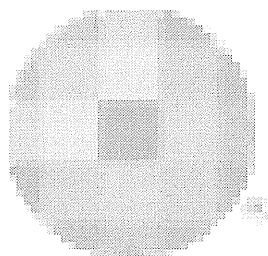


- human lung carcinoma growth. *Anal Cell Pathol* 1990;2:167-78.
235. Kayser K, Biechele U, Kayser G, Dienemann H, André S, Bovin NV, et al. Pulmonary metastases of breast carcinomas: Ligandohistochemical, nuclear, and structural analysis of primary and metastatic tumors with emphasis on period of occurrence of metastases and survival. *J Surg Oncol* 1998;69:137-46.
 236. Kayser K, Bovin NV, Korchagina EY, Zeilinger C, Zeng FY, Gabius HJ. Correlation of expression of binding sites for synthetic blood group A-, B- and H-trisaccharides and for sarcolectin with survival of patients with bronchial carcinoma. *Eur J Cancer* 1994;30A: 653-7.
 237. Kayser K, Bubbenzer J, Kayser G, Eichhorn S, Zemlyanukhina TV, Bovin NV, et al. Expression of lectin, interleukin-2 and histopathologic blood group binding sites in prostate cancer and its correlation with integrated optical density and syntactic structure analysis. *Anal Quant Cytol Histol* 1995;17:135-42.
 238. Kayser K, Görtler J, Bogovac M, Bogovac A, Goldmann T, Vollmer E, et al. AI (artificial intelligence) in histopathology: From image analysis to automated diagnosis. *Folia Histochem Cytobiol* 2009;47:355-61.
 239. Kayser K, Görtler J, Borkenfeld S, Kayser G. Grid computing in image analysis. *Diagn Pathol* 2011;6:S12.
 240. Kayser K, Görtler J, Borkenfeld S, Kayser G. How to measure diagnosis-associated information in virtual slides. *Diagn Pathol* 2011;6:S9.
 241. Kayser K, Görtler J, Goldmann T, Vollmer E, Hufnagl P, Kayser G. Image standards in tissue-based diagnosis (diagnostic surgical pathology). *Diagn Pathol* 2008;3:17.
 242. Kayser K, Molnar B, Weinstein RS. Virtual microscopy-fundamentals-applications-perspectives of electronic tissue-based diagnosis. Berlin: VSV Interdisciplinary Medical Publishing; 2006.
 243. Swett HA, Holaday L, Leffell D, Merrell RC, Morrow JS, Rosser JC, et al. Telemedicine: Delivering medical expertise across the state and around the world. *Conn Med* 1995;59:593-602.
 244. Walter GF, Matthies HK, Brandis A, von Jan U. Telemedicine of the future: Teleneuropathology. *Technol Health Care* 2000;8:25-34.
 245. Weinstein RS. Innovations in medical imaging and virtual microscopy. *Hum Pathol* 2005;36:317-9.
 246. Weinstein RS. Time for a reality check. *Arch Pathol Lab Med* 2008;132:777-80.
 247. Wootton R. Realtime telemedicine. *J Telemed Telecare* 2006;12:328-36.
 248. Kayser K. Telepathology – Visual telecommunication in pathology. An introduction. *Zentralbl Pathol* 1992;138:381-2.
 249. Kayser K. Progress in telepathology. *In Vivo* 1993;7:331-3.
 250. Molinari G, Reboa G, Frascio M, Leoncini M, Rolandi A, Balzan C, et al. The role of telecardiology in supporting the decision-making process of general practitioners during the management of patients with suspected cardiac events. *J Telemed Telecare* 2002;8:97-101.
 251. Rovetta A, Sala R, Bressanelli M, Garavaldi ME, Lorini F, Pegoraro R, et al. Demonstration of surgical telerobotics and virtual telepresence by Internet+ISDN from Monterey (USA) to Milan (Italy). *Stud Health Technol Inform* 1998;50:79-83.
 252. Weinstein RS, Bloom KJ, Krupinski EA, Rozek LS. Human performance studies of the video microscopy component of a dynamic telepathology system. *Zentralbl Pathol* 1992;138:399-403.
 253. Weinstein RS, Bloom KJ, Rozek LS. Telepathology. Long-distance diagnosis. *Am J Clin Pathol* 1989;91:S39-42.
 254. Weinstein RS, Descour MR, Liang C, Bhattacharyya AK, Graham AR, Davis JR, et al. Telepathology overview: From concept to implementation. *Hum Pathol* 2001;32:1283-99.
 255. Williams BH. The AFIP center for telemedicine application – Pathology for the twenty-first century. *Telemed Virtual Real* 1998;3:64-5.
 256. Bogomolov VV, Castrucci F, Comtois JM, Damann V, Davis JR, Duncan JM, et al. International Space Station medical standards and certification for space flight participants. *Aviat Space Environ Med* 2007;78:1162-9.
 257. McFarlin K, Sargsyan AE, Melton S, Hamilton DR, Dulchavsky SA. A surgeon's guide to the universe. *Surgery* 2006;139:587-90.
 258. Otto C, Comtois JM, Sargsyan A, Dulchavsky A, Rubinfeld I, Dulchavsky S. The Martian chronicles: Remotely guided diagnosis and treatment in the Arctic Circle. *Surg Endosc* 2010;24:2170-7.
 259. Otto C, Hamilton DR, Levine BD, Hare C, Sargsyan AE, Altshuler P, et al. Into thin air: Extreme ultrasound on Mt Everest. *Wilderness Environ Med* 2009;20:283-9.
 260. Sargsyan AE, Doarn CR, Simmons SC. Internet and World Wide Web technologies for medical data management and remote access to clinical expertise. *Tex Med* 1998;94:75-80.
 261. Sargsyan AE, Doarn CR, Simmons SC. Internet and World Wide Web technologies for medical data management and remote access to clinical expertise. *Aviat Space Environ Med* 1999;70:185-90.
 262. Sargsyan AE, Hamilton DR, Jones JA, Melton S, Whitson PA, Kirkpatrick AW, et al. FAST at MACH 20: Clinical ultrasound aboard the International Space Station. *J Trauma* 2005;58:35-9.
 263. Sargsyan AE, Hamilton DR, Nicolaou S, Kirkpatrick AW, Campbell MR, Billica RD, et al. Ultrasound evaluation of the magnitude of pneumothorax: A new concept. *Am Surg* 2001;67:232-5.
 264. Kayser K. Telemedicine. *Wien Klin Wochenschr* 1996;108:532-40.
 265. Molinari G, Valbusa A, Terrizzano M, Bazzano M, Torelli L, Girardi N, et al. Nine years' experience of telecardiology in primary care. *J Telemed Telecare* 2004;10:249-53.
 266. Weinstein RS. Telepathology: Practicing pathology in two places at once. *Clin Lab Manage Rev* 1992;6:171-3.
 267. Weinstein RS, Bloom KJ, Rozek LS. Telepathology and the networking of pathology diagnostic services. *Arch Pathol Lab Med* 1987;111:646-52.
 268. Frost JK. Cytology's challenge today – For a better tomorrow. *Acta Cytol* 1966;10:311-5.
 269. Frost JK. Education and training of the pathologist in cytopathology. *Acta Cytol* 1977;21:661-5.
 270. Frost JK. The cell in health and disease. An evaluation of cellular morphologic expression of biologic behavior. 2nd, revised edition. *Monogr Clin Cytol* 1986;2:1-304.
 271. Frost JK, Ball WC Jr, Levin ML, Tockman MS, Erozan YS, Gupta PK. Sputum for cytologic diagnosis of lung cancer. *N Engl J Med* 1982;306:109-10.
 272. Weinberg DS, Allaert FA, Dusserre P, Drouot F, Retailiau B, Welch VVR, et al. Telepathology diagnosis by means of digital still images: An international validation study. *Hum Pathol* 1996;27:111-8.
 273. Weinstein RS. Prospects for telepathology. *Hum Pathol* 1986;17:433-4.
 274. Kayser K, Gabius HJ, Ciesiolka T, Ebert VW, Bach S. Histopathologic evaluation of application of labeled neoglycoproteins in primary bronchus carcinoma. *Hum Pathol* 1989;20:352-60.
 275. Eide TJ, Nordrum I, Engum B, Rinde E. Use of telecommunications in pathology and anatomy services. *Tidsskr Nor Laegeforen* 1991;111:17-9.
 276. Busch C. Telepathology in Sweden. A national study including all histopathology and cytology laboratories. *Zentralbl Pathol* 1992;138:429-30.
 277. Martin E, Dusserre P, Fages A, Hauri P, Vieillefond A, Bastien H. Telepathology: A new tool of pathology? Presentation of a French national network. *Zentralbl Pathol* 1992;138:419-23.
 278. Miaoulis G, Protopapa E, Skourlas C, Deldis G. Telepathology in Greece. Experience of the Metaxas Cancer Institute. *Zentralbl Pathol* 1992;138:425-8.
 279. Schwarzmann P. Telemicroscopy. Design considerations for a key tool in telepathology. *Zentralbl Pathol* 1992;138:383-7.
 280. Dunn BE, Almagro UA, Choi H, Recla DL, Weinstein RS. Use of telepathology for routine surgical pathology review in a test bed in the Department of Veterans Affairs. *Telemed J* 1997;3:1-10.
 281. Dunn BE, Almagro UA, Choi H, Sheth NK, Arnold JS, Recla DL, et al. Dynamic-robotic telepathology: Department of Veterans Affairs feasibility study. *Hum Pathol* 1997;28:8-12.
 282. Dusserre P, Allaert FA, Dusserre L. Basic rules for the security of frozen section diagnosis through image transmission between anatomic-pathologists. *Stud Health Technol Inform* 1997;43:171-5.
 283. Weinstein RS, Bhattacharyya AK, Graham AR, Davis JR. Telepathology: A ten-year progress report. *Hum Pathol* 1997;28:1-7.
 284. Della Mea V. Image acquisition devices for telepathology. *Adv Clin Pathol* 1998;2:169-170.
 285. Kayser K. Telepathology, images, and multimedia archives. *Adv Clin Pathol* 1998;2:157.
 286. Mairinger T, Netzer TT, Schoner W, Gschwendtner A. Pathologists' attitudes to implementing telepathology. *J Telemed Telecare* 1998;4:41-6.
 287. Park S, Pantanowitz L, Parwania AV. Digital imaging in pathology. *Clin Lab Med* 2012;32:557-84.
 288. Garcia Rojo M, Punys V, Slodkowska J, Schrader T, Daniel C, Blobel B. Digital pathology in Europe: Coordinating patient care and research efforts. *Stud*

- Health Technol Inform 2009;150:997-1001.
289. Kidiashvili E, Schrader T. Implementation of telepathology in the republic of georgia. *Telemed J E Health* 2009;15:479-83.
290. Massone C, Wurm EM, Soyer HP. Teledermatology. *G Ital Dermatol Venereol* 2008;143:213-8.
291. Hartvigsen G, Johansen MA, Hasvold P, Bellika JG, Arsand E, Arild E, et al. Challenges in telemedicine and eHealth: Lessons learned from 20 years with telemedicine in Tromsø. *Stud Health Technol Inform* 2007;129:82-6.
292. Jones SM, Banwell PE, Shakespeare PG. Telemedicine in wound healing. *Int Wound J* 2004;1:225-30.
293. Della Mea V. Pre-recorded telemedicine. *J Telemed Telecare* 2005;11:276-84.
294. Pálsson T, Valdimarsdóttir M. Review on the state of telemedicine and eHealth in Iceland. *Int J Circumpolar Health* 2004;63:349-55.
295. Kropf R, Cipolat C, Burg G. Telemedicine in Europe. *Curr Probl Dermatol* 2003;32:247-51.
296. Aas IH. Telemedicine and changes in the distribution of tasks between levels of care. *J Telemed Telecare* 2002;8 (Suppl 2):1-2.
297. Aas IH. Telemedical work and cooperation. *J Telemed Telecare* 2001;7:212-8.
298. Eilford DR. Telemedicine in northern Norway. *J Telemed Telecare* 1997;3:1-22.
299. Smith L. Telemedicine applications clear time and distance barriers easily. *Health Manag Technol* 1996;17:22, 24, 27-9.
300. Massone C, Brunasso AM, Campbell TM, Soyer HP. State of the art of teledermatopathology. *Am J Dermatopathol* 2008;30:446-50.
301. Cipolat C, Bader U, Ruffli T, Burg G. Teledermatology in Switzerland. *Curr Probl Dermatol* 2003;32:257-60.
302. Pak HS. Teledermatology and teledermatopathology. *Semin Cutan Med Surg* 2002;21:179-89.
303. Mizushima H, Uchiyama E, Nagata H, Matsuno Y, Sekiguchi R, Ohmatsu H, et al. Japanese experience of telemedicine in oncology. *Int J Med Inform* 2001;61:207-15.
304. Kovai L, Lonari S, Paladino J, Kern J. The Croatian telemedicine. *Stud Health Technol Inform* 2000;77:146-50.
305. Plinkert PK, Plinkert B, Fuchs M, Zenner HP. Telemedicine in otorhinolaryngology exemplified by a Tübingen-Leipzig video conference. *HNO* 2000;48:728-34.
306. Buckner F. Telemedicine: The state of the art and current issues. *J Med Pract Manage* 1998;14:145-9.
307. Rosen E. Telemedicine German-style. *Telemed Today* 1999;7:13-5.
308. Schlag PM, Moesta KT, Rakovsky S, Graszew G. Telemedicine: The new must for surgery. *Arch Surg* 1999;134:1216-21.
309. Smits HL, Baum A. Health Care Financing Administration (HCFA) and reimbursement in telemedicine. *J Med Syst* 1995;19:139-42.
310. Gonçalves L, Cunha C. Telemedicine project in the Azores Islands. *Arch Anat Cytol Pathol* 1995;43:285-7.
311. Proceedings of the 1st European Symposium on Telepathology. Heidelberg, July 20-21, 1992. *Zentralbl Pathol* 1992;138:381-434.
312. Scalvini S, Glisenti F. Centenary of tele-electrocardiography and telephonocardiography-where are we today? *J Telemed Telecare* 2005;11:325-30.
313. Scalvini S, Zanelli E. Telecardiology: A new support for general practitioners in the management of elderly patients. *Age Ageing* 2002;31:153.
314. Scalvini S, Zanelli E, Domenighini D, Massarelli G, Zampini P, Giordano A, et al. Telecardiology community: A new approach to take care of cardiac patients. "Boario Home-Care" Investigators. *Cardiologia* 1999;44:921-4.
315. Sierdzinski J, Bala P, Rudowski R, Grabowski M, Karpinski G, Kaczynski B. KARDIONET: Telecardiology based on GRID technology. *Stud Health Technol Inform* 2009;150:463-7.
316. Shanit D. The Israel Center of Telemedicine. Telecardiology in the Negev. *Telemed Today* 1996;4:43-4.
317. Rendina MC. The effect of telemedicine on neonatal intensive care unit length of stay in very low birthweight infants. *Proc AMIA Symp* 1998;p.111-5.
318. Roth A, Rogowski O, Yanay Y, Kehati M, Malov N, Golovner M. Teleconsultation for cardiac patients: A comparison between nurses and physicians: The SHL experience in Israel. *Telemed J E Health* 2006;12:528-34.
319. Scalvini S, Capomolla S, Zanelli E, Benigno M, Domenighini D, Paletta L, et al. Effect of home-based telecardiology on chronic heart failure: Costs and outcomes. *J Telemed Telecare* 2005;11:16-8.
320. Scalvini S, Zanelli E, Conti C, Volterrani M, Pollina R, Giordano A, et al. Assessment of prehospital chest pain using telecardiology. *J Telemed Telecare* 2002;8:231-6.
321. Angelini A, Andersen CB, Bartoloni G, Black F, Bishop P, Doran H, et al. A web-based pilot study of inter-pathologist reproducibility using the ISHLT 2004 working formulation for biopsy diagnosis of cardiac allograft rejection: The European experience. *J Heart Lung Transplant* 2011;30:1214-20.
322. Ayad E. Virtual telepathology in Egypt, applications of WSI in Cairo University. *Diagn Pathol* 2011;6 Suppl 1:S1.
323. Biswas J, Das D, Vijayanthi P. Ophthalmic telepathology: Concept and practice. *Indian J Pathol Microbiol* 2010;53:571.
324. Della Mea V. Camera phones: An emergency solution. *J Telemed Telecare* 2010;16:165-6.
325. Giansanti D, Cerroni F, Amodeo R, Filoni M, Giovagnoli MR. A pilot study for the integration of cytometry reports in digital cytology telemedicine applications. *Ann Ist Super Sanita* 2010;46:138-43.
326. Intersimone D, Snoj V, Riosa F, Bortolotti N, Sverko S, Beltrami CA, et al. Transnational telepathology consultations using a basic digital microscope: Experience in the Italy-Slovenia INTERREG project "patient without borders". *Diagn Pathol* 2011;6 Suppl 1:S25.
327. Gortler J, Berghoff M, Kayser G, Kayser K. Grid technology in tissue-based diagnosis: fundamentals and potential developments. *Diagn Pathol* 2006;1:23.
328. Kayser K, Ogilvie R, Borkenfeld S, Kayser G. E-education in pathology including certification of e-institutions. *Diagn Pathol* 2011;6:S11.
329. Kumar N, Busarla SV, Sayed S, Kirimi JM, Okiro P, Gakinya SM, et al. Telecytology in East Africa: A feasibility study of forty cases using a static imaging system. *J Telemed Telecare* 2012;18:7-12.
330. Rojo MG, Castro AM, Gonçalves L. COST Action "EuroTelepath": Digital pathology integration in electronic health record, including primary care centres. *Diagn Pathol* 2011;6:S6.
331. Romo D, Romero E, González F. Learning regions of interest from low level maps in virtual microscopy. *Diagn Pathol* 2011;6:S22.
332. Słodkowska J, Markiewicz T, Grala B, Kozłowski W, Papierz W, Pleskacz K, et al. Accuracy of a remote quantitative image analysis in the whole slide images. *Diagn Pathol* 2011;6:S20.
333. Wałkowski S, Szymas J. Quality evaluation of virtual slides using methods based on comparing common image areas. *Diagn Pathol* 2011;6:S14.
334. Nordrum I, Engum B, Rinde E, Finseth A, Ericsson H, Kearney M, et al. Remote frozen section service: A telepathology project in northern Norway. *Hum Pathol* 1991;22:514-8.
335. Weinstein RS. Telepathology comes of age in Norway. *Hum Pathol* 1991;22:511-3.
336. Allen A, Hayes J, Sadasivan R, Williamson SK, Wittman C. A pilot study of the physician acceptance of tele-oncology. *J Telemed Telecare* 1995;1:34-7.
337. Kayser K. Telepathology in Europe. Its practical use. *Arch Anat Cytol Pathol* 1995;43:196-9.
338. Kayser K, Fritz P, Drlicek M. Aspects of telepathology in routine diagnostic work with specific emphasis on ISDN. *Arch Anat Cytol Pathol* 1995;43:216-8.
339. Telemed Virtual Real. Swedish hospitals field test telepathology. *Telemed Virtual Real* 1998;3:9.
340. Camby I I, Rimmelink M, Nagy N, Rombaut K, Kiss R, Salmon I I. Neuropathological consultation by means of telepathology: A clinical tool for imposing diagnosis of rare and difficult cases. *Adv Clin Path* 1998;2:152-3.
341. Dzubur A, Seiwerth S, Danilovic Z. Benefits of image databank supporting the telepathology system. *Adv Clin Path* 1998;2:158-159.
342. Bocker PB. ISDN- Digitale Netze für Sprach-, Text-, Daten-, Video- und Multimediakommunikation. Berlin, Heidelberg: Springer Verlag; 1997.
343. Demichelis F, Barbareschi M, Boi S, Clemente C, Dalla Palma P, Eccher C, et al. Robotic telepathology for intraoperative remote diagnosis using a still-imaging-based system. *Am J Clin Pathol* 2001;116:744-52.
344. Kayser K, Beyer M, Blum S, Kayser G. Recent developments and present status of telepathology. *Anal Cell Pathol* 2000;21:101-6.
345. Kayser K, Beyer M, Blum S, Kayser G. Telecommunication: A new tool for quality assurance and control in diagnostic pathology. *Folia Neuropathol* 2000;38:79-83.
346. Della Mea V, Cataldi P, Pertoldi B, Beltrami CA. Combining dynamic and static robotic telepathology: A report on 184 consecutive cases of frozen sections, histology and cytology. *Anal Cell Pathol* 2000;20:33-9.
347. Della Mea V, Cataldi P, Pertoldi B, Beltrami CA. Dynamic robotic

- telepathology: A preliminary evaluation on frozen sections, histology and cytology. *J Telemed Telecare* 1999;5:555-6.
348. Szymaś J, Wolf G. Remote microscopy through the internet. *Pol J Pathol* 1999;50:37-42.
349. Wolf G, Petersen I, Dietel M. Microscope remote control with an Internet browser. *Anal Quant Cytol Histol* 1998;20:127-32.
350. Nordrum I, Eide TJ. Remote frozen section service in Norway. *Arch Anat Cytol Pathol* 1995;43:253-6.
351. Oberholzer M, Fischer HR, Christen H, Gerber S, Brühlmann M, Mihatsch MJ, et al. Telepathology: Frozen section diagnosis at a distance. *Virchows Arch* 1995;426:3-9.
352. Della Mea V, Cortolezzi D, Beltrami CA. The economics of telepathology – A case study. *J Telemed Telecare* 2000;6:S168-9.
353. Reith A. Experience with the use of telemedicine in Norway. *Int J Comput Dent* 2002;5:115-7.
354. Schwarzmann P, Binder B, Käser M, Klose R. European field tests with HISTKOM telepathology equipment. *Stud Health Technol Inform* 1999;64:192-207.
355. Schwarzmann P, Binder B, Klose R. Technical aspects of telepathology with emphasis on future development. *Anal Cell Pathol* 2000;21:107-26.
356. Schwarzmann P, Binder B, Klose R, Kaeser M. Histkom-evaluation of active telepathology in fieldtests. *Adv Clin Path* 1998;2:135-138.
357. Schwarzmann P, Schmid J, Schnörr C, Strässle G, Witte S. Telemicroscopy stations for telepathology based on broadband and ISDN connections. *Arch Anat Cytol Pathol* 1995;43:209-15.
358. Nordrum I, Isaksen V, Arvola L. Breast carcinoma diagnosed by telepathology. *J Telemed Telecare* 1997;3:172-3.
359. Nordrum I, Johansen M, Amin A, Isaksen V, Ludvigsen JA. Diagnostic accuracy of second-opinion diagnoses based on still images. *Hum Pathol* 2004;35:129-35.
360. Kayser K. Telepathology in Europe. *Anal Cell Pathol* 2000;21:95-6.
361. Kayser K, Drlicek M, Rahn W. Aids of telepathology in intra-operative histomorphological tumor diagnosis and classification. *In Vivo* 1993;7:395-8.
362. Kayser K, Kayser G, Radziszowski D, Oehmann A. From telepathology to virtual pathology institution: The new world of digital pathology. *Rom J Morphol Embryol* 1999;45:3-9.
363. Weinstein RS. Static image telepathology in perspective. *Hum Pathol* 1996;27:99-101.
364. De Michelis F, Eccher C, Clemente C, Migliore G, Dalla Palma P, Forti S. A feasibility study of a static-robotic telepathology system for remote diagnosis. *Adv Clin Path* 1998;2:138-9.
365. Ferrer Roca OF, Ramos A, Diaz Cardama A. Immunohistochemical correlation of steroid receptors and disease-free interval in 206 consecutive cases of breast cancer: Validation of telequantification based on global scene segmentation. *Anal Cell Pathol* 1995;9:151-63.
366. Ferrer-Roca O. Telepathology and optical biopsy. *Int J Telemed Appl* 2009;2009:740712.
367. Galvez J, Howell L, Costa MJ, Davis R. Diagnostic concordance of telecytology and conventional cytology for evaluating breast aspirates. *Acta Cytol* 1998;42:663-7.
368. Gombas P. Informational aspects of telepathology in routine surgical pathology. *Anal Cell Pathol* 2000;21:141-7.
369. Kayser K, Kayser G, Becker HD, Herth F. Tele diagnosis of transbronchial fine needle aspirations: A feasibility study. *Anal Cell Pathol* 2000;21:207-12.
370. Kldiashvili E, Schrader T. Diagnostic accuracy and image quality using a USB digital eyepiece camera for telecytology-Georgian experience. *Telemed J E Health* 2010;16:1051-2.
371. Lee ES, Kim IS, Choi JS, Yeom BW, Kim HK, Han JH, et al. Accuracy and reproducibility of telecytology diagnosis of cervical smears. A tool for quality assurance programs. *Am J Clin Pathol* 2003;119:356-60.
372. Leong FJ, Graham AK, Schwarzmann P, McGee JO. Clinical trial of telepathology as an alternative modality in breast histopathology quality assurance. *Telemed J E Health* 2000;6:373-7.
373. Leong FJ, Nicholson AG, McGee JO. Robotic telepathology: Efficacy and usability in pulmonary pathology. *J Pathol* 2002;197:211-7.
374. Mairinger T. Acceptance of telepathology in daily practice. *Anal Cell Pathol* 2000;21:135-40.
375. Mairinger T, Gschwendtner A. Telecytology using preselected fields of view: The future of cytodiagnosis or a dead end? *Am J Clin Pathol* 1997;107:620-1.
376. Martin E, Dusserre P, Got C, Vieillefond A, Franc B, Brugal G, et al. Telepathology in France. Justifications and developments. *Arch Anat Cytol Pathol* 1995;43:191-5.
377. Martin ED, Dusserre P, Flandrin G, Got C, Vieillefond A, Vacher-Lavenu MC. Contribution of computers and telepathology in cancerologic pathology. *Bull Cancer* 1995;82 Suppl 5:565s-8.
378. Morgan MB, Tannenbaum M, Smoller BR. Telepathology in the diagnosis of routine dermatopathologic entities. *Arch Dermatol* 2003;139:637-40.
379. Schwarzmann P, Schmid J, Binder B, Burkart J. Field test to evaluate telepathology in telemedicine. *J Telemed Telecare* 1996;2 (Suppl 1):17-20.
380. Stauch G, Schweppe KV, Kayser K. Diagnostic errors in interactive telepathology. *Anal Cell Pathol* 2000;21:201-6.
381. Szymaś J, Papierz W, Danilewicz M. Real-time teleneuropathology for a second opinion of neurooncological cases. *Folia Neuropathol* 2000;38:43-6.
382. Kayser K, Fritz P, Drlicek M, Rahn W. Expert consultation by use of telepathology – The Heidelberg experiences. *Anal Cell Pathol* 1995;9:53-60.
383. Kayser K, Kayser G. Basic aspects of and recent developments in telepathology in Europe, with specific emphasis on quality assurance. *Anal Quant Cytol Histol* 1999;21:319-28.
384. Kayser K, Kayser G, Radziszowski D, Oehmann A. New developments in digital pathology: From telepathology to virtual pathology laboratory. *Stud Health Technol Inform* 2004;105:61-9.
385. Kuakpaetoon T, Stauch G, Visalsawadi P. Image quality and acceptance of Telepathology. *Adv Clin Path* 1998;2:305-12.
386. Brauchli K, Oberholzer M. Comparison of telepathology services. *J Telemed Telecare* 2004;10:307-8.
387. Brauchli K, Oberholzer M. The iPath telemedicine platform. *J Telemed Telecare* 2005;11 Suppl 2:S3-7.
388. Friedrich K, Scheithauer J, Dimmer V, Meyer W, Theissig F, Haroske G, et al. DNA ploidy and chromosomal imbalances in invasive ductal breast cancer. A comparative study of DNA image cytometry and comparative genomic hybridization (CGH). *Anal Cell Pathol* 2000;20:69-82.
389. Brauchli K, Christen H, Meyer P, Haroske G, Meyer W, Kunze KD, et al. Telepathology: Design of a modular system. *Anal Cell Pathol* 2000;21:193-9.
390. Giroud F, Haroske G, Reith A, Böcking A. 1997 ESACP consensus report on diagnostic DNA image cytometry. Part II: Specific recommendations for quality assurance. European Society for Analytical Cellular Pathology. *Anal Cell Pathol* 1998;17:201-8.
391. Kayser K, Blum S, Beyer M, Haroske G, Kunze KD, Meyer W. Routine DNA cytometry of benign and malignant pleural effusions by means of the remote quantitation server Euroquant: A prospective study. *J Clin Pathol* 2000;53:760-4.
392. Haroske G, Meyer W, Kunze D, Boeking A. Quality control measures for dna image cytometry in a telepathology network. *Adv Clin Path* 1998;2:143-5.
393. Schwarzmann P, Schenck U, Binder B, Schmid J. Is today's telepathology equipment also appropriate for telecytology? A pilot study with pap and blood smears. *Adv Clin Path* 1998;2:176-178.
394. Dietel M, Dierks C, Hufnagl P, Schlag PM. Automobile versus horse: Immediate diagnostic section by telepathology. *Pathologe* 2000;21:391-5.
395. Mireskandari M, Kayser G, Hufnagl P, Schrader T, Kayser K. Teleconsultation in diagnostic pathology: Experience from Iran and Germany with the use of two European telepathology servers. *J Telemed Telecare* 2004;10:99-103.
396. Kunze KD, Boeking A, Haroske G, Kayser K, Meyer W, Oberholzer M. Remote quantitation in the framework of telepathology. *Adv Clin Path* 1998;2:141-3.
397. van den Tweel JG, Bosman FT. The use of virtual slides in the EUROPALS examination. *Diagn Pathol* 2011;6 Suppl 1:S23.
398. Tsuchihashi Y. Expanding application of digital pathology in Japan: From education, telepathology to autodiagnosis. *Diagn Pathol* 2011;6 Suppl 1:S19.
399. Zerbe N, Hufnagl P, Schliuns K. Distributed computing in image analysis using open source frameworks and application to image sharpness assessment of histological whole slide images. *Diagn Pathol* 2011;6:S16.
400. Szymas J, Lundin M. Five years of experience teaching pathology to dental students using the VWebMicroscope. *Diagn Pathol* 2011;6 Suppl 1:S13.
401. Têtu B, Boulanger J, Houde C. Telepathology project on virtual slides of eastern Quebec: A clinical project carried out in 21 areas. *Ann Pathol* 2010;30 Suppl 1:25-7.
402. Johnson JR, Krupinski EA, Yan M, Roehrig H, Graham AR, Weinstein RS. Using

- a visual discrimination model for the detection of compression artifacts in virtual pathology images. *IEEE Trans Med Imaging* 2011;30:306-14.
403. Krupinski EA. Virtual slide telepathology workstation-of-the-future: Lessons learned from teleradiology. *Semin Diagn Pathol* 2009;26:194-205.
404. Glatz-Krieger K, Glatz D, Mihatsch MJ. Virtual slides: High-quality demand, physical limitations, and affordability. *Hum Pathol* 2003;34:968-74.
405. Glatz-Krieger K, Spornitz U, Spatz A, Mihatsch MJ, Glatz D. Factors to keep in mind when introducing virtual microscopy. *Virchows Arch* 2006;448:248-55.
406. Hufnagl P, Schlüns K. Virtual microscopy and routine diagnostics. A discussion paper. *Pathologie* 2008;29 Suppl 2:250-4.
407. Hipp J, Flotte T, Monaco J, Cheng J, Madabhushi A, Yagi Y, et al. Computer aided diagnostic tools aim to empower rather than replace pathologists: lessons learned from computational chess. *J Pathol Inform* 2011;2:25.
408. Kayser K, Görtler J, Borkenfeld S, Kayser G. Interactive and automated application of virtual microscopy. *Diagn Pathol* 2011;6 Suppl 1:S10.
409. Kayser K, Hoshang SA, Metzke K, Goldmann T, Vollmer E, Radziszowski D, et al. Texture- and object-related automated information analysis in histological still images of various organs. *Anal Quant Cytol Histol* 2008;30:323-35.
410. Oberholzer M, Christen H, Haroske G, Helfrich M, Oberli H, Jundt G, et al. Modern telepathology: A distributed system with open standards. *Curr Probl Dermatol* 2003;32:102-14.
411. Williams BH, Hong IS, Mullick FG, Butler DR, Herring RF, O'Leary TJ. Image quality issues in a static image-based telepathology consultation practice. *Hum Pathol* 2003;34:1228-34.
412. Williams BH, Mullick FG, Becker RL, Kyte RT, Noe A. A national treasure goes online: The Armed Forces Institute of Pathology. *MD Comput* 1998;15:260-5.
413. Williams BH. Virtual slides: the AFIP experience. In: Ogilvie RW, editor. *Virtual Microscopy and Virtual Slides in Teaching, Diagnosis, and Research*. Boca Raton: CRC Press; 2005. p. 227-300.
414. Williams S, Henricks WH, Becich MJ, Toscano M, Carter AB. Telepathology for patient care: What am I getting myself into? *Adv Anat Pathol* 2010;17:130-49.
415. Dietel M, Nguyen-Dobinsky TN, Hufnagl P. The UICC Telepathology Consultation Center. International Union Against Cancer. A global approach to improving consultation for pathologists in cancer diagnosis. *Cancer* 2000;89:187-91.
416. Fontelo PA. Telepathology and the Internet. *Adv Clin Path* 1997;1:95-96.
417. Brauchli K, Jagilly R, Oberli H, Kunze KD, Phillips G, Hurwitz N, et al. Telepathology on the Solomon Islands – Two years' experience with a hybrid Web- and email-based telepathology system. *J Telemed Telecare* 2004;10:14-7.
418. Kayser K. Medical telecommunication systems today: what has been done – what should be done. *J Telecommun Syst Manage* 2012;1:1.
419. Collen MF. Origins of medical informatics. *West J Med* 1986;145:778-85.
420. Wilkerson MJ. Review of "Pathology informatics: Theory and practice" by L Pantanowitz, JM Tuthill, and UGJ Balis (Editors). *J Pathol Inform* 2012;3:38.
421. Blum BI, Duncan K. *A History of Medical Informatics*. New York: ACM Press; 1990.



陸沿岸部震災被災地域との皮膚科遠隔診療の試み —陸前高田診療所（岩手県医師会）と岩手医科大学皮膚科との遠隔皮膚科診療—

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The trial of dermatological tele-medicine with Rikuzen-Takata clinic (Iwate medical association) in the Sanriku shore area where is the earthquake disaster stricken area and the dermatology of Iwate Medical University

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要旨

東日本大震災津波により壊滅的な被害を受けた陸前高田地域の皮膚科医は皆無となった。そこで、陸前高田診療所（岩手県医師会）と岩手医科大学皮膚科との遠隔皮膚科診療を試み、遠隔診療が医療過疎の解消の一助となるか検討した。通信方法は専用回線を用い、診療現場に検査機器、ムービーカメラ、照明器具を用い、岩手医科大学には皮膚科専門医が待機するシステムを構築して行った。その結果、1) 遠隔医療機器システム立ち上げまでの時間は平均 40 分であった。2) 1 人の患者に要する遠隔医療の時間は平均 34 分であった。患者への説明と同意取得、診断機器や映像機器の切り替えに時間を要した。3) 診断一致率は 22 例中 21 例が一致 (95%) していた。診断確定に苦慮した例の多くは、①頭皮の毛髪間や指間、口腔内、陰部・皸裂部などの皮疹の焦点が合わない、②蕁麻疹など淡い紅斑の色調あるいは常色の軽い扁平な盛り上がりが見えにくい、③アナフィラキシー紫斑病など微小点状出血は映像では不明瞭である、④悪性黒色腫の初期病変や軽症の太田母斑の淡い黒色斑や青色斑は映像で不明瞭である、⑤真菌検査の菌糸の画像が不鮮明である、などであった。これらの問題は診断を補助する機器の充実で改善すると考えられた。4) 患者からの遠隔診療に対する評価は VAS で 66% であった。①大きなモニター画像に映し出され、おどろいた、②診察のスキンシップが感じられない、③診療時間が長すぎる、④カメラに追い回されている感じがする、などの意見があった。しかし、意見の多くは専門医の診療・判断を仰ぐことができ、安心感を示すものが多くみられた。本研究によって、他科の医師と機器操作に熟練した技術員の存在のもとに皮膚科遠隔医療が可能であることが示唆された。しかし、緊急に改善すべき、①遠隔医療に関する受診者の理解、②運用性に優れたムービーカメラの精度向上、などの問題点が提起された。

キーワード：東日本大震災津波、陸前高田地域、皮膚科遠隔診療、高性能ムービーカメラ、NTT 専用回線

1. はじめに

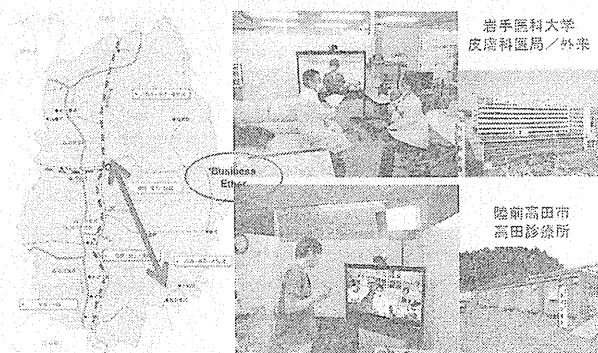
岩手県三陸沿岸地域は以前から医療過疎地域であった。皮膚科診療を有する総合病院が 5 カ所（うち皮膚科常勤医は 1 人）、皮膚科開業医診療所が 3 カ所と皮膚科領域についても医療過疎地域であった。東日本大震災津波により壊滅的な被害を受けた三陸沿岸地域、特に陸前高田地域では開業医 1 人によって皮膚科診療が行われていたが、震災に被災し大都市に避難したため同地域には皮膚科医は皆無となった。一方、岩手医科大学附属病院は、「岩手県東日本大震災津波復興計画」のなかで、被災した医療過疎地

域に対して皮膚科領域も含め医療情報機器等を活用した遠隔医療によって高度な専門医療を提供する役割を求められている。

一方、従来の皮膚科遠隔医療は個別的な支援や簡単な疾患の診断に留まっており、検査や診断・治療など総合的医療の提供はできていない。本研究では三陸沿岸部震災被災地域である陸前高田診療所（岩手県医師会）と岩手医科大学皮膚科との遠隔皮膚科診療を試み、遠隔診療が医療過疎の解消の一助となるかを検討した。

2. 目的

本研究では、①被災した医療過疎地域において災害拠点病院である大学病院が皮膚科遠隔医療によって高度医療を安定的に提供するための施設・設備・人員体制・コスト等についての検討と②対面診療と比較した遠隔医療の質についての検討を行う。①においては、専用回線を用い、診療現場に検査機器、ムービーカメラ、照明器具を用い、これらの器材の使用法に熟練した人材がいることと、また、岩手医科大学には皮膚科専門医が 2 名待機するシステムを構築する。②においては、皮膚疾患患者を対象として、陸前高田診療所における皮膚科専門医による対面診療と遠隔診療とを比較検討する【図 1】。



【図 1】 遠隔医療実証実験プロジェクト概要

3. 方法

1. 研究倫理および記録保存

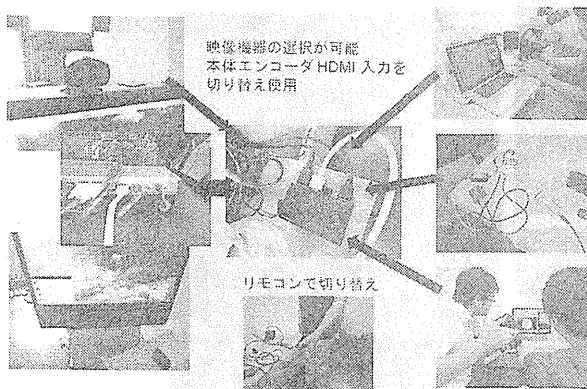
本研究は岩手医科大学倫理委員会の許可を得た。実験は患者のインフォームドコンセントを得て行う。患者情報や画像は匿名化し、個人を特定できないようにする。また、各患者の対面診療の動画は岩手医科大学情報センターにサーバー室を設けて保管した。医療情報は高田診療所の診療録に同診療所医師が記載し、同診療所に保管した。診療録の一部は患者および高田診療所の許可のもと、研究材料として用いた。

2. 利用回線および診療現場の器材

画像および医療情報の更新はNTT専用回線 (NTT Business Ether) を使用した。実験に先駆けて、対面診療による問診のためにテレビ電話付き大型モニターを含むテレビ会議伝信システム (フルHD (1080P/30fps)) 【図2】、患部の撮影のため2機の高性能ムービーカメラ、1機の接写カメラ、真菌検査および病理組織検査標本確認のためにオリンパス顕微鏡、患者情報記録のためノートパソコン、FAX機を設置した【図3】。それぞれを接続し、必要に応じてこれらの機器を切り替えて使用した。また、画像の色調を統一化、一定化するためにLED照明システムを使用した。これらのシステムで遠隔診断と医療提供が可能かを評価すると共にシステム設定にかかる時間も計測した。



【図2】高田診療所のシステム機器

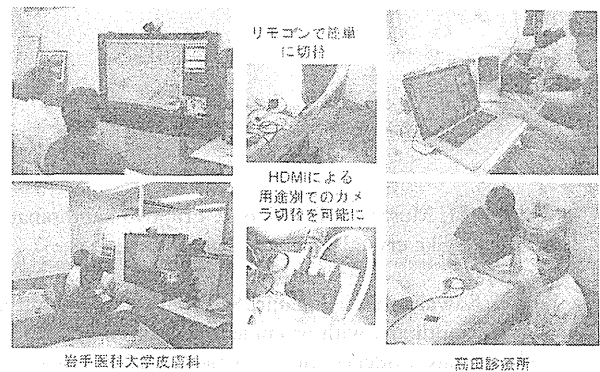


【図3】高田診療所カメラの切り替え状況

3. 遠隔対面診療の評価方法

研究の大半は高田診療所に皮膚科専門医が外向き、インフォームドコンセントの取得、患者の問診、診療録記載、処方箋発行、皮膚検査、機器の設定、皮膚病変の撮影、岩手医科大学皮膚科専門医との交信を行い、以下について評価した。平成25年1月末まで計22人の皮膚科患者の診療を行った【図4】。

- 1) 患者1人の診察時間
- 2) 診断名：高田診療所と岩手医科大学の皮膚科専門医の



【図4】皮膚科患者画像の送受信状況

診断の一致率

- 3) 皮疹の部位で診断しにくい部位
- 4) 皮疹の形態で診断しにくい皮疹
- 5) 患者満足度 (通常の対面診療と比較したVASで表示：100%が通常対面診療と同等、0%が全く対面診療に値しない)

4. 結果

- 1) 診療前の遠隔医療機器システム立ち上げまでにかかる時間は平均40分であった。熟練すると短縮可能と考えられた。
- 2) 1人の患者に要する遠隔医療の時間は最短26分、最長52分、平均34分であった。患者への説明と同意取得、診断機器や映像機器の切り替えに時間を要した。
- 3) 診断一致率は22例中21例が一致 (95%) していた。診断確定に苦慮した例の多くは、①頭皮の毛髪間や指間、口腔内、陰部・皸裂部などの皮疹の映像の焦点が合わない、②尋麻疹など淡い紅斑の色調あるいは常色の軽い扁平な盛り上がり画像で認識しがたい、③アナフィラキシー紫斑病など微小点状出血は映像では不明瞭である、④悪性黒色腫の初期病変や軽症の太田母斑の淡い黒色斑や青色斑は映像で不明瞭である、⑤真菌検査の菌糸の画像が不鮮明である、などによるものであった。これらの問題は診断を補助する機器の充実で改善すると考えられた。
- 4) 患者からの遠隔診療に対する評価はVisual Analog Scale (VAS) で66%であった。①大きなモニター画像に映し出され、おどろいた、②診察のスキンシップが感じられない、⑤診療時間が長すぎる、⑥カメラに追い回されている感じがする、などの意見があった。しかし、意見の多くは専門医の診療・判断を仰ぐことができ、安心感を示すものが多くみられた。

5. 考察

本研究の最終目標は遠隔地に皮膚科専門医がいない状況での遠隔診療である。他科の医師と機器操作に熟練した技術員の存在のもとに皮膚科遠隔医療が可能であることが示唆された。しかし、緊急に改善すべき以下の問題点が提起された。①遠隔医療に関する受診者の理解、②他科の医師の皮膚科遠隔医療に対する理解、③カメラ、検査機器、コンピュータの操作に熟練した技術員の存在、④患者誘導や発疹の選択に熟練した看護師の存在、⑤運用性に優れたムービーカメラの精度向上、⑥診断精度向上のための機器 (皮膚温検査機、エコー機器など) の必要性、⑦画像および遠隔診療カルテの保存方法の改善、⑧診療費用の配分。

References

- 1 Walsh NM, Murray S, D'Intino Y. Eruptive xanthomata with urate-like crystals. *J Cutan Pathol* 1994; 21: 350-355.
- 2 Bito T, Kawakami C, Shimajiri S, Tokura Y. Generalized eruptive xanthoma with prominent deposition of naked chylomicrons: evidence for chylomicrons as the origin of urate-like crystals. *J Cutan Pathol* 2010; 37: 1161-1163.
- 3 Kodama H, Akiyama H, Nagao Y, *et al.* Persistence of foam cells in rabbit xanthoma after normalization of serum cholesterol level. *Arch Dermatol Res* 1988; 280: 108-113.
- 4 Bergman R, Aviram M, Shemer A, *et al.* Enhanced low-density lipoprotein degradation and cholesterol synthesis in monocyte-derived macrophages of patients with adult xanthogranulomatosis. *J Invest Dermatol* 1993; 101: 880-882.

Annular elastolytic giant cell granuloma developing on lesions of vitiligo

Dear Editor,

A 74-year-old woman presented with a two-year history of reddish annular nodules on the neck, trunk, and forearms. She had a 10-year history of generalized vitiligo that had been untreated. The nodules with elevated borders and central atrophy and hypopigmentation, 0.5-1 cm in diameter, were located on the neck, trunk, and forearms (Fig. 1). These nodules had developed mainly on the preexisting vitiligo lesions. The patient had been taking benidipine hydrochloride for hypertension for the last 14 years. She had no history of diabetes mellitus or sarcoidosis.

A skin biopsy was taken from a nodule on the forearm. Hematoxylin and eosin staining of the specimen showed a granulomatous infiltrate of lymphocytes, histiocytes,

and multinucleated giant cells in the upper and middle dermis without palisading (Fig. 2a). Immunohistochemistry showed that the lymphocytes in the inflammatory infiltrate were positive for CD3. CD4⁺ T-cells predominated over CD8⁺ T-cells, and the histiocytes and multinucleated giant cells were positive for CD68. Almost all of the epidermal cells were negative for Melan-A (Fig. 2b). Slight mucin deposition was evident. Elastica van Gieson staining showed that elastic fibers were absent from the reticular dermis in the area surrounded by the granulomatous infiltrate (Fig. 2c). Fragmented elastic fibers were present within some of the histiocytes and multinucleated giant cells (Fig. 2d). Ziehl-Neelsen and PAS staining revealed no acid-fast bacilli or fungal organisms. Laboratory examinations, including a full blood cell count, routine biochemistry, fasting blood sugar level, and urinalysis gave normal results. These findings were consistent with

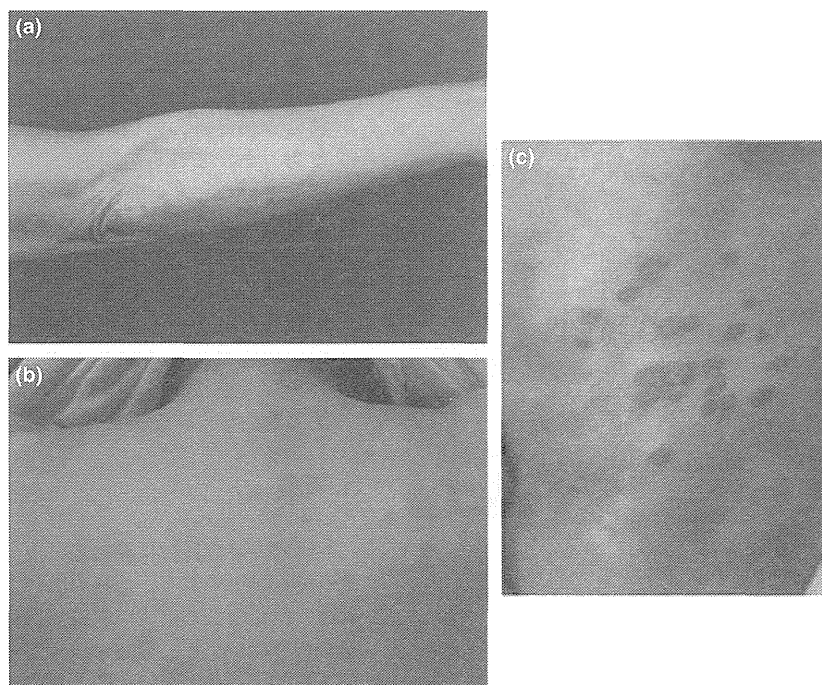


Figure 1 Multiple, erythematous, marginally elevated, annular nodules and round papules, developed mainly on lesions of preexisting vitiligo, distributed on the back (a), forearms (b) and abdomen (c)

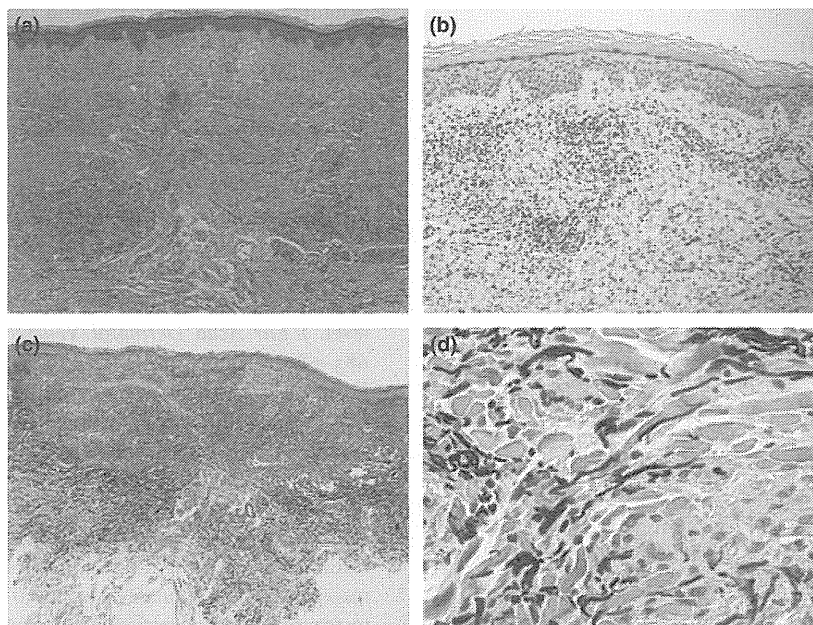


Figure 2 (a) Granulomatous infiltrates of lymphocytes, histiocytes and multinucleated giant cells in the upper and middle dermis without palisading (hematoxylin and eosin, original magnification $\times 40$). (b) Almost all of the epidermal cells were negative for Melan-A (original magnification $\times 100$). (c) Elastica van Gieson staining demonstrates the absence of elastic fibers in the reticular dermis in the area surrounded by the granulomatous infiltrate (original magnification $\times 40$). (d) Fragmented elastic fibers are present within some of the histiocytes and multinucleated giant cells (original magnification $\times 400$)

annular elastolytic giant cell granuloma (AEGCG). Thereafter, the patient was treated with topical corticosteroid (clobetasol propionate). This resulted in gradual flattening of the nodules, some of which finally disappeared.

Annular elastolytic giant cell granuloma is an entity that was proposed originally by Hanke *et al.*¹ for cutaneous annular lesions characterized by elastolysis, elastophagocytosis, and an infiltrate of multinucleated giant cells. The pathogenesis of AEGCG remains unclear. It has been postulated that exposure to the sun or other unknown factors alters the antigenicity of the elastic fibers and induces cell-mediated immune reactions.²

Interestingly, in the present case, AEGCG occurred mainly on pre-existing lesions of vitiligo. This anatomical co-localization suggests that the association between the two disorders is not accidental. The pathogenesis of vitiligo is also still unknown, although recently it has been shown that oxidative stress and accumulation of free radicals (FRs) acting as a trigger of melanocyte degeneration in the epidermis of affected skin are involved.³ Oxidative stress can be induced by an increase in the generation of reactive oxygen species (ROS) and other radicals. Recent studies of vitiligo have shown that FRs are increased and that antioxidant systems are deficient. It has been demonstrated both *in vivo* and *in vitro* that patients with vitiligo accumulate high levels of hydrogen peroxide (H_2O_2), which

leads to the destruction of melanocytes in the epidermis.⁴ On the other hand, high superoxide dismutase activity in the serum and skin of patients with stable vitiligo has been reported.^{5,6} These changes would lead to the accumulation of H_2O_2 .

Recent studies of photoaging have revealed that ultraviolet (UV) irradiation induces the formation of ROS in skin tissue. Dermal fibroblasts exposed to ROS show increased expression of mRNA for matrix metalloproteinases (MMP)-1 and MMP-2, which have the ability to degrade collagen and elastic fibers.^{7,8} Igawa *et al.*⁹ have reported that oral Dapsone, which has an anti-oxidative effect, is effective for treatment of AEGCG. In our present patient, annular nodules were located on covered, as well as sun-exposed areas, and the patient had not been treated with UV irradiation. We postulate that in this case, degradation of dermal elastic fibers in the vitiligo lesions may have triggered the accumulation of lymphocytes and macrophages, elastophagocytosis, and subsequent granuloma formation.

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References

- 1 Hanke CW, Bailin PL, Roenigk HH Jr. Annular elastolytic giant cell granuloma. A clinicopathologic study of five cases and a review of similar entities. *J Am Acad Dermatol* 1979; 1: 413-421.
- 2 Ozkaya-Bayazit E, Buyukbabani N, Baykal C, et al. Annular elastolytic giant cell granuloma: sparing of a burn scar and successful treatment with chloroquine. *Br J Dermatol* 1999; 140: 525-530.
- 3 Dell'ana ML, Picardo M. A review and a new hypothesis for non-immunological pathogenetic mechanisms in vitiligo. *Pigment Cell Res* 2006; 19: 406-411.

Spiny keratoderma of the palms in an insulin-treated diabetic patient

Spiny keratoderma is a rare disease characterized by multiple discrete keratotic plugs, resembling a "music box spine", arising from the palms, soles, or both.¹ It was first described in 1971 by Brown², who called it punctate keratoderma. These spiny lesions have been described as filiform hyperkeratosis,³ minute digitate hyperkeratosis,⁴ punctate keratoderma,² punctate porokeratotic keratoderma⁵ and, most recently, spiny keratoderma of the palms and soles.¹ Most of the cases described represent acquired disease, but there are also familial cases. Associations with a systemic disease or malignancy occur in some acquired cases. We present the case of a man with acquired spiny keratoderma who was receiving insulin treatment for type 2 diabetes mellitus.

A 58-year-old Japanese male was referred to us because of multiple asymptomatic, keratotic papules on his fingers and palms. He had a more than 20-year history of diabetes mellitus. He had noted the lesions more than 10 years ago and had started insulin treatment around the same time. Family history was negative for keratoderma, ichthyosis, or other dermatological diseases. He had no history of renal failure or hyperlipidemia and was unaware of any arsenic exposure.

On physical examination, numerous firm keratotic spicules were seen on the volar surface of the palms and

- 4 Schallreuter KU, Moore J, Wood JM, et al. *In vivo* and *in vitro* evidence for hydrogen peroxide (H₂O₂) accumulation in the epidermis of patients with vitiligo and its successful removal by a UVB-activated pseudocatalase. *J Invest Dermatol Symp Proc* 1999; 4: 91-96.
- 5 Ines D, Sonia B, Riadh BM, et al. A comparative study of oxidant-antioxidant status in stable and active vitiligo patients. *Arch Dermatol Res* 2006; 298: 147-152.
- 6 Yildirim M, Baysal V, Inaloz HS, Can M. The role of oxidants and antioxidants in generalized vitiligo at tissue level. *J Eur Acad Dermatol Venereol* 2004; 18: 683-686.
- 7 Kawaguchi Y, Tanaka H, Okada T, et al. The effects of ultraviolet A and reactive oxygen species on the mRNA expression of 72-kDa type IV collagenase and its tissue inhibitor in cultured human dermal fibroblasts. *Arch Dermatol Res* 1996; 288: 39-44.
- 8 Zaw KK, Yokoyama Y, Abe M, Ishikawa O. Catalase restores the altered mRNA expression of collagen and matrix metalloproteinases by dermal fibroblasts exposed to reactive oxygen species. *Eur J Dermatol* 2006; 16: 375-379.
- 9 Igawa K, Maruyama R, Katayama I, Nishioka K. Anti-oxidative therapy with oral dapsone improved HCV antibody-positive annular elastolytic giant cell granuloma. *J Dermatol* 1997; 24: 328-331.

fingers (Fig. 1). The lesions were 0.5-1 mm in diameter, 1-2 mm in length, and skin-colored. There were no similar lesions at other sites, including the soles.

Histopathological examination of skin biopsy specimens of the palms showed large columns of keratin arising from the epidermis in an area with a focally decreased granular layer and parakeratosis (Fig. 2). No dyskeratotic or vacuolated keratinocytes were seen in the underlying epidermis. Blood tests and computer tomography scan showed no abnormalities. Results of upper and lower endoscopy did not reveal any malignant neoplasms.

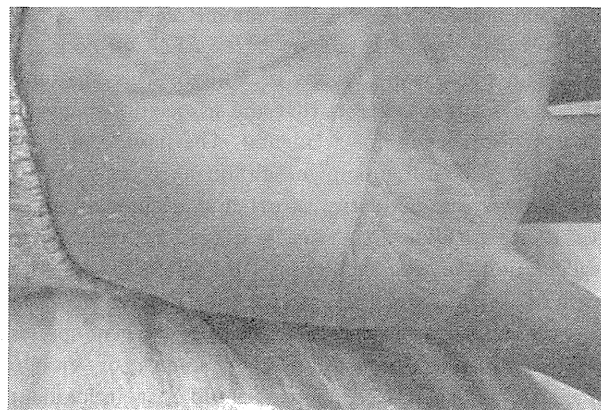


Figure 1 Multiple yellowish filiform keratotic projections on the left palm

in the antimelanogenic action down-regulating the expression of MITF or tyrosinase gene in cAMP-elevated melanocyte cultures and UV-irradiated dorsal skins of guinea pigs (Supplementary Figure S12 online). Finally, this study suggests a potential application of 4H3MC in the treatment of hyperpigmented skin disorders.

Animal experiments were carried out according to the protocols approved by Animal Experimentation Ethics Committee in CBNU institute.

CONFLICT OF INTEREST

The authors state no conflict of interest.

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SUPPLEMENTARY MATERIAL

Supplementary material is linked to the online version of the paper at <http://www.nature.com/jid>

REFERENCES

- Busca R, Ballotti R (2000) Cyclic AMP a key messenger in the regulation of skin pigmentation. *Pigment Cell Res* 13:60–9
- Hummeler E, Cole TJ, Blendy JA et al. (1994) Targeted mutation of the CREB gene: compensation within the CREB/ATF family of transcription factors. *Proc Natl Acad Sci USA* 91:5647–51
- Kim C, Cheng CY, Saldanha SA et al. (2007) PKA-I holoenzyme structure reveals a mechanism for cAMP-dependent activation. *Cell* 130:1032–43
- Lee HS (2002) Tyrosinase inhibitors of *Pulsatilla cernua* root-derived materials. *J Agric Food Chem* 50:1400–3
- Lochner A, Moolman JA (2006) The many faces of H89: a review. *Cardiovasc Drug Rev* 24:261–74
- Maeda K, Fukuda M (1996) Arbutin: mechanism of its depigmenting action in human melanocyte culture. *J Pharmacol Exp Ther* 276:765–9

- Moll D, Prinz A, Brendel CM et al. (2008) Biochemical characterization and cellular imaging of a novel, membrane permeable fluorescent cAMP analog. *BMC Biochem* 9:18
- Ortonne JP, Passeron T (2005) Melanin pigmentary disorders: treatment update. *Dermatol Clin* 23:209–26
- Roh E, Yun CY, Young Yun J et al. (2013) cAMP-binding site of PKA as a molecular target of bisabolangelone against melanocyte-specific hyperpigmented disorder. *J Invest Dermatol* 133:1072–9
- Takasao N, Tsuji-Naito K, Ishikura S et al. (2012) Cinnamon extract promotes type I collagen biosynthesis via activation of IGF-I signaling in human dermal fibroblasts. *J Agric Food Chem* 60:1193–200
- Taylor SS, Kim C, Cheng CY et al. (2008) Signaling through cAMP and cAMP-dependent protein kinase: diverse strategies for drug design. *Biochim Biophys Acta* 1784:16–26
- Vachtenheim J, Borovansky J (2010) "Transcription physiology" of pigment formation in melanocytes: central role of MITF. *Exp Dermatol* 19:617–27
- Wu J, Jones JM, Nguyen-Huu X et al. (2004) Crystal structures of R1 α subunit of cyclic adenosine 5'-monophosphate (cAMP)-dependent protein kinase complexed with (Rp)-adenosine 3',5'-cyclic monophosphothioate and (Sp)-adenosine 3',5'-cyclic monophosphothioate, the phosphothioate analogues of cAMP. *Biochemistry (Mosc)*. 43:6620–9

A Somatic Mutation of the *KEAP1* Gene in Malignant Melanoma Is Involved in Aberrant NRF2 Activation and an Increase in Intrinsic Drug Resistance

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TO THE EDITOR

Among the several characteristics of malignant melanoma, insensitivity to anti-cancer agents is a frequent clinical problem in the treatment of patients (Grossman and Altieri, 2001). In fact, the most commonly used chemotherapy agents cisplatin and dacarbazine for malignant melanoma elicit a response rate of only 10% (Flaherty, 2010).

The small-molecule inhibitor of BRAF, vemurafenib (also known as

PLX4032), elicits potent tumor regression in patients with BRAF-positive stage IV melanoma (Huang et al., 2012), and its use is expected in patients harboring the *BRAF*^{V600E} mutation (Tsai et al., 2008). However, vemurafenib is not effective against melanomas with wild-type BRAF protein (Joseph et al., 2010).

Acral lentiginous melanoma (ALM) is one of the subtypes of cutaneous melanoma most frequent in colored races (Bradford et al., 2009). In fact, only

about 10% of ALM cases harbor the *BRAF*^{V600E} mutation, compared with over 60% of cases of superficial spreading melanoma (SSM), which is most frequent in Caucasian populations (Saldanha et al., 2006). Because these melanomas are insensitive to BRAF inhibitors (Joseph et al., 2010), a search for molecular targets that would enhance sensitivity to standard treatment with cisplatin or dacarbazine would seem justified.

To address the genes responsible for drug resistance in melanoma, whole-exome sequencing was performed. We identified a single-nucleotide deletion in codon 507 from exon 4 of the *KEAP1* gene, common to MM-RU and PM-WK, as a candidate gene for drug resistance

Abbreviations: ALM, acral lentiginous melanoma; CDDP, cis-diamminedichloro-platinum (III); DTIC, 5-(3,3-dimethyl-1-triazenyl) imidazole-4-carboxamide; FSM, frameshift mutant; ROS, reactive oxygen species; SSM, superficial spreading melanoma

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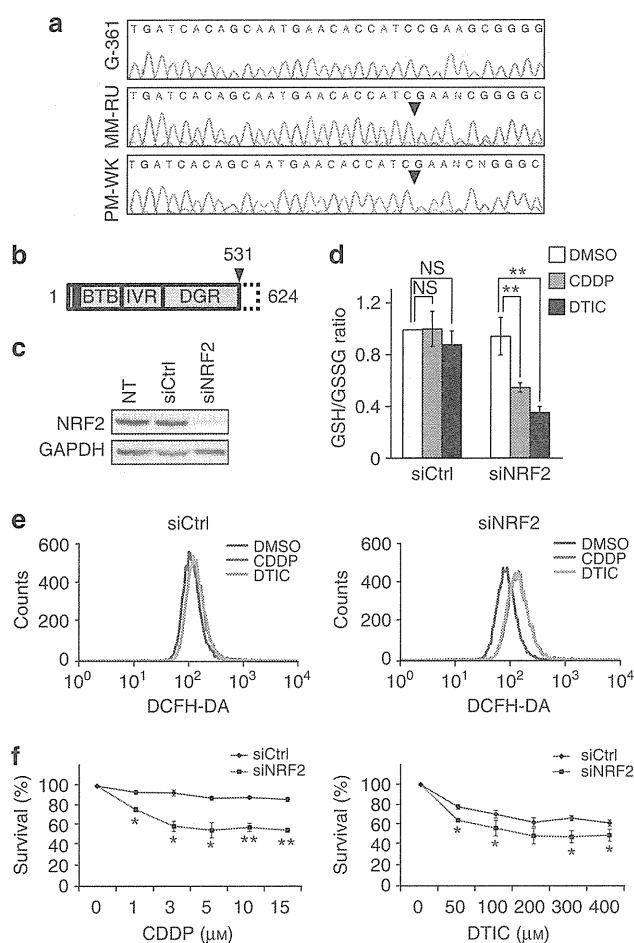


Figure 1. NRF2 contributes to chemoresistance in KEAP1-F5M cells. (a) Representative electropherograms of *KEAP1* gene exon 4 by capillary sequencing in melanoma cell lines. Arrowheads indicate the single-nucleotide deletion at codon 507 (1518delC). (b) Schematic representation of the *KEAP1* protein. Arrowheads indicate the appearance of a stop codon. (c) Expression of NRF2 protein in siCtrl- or siNRF2-transfected PM-WK cells. (d) Quantification of the ratio of reduced-to-oxidized glutathione (GSH/GSSG) or (e) intracellular reactive oxygen species levels after treatment with vehicle, CDDP, or DTIC in siCtrl- or siNRF2-transfected PM-WK cells. (f) Cell viability at 24 hours after treatment with vehicle, CDDP, or DTIC in siCtrl- or siNRF2-transfected PM-WK cells. NS, no significant difference; * $P < 0.05$ and ** $P < 0.01$ versus control by *t*-test. CDDP, *cis*-diamminedichloro-platinum (II); DTIC, 5-(3, 3-dimethyl-1-triazenyl) imidazole-4-carboxamide.

(Figure 1a). The KEAP1 protein produced by this mutant allele partially lacks the DGR/Kelch domain that is important for interacting with NRF2, a key transcriptional regulator of oxidative stress such as that resulting from reactive oxygen species (ROS; Figure 1b). Under normal conditions, NRF2 is kept transcriptionally inactive through binding to the DGR domains of KEAP1 and constitutively degraded by the ubiquitin proteasome system (Sekhar *et al.*, 2002). In contrast, in the cell lines harboring the *KEAP1* frameshift mutation (KEAP1-F5M), constitutive NRF2 stabilization, nuclear localization (Supple-

mentary Figure S1 online), and target-gene activation were observed (Supplementary Figure S2 online). We also observed strong expression of the NRF2 protein in HMVI cells, which do not harbor *KEAP1* mutation, suggesting that another signaling pathway, such as the PI3K pathway, may regulate the constitutive expression of NRF2 in HMVI cells (Mitsuishi *et al.*, 2012).

Thus far, several somatic mutations of *KEAP1* that affect its NRF2-inhibitory activity have been identified in patients with cancers of the lungs, gallbladder, and liver (Taguchi *et al.*, 2011). Aberrant activation of NRF2 induced by *KEAP1*

gene mutation in gallbladder cancer has been reported to lead to 5-fluorouracil (5-FU) resistance (Shibata *et al.*, 2008). However, the relationship between the KEAP1-NRF2 pathway and drug resistance in malignant melanoma still remains to be elucidated.

Cisplatin (*cis*-diamminedichloro-platinum (II), CDDP) and dacarbazine (5-(3, 3-dimethyl-1-triazenyl) imidazole-4-carboxamide, DTIC) have been shown to increase oxidative stress by raising the levels of intracellular ROS such as H_2O_2 (Zhang *et al.*, 2010; Deavall *et al.*, 2012). Therefore, we investigated whether NRF2 is involved in sensitivity to these drugs. The ratio of reduced-to-oxidized glutathione (GSH/GSSG) was decreased by treatment with CDDP or DTIC in siNRF2-transfected PM-WK cells (Figure 1c and d). Furthermore, intracellular ROS levels were increased by treatment with CDDP or DTIC in siNRF2-transfected cells, suggesting that the H_2O_2 detoxification pathway is active in these cell lines (Figure 1e; Supplementary Figure S3a online). Notably, knockdown of *NRF2* by small interfering RNA in PM-WK and MM-RU cells significantly enhanced their sensitivity to apoptosis by CDDP or DTIC (Figure 1f; Supplementary Figures S3b and S4 online). Conversely, the overexpression of NRF2 or activation of NRF2 by sulforaphane in G-361 cells reduced their sensitivity to CDDP or DTIC (Supplementary Figure S5 online). Moreover, the NRF2-positive cell lines tended to show higher IC_{50} values compared with NRF2-negative cell lines (Supplementary Figure S6 online). Taken together, the data indicate that, in addition to the mechanisms of drug resistance in melanoma, such as drug trapping by melanosomes and elevated expression of ATP-dependent transporters (Gottesman *et al.*, 2002; Chen *et al.*, 2006), aberrantly expressed NRF2 is one of the causes of resistance to CDDP and DTIC.

Next, we examined *KEAP1* gene mutation and NRF2 expression in specimens of primary melanoma. The histological types, patient genders and ages, and the stages of the primary melanomas we examined are shown in Supplementary Table online. Ten (48%), eight (38%), two (10%), and one (4%) of the

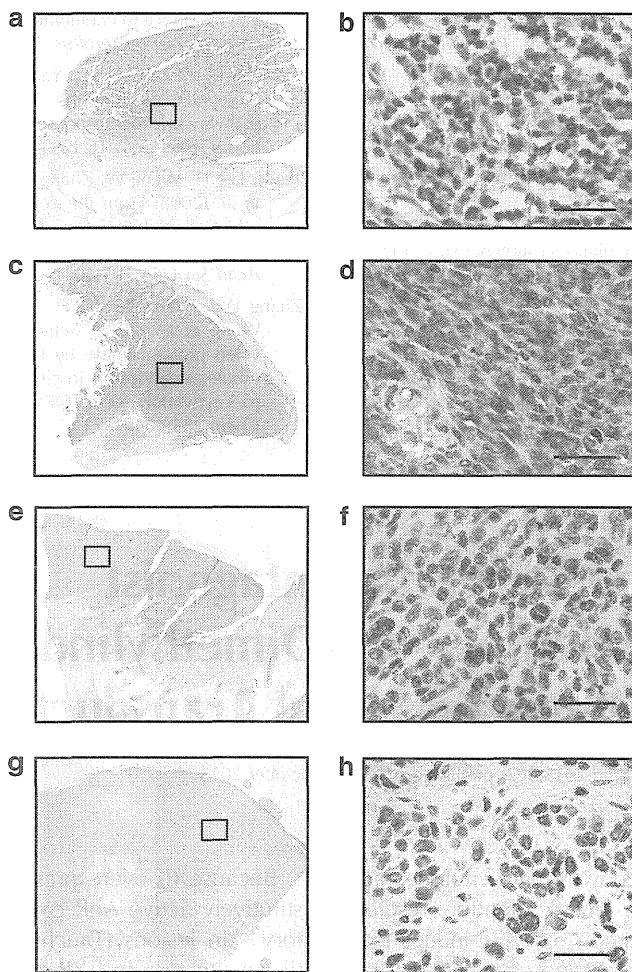


Figure 2. Aberrant NRF2 protein accumulation in primary melanomas harboring *KEAP1* frameshift mutation (FSM). Immunohistochemical analysis of endogenous NRF2 protein expression in melanomas harboring the *KEAP1* FSM (a and b, #02-2078; c and d, #03-8008) and with wild-type *KEAP1* (e and f, #09-7356; g and h, #07-7252). Cells were stained with NRF2 antibody. High-magnification images correspond to the respective boxed areas in the loupe images. Bar = 100 μm.

21 cases were ALM, NM, LMM, and SSM, respectively. We performed direct sequencing of *KEAP1* gene exon 4 using the genomic DNAs extracted from these paraffin-embedded specimens. In two of the 21 cases (~10%), we identified a homozygous single-nucleotide deletion that was the same as that identified in melanoma cell lines (#02-2078, 1518delC) and a heterozygous single-nucleotide deletion in a neighboring position (#03-8008, 1519delG) (Supplementary Table online). The stop codon appeared 23 nucleotides after the 1519delG mutation and also produced the same type of truncated *KEAP1* protein as that resulting from 1518delC mutation. Immunohistochemical staining revealed that NRF2 was positive in

specimens #02-2078 and #03-8008 (Figure 2b and d, respectively), which were ALM specimens harboring *KEAP1*-FSM mutation, whereas NRF2 was negative in specimens #09-7356 and #07-7252, which were ALM specimen without *KEAP1* mutation (Figure 2, Supplementary Table online). We also performed direct sequencing of *BRAF* gene exon 15 using DNAs from #02-2078, #03-8008, #08-6115, and #09-7356, but the typical *BRAF*^{V600E} mutation was not identified (Supplementary Table online). Taking these results together, it appears that the loss of function of *KEAP1* caused by single-nucleotide deletions is indeed correlated with aberrant NRF2 expression in primary melanoma.

Missense mutations of the *KEAP1* gene have been identified in various cancers. Interestingly, the *KEAP1*-FSMs are most frequently identified in the DGR domain (65%), which is important for interaction with NRF2 (Taguchi et al., 2011). Our results also indicate that the DGR domain of the *KEAP1* gene may be a “hot-spot” for FSM that causes aberrant activation of NRF2 and causes melanomas to become more resistant to CDDP or DTIC.

In conclusion, our present results and available data fully support the possibility that decreased expression or inhibition of NRF2 may provide an avenue for treatment of patients with malignant melanoma, by improving the rate of response to the standard chemotherapy agents, namely, cisplatin and dacarbazine.

CONFLICT OF INTEREST

The authors state no conflict of interest.

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SUPPLEMENTARY MATERIAL

Supplementary material is linked to the online version of the paper at <http://www.nature.com/jid>

REFERENCES

- Bradford PT, Goldstein AM, McMaster ML, Tucker MA (2009) Acral lentiginous melanoma: incidence and survival patterns in the United States, 1986-2005. *Arch Dermatol* 145: 427-34
- Chen KG, Valencia JC, Lai B, Zhang G, Paterson JK, Rouzaud F et al. (2006) Melanosomal

- sequestration of cytotoxic drugs contributes to the intractability of malignant melanomas. *Proc Natl Acad Sci USA* 103:9903–7
- Deavall DG, Martin EA, Horner JM, Roberts R (2012) Drug-induced oxidative stress and toxicity. *J Toxicol* 2012:1–13
- Flaherty KT (2010) Narrative review: BRAF opens the door for therapeutic advances in melanoma. *Ann Intern Med* 153:587–91
- Gottesman MM, Fojo T, Bates SE (2002) Multidrug resistance in cancer: role of Atp-dependent transporters. *Nat Rev Cancer* 2:48–58
- Grossman D, Altieri DC (2001) Drug resistance in melanoma: mechanisms, apoptosis, and new potential therapeutic targets. *Cancer Metastasis Rev* 20:3–11
- Huang V, Hepper D, Anadkat M, Cornelius L (2012) Cutaneous toxic effects associated with vemurafenib and inhibition of the BRAF pathway. *Arch Dermatol* 148:628–33
- Joseph EW, Pratilas CA, Poulidakos PI, Tadi M, Wang W, Taylor BS et al. (2010) The RAF inhibitor PLX4032 inhibits ERK signaling and tumor cell proliferation in a V600E BRAF-selective manner. *Proc Natl Acad Sci USA* 107:14903–8
- Mitsuishi Y, Taguchi K, Kawatani Y, Shibata T, Nukiwa T, Aburatani H et al. (2012) Nrf2 redirects glucose and glutamine into anabolic pathways in metabolic reprogramming. *Cancer Cell* 22:66–79
- Saldanha G, Potter L, DaForno P, Pringle JH (2006) Cutaneous melanoma subtypes show different BRAF and NRAS mutation frequencies. *Clin Cancer Res* 12:4499–505
- Sekhar KR, Yan XX, Freeman ML (2002) Nrf2 degradation by the ubiquitin proteasome pathway is inhibited by KIAA0132, the human homolog to INrf2. *Oncogene* 21:6829–34
- Shibata T, Kokubu A, Gotoh M, Ojima H, Ohta T, Yamamoto M et al. (2008) Genetic alteration of Keap1 confers constitutive Nrf2 activation and resistance to chemotherapy in gallbladder cancer. *Gastroenterology* 135(1358-68):e4
- Taguchi K, Motohashi H, Yamamoto M (2011) Molecular mechanisms of the Keap1-Nrf2 pathway in stress response and cancer evolution. *Genes Cells* 16:123–40
- Tsai J, Lee JT, Wang W, Zhang J, Cho H, Mamo S et al. (2008) From the cover: discovery of a selective inhibitor of oncogenic B-Raf kinase with potent antimelanoma activity. *Proc Natl Acad Sci USA* 105:3041–6
- Zhang WB, Wang Z, Shu F, YH Jin, HY Liu, QJ Wang et al. (2010) Activation of AMP-activated protein kinase by temozolomide contributes to apoptosis in glioblastoma cells via p53 activation and mTORC1 inhibition. *J Biol Chem* 285:40461–71

The New Aryl Hydrocarbon Receptor Antagonist E/Z-2-Benzylindene-5,6-Dimethoxy-3,3-Dimethylindan-1-One Protects against UVB-Induced Signal Transduction

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TO THE EDITOR

The aryl hydrocarbon receptor (AhR) is a ligand-activated transcription factor that mediates the toxicity of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD), polycyclic aromatic hydrocarbons, and related environmental contaminants (Abel and Haarmann-Stemmann, 2010). The unligated AhR is trapped in a cytosolic multiprotein complex, which rapidly dissociates upon ligand binding. Subsequently, the AhR shuttles into the nucleus, dimerizes with ARNT, and binds to xenobiotic-responsive elements (XREs) in the promoter of target genes, e.g., encoding cytochrome P450 (CYP) 1 monooxygenases, to enforce transcription (Abel and Haarmann-Stemmann, 2010). Furthermore, AhR-triggered activation of c-src tyrosine kinase stimulates

EGFR and downstream mitogen-activated protein kinase signaling, resulting in the induction of XRE-independent genes, such as cyclooxygenase-2 (COX-2; Abel and Haarmann-Stemmann, 2010). We have previously shown that the AhR in keratinocytes is not only activated by anthropogenic chemicals but also by UVB irradiation, which leads to the intracellular formation of the tryptophan photoproduct and high-affinity AhR ligand 6-formylindolo[3,2-*b*]carbazole (FICZ; Rannug et al., 1995; Fritsche et al., 2007). Indeed, UVB exposure enhances AhR/XRE binding (Supplementary Figure 1 online) and accompanied CYP1A1/1B1 expression (Katiyar et al., 2000), as well as XRE-independent COX-2 expression (Fritsche et al., 2007).

Because (i) overexpression of a constitutively active AhR causes inflammatory skin lesions (Tauchi et al., 2005), (ii) an increase in CYP activity leads to reactive oxygen species formation (Puntarulo and Cederbaum, 1998), (iii) CYP1 enzymes are critical for chemical-induced skin carcinogenesis (Shimizu et al., 2000), and (iv) COX-2 is involved in UV-induced inflammation and carcinogenesis (Elmets et al., 2010), it was postulated that a transient inhibition of AhR may protect human skin against the detrimental effects of UVB irradiation (Agostinis et al., 2007; Haarmann-Stemmann et al., 2012). Moreover, we have shown that the expression of matrix metalloproteinase-1 (MMP-1), which is critically involved in extrinsic skin aging, is upregulated in an AhR-dependent manner in tobacco smoke extract-exposed keratinocytes (Ono et al., 2013). Therefore, we decided to develop an AHR antagonist that is suitable for topical UV-protection. We screened a library of compounds that possess the structural prerequisites to

Abbreviations: AhR, aryl hydrocarbon receptor; BDDI, E/Z-2-benzylidene-5,6-dimethoxy-3,3-dimethylindan-1-one; COX-2, cyclooxygenase-2; CYP, cytochrome P450; EROD, 7-O-ethoxyresorufin-deethylase; FICZ, 6-formylindolo[3,2-*b*]carbazole; MMP-1, matrix metalloproteinase-1; MNF, 3'-methoxy-4'-nitroflavone; NHEK, normal human epidermal keratinocyte; TCDD, 2,3,7,8-tetrachlorodibenzo-*p*-dioxin; XRE, xenobiotic-responsive element

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Case Study

Subcutaneous Panniculitis-Like T-Cell Lymphoma (SPTCL) with Hemophagocytosis (HPS) : Successful Treatment Using High-Dose Chemotherapy (BFM-NHL & ALL-90) and Autologous Peripheral Blood Stem Cell Transplantation

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Kanako Tsunoda,^{1,2)} Mikiya Endo,³⁾ Toshihide Akasaka,¹⁾ and Tomoyuki Masuda²⁾

Subcutaneous panniculitis-like T-cell lymphoma (SPTCL) is a rare form of non-Hodgkin lymphoma, in which lymphoma cells infiltrate preferentially into subcutaneous adipose tissue. Although various treatment trials for SPTCL have been attempted, no standardized therapy has been established. Here, we report a case of α/β^+ T-cell-phenotype SPTCL (SPTCL-AB) with hemophagocytosis (HPS) in a 14-year-old girl, who presented with low-grade fever, general fatigue and chest swelling. Laboratory examinations revealed leukocytopenia, and bone marrow aspiration cytology showed HPS. The diagnosis of SPTCL-AB was made by biopsy on the basis of thickened subcutaneous tissue in the chest wall. Following high-dose chemotherapy (HDT) of BFM-NHL & ALL-90, autologous peripheral blood stem cell transplantation (auto-PBSCT) was performed. The patient responded to the treatment and has remained asymptomatic for 2 years. Our results suggest that a combination of HDT of BFM-NHL & ALL-90 and auto-SCT treatment is effective for SPTCL associated with HPS. [*J Clin Exp Hematop* 53(2) : 135-140, 2013]

Keywords: SPTCL, hemophagocytosis, BFM-NHL & ALL-90, auto-PBSCT

INTRODUCTION

In 1991, the distinct clinicopathological features of a T-cell lymphoma in which the lymphoma cells preferentially invade the subcutaneous tissue were described by Gonzalez *et al.*¹ Under the term subcutaneous panniculitis-like T-cell lymphoma (SPTCL), this new condition was established as a distinct disease entity in the World Health Organization (WHO) classification.² Because of its peculiar pathological features, SPTCL may be initially misdiagnosed as Weber-Christian disease, a benign inflammatory panniculitis and a granulomatous disease.^{3,4} Recent studies have disclosed that cases with an α/β^+ T-cell phenotype (SPTCL-AB) and a γ/δ^+ T-cell phenotype (SPTCL-GD) can be distinguished within

the group of SPTCL.⁵ SPTCL-ABs have a CD4⁻, CD8⁺, CD56⁻ phenotype, and SPTCL-GDs have a CD4⁻, CD8⁻ phenotype with frequent expression of CD56. Compared with SPTCL-ABs, SPTCL-GDs have a poor prognosis.^{6,7} On the basis of these observations, the term SPTCL is used only for SPTCL-ABs, and SPTCL-GDs are included within the cutaneous γ/δ^+ T-cell lymphomas.^{6,8} Although SPTCL-AB patients without hemophagocytic syndrome (HPS) have a favorable prognosis, the clinical course of cases associated with HPS is generally aggressive, and a delay in diagnosis or treatment may result in a fatal outcome. There have been few reports of successful treatment of SPTCL-AB patients with HPS, and no therapeutic regimen has been established.

Here, we report a case of SPTCL-AB with HPS that was treated successfully with a combination of high-dose chemotherapy (HDT) of Berlin-Frankfurt-Münster-non-Hodgkin lymphoma-90 (BFM-NHL-90) and autologous peripheral blood stem cell transplantation (auto-PBSCT).

CASE REPORT

A previously healthy 14-year-old Japanese girl visited a physician because of a 1-month history of chest swelling.

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She also had a 3-month history of general fatigue and low-grade fever. Initial laboratory examinations revealed a white blood cell count of $2.82 \times 10^3/\mu\text{L}$, hemoglobin of 11.6 g/dL, hematocrit of 33.8 g/dL and platelet count of $16.4 \times 10^4/\mu\text{L}$. Histopathologic examination of a biopsy specimen of subcutaneous adipose tissue from the chest wall revealed diffuse infiltration of medium-sized lymphocytes. Under a diagnosis of panniculitis due to lupus profundus, administration of low-dose prednisolone (PSL) was started. Several weeks later, the chest swelling expanded gradually. She was therefore referred and admitted to our hospital for further evaluation and treatment.

On physical examination, a massive tumor was evident in her chest subcutaneous tissue (Fig. 1). The findings from laboratory examinations were as follows: aspartate aminotransferase 27 IU/L (normal: 10-40 IU/L), alanine aminotransferase 15 IU/L (normal: 5-35 IU/L), lactic dehydrogenase 235 U/L (normal: 115-359 IU/L), soluble interleukin-2 receptor 641 U/mL (normal: 220-530 U/mL) and ferritin 56 ng/mL. Moreover, previous infection with Epstein-Barr virus was evident. Magnetic resonance imaging revealed noticeable thickening of the subcutaneous tissue, compatible with the region of the massive tumor in the chest (Fig. 2). Histopathology of skin biopsy specimens showed medium-sized lymphocytes diffusely infiltrating into the subcutaneous fat tissue (Fig. 3a). Rimming of the lymphocytes around individual fat cells was observed (Fig. 3b).

Immunohistochemical staining revealed that the infiltrating lymphocytes were CD3⁺, CD4⁻, CD5⁺, CD8⁺, CD20⁻, CD30⁻, CD45RO⁺, CD56⁻ and CD79a⁻ (Fig. 4). They were positive for T-cell receptor- β (Fig. 5a). Cytotoxic molecules such as granzyme B, T-cell intracellular antigen-1 (TIA-1) and perforin were positive (Fig. 5b). Latent membrane protein 1 (LMP-1) and EBER *in situ* hybridization were negative (Fig. 5c & 5d). Bone marrow smears showed hemophagocytosis.

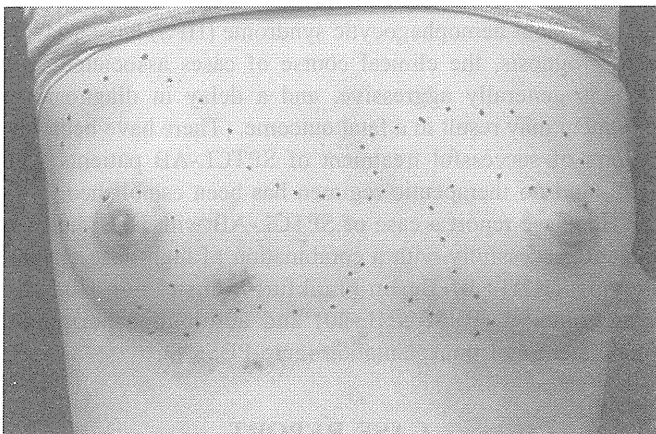


Fig. 1. A massive tumor, indicated by black dots, was present in the subcutaneous tissue of the chest.

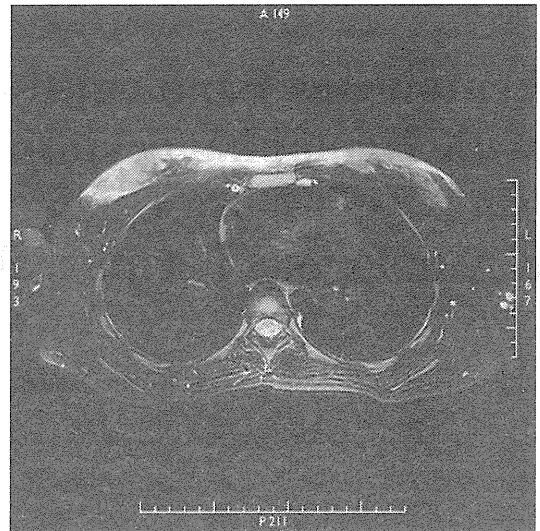


Fig. 2. Magnetic resonance imaging of the chest (T2-weighted). The subcutaneous tumor of the chest wall showed high signal intensity.

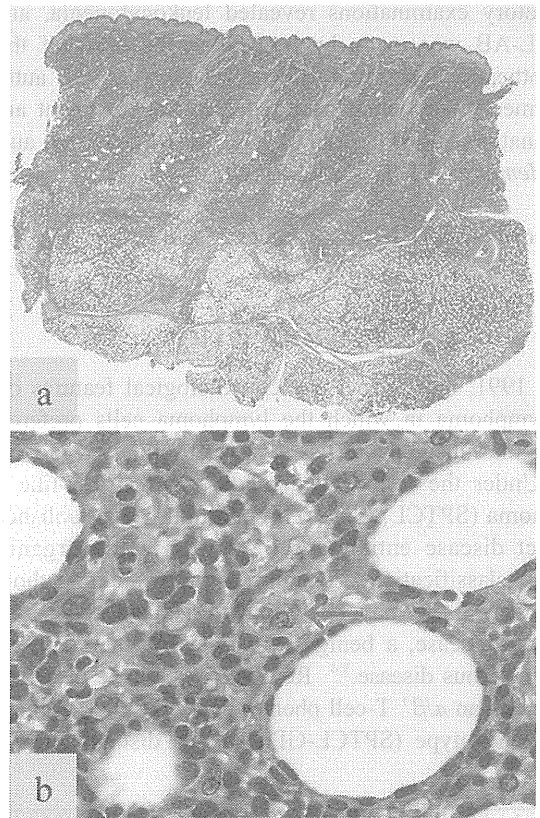


Fig. 3. Histopathology of the skin biopsy specimen (H&E stain). (3a) Diffuse cellular infiltration occurred in the subcutaneous fat tissue. (3b) Rimming of lymphocytes around fat cells was observed (arrow). Bean bag cells, phagocytosing erythrocytes, were found among the lymphocytes. (3a) $\times 10$, (3b) $\times 400$.

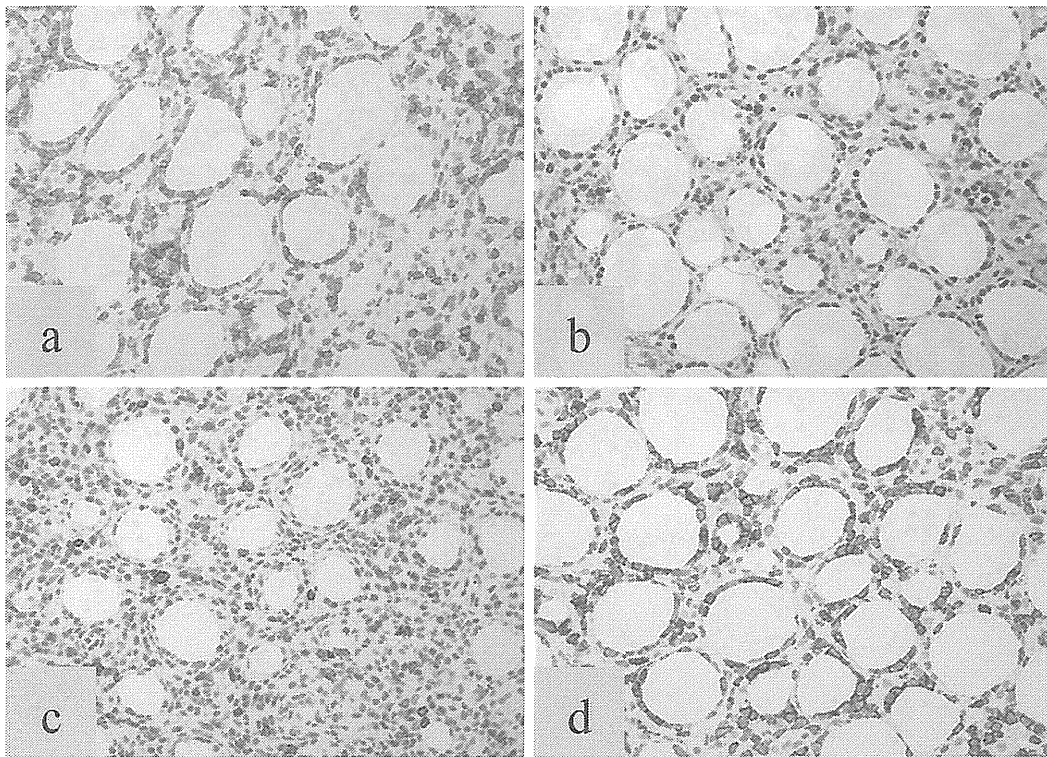


Fig. 4. Immunohistochemistry of CD3 (4a), CD4 (4b), CD5 (4c) and CD8 (4d). The infiltrating lymphocytes were positive for CD3, CD5 and CD8, but negative for CD4. $\times 400$.

tosis of red blood cells and neutrophils by histiocytes (Fig. 6).

On the basis of these results, the patient was diagnosed as having SPTCL accompanied by HPS in spite of the low levels of lactic dehydrogenase and ferritin, and was treated with high-dose PSL (75 mg/day internally), starting on day 9 of admission.

Nine days later (on day 17 of admission), despite the PSL therapy, the chest tumor was unchanged in size. Additional HDT of BFM-NHL-90 (protocol I) was therefore applied. After starting the first course of the chemotherapy, the tumor in the chest gradually decreased in size. During the phase of recovery from the chemotherapy, peripheral blood stem cells (CD34⁺ cells) were harvested. Moreover, chemotherapy of BFM-NHL-90 (Protocol M) was applied as a consolidation therapy. In addition, one course of the BFM-acute lymphoblastic leukemia (ALL)-90 regimen (HR3) was applied for further treatment.

Following pretreatment with the MCVAC (ranimustine, cytarabine, etoposide and cyclophosphamide) regimen, the patient was treated with auto-PBSCT. The clinical course thereafter was uneventful. She has been in complete remission for more than 2 years, and there was no evidence of local recurrence in the chest wall and HPS in the bone marrow at the final follow-up examination.

DISCUSSION

SPTCL is a type of skin lymphoma characterized by the infiltration of subcutaneous tissue by pleomorphic T cells and benign macrophages, mimicking lobular panniculitis. This malignancy typically presents in the form of skin nodules that involve the extremities and can become ulcerated.⁹ The clinical course associated with HPS is aggressive, and a delay in diagnosis and treatment may result in a fatal outcome.

Although the mechanism of HPS in SPTCL has not been clarified, the phenomenon of HPS results from overproduction of cytokines, including interferon- γ , interleukin-2 (IL-2), IL-6, IL-12, IL-18 and tumor necrosis factor- α , produced by activated T cells (Th1 cells) and macrophages, which leads to a chain reaction of cytokines.¹⁰⁻¹³ Control of HPS in SPTCL-AB patients improves the prognosis.

Various therapies such as radiotherapy, PSL, CHOP (cyclophosphamide, hydroxydaunorubicin, vincristine and prednisolone) (-like) chemotherapy and auto/allo-SCT have been applied for SPTCL-AB.⁹ For relapsed or refractory disease, various regimens have been attempted as salvage chemotherapy, including cladribine, DHAP (dexamethasone, cytarabine and cisplatin), ESHAP (etoposide, methylprednisolone, cytarabine and cisplatin), FLAG (fludarabine, cytarabine and granulocyte-colony stimulating factor), mini-BEAM (carmus-

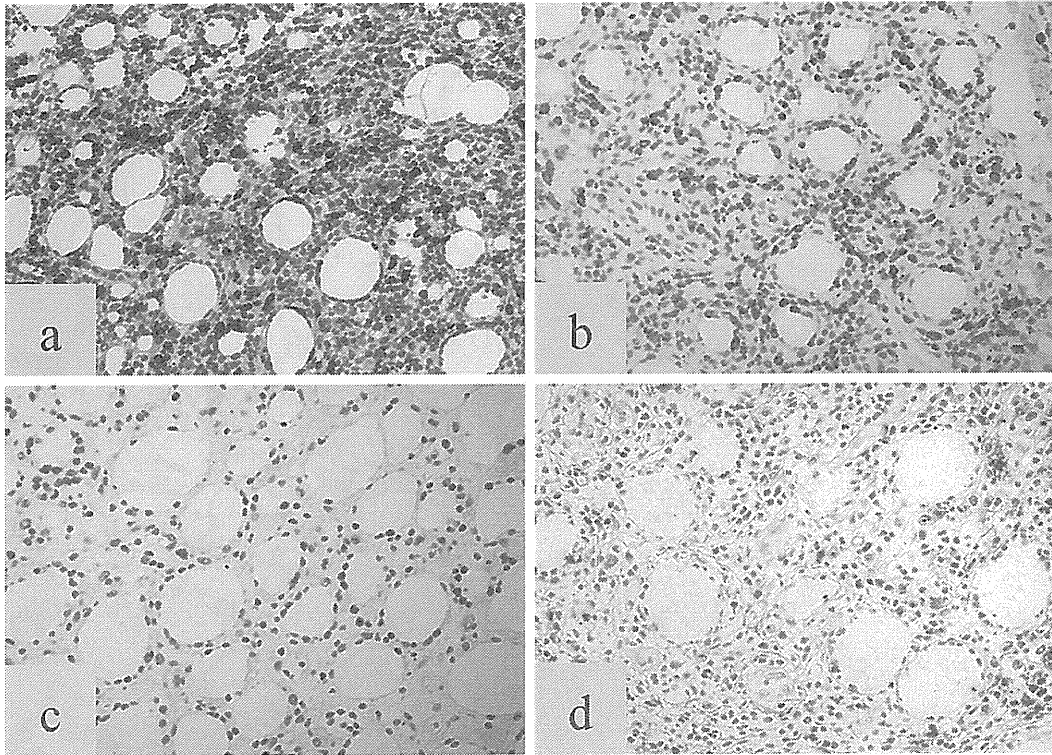


Fig. 5. Immunohistochemistry for T-cell receptor (TCR)- β (5a), granzyme B (5b), latent membrane protein-1 (LMP-1, 5c) and Epstein-Barr virus (EBV)-encoded RNA *in situ* hybridization (5d). Lymphocytes were positive for TCR- β , and expression of the cytotoxic molecule granzyme B was also found. LMP-1 and EBV were negative. $\times 200$.

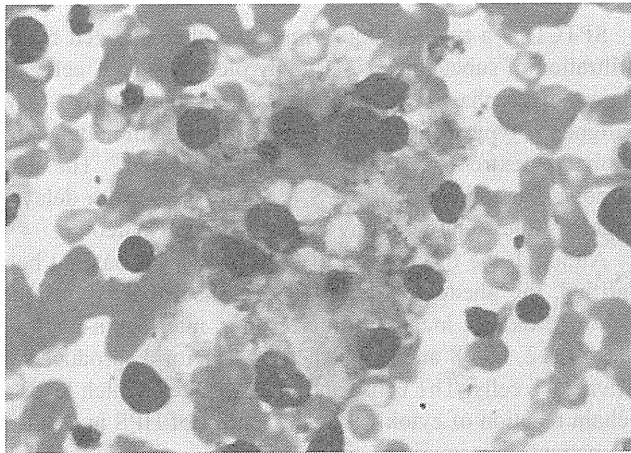


Fig. 6. Hemophagocytosis was seen in a smear of the bone marrow. Giemsa stain, $\times 1000$.

tine, etoposide, cytarabine and melphalan) and VEPPB (vincristine, etoposide, prednisone, pocarbazine and bleomycin).⁹ However, no standardized treatment has yet emerged. In SPTCL-AB patients with HPS, a recent report has indicated that CHOP (-like) chemotherapy is not very effective. Auto-SCT or allo-SCT following HDT has been suggested as an

important option for patients with refractory or recurrent SPTCL.^{4,14}

In the present case, high-dose PSL was initially used, but no response was obtained. Therefore, we treated the patient according to the BFM-NHL & ALL-90 protocol.

BFM-NHL-90 is a protocol for pediatric malignant lymphoma and yields a significantly better outcome for non-Hodgkin lymphoma at stage I or II.¹⁵ BFM-NHL-90 is also effective for pediatric anaplastic large-cell lymphoma. BFM-ALL-90 is a superior regimen for high-risk childhood T-cell acute lymphoblastic leukemia.¹⁶

The present case was treated successfully with a combination of HDT of BFM-NHL & ALL-90 followed by auto-PBSCT, and achieved clinical complete remission (CR). This result suggests that the BFM protocol is applicable and can yield complete remission in cases of SPTCL with HPS. Medhi *et al.* have also reported the value of the BFM-90 protocol for the treatment of patients with SPTCL and HPS.¹⁷

To date, there have been few reports of effective treatment for SPTCL using HDT following SCT. In almost all of the reported cases, the patients underwent HDT-allo-SCT and its effectiveness was impressive, 92% achieving CR, with a median response duration of ≥ 14 months.⁹ In intermediate- and high-grade lymphomas, myeloablative allo-SCT is associated

with a lower relapse rate than auto-SCT for the graft-versus-leukemia effect.^{18,19} However, chronic graft-versus-host disease (cGVHD) is a very common complication, with a reported incidence of between 40% and 70%,²⁰ and is the leading cause of late death in allo-SCT survivors.^{20,21}

The median age of patients with SPTCL-AB at diagnosis is 36 years (range: 9-79 years), and about 19% are in the second decade or younger.⁸ In children, cGVHD may reduce the quality of life because of the induction of growth irregularity. Mukai *et al.* described a patient with SPTCL and HPS who received HDT of BFM-NHL & ALL-90 and auto-SCT.⁴ The present case received a combination of HDT of BFM-NHL & ALL-90 and auto-SCT, and has been in complete remission for more than 2 years. This suggests that auto-SCT might be a feasible option following HDT.

In summary, we have reported a case of SPTCL complicated by HPS, which responded to treatment with HDT of BFM-NHL & ALL-90 and auto-SCT. Although the value of the BFM-NHL & ALL-90 protocol has to be further evaluated in SPTCL cases, our findings suggest that a combination of HDT of BFM-NHL & ALL-90 and auto-SCT is applicable for the treatment of SPTCL with HPS.

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DISCLOSURE/CONFLICT OF INTEREST

The authors state that they have no financial interest in the products mentioned within this article.

REFERENCES

- Gonzalez CL, Medeiros LJ, Brazier RM, Jaffe ES: T-cell lymphoma involving subcutaneous tissue. A clinicopathologic entity commonly associated with hemophagocytic syndrome. *Am J Surg Pathol* 15:17-27, 1991
- Jaffe ES, Gaulard P, Ralfkiaer E, Cerroni L, Meijer CJLM: Subcutaneous panniculitis-like T-cell lymphoma. In: Swerdlow SH, Campo E, Harris NL, Jaffe ES, Pileri SA, *et al.* (eds): *World Health Organization Classification of Tumours, WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues*. 4th ed, Lyon, International Agency for Research on Cancer (IARC), pp.294-295, 2008
- Tsakamoto Y, Katsunobu Y, Omura Y, Maeda I, Hirai M, *et al.*: Subcutaneous panniculitis-like T-cell lymphoma: successful initial treatment with prednisolone and cyclosporin A. *Intern Med* 45:21-24, 2006
- Mukai HY, Okoshi Y, Shimizu S, Katsura Y, Takei N, *et al.*: Successful treatment of a patient with subcutaneous panniculitis-like T-cell lymphoma with high-dose chemotherapy and total body irradiation. *Eur J Haematol* 70:413-416, 2003
- Willemze R, Jaffe ES, Burg G, Cerroni L, Berti E, *et al.*: WHO-EORTC classification for cutaneous lymphomas. *Blood* 105:3768-3785, 2005
- Massone C, Chott A, Metzger D, Kerl K, Citarella L, *et al.*: Subcutaneous, blastic natural killer (NK), NK/T-cell, and other cytotoxic lymphomas of the skin: a morphologic, immunophenotypic, and molecular study of 50 patients. *Am J Surg Pathol* 28:719-735, 2004
- Massone C, Lozzi GP, Egberts F, Fink-Puches R, Cota C, *et al.*: The protean spectrum of non-Hodgkin lymphomas with prominent involvement of subcutaneous fat. *J Cutan Pathol* 33:418-425, 2006
- Willemze R, Jansen PM, Cerroni L, Beritelli E, Santucci M, *et al.*: Subcutaneous panniculitis-like T-cell lymphoma: definition, classification, and prognostic factors: an EORTC Cutaneous Lymphoma Group Study of 83 cases. *Blood* 111:838-845, 2008
- Go RS, Wester SM: Immunophenotypic and molecular features, clinical outcomes, treatments, and prognostic factors associated with subcutaneous panniculitis-like T-cell lymphoma: a systematic analysis of 156 patients reported in the literature. *Cancer* 101:1404-1413, 2004
- Imashuku S: Advances in the management of hemophagocytic lymphohistiocytosis. *Int J Hematol* 72:1-11, 2000
- Akashi K, Hayashi S, Gondo H, Mizuno S, Harada M, *et al.*: Involvement of interferon- γ and macrophage colony-stimulating factor in pathogenesis of hemophagocytic lymphohistiocytosis in adults. *Br J Haematol* 87:243-250, 1994
- Osugi Y, Hara J, Tagawa S, Takai K, Hosoi G, *et al.*: Cytokine production regulating Th1 and Th2 cytokines in hemophagocytic lymphohistiocytosis. *Blood* 89:4100-4103, 1997
- Aricò M, Danesino C, Pende D, Moretta L: Pathogenesis of hemophagocytic lymphohistiocytosis. *Br J Haematol* 114:761-769, 2001
- Alaibac M, Berti E, Pigozzi B, Chiarion V, Aversa S, *et al.*: High-dose chemotherapy with autologous blood stem cell transplantation for aggressive subcutaneous panniculitis-like T-cell lymphoma. *J Am Acad Dermatol* 52 (Suppl):121-123, 2005
- Kavan P, Kabicková E, Gajdos P, Koutecký J, Smelhaus V, *et al.*: Treatment of children and adolescents with non-Hodgkin's lymphoma (results based on the NHL Berlin-Frankfurt-Münster 90 protocols). *Cas Lek Cesk* 138:40-46, 1999
- Schrauder A, Reiter A, Gardner H, Niethammer D, Klingebiel T, *et al.*: Superiority of allogeneic hematopoietic stem-cell transplantation compared with chemotherapy alone in high-risk childhood T-cell acute lymphoblastic leukemia: results from ALL-BFM 90 and 95. *J Clin Oncol* 24:5742-5749, 2006
- Medhi K, Kumar R, Rishi A, Kumar L, Bakhshi S: Subcutaneous panniculitis-like T-cell lymphoma with hemophagocytosis: complete remission with BFM-90 protocol. *J Pediatr Hematol Oncol* 30:558-561, 2008
- Ratanatharathorn V, Uberti J, Karanes C, Abella E, Lum LG: Prospective comparative trial of autologous versus allogeneic bone marrow transplantation in patients with non-Hodgkin's lymphoma.

Sakurai E, *et al.*

Blood 84:1050-1055, 1994

19 Jones RJ, Ambinder RF, Piantadosi S, Santos GW: Evidence of a graft-versus-lymphoma effect associated with allogeneic bone marrow transplantation. *Blood* 77:649-653, 1991

20 Lee SJ, Klein JP, Barrett AJ, Ringden O, Antin JH, *et al.*: Severity of chronic graft-versus-host disease : association with

treatment-related mortality and relapse. *Blood* 100:406-414, 2002

21 Socié G, Stone JV, Wingard JR, Weisdorf D, Henslee-Downey PJ, *et al.*: Long-term survival and late deaths after allogeneic bone marrow transplantation. Late Effects Working Committee of the International Bone Marrow Transplant Registry. *N Engl J Med* 341:14-21, 1999

SHORT COMMUNICATION

A Case of Atypical Fibrous Histiocytoma with Positivity for CD163 and CD44

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Atypical fibrous histiocytoma (AFH) is a variant of dermatofibroma (DF) that was first described by Fukamizu et al. (1) in 1983. Histologically AFH is characterized by proliferation of dermal spindle cells composed mainly of atypical histiocytic cells with striking nuclear pleomorphism and atypia, in a background of classic fibrous histiocytoma (2). It is known that many cases of AFH follow a benign course if complete excision is carried out (2, 3). However, because the tumour cells are atypical, AFH must be differentiated from tumours of intermediate malignancy, such as dermatofibrosarcoma protuberans (DFSP) or atypical fibrous xanthoma (AFX), as well as more malignant tumours, such as pleomorphic dermal sarcoma (PDS)/malignant fibrous histiocytoma (MFH).

We report here a case of AFH on the left upper arm of a 63-year-old woman and describe its immunoreactivity in detail. We also discuss the points of histological and immunohistological differentiation between AFH and other cutaneous spindle cell tumours.

CASE REPORT

A 63-year-old woman presented with an 8-month history of a symptomless, slowly growing swelling on the left upper arm. The patient had no unusual medical or family history. Clinical examination revealed an 8-mm black-purplish hard mass with peripheral erythema (Fig. 1A). The tumour had arisen at a site without any known previous history of injury. A haemangioma was clinically suspected, and surgical excision was performed. Microscopic examination revealed a well-defined lesion, located in the dermis and extending to the subcutaneous tissue, with epidermal hyperplasia and a grenz zone (Fig. 1B). The lesion was composed largely of interlacing fascicles of predominant histiocyte-like eosinophilic spindle cells with elongated or plump vesicular nuclei, arranged in a storiform pattern. Abundant pleomorphic giant cells with huge bizarre nuclei (bi-lobed and multi-lobed) and histiocytes with large vesicular nuclei and prominent eosinophilic nucleoli were observed (Fig. 1C). In the peripheral region of the tumour, fibroblast-like spindle cells arranged in a storiform or fascicular pattern with collagen bundles were observed, resembling the classic features of DF. No necrosis was present. Foci of chronic inflammatory cells, including lymphocytes and plasma cells, were also evident. As a typical feature, we noted individual prominent hyalinized collagen bundles surrounded by tumour cells, predominantly in the periphery of the lesion (Fig. 1D). The mitotic count was 3 per 10 high-power fields (HPF).

Immunohistochemical staining revealed diffuse positivity for vimentin, factor XIIIa, CD68, CD163 (Fig. 1E) and CD44 (Fig. 1F). The lesion showed no reactivity for desmin, CD34, AE1/AE3, desmin, S-100 protein, α 1-antitrypsin or α 1-antichymotrypsin. Ki-67 staining showed less than 5% positive reactivity. Based on these findings, a diagnosis of atypical fibrous histiocytoma was made. As it was suspected that the initial resection may have left some residual tumour cells at depth, expanded excision with a 3-cm margin was performed one month later. At 30 months of follow-up, the patient was asymptomatic with no evidence of tumour recurrence.

Fig. 1. (A) Macroscopic view of the blackish-purplish skin tumour on the right arm. (B) Dermal-to-subcutaneous tumour with focal extension into the subcutaneous tissue. The epidermis was hyperplastic and a grenz zone was evident (haematoxylin and eosin; HE \times 1). (C) The tumour is composed of a proliferation of interlacing fascicles of predominantly histiocyte-like eosinophilic spindle cells with vesicular nuclei. Abundant pleomorphic giant cells with huge bizarre nuclei are present (HE \times 100). (D) At the border of the lesion, the cells are interspersed with hyaline collagen bundles (HE \times 40). Immunohistochemical studies of the tumour cells. The cells were positive for (E) CD163 and (F) CD44.

