Table VI. Determination of Lipid Concentration in Four Lots of DOXIL®

		Calculated concentration (n=3; µg/ml, RSD)			
Lipid	Ingredient amount on labeling (µg/ml)	Lot#011AFL	Lot#012AGD	Lot#029BJD	Lot#032BKA
Chol ^a	3,190	3,438.13 (0.29)	3,561.10 (0.41)	3,269.43 (0.24)	3,416.59 (0.26)
$HSPC^b$	9,580	8,420.41 (1.18)	8,859.23 (0.48)	8,836.49 (1.53)	9,328.49 (1.22)
DSPE-PEG	3,190	2,435.90 (0.76)	2,552.80 (1.18)	2,686.19 (1.09)	2,757.95 (1.22)
Calculated molar ratio Chol/HSPC/DSPE-PEG		43.49/52.17/4.34	43.12/52.53/4.35	40.99/54.27/4.74	40.80/54.56/4.64

[&]quot;Chol was detected by a UV detector

starting material. When compared lot-to-lot, while there were almost no differences between the two older lots or the two relatively newer lots, 0.4–2.4% differences were observed between the old and new lots. The ratio of Chol was higher in the old lots and those of HSPC and DSPE-PEG were lower. This may be due to the hydrolysis of phospholipids, which resulted in a reduced amount of HSPC and DSPC-PEG.

Analysis of Lipid Composition After Storage at Different Temperatures

Thus, the developed method was used to assess liposome stability. It is known that the hydrolysis of phospholipid follows a pseudo first-order kinetics model and that the hydrolysis rate correspondingly increases with temperature (7). First, the prepared liposomes were incubated for 3 days at 4, 37, or 57°C and the concentrations of the main hydrolysis products, S-LysoPC and SA, were measured. A significant increase in S-LysoPC and SA was observed with a rise in temperature (Fig. 3). Consistent

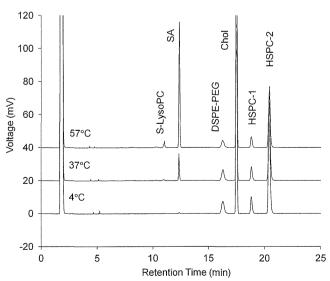


Fig. 3. A stability evaluation of the PEG liposome under each thermal condition (4°C, 37°C, and 57°C). The lipid composition of the PEG liposome, which was dialyzed against 10% sucrose, is presented in Table VI. The PEG liposome was incubated at each temperature for 3 days and diluted with methanol, and 20- μ l aliquots of the diluted solution were injected

with this observation, the concentrations of HSPC and DSPE-PEG decreased and that of Chol was stable (data not shown). Next, the concentration of the hydrolysis products in the four lots of DOXIL® were measured (Table VII). The concentrations of S-LysoPC and SA significantly increased in the two older lots (long past the expiration date). It was confirmed that even under storage at 4°C and solution of pH 6.5, i.e., conditions in which the hydrolysis rate was the least (8), the lysophospholipid and free fatty acids were produced by hydrolysis during long-term storage.

CONCLUSIONS

A simple reversed-phase HPLC-ELSD method was developed for the quantification of lipids in PEGylated liposomes. Although the linearity and accuracy obtained with UV detection of Chol were better those obtained with ELSD, the HPLC-ELSD method was validated to be linear, precise, accurate, and sensitive. Additionally, the HPLC-UV/ELSD method was found to be suitable for simultaneous determination of HSPC, DSPC-PEG, and Chol as well as their hydrolysis products in PEGylated liposomal products. It was also suggested that accurate quantification of the lipid component enables assessment of changes in lipid composition during the preparation process. In addition, the increase of hydrolysis products of phospholipids under a heat-accelerated condition may be observed. This method will be useful for quantifying the hydrolysis products in liposomal products in a stability test, such as a long-term storage test or an accelerated test, as well as for quantifying the lipid composition of liposomal products.

Table VII. Determination of S-LysoPC and SA Concentration in Four Lots of DOXIL by the HPLC-ELSD

	Calculated concentration (n=3; μg/ml, RSD)					
	Lot#011AFL	Lot#012AGD	Lot#029BJD	Lot#032BKA		
	Approximate elapsed time after manufacture					
Lipid	68 months	65 months	29 months	26 months		
S - LysoPC	442.83 (3.36)	425.00 (5.56)	238.06 (1.66)	202.61 (2.74)		
SA	374.50 (2.10)	376.46 (3.04)	215.19 (2.30)	175.17 (4.59)		

^b The value of HSPC was calculated from the peak of HSPC-1

ACKNOWLEDGMENTS

We thank Professor Maitani, Hoshi University, and Professor Mruyama and Dr. Suzuki, Teikyo University, for their advice regarding the preparation of PEGylated liposomes. This study was supported by the Research on Publicly Essential Drugs and Medical Devices from the Japan Health Sciences Foundation (KHB1005 and KHB1206).

REFERENCES

- 1. Allen TM, Hansen C, Martin F, Redemann C, Yau-Young A. Liposomes containing synthetic lipid derivatives of poly(ethylene glycol) show prolonged circulation half-lives *in vivo*. Biochim Biophys Acta. 1991;1066(1):29–36.
- Papahadjopoulos D, Allen TM, Gabizon A, Mayhew E, Matthay K, Huang SK, et al. Sterically stabilized liposomes: improvements in pharmacokinetics and antitumor therapeutic efficacy. Proc Natl Acad Sci U S A. 1991;88(24):11460–4.
- Adler-Moore J, Proffitt RT. Effect of tissue penetration on AmBisome efficacy. Curr Opin Investig Drugs. 2003;4(2):179–85.
 Zhao Y, Zhang S, Cui S, Wang B. Peptide-based cationic lipo-
- Zhao Y, Zhang S, Cui S, Wang B. Peptide-based cationic liposome-mediated gene delivery. Expert Opin Drug Deliv. 2012;9(1):127-39. doi:10.1517/17425247.2011.630387.
- 5. Inoue K, Kitagawa T. Effect of exogenous lysolecithin on liposomal membranes. Its relation to membrane fluidity. Biochim Biophys Acta. 1974;363(3):361–72.
- Zuidam NJ, Gouw HK, Barenholz Y, Crommelin DJ. Physical (in) stability of liposomes upon chemical hydrolysis: the role of lysophospholipids and fatty acids. Biochim Biophys Acta. 1995;1240(1):101-10.
- Zuidam NJ, Crommelin DJ. Chemical hydrolysis of phospholipids. J Pharm Sci. 1995;84(9):1113–9.
- Grit M, Underberg WJ, Crommelin DJ. Hydrolysis of saturated soybean phosphatidylcholine in aqueous liposome dispersions. J Pharm Sci. 1993;82(4):362-6.
- Zhang JA, Pawelchak J. Effect of pH, ionic strength and oxygen burden on the chemical stability of EPC/cholesterol liposomes under accelerated conditions. Part 1: lipid hydrolysis. Eur J Pharm Biopharm. 2000;50(3):357-64.
- FDA US (2002). Draft Guidance for Industry "Liposome Drug Products: Chemistry, Manufacturing, and Controls;

- Human Pharmacokinetics and Bioavailability; and Labeling Documentation". http://www.fda.gov/downloads/Drugs/GuidanceComplianceRegulatoryInformation/Guidances/UCM070570.pdf.
- 11. Mazzella N, Molinet J, Syakti AD, Dodi A, Doumenq P, Artaud J, et al. Bacterial phospholipid molecular species analysis by ion-pair reversed-phase HPLC/ESI/MS. J Lipid Res. 2004;45(7):1355-63. doi:10.1194/jlr.D300040-JLR200.
- 12. Rodriguez-Alcala LM, Fontecha J. Major lipid classes separation of buttermilk, and cows, goats and ewes milk by high performance liquid chromatography with an evaporative light scattering detector focused on the phospholipid fraction. J Chromatogr A. 2010;1217(18):3063–6. doi:10.1016/j.chroma.2010.02.073.
- Schonherr C, Touchene S, Wilser G, Peschka-Suss R, Francese G. Simple and precise detection of lipid compounds present within liposomal formulations using a charged aerosol detector. J Chromatogr A. 2009;1216(5):781-6. doi:10.1016/ j.chroma.2008.11.080.
- 14. Mengesha AE, Bummer PM. Simple chromatographic method for simultaneous analyses of phosphatidylcholine, lysophosphatidylcholine, and free fatty acids. AAPS PharmSciTech. 2010;11(3):1084-91. doi:10.1208/s12249-010-9470-4.
- Holland WL, Stauter EC, Stith BJ. Quantification of phosphatidic acid and lysophosphatidic acid by HPLC with evaporative lightscattering detection. J Lipid Res. 2003;44(4):854–8. doi:10.1194/ jlr.D200040-JLR200.
- Simonzadeh N. An isocratic HPLC method for the simultaneous determination of cholesterol, cardiolipin, and DOPC in lyophilized lipids and liposomal formulations. J Chromatogr Sci. 2009;47(4):304–8.
- 17. Jaaskelainen I, Urtti A. Liquid chromatography determination of liposome components using a light-scattering evaporative detector. J Pharm Biomed Anal. 1994;12(8):977–82.
- Zhong Z, Ji Q, Zhang JA. Analysis of cationic liposomes by reversed-phase HPLC with evaporative light-scattering detection. J Pharm Biomed Anal. 2010;51(4):947-51. doi:10.1016/j.jpba.2009.10.001.
- 19. Maitani Y, Soeda H, Junping W, Takayama K. Modified ethanol injection method for liposomes containing beta-sitosterol beta-D-glucoside. J Liposome Res. 2001;11(1):115-25. doi:10.1081/LPR-100103174.
- Shimizu Y, Nakata M, Matsunuma J, Mizuochi T. Simultaneous quantification of components of neoglycolipid-coated liposomes using high-performance liquid chromatography with evaporative light scattering detection. J Chromatogr B Biomed Sci Appl. 2001;754(1):127–33.

