# 研究成果の刊行に関する一覧表レイアウト

# 書籍

著者氏名	論文タイトル名	書籍全体の 編集者名	書籍名	出版社名	出版地	出版年	ページ
○佐藤貴浩	2.紅斑の発症メカニズム	古江増隆横関博雄	皮膚科臨床ア セット18「紅班 症と痒疹群」	中山書店.	東京	2013	p7-11
o高山かおる	17.成人Still病に伴う紅 斑の症状・診断・治療	古江増隆横関博雄	皮膚科臨床ア セット18「紅班 症と痒疹群」	中山書店.	東京	2013	p91-95
o岡 恵子	27.点状紅斑の概念・病態・診断・治療	古江増隆横関博雄	皮膚科臨床ア セット18「紅班 症と痒疹群」	中山書店.	東京	2013	p149-152
o沢田泰之	28.手掌紅斑の概念・病態・診断・治療	古江増隆横関博雄	皮膚科臨床ア セット18「紅班 症と痒疹群」	中山書店.	東京	2013	p153-157
○佐藤貴浩	30.痒疹の定義・分類	古江増隆横関博雄	皮膚科臨床ア セット18「紅班 症と痒疹群」	中山書店.	東京	2013	p166
o宇賀神つかさ	31.痒疹発症における 好塩基球の役割.	古江増隆横関博雄	皮膚科臨床ア セット18「紅班 症と痒疹群」	中山書店.	東京	2013	p167-171
○佐藤貴浩	32.急性痒疹・亜急性痒疹の概念・病態・症状.	古江増隆横関博雄	皮膚科臨床ア セット18「紅班 症と痒疹群」	中山書店.	東京	2013	p172-173
○西澤 綾	33.急性痒疹・亜急性痒 疹の診断・治療・生活 指導.	古江増隆横関博雄	皮膚科臨床ア セット18「紅班 症と痒疹群」	中山書店.	東京	2013	p174-178
○横関博雄	34.慢性痒疹の定義・分類・症状・病理・診断・ 鑑別診断	古江増隆 横関博雄	皮膚科臨床ア セット18「紅班 症と痒疹群」	中山書店.	東京	2013	p179-183
○佐藤貴浩	第1章 各疾患の診断 と治療. I.湿疹と類症. 1.皮膚瘙痒症	横関博雄片山一朗	高齢者によく みられる皮膚 疾患アトラス	医薬ジャーナル	東京	2013	p12.
o高山かおる	第1章 各疾患の診断 と治療. I.湿疹と類症. 2.接触皮膚炎	横関博雄 片山一朗	高齢者によく みられる皮膚 疾患アトラス	医薬ジャーナル	東京	2013	p14.
○井川 健	第1章 各疾患の診断 と治療. I.湿疹と類症. 6.ビダール苔癬.	横関博雄 片山一朗	高齢者によく みられる皮膚 疾患アトラス	医薬ジャーナル	東京	2013	p28.
○佐藤貴浩	第1章 各疾患の診断 と治療. I.湿疹と類症. 9.多形慢性痒疹.	横関博雄 片山一朗	高齢者によく みられる皮膚 疾患アトラス	医薬ジャーナル	東京	2013	p41-43.
○沢田泰之	第1章 各疾患の診断 と治療. III.物理的障 害および薬剤による疾 患 1.下腿潰瘍、静脈 瘤、慢性色素性紫斑	横関博雄片山一朗	高齢者によく みられる皮膚 疾患アトラス	医薬ジャーナル	東京	2013	p70-78.

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o高山かおる	第1章 各疾患の診断 と治療. III.物理的障 害および薬剤による疾 患 6.胼胝・鶏眼	横関博雄片山一朗	高齢者によく みられる皮膚 疾患アトラス	医薬ジャーナル	東京	2013	p97-99.
○西澤 綾	第1章 各疾患の診断 と治療. III.物理的障 害および薬剤による疾 患 8.扁平苔癬	横関博雄片山一朗	高齢者によく みられる皮膚 疾患アトラス	医薬ジャーナル	東京	2013	p104-107.
o高山かおる	第1章 各疾患の診断 と治療. IV老化に伴 う皮膚変化 6.爪の変 化	横関博雄片山一朗	高齢者によく みられる皮膚 疾患アトラス	医薬ジャーナル	東京	2013	p127-129.
○高河慎介・ 沢田泰之	第1章 各疾患の診断 と治療. VIII.デルマ ドローム 1.糖尿病性 皮膚症	横関博雄片山一朗	高齢者によく みられる皮膚 疾患アトラス	医薬ジャーナル	東京	2013	p206-211.
○井川 健	第2章 外用剤の種類 と使い方. I.ステロイ ド外用剤.	横関博雄 片山一朗	高齢者によく みられる皮膚 疾患アトラス	医薬ジャーナル	東京	2013	P224-230.
○井川 健	肉芽腫性皮膚疾患 サルコイドーシス・他の肉芽腫. Vリポイド類壊死症. 38. リポイド類壊死症の治療と経過.	横関博雄片山一朗	皮膚科臨床ア セット14	中山書店	東京	2013	P221-223.
椛島健治	アレルギー学への招待状	椛島健治	実験医学増 刊・アレルギー 疾患研究	羊土社	東京都	2013 年	2698-2705
宮地良樹	保湿薬の使い方	五十嵐敦之、 宮地良樹、 清水宏	一冊で分かる 最新皮膚科治療、皮膚科サブスペシャリティーシリーズ	文光堂	東京都	2013 年	13-16
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宮地良樹	抗ヒスタミン薬の効果 と眠気は相関するの か?	宮地良樹	抗ヒスタミン 薬:達人の処方 箋	メディカル レビュー社	東京	2013 年	38-39
宮地良樹	なぜ血が出るまで掻い ても掻破をやめないの か?	宮地良樹	抗ヒスタミン 薬:達人の処方 箋	メディカル レビュー社	東京	2013 年	70-71
戸倉新樹	皮膚科疾患 最近の動 向	山口徹, 北原光夫, 福井次矢	今日の治療指 針2013年版	医学書院	東京	2013	pp1024- 1026
織茂弘志, 戸倉新樹	皮膚科用薬	高久史磨(監), 堀正二, 菅野 健太郎, 門脇 孝, 乾賢一, 林昌洋	治療薬ハンド ブック2013	じほう	東京	2013	pp240-243

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戸倉新樹	Gibertばら色粃糠疹	瀧川雅治, 渡辺晋一	皮膚疾患最新 の治療 2013-2014	南江堂	東京	2013	P142
島内隆寿, 戸倉新樹	FILE008 誤診?アトピー性皮膚炎→本当は 悪性リンパ腫(禁状息 肉症)	宮地良樹	皮膚科フォト ニクスシリー ズ 誤診され ている皮膚疾 患	メディカル レビュー社	東京	2013	pp44-47
戸倉新樹	紅皮症	富田靖(監), 橋本隆,岩月 啓氏,照井正	標準皮膚科学 第10版	医学書院	東京	2013	pp119-123
戸倉新樹	皮膚悪性腫瘍/悪性リンパ腫とその類症	富田靖(監), 橋本隆,岩月 啓氏,照井正	標準皮膚科学 第10版	医学書院	東京	2013	pp380-398
戸倉新樹	抗ヒスタミン薬の抗炎 症作用はどのようなも のが期待できるのか?	宮地良樹	抗ヒスタミン 薬〜達人の処 方箋R x 〜	メディカル レビュー社	東京	2013	pp68-69
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戸倉新樹	職業性皮膚疾患の定義は?	(日本職業・ 環境アレルギ 一学会(監))	職業性アレル ギー疾患診療 ガイドライン 2013	協和企画	東京	2013	p63
戸倉新樹	職業性皮膚疾患の分類は?	日本職業・環 境アレルギー 学会(監))	職業性アレル ギー疾患診療 ガイドライン 2013	協和企画	東京	2013	pp63-64
戸倉新樹	職業性刺激性接触皮膚 炎とは?	日本職業・環 境アレルギー 学会(監))	職業性アレル ギー疾患診療 ガイドライン 2013	協和企画	東京	2013	pp64-65
戸倉新樹	接触皮膚炎ー病態と治療戦略が見えるー	田中良哉	免疫・アレルギ 一疾患イラス トレイテッド	羊土社	東京	2013	pp332-339

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# $\alpha(1,3)$ Fucosyltransferases IV and VII Are Essential for the Initial Recruitment of Basophils in Chronic Allergic Inflammation

Kazumi Saeki<sup>1</sup>, Takahiro Satoh<sup>2</sup> and Hiroo Yokozeki<sup>1</sup>

Basophils act as initiator cells for the development of IgE-mediated chronic allergic inflammation (IgE-CAI). However, detailed mechanisms of initial recruitment of basophils into the skin have yet to be clarified. Selectins mediate leukocyte capture and rolling on the vascular endothelium for extravasation. Counter-receptor activity of selectins is regulated by  $\alpha(1,3)$  fucosyltransferases (FTs) IV and VII. To clarify the contribution of selectin ligands regulated by FTs for initial basophil recruitment, IgE-CAI was induced in mice deficient in *FT-IV* and/or *FT-VII* genes. Although FT-IV(-/-) and FT-VII(-/-) mice exhibited comparable skin responses to wild-type mice, the FT-IV(-/-)/FT-VII(-/-) mice showed significantly impaired inflammation. Although the transfer of basophils to FcR $\gamma(-/-)$  mice induced IgE-CAI, this induction was completely absent when basophils from FT-IV(-/-)/FT-VII(-/-) mice were transferred. L-selectin, but not P- and E-selectin, blocking Abs inhibited skin inflammation *in vivo*. P-selectin glycoprotein-1 (PSGL-1) antibody also ameliorated skin inflammation, and basophils were bound to L-selectin in a PSGL-1-dependent manner, which was regulated by FT-IV/VII. Functional PSGL-1 generated by basophil FT-IV/VII and its subsequent binding to L-selectin could be one of the essential steps required for initial basophil recruitment and the development of IgE-CAI in mice.

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### **INTRODUCTION**

Leukocyte recruitment from the vasculature to the inflammatory sites is a multistep process. The first step of extravasation is leukocyte capture and rolling along the endothelial surfaces, a process that is mediated by selectins. P- and E-selectins on the endothelial cells contribute to the primary capture of leukocytes via binding to their ligands. Conversely, L-selectin is constitutively expressed on most types of circulating leukocytes. L-selectin binds to its ligands on activated endothelial cells (Spertini *et al.*, 1992; Luscinskas *et al.*, 1994; Tu *et al.*, 1999), and also mediates binding to leukocytes already adhering to endothelial cells (secondary capture) (Guyer *et al.*, 1996; Walcheck *et al.*, 1996).

The glycans that contribute to selectin counter-receptor activity arise through glycosylation reactions in which the terminal steps are catalyzed by  $\alpha(1,3)$  fucosyltransferases (FTs) (Lowe, 2002). Mice deficient in the *FT-VII* gene (FT-VII(-/-) mice) are characterized by absent P-, E-, and L-selectin ligand

activities (Maly *et al.*, 1996). Although the contribution of FT-IV is somewhat subtle when FT-VII is expressed (Weninger *et al.*, 2000), the inflammation-dependent leukocyte recruitment is retained in the FT-VII(-/-) mice. However, it is extinguished in the FT-IV(-/-)/FT-VII(-/-) mice, indicating that FT-IV contributes to E-, P-, and L-selectin ligand generation (Homeister *et al.*, 2001).

Basophils represent <1% of the peripheral blood leukocytes. Under physiological conditions, basophils do not reside in the peripheral tissues. However, basophils can infiltrate into the skin during inflammatory conditions (Ito et al., 2011). Despite the similarities of basophils and mast cells, recent studies have revealed unique functions for basophils, such as producing IL-4 and IL-13 (Redrup et al., 1998; Sokol et al., 2008; Watanabe et al., 2008), and functioning as antigen-presenting cells that induce Th2 cells (Sokol et al., 2009). Basophils also mediate protective immunity against helminthes and ticks (Voehringer, 2009; Wada et al., 2010), in addition to being indispensable for IgG-mediated anaphylactic reactions in mice (Tsujimura et al., 2008).

IgE-mediated chronic allergic inflammation (IgE-CAI) is a long-lasting inflammation that follows immediate-type reactions and late-phase responses. It is histopathologically characterized by numerous eosinophils and mast cells (Mukai et al., 2005; Obata et al., 2007). Although tissue basophils constitute only a minor population of total cellular infiltrate, they have a critical role in the development of IgE-CAI. After a depletion of basophils but not the mast cells, it has been

<sup>&</sup>lt;sup>1</sup>Department of Dermatology, Tokyo Medical and Dental University, Tokyo, Japan and <sup>2</sup>Department of Dermatology, National Defense Medical College, Tokorozawa, Japan

Correspondence: Takahiro Satoh, Department of Dermatology, National Defense Medical College, 3-2 Namiki, Tokyo, Tokorozawa 359-8513, Japan. E-mail: tasaderm@ndmc.ac.jp

Abbreviations: CHS, contact hypersensitivity; FT, α(1, 3) fucosyltransferase; IgE-CAI, IgE-mediated chronic allergic inflammation; mRNA, messenger RNA; PSGL-1, P-selectin glycoprotein-1; TNP, trinitrophenyl; WT, wild type

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shown that there is an almost complete abrogation of IgE-CAI (Obata *et al.*, 2007). Thus, basophils are now considered to initiate inflammation of IgE-CAI. Nevertheless, the current understanding of early events involving basophil recruitment

to the skin remains limited. This study was designed to determine the requirements of selectin ligand activity for initial basophil recruitment to the skin controlled by FT-IV and -VII during IgE-CAI.

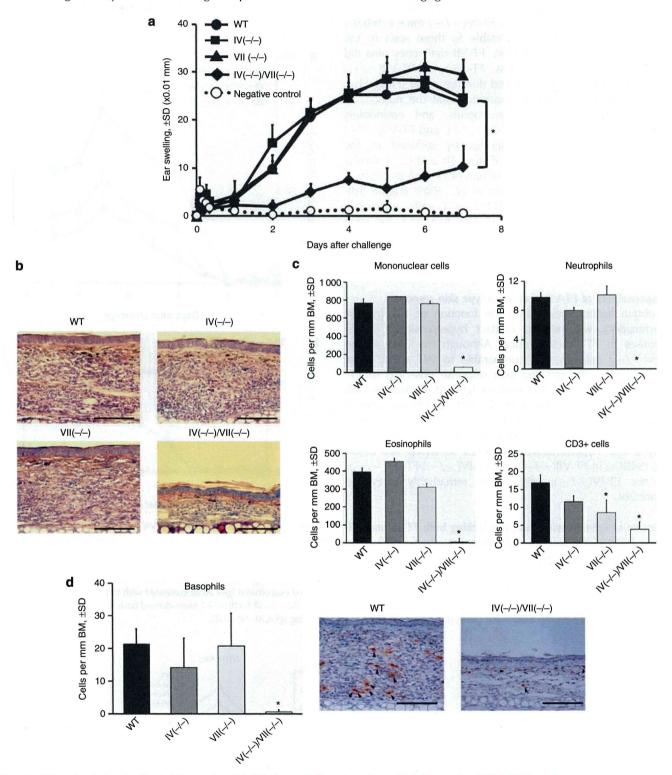


Figure 1. IgE-mediated chronic allergic inflammation (IgE-CAI) in  $\alpha(1,3)$  fucosyltransferase-IV (FT-IV)- and/or FT-VII-deficient mice. (a) IgE-CAI was induced in mice lacking FT-IV and/or FT-VII. Negative control mice were challenged with trinitrophenyl-OVA (TNP-OVA) without TNP-IgE injection. (b) Histopathological features of the skin (Giemsa's staining). (c) Cell populations in inflammatory skin. (d) Basophil numbers in inflammatory skin. Basophils were detected by mouse mast cell protease-8 mAb (arrows in the right panel). \*P<0.05 compared with wild-type (WT) mice. BM, basement membrane. Bar=100  $\mu$ m.

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### **RESULTS**

# Dependency of IgE-CAI on the collaborative functions of FT-IV and FT-VII

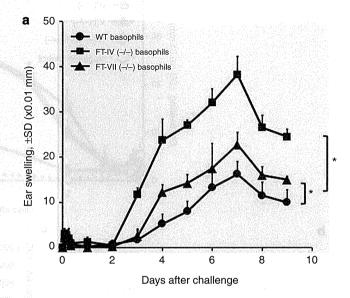
To determine selectins and FTs contribution to skin inflammation, IgE-CAI was induced in FT-IV(-/-), FT-VII(-/-), and FT-IV(-/-)/FT-VII(-/-) mice. FT-IV(-/-) mice exhibited levels of skin responses comparable to those seen in the wild-type (WT) mice. In addition, FT-VII deficiency also did not affect IgE-CAI. Nevertheless, FT-IV(-/-)/FT-VII(-/-)mice showed remarkably reduced skin responses (Figure 1a). Histological examination demonstrated that the number of dermal mononuclear cells, neutrophils, and eosinophils were similar among the WT, FT-IV(-/-), and FT-VII(-/-)mice, although they were significantly reduced in the FT-IV(-/-)/FT-VII(-/-) mice (Figure 1b and c). A similar trend was noted for the number of basophils as detected by a basophil-specific antibody (Ugajin et al., 2009) (Figure 1d). Conversely, the number of CD3 (+) T cells apparently decreased in FT-VII(-/-) mice, with this decrease even prominent in FT-IV(-/-)/FT-VII(-/-)(Figure 1c). These findings demonstrate that IgE-CAI is dependent on both FT-IV and FT-VII.

## Indispensability of FT-VII in delayed-type skin responses

To obtain further insight into the function of FTs in skin inflammation, we induced contact hypersensitivity (CHS) responses in FT-deficient mice. Although the CHS of the FT-IV(-/-) animals was comparable to WT mice, there was a significantly reduced skin response observed in FT-VII(-/-) mice, unlike that seen for IgE-CAI. Consistent with a previous report (Smithson *et al.*, 2001), CHS was almost completely absent in FT-IV(-/-)/FT-VII(-/-) mice (Figure 2a). Similarly, as compared with the WT mice, delayed-type hypersensitivity reactions to sheep red blood cells (SRBCs) in FT-VII(-/-) and FT-IV(-/-)/FT-VII(-/-), but not FT-IV(-/-), mice were remarkably alleviated (Figure 2b).

# Induction of IgE-CAI with basophils lacking both FT-IV and VII On the basis of the fact that IgE-CAI is entirely dependent on basophils (Mukai *et al.*, 2005; Obata *et al.*, 2007), we attempted to determine the contribution of selectin ligands generated by basophil FTs to the development of skin responses. Basophil transfer from WT mice to irradiated FcRy(-/-) mice lacking FcRI successfully induced IgE-CAI

(Figure 3a), which was consistent with a prior report (Mukai *et al.*, 2005). Basophil-enriched cell suspension consisted of ~20% primary basophils and ~80% other cells, including CD49b (+) natural killer (NK) cells. Nevertheless, NK cells, T cells, NKT cells, B cells, and



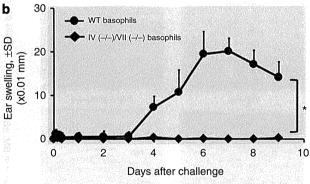


Figure 3.  $\alpha(1,3)$  Fucosyltransferase-IV/VII (FT-IV/VII) in basophils are indispensable for IgE-mediated chronic allergic inflammation (IgE-CAI). IgE-CAI was induced in FcR $\gamma(-/-)$  mice that received primary basophils from wild-type (WT), FT-IV(-/-), FT-VII(-/-), and FT-IV(-/-)/FT-VII(-/-) mice. (a) Although basophils from FT-IV(-/-) and FT-VII(-/-) mice induced exacerbated IgE-CAI as compared with WT basophils, (b) FT-IV(-/-)/FT-VII(-/-) mice-derived basophils were incapable of inducing IgE-CAI. \*P<0.05.

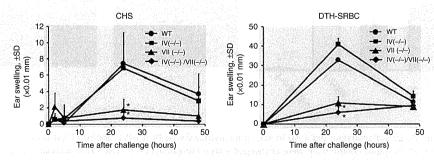


Figure 2. Delayed-type hypersensitivity (DTH) reactions in  $\alpha(1, 3)$  fucosyltransferase-IV (FT-IV)- and/or FT-VII-deficient mice. Contact hypersensitivity (CHS) and DTH to sheep red blood cells (DTH-SRBCs) were induced in mice lacking FT-IV and/or -VII. \*P < 0.05 compared with wild-type (WT) mice.

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dendritic cells are dispensable for IgE-CAI (Mukai et al., 2005), and thus the development of IgE-CAI in  $FcR\gamma(-/-)$  mice in this experiment could be exclusively mediated by primary basophils. This was also confirmed by the results that IgE-CAI in mice receiving basophil-enriched cell suspension was remarkably alleviated when recipient mice were treated with basophil-depletion antibody (Ba103, kindly provided by Dr Karasuyama (Obata et al., 2007)) (Supplementary Figure S1 online). Basophils from FT-VII(-/-) mice were also capable of inducing IgE-CAI, and interestingly there were higher induction levels as compared with those seen for the WT basophils. This exacerbation was even more marked when basophils were transferred from FT-IV(-/-) mice. Conversely, skin responses in  $FcR\gamma(-/-)$  mice that underwent transfers of primary basophils from FT-IV(-/-)/FT-VII(-/-) mice were completely absent (Figure 3b). Thus, IgE-CAI is entirely dependent on basophil selectin ligands that are collaboratively generated by FT-IV and FT-VII.

# Expression of functional selectin ligands on basophils is not sufficient for the full development of IgE-CAI

As inflammatory cells, such as T cells, neutrophils, and eosinophils, have FT-IV and/or FT-VII and are recruited to the skin in a selectin-dependent manner (Homeister et al., 2001; Smithson et al., 2001; Satoh et al., 2005), we examined the development of IgE-CAI by performing experiments designed to assess the contribution of selectin ligands generated by FT-IV/VII in cells other than basophils. WT basophils together with CD49b (-) bone marrow cells (effector cells) from either WT or FT-IV(-/-)/FT-VII(-/-)mice were transferred to irradiated FT-IV(-/-)/FT-VII(-/-)mice. Transfers with the CD49b (-) effector cells from WT mice resulted in a successful induction of IgE-CAI in FT-IV(-/-)/FT-VII(-/-) mice (Figure 4a). Although the CD49b (-) effector cells from FT-IV(-/-)/FT-VII(-/-)mice also induced IgE-CAI responses, induction levels were lower than those of the mice receiving WT mice-derived

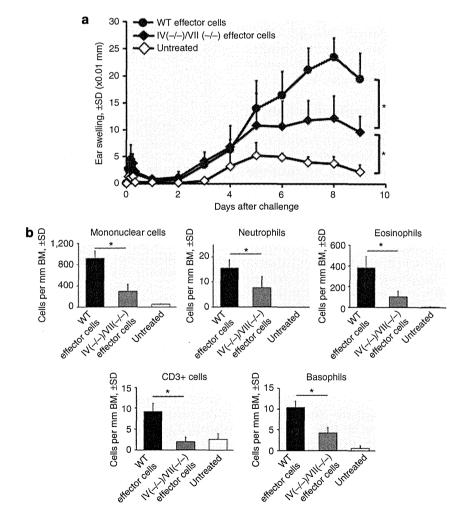


Figure 4. Selectin-dependent cooperative recruitment of basophils and effector cells. (a) Irradiated  $\alpha(1,3)$  fucosyltransferase-IV (FT-IV)(-/-)/FT-VII(-/-) mice received wild-type (WT) basophils in combination with CD49b(-) bone marrow cells (effector cells) from either WT or FT-IV(-/-)/FT-VII(-/-) mice. They were then immunized with trinitrophenyl-IgE (TNP-IgE) and challenged with TNP-OVA. The untreated group comprised FT-IV(-/-)/FT-VII(-/-) mice without cell transfer. (b) Cell populations in inflammatory skin. \*P<0.05 compared with WT effector cells. BM, basement membrane.

CD49b (-) effector cells. When cell populations from inflammatory skin were analyzed, it was shown that, even in the presence of WT basophils, there was an impairment of the recruitment of mononuclear cells, neutrophils, CD3 (+) T cells, and eosinophils in mice transferred with FT-IV(-/-)/FT-VII(-/-) mice–derived CD49b(-) effector cells (Figure 4b). More importantly, when WT basophils were cotransferred with CD49b (-) effector cells from FT-IV (-/-)/FT-VII(-/-) mice, complete recruitment into the skin was not achieved. These data suggest that selectin-dependent recruitment of the effector cells appears to be necessary for sufficient responses of IgE-CAI and effective basophil infiltration to occur, even though functional selectin ligand generation in basophils by FT-IV/VII is essential for skin inflammation.

## Binding of E- and P-selectins to basophils in vitro

Primary basophils expressed transcripts of FT-IV and FT-VII messenger RNA (mRNA; Figure 5a). This was in contrast to bone marrow–derived mast cells, which only expressed extremely low levels of FT mRNA. Although bone marrow–derived basophils had FT transcripts, the levels were much lower than those seen for the primary basophils. Flow cytometry results showed that E- and P-selectin chimeras could bind to primary basophils from WT but not to FT-IV(-/-)/FT-VII(-/-) mice *in vitro* (Figure 5b).

Blockade of E- and/or P-selectins and the amelioration of IgE-CAI Given the evidence that basophil expression of both E- and P-selectin ligands was dependent upon FT-IV/VII expression, we next attempted to determine the contribution of E- and P-selectins to the actual basophil recruitment. To determine this, we initially examined the effects of blocking Abs against selectins on the development of IgE-CAI. Unexpectedly, we found that blocking of either the E- (clone 10E9.6, BD Bioscience Pharmingen (San Jose, CA), 100 µg per mouse, intravenous ) or P- (clone RB40.34, BD Bioscience Pharmingen, 100 µg per mouse, intravenous) selectins did not result in amelioration of IgE-CAI (Supplementary Figure S2 online). Similarly, dual blocking of P- and E-selectins by coadministration of these two Abs also failed to suppress IgE-CAI. These results were in a striking contrast to prior reports demonstrating that the same antibody clones against P- and E-selectins clearly alleviated eotaxin-induced eosinophil accumulation (Satoh et al., 2005) and cutaneous arthus reaction (Yanaba et al., 2003).

# Role of P-selectin glycoprotein-1 and L-selectin interaction in basophil recruitment and development of IgE-CAI

Leukocytes express L-selectin, which then interacts with inducible endothelial ligands and contributes to leukocyte rolling (Spertini *et al.*, 1991, 1992; Luscinskas *et al.*, 1994; Tu *et al.*, 1999). Counter-receptor activity of the L-selectin ligand on endothelial cells has been shown to be dependent on the modification by FT-IV and/or VII (Maly *et al.*, 1996; Tu *et al.*, 1999). However, the interaction of basophil L-selectin with ligands modified by endothelial FTs did not seem to be part of the essential pathway for the development of IgE-CAI, as

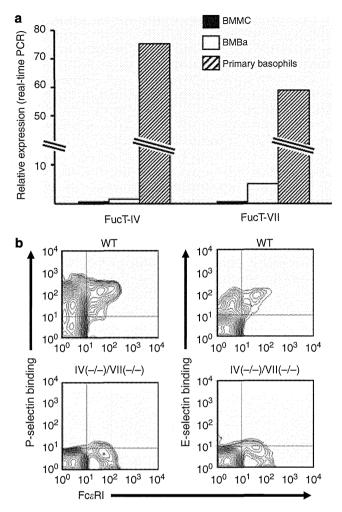


Figure 5. Expression of α(1, 3) fucosyltransferase (FT) messenger RNA (mRNA) and FT-dependent selectin binding in basophils. (a) Primary basophils were subjected to further purification by positive selection with CD123 (purity >99%). Transcripts for FT-IV and FT-VII mRNA were quantified by real-time PCR. (b) Binding of soluble P- and E-selectins to primary basophils assessed by flow cytometry. BMBa, bone marrow–derived basophil; BMMCs, bone marrow–derived mast cells.

basophils from WT mice were able to successfully induce skin inflammation in FT-IV(-/-)/FT-VII(-/-) mice lacking counter-receptor activity for L-selectin on endothelial cells (Figure 4a). Prior evidence has also shown that leukocyte PGSL-1, which is a major ligand for P-selectin, can function as a counter-receptor for L-selectin in an FT-dependent manner, thereby contributing to secondary tethering(Guyer et al., 1996; Walcheck et al., 1996). These findings led us to hypothesize that modification of P-selectin glycoprotein-1 (PSGL-1) by FTs in basophils combined with the subsequent binding to L-selectin was an essential pathway for the development of IgE-CAI. To test this hypothesis, we initially confirmed that primary basophils from both WT and FT-IV(-/-)/FT-VII(-/-) mice expressed PSGL-1 and L-selectin on their cell surface (Figure 6a). PSGL-1 Ab (4RA10, BD Bioscience Pharmingen) almost completely inhibited the in vitro binding

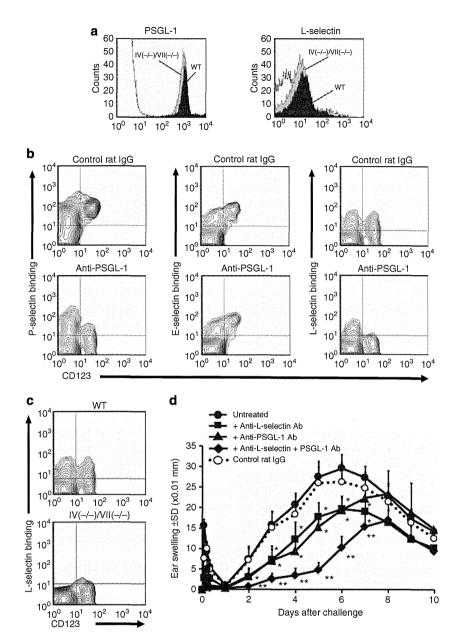


Figure 6. Basophil P-selectin glycoprotein-1 (PSGL-1)–L-selectin interaction involvement in IgE-mediated chronic allergic inflammation (IgE-CAI). (a) PSGL-1 and L-selectin expressions on basophils. (b) PSGL-1 Ab effect on selectin binding to primary basophils. (c) L-selectin binding to basophils from wild-type (WT) and  $\alpha(1, 3)$  fucosyltransferase-IV (FT-IV)(-/-)/FT-VII(-/-) mice. (d) Effects of L-selectin and/or PSGL-1 Abs on IgE-CAI in WT mice. \*P<0.05 compared with control IgG. \*\*P<0.05 compared with L-selectin or PSGL-1 Ab alone.

of both the L- and P-selectins to the WT basophils (Figure 6b). Counter-receptor activity of PSGL-1 for L-selectin appears to be dependent on FT-VI/VII, as L-selectin failed to bind the primary basophils from FT-IV(-/-)/FT-VII(-/-) mice (Figure 6c). We then assessed whether blocking L-selectin could ameliorate IgE-CAI. As expected, the administration of the L-selectin blocking Ab (MEL-14, eBioscience, San Diego, CA) partly but significantly inhibited IgE-CAI. Similarly, PSGL-1 blocking Ab (4RA10) also inhibited IgE-CAI. When there was concomitant administration of these two Abs, further suppression of skin responses was also observed (Figure 6d).

#### **DISCUSSION**

Extravasation and recruitment of basophils to the skin are an essential step for the development of IgE-CAI (Mukai *et al.*, 2005; Obata *et al.*, 2007). This study examined the FT-IV/VII-dependent basophil recruitment and induction of IgE-CAI.

Although a single deficiency of the *FT-IV* or *FT-VII* genes did not affect IgE-CAI, greatly impaired skin responses were seen in the FT-IV(-/-)/FT-VII(-/-) mice. To elucidate the contribution of FTs in basophils during IgE-CAI, we transferred basophils into FcR $\gamma$ (-/-) mice lacking Fc $\epsilon$ RI. Unlike those from WT mice, basophils from the FT-IV(-/-)/FT-VII(-/-)

mice failed to induce IgE-CAI in FcR $\gamma(-/-)$  mice. These data confirm the critical contribution of basophils for the development of IgE-CAI (Mukai *et al.*, 2005) and suggest that impaired skin responses in FT-IV(-/-)/FT-VII(-/-) mice is largely due to the inability to recruit basophils into the skin.

Leukocytes other than basophils may also require selectins for their recruitment to the skin during IgE-CAI. Our results indicated that the transfer of WT basophils with basophildepleted bone marrow cells (effector cells) from FT-IV(-/-)/FT-VII(-/-) mice were not able to fully develop IgE-CAI as compared with the WT basophils that were cotransferred with WT effector cells (Figure 4a). In addition, WT basophils themselves were not effectively recruited into the skin when in the presence of effector cells from FT-IV(-/-)/FT-VI(-/-) mice. Thus, it appears that some effector cells require FT-IV/VII-dependent modification of selectin ligands in order to be recruited to the skin. In addition, these cells appeared to increase the effectiveness of basophil recruitment to the skin. Once basophils are recruited into the skin, they can promote the accumulation of other effector cells. These cells, in turn, may then assist in further basophil recruitment into the skin.

Basophils from FT-IV(-/-)/FT-VII(-/-) mice did not show avidity to soluble E- and P-selectins, which indicates that these are dependent on the FT function (Figure 5b). However, IgE-CAI was unexpectedly not suppressed after the use of blocking Abs against P- and E-selectins, despite the inability of basophils from FT-IV(-/-)/VII(-/-) mice to induce IgE-CAI (Figure 3b). Conversely, blockade of L-selectin resulted in a moderate suppression of IgE-CAI. It is possible that PSGL-1 on basophils could be a counter-receptor of the basophil L-selectin. On the basis of our results that showed that WT basophils could successfully induce IgE-CAI in FT-IV(-/-)/FT-VII(-/-) mice, it appears that endothelial L-selectin ligands might not be essential for basophil recruitment. We demonstrated that L-selectin bound PSGL-1 in vitro, and this binding was dependent on the basophil FT-IV/VII. In addition, when we blocked PSGL-1, this alleviated IgE-CAI in vivo. These were similar to the level of suppression that was seen when using anti-L-selectin Ab. Thus, FT-mediated modification of basophil PSGL-1 and the binding to L-selectin appear to be one of the important steps required for the development of IgE-CAI.

Intriguingly, we also noted that coadministration of anti-PSGL-1 and L-selectin Abs was able to more efficiently inhibit IgE-CAI than the injection of a single Ab. Although we have not been able to completely assure that optimal doses of each antibody were used, this suggests that an adhesion pathway other than PSGL-1-L-selectin interaction might contribute to the development of IgE-CAI. Several lines of evidence have suggested that an L-selectin-dependent leukocyte–leukocyte interaction facilitates the subsequent direct interaction of leukocytes with endothelial selectins, which leads to the amplification of initial leukocyte recruitment (Alon *et al.*, 1996; Walcheck *et al.*, 1996; Sperandio *et al.*, 2003). In this respect, endothelial L-selectin ligands and P-selectin might assist in the capture and rolling of basophils and effector cells

on the endothelial cells following the PSGL-1-L-selectin interaction, although the blocking of P-selectin alone is not sufficient for the inhibition of basophil recruitment and the development of IgE-CAI. The roles of E-, P-, and L-selectins in leukocyte capture and/or rolling on endothelial cells have been shown to be partially redundant, and these three selectins can also function synergistically (Ley *et al.*, 1993, 1995; Ley and Tedder, 1995; Lowe, 2002).

IgE-CAI offers a unique mouse model of skin inflammation, in that it is dependent on IgE and FceRI of basophils, but independent of FceRI of mast cells and other cells that usually have central roles in some human allergic inflammations (von Bubnoff et al., 2003). In addition, the characteristics of mouse basophils differ from those of human basophils in many respects (Lee and McGarry, 2007). Another difference between humans and mice is seen in the regulatory functions of FTs. Human FT-VII, but not FT-IV, modifies PSGL-1 of leukocytes, leading to the expression of cutaneous lymphocyte-associated antigen, which acts as a functional selectin ligand and skin-homing receptor (Kieffer et al., 2001). On the other hand, murine leukocytes express barely detectable levels of cutaneous lymphocyte-associated antigen epitope despite the expression of FT-VII, but still efficiently bind to E- and P-selectins. Murine FT-VII appears to fucosylate only a few quite specific glycans that interact preferentially with selectins (Kobzdej et al., 2002). Thus, it would be difficult to consider the present findings for IgE-CAI in mice as directly applicable to human allergic skin diseases.

Collectively, basophil recruitment and development of IgE-CAI are entirely dependent on collaborative control by FT-IV and VII in the basophils. L-selectin binding to basophil PSGL-1 modified by the FTs could be a central event that ultimately leads to the subsequent inflammatory steps of IgE-CAI.

### MATERIALS AND METHODS

#### Mice

C57BL/6 mice were purchased from Sankyo Labo Service (Tokyo, Japan). FcR $\gamma$  chain(-/-) C57BL/6 mice (Takai *et al.*, 1994) were kindly provided by Dr Takai of Tohoku University, Japan. FT-IV(-/-) mice, FT-VII(-/-) mice, and FT-IV(-/-)/FT-VII(-/-) mice (Maly *et al.*, 1996; Homeister *et al.*, 2001) were originally established at the University of Michigan (Dr Lowe), with colonies maintained at Case Western Reserve University (Dr Myers), which provided animals to our department. The use of animals was in full compliance with the Committee for Animal Experiments of Tokyo Medical and Dental University.

## **Antibodies**

Isotype-matched control Ab (rat IgG2aκ), rat anti-CD16/CD32 (2.4G2), biotinylated anti-CD49b (Dx5), and PE-labeled anti-PSGL-1 (P-selectin glycoprotein-1) (2PH1) Abs were from BD Bioscience Pharmingen. PE/Cy5-labeled anti-L-selectin Ab (MEL-14) was from BioLegend (San Diego, CA). FITC-conjugated anti-CD49b (Dx5), FITC- and PE-conjugated anti-FcεRI Ab (MAR-1), FITC- and PE-labeled anti-mouse CD123 (IL-3Rα), and PE-labeled anti-c-kit (ACK2), biotinylated anti-c-kit (2B8) Abs were purchased from eBioscience. Anti-CD3e (M-20) was from Santa Cruz Biotechnology