25	5	(-)	(+)	(-)	(-)	(+)	ND	4	6	15	ND	(+)		ND	(-)		60(6)
26	7	(+)	(+)	(-)	(+)	(-)	(+)	3	12	24	18	(+)		53.4	(-)		46(7)
27	11	(-)	(-)	(-)	ND	(-)	ND	4	8	16	ND	ND		ND	(-)		ND
28	18	(+)	(+)	(-)	(+)	(-)	ND	ND	20	36	24	(+)		ND	(+)	17	ND
29	2	(+)	(-)	(-)	ND	(+)	(-)	4	8	19	ND	(+)		ND	ND		ND
30	7	(-)	(-)	(-)	(-)	(-)	ND	3	8	13	24	(+)		ND	(-)		ND
31	1	(-)	(-)	(-)	(+)	(-)	(+)	6	14	ND	ND	ND		ND	ND		ND
32	6	(-)	(-)	(-)	(-)	(-)	ND	5	9	18	22	(+)		ND	(-)		40(4)
33	1	(+)	(-)	(+)	(-)	(+)	(-)	7	(-)	ND	ND	ND		ND	(-)		5(1)
		15/32 (46.9%)	13/32 (40.6%)	12/33 (36.4%)	10/29 (34.5%)	9/33 (27.3%)	5/25 (20%)										

Pt 18 was reported to walk with knee-ankle-foot orthoses at age 11 years. Pts 20 and 30 were reported to be able to walk independently but they respectively required a wheelchair at ages 13 years and 6 years. Pts 8 and 27 showed 32% and 70.4% of predicted VC at ages 8 years and 6 years, respectively. Pt 28 was reported to suffer from marked scoliosis but no data of Cobb angle was available.

AMC, arthrogryposis multiplex congenita; CHD, congenital hip dislocation; ND, no data; NIV, noninvasive ventilation; Pt, patient; %VC, % vital capacity; (+), present; (-), absent.

## Neuromuscular

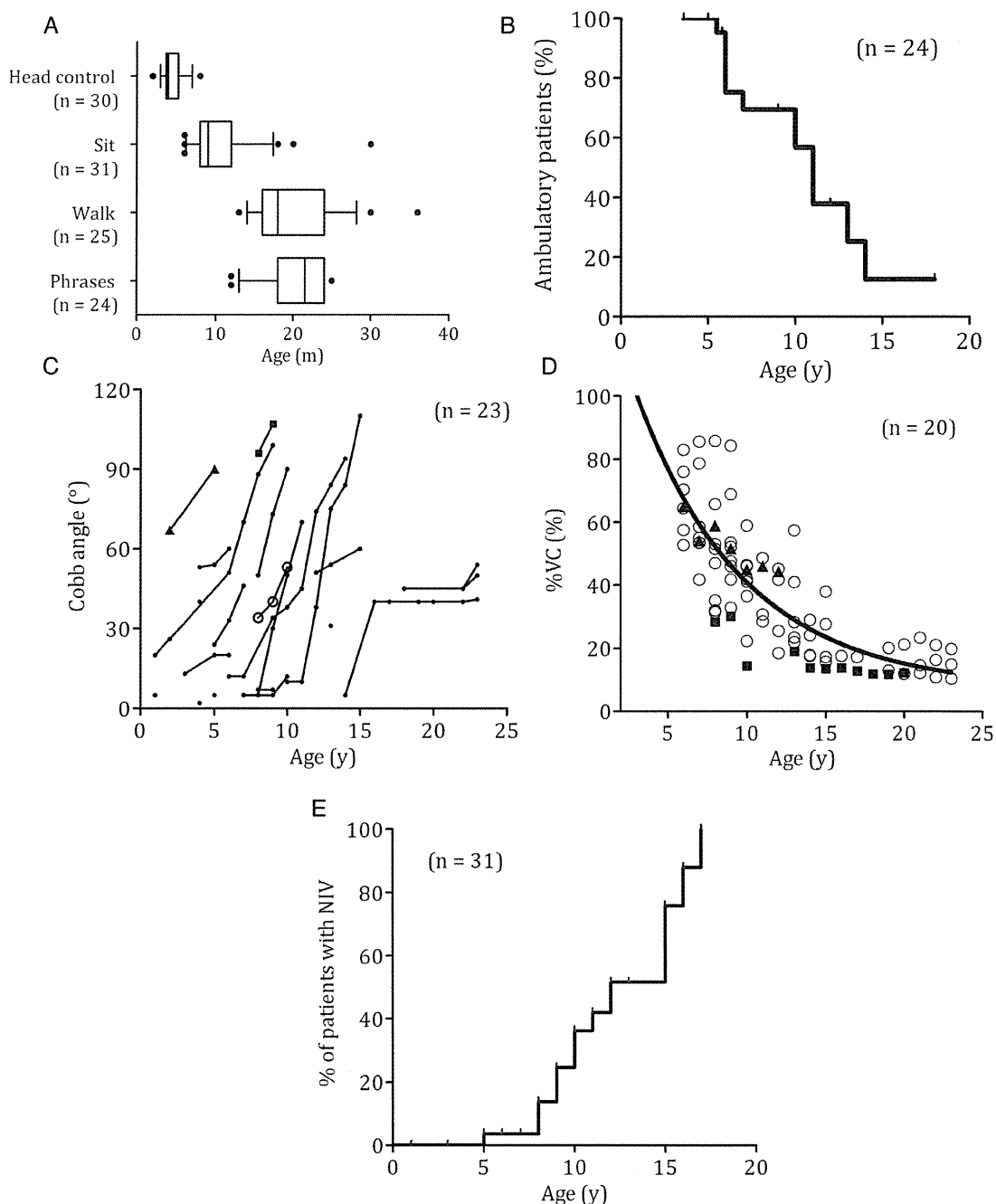


Figure 1 (A) Age ranges at completion of neck control, sit, independent ambulation and phrases. The boxes represent the range from the 25–75th percentile, while the bars span the 10–90th percentile. (B) Kaplan-Meier curve showing deterioration of walking ability in Ullrich congenital muscular dystrophy (n=24). Patients 20 and 30, respectively, become wheelchair-bound at ages 13 years and 6 years. (C) Severity and progression of scoliosis (n=23). Open circles, solid squares and triangles indicate preoperative Cobb angles from Patients 8, 9 and 21 who underwent scoliosis surgery at ages 10 years, 9 years and 5 years, respectively. (D) %Vital capacity (%VC) (n=20). Solid line represents the regression curve ( $\%VC = 144.8 * \exp(-0.146 * \text{Age}) + 7.386$ ,  $R^2 = 0.6684$ ). Solid squares and triangles respectively represent values from Patients 9 and 21 who underwent scoliosis surgery at ages 9 years and 5 years. (E) Kaplan-Meier curve showing the percentage of patients with non-invasive ventilation (NIV) (n=31).

clear-cut definition of two major phenotypes.<sup>8 15</sup> According to the clinical classification of early onset COL6-related myopathies, all the patients in our series can be classified into the most severe (early-severe) or moderate-progressive groups.<sup>16 17</sup> The age at loss of ambulation was slightly younger compared with the previous observations ( $10.7 \pm 4.8$  years and  $10.1 \pm 4.4$  years).<sup>13 17</sup> Interestingly, patients with CD never walked independently or became unable to walk by age 6 years,

indicating that CD is most likely to be associated with the more severe phenotype than SSCD. On the other hand, 3 (10.7%) of 28 patients with SSCD did not acquire independent ambulation. Unlike patients with CD, a great heterogeneity in the maximal motor capacity was observed in those with SSCD, ranging from no acquisition of walking ability to retaining ambulation throughout childhood. Four patients with a heterozygous c.850G>A (p.Gly284Arg) mutation in *COL6A1* showed a wide

Table 4 Data of Cobb angle in 23 patients

Pt	COL6 deficiency on IHC	Age at loss of ambulation (years)	Age at assessment (years)																			
			<3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	22	23
1	CD	NW																				45
2	CD	NW						5	5	5	12											
5	CD	6																			45	50
6	SSCD	NW																	45		45	54
7	SSCD	NW	26 (2)			51	70	88	99													
8	SSCD	NW						34	40	50												
9	SSCD	6						96	107					80								
10	SSCD	6	13 (3)		20	20																
11	SSCD	7						7	7	10												
12	SSCD	10												31								
13	SSCD	10											51	54			60					
14	SSCD	11						0		10	10	38	75	84	110							
15	SSCD	11				12	12			34	38	45	74	84	94							
16	SSCD	11						5	30	50	70											
17	SSCD	14												5		40	40		40	40	40	41
20	SSCD	W						50	73	90												
21	SSCD	W	67 (2)		90	40																
23	SSCD	W		2																		
24	SSCD	W			5																	
25	SSCD	W			53	54	60															
26	SSCD	W			24	33	46															
32	SSCD	W		40																		
33	SSCD	NW	5 (1)																			

Pts 8, 9 and 21 respectively underwent scoliosis surgery at age 10, 9 and 5 years.

CD, complete deficiency; COL6, collagen VI; IHC, immunohistochemistry; NW, not walk; Pt, patient; SSCD, sarcolemma specific collagen VI deficiency; W, walk.

variety in their ability to walk (table 3). In this study we were not able to confirm recessive mutations and a heterozygous mutation in 2 with CD and 13 with SSCD, respectively. The mutation detection rate (59.4%) was comparable with those reported to be up to 60% in other groups,<sup>15</sup> and those patients without a putative mutation identified may carry deletions or duplications of one or more exons as well as intronic, regulatory mutations.

The onset of scoliosis preceded loss of ambulation in UCMD. This pattern of scoliosis progression was also pointed out by Nadeau *et al.*<sup>13</sup> Development of scoliosis in Duchenne muscular dystrophy, on the other hand, is strongly related to the loss of walking ability.<sup>18</sup> In Duchenne muscular dystrophy, typically, scoliosis is not evident in ambulatory patients and starts after patients become wheelchair dependent. In UCMD, in contrast, scoliosis developed even when patients were still ambulant and is characterised by marked progression from early stage. For the first time, we characterised scoliosis progression in this study. It is noteworthy that scoliosis progresses rapidly, within years, once it starts. The early-onset and rapidly-progressive scoliosis in UCMD may well accelerate physical disability, such as difficulty in sitting, standing and walking, and cause pain. More importantly, scoliosis may well compromise respiratory function by reducing chest wall compliance.

VC declined exponentially with age, with a sharp decrease by age 10 years. Nadeau *et al* showed that forced VC (%predicted) in UCMD declined by  $6.6 \pm 1.9\%$ /year from age 6 years to 10 years compared with by  $0.4 \pm 3\%$ /year from age 11 years to 15 years.<sup>13</sup> Although the parameters were different, both studies indicate that UCMD patients develop restrictive respiratory dysfunction rapidly in the first decade of life. This decay in VC

might be associated with proximal joint and vertebral contractures together with weakness of the diaphragm. Considering the slower decline of %VC in the youngest patient after surgical correction of scoliosis, earlier surgical intervention to correct spinal deformity may be beneficial for maintaining chest wall compliance, thus preventing progressive respiratory dysfunction. Takaso *et al* successfully performed scoliosis surgery in three patients with UCMD at ages 11 years, 13 years and 17 years, respectively (not enrolled in the present study).<sup>19</sup> However, in these patients, surgery did not prevent deterioration of respiratory function suggesting that at such older ages pulmonary and chest wall compliance might be too severely compromised for patients to benefit from scoliosis surgery, and earlier surgical intervention may be more beneficial. However, further studies are necessary to conclude the efficacy of early scoliosis surgery.

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**Contributors** TY: designed the study, performed literature search, analysed the data and wrote the manuscript. HK, MO and YKH: supervised all aspects of this study, including the study design, interpretation and manuscript preparation. IN, KS and MS gave valuable comments for the manuscript. IN was involved in analysing and interpreting all the data and also supervised the study design, execution and manuscript preparation.

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**Competing interests** None.

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## Neuromuscular

## REFERENCES

- 1 Okada M, Kawahara G, Noguchi S, *et al*. Primary collagen VI deficiency is the second most common congenital muscular dystrophy in Japan. *Neurology* 2007;69:1035–42.
- 2 Ullrich O. Kongenitale atonisch-sklerotische Muskeldystrophie ein weiterer Typus der hereditären degenerativen Erkrankungen des neuromuskulären systems. *Z Ges Neurol Psychiat* 1930;126:171–201.
- 3 Nonaka I, Une Y, Ishihara T, *et al*. A clinical and histological study of Ullrich's disease (congenital atonic-sclerotic muscular dystrophy). *Neuropediatrics* 1981;12:197–208.
- 4 Mercuri E, Yuva Y, Brown SC, *et al*. Collagen VI involvement in Ullrich syndrome: a clinical, genetic, and immunohistochemical study. *Neurology* 2002;58:1354–9.
- 5 Norwood FL, Harling C, Chinnery PF, *et al*. Prevalence of genetic muscle disease in Northern England: in depth analysis of a muscle clinic population. *Brain* 2009;132:3175–86.
- 6 Ishikawa H, Sugie K, Murayama K, *et al*. Ullrich disease: collagen VI deficiency: EM suggests a new basis for muscular weakness. *Neurology* 2002;59:920–3.
- 7 Ishikawa H, Sugie K, Murayama K, *et al*. Ullrich disease due to deficiency of collagen VI in the sarcolemma. *Neurology* 2004;62:620–3.
- 8 Allamand V, Briñas L, Richard P, *et al*. ColVI myopathies: where do we stand, where do we go? *Skelet Muscle* 2011;1:30–42.
- 9 Grumati P, Coletto L, Sabatelli P, *et al*. Autophagy is defective in collagen VI muscular dystrophies, and its reactivation rescues myofiber degeneration. *Nat Med* 2010;16:1313–20.
- 10 ClinicalTrials.gov Identifier: NCT01438788. Clinical Trials.gov Website. <http://www.clinicaltrials.gov/> (accessed 4 Dec 2012).
- 11 Baker NL, Morgelin M, Pace RA, *et al*. Molecular consequences of dominant Bethlem myopathy collagen VI mutations. *Ann Neurol* 2007;62:390–405.
- 12 Gualandi F, Manzati E, Sabatelli P, *et al*. Antisense-induced messenger depletion corrects a COL6A2 dominant mutation in Ullrich myopathy. *Hum Gene Ther* 2012;23:1313–18.
- 13 Nadeau A, Kinali M, Main M, *et al*. Natural history of Ullrich congenital muscular dystrophy. *Neurology* 2009;73:25–31.
- 14 Pepe G, Bertini E, Bonaldo P, *et al*. Bethlem myopathy (BETHLEM) and Ullrich scleroatonic muscular dystrophy: 100th ENMC International Workshop, 23–24 November 2001, Naarden, the Netherlands. *Neuromuscul Disord* 2002;12:984–93.
- 15 Allamand V, Merlini L, Bushby K. 166th ENMC International Workshop on Collagen type VI- related Myopathies, 22–24 May 2009, Naarden, the Netherlands. *Neuromuscul Disord* 2010;20:346–54.
- 16 Quijano-Roy S, Allamand V, Riahi N, *et al*. Predictive factors of severity and management of respiratory and orthopaedic complications in 16 Ullrich CMD patients [abstract]. *Neuromuscul Disord* 2007;17:844.
- 17 Briñas L, Richard P, Quijano-Roy S, *et al*. Early onset collagen VI myopathies: Genetic and clinical correlations. *Ann Neurol* 2010;68:511–20.
- 18 Mullender MG, Blom NA, De Kleuver M, *et al*. A Dutch guideline for the treatment of scoliosis in neuromuscular disorders. *Scoliosis* 2008;3:14–27.
- 19 Takaso M, Nakazawa T, Imura T, *et al*. Surgical correction of spinal deformity in patients with congenital muscular dystrophy. *J Orthop Sci* 2010;15:493–501.



## Rapidly progressive scoliosis and respiratory deterioration in Ullrich congenital muscular dystrophy

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Case report

# Congenital generalized lipodystrophy type 4 with muscular dystrophy: Clinical and pathological manifestations in early childhood

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## Abstract

A boy with congenital generalized lipodystrophy type 4 with muscular dystrophy presented in infancy with delay in motor milestones and a persistent elevation of CK. There was no associated mental retardation. He was followed up to 3 years and 11 months; he had a homozygous c.696\_697insC mutation in *polymerase I and transcript release factor* (PTRF). He started to walk at 2 years and 6 months although he did not have mental retardation. Insulin resistance appeared at 3 years and 11 months of age. PTRF immunostaining positivity was absent in the muscle but caveolin-3 was preserved in the sarcolemma at 16 months of age. Secondary deficiency of caveolins may be closely associated with disease progression.

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**Keywords:** PTRF; Generalized lipodystrophy; Muscular dystrophy; Insulin resistance; Muscle mounding

## 1. Introduction

Congenital generalized lipodystrophies are rare autosomal recessive disorders that are characterized by an almost total loss of subcutaneous adipose tissues from birth, insulin resistance, diabetes, hypertriglyceridemia, and hepatic steatosis [1,2]. Recently, we first described muscular dystrophy with generalized lipodystrophy caused by *polymerase I and transcript release factor* (PTRF) mutations [3], and this disease was categorized as congenital generalized lipodystrophy type 4 (CGL4) (OMIM #613327). Patients with PTRF deficiency can show various symptoms that include arrhythmia,

atlantoaxial instability, and pyloric stenosis in addition to manifestations of congenital generalized lipodystrophy and muscular dystrophy [3–6].

Only a limited number of patients with this condition have been reported. Therefore, the accumulation of detailed clinical information, especially in early childhood, is important to understand this disease and to facilitate early diagnosis. Herein, we describe the detailed clinical course of a 3-year-old Japanese boy with CGL4 with muscular dystrophy.

## 2. Case report

Our patient was a Japanese boy aged 3 years and 11 months from healthy non-consanguineous parents. He was born via normal delivery at 39 weeks and 4 days of gestational age. His body height, body weight, and head circumference were 47 cm (−1.0 SD), 3070 g (−0.3 SD), and 34 cm (0.3 SD), respectively. He gained body weight

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slowly and weighed 4.5 kg at 4 months of age, upon which he was diagnosed with hypothyroidism. An elevated serum creatine kinase (CK) level (812 IU/L; normal <200) was also noted. He received thyroid hormone therapy consisting of 10 µg/day levothyroxine sodium hydrate. The subcutaneous fat of his face began to decrease at 7 months of age. He was referred to our hospital at 11 months of age because of continuous elevation of serum CK levels. Although he had head control at 4 months of age, he showed delayed motor milestones. He could crawl at 14 months and sit at 16 months of age. He showed normal mental development and spoke several meaningful words at 14 months of age.

At 16 months of age, his body height and weight were 78.5 cm (−0.1 SD) and 9.6 kg (−0.5 SD), respectively. He had a saddle nose, prominent ears, curled hair, and mild macroglossia. Loss of subcutaneous fat was marked on his face and limbs, which exposed prominent blood vessels in his extremities. His abdomen was distended without evidence of hepatosplenomegaly or tumor. He did not have hypertrophic tonsils or scoliosis and facial muscle involvement or a high arched palate was not observed. He had mild hypertrophic muscles, especially in his extremities, and mild proximal muscle weakness was seen with normal deep tendon reflexes. He could crawl and stand with support. Although he demonstrated percussion-induced muscle mounding, he did not demonstrate the rippling phenomenon.

His serum CK level had increased to 2293 IU/L, but his serum immunoglobulin level was normal. Chest radiography, electrocardiography, cardiac echocardiography, and bone radiography showed normal findings. A muscle computed tomography (CT) showed decreased subcutaneous adipose tissue and hypertrophic muscles with normal intensity (Fig. 1A).

A muscle biopsy taken from his left biceps brachii showed dystrophic changes including variation in fiber size and scattered necrotic and regenerating fibers (Fig. 2A and B). Immunohistochemistry for PTRF was negative in both the sarcolemma of the muscle fibers and blood vessels (Fig. 2C). The caveolin-3 stain was slightly irregular but well preserved (Fig. 2D), whereas the immunoreactions of caveolins-1 and -2 were barely detectable in the blood vessels (data not shown). Antibodies for dystrophin and other major proteins

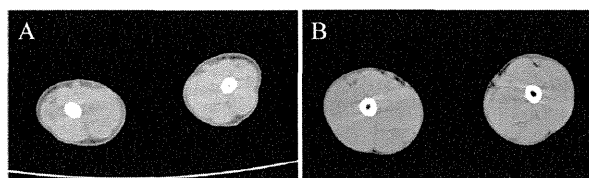


Fig. 1. Computed tomographic scan at the thigh level at 1 year (A) and 2 years and 10 months (B) of age. The thigh muscles showed hypertrophy without abnormal intensity. Note the progressive loss of subcutaneous adipose tissues.

associated with muscular dystrophies showed normal staining (data not shown). Genetic analysis revealed a homozygous mutation of c.696\_697insC in the *PTRF* gene, which is a common mutation in Japanese CGL4 patients [3]. This mutation resulted in substitution of the last 158 amino acids with an unrelated 191-amino acid sequence; moreover, the mutant protein was mislocalized, as shown by a previous *in vitro* experiment [3].

He started to walk without support at 2 years and 6 months of age and he could speak 2-word sentences. At the age of 2 years and 10 months, his body height and weight were 92.7 cm (+0.2 SD) and 13.2 kg (−0.1 SD), respectively (Table 1). He could run and climb stairs slowly, but he could not jump. The Gowers' sign was negative. The endocrinological examination results at that time were as follows: total cholesterol (T-cho), 213 (140–220) IU/L; triglyceride (TG), 309 (30–150) mg/dL; high-density lipopolysaccharide cholesterol (HDL-C), 27 (40–76) mg/dL; low-density lipopolysaccharide cholesterol (LDL-C), 125 (70–139) mg/dL; glucose, 98 mg/dL; HbA1c, 5.1 (3.8–5.5)%; insulin, 1.5 (5.0–20.0) µU/mL; fT4, 1.42 (0.97–1.80) ng/dL; fT3, 4.36 (2.73–4.50) pg/mL; and thyroid-stimulating hormone, 4.560 (0.300–3.000) µU/mL. The oral glucose tolerance test showed a normal reaction and his blood sugar level was 123 mg/mL 2 h after administration. The calculated insulin resistance index (HOMA-R) was 1.24 (<1.6). Serum adiponectin, total PAI-1, and leptin levels were 1.2 (>4.0) µg/mL, 23 (<50) ng/mL, and 1.9 (male: 0.9–13.0, female: 2.5–21.8) ng/mL, respectively.

At 3 years and 11 months of age, his body height and weight were 99.2 cm (−0.1 SD) and 15.3 kg (−0.1 SD), respectively. The endocrinological examination results at that time were as follows: T-cho, 154 IU/L; TG, 680 mg/dL; HDL-C, 20 mg/dL; LDL-C, 57 mg/dL; glucose, 98 mg/dL; HbA1c, 5.0%; and insulin, 12.3 µU/mL. His HOMA-R was increased to 2.98. Serum adiponectin, total PAI-1, and leptin levels were 1.1 µg/mL, 96 ng/mL, and 1.6 ng/mL, respectively. His body fat percentage was 12.2% according to dual energy X-ray absorptiometry (DEXA) measurement.

### 3. Discussion

We previously reported that the clinical features observed in patients with *PTRF* mutations were closely associated with a secondary deficiency of caveolin [3].

In various types of congenital generalized lipodystrophy, body fat loss and insulin resistance are usually noticed at birth. In CGL4, however, the loss of adipose tissue in the face is observed after several months of age and insulin resistance appears from early childhood [3–6]. The boy described herein showed decreased subcutaneous fat in his face at 7 months of age. Lipodystrophy was progressive, and loss of adipose tissue in the lower legs became more apparent with age. This finding was confirmed by CT images. Metabolic

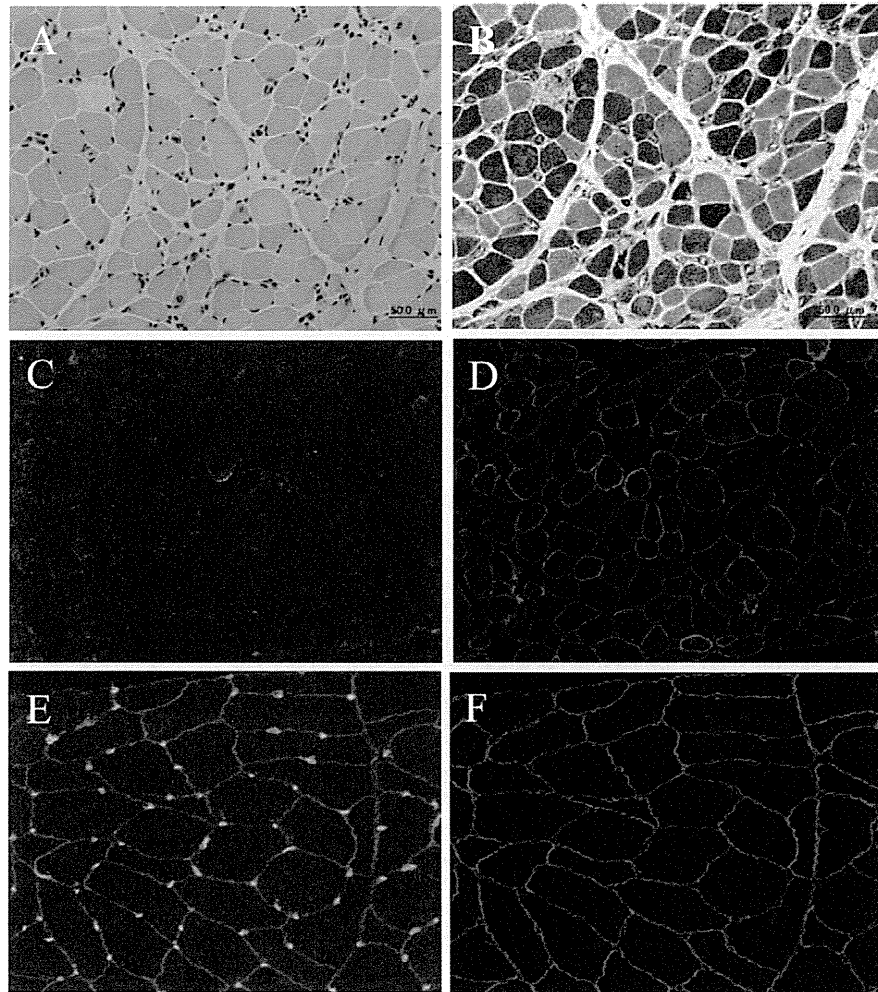


Fig. 2. The biceps brachii muscle showed moderate variation in fiber size, a few necrotic fibers, and increased endomysial and perimysial fibrosis (A, B). Negative immunoreactivity for PTRF (C) but almost normal caveolin-3 levels was observed (D). A: hematoxylin and eosin stain, B: NADH-tetrazolium reductase stain, C, E: PTRF immunohistochemistry, D, F: caveolin-3 immunohistochemistry. E, F: control muscle. Scale bar: 50  $\mu$ m.

abnormalities were also progressive. Although he did not have insulin resistance at 2 years and 10 months of age, he already showed high levels of T-cho and TG. He demonstrated insulin resistance at 3 years and 11 months.

PTRF is an essential component for the stabilization of caveolae. PTRF-deficient mice do not have detectable caveolae and show decreased insulin receptor levels in fat tissue [10]. Similarly, caveolin-1-deficient mice that show loss of caveolae have been reported to show insulin resistance due to decreased insulin receptor levels in adipose tissues [7–9]. This and a previous report [3] have shown that caveolin-1 and caveolin-2 levels greatly reduced in the intramuscular blood vessels from PTRF-deficient patients. Lipodystrophy and insulin resistance can be closely associated with secondary deficiency of caveolins.

This boy had delayed motor milestones associated with dystrophic changes in his muscles. Interestingly,

sarcolemmal caveolin-3 staining was well preserved in this patient compared to previously reported results in older patients, although his immunoreactivity for PTRF was defective. In the case of a 3-year-old Japanese girl who did not show muscle weakness accompanied by high serum levels of CK [5], caveolin-3 immunoreactivity was well preserved despite negative reactivity of PTRF. Secondary reduction of caveolin-3 may progress with age or disease progression [3]; and further studies are necessary to confirm this.

Percussion-induced muscle mounding is a characteristic finding in patients with PTRF deficiency as well as in some patients with caveolin-3 deficiency. Although the detailed mechanism involved in muscle mounding remains to be elucidated, it may be closely related to deficiencies of both caveolin-3 and PTRF.

CGL4 with muscular dystrophy is a progressive disorder, and cardiac problems, including arrhythmia,

Table 1  
Clinical and biological summary.

Age	Height (SD)	Weight (SD)	Clinical and biological signs	CK (normal < 200 IU/L)	T-Cho (140–220 IU/L)	TG (30–150 mg/dL)	Adiponectin (>4.0 µg/mL)	Leptin (0.9–13.0 ng/mL)
Birth	47.0 cm (−1.0)	3.1 kg (−0.3)	Normal amount of subcutaneous fat	ND	ND	ND	ND	ND
4 m	59.0 cm (−2.3)	4.5 kg (−2.9)	Head control (+), hypothyroidism	812	ND	ND	ND	ND
7 m	66.4 cm (−1.2)	5.8 kg (−2.7)	Decreased subcutaneous fat on his face	1779	ND	ND	ND	ND
14 m	76.2 cm (−0.3)	8.6 kg (−1.2)	Crawl (+), stand with support (+), meaningful words (+)	1973	ND	ND	ND	ND
16 m	78.5 cm (−0.1)	9.6 kg (−0.5)	Sit alone (+), loss of subcutaneous fat on his face and extremities	2293	252	ND	ND	ND
2 y 6 m	88.0 cm (−0.5)	12.9 kg (+0.2)	Walk alone (+), 2-word sentences (+)	ND	ND	ND	ND	ND
2 y 10 m	92.7 cm (+0.2)	13.2 kg (−0.1)	Run (+), jump (−), normal glucose tolerance and no insulin resistance, low level of adipokines	1715	213	309	1.2	1.9
3 y 11 m	99.2 cm (−0.1)	15.3 kg (−0.1)	Insulin resistance (+)	1777	154	680	1.1	1.6

ND: not done.

supraventricular, and ventricular tachycardia, may develop after 8–10 years of age [3,4,6]. Careful follow-up is necessary for a better prognosis.

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### References

- [1] Garg A, Agarwal AK. Lipodystrophies: disorders of adipose tissue biology. *Biochim Biophys Acta* 2009;1791:507–13.
- [2] Garg A. Lipodystrophies: genetic and acquired body fat disorders. *J Clin Endocrinol Metab* 2011;96:3313–25.
- [3] Hayashi YK, Matsuda C, Ogawa M, et al. Human *PTRF* mutations cause secondary deficiency of caveolins resulting in muscular dystrophy with generalized lipodystrophy. *J Clin Invest* 2009;119:2623–33.
- [4] Rajab A, Straub V, McCann LJ, et al. Fatal cardiac arrhythmia and long-QT syndrome in a new form of congenital generalized lipodystrophy with muscle rippling (CGL4) due to *PTRF-CAVIN* mutations. *PLoS Genet* 2010;6:e1000874.
- [5] Dwianingsih EK, Takeshima Y, Itoh K, et al. A Japanese child with asymptomatic elevation of serum creatine kinase shows *PTRF-Cavin* mutation matching with congenital generalized lipodystrophy type 4. *Mol Genet Metab* 2010;101:233–7.
- [6] Shastry S, Delgado MR, Dirik E, Mehmet T, Agrarwal AK, Garg A. Congenital generalized lipodystrophy, type 4 (CGL4) associated with myopathy due to novel *PTRF* mutations. *Am J Med Genet A* 2010;152A:2245–53.
- [7] Ranzani B, Combs TP, Wang XB, et al. Cavoline-1-deficient mice are lean, resistant to diet-induced obesity, and show hypertriglyceridemia with adipocyte abnormalities. *J Biol Chem* 2002;277:8635–47.
- [8] Cohen AW, Razani B, Wang XB, et al. Caveolin-1-deficient mice show insulin resistance and defective insulin receptor protein expression in adipose tissue. *Am J Physiol Cell Physiol* 2003;285:c222–35.
- [9] Gnozález-Muñoz E, López-Iglesias C, Calvo M, Palacín M, Zorzano A, Camps M. Cavolin-1 loss of function accelerates glucose transporter 4 and insulin receptor degradation in 3T3-L1 adipocytes. *Endocrinology* 2003;150:3493–502.
- [10] Liu L, Brown D, McKee M, et al. Deletion of *Cavin/PTRF* causes global loss of caveolae, dyslipidemia and glucose intolerance. *Cell Metab* 2008;8:310–7.