

micrognathia and sparse and depigmented scalp hair), hypoplasia of the right clavicle and other long bones, aplasia of the left clavicle, absence of bilateral thumbs and halluces, hypoplasia of the distal phalanges, multiple appendicular bone fracture and proctatresia (Figures 1a-c). Congenital cataract and hearing loss were also noted. Head computed tomography showed severe hypoplasia of the cerebrum and enlargement of lateral, third and fourth ventricles (Figure 1d). At the age of 2 years, her height was 67.4 cm (−4.9 SD), body weight was 9.1 kg (−1.9 SD) and head was circumference 46.2 cm (−0.2 SD). Her developmental mile stones were severely retarded. She was bedridden and unable to support her head. She was therefore diagnosed with YVS.

To identify the genetic cause of the proband, we performed whole-exome sequencing on the patient and her parents as *FIG4* mutations were not described in YVS when we started the genetic analysis. Peripheral blood samples were collected from the trio (patient and both parents) after obtaining written informed consent. This study was approved by the institutional review board of Yokohama City University School of Medicine. Each individual's DNA was captured with the SureSelect Human All Exon v4 Kit (Agilent Technologies, Santa Clara, CA, USA) and sequenced on a HiSeq2000 with 101 bp paired-end reads and 7 bp index reads (Illumina, San Diego, CA, USA). Image analysis and base calling were performed by sequence control software real-time analysis and CASAVA software v1.8 (Illumina). The reads were aligned to a human reference genome (hg19) with Novoalign 2.08.02 (<http://www.novocraft.com/>). After the removal of PCR duplication by Picard, the variants were called by Genome Analysis Toolkit 1.6-5 (GATK: <http://www.broadinstitute.org/gatk/>)

and annotated by ANNOVAR (2012feb) (<http://www.openbioinformatics.org/annovar/>). Through this flow, common variants registered in common dbSNP137 (minor allele frequency ≥ 0.01) (<http://genome.ucsc.edu/cgi-bin/hgTrackUi?hgsid=335665349&c=chr6&g=snp137-Common>) were removed.

More than 93% of coding sequence was covered by at least 20 reads in each individual. Because this syndrome was suspected as an autosomal recessive disease because of the consanguinity and/or affected siblings in reported families,^{1,11,12} we extracted homozygous or compound heterozygous variants using whole-exome sequencing data. Through data processing synonymous variants, variants in segmental duplications, variants registered in dbSNP137 or our in-house database (exome data of 408 Japanese individuals) were all excluded from the candidates (Figure 2). As a result, only one compound heterozygous mutation in *FIG4* remained: c.1750 + 1delG and c.2285_2286delCT (p.S762Wfs*3), which were also confirmed by Sanger sequencing. The c.1750 + 1delG at intron 15 and c.2283_2284delCT at exon 20 were inherited from her mother and father, respectively (Figure 3).

Campeau *et al.*² proposed a model in which the clinical difference among the above-mentioned three conditions regarding *FIG4* abnormalities (that is, YVS, CMT4J and ALS/PLS) depends on how much residual *FIG4* function remains based on the following evidence: (1) all previously reported CMT4J patients had compound heterozygous mutations but carried one null allele and one missense allele, whereas all YVS patients had biallelic null mutations.^{7,13–15} (2) ALS/PLS patients had heterozygous null or partially loss-of-function mutations.⁵ (3) In functional assays,

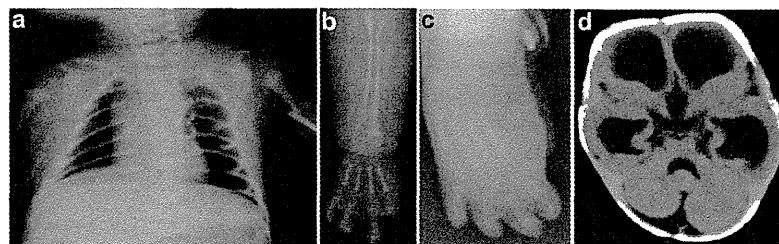


Figure 1 Imaging studies of the proband. (a) Thoracoabdominal X-ray on the first day after birth (dorsal position). Hypoplasia of both clavicle and the fracture of left humerus were evident. (b) X-ray of left forearm and hand. Absence of thumbs and hypoplastic phalangeal bones were observed. (c) X-ray of right foot. Absence of halluces and hypoplastic phalangeal bones was noted. (d) Head CT. Extended hypoplasia of the cerebrum and enlargement of the third and the lateral ventricles were noted. The fourth ventricle was also slightly enlarged.

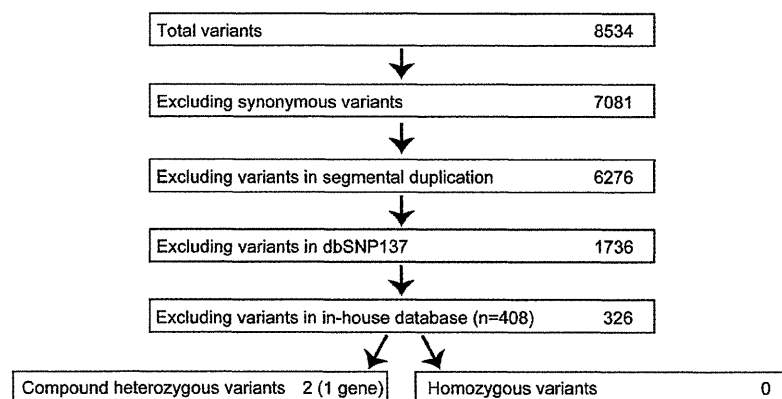


Figure 2 Priority scheme of whole-exome sequencing data. Numbers of variants surviving after each selection are shown.

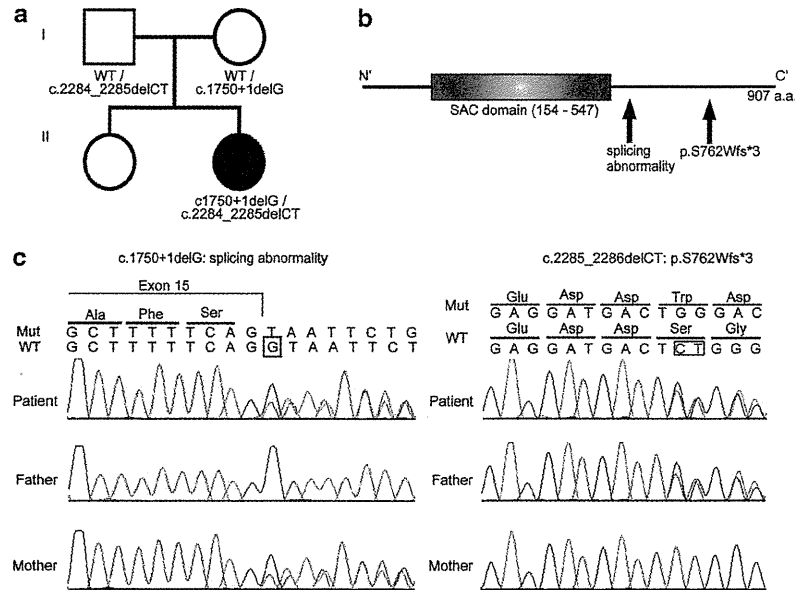


Figure 3 *FIG4* mutations in the patient. (a) Pedigree and mutations. (b) Schematic presentation of the *FIG4* protein and mutations found in the patient. (c) Electropherograms corresponding to each mutation in the patient, her father and mother. WT: wild-type allele, Mut: mutant allele. The altered or deleted bases were marked by square.

CMT4J patients had less *FIG4* activity than ALS/PLS patients.⁵ Biallelic null mutations in our YVS patient support this model.

The clinical effects of heterozygous *FIG4* mutations remain unclear. Theoretically, all parents (excluding the patient having a *de novo* truncation mutation) of YVS patients should carry a heterozygous null mutation. However, none of the parents of identified CMT4J patients or YVS patients, who should be heterozygous carriers, have been reported to show ALS/PLS. The current age of the father and mother in this study were 27 and 34 years old, respectively, and they did not show any ALS/PLS phenotype. Furthermore, the patient's grandparents did not show any sign of ALS/PLS. This discrepancy could be presymptomatic status of the family members carrying a heterozygous mutation as the average onset age of ALS/PLS in patients with *FIG4* mutation was reported to be 56 ± 14 years (mean \pm SD).⁵ It would be strongly encouraged to observe their longitudinal clinical course of heterozygous carriers in human. The other possibility for this discrepancy would be unknown genetic modifier(s) leading to the incomplete penetrance. To clarify the genotype–phenotype correlation and the diverse clinical phenotypes by *FIG4* mutations, further studies of genetic and clinical features are necessary.

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- Yunis, E. & Varon, H. Cleidocranial dysostosis, severe micrognathism, bilateral absence of thumbs and first metatarsal bone, and distal aphalangia: a new genetic syndrome. *Am. J. Dis. Child.* **134**, 649–653 (1980).
- Campeau, P. M., Lenk, G. M., Lu, J. T., Bae, Y., Burrage, L., Turpenney, P. *et al.* Yunis–Varon syndrome is caused by mutations in *FIG4*, encoding a phosphoinositide phosphatase. *Am. J. Hum. Genet.* **92**, 781–791 (2013).
- Martyn, C. & Li, J. *Fig4* deficiency: a newly emerged lysosomal storage disorder? *Prog. Neurobiol.* **101–102**, 35–45 (2013).
- Huotari, J. & Helenius, A. Endosome maturation. *EMBO J.* **30**, 3481–3500 (2011).
- Chow, C. Y., Landers, J. E., Bergren, S. K., Sapp, P. C., Grant, A. E., Jones, J. M. *et al.* Deleterious variants of *FIG4*, a phosphoinositide phosphatase, in patients with ALS. *Am. J. Hum. Genet.* **84**, 85–88 (2009).
- Zhang, X., Chow, C. Y., Sahenk, Z., Sny, M. E., Meisler, M. H. & Li, J. Mutation of *FIG4* causes a rapidly progressive, asymmetric neuronal degeneration. *Brain* **131**, 1990–2001 (2008).
- Nicholson, G., Lenk, G. M., Reddel, S. W., Grant, A. E., Towne, C. F., Ferguson, C. J. *et al.* Distinctive genetic and clinical features of CMT4J: a severe neuropathy caused by mutations in the PI(3,5)P(2) phosphatase *FIG4*. *Brain* **134**, 1959–1971 (2011).
- Garrett, C., Berry, A. C., Simpson, R. H. & Hall, C. M. Yunis–Varon syndrome with severe osteodysplasia. *J. Med. Genet.* **27**, 114–121 (1990).
- Walch, E., Schmidt, M., Brenner, R. E., Ermons, D., Dame, C., Pontz, B. *et al.* Yunis–Varon syndrome: evidence for a lysosomal storage disease. *Am. J. Med. Genet.* **95**, 157–160 (2000).
- Dworzak, F., Mora, M., Borroni, C., Cornelio, F., Blasevich, F., Cappellini, A. *et al.* Generalized lysosomal storage in Yunis Varon syndrome. *Neuromuscul. Disord.* **5**, 423–428 (1995).
- Rabe, H., Brune, T., Rossi, R., Steinhilber, V., Jorch, G., Horst, J. *et al.* Yunis–Varon syndrome: the first case of German origin. *Clin. Dysmorphol.* **5**, 217–222 (1996).
- Pfeiffer, R. A., Diekmann, L. & Stock, H. J. Aplasia of the thumbs and great toes as the outstanding feature of Yunis and Varon syndrome. A new entity. A new observation. *Ann. Genet.* **31**, 241–243 (1988).
- Chow, C. Y., Zhang, Y., Dowling, J. J., Jin, N., Adamska, M., Shiga, K. *et al.* Mutation of *FIG4* causes neurodegeneration in the pale tremor mouse and patients with CMT4J. *Nature* **448**, 68–72 (2007).
- de Leeuw, C. N. CMT4J: Charcot–Marie–Tooth disorder caused by mutations in *FIG4*. *Clin. Genet.* **73**, 318–319 (2008).
- Ikonomov, O. C., Sbrissa, D., Fligger, J., Delvecchio, K. & Shisheva, A. ArPIKfyve regulates Sac3 protein abundance and turnover: disruption of the mechanism by Sac3I41T mutation causing Charcot–Marie–Tooth 4J disorder. *J. Biol. Chem.* **285**, 26760–26764 (2010).

