



45. Clower, C. V. *et al.* The alternative splicing repressors hnRNP A1/A2 and PTB influence pyruvate kinase isoform expression and cell metabolism. *Proc Natl Acad Sci U S A* **107**, 1894–1899 (2010).
46. Chiou, N. T., Shankarling, G. & Lynch, K. W. hnRNP L and hnRNP A1 induce extended U1 snRNA interactions with an exon to repress spliceosome assembly. *Mol Cell* **49**, 972–982 (2013).
47. Rideau, A. P. *et al.* A peptide motif in Raver1 mediates splicing repression by interaction with the PTB RRM2 domain. *Nat Struct Mol Biol* **13**, 839–848 (2006).
48. Shankarling, G. & Lynch, K. W. Minimal functional domains of paralogs hnRNP L and hnRNP LL exhibit mechanistic differences in exonic splicing repression. *Biochem J* **453**, 271–279 (2013).
49. Sorek, R., Ast, G. & Graur, D. Alu-containing exons are alternatively spliced. *Genome Res* **12**, 1060–1067 (2002).
50. Krull, M., Petrusma, M., Makalowski, W., Brosius, J. & Schmitz, J. Functional persistence of exonized mammalian-wide interspersed repeat elements (MIRs). *Genome Res* **17**, 1139–1145 (2007).
51. Duclert, A., Savatier, N., Schaeffer, L. & Changeux, J. P. Identification of an element crucial for the sub-synaptic expression of the acetylcholine receptor epsilon-subunit gene. *J Biol Chem* **271**, 17433–17438 (1996).
52. Koike, S., Schaeffer, L. & Changeux, J. P. Identification of a DNA element determining synaptic expression of the mouse acetylcholine receptor delta-subunit gene. *Proc Natl Acad Sci U S A* **92**, 10624–10628 (1995).
53. Hutchinson, D. O. *et al.* Congenital endplate acetylcholinesterase deficiency. *Brain* **116** (Pt 3), 633–653 (1993).
54. Engel, A. G., Nagel, A., Walls, T. J., Harper, C. M. & Waisburg, H. A. Congenital myasthenic syndromes: I. Deficiency and short open-time of the acetylcholine receptor. *Muscle Nerve* **16**, 1284–1292 (1993).
55. Engel, A. G., Ohno, K., Bouzat, C., Sine, S. M. & Griggs, R. C. End plate acetylcholine receptor deficiency due to nonsense mutations in the epsilon subunit. *Ann Neurol* **40**, 810–817 (1996).
56. Engel, A. G., Lindstrom, J. M., Lambert, E. H. & Lennon, V. A. Ultrastructural localization of the acetylcholine receptor in myasthenia gravis and in its experimental autoimmune model. *Neurology* **27**, 307–315 (1977).
57. Uchitel, O. *et al.* Congenital myasthenic syndromes: II. Syndrome attributed to abnormal interaction of acetylcholine with its receptor. *Muscle Nerve* **16**, 1293–1301 (1993).
58. Ohno, K. *et al.* Congenital myasthenic syndrome caused by decreased agonist binding affinity due to a mutation in the acetylcholine receptor epsilon subunit. *Neuron* **17**, 157–170 (1996).
59. Das, R., Zhou, Z. & Reed, R. Functional association of U2 snRNP with the ATP-independent spliceosomal complex E. *Mol Cell* **5**, 779–787 (2000).

Acknowledgements

We are grateful to Robin Reed (Harvard Medical School, Boston, MA) for kindly providing MS2-MBP fusion protein and to Kentaro Taki (Nagoya University) for his technical assistance on the mass spectrometry analysis. This work was supported by Grants-in-Aid from the MEXT and MHLW of Japan to AM¹, KeO, MI, AM⁴, and KiO; and by NIH Research Grant NS6277 from the NINDS and by Research Grant from the MDA to AGE.

Author contributions

A.G.E. and Ki.O. conceived the project. M.A.R., A.M.,¹ Ke.O. and M.I. designed experiments; M.A.R. performed most of the experiments; Ki.O., D.O.H., Ke.O. contributed to genetic studies, electrophysiological studies, and *in vitro* spliceosome studies, respectively. M.A.R., Ke.O., A.M.,⁴ A.G.E. and Ki.O. wrote the paper.

Additional information

Supplementary information accompanies this paper at <http://www.nature.com/scientificreports>

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Rahman, M.A. *et al.* HnRNP L and hnRNP LL antagonistically modulate PTB-mediated splicing suppression of *CHRNA1* pre-mRNA. *Sci. Rep.* **3**, 2931; DOI:10.1038/srep02931 (2013).



This work is licensed under a Creative Commons Attribution 3.0 Unported license. To view a copy of this license, visit <http://creativecommons.org/licenses/by/3.0>

Teaching NeuroImages:

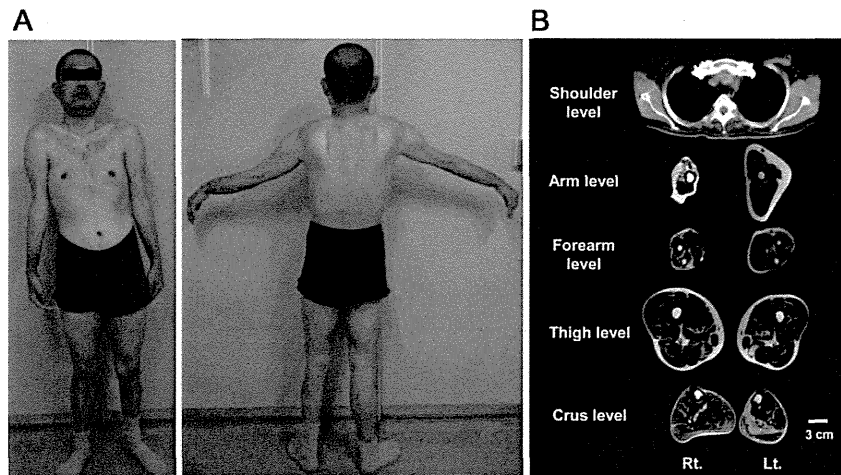
Unilateral arm and contralateral leg amyotrophy in FSHD

Unusual presentation

Kazuma Sugie, MD,
PhD
Yukiko K. Hayashi,
MD, PhD
Kanako Goto, BS
Ichizo Nishino, MD,
PhD
Satoshi Ueno, MD, PhD

Correspondence & reprint
requests to Dr. Sugie:
ksugie@naramed-u.ac.jp

Figure Photographs and muscle imaging findings of the whole body in a patient with facioscapulohumeral muscular dystrophy (FSHD)



(A) The patient predominantly shows remarkable atrophy of the right facial, shoulder girdle, and arm muscles and of the left leg muscles. He put his weight on his right leg (with permission). (B) CT of the shoulder and T2-weighted MRI of the 4 limbs. CT shows atrophy of the right shoulder girdle and greater pectoral muscles. Amyotrophy and replacement of muscle tissue by fat is asymmetrically pronounced in the right sides of the biceps and triceps brachii and forearm, and the left sides of the femoral and calf muscles on MRI.

A 43-year-old, right-handed man noticed right arm weakness at age 23, followed by the development of left leg weakness and claudication. Although his deceased mother was considered to have had facioscapulohumeral muscular dystrophy (FSHD), her clinical symptoms were unclear. Neurologic examinations and imaging showed predominant weakness/atrophy in the right arm and left leg (figure). EMG demonstrated predominant myopathic changes in the right arm and left leg. No involvement of the CNS or peripheral nerves was apparent. Southern blotting analysis for FSHD revealed a 20-kb *EcoRI* fragment on 4q35 (normal >35 kb).

Asymmetric muscle involvement is a characteristic feature of FSHD.^{1,2} Asymmetry might depend not only on handedness, but also on genetic predisposition.

REFERENCES

- Olsen DB, Gideon P, Jeppesen TD, Vissing J. Leg muscle involvement in facioscapulohumeral muscular dystrophy assessed by MRI. *J Neurol* 2006;253:1437–1441.
- Brouwer OF, Padberg GW, van der Ploeg RJ, Ruys CJ, Brand R. The influence of handedness on the distribution of muscular weakness of the arm in facioscapulohumeral muscular dystrophy. *Brain* 1992;115:1587–1598.

From the Department of Neurology (K.S., S.U.), Nara Medical University School of Medicine, Nara; and Department of Neuromuscular Research (K.S., Y.K.H., K.G., I.N.), National Institute of Neuroscience, National Center of Neurology and Psychiatry (NCNP), Kodaira, Tokyo, Japan. The authors report no disclosures relevant to the manuscript. Go to Neurology.org for full disclosures.

Provided for non-commercial research and education use.
Not for reproduction, distribution or commercial use.



This article appeared in a journal published by Elsevier. The attached copy is furnished to the author for internal non-commercial research and education use, including for instruction at the authors institution and sharing with colleagues.

Other uses, including reproduction and distribution, or selling or licensing copies, or posting to personal, institutional or third party websites are prohibited.

In most cases authors are permitted to post their version of the article (e.g. in Word or Tex form) to their personal website or institutional repository. Authors requiring further information regarding Elsevier's archiving and manuscript policies are encouraged to visit:

<http://www.elsevier.com/copyright>

suggest a differing pattern of risk factors in CA-MRSA infection in HIV patients in our area resulting from the combination of risk factors. Those patients with poorer control of HIV infection and immigrants, mainly from South America, have the higher risk for such infections, although high-risk sexual behavior seems to be also associated.

Conflicts of interest

All authors declare no conflict of interest.

Acknowledgments

This study was presented in part Presented in part at the 13th European AIDS Conference (EACS), Belgrade, Serbia, 2011 (abstract PE 11.5/1).

References

1. Popovich KJ, Weinstein RA, Aroutcheva A, Rice T, Hota B. Community-associated methicillin-resistant *Staphylococcus aureus* and HIV: intersecting epidemics. *Clin Infect Dis* 2010;**50**(7): 979–87.
2. Crum-Cianflone NF, Grandits G, Echols S, Ganesan A, Landrum M, Weintrob A, et al. Trends and causes of hospitalizations among HIV-infected persons during the late HAART era: what is the impact of CD4 counts and HAART use. *J Acquir Immune Defic Syndr* 2010;**54**:248–57.
3. Lee NE, Taylor MM, Bancroft E, Ruane PJ, Morgan M, McCoy L, et al. Risk factors for community-associated methicillin-resistant *Staphylococcus aureus* skin infections among HIV-positive men who have sex with men. *Clin Infect Dis* 2005;**40**:1529–34.
4. Atkinson SR, Paul J, Sloan E, Curtis S, Miller R. The emergence of methicillin-resistant *Staphylococcus aureus* among injecting drug users. *J Infect* 2009;**58**(5):339–45.
5. Imaz A, Pujol M, Barragán P, Domínguez MA, Tiraboschi JM, Podzamczar D. Community associated methicillin-resistant *Staphylococcus aureus* in HIV-infected patients. *AIDS Rev* 2010;**12**:153–63.
6. Tenover FC, Arbeit RD, Goering RV, Mickelsen PA, Murray BE, Persing DH, et al. Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing. *J Clin Microbiol* 1995;**33**:2233–9.
7. Enright MC, Day NP, Davies CE, Peacock SJ, Spratt BG. Multilocus sequence typing for characterization of methicillin-resistant and methicillin-susceptible clones of *Staphylococcus aureus*. *J Clin Microbiol* 2000;**38**:1008–15.
8. Oliveira DC, De Lencastre H. Multiplex PCR strategy for rapid identification of structural types and variants of the *mec* element in methicillin-resistant *Staphylococcus aureus*. *Antimicrob Agents Chemother* 2002;**46**:2155–61.
9. Manzur A, Dominguez AM, Pujol M, González MP, Limon E, Hornero A, et al. Community-acquired methicillin-resistant *Staphylococcus aureus* infections: an emerging threat in Spain. *Clin Microbiol Infect* 2008;**14**:377–80.
10. Marimón JM, Villar M, García-Arenzana JM, Caba Ide L, Pérez-Trallero E. Molecular characterization of *Staphylococcus aureus* carrying the *panton-valentine leucocidin* genes in northern Spain. *J Infect* 2012;**64**:47–53.
11. Blanco R, Tristan A, Ezpeleta G, Larsen AR, Bes M, Etienne J, et al. Molecular epidemiology of *panton-valentine leucocidin*-positive *Staphylococcus aureus* in Spain: emergence of the USA300 clone in an autochthonous population. *J Clin Microbiol* 2011;**49**:433–6.

Arkaitz Imaz*
HIV Unit, Infectious Diseases Service,
Hospital Universitari de Bellvitge,
L'Hospitalet de Llobregat, Barcelona, Spain
E-mail address: aimaz@bellvitgehospital.cat

Nazaret Cobos-Trigueros
Infectious Diseases Service, Hospital Clinic-IDIBAPS,
Barcelona, Spain

Vicenç Falcó
Infectious Diseases Service, Hospital Universitari
Vall d'Hebron, Barcelona, Spain

M. Angeles Dominguez
Microbiology Service, Hospital Universitari de Bellvitge,
L'Hospitalet de Llobregat, Barcelona, Spain

Christian Manzardo
Infectious Diseases Service, Hospital Clinic-IDIBAPS,
Barcelona, Spain

Miquel Pujol
Infectious Diseases Service, Hospital Universitari
de Bellvitge, L'Hospitalet de Llobregat, Barcelona, Spain

Jordi Curto
HIV Unit, Infectious Diseases Service,
Hospital Universitari de Bellvitge,
L'Hospitalet de Llobregat, Barcelona, Spain

Rosa Bartolomé
Microbiology Service, Hospital Universitari
Vall d'Hebron, Barcelona, Spain

Alex Soriano
Infectious Diseases Service, Hospital Clinic-IDIBAPS,
Barcelona, Spain

Daniel Podzamczar
HIV Unit, Infectious Diseases Service,
Hospital Universitari de Bellvitge,
L'Hospitalet de Llobregat, Barcelona, Spain

*Corresponding author. HIV Unit, Infectious Disease
Department, Hospital Universitari de Bellvitge,
Feixa Llarga s/n, 08907 L'Hospitalet de Llobregat,
Barcelona, Spain. Tel./fax: +34 932607667.

Accepted 7 October 2012

© 2012 The British Infection Association. Published by Elsevier Ltd.
All rights reserved.

<http://dx.doi.org/10.1016/j.jinf.2012.10.013>

Predictors of outcomes in acyclovir-treated limbic encephalitis

Recently published national guidelines for suspected viral encephalitis cover a wide range of encephalitis, including viral infections, especially herpes simplex virus (HSV), and autoimmune inflammatory disorders.¹ Subacute

Table 1 Detailed clinical and neuroradiological and neurophysiological independent variables of 31 patients with subacute limbic encephalitis.

Patient no.	(1) Sex	(2) Age (years)	(3) Detect of viral etiology ^a	(4) Days ^b	(5) GCS score ^h	(6) Maximum monocytes in CSF (/mm ³)	(7) Detection of focal lesion by initial CT ^b	(8) MRI lesions ^c	(9) Detection of PLEDs ^d	(10) Use of steroid treatments ^e	Outcome ^f
1	M	57	0	5	7	128	1	1	1	1	Poor (severe sequela)
2	F	60	0	2	6	28	1	1	1	0	Good (mild sequela)
3	F	30	0	8	7	9	0	1	0	0	Poor (moderate sequela)
4	F	31	0	9	7	296	1	1	1	0	Poor (severe sequela)
5	F	64	0	3	13	49	1	0	0	1	Poor (severe sequela)
6	M	33	0	2	13	54	0	1	1	0	Poor (death)
7	F	51	0	4	13	48	0	0	0	0	Good (mild sequela)
8	M	45	0	4	13	8	1	1	1	0	Good (mild sequela)
9	F	64	0	4	10	257	0	0	1	0	Poor (severe sequela)
10	M	74	0	2	14	426	0	0	1	0	Good (complete recovery)
11	M	29	1	3	13	3	0	0	0	0	Good (complete recovery)
12	M	73	1	3	13	108	1	1	1	0	Poor (severe sequela)
13	M	58	1	10	12	25	1	1	1	0	Poor (moderate sequela)
14	M	62	0	6	10	526	0	0	1	0	Poor (severe sequela)
15	F	34	1	4	11	161	0	1	0	0	Good (mild sequela)
16	F	18	1	2	8	38	0	0	0	1	Good (mild sequela)
17	F	59	0	5	12	50	0	1	0	0	Good (mild sequela)
18	F	59	1	3	11	73	0	0	0	0	Good (mild sequela)
19	F	56	1	12	12	13	0	0	0	1	Poor (moderate sequela)
20	F	28	1	8	11	4	0	0	0	1	Poor (moderate sequela)
21	F	68	1	14	9	6	0	1	1	0	Poor (severe sequela)
22	M	36	1	4	13	16	1	1	0	0	Poor (moderate sequela)
23	F	23	1	5	14	11	0	0	0	0	Good (mild sequela)
24	F	25	1	6	10	56	0	1	0	0	Poor (moderate sequela)
25	M	24	1	2	7	29	0	0	0	0	Poor (moderate sequela)
26	F	23	1	15	14	123	0	1	1	0	Poor (severe sequela)
27	M	29	1	7	14	282	0	0	0	0	Good (mild sequela)
28	F	27	1	3	7	4	0	1	0	0	Poor (severe sequela)
29	M	88	1	26	12	4	0	0	1	0	Poor (death)
30	M	61	1	2	11	98	1	1	1	1	Poor (severe sequela)
31	F	32	1	8	14	98	0	0	0	1	Good (complete recovery)

GCS: Glasgow coma scale CSF: cerebral spinal fluid, CT: computed tomography, MRI: magnetic resonance imaging, PLEDs :periodic lateralized epileptiform discharges.

^a 0 = absent, 1 = present.

^b 0 = absent, 1 = present.

^c Abnormal T2/FLAIR signal hyperintensity on brain MRI restricted to medial temporal areas (=0) or in that plus other areas as shown in

Supplemental Table 2 (=1).

^d 0 = absent, 1 = present.

^e At acute stage, 0 = given, 1 = not given.

^f Outcome 3 months after completion of acyclovir treatments.

^g Duration from neurological onset to acyclovir treatment.

^h When acyclovir treatment was started.

limbic encephalitis (LE) is a subtype of encephalitis that includes a wide differential diagnosis because of the rapid clinical presentation and lack of specificity. Outcomes of LE have been reported according to the underlying cause. In HSV encephalitis, age, consciousness level, the delay between hospital admission and initiation of acyclovir, and the detection of lesions on computed tomography (CT) are predictors of outcome.^{2–4} In encephalitis associated with neuronal antibodies, whether the antibodies are against intraneuronal antigens or neutrophil antibodies⁵ and the presence of tumor⁶ are important prognostic factors, and antibody suppression and tumor resection are effective treatments. Many patients with LE have initially receive acyclovir with or without steroids if a causal factor is not detected early after disease onset, even in the presence of medial temporal lesions on magnetic resonance imaging (MRI). Since the main predictors of outcome in LE remain unclear, we studied 31 patients with LE who showed medial temporal lesions on MRI to identify potential predictors of outcome.

Subjects and methods

We studied 31 patients with LE (45.8 ± 19 years, range 18–88 years) from among 97 patients with encephalitis between March 1993 and May 2012. Diagnostic criteria for LE and information on detectable viruses and acyclovir treatment are shown in Supplemental material 1.

Statistical analysis

A total of 10 variables divided into the following two sets were evaluated: clinical independent variables and neuro-radiological and neurophysiological independent variables (Supplemental material 2).

Variables related to outcomes on univariate logistic regression analysis were entered into multiple logistic regression analysis using forced entry. Odds ratios (OR) and 95% confidence intervals (CI) were calculated. Correlations of each variable were also evaluated by Spearman's rank correlation test. Receiver operating characteristic (ROC) analysis was used to determine cutoff values of the significant variables on multiple logistic regression analysis, and the cutoff value derived from the ROC curves at the point of highest accuracy was used to calculate mean sensitivity and specificity. To statistically analyze differences in clinical characteristics between patients with good outcomes and those with poor outcomes, variables were evaluated with Mann–Whitney tests, followed by Fisher's exact probability test. SPSS software (Version 18) was used for statistical analysis.

Results

The clinical and other independent variables of the 31 patients with LE are shown in Table 1. Nineteen patients (61%) had poor outcomes (Table 2). On multiple logistic regression analysis, the Glasgow coma scale (GCS) score at initiation of acyclovir treatment was the best predictor of outcomes (OR = 2.158, $p = 0.032$, 95% CI = 1.07–4.352) (Table 3). No interactions were found between the GCS

Table 2 Detailed clinical and neuroradiological and neurophysiological independent variables of 19 patients with subacute limbic encephalitis who had poor outcomes.

Subacute limbic encephalitis with poor outcomes ($n = 19$)	
Age (years)	Mean 47.7 (SD, 20.1; median, 56)
Sex (Men/Female)	9/10.
No detection of viral etiology (n)	12
Duration from neurological onset to initiation of acyclovir treatment (days)	Mean 7.4 (SD, 6; median, 5)
GCS score at initiation of acyclovir treatment	Mean 10.4 (SD, 2.4; median, 12)
Maximum monocytes in CSF ($/\text{mm}^3$)	Mean 95 (SD, 133.7; median 49)
Presence of focal lesions on initial CT	7
Cranial MRI lesions ^a	12
Presence of PLEDs on electroencephalogram	2
Not given steroid treatments at the acute stage	5

GCS: Glasgow coma scale CSF: cerebrospinal fluid, CT: computed tomography.

MRI: magnetic resonance imaging, PLEDs: periodic lateralized epileptiform discharges, n : number.

^a Abnormal T2/FLAIR signal hyperintensity on brain MRI restricted to the limbic areas plus other areas.

score at initiation of acyclovir treatment and the 9 other independent variables. Furthermore, when we additionally entered age, a factor that has been identified as a major determinant of prognosis in encephalitis,^{3,4} the results similarly showed that the GCS score was the best predictor of outcomes, with higher odds ratios than the other variables (OR = 1.765, $p = 0.029$, 95% CI = 1.061–2.936). The GCS score at initiation of acyclovir treatment was higher, and the interval from neurological onset including headache to initiation of acyclovir treatment was shorter in patients with good outcomes than in those with poor outcomes ($p = 0.062$ and $p = 0.072$, respectively). No other independent variables differed significantly according to outcome. On ROC analysis, the GCS score cutoff value at initiation of acyclovir treatment was 12.5, with 58.3% sensitivity and 73.7% specificity for outcome; the area under the ROC curve was 0.7 ($P = 0.065$). Among the 19 patients with a GCS score of <12.5 at the initiation of acyclovir treatment, 14 had poor outcomes.

Discussion

GCS score at admission or treatment initiation was previously reported to be a predictor of outcomes in central infectious diseases such as encephalitis without confirmed viral findings,⁷ and HSV encephalitis,^{1–3} but not in LE. Since our subjects were evaluated starting in 1993, viral diagnosis might not have been timely. Moreover, our subjects included LE patients with confirmed HSV or varicella

Table 3 Independent variables predictive of outcomes in 31 patients with subacute limbic encephalitis.

	Crude odds ratio (95% CI)	P	Adjusted odds ratio (95% CI) ^a	P
Age (years)	1.066 (0.928–1.225)	0.365		
Sex	0.02 (0–3.236)	0.132	0.075 (0.005–1.097)	0.058
Detection of viral etiology	0.317 (0.01–9.764)	0.511		
Duration from neurological onset to initiation of acyclovir treatment (days)	2.332 (1.043–5.214)	0.039*	0.611 (0.396–0.943)	0.026*
GCS score at initiation of acyclovir treatment	0.311 (0.104–0.934)	0.037*	2.158 (1.07–4.352)	0.032*
Maximum monocytes in CSF (/mm ³)	0.996 (0.997–1.017)	0.717		
Detection of focal lesions on initial CT	1.31 (0.013–128.526)	0.908		
Cranial MRI lesions ^b	4.214 (0.04–440.03)	0.544		
Detection of PLEDs on electroencephalogram	0.001 (0–1.414)	0.063	0.041 (0–3.799)	0.201
Use of steroid treatments at the acute stage	9.75 (0.033–2913.511)	0.434		

GCS: Glasgow coma scale CSF: cerebrospinal fluid, CT: computed tomography, MRI: magnetic resonance imaging.

* $P < 0.05$.

PLEDs: periodic lateralized epileptiform discharges.

^a Adjusted for the duration from neurological onset to acyclovir treatment, Glasgow coma scale score, sex and the detection of PLEDs.

^b Abnormal T2/FLAIR signal hyperintensity on brain MRI restricted to the limbic areas or in that plus other areas.

zoster virus who received acyclovir, which is likely to be unsuitable for the evaluation of outcomes. However, all of our patients received acyclovir, and a significant independent variable providing evidence of a viral etiology was not found on single or multiple logistic regression analysis. Because the etiology of LE is unclear in most cases,⁶ it is important to identify predictors of outcomes in LE as a whole, including cases in which a cause is not detected. Because the sensitivity was relatively low, whether a cutoff value of 12.5 for the GCS score is useful remains unclear. The initial GCS scores of patients with HSV encephalitis were ≥ 12 in most cases.⁸ The low sensitivity of the GCS score in our study may be attributed to the fact that some patients showed rapid neurological deterioration despite having a high GCS score at admission.⁹ The interval from neurological onset to initiation of acyclovir treatment was likely a predictor of outcomes in our study. The delay between hospital admission and the initiation of acyclovir therapy was twice as long for patients with poor outcomes than those with favorable outcomes.²

Age was not a significant predictor of outcomes in our study. A possible reason for lack of significance for age may be related to the fact that one-third of the patients were relatively young, between the ages of 18 and 34 years.

When initially treating a patient with LE, the detection of a virus or neuronal antibodies must be the best predictor of outcomes. However, if virus or neuronal antibody is not detected, the identification of early factors that predict outcomes might contribute to better disease management. We hope that our results will enhance the accuracy of predicting outcomes in patients with LE.

Financial disclosure

There was no financial disclosure related with our paper.

Disclosure

The authors report no conflicts of interest related with our paper.

Appendix A. Supplementary data

Supplementary data related to this article can be found at <http://dx.doi.org/10.1016/j.jinf.2012.10.007>.

References

- Solomon T, Michael BD, Smith PE, Sanderson F, Davies NW, Hart IJ, et al. Management of suspected viral encephalitis in adults—Association of British neurologists and British infection association national guidelines. *J Infect* 2012;**64**:347–73.
- McGrath N, Anderson NE, Croxson MC, Powell KF. Herpes simplex encephalitis treated with acyclovir: diagnosis and long term outcome. *J Neurol Neurosurg Psychiatry* 1997;**63**:321–6.
- Whitley RJ, Alford CA, Hirsch MS, Schooley RT, Luby JP, Aoki FY, et al. Vidarabine versus acyclovir therapy in herpes simplex encephalitis. *N Engl J Med* 1986;**314**:144–9.
- Kamei S, Sekizawa T, Shiota H, Mizutani T, Itoyama Y, Takasu T, et al. Evaluation of combination therapy using acyclovir and corticosteroid in adult patients with herpes simplex virus encephalitis. *J Neurol Neurosurg Psychiatry* 2005;**76**:1544–9.
- Ances BM, Vitaliani R, Taylor RA, Liebeskind DS, Voloschin A, Houghton DJ, et al. Treatment-responsive limbic encephalitis identified by neuropil antibodies: MRI and PET correlates. *Brain* 2005;**128**:1764–77.
- Gultekin SH, Rosenfeld MR, Voltz R, Eichen J, Posner JB, Dalmau J. Paraneoplastic limbic encephalitis: neurological symptoms, immunological findings and tumour association in 50 patients. *Brain* 2000;**123**:1481–94.
- Berlit P. The prognosis and long-term course of viral encephalitis. *J Neuroimmunol* 1988;**20**:117–25.

8. Domingues RB, Fink MC, Tsanaclis AM, de Castro CC, Cerri GG, Mayo MS, et al. Diagnosis of herpes simplex encephalitis by magnetic resonance imaging and polymerase chain reaction assay of cerebrospinal fluid. *J Neurol Sci* 1998;157:148–53.
9. Raschilas F, Wolff M, Delatour F, Chaffaut C, De Broucker T, Chevret S, et al. Outcome of and prognostic factors for herpes simplex encephalitis in adult patients: results of a multicenter study. *Clin Infect Dis* 2002;35:254–60.

Hiroshi Kataoka*

*Department of Neurology, Nara Medical University, 840
Shijo-cho, Kashihara, Nara 634-8522, Japan
E-mail address: hk55@naramed-u.ac.jp*

Keigo Saeki

Community Health and Epidemiology, Nara Medical University, Kashihara, Nara, Japan

Yasuyo Kobayashi
Takao Kiriyaama
Kazuma Sugie
Satoshi Ueno

*Department of Neurology, Nara Medical University, 840
Shijo-cho, Kashihara, Nara 634-8522, Japan*

*Corresponding author. Tel.: +81 744 29 8860;
fax: +81 744 24 6065.

Accepted 7 October 2012

Crown Copyright © 2012 Published by Elsevier Ltd on behalf of The British Infection Association. All rights reserved.

<http://dx.doi.org/10.1016/j.jinf.2012.10.007>

Characterization of Dermatomyositis with Coexistence of Anti-Jo-1 and Anti-SRP Antibodies

Kazuma Sugie, Yasuyo Tonomura and Satoshi Ueno

Abstract

We describe a patient with dermatomyositis who presented with rapidly developing severe muscle weakness complicated by massive pleural effusion with interstitial lung disease. Myopathological analysis was suggestive of dermatomyositis. This patient showed both anti-Jo-1 and anti-SRP antibodies in serum. To our knowledge, the coexistence of these two myositis-specific autoantibodies (MSA) is considered extremely rare and is clearly an exception to the rule of having only one MSA. Our findings provide compelling evidence that the coexistence of these two MSAs may lead to more severe clinical symptoms, interacting in a complex fashion, thus expanding the clinical spectrum of idiopathic inflammatory myopathy.

Key words: idiopathic inflammatory myopathy, dermatomyositis, pleural effusion, myositis-specific autoantibody, anti-Jo-1 antibody, anti-SRP antibody

(Intern Med 51: 799-802, 2012)

(DOI: 10.2169/internalmedicine.51.6566)

Introduction

Idiopathic inflammatory myopathies such as polymyositis (PM) and dermatomyositis (DM) are systemic autoimmune connective tissue diseases characterized by chronic muscle inflammation with involvement of various organs (1). The pathogenesis of PM/DM is unknown, but autoantibodies directed against various cellular constituents have been identified in patients with PM/DM. Some autoantibodies found almost exclusively in PM/DM are known as myositis-specific autoantibodies (MSA), including anti-Jo-1 (histidyl tRNA synthetase) antibody, anti-PL-7 antibody, anti-signal recognition particle (SRP) antibody, anti-Mi-2 antibody, and anti-CADM-140 antibody. Each MSA is associated with a set of unique clinical features (2, 3).

We describe a 61-year-old man with DM who presented with severe muscle involvement characterized by rapidly developing proximal weakness, culminating in severe disability. He also showed massive pleural effusion with interstitial lung disease (ILD). Interestingly, both anti-Jo-1 and anti-SRP antibodies were positive in his serum. To our knowledge, the coexistence of these two types of MSA is considered extremely rare. Only one other case of idiopathic in-

flammatory myopathy with these two MSAs has been described in a recent report (4). Our findings suggest that coexistence of these two MSAs is associated with specific clinicopathological features.

Case Report

A 61-year-old man was admitted in June because of a 1-month history of rapidly progressive severe weakness of all four extremities. His skin was discolored, and he had dyspnea. The past history was noncontributory to the present illness. On admission, he presented with difficulty in getting up from bed and lifting his arms above his head. Physiological examination showed severe symmetric proximal weakness (less than grade 3 according to the Medical Research Council scale) of all four extremities. There were no other motor deficits. Sensory and stretch reflexes were normal. Erythematous rashes were present on the arms, trunk, legs, and face, including a typical heliotrope rash and Gottron's papules.

Laboratory examinations showed very high levels of creatine kinase (CK) (5,685 IU/L; normal: <160) in serum. The erythrocyte sedimentation rate and C-reactive protein were slightly elevated (80 mm/hr, <10; 2.6 IU/L, <0.1). Serum

Department of Neurology, Nara Medical University School of Medicine, Japan

Received for publication September 15, 2011; Accepted for publication December 5, 2011

Correspondence to Dr. Kazuma Sugie, ksugie@naramed-u.ac.jp

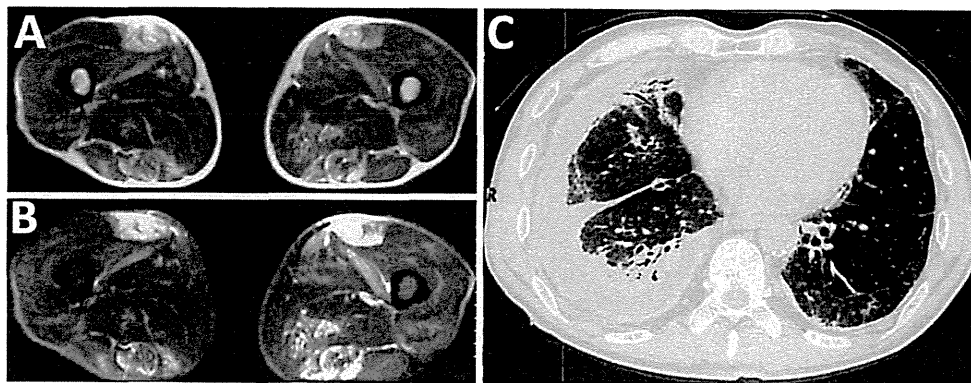


Figure 1. Magnetic resonance images of the thighs (A, B) and computed tomographic scans of the chest (C). T2-weighted (A) and T2-short-tau inversion recovery (STIR) images (B) showed diffuse high intensity in the frontal and dorsal aspects of both thigh muscles, suggesting intramuscular inflammation and edema. Chest images showed massive pleural effusion with interstitial lung disease (C).

antinuclear antibody was detected, accompanied by positivity for both anti-Jo-1 and anti-SRP antibodies, but negativity for other MSA, such as anti-PL-7 antibody. Among myositis-associated autoantibodies, anti-SS-A, anti-SS-B, anti-U1-RNP, and anti-Scl-70 antibodies were not detected. Electromyography showed short duration, small amplitude, polyphasic motor unit potentials with fibrillation potentials in the upper and lower limb muscles. Magnetic resonance images of the skeletal muscle showed diffuse inflammation and edema, most prominent in the proximal muscles of all four extremities (Fig. 1A, 1B). Computed tomography of the lung showed severe changes characteristic of ILD, with massive pleural effusion (Fig. 1C). The pleural fluid revealed exudate with no evidence of malignancy. A biopsy of the femoral muscle showed many necrotic and regenerative fibers with marked perimysial cell infiltration (Fig. 2). The infiltrating CD4+/CD8+ T cell ratio at perimysial sites (mean \pm SD) was 1.58 ± 0.28 . Characteristic perifascicular muscle fiber atrophy was seen. Strong major histocompatibility complex class I (MHC-I) expression, especially in perifascicular atrophic fibers, was positive in cytoplasm. There was no expression of CD8/MHC-I complex, which suggested that CD8+ T cells invaded non-necrotic fibers that express MHC-I antigen. Expression of membrane attack complex (MAC) was present on endomysial capillaries, but not on necrotic fibers. These pathological findings of muscle suggested DM rather than PM.

The patient was given a diagnosis of DM with ILD. He initially received oral prednisone (1 mg/kg/day) for a month, with tapering to 20 mg/day over the course of the next three months. His muscle strength gradually improved, but he was still unable to move independently. Respiratory difficulties and pleural effusion were mildly decreased. The erythematous rashes decreased, but persisted slightly. Four months after the start of treatment, a progressive gastric cancer (papillary adenocarcinoma, stage IIIA) was diagnosed. A gastrectomy was thus performed. Subsequently, the muscle weak-

ness and respiratory difficulties worsened despite an increase in the dose of steroids. One year after gastrectomy, the patient died of progressive ILD with massive pleural effusion and multiple liver metastases from gastric cancer.

Discussion

We described a patient with idiopathic inflammatory myopathy accompanied by ILD with massive pleural effusion, who presented with rapidly developing severe proximal weakness and respiratory difficulty. His skin lesions were suggestive of DM. Histopathological examination of a muscle specimen revealed many necrotic and regenerative fibers with marked perimysial cell infiltration, predominantly involving CD4+ T cells. Strong MHC-I expression by perifascicular atrophic fibers was consistent with DM (5). In addition, expression of MAC on endomysial capillaries, but not on necrotic fibers in our patient distinguished DM from paraneoplastic necrotizing myopathy (6). Collectively, these histopathological findings of muscle, including no expression of CD8/MHC-I complex, suggested DM rather than PM.

Interestingly, the present patient showed both anti-Jo-1 and anti-SRP antibody in his serum. The presence of these two MSAs is considered extremely rare and is clearly an exception to the rule of having only one MSA in association with PM/DM (7). To our knowledge, the coexistence of these MSAs has only been documented one time previously (4). That patient had severe muscle weakness and ILD, characterized by the presence of both anti-Jo-1 and anti-SRP antibody. Although the reason for this association and the pathogenic roles of these two MSAs are unclear, MSA may play a key, yet indirect part in the etiology of PM/DM.

Each MSA is associated with a set of unique clinical features (2, 3). Anti-Jo-1 antibody, one of the aminoacyl tRNA synthetases antibodies, is closely related to PM/DM, which