

**Figure 6. SPCS1 forms a complex with NS2 and E2.** (A) Lysates of cells, which were co-transfected with Core-p7, FLAG-NS2, and SPCS1-myc expression plasmids, were immunoprecipitated with anti-myc or anti-FLAG antibody. The resulting precipitates and whole cell lysates used in IP were examined by immunoblotting using anti-E2, anti-FLAG, or anti-myc antibody. An empty plasmid was used as a negative control. (B) Cells were transfected with Core-p7 expression plasmid in the presence or absence of SPCS1-myc expression plasmid. The cell lysates of the transfected cells were immunoprecipitated with anti-myc antibody. The resulting precipitates and whole cell lysates used in IP were examined by immunoblotting using anti-E2 or anti-myc antibody. An empty plasmid was used as a negative control. The bands corresponding to immunoglobulin heavy chain are marked by an asterisk. (C) Cells were co-transfected with Core-p7 and SPCS1-myc expression plasmids. The cell lysates of the transfected cells were immunoprecipitated with anti-myc antibody. The resulting precipitates and whole cell lysates used in IP were examined by immunoblotting using anti-E2 or anti-myc antibody. (D) Huh7.5.1 cells were transfected with SPCS1 siRNA or control siRNA at a final concentration of 20 nM. After 24 h, Huh7.5.1 cells were then co-transfected with FLAG-NS2 and Core-p7 expression plasmids. The lysates of transfected cells were immunoprecipitated with anti-FLAG antibody, followed by immunoblotting with anti-FLAG and anti-E2 antibodies. Immunoblot analysis of whole cell lysates was also performed. Intensity of E2 bands was quantified, and the ratio of immunoprecipitated E2 to E2 in cell lysate was shown. Similar results were obtained in 2 independent experiments. (E) KD#31 cells and parental Huh-7 cells were co-transfected with FLAG-NS2, Core-p7, and NS3 expression plasmids. The lysates of transfected cells were immunoprecipitated with anti-FLAG antibody followed by immunoblotting with anti-FLAG, anti-E2, and anti-NS3 antibodies. Immunoblot analysis of whole cell lysates was also performed. The ratio of immunoprecipitated E2 or NS3 to E2 or NS3 in cell lysate, respectively, were shown.

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nucleus [37]. They further screened a human liver cDNA library using NS2 with deletion of the N-terminal TM domain, and CIDE-B protein, a member of the CIDE family of apoptosis-inducing factors, was identified. However, whether CIDE-B is involved in the HCV lifecycle and/or viral pathogenesis is unclear. de Chasse et al. reported several cellular proteins as potential NS2 binding proteins using NS2 with N-terminal deletion as a bait [38]. Involvement of these proteins in the HCV lifecycle is also unclear. In our study, to screen for NS2-binding partners using full-length NS2 as a bait, we utilized a split-ubiquitin yeast two-hybrid system that allows for the identification of interactions between full-length integral membrane proteins or between a full-length membrane-associated protein and a soluble protein [39]. SPCS1 was identified as a positive clone of an NS2-binding protein, but proteins that have been reported to interact with NS2 were not selected from our screening.

SPCS1 is a component of the signal peptidase complex that processes membrane-associated and secreted proteins in cells. The mammalian signal peptidase complex consists of five subunits, SPCS1, SPCS2, SPCS3, SEC11A, and SEC11C [27]. Among them, the functional role of SPCS1 is still unclear, and SPCS1 is considered unlikely to function as a catalytic subunit according to membrane topology [40]. The yeast homolog of SPCS1, Spclp, is also known to be nonessential for cell growth and enzyme activity [28,41]. Interestingly, these findings are consistent with the results obtained in this study. Knockdown of SPCS1 did not impair processing of HCV structural proteins (Fig. 4A) or secretion of apoE and albumin (Fig. 4B and C), which are regulated by ER membrane-associated signal peptidase activity. The propagation of JEV, whose structural protein regions are cleaved by signal peptidase, was also not affected by the knockdown of SPCS1 (Fig. 3B). SPCS1, SPCS2, and SPCS3 are among the host factors that function in HCV production identified from genome-wide siRNA screening [42]. It seemed that knockdown of SPCS1 had a higher impact on the later stage of viral infection compared to either SPCS2 or SPCS3, which are possibly involved in the catalytic activity of the signal peptidase.

Further analyses to address the mechanistic implication of SPCS1 on the HCV lifecycle revealed that SPCS1 knockdown impaired the assembly of infectious viruses in the cells, but not cell entry, RNA replication, or release from the cells (Fig. 5). We thus considered the possibility that the SPCS1-NS2 interaction is important for the role of NS2 in viral assembly. Several studies have reported that HCV NS2 is associated biochemically or genetically with viral structural proteins as well as NS proteins [10,18–25]. As an intriguing model, it has been proposed that NS2 functions as a key organizer of HCV assembly and plays a key role in recruiting viral envelope proteins and NS protein(s) such as NS3 to the assembly sites in close proximity to lipid droplets [21]. The interaction of NS2 with E2 has been shown by use of an HCV genome encoding tagged-NS2 protein in virion-producing cells. Furthermore, the selection of an assembly-deficient NS2 mutation located within its TM3 for pseudoreversion leads to a rescue mutation in the TM domain of E2, suggesting an in-membrane interaction between NS2 and E2 [21]. Another study identified two classes of NS2 mutations with defects in virus assembly; one class leads to reduced interaction with NS3, and the other, located in the TM3 domain, maintains its interaction with NS3 but shows impaired interaction between NS2 and E1-E2 [20]. However, the precise details of the NS2-E2 interaction, such as direct protein-protein binding or participating host factors, are unknown. Our results provide evidence that SPCS1 has an important role in the formation of the NS2-E2 complex by its interaction with both NS2 and E2, most likely via their transmembrane domains, including

TM3 of NS2. As knockdown of SPCS1 reduced the interaction of NS2 and E2 as shown in Fig. 6D and E, it may be that SPCS1 contributes to NS2-E2 complex formation or to stabilizing the complex. Based on data obtained in this study, we propose a model of the formation of an E2-SPCS1-NS2 complex at the ER membrane (Fig. 7).

In summary, we identified SPCS1 as a novel NS2-binding host factor required for HCV assembly by split-ubiquitin membrane yeast two-hybrid screening. Our data demonstrate that SPCS1 plays a key role in the E2-NS2 interaction via formation of an E2-SPCS1-NS2 complex. These findings provide clues for understanding the molecular mechanism of assembly and formation of infectious HCV particles.

## Materials and Methods

### Split ubiquitin-based yeast two-hybrid screen

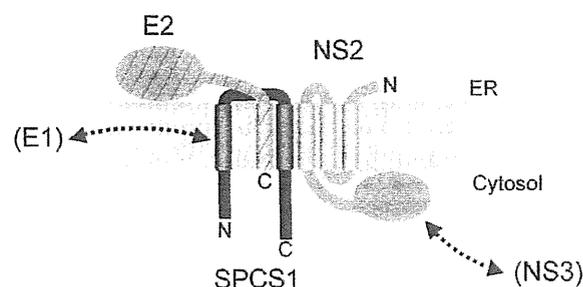
A split-ubiquitin membrane yeast two-hybrid screen was performed to identify possible NS2 binding partners. This screening system (DUALmembrane system; Dualsystems Biotech, Schlieren, Switzerland) is based on an adaptation of the ubiquitin-based split protein sensor [26]. The full-length HCV NS2 gene derived from the JFH-1 strain [29] was cloned into pBT3-SUC bait vector to obtain bait protein fused to the C-terminal half of ubiquitin (NS2-Cub) along with a transcription factor. Prey proteins generated from a human liver cDNA library (Dualsystems Biotech) were expressed as a fusion to the N-terminal half of ubiquitin (NubG). Complex formation between NS2-Cub and NubG-protein from the library leads to cleavage at the C-terminus of reconstituted ubiquitin by ubiquitin-specific protease(s) with consequent translocation of the transcription factor into the nucleus. Library plasmids were recovered from positive transformants, followed by determining the nucleotide sequences of inserted cDNAs, which were identified using the BLAST algorithm with the GenBank database.

### Cell culture

Human embryonic kidney 293T cells, and human hepatoma Huh-7 cells and its derivative cell lines Huh7.5.1 [43] and Huh7-25 [36], were maintained in Dulbecco's modified Eagle medium supplemented with nonessential amino acids, 100 U of penicillin/ml, 100 µg of streptomycin/ml, and 10% fetal bovine serum (FBS) at 37°C in a 5% CO<sub>2</sub> incubator.

### Plasmids

Plasmids pCAGC-NS2/JFH1am and pHHJFH1am were previously described [33]. The plasmid pCAGC-p7/JFHam, having



**Figure 7. A proposed model for a complex consisting of NS2, SPCS1 and E2 associated with ER membranes.**  
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adaptive mutations in E2 (N417S) and p7 (N765D) in pCAG/C-p7 [44], was constructed by oligonucleotide-directed mutagenesis.

To generate the NS2 expression plasmid pCAG F-NS2 and the NS2-deletion mutants, cDNAs encoding the full-length or parts of NS2 possessing the FLAG-tag and spacer sequences (MDYKDDDDKGGGGS) were amplified from pCAGC-NS2/JFH1am by PCR. The resultant fragments were cloned into pCAGGS. For the NS2-NS3 expression plasmid pEF F-NS2-3, a cDNA encoding the entire NS2 and the N-terminal 226 amino acids of NS3 with the N-terminal FLAG-tag sequence as above was amplified by PCR and was inserted into pEF1/myc-His (Invitrogen, Carlsbad, CA). The plasmid pEF F-NS2-3 H956A, having a defective mutation in the protease active site within NS2, was constructed by oligonucleotide-directed mutagenesis.

To generate the NS3 expression plasmid pCAGN-HANS3JFH1, a cDNA encoding NS3 with an HA tag at the N terminus, which was amplified by PCR with pHHJFHam as a template, was inserted downstream of the CAG promoter of pCAGGS.

To generate the SPCS1-expressing plasmid pCAG-SPCS1-myc and its deletion mutants, cDNAs encoding all of or parts of SPCS1 with the Myc tag sequence (EQKLISEEDL) at the C-terminus, which was amplified by PCR, was inserted into pCAGGS. pSilencer-shSPCS1 carrying a shRNA targeted to SPCS1 under the control of the U6 promoter was constructed by cloning the oligonucleotide pair 5'-GATCCGCAATAGTTGGATTTATCTTTCAAGAGAAGATAAATCCAACCTATTGCTTTTITGGGAA-3' and 5'-AGCTTTTCCAAAAAAGCAATAGTTGGATT-TATCTTCTCTTGAAAGATAAATCCAACCTATTGCGG-3' between the BamHI and HindIII sites of pSilencer 2.1-U6 hygro (Ambion, Austin, TX). To generate a construct expressing shRNA-resistant SPCS1 pSPCS1-sh<sup>r</sup>, a cDNA fragment coding for SPCS1, in which the 6 bp within the shRNA targeting region (5'-GCAATAGTTGGATTTATCT-3') was replaced with GCTATTGTCGGCTTCATAT that causes no aa change, was amplified by PCR. The resulting fragment was confirmed by sequencing and then cloned into pCAGGS.

Full-length SPCS1 and N-terminal region of NS2 (aa 1-94) were amplified by PCR and cloned onto EcoRI and HindIII sites of phmKGN-MN and phmKGC-MN, which encode the mKG fragments (CoralHue Fluo-chase Kit; MBL, Nagoya, Japan), designated as pSPCS1-mKG(N) and pNS2-mKG(C), respectively. Transmembrane domain of the E1 to E2 was also amplified by PCR and cloned onto EcoRI and HindIII sites of phmKGC-MN. To avoid the cleavage of E2-mKG(C) fusion protein in the cells, last alanine of the E2 protein was deleted. Positive control plasmids for mKG system, pCONT-1 and pCONT-2, which encode p65 partial domain from NF- $\kappa$ B complex fused to mKG(N) and p50 partial domain from NF- $\kappa$ B complex fused to mKG(C) respectively, were supplied from MBL. For PLA experiments, cDNA for SPCS1 d2-myc with the V5 tag at the N-terminus was amplified by PCR, and inserted into pCAGGS. For expression of HCV E2, cDNA from E1 signal to the last codon of the transmembrane domain of the E2, in which part of the hypervariable region-1 (aa 394-400) were replaced with FLAG-tag and spacer sequences (DYKDDDDKGGG), was amplified by PCR, and inserted into pCAGGS. For expression of FLAG-core, cDNAs encoding Core (aa 1-152) possessing the FLAG-tag and spacer sequences (MDYKDDDDKGGGGS) were amplified from pCAGC191 [45] by PCR. The resultant fragments were cloned into pCAGGS.

#### DNA transfection

Monolayers of 293T cells were transfected with plasmid DNA using FuGENE 6 transfection reagent (Roche, Basel, Switzerland) in accordance with the manufacturer's instructions. Huh-7,

Huh7.5.1, and Huh7-25 cells were transfected with plasmid DNA using TransIT LT1 transfection reagent (Mirus, Madison, WI).

#### PLA

The assay was performed in a humid chamber at 37°C according to the manufacturer's instructions (Olink Bioscience, Uppsala, Sweden). Transfected 293T cells were grown on glass coverslips. Two days after transfection, cells were fixed with 4% paraformaldehyde in phosphate-buffered saline (PBS) for 20 min, then blocked and permeabilized with 0.3% Triton X-100 in a nonfat milk solution (Block Ace; Snow Brand Milk Products Co., Sapporo, Japan) for 60 min at room temperature. Then the samples were incubated with a mixture of mouse anti-FLAG monoclonal antibody M2 and rabbit anti-V5 polyclonal antibody for 60 min, washed three times, and incubated with plus and minus PLA probes. After washing, the ligation mixture containing connector oligonucleotide was added for 30 min. The washing step was repeated, and amplification mixture containing fluorescently labeled DNA probe was added for 100 min. Finally, the samples were washed and mounted with DAPI mounting medium. The signal representing interaction was analyzed by Leica TCS SPE confocal microscope.

#### mKG system

The assay was performed according to the manufacturer's instructions (CoralHue Fluo-chase Kit; MBL). 293T cells were transfected by a pair of mKG fusion constructs. Twenty-four hours after transfection, cell were fixed and stained with DAPI. The signal representing interaction was analyzed by Leica TCS SPE confocal microscope.

#### Gene silencing by siRNA

The siRNAs were purchased from Sigma-Aldrich (St. Louis, MO) and were introduced into the cells at a final concentration of 10 to 30 nM using Lipofectamine RNAiMAX (Invitrogen). Target sequences of the siRNAs were as follows: SPCS1 #1 (5'-CAGUUCGGGUGGACUGUCU-3'), SPCS1 #2 (5'-GCAAUA GUUGGAUUUAUCU-3'), SPCS1 #3 (5'-GAUGUUUCAGG-GAAUUUU-3'), SPCS1 #4 (5'-GUUAUGGCCGGGAUUUG-CUU-3'), claudin-1 (5'-CAGUCAAUCCAGGUACGA-3'), PI4K (5'-GCAAUGUGCUUCGCGAGAA-3') and scrambled negative control (5'-GCAAGGGAAACCGUGUAAU-3'). Additional control siRNAs for SPCS1 were as follows: C911-#2 (5'-GCAAUAGUaccAUUUUAUCU-3'), C911-#3 (5'-GAUGUUU-CuccGAAUUUU-3') and C911-#4 (5'-GUUAUGGCCgccAUU UGCUU-3'). Bases 9 through 11 of the siRNAs replaced with their complements were shown in lower cases.

#### Establishment of a stable cell line expressing the shRNA

Huh-7 cells were transfected with pSilencer-SPCS1, and drug-resistant clones were selected by treatment with hygromycin B (Wako, Tokyo, Japan) at a final concentration of 500  $\mu$ g/ml for 4 weeks.

#### Virus

HCVtcp and HCVcc derived from JFH-1 having adaptive mutations in E2 (N417S), p7 (N765D), and NS2 (Q1012R) were generated as described previously [33]. The rAT strain of JEV [46] was used to generate virus stock.

#### Antibodies

Mouse monoclonal antibodies against actin (AC-15) and FLAG (M2) were obtained from Sigma-Aldrich (St. Louis, MO). Mouse

monoclonal antibodies against flavivirus group antigen (D1-4G2) were obtained from Millipore (Billerica, MA). Rabbit polyclonal antibodies against FLAG and V5 were obtained from Sigma-Aldrich. Rabbit polyclonal antibodies against SPCS1, claudin-1, PI4K and myc were obtained from Proteintech (Chicago, IL), Life Technologies (Carlsbad, CA), Cell Signaling (Danvers, MA) and Santa Cruz Biotechnology (Santa Cruz, CA), respectively. An anti-apoE goat polyclonal antibody was obtained from Millipore. Rabbit polyclonal antibodies against NS2 and NS3 were generated with synthetic peptides as antigens. Mouse monoclonal antibodies against HCV Core (2H9) and E2 (8D10-3) and rabbit polyclonal antibodies against NS5A and JEV are described elsewhere [47].

### Titration

To determine the titers of HCVcc, Huh7.5.1 cells in 96-well plates were incubated with serially-diluted virus samples and then replaced with media containing 10% FBS and 0.8% carboxymethyl cellulose. Following incubation for 72 h, the monolayers were fixed and immunostained with the anti-NS5A antibody, followed by an Alexa Fluor 488-conjugated anti-rabbit secondary antibody (Invitrogen). Stained foci were counted and used to calculate the titers of focus-forming units (FFU)/ml. For intracellular infectivity of HCVcc, the pellets of infected cells were resuspended in culture medium and were lysed by four freeze-thaw cycles. After centrifugation for 5 min at 4,000 rpm, supernatants were collected and used for virus titration as above. For titration of JEV, Huh7.5.1 cells were incubated with serially-diluted virus samples and then replaced with media containing 10% FBS and 0.8% carboxymethyl cellulose. After a 24 h incubation, the monolayers were fixed and immunostained with a mouse monoclonal anti-flavivirus group antibody (D1-4G2), followed by an Alexa Fluor 488-conjugated anti-mouse secondary antibody (Invitrogen).

### Immunoprecipitation

Transfected cells were washed with ice-cold PBS, and suspended in lysis buffer (20 mM Tris-HCl [pH 7.4] containing 135 mM NaCl, 1% TritonX-100, and 10% glycerol) supplemented with 50 mM NaF, 5 mM Na<sub>2</sub>VO<sub>4</sub>, and complete protease inhibitor cocktail, EDTA free (Roche). Cell lysates were sonicated for 10 min and then incubated for 30 min at 4°C, followed by centrifugation at 14,000 × *g* for 10 min. The supernatants were immunoprecipitated with anti-Myc-agarose beads (sc-40, Santa Cruz Biotechnology) or anti-FLAG antibody in the presence of Dynabeads Protein G (Invitrogen). The immunocomplexes were precipitated with the beads by centrifugation at 800 × *g* for 30 s, or by applying a magnetic field, and then were washed four times with the lysis buffer. The proteins binding to the beads were boiled with SDS sample buffer and then subjected to SDS polyacrylamide gel electrophoresis (PAGE).

### Immunoblotting

Transfected cells were washed with PBS and lysed with 50 mM Tris-HCl, pH 7.4, 300 mM NaCl, 1% Triton X-100. Lysates were then sonicated for 10 min and added to the same volume of SDS sample buffer. The protein samples were boiled for 10 min, separated by SDS-PAGE, and transferred to polyvinylidene difluoride membranes (Millipore). After blocking, the membranes were probed with the primary antibodies, followed by incubation with peroxidase-conjugated secondary antibody. Antigen-antibody complexes were visualized by an enhanced chemiluminescence detection system (Super Signal West Pico Chemiluminescent

Substrate; PIERCE, Rockford, IL) according to the manufacturer's protocol and were detected by an LAS-3000 image analyzer system (Fujifilm, Tokyo, Japan).

### Albumin measurement

To determine the human albumin level secreted from cells, culture supernatants were collected and passed through a 0.45-μm pore filter to remove cellular debris. The amounts of human albumin were quantified using a human albumin ELISA kit (Bethyl Laboratories, Montgomery, TX) according to the manufacturer's protocol.

### Supporting Information

**Figure S1** Effects of SPCS1-siRNAs and the C911 mismatch control siRNAs on the expression of SPCS1 and production of HCV. (A) Huh7.5.1 cells were transfected with either siRNAs targeted for SPCS1 (SPCS1-#2, -#3, and -#4), scrambled control siRNA (Scrambled) or C911 siRNA in which bases 9 through 11 of each SPCS1 siRNA were replaced with their complements (C911-#2, -#3, and -#4) at a final concentration of 15 nM, and were infected with HCVcc at a multiplicity of infection (MOI) of 0.05 at 24 h post-transfection. Expression levels of endogenous SPCS1 and actin in the cells were examined by immunoblotting using anti-SPCS1 and anti-actin antibodies at 3 days post-infection. (B) Infectious titers of HCVcc in the supernatant of the infected cells were determined at 3 days postinfection.

(TIF)

**Figure S2** 293T cells were transfected with E2 expression plasmid in the presence or absence of SPCS1-myc expression plasmid. The cell lysates of the transfected cells were immunoprecipitated with anti-myc antibody. The resulting precipitates and whole cell lysates used in IP were examined by immunoblotting using anti-E2 or anti-myc antibody. An empty plasmid was used as a negative control.

(TIF)

**Figure S3** Interaction of HCV E2 with SPCS1 in mammalian cells. (A) 293T cells were transfected with indicated plasmids. 2 days posttransfection, cells were fixed and permeabilized with Triton X-100, then subjected to in situ PLA (Upper) or immunofluorescence staining (Lower) using anti-FLAG and anti-V5 antibodies. (B) Detection of the SPCS1-E2 interaction in transfected cells using the mKG system. 293T cells were transfected by indicated pair of mKG fusion constructs. Twenty-four hours after transfection, cell were fixed and stained with DAPI, and observed under a confocal microscope.

(TIF)

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### Author Contributions

Conceived and designed the experiments: RS TS. Performed the experiments: RS MM. Analyzed the data: RS KW HA TS. Contributed reagents/materials/analysis tools: YM TW. Wrote the paper: RS TS.

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# Interleukin-1 and Tumor Necrosis Factor- $\alpha$ Trigger Restriction of Hepatitis B Virus Infection via a Cytidine Deaminase Activation-induced Cytidine Deaminase (AID)\*

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**Background:** Cytokines and host factors triggering innate immunity against hepatitis B virus (HBV) are not well understood.

**Results:** IL-1 and TNF $\alpha$  induced cytidine deaminase AID, an anti-HBV host factor, and reduced HBV infection into hepatocytes.

**Conclusion:** IL-1/TNF $\alpha$  reduced host susceptibility to HBV infection through AID up-regulation.

**Significance:** Proinflammatory cytokines modulate HBV infection through a novel innate immune pathway involving AID.

Virus infection is restricted by intracellular immune responses in host cells, and this is typically modulated by stimulation of cytokines. The cytokines and host factors that determine the host cell restriction against hepatitis B virus (HBV) infection are not well understood. We screened 36 cytokines and chemokines to determine which were able to reduce the susceptibility of HepaRG cells to HBV infection. Here, we found that pretreatment with IL-1 $\beta$  and TNF $\alpha$  remarkably reduced the host cell susceptibility to HBV infection. This effect was mediated by activation of the NF- $\kappa$ B signaling pathway. A cytidine deaminase, activation-induced cytidine deaminase (AID), was up-regulated by both IL-1 $\beta$  and TNF $\alpha$  in a variety of hepatocyte cell lines and primary human hepatocytes. Another deaminase APOBEC3G was not induced by these proinflammatory cytokines. Knockdown of AID expression impaired the anti-HBV effect of IL-1 $\beta$ , and overexpression of AID antagonized HBV infection, suggesting that AID was one of the responsible factors for the anti-HBV activity of IL-1/TNF $\alpha$ . Although AID induced hypermutation of HBV DNA, this activity was dispensable for the anti-

HBV activity. The antiviral effect of IL-1/TNF $\alpha$  was also observed on different HBV genotypes but not on hepatitis C virus. These results demonstrate that proinflammatory cytokines IL-1/TNF $\alpha$  trigger a novel antiviral mechanism involving AID to regulate host cell permissiveness to HBV infection.

The intracellular immune response can eliminate pathogens from a host, and host cells possess different mechanisms to counteract viral infection depending on the virus type. Human immunodeficiency virus (HIV) infection is restricted by cellular proteins designated as restriction factors, including APOBEC3G (A3G),<sup>3</sup> TRIM5 $\alpha$ , tetherin/BST-2, and SAMHD1 (1, 2). All of these factors can be induced by stimulation with interferon (IFN). Hepatitis C virus (HCV) is eliminated by type I and III IFNs derived from dendritic cells or infected hepatocytes (3–6). In hepatocytes, this process involves a series of antiviral factors that are downstream genes of IFN, IFN-stimulated genes (ISGs). Influenza virus spread and virulence is inhibited by cytokines such as IFNs and TNF $\alpha$ . Responsive genes for these mechanisms include IFN-induced cellular Mx proteins that are dynamin-like GTPases (7, 8). However, these cytokine-induced antiviral immune responses are poorly understood in hepatitis B virus (HBV) infection.

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<sup>3</sup> The abbreviations used are: A3G, APOBEC3G; AID, activation-induced cytidine deaminase; HBV, hepatitis B virus; HCV, hepatitis C virus; ISG, IFN-stimulated gene; QNZ, 6-amino-4-(4-phenoxyphenylethylamino)quinazolinone; GEQ, genome equivalent; PHH, primary human hepatocyte; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; ISRE, interferon sensitivity-responsive element; cccDNA, covalently closed circular DNA.

## Anti-HBV Activity of IL-1 and TNF $\alpha$ Mediated by AID

HBV infection is a worldwide health problem affecting more than 350 million people and is a major cause of the development of liver cirrhosis and hepatocellular carcinoma (9–11). During the course of infection, a number of cytokines and chemokines are up-regulated in HBV-infected patients, including IFN $\alpha$ / $\gamma$ / $\lambda$ , TNF $\alpha$ , IL-1, IL-6, IL-10, IL-12, IL-15, and IL-8 (12–15). Some of these cytokines are reported to suppress HBV replication (3, 16–21). In particular, type I, II, and III IFNs suppress the replication of HBV *in vitro* and *in vivo* (19, 20, 22–26). Although one of the downstream genes of IFN, A3G, has the potential to reduce HBV replication (27–34), it is still under discussion whether this protein is responsible for the anti-HBV activity of type I IFN, because it has been previously reported by Trono and co-workers (28, 35) that the induction of A3G does not explain the IFN-induced inhibition of HBV replication. Moreover, these studies were carried out using an HBV transgene that only reproduces a portion of the whole HBV life cycle, mainly focusing on intracellular HBV replication.

Here, we screened for cytokines and chemokines that affected HBV infection in HepaRG cells, a human hepatocyte cell line susceptible to HBV infection and reproducing the whole HBV life cycle (36, 37). IL-1 and TNF $\alpha$  decreased the host cell permissiveness to HBV infection, and this effect was at least partly mediated by the induction of activation-induced cytidine deaminase (AID). The anti-HBV activity of IL-1/TNF $\alpha$  was mechanistically different from that of IFN $\alpha$ . This study presents the activity of IL-1/TNF $\alpha$  to suppress HBV infection into hepatocytes independent of the effect on immune cells and the physiological role of AID in this machinery. Moreover, as far as we know, this is the first report to show the AID function to inhibit the infection of human pathogenic virus.

### EXPERIMENTAL PROCEDURES

**Reagents**—All cytokines were purchased from PeproTech or R & D Systems. Heparin was obtained from Mochida Pharmaceutical. Lamivudine, PD98059, SP600125, SB203580, and Bay11-7082 were obtained from Sigma. Entecavir was obtained from Santa Cruz Biotechnology. BMS-345541 and 6-amino-4-(4-phenoxyphenylethylamino)quinazoline (QNZ) were purchased from Merck.

**Cell Culture**—HepaRG cells (Biopredic) were cultured with Williams' medium E (Invitrogen) supplemented with 2 mM L-glutamine, 200 units/ml penicillin, 200  $\mu$ g/ml streptomycin, 10% FBS, 5  $\mu$ g/ml insulin (Wako), 20 ng/ml EGF (PeproTech), 50  $\mu$ M hydrocortisone (Sigma), and 2% DMSO (Sigma). HepG2, HepAD38 (kindly provided by Dr. Seeger at Fox Chase Cancer Center) (38), and HepG2.2.15 cells (a kind gift from Dr. Urban at Heidelberg University) (39) were cultured with DMEM/F-12 + GlutaMAX (Invitrogen) supplemented with 10 mM HEPES (Invitrogen), 200 units/ml penicillin, 200  $\mu$ g/ml streptomycin, 10% FBS, 50  $\mu$ M hydrocortisone, and 5  $\mu$ g/ml insulin in the presence (HepAD38 and HepG2.2.15) or absence (HepG2) of 400  $\mu$ g/ml G418 (Nacalai Tesque). HepAD38 cells were cultured with 0.3  $\mu$ g/ml tetracycline when terminating HBV induction. Huh-7.5.1 cells (kindly provided from Dr. Chisari at Scripps Research Institute) were cultured as described previously (40). Primary human hepatocytes (PHH) isolated from urokinase-type plasminogen activator transgen-

ic/SCID mice inoculated with PHH (PhoenixBio) or purchased from Lonza were cultured with DMEM supplemented with 20 mM HEPES, 100 units/ml penicillin, 100  $\mu$ g/ml streptomycin, 10% FBS, and 44 mM NaHCO<sub>3</sub> or with 1 mM pyruvate, nonessential amino acids, 20 mM HEPES, 200 units/ml penicillin, 200  $\mu$ g/ml streptomycin, 10% FBS, 0.25  $\mu$ g/ml insulin (Wako), 5 ng/ml EGF, and 50 nM dexamethasone.

**HBV Preparation and Infection**—HBV used in this study was mainly derived from HepAD38 cells, which is classified as genotype D (38). Media from HepAD38 cells at days 7–31 post-induction of HBV by depletion of tetracycline were recovered every 3 days. Media were cleared through a 0.45- $\mu$ m filter and precipitated with 10% PEG8000 and 2.3% NaCl. The precipitates were washed and resuspended with medium at  $\sim$ 200-fold concentration. The HBV DNA was quantified by real time PCR. HBV genotype A and C in Fig. 7B was recovered from the media of HepG2 cells transfected with the plasmid pHBV/Aeus and pHBV/C-AT (41).

HepaRG cells were infected with HBV at 2000 (Fig. 7B) or 6000 (other figures) genome equivalent (GEq)/cell in the presence of 4% PEG8000 for 16 h as described previously (36). Urban and co-workers (42) reported that more than 10<sup>3</sup> GEq/cell amount of HBV derived from HepAD38 or HepG2.2.15 cells (*i.e.* 1.25–40  $\times$  10<sup>4</sup> GEq/cell) as inoculum was required for efficient infection into HepaRG cells. The anti-HBV effect of IL-1/TNF $\alpha$  shown in this study was also observed when inoculated with HBV at 300 GEq/cell (data not shown).

**Extraction of DNA and RNA**—HBV DNA was extracted from the cells or from the medium using a DNA kit (Qiagen) according to the manufacturer's protocol. Total RNA was recovered with RNeasy mini kit (Qiagen) according to the manufacturer's protocol.

**Real Time PCR and RT-PCR**—HBV DNA was quantified by real time PCR analysis using the primer set 5'-ACTCACC-AACCTCCTGTCCCT-3' and 5'-GACAAACGGGCAACAT-ACCT-3' and probe 5'-carboxyfluorescein (FAM)-TATCG-CTGGATGTGTCTGCGGCGT-carboxytetramethylrhodamine (TAMRA)-3' (43). The PCR was performed at 50 °C for 2 min, 94 °C for 10 min, and 50 cycles of 94 °C for 15 s and 60 °C for 1 min. Detection of cccDNA was achieved using 5'-CGTCTGTGCCTTCTCATCTGC-3' and 5'-GCACAG-CTTGAGGCTTGAA-3' as primers and 5'-CTGTAGGC-ATAAA'TGGT (MGB)-3' as a probe (44). This primer-probe set theoretically detected neither relaxed circular DNA nor HBV DNA integrated into host genome but can capture cccDNA as described previously (44). For quantification of cellular mRNA, cDNA was synthesized from extracted RNA using SuperScriptIII (Invitrogen), followed by PCR with TaqMan Gene Expression Master Mix (Applied Biosystems) and primer-probe set (TaqMan Gene Expression Assay, Applied Biosystems) or with Power SYBR Green PCR Master Mix (Applied Biosystems) and 5'-AAATGTC-CGCTGGGCTAAGG-3' and 5'-GGAGGAAGAGCAATT-CCACGT-3' as primers for *AID*.

RT-PCR was performed as described previously (45) using a one-step RNA PCR kit (Takara). Primers for amplifying each gene were as follows: 5'-CTCTGAGGTTTAGCATTTCA-3' and 5'-CTCCAGGTCCAAAATGAATA-3' for *cIAP*; 5'-GCA-

GATTTATCAACGGCTTT-3' and 5'-CAGTTTTCCACCA-CAACAAA-3' for XIAP; 5'-TAGCCAACATGTCTCACA-GAC-3' and 5'-TCTTCTACCACTGGTTTCATGC-3' for ISG56; 5'-GCCTTTTCATCCAAATGGAATTC-3' and 5'-GAAATCTGTTCTGGGCTCATG-3' for PKR; and 5'-CCATG-GAGAAGGCTGGGG-3' and 5'-CAAAGTTGTCATGGATG-ACC-3' for GAPDH, respectively.

**ELISA**—HBs protein was quantified by ELISA using plates incubated at 4 °C overnight with a sheep anti-HBs antibody at 1:5000 dilution (Maxisorp nunc-immuno plate, Nunc catalog no. 439454) followed by coating with 0.2% BSA, 0.02% Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>, 1× PBS at 4 °C until use. Samples were incubated with the plates for 2 h and after washing with TBST four times, horseradish peroxidase-labeled rabbit anti-HBs antibody was added for 2 h. The substrate solution (HCV core ELISA kit: Ortho) was reacted for 30 min before the A<sub>450</sub> values were measured.

**Indirect Immunofluorescence Analysis**—Indirect immunofluorescence analysis was performed essentially as described previously (45). After fixation with 4% paraformaldehyde and permeabilization with 0.3% Triton X-100, an anti-HBc antibody (DAKO, catalog no. B0586) was used as the primary antibody.

**MTT Assay**—The MTT assay was performed as described previously (46).

**Immunoblot Analysis**—Immunoblot analysis was performed as described previously (47). The polyclonal antibody against AID was generated using a peptide derived from AID protein as an immunogen as described previously for preparation of the anti-AID antibody 1 (48). The specificity of the antibody was described previously (48, 49).

**Lentiviral Vector-mediated Gene Transduction**—Lentivirus carrying shRNAs was prepared with 293T cells transfected with expression plasmids for HIV-1 Gag-Pol, VSV G, and shRNAs (sh-control, sh-cyclophilin A, sh-AID(1), sh-AID(2); Mission shRNA) (Sigma) with Lipofectamine 2000 (Invitrogen). Recovered lentiviral vector was transduced into HepaRG cells followed by selection with 1.5  $\mu$ g/ml puromycin. Lentivirus overexpressing AID, AID mutant, A3G, or the control lentivirus was recovered using expression plasmids for HIV-1 Gag-Pol, Rev, VSV G, and the corresponding expression vector as described previously (50).

**Southern Blot Analysis**—Southern blot was performed as described previously (41). After digestion of free nucleic acids with DNase I and RNase A, cell lysates were digested with proteinase K, and HBV DNA in the core particles was extracted with phenol/chloroform, followed by isopropyl alcohol precipitation. Probe was prepared by cutting pHBV/D-IND60 (41) with SacII and BspHI to generate a full-length HBV DNA probe and labeled with AlkPhos direct labeling reagents (GE Healthcare). Labeled bands were visualized with CDP-star detection reagent (GE Healthcare).

**Quantification of Nucleocapsid-associated HBV RNA**—After digestion of free nucleic acids with DNase I and RNase A, nucleocapsid was precipitated with PEG8000 (41). Total RNA was then extracted from the resuspended precipitates. HBV RNA was quantified by real time RT-PCR with 5'-TCCCTCGCCTCGCAGACG-3' and 5'-GTTTCCCACCTTAT-

GAGTC-3' as primers with Power SYBR Green PCR Master Mix (Applied Biosystems).

**Co-immunoprecipitation Assay**—Co-immunoprecipitation assay was essentially performed as described (45).

**Differential DNA Denaturation PCR**—Differential DNA denaturation PCR was performed as described previously (51).

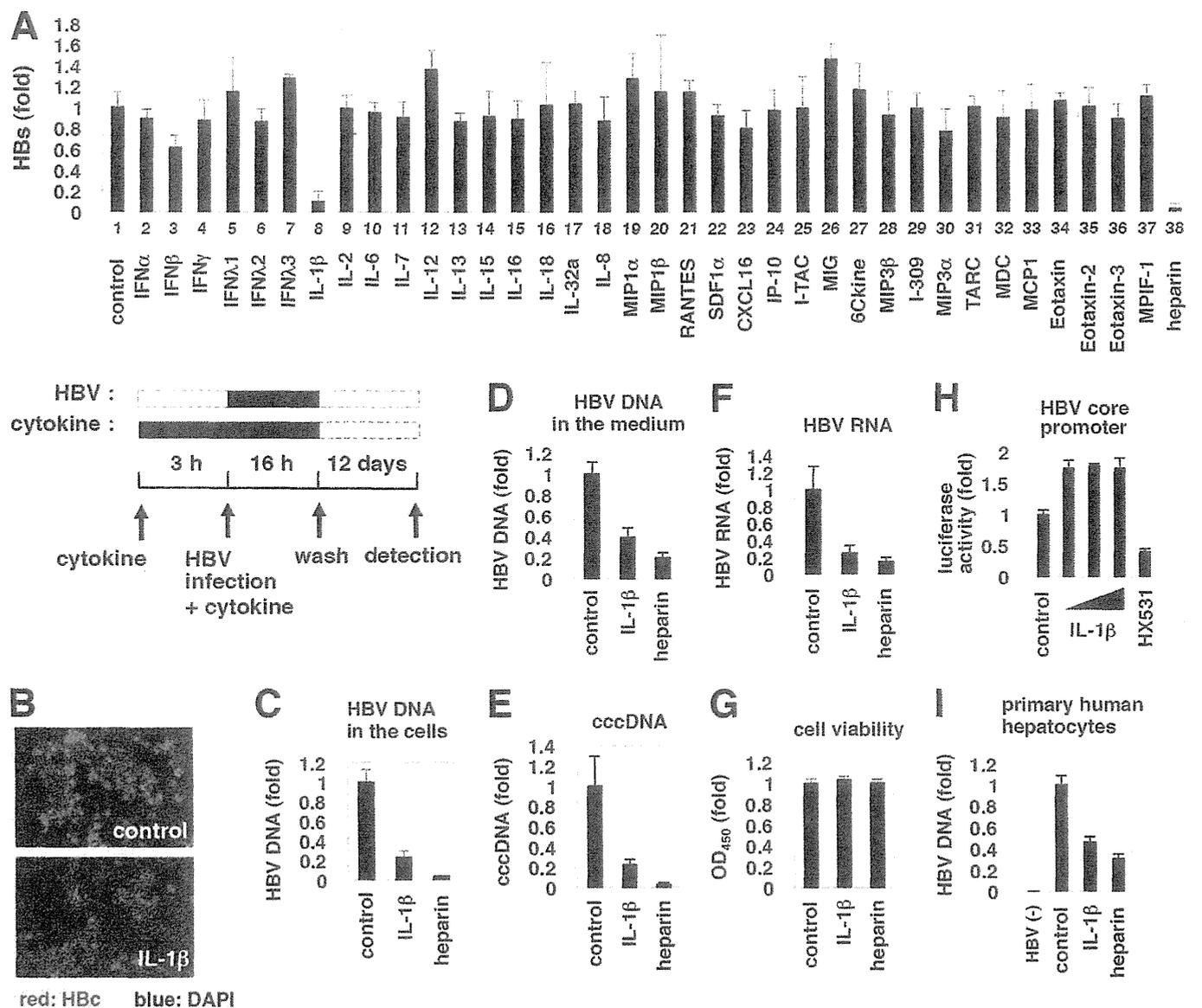
**Reporter Assay**—DNA transfection was performed with pNF- $\kappa$ B-luc or pISRE-TA-luc (Stratagene) and pRL-TK (Promega), which express firefly luciferase driven by NF- $\kappa$ B or ISRE and *Renilla* luciferase by herpes simplex virus thymidine kinase promoter, respectively, and Polyethylenimine Max (Polysciences Inc., catalog no. 24765). After compound or cytokine treatment, cells were lysed, and luciferase activities were measured as described previously (52). A reporter carrying HBV core promoter was constructed by inserting the DNA fragment (1413–1788 nucleotide number) of HBV DNA (D-IND60) into pGL4.28 vector (Promega) (41). In the reporter assay using this construct (Fig. 1H), HX531, a retinoid X receptor antagonist was used as a positive control as retinoid X receptor was involved in the transcription from the core promoter (53).

## RESULTS

**IL-1 Reduced Host Cell Susceptibility to HBV Infection**—To evaluate the effect of cytokines and chemokines on susceptibility to HBV infection, we treated HepaRG cells (36) with cytokines for 3 h prior to and 16 h during HBV infection, followed by culture without stimuli for an additional 12 days (Fig. 1A, lower scheme). Heparin, a competitive inhibitor of HBV attachment (54), was used as a positive control and decreased secretion of the viral envelope surface protein (HBs) from HBV-infected cells (Fig. 1A, upper graph, lane 38), which suggests a successful HBV infection in this experiment. Examination of 36 cytokines and chemokines revealed that IL-1 $\beta$  drastically decreased protein secretion from HBs (Fig. 1A, upper graph, lane 8). Although IFNs had a strong anti-HBV effect by a continuous treatment after HBV infection (Fig. 3C, panel b, and data not shown), they had only a limited effect in this screening where cytokines were only pretreated and cotreated with HBV (Fig. 1A, lanes 2–7). HBc protein expression (Fig. 1B) and HBV DNA (Fig. 1C) in the cells and medium (Fig. 1D) were significantly decreased by treatment with IL-1 $\beta$  without cytotoxicity (Fig. 1G). HBV cccDNA and HBV RNA was also decreased in infected cells treated with IL-1 $\beta$  (Fig. 1, E and F). IL-1 $\beta$  did not decrease HBV core promoter activity at least in HepG2 cells (Fig. 1H). These results suggest that IL-1 $\beta$  suppressed HBV infection to HepaRG cells. IL-1 $\beta$  did not decrease the expression of sodium taurocholate cotransporting polypeptide (*NTCP*), a recently reported HBV entry receptor (data not shown) (55). Similar results were obtained using primary human hepatocytes (Fig. 1J).

**NF- $\kappa$ B Signaling Was Critical for Anti-HBV Activity**—As shown in Fig. 2A, IL-1 $\beta$  suppressed HBV infection in a dose-dependent manner. This anti-HBV effect was reversed by cotreatment with a neutralizing antibody for the IL-1 receptor, IL-1RI (Fig. 2B), suggesting that receptor engagement was required for anti-HBV activity. IL-1Ra is a natural antagonist that associates with IL-1RI but does not trigger downstream signal transduc-

## Anti-HBV Activity of IL-1 and TNF $\alpha$ Mediated by AID

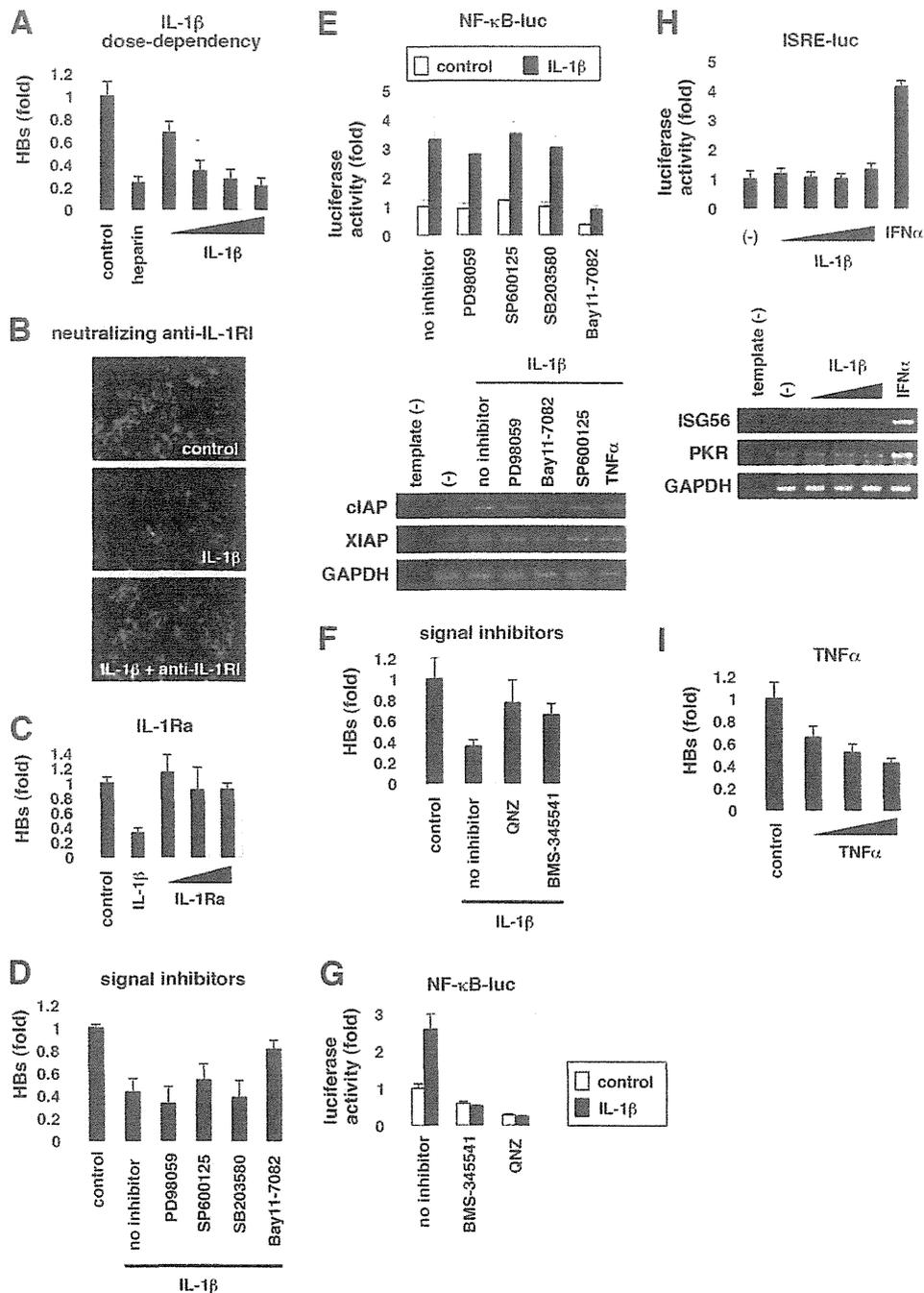


**FIGURE 1. Suppression of HBV infection by IL-1 $\beta$ .** *A*, upper graph, HepaRG cells were pretreated with cytokines at 100 ng/ml (except for IFN $\alpha$  and IFN $\beta$  at 100 IU/ml) or heparin at 25 units/ml as a positive control or were left untreated (*control*) for 3 h and then infected with HBV in the presence of each stimuli for 16 h. After washing, cells were cultured in normal growth medium for 12 days. HBs protein secreted into the medium was quantified by ELISA. Lower scheme indicates the treatment procedure for HepaRG cells. Black and dashed line boxes indicate the periods with and without treatment, respectively. *B–G* and *I*, HepaRG cells (*B–G*) or PHH (*I*) were treated as shown in *A* with or without 100 ng/ml IL-1 $\beta$  or 25 units/ml heparin as a positive control. HBc protein in the cells (*red*) was detected by indirect immunofluorescence analysis, and the nucleus was stained with DAPI (*blue*) at 12 days post-infection (*B*). HBV DNA (*C* and *I*), cccDNA (*E*), and HBV RNA (*F*) in the cells as well as HBV DNA in the medium (*D*) were detected. Cell viability was quantified by MTT assay (*G*). *HBV*(–) in *I* indicates uninfected cells. All of the data, except in *I*, are based on the average of three independent experiments. *I* shows the average results from one representative experiment, but the reproducibility of the data were confirmed in three independent experiments. *H*, reporter plasmid carrying the HBV core promoter was transfected with HepG2 cells and then treated with or without IL-1 $\beta$  (1, 10, and 100 ng/ml) and an retinoid X receptor antagonist HX531 as a positive control for 6 h. Luciferase activity was measured.

tion (56). Treatment with IL-1Ra did not decrease HBV infectivity (Fig. 2C), suggesting that signal transduction triggered by IL-1 was required for anti-HBV activity.

To identify the signal transduction pathway essential for anti-HBV activity, we treated HepaRG cells with PD98059, SP600125, SB203580, and Bay11-7082, which are inhibitors for MEK, JNK, p38, and NF- $\kappa$ B, respectively (57). As shown in Fig. 2D, only cotreatment with Bay11-7082 significantly removed the anti-HBV effect of IL-1 $\beta$ . Luciferase assay and RT-PCR analysis indicated that Bay11-7082, but not other inhibitors, blocked the transactivation of NF- $\kappa$ B (Fig. 2E, upper panels) and NF- $\kappa$ B downstream genes, *cIAP* and *XIAP* (Fig. 2E, lower

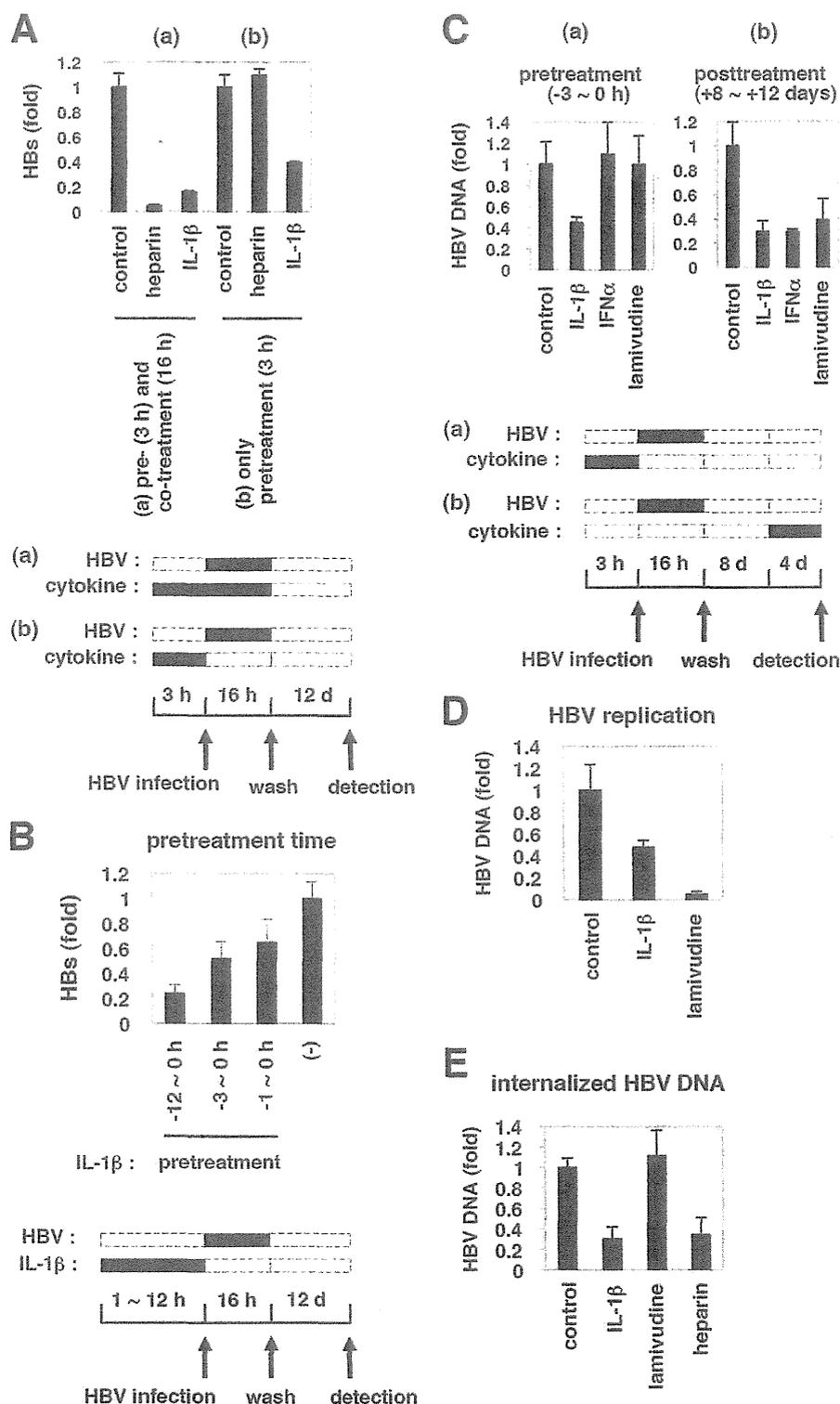
panels). Additional NF- $\kappa$ B inhibitors, BMS-345541 and QNZ (Fig. 2G), also reversed the anti-HBV effect of IL-1 $\beta$  (Fig. 2F). These data suggest a critical role for NF- $\kappa$ B activation in the anti-HBV activity. Additionally, IL-1 $\beta$  did not augment the activity of interferon sensitivity-responsive element (ISRE) and mRNAs for ISGs, *ISG56*, and double-stranded RNA-dependent protein kinase (*PKR*) in HepaRG cells (Fig. 2H), suggesting that the anti-HBV activity is independent of ISG up-regulation. TNF $\alpha$ , another cytokine that activates NF- $\kappa$ B signaling (Fig. 2E, lower panels), also inhibited HBV infection (Fig. 2I). Thus, NF- $\kappa$ B activation in host hepatocytes was critical for the anti-HBV activity of proinflammatory cytokines.



**FIGURE 2. NF- $\kappa$ B activation triggered by IL-1 and TNF $\alpha$  was critical for anti-HBV activity.** A–D, F, and I, HepaRG cells were left untreated (control) or treated with varying concentrations of IL-1 $\beta$  (1, 10, 30, and 100 ng/ml) or 25 units/ml heparin (A), with 30 ng/ml IL-1 $\beta$  together with or without a neutralizing anti-IL-1RI antibody at 20  $\mu$ g/ml (B), with 10 ng/ml IL-1 $\beta$  or varying concentrations of IL-1Ra (10, 30, and 100 ng/ml) (C), with 3 ng/ml IL-1 $\beta$  together with or without PD98059, SP600125, SB203580, or Bay11-7082 (D), or QNZ or BMS-345541 (F), or with TNF $\alpha$  (10, 100, and 300 ng/ml) (I) according to the treatment schedule shown in Fig. 1A. HBV infection was monitored by HBs protein secretion into the medium in A, C, D, F, and I and with Hbc protein in the cells in B, E, G, and H. NF- $\kappa$ B (E and G) and ISRE activity (H) were measured by reporter assay in the cells transfected with the reporter plasmid expressing Luciferase driven from five tandem repeats of NF- $\kappa$ B elements (E, upper graph, and G) or ISRE (H, upper graph) or by RT-PCR in the cells (E and H, lower panels) upon signaling inhibitors used in D and F together with or without IL-1 $\beta$  (E and G), or upon IL-1 $\beta$  (10, 30, and 100 ng/ml) or IFN $\alpha$  100 IU/ml as a positive control (H) for 6 h. The white and black bars in the upper graph of E and G show the data in the absence or presence of IL-1 $\beta$ , respectively. Bands for mRNA for cIAP, XIAP, and GAPDH (E) or ISG56, PKR, or GAPDH (H) are presented in the lower panels. All of the data are based on averages of three independent experiments.

**Early Phase of HBV Infection as Well as HBV Replication Were Impaired by IL-1 Treatment**—Although heparin, an attachment inhibitor, could block HBV infection only if added together with the HBV inoculum, pretreatment with IL-1 $\beta$  before HBV infection was sufficient to show anti-HBV activity (Fig. 3A, panel b). This activity was amplified by a prolonged

treatment time of up to 12 h (Fig. 3B). Intriguingly, HBV cellular DNA was also reduced by IL-1 $\beta$  treatment following HBV infection (Fig. 3C, panel b). In contrast, IFN $\alpha$  was not effective by pretreatment (Figs. 3C, panel a, and 1A), although it did decrease HBV DNA by treatment after HBV infection (Fig. 3C, panel b), consistent with previous reports that IFN $\alpha$  can sup-



**FIGURE 3. Defining the steps of the HBV life cycle targeted by IL-1 $\beta$ .** *A*, HepaRG cells were pretreated with IL-1 $\beta$  or heparin for 3 h and then infected with HBV in the presence (*A, panel a*) or absence (*A, panel b*) of IL-1 $\beta$  or heparin for 16 h. HBV infection was monitored with HBs protein secretion from the infected cells. Only pretreatment with IL-1 $\beta$  and not heparin could inhibit HBV infectivity. *d*, day. *B*, HepaRG cells were pretreated with IL-1 $\beta$  or left untreated (-) for the indicated time (*h*) and infected with HBV without IL-1 $\beta$ . Anti-HBV activity was amplified by a prolonged treatment time. *C, panel a*, HepaRG cells were pretreated with 10 ng/ml IL-1 $\beta$ , 100 IU/ml IFN $\alpha$ , or 1  $\mu$ M lamivudine for 3 h, followed by infection with HBV for 16 h in the absence of cytokines (*pretreatment*). *C, panel b*, HepaRG cells were infected with HBV for 16 h without pretreatment. After washing out the input virus, cells were cultured in normal medium for the first 8 days and then cultured with IL-1 $\beta$ , IFN $\alpha$ , or lamivudine for the following 4 days (*post-treatment*). HBV DNA in the cells was measured by real time PCR. IL-1 $\beta$  showed an anti-HBV activity in both pretreatment and post-treatment, although an anti-HBV effect of IFN $\alpha$  was seen only with post-treatment. *D*, HepAD38 cells were treated with 100 ng/ml IL-1 $\beta$  or 1  $\mu$ M lamivudine, or left untreated for 6 days in the absence of tetracycline. HBV replication was evaluated by measurement of HBV DNA in the medium. *E*, HepaRG cells were pretreated with IL-1 $\beta$ , lamivudine, or heparin for 3 h or left untreated and infected with HBV for 16 h in the presence or absence of each compound. After trypsinization and extensive washing of the cells, cellular DNA was immediately recovered to detect HBV DNA. HBV DNA at 16 h post-infection was decreased by treatment with IL-1 $\beta$  but not lamivudine.

press HBV replication (19, 20, 26). Thus, the anti-HBV activity of IL-1 $\beta$  is likely to be mechanistically different from that of IFN $\alpha$ .

The HBV life cycle can be divided into at least two phases as follows: 1) the early phase of infection that includes attachment, entry, nuclear import, and cccDNA formation; and 2) the late phase representing HBV replication, including transcription, assembly, reverse transcription, DNA synthesis, and viral release (58). The early phase of HBV infection is not supported, but HBV DNAs persistently replicate in HepAD38 cells in the presence of tetracycline (38). IL-1 $\beta$  decreased the HBV DNA levels in HepAD38 cells (Fig. 3D), suggesting suppression of HBV replication. In addition, to examine the early phase preceding HBV replication, we infected HepaRG cells with HBV in the presence of IL-1 $\beta$  for 16 h and then immediately recovered cellular DNA in the trypsinized cells for quantification of HBV DNA (Fig. 3E). This procedure likely detected HBV DNA that had been internalized and evaded the host restriction before initiation of HBV replication because lamivudine showed no effect on the amount of DNA detected (Fig. 3E). In this experiment, IL-1 $\beta$  significantly decreased HBV DNA (Fig. 3E). cccDNA was also decreased by IL-1 $\beta$ , suggesting that the early phase of HBV infection before cccDNA formation was also interrupted by IL-1 $\beta$ .

**IL-1 and TNF $\alpha$  Induced the Expression of AID**—The innate immune pathway against HBV infection remains largely unknown. Recently, accumulating evidence suggested that several APOBEC family proteins, especially A3G, suppressed HBV replication when overexpressed (27–33). In contrast, there was no report available suggesting the anti-HBV function of other restriction factors against HIV, TRIM5 $\alpha$ , tetherin/BST-2, and SAMHD1. We then investigated APOBEC family proteins as a candidate for an anti-HBV effector. The APOBEC family includes APOBEC1 (A1), A2, A3s, A4, and AID (59). Because some of these proteins are reported to be up-regulated in cytokine-stimulated hepatocytes (27, 28, 60, 61), we examined the expression of these genes in cells treated with IL-1 $\beta$ , TNF $\alpha$ , and IFN $\alpha$  as a control for 12 h. The mRNA levels of A1, A2, and A3A were below the detection threshold. A3G and A3F mRNA were significantly expressed in HepaRG cells, and their expression levels were remarkably increased by IFN $\alpha$  treatment (Fig. 4A), as observed in other reports (27, 28, 61). IL-1 $\beta$  and TNF $\alpha$  did not significantly up-regulate A3s, and only AID was up-regulated 6–10-fold by both cytokines (Fig. 4A). Induction of A3s by both IL-1 $\beta$  and TNF $\alpha$  was not observed at any time point examined until 12 h (data not shown). In contrast, induction of AID mRNA by IL-1 $\beta$  and TNF $\alpha$  was conserved in human hepatocyte cell lines, such as HepG2 and FLC4 cells, and in primary human hepatocytes (Fig. 4B). AID protein production was also increased in primary human hepatocytes by treatment with IL-1 $\beta$  and TNF $\alpha$  (Fig. 4C). This AID induction by IL-1 $\beta$  was suggested to be NF- $\kappa$ B-dependent, because the up-regulation of AID mRNA was canceled by addition of NF- $\kappa$ B inhibitors, Bay11-7082 or QNZ (Fig. 4D).

**AID Played a Significant Role in the IL-1-mediated restriction of HBV**—To examine the function of AID during HBV infection, we transduced AID ectopically into HepaRG cells using a lentiviral vector (Fig. 5A, left panel). The susceptibility of these

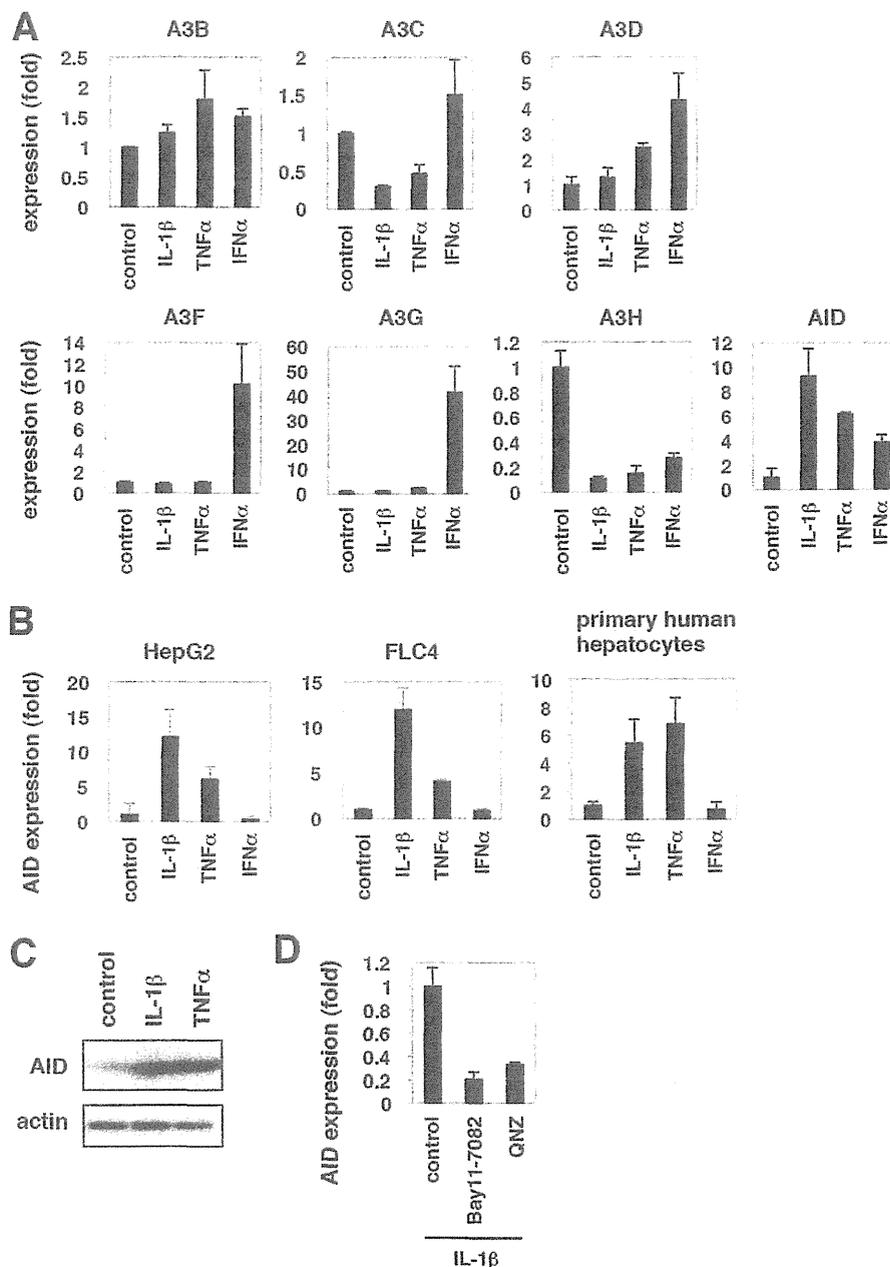
AID-overexpressing cells to HBV was decreased by approximately one-third compared with the parental or empty vector-transduced HepaRG cells (Fig. 5A, right panel), suggesting that AID can restrict HBV infection. An AID mutant AID(M139V), with reported diminished activity to support class switching (48), also decreased the susceptibility to HBV infection, although the reduction in HBV susceptibility was moderate compared with the case of the wild type AID (Fig. 5B).

To examine the relevance of endogenous AID in the anti-HBV activity of IL-1, we transduced a lentiviral vector carrying a short hairpin RNA (shRNA) against AID (sh-AID) or a non-relevant protein cyclophilin A (Fig. 5C), and we observed the anti-HBV activity of IL-1 $\beta$  in these cells. IL-1 $\beta$  decreased HBV infection in the control and sh-cyclophilin A-transduced cells by ~3.0-fold as determined by HBs secretion (Fig. 5D, lanes 1 and 2, black bars). In contrast, anti-HBV activity of IL-1 $\beta$  was limited to only 1.6–1.7-fold in the cells transduced with sh-AIDs (Fig. 5D, lanes 3 and 4, black bars). Such relieved anti-HBV activity following AID knockdown was not observed in the case for heparin treatment (Fig. 5D, lanes 1–4, gray bars). Similar results were obtained by monitoring intracellular HBV DNA after infection (data not shown). Although the anti-HBV effect of IL-1 $\beta$  was not completely blunted, these data suggest that AID plays a significant role in mediating the anti-HBV effect of IL-1 $\beta$ .

Similar observations were obtained in HBV-replicating cells overexpressing AID (Fig. 5, E and F). Core particle-associated HBV DNA in HepG2 cells transfected with an HBV-encoding plasmid was decreased by overexpression with AID as well as with A3G (Fig. 5E, lanes 1 and 3). Intriguingly, HBV DNA in core particles was also decreased by expression of an AID mutant AID(H56Y), which contains a mutation in the cytidine deaminase motif and is derived from a class switch deficiency patient (Fig. 5E, lane 2) (48). Southern blot also showed that the HBV rcDNA level in HepG2.2.15 cells was reduced by transduction with AID and another mutant AID(M139V), with diminished activity to support class switching (Fig. 5F) (48). These data suggest that AID could suppress HBV replication, and this restriction activity can be still observed with reduced enzymatic activity. In addition, AID was shown to interact with HBV core protein by coimmunoprecipitation assay (Fig. 5G). Moreover, overexpression of AID reduced the levels for nucleocapsid-associated HBV RNA (Fig. 5H). These results further suggest an antiviral activity of AID against HBV replication.

**AID Could Induce Hypermutation of HBV DNA**—Major enzymatic activity for APOBEC family proteins is the introduction of hypermutation in target DNA/RNA, and hypermutation accounts for antiviral activity for A3G against HIV-1 to some extent (2). Several groups reported that APOBEC family proteins could induce hypermutation in HBV DNA (27, 30, 32, 34). Next we asked whether AID could induce hypermutations in HBV DNA. In differential DNA denaturation PCR analysis, a high content of A/T bases introduced by hypermutation decreased denaturation temperatures (51). As shown in Fig. 6A, ectopic expression of AID decreased the denaturation temperature of HBV DNA as shown by that of A3G. Sequence analyses of the HBV DNA X region amplified at 83 °C by differential DNA denaturation PCR indicated a massive accumulation of

## Anti-HBV Activity of IL-1 and TNF $\alpha$ Mediated by AID



**FIGURE 4. AID expression was induced by IL-1 $\beta$  and TNF $\alpha$ .** *A*, mRNAs for A3B, -C, -D, -F, -G, -H and AID were quantified by real time RT-PCR analysis in HepaRG cells treated with 100 ng/ml IL-1 $\beta$ , 100 ng/ml TNF $\alpha$ , or 100 IU/ml IFN $\alpha$  for 12 h or left untreated. *Graphs* show the relative expression levels compared with the controls set at 1. *B*, AID mRNA was detected in HepG2, FLC4 cells, and PHH treated with IL-1 $\beta$ , TNF $\alpha$ , or IFN $\alpha$  or left untreated. Induction of AID by IL-1 $\beta$  and TNF $\alpha$  was observed in HepG2 and FLC4 cells and primary human hepatocytes. *C*, AID protein (upper panel) and actin levels as an internal control (lower panel) were examined by immunoblot of primary human hepatocytes treated with IL-1 $\beta$  or TNF $\alpha$  or left untreated. *D*, AID mRNA was detected in PHH treated with 100 ng/ml IL-1 $\beta$  in the presence or absence of NF- $\kappa$ B inhibitors, Bay11-7082, or QNZ for 12 h.

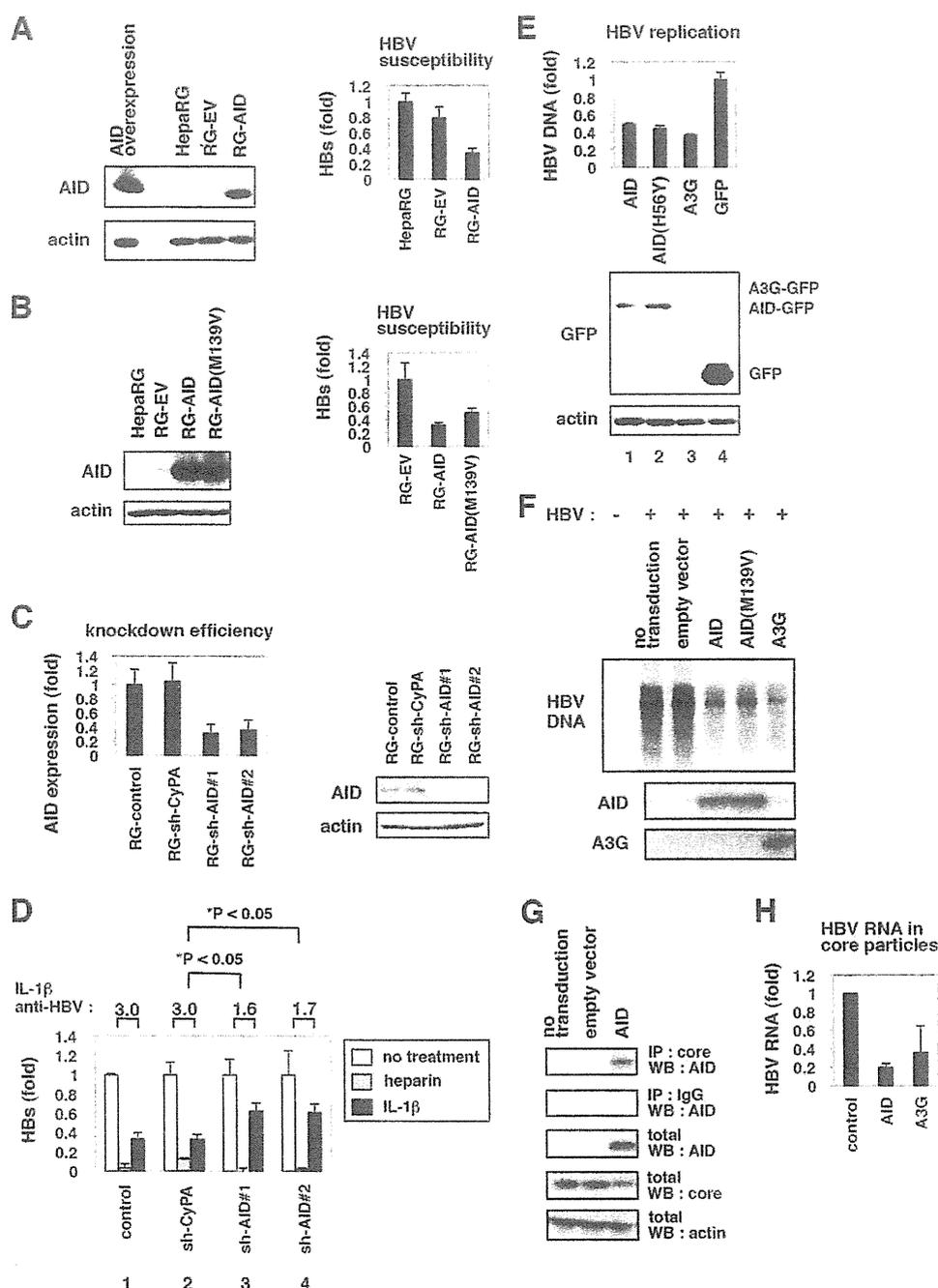
G-to-A mutations by AID (Fig. 6B). The frequency of G-to-A mutations was augmented by AID expression (Fig. 6C). In this experiment, AID(JP8Bdel), a hyper-active mutant of AID (62), further promoted the accumulation of the G-to-A and C-to-T mutations, although AID(H56Y) showed mutations in HBV DNA equivalent with mock GFP control sample (Fig. 6C). Thus, AID had the potential to introduce hypermutation in nucleocapsid-associated HBV DNA.

**IL-1 Suppressed the Infection of Different HBV Genotypes but Not That of HCV**—We examined whether the antiviral activity of IL-1 $\beta$  and TNF $\alpha$  could be generalized to other viruses or was specific to HBV. As shown in Fig. 7A, the production of infec-

tious HCV and HCV core proteins in the medium was not significantly altered by treatment with these cytokines in HCV-infected cells, compared to when IFN $\alpha$  was used as a positive control (Fig. 7A). In contrast, IL-1 suppressed the infection of HBV genotype A and C into HepaRG cells (Fig. 7B) as well as genotype D (Fig. 1C). These data suggest that the antiviral activity of proinflammatory cytokines IL-1 and TNF $\alpha$  is specific to HBV.

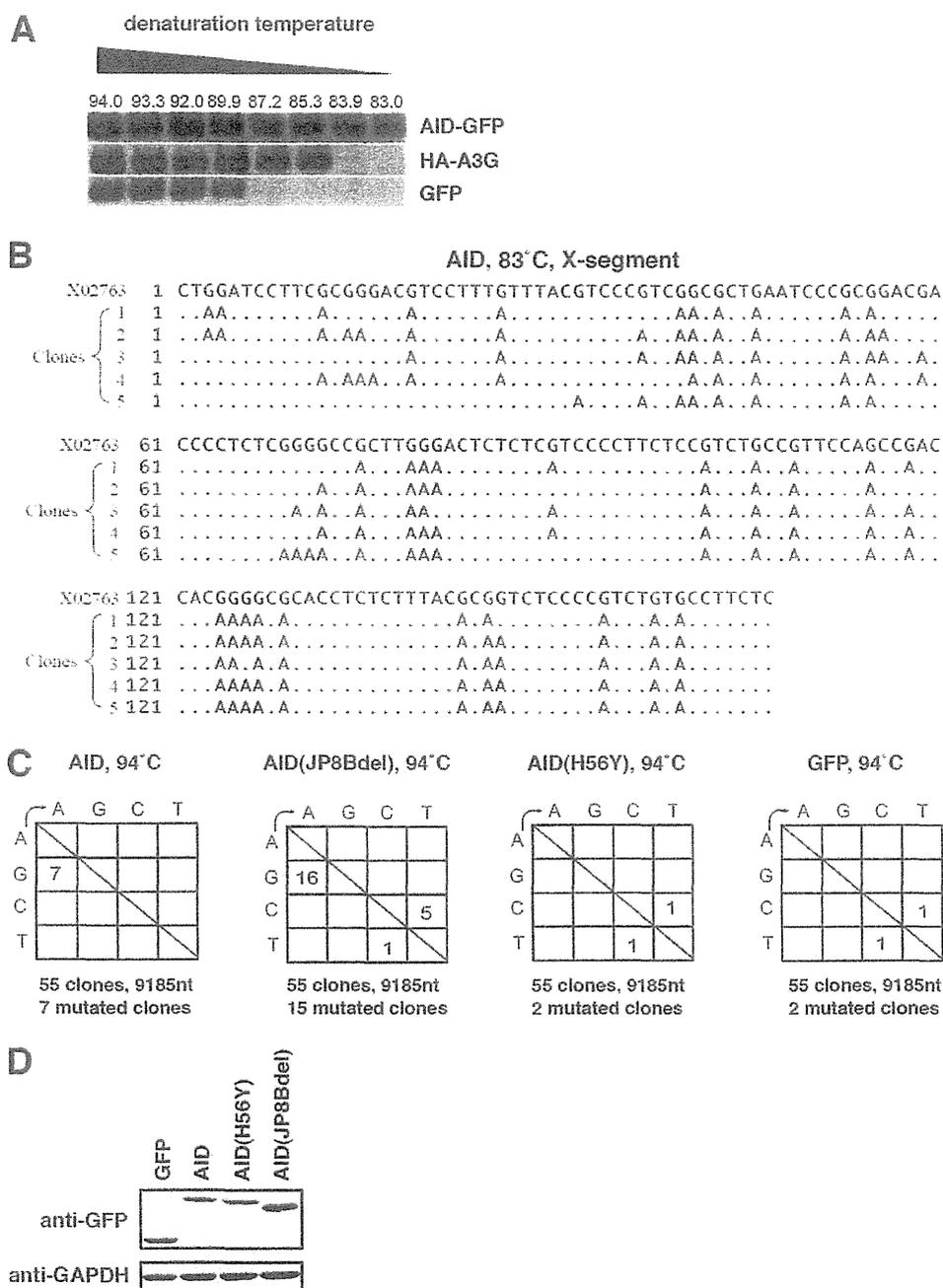
## DISCUSSION

In this study, cytokine screening revealed that IL-1 and TNF $\alpha$  decreased the host cell susceptibility to HBV infection.



**FIGURE 5. AID played a significant role in IL-1-mediated anti-HBV activity.** *A* and *B*, left panels, HepaRG cells were transduced with a lentiviral vector carrying the expression plasmid for AID (RG-AID), AID(M139V) mutant (RG-AID(M139V)) (*B*), or the control vector (RG-EV). Protein expression for AID (upper panel) and actin (lower panel) in these cells, the parental HepaRG cells (HepaRG), and those transiently transfected with AID expression plasmid (AID overexpression) (*A*) was examined by immunoblot. Right panels, these cells were infected with HBV followed by detection of secreted HBs protein as Fig. 1A. AID-transduced cells were less susceptible to HBV infection. *C*, HepaRG cells were transduced with lentiviral vector carrying shRNAs for AID (RG-shAID#1 and RG-shAID#2) or for cyclophilin A (RG-shCyPA) as a control. AID mRNA (left panel) and protein (right panel) were quantified by real time RT-PCR and immunoblot analysis. *D*, cells produced in *C* were infected with HBV in the absence or presence of IL-1 $\beta$  or heparin, and HBs was detected in the medium as in Fig. 1A to examine the anti-HBV effect of IL-1 $\beta$  and heparin. The fold reduction of HBV infection by IL-1 $\beta$  treatment is shown as IL-1 $\beta$  anti-HBV above the graph. The white, gray, and black bars indicate HBs value of the cells without treatment and with heparin and IL-1 $\beta$  treatment, respectively. The anti-HBV activity of IL-1 $\beta$  but not heparin was reduced in the AID-knockdown cells. *E*, AID and its mutant suppressed HBV replication. HepG2 cells were cotransfected with GFP-tagged AID, AID(H56Y), A3G, and GFP itself along with an HBV-encoding plasmid. Following 3 days, cytoplasmic nucleocapsid HBV DNA was quantified (upper graph), and the overexpressed proteins as well as actin were detected (lower panels). *F*, lentiviral vectors carrying AID, AID(M139V) mutant, A3G, or an empty vector (empty vector) were transduced or left untransduced (no transduction) into HepG2.2.15 cells. Nucleocapsid associated HBV DNA in these cells or in HepG2 cells (HBV-) was detected by Southern blot (upper panel). AID (middle panel) and A3G protein (lower panel) were also detected by immunoblot. *G*, HBV core interacted with AID. HepAD38 cells transduced without (no transduction) or with AID-expressing vector or the empty vector (empty vector) were lysed and treated with anti-core antibody (1st panel) or control normal IgG (2nd panel) for immunoprecipitation (IP). Total fraction without immunoprecipitation (3rd to 5th panels) was also recovered to detect AID (1st to 3rd panels), HBV core (5th panel), and actin (5th panel) by immunoblot. WB, Western blot. *H*, HBV RNA in core particles was extracted as shown under "Experimental Procedures" in HepG2 cells overexpressing HBV DNA together with or without AID or A3G.

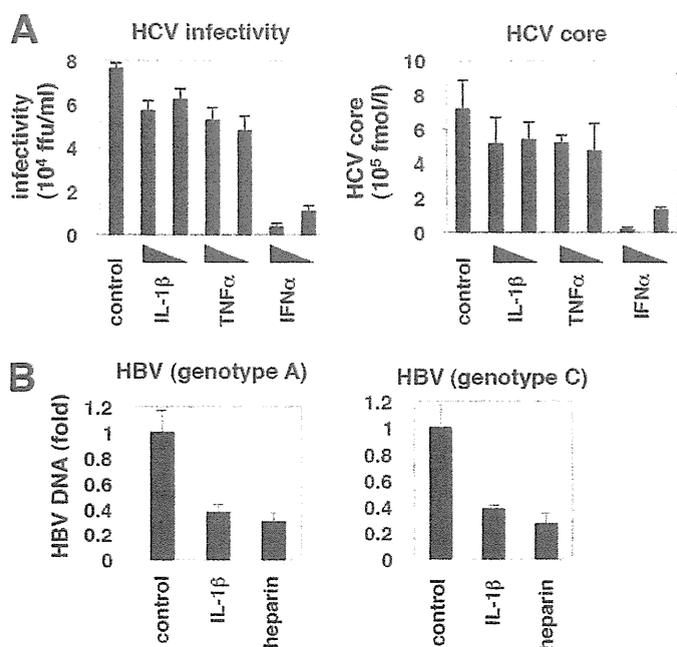
## Anti-HBV Activity of IL-1 and TNF $\alpha$ Mediated by AID



**FIGURE 6. AID could induce hypermutation of HBV DNA.** *A* and *B*, HepG2 cells were cotransfected with an expression vector for GFP-tagged AID, HA-tagged A3G, or GFP along with an HBV-encoding plasmid. 3 days after transfection, nucleocapsid-associated HBV DNA was extracted, and differential DNA denaturation PCR was performed to amplify the X gene segments. The numbers above the panels in *A* show denaturing temperatures. The X gene fragment amplified at 83°C in the AID sample was cloned in to a T vector and sequenced in *B*. Alignment of independent five clones with reference sequence (X02763) is indicated. *C*, AID and its mutant (JP8Bdel) induced G-to-A and C-to-T hypermutations in HBV DNA. HepG2 cells were transfected with expression vectors of GFP-tagged AID, AID(H56Y), AID(JP8Bdel), or GFP itself together with HBV encoding plasmid. Three days after transfection, cells were harvested, and nucleocapsid-associated HBV DNA was extracted. X gene fragments were amplified at 94°C and cloned in T vector. 55 clones were sequenced as described under "Experimental Procedures." The numbers indicate the clone numbers carrying the mutation. *D*, expression of GFP, GFP-tagged AID, AID(H56Y), and AID(JP8Bdel) is shown by immunoblot.

This antiviral mechanism is rather unique, given that the intracellular immune response against viruses is typically triggered by IFNs. So far, type I, II, and III IFNs are reported to suppress the replication step of the HBV life cycle (19, 20, 25, 26). In contrast, we suggest that IL-1 and TNF $\alpha$  inhibit the early phase of HBV infection as well as the replication. This is consistent with cumulative clinical evidence suggesting that these proinflammatory cytokines contribute to HBV elimination (63–65).

IL-1 and TNF $\alpha$  are generally produced mainly in macrophages and also in other cell types, including T cells and endothelial cells (66). Although the main producer cells of these cytokines in hepatitis B patients are not defined, it has been reported that the secretion of IL-1 and TNF $\alpha$  in nonparenchymal cells were increased by HBV infection into hepatocytes (67). TNF $\alpha$  production in macrophages was augmented by addition of recombinant HBc (68). A number of clinical studies cumulatively



**FIGURE 7. Antiviral activity of AID was specific to HBV.** A, Huh-7.5.1 cells were pretreated with IL-1 $\beta$ , TNF $\alpha$ , or IFN $\alpha$  for 3 h or left untreated and then coincubated with HCV for 4 h. After washing HCV and cytokines and culturing the cells with normal medium for 72 h, the infectivity of HCV (left panel) as well as HCV core protein (right panel) in the medium was quantified. B, HepaRG cells were treated with IL-1 $\beta$  or heparin or left untreated for 3 h prior to and 16 h during infection of HBV genotype A (left graph) or C (right graph) as shown in Fig. 1A. HBV infection was monitored with cellular HBV DNA at 12 days after the infection as Fig. 1C.

show that serum levels of IL-1 and TNF $\alpha$  are increased in hepatitis B patients (12). Recently, it has been a significant clinical problem that HBV reactivates during the course of treatment with immunosuppressants such as anti-TNF $\alpha$  agents (64, 65). Taken together, it is proposed that acute or chronic HBV infection induces IL-1/TNF $\alpha$  from macrophages or other cells in the liver of infected patients, which can directly suppress HBV infection in hepatocytes, in addition to their immunomodulatory effects to the host immune cells. Although IL-1 level in HBV-infected patients varies between papers, Daniels *et al.* (63) reported that the peak IL-1 $\beta$  level in HBV-infected patients was 9–36 ng/ml under Toll-like receptor stimulation, at which concentration IL-1 $\beta$  showed significant anti-HBV effects in this study. In general, downstream genes of NF- $\kappa$ B include a number of antiviral factors such as *viperin*, *iNOS*, and *RANTES* (69). Although some of these genes may function cooperatively for IL-1- and TNF $\alpha$ -induced anti-HBV machinery, our data suggest that AID, at least in part, plays a role in the elimination of HBV that was potentiated by proinflammatory cytokines IL-1 and TNF $\alpha$ .

AID belongs to APOBEC family proteins that share enzyme activity to convert cytidine to uracil in mainly DNA, and occasionally RNA (51, 70, 71). Although AID was initially identified in B cells, chronic inflammation can trigger its expression in hepatocytes (60). The induction of AID was reportedly mediated by NF- $\kappa$ B (60), consistent with the results in this study. Although AID in B cells is essential for class switch recombination and somatic hypermutation of immunoglobulin genes (70, 72), the physiological role of AID in hepatocytes is unknown.

Although expression of AID in hepatocytes is still lower than in B cells, AID is reportedly expressed in the liver both in cell culture and *in vivo* settings (34, 60). Our results raise the idea that AID plays a role in innate antiviral immunity. AID also has a role in virus-induced pathogenesis as it was reported to counteract oncogenesis induced by Abelson-murine leukemia virus (73). In addition, AID was reported to restrict L1 retrotransposition, which can predict the role of AID in innate immunity (74). This study is significant in that it revealed a biological function of AID in viral infection itself, linking it to the restriction of a pathogenic human virus. It will be interesting to analyze the role of AID in the infection process of other viruses in the future.

Although the mechanism for AID suppression of the HBV life cycle is the subject of future study, AID possibly targets the early phase of HBV infection, including entry as well as the replication stage, including assembly and reverse transcription (Fig. 3). It has been recently reported that chicken AID reduced cccDNA of duck HBV possibly through targeting cccDNA as well as nucleocapsid-associated HBV DNA (75). This study is likely to support the idea that AID may target cccDNA formed after HBV entry into hepatocytes, and also associates with nucleocapsid-associated HBV DNA during HBV replication, although it is not clear whether the innate immune machinery against HBV/duck HBV is conserved in human and chicken cells. A3G blocked HBV replication through the inhibition of reverse transcriptase (29), packaging of pregenomic RNA (33), and the destabilization of packaged pregenomic RNA (31) independently of its deaminase activity, and it also induced hypermutation of HBV DNA (27, 30, 32, 34). It was recently reported that AID was packaged into the HBV nucleocapsid (51). Moreover, AID induced C-to-T and G-to-A hypermutations in HBV DNA/RNA, although the anti-HBV activity has not been demonstrated so far (51). The hypermutation activity of AID was likely to be dispensable for its anti-HBV replication function (Figs. 5 and 6), as reported for APOBEC3G by several groups (29, 30, 33). Further analysis is required to elucidate the precise mechanisms for AID-mediated suppression of the HBV life cycle.

In conclusion, we have identified that host cell susceptibility to HBV infection is modulated by IL-1 and TNF $\alpha$ , and AID is involved in this machinery. This sheds new light on the link between proinflammatory cytokines and the development of the innate antiviral defense.

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# Antiviral Activity of Glycyrrhizin against Hepatitis C Virus *In Vitro*

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## Abstract

Glycyrrhizin (GL) has been used in Japan to treat patients with chronic viral hepatitis, as an anti-inflammatory drug to reduce serum alanine aminotransferase levels. GL is also known to exhibit various biological activities, including anti-viral effects, but the anti-hepatitis C virus (HCV) effect of GL remains to be clarified. In this study, we demonstrated that GL treatment of HCV-infected Huh7 cells caused a reduction of infectious HCV production using cell culture-produced HCV (HCVcc). To determine the target step in the HCV lifecycle of GL, we used HCV pseudoparticles (HCVpp), replicon, and HCVcc systems. Significant suppressions of viral entry and replication steps were not observed. Interestingly, extracellular infectivity was decreased, and intracellular infectivity was increased. By immunofluorescence and electron microscopic analysis of GL treated cells, HCV core antigens and electron-dense particles had accumulated on endoplasmic reticulum attached to lipid droplet (LD), respectively, which is thought to act as platforms for HCV assembly. Furthermore, the amount of HCV core antigen in LD fraction increased. Taken together, these results suggest that GL inhibits release of infectious HCV particles. GL is known to have an inhibitory effect on phospholipase A2 (PLA2). We found that group 1B PLA2 (PLA2G1B) inhibitor also decreased HCV release, suggesting that suppression of virus release by GL treatment may be due to its inhibitory effect on PLA2G1B. Finally, we demonstrated that combination treatment with GL augmented IFN-induced reduction of virus in the HCVcc system. GL is identified as a novel anti-HCV agent that targets infectious virus particle release.

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## Introduction

Hepatitis C virus (HCV) infection is a major public health problem since most cases cause chronic hepatitis, hepatic cirrhosis and hepatocellular carcinoma. Current treatment of chronic hepatitis C is based on the combination of pegylated interferon-alpha (IFN- $\alpha$ ) and ribavirin. However, approximately 50% of treated patients infected with genotype 1 do not respond, or show only a partial or transient response, and therapy causes significant side effects [1]. In Japan, glycyrrhizin (GL) preparations (stronger neo-minophagen C

(SNMC)) have been used for more than 20 years as a treatment for chronic hepatitis patients who do not respond to IFN therapy.

GL is the major component of licorice root extract, and is composed of glycyrrhetic acid. GL has been shown to possess several beneficial pharmacological activities, including anti-inflammatory activity [2], anti-tumor activity [3], anti-allergic activities [4], and anti-viral activities [5]. Several mechanisms of the GL-induced anti-inflammatory effect are reported, such as inhibition of thrombin-induced platelet aggregation [6], inhibition

of prostaglandin E2 production [7] and inhibition of phospholipase A2 (PLA2) [8].

Many anti-viral effects of GL have been reported previously, for example, against herpes simplex type 1 (HSV-1) [9], varicella-zoster virus (VZV) [10], hepatitis A (HAV) [11] and B virus (HBV) [12], human immunodeficiency virus (HIV) [13], severe acute respiratory syndrome (SARS) and coronavirus [14], Epstein-Barr virus (EBV) [15], human cytomegalovirus [16] and influenza virus [17]. GL has been considered as a potential treatment for patients with chronic hepatitis C, and long term administration of GL to patients is effective in suppressing serum alanine aminotransferase (ALT) levels and histological change [18]. However, a direct anti-viral effect of GL against HCV has never been reported.

In this study, we evaluated the anti-HCV effects of GL, and demonstrated that GL targeted the release step of infectious HCV particles from infected cells. We found that the suppression of virus release by GL may be derived from its inhibitory effect on group 1B PLA2 (PLA2G1B). These findings suggest possible novel roles for GL in the treatment of patients with chronic hepatitis C.

## Materials and Methods

### Cell culture and reagents

The human hepatoma cell line, Huh7, and its derivative cell line, Huh7.5.1, provided by Francis Chisari (Scripps Research Institute, La Jolla, CA), were maintained in Dulbecco's modified Eagle's medium (DMEM) containing 10% fetal bovine serum (FBS) [19]. Huh7 cells harboring the subgenomic replicon [20] [21] were maintained in complete DMEM supplemented with 0.5 mg/ml G418 (Geneticin, Life Technologies Japan Ltd., Tokyo, Japan). GL (20 $\beta$ -carboxyl-11-oxo-30-norolean-12-en-3 $\beta$ -yl-2-O- $\beta$ -D-glucopyranuronosyl- $\beta$ -D-glucopyranosiduronic acid) and IFN- $\alpha$  were kindly provided by the Minophagen Pharmaceutical Co., Ltd., (Tokyo, Japan) and MSD K.K., (Tokyo, Japan) respectively. Oleyloxyethyl phosphorylcholine (OPC) (Cayman Chemical Company, Ann Arbor, MI), sPLA2IIA Inhibitor I (MERCK, Darmstadt, Germany), anti-Actin (Santa Cruz Biotechnology, Santa Cruz, CA) and anti-Human CD81 (BD Pharmingen, San Jose, CA) antibodies were purchased. The solvents were distilled water (GL), ethanol (OPC), and DMSO (sPLA2IIA inhibitor).

### Quantification of HCV core antigen and cell viability

The production of cell culture-produced HCV (HCVcc) has been previously reported [22]. Purification of LD has been previously reported [23]. The concentration of HCV core antigen in filtered culture medium, in cell lysates and in LD fraction of infected cells was determined using the Lumipulse Ortho HCV antigen kit (Ortho Clinical Diagnostics, Tokyo, Japan). Cell viability was analyzed by using Cell Titer-Glo Luminescent Cell Viability Assay (Promega, Madison, WI) according to the manufacturers' protocol.

Electroporation of HCV RNA lacking E1 and E2

In vitro synthesis of HCV RNA JFH1 lacking E1 and E2 (JFH1delE1E2), and electroporation were performed as described previously [22].

### HCV pseudoparticle (HCVpp) assay

HCVpp harboring E1 and E2 glycoproteins of the JFH-1 clone (genotype 2a) (HCVpp2a) and the TH clone (genotype 1b) (HCVpp1b) were produced as previously described [24]. Pseudotype virus with VSV G glycoprotein (VSVpp) were also generated [24]. Huh7 or Huh7.5.1 cells were seeded into 48-well plates, incubated overnight at 37°C, and then infected with the HCVpp in the presence of various concentration of GL. Several hours post-infection, medium was replaced with DMEM with 10% FBS, and the cells were harvested 48 hours later to determine intracellular luciferase activity (Luciferase Assay System, Promega).

### HCV subgenomic replicon assay

The assay for the genotype 1b and 2a subgenomic reporter replicon has been previously reported [20] [21]. After 72 hours of treatment with GL, the replicon-transfected cells were harvested for either measurement of luciferase activity (Promega) or HCV RNA titer, as described previously [25]. The replication efficiency of HCV in each preparation was calculated as the percentage of luciferase activity or HCV RNA titer compared with that of cells subjected to the control treatment.

### Extra- and intracellular infectivity

To determine extracellular HCV infectivity, naïve Huh7 cells were inoculated with cell culture supernatant medium containing HCVcc. After 3 hours of incubation, the medium was replaced with DMEM containing 10% FBS, and the cells were cultured for an additional 72 hours. The infectious HCV titer in the culture medium was determined by quantification using the Lumipulse Ortho HCV antigen kit or by immunostaining of the HCV core antigen. Using an immunoassay that also provided results indicative of HCV infectivity [26], we confirmed a good correlation between the levels of core antigen and infectious titers (data not shown). To estimate intracellular infectivity, cells in the culture plates filled with DMEM containing 10% FBS were subjected to four cycles of freezing and thawing, using dry ice and a 37°C water bath. Cells in the culture plates were centrifuged at 1,200 rpm for 5 min at 4°C to remove cell debris, and the supernatants were collected to evaluate infectivity as above.

### RNA interference

The siRNA targeted to PLA2G1B, 5'-GCUGGACAGCUGUAAAUUUTT-3', and scramble negative control siRNA to PLA2G1B were purchased from Sigma (Tokyo, Japan). Cells in a 24-well plate were transfected with siRNA using HiPerFect transfection reagent (Qiagen, Tokyo, Japan) following the manufacturer's instructions.

Quantification of triglyceride

Triglyceride (TG) was measured with a Triglyceride kit (Wako, Tokyo, Japan) according to the manufacturer's instructions.