

球細胞の門脈内投与療法、培養自己骨髄由来細胞を用いた細胞療法などの臨床研究の論文報告がある。本稿では、われわれのABMi療法を含む肝硬変症に対する肝臓再生療法の状況や、今後の展望について概説する。

A. 基礎研究；マウス GFP/CCl₄モデル

われわれは、四塩化炭素 (CCl₄) による肝細胞直接障害モデルを用いて基礎研究を進めてきた。われわれのマウス GFP/CCl₄モデルの特徴は、1) 四塩化炭素の反復投与により慢性肝障害環境下にあること、2) 骨髄細胞投与後も四塩化炭素投与を継続しこの炎症環境を維持すること、3) 自家骨髄細胞投与を想定して同種同系 GFPトランスジェニックマウスをドナーとしたことである^{9,10)}。また本モデルでは、6週齢の C57BL/6 マウスに四塩化炭素 (1.0mL/kg) を4週間 (計8回) 腹腔内反復投与することで慢性肝障害 (肝硬変) 状態とし、これに同種同系 GFPトランスジェニックマウス大腿骨から分離した全骨髄細胞を洗浄後に末梢静脈である尾静脈から投与し、その後も四塩化炭素投与は継続したうえで、経時的に肝機能改善効果を評価している。結果、骨髄細胞を投与することにより、血清アルブミン値の改善、生存率の有意な上昇さらにはシリウスレッド染色で評価した肝線維の減少が認められた⁹⁻¹¹⁾。投与した骨髄細胞は肝障害がないマウス肝臓には生着しないものの、四塩化炭素による持続肝障害環境下で投与した骨髄細胞は投与後1日目から門脈域周囲の線維に沿って生着し、さらに週を追うごとに既存の線維の中にも観察された¹¹⁾。またこの過程で、障害肝に生着した骨髄由来 GFP陽性細胞が matrix metalloproteinase (MMP) 9などのコラゲナーゼを産生し、肝線維の溶解に働くことを確認した¹¹⁾。これらの基礎研究結果から、慢性肝障害環境下に自己骨髄細胞を末梢静脈から投与す

ることにより、レシピエントの肝合成能・肝線維化さらには生命予後までも改善させたと考えた。さらにその後の検討により、本過程に関与する因子として FGF2 (fibroblast growth factor 2) が促進的に働くことも明らかとなった¹²⁾。さらに高発癌肝硬変マウスに対して骨髄細胞を頻回投与したモデル系において肝発癌は有意に抑制されており、骨髄細胞投与により酸化ストレスが制御されていた可能性が示唆された¹³⁾。

B. 臨床研究

1. 自己骨髄細胞を用いた ABMi 療法

われわれの自己骨髄細胞 (非培養) を用いた ABMi 療法の適応条件などの詳細は以下である。

【対象疾患】肝硬変症

【治療適応基準】1) 総ビリルビン値 3.0mg/dL 以下、2) 血小板数 $5.0 \times 10^{10}/L$ 以上、3) 食道胃静脈瘤および肝細胞癌コントロール良好、4) 心肺機能良好で重篤な併存疾患を認めない、5) CT や MRI などの画像診断で肝細胞癌が存在しない。

【プロトコール】全身麻酔下に約 400mL の自己骨髄細胞を採取し、GMP グレード設備が完備された再生・細胞療法センターで SOP (Standard Operating Procedures) に準じて骨髄単核球細胞を精製濃縮し、平均 5.2×10^9 個の自己骨髄単核球細胞を採取同日に本人の末梢静脈から点滴投与した。細胞投与後は6カ月間経過観察を行い、血液生化学検査、肝組織検査 (肝生検)、腹部超音波検査や腹部 CT 検査により安全性および有効性の評価を行った。また経過観察中は、内服薬剤や抗ウイルス剤などの変更は行っていない¹⁴⁾。

【結果】骨髄細胞投与後6カ月間経過観察可能であった症例において、投与6カ月後の血清アルブミン値、総蛋白値や Child-Pugh スコアは有意に改善し¹⁴⁾、さらに15カ月間経過観察可能であった9例でも同様の改善効果を認めた。なお2013

年7月現在において、特に問題となる有害事象の発生は認めていない¹⁴⁻¹⁶⁾。さらに2011年には、非代償性B型肝炎を対象としたABMi療法が血清アルブミン値やChild-Pughスコアを改善させたとの報告がKimらによりなされ、そのメカニズムとして経時的な肝生検からhepatic progenitor cell(HPC)を活性化させる可能性が示唆された¹⁷⁾。さらにアルコール性肝硬変に対するABMi療法の有効性と安全性が齊藤らにより報告された¹⁸⁾。

2. 先進医療B「C型肝炎ウイルスに起因する肝硬変患者に対するABMi療法(ランダム化比較試験)」

これまでのABMi療法の実績により、「C型肝炎ウイルスに起因する肝硬変患者に対するABMi療法の有効性と安全性に関する研究(ランダム化比較試験)」が2011年12月に「ヒト幹細胞を用いる臨床研究に関する指針」の承認を、2013年6月には先進医療Bの認可を受け、今後、実施していく。なお、本ランダム化比較試験の概要は以下のとおりである。

【目的】C型肝炎ウイルスに起因する肝硬変患者に対する自己骨髄細胞投与療法の有効性と安全性の検討

【適格基準】以下の選択基準をすべて満たし、かつ、以下の除外基準のいずれにも該当しない症例を適格症例とする。

【選択基準】

- (1) C型肝炎ウイルスに起因する肝硬変症例
- (2) 90日以上離れた2点において、Child-Pughスコアが7点(Child-Pugh B)以上の状態にあり、現行の内科的な治療法では改善が見込めない症例
- (3) 20歳以上75歳以下の症例
- (4) インフォームドコンセントを取得可能で、研究参加の同意が得られた症例

【除外基準】

- (1) C型肝炎ウイルス以外の原因で肝硬変へ至った症例、もしくは肝硬変へ至った原因が不明の症例
- (2) 悪性新生物を合併する、または既往を有する症例
- (3) 破裂の危険性を有する食道・胃静脈瘤を有する症例
- (4) 血清クレアチニン値 2mg/dL以上の腎機能障害を合併する症例
- (5) ヘモグロビン値が 8g/dL未満、あるいは血小板数が 50,000/ μ L未満の症例
- (6) 血清総ビリルビン値が3.0mg/dL以上の症例
- (7) Performance Status 3あるいは4の症例
- (8) 同種血輸血に関する同意を得られない症例
- (9) B型肝炎ウイルス感染症、ヒト免疫不全ウイルス感染症、成人T細胞白血病ウイルス感染症、パルボウイルスB19感染症が否定できない症例
- (10) 妊娠中の女性
- (11) 全身麻酔を行うことが適切でないと担当医が総合的に判断した症例
- (12) 造影剤に対する重篤なアレルギーのある症例もしくは造影剤に対する重篤なアレルギーの既往を有する症例
- (13) その他、担当医が不相当と判断した症例

【主要評価項目】細胞投与群は細胞投与後、標準的治療群は登録後24週の時点でChild-Pughスコアの1点以上改善する割合

【副次評価項目】細胞投与群は細胞投与後、標準的治療群は登録後24週の時点の以下の指標を副次エンドポイントとする。

- (1) 効果維持率の推移(効果維持の定義: Child-Pughスコアが悪化しない)
- (2) 血清アルブミン値の推移
- (3) 血清線維化マーカー値の推移

- (4) 腹水量の推移
- (5) 下腿浮腫の改善率及び消失率の推移
- (6) 自覚症状の推移
- (7) 有害事象の発生頻度

【有害事象・重大な事態の評価】有害事象とは、臨床研究参加期間中（同意取得時～プロトコル治療後観察期終了もしくは中止時）に被験者に生じたあらゆる好ましくない、意図しない徴候（臨床検査値の異常変動を含む）、症状又は病気をいい、当該プロトコル治療との因果関係の有無は問わない。

3. ABMi療法以外の臨床研究

まずG-CSF (granulocyte-colony stimulating factor) を使った肝硬変に対する臨床研究論文としては、GordonらのG-CSFで誘導した自己末梢血CD34陽性細胞を門脈または肝動脈から投与したところ血清アルブミン値が上昇したとの報告¹⁹⁾、アルコール性肝硬変症に対するG-CSF投与が肝前駆細胞の増殖を促進させたとのSpahrらの報告²⁰⁾、Child-Pughスコアを改善させたとのPaiらの報告や²¹⁾、HanらによるB型肝硬変症に対するG-CSF投与の報告がある²²⁾。しかしながら、G-CSF投与により健常人でも脾破裂を起こしたとの報告があることから、脾腫を伴う肝硬変症例へのG-CSF投与には注意が必要である²³⁾。最近では、Spahrらが非代償性アルコール性肝硬変症を対象にG-CSF投与しその後に採取した自己骨髓単核球細胞を投与したものの、標準治療群であるステロイド投与群と同等の改善であったと報告している²⁴⁾。またG-CSFを使用せず200mLの自己骨髓液から分離した濃縮CD34陽性細胞を肝動脈から投与するPhase I臨床研究の報告があるが、本臨床研究では造影剤投与が原因と考えられる腎不全による死亡例の報告がある²⁵⁾。このことは、投与細胞の種類、細胞濃度や投与速度を最適化することの重要を示している。

その他にはドイツから、肝悪性腫瘍切除術後の残肝に自己CD133陽性細胞を経門脈投与する細胞療法の有効性も報告されている^{26,27)}。

一方、ABMi療法と同様に自己骨髓細胞を用いる肝臓再生療法には以下のような論文報告がある。PengらはB型肝硬変症を対象に120mLの自己骨髓単核球細胞（間葉系幹細胞を含む）を肝動脈から投与する群（n=53）と、投与しないコントロール群（n=105）で解析した。その結果、骨髓投与により副作用はなく、早期には肝機能の改善を確認した。また長期観察では、骨髓細胞投与後の肝細胞癌の発生率は低い傾向で（p=0.107）、われわれの高発癌肝硬変マウスと矛盾しない結果であった²⁸⁾。その他、ブラジルのLyraらによる、肝移植待機例10例に対して腸骨から採取した自己骨髓単核球細胞を肝動脈から投与したところ血清アルブミン値の上昇と血清ビリルビン値の低下を認め、投与経路は末梢静脈より肝動脈投与が有効であったとの報告がある^{29,30)}。

さらには、培養骨髓由来細胞を用いた臨床研究の論文報告もある。少量の骨髓液から肝再生・修復作用を有する細胞を分離培養し再投与することができれば、全身麻酔下に骨髓液を採取する必要がなくなるため適応は拡大し、さらには凍結保存した培養細胞を分割投与することも不可能ではなくなり、患者負担は大きく軽減される。まず2007年にMohamadnejadらはPhase I臨床研究として4例の非代償性肝硬変症に対して、平均 3.2×10^7 個の培養自己骨髓間葉系幹細胞を末梢静脈から投与し、うち2例でMELD (the model for end-stage liver disease) スコアが改善したと報告している³¹⁾。さらにKharazihaらは8例の肝硬変症に対して、局所麻酔下に腸骨から採取した約20mLの骨髓液から単核球をフィコール法で分離し約2週間通常培養した。この培養細胞はCD44/CD73/CD105（間葉系細胞マーカー）陽性で、 $3 \sim 5 \times 10^7$ 個を末梢静脈または門脈から

表1 論文報告されたおもな肝臓再生療法

投与細胞の種類	投与細胞数	改善効果	対象症例	文献番号
CD34陽性細胞 (G-CSF誘導末梢血由来)	$1 \times 10^6 \sim 2 \times 10^8$	血清アルブミン・ ビリルビン改善	5 Alcohol	19
5日間G-CSF投与のみ	記載なし	血清HGF上昇 肝Ki67 ⁺ /CK7 ⁺ 細胞増加	24 Alcohol (うち11コントロール)	20
培養CD34陽性細胞 (G-CSF誘導末梢血由来)	平均 2.3×10^8	血清アルブミン・ Child-Pughスコア改善	9 Alcohol	21
末梢血単核球細胞 (G-CSF誘導末梢血由来)	$10^7 \sim 10^8/\text{kg}$	血清アルブミン・ Child-Pughスコア改善	40 HBV (うち20コントロール)	22
G-CSF+単核球細胞	$4.7 \pm 10^7/\text{kg}$	MELDスコア, 肝脂肪化 (標準治療群でも改善)	48 Alcohol (うち30コントロール; ステロイド投与)	24
骨髓単核球細胞; ABM/療法	$5.20 \pm 0.63 \times 10^9$	血清アルブミン・ Child-Pughスコア改善	5 HCV, 3 HBV, 1 成因不明	14
骨髓単核球細胞; ABM/療法	$0.48 \sim 1.48 \times 10^8/\text{kg}$	肝CK7陽性細胞増加 肝容量増加 Child-Pughスコア改善	10 HBV	17
骨髓単核球細胞; ABM/療法	$8.0 \pm 7.3 \times 10^9$	血清アルブミン・PT% Child-Pughスコア改善	10 Alcohol (うち5コントロール)	18
骨髓単核球細胞(MSC)	記載なし	血清アルブミン・ ビリルビン・PT% MELDスコア改善 HCC増加なし	158 HBV (うち105コントロール)	28
骨髓単核球細胞	$> 1 \times 10^8$	血清アルブミン・ ビリルビン改善	10	29
骨髓単核球細胞	$3.78 \pm 2.69 \times 10^8$	血清アルブミン・ ビリルビン・ Child-Pughスコア改善	30 (うち15コントロール)	30
骨髓由来CD133陽性細胞	$2.4 \sim 12.3 \times 10^6$	肝切除後の肝容量増加	6 肝癌 (うち3コントロール)	26
骨髓由来CD34陽性細胞	平均 5.25×10^6 (CD34 ⁺ , 90.5%)	造影剤による腎不全死亡 (1例)	1 HBV, 1 PBC 1 AIH, 1 成因不明	27
培養骨髓由来MSC	平均 31.7×10^6	MELDスコア改善 (ただし2例のみ)	3 成因不明, 1 AIH	31
培養骨髓由来MSC	$3 \sim 5 \times 10^7$	MELDスコア・ 血清クレアチニン改善	4 HBV, 2 成因不明 1 HCV	32
HGF含有培地培養MSC	2×10^8	MELDスコア改善	40 HCV (うち20コントロール)	33

G-CSF, granulocyte colony-stimulating factor; HGF; hepatocytes growth factor; PT, prothrombin time; MSC, mesenchymal stem cell; MELD, Model for End Stage Liver Disease [Takami T, et al. Curr Opin Gastroenterol. 2012; 28(3): 203-8⁷⁾ より改変]

投与したところMELDスコアが平均17.9から10.7へと改善したと報告している³²⁾。Amerらは20例のC型肝硬変症から局所麻酔下に腸骨から約120mLの骨髓液を採取し、HGF (hepatocyte growth factor) 含有培地で培養した間葉系幹細胞をエコーガイド下に肝臓または脾臓内へ直接注入し、コントロール20例に比べて有意にMELDスコアおよびChildスコアが投与後2週から6カ月の間は改善したと報告している³³⁾。今後は、エビデンスレベルの高い有効性を示すためにランダム化比較試験の実施などが求められよう。

なお、以上のおもな肝臓再生療法の概要を表1にまとめているので参照いただきたい⁷⁾。

むすび

これまでの基礎研究や臨床研究成績から、肝硬変症に対する(自己)骨髓細胞を用いた肝臓再生療法の有効性は強く示唆されている。今後は、エビデンスレベルの高い有効性を示すためにランダム化比較試験を推進することや、低侵襲な骨髓由来培養細胞を用いた治療法の開発が必要となっていくであろう。

文献

- 1) Petersen BE, Bowen WC, Patrene KD, et al. Bone marrow as a potential source of hepatic oval cells. *Science*. 1999; 284(5417): 1168-70.
- 2) Theise ND, Nimmakayalu M, Gardner R, et al. Liver from bone marrow in humans. *Hepatology*. 2000; 32(1): 11-6.
- 3) Houlihan DD, Newsome PN. Critical review of clinical trials of bone marrow stem cells in liver disease. *Gastroenterology*. 2008; 135(2): 438-50.
- 4) Gilchrist ES, Plevris JN. Bone marrow-derived stem cells in liver repair: 10 years down the line. *Liver Transpl*. 2010; 16(2): 118-29.
- 5) Stutchfield BM, Forbes SJ, Wigmore SJ. Prospects for stem cell transplantation in the treatment of hepatic disease. *Liver Transpl*. 2010; 16(7): 827-36.
- 6) Souza BS, Nogueira RC, de Oliveira SA, et al. Current status of stem cell therapy for liver diseases. *Cell Transplant*. 2009; 18(12): 1261-79.
- 7) Takami T, Terai S, Sakaida I. Stem cell therapy in chronic liver disease. *Curr Opin Gastroenterol*. 2012; 28(3): 203-8.
- 8) Pai M, Spalding D, Xi F, et al. Autologous bone marrow stem cells in the treatment of chronic liver disease. *Int J Hepatol*. 2012; 2012: 307165. doi: 10.1155/2012/307165. Epub 2011 Nov 3.
- 9) Terai S, Sakaida I, Yamamoto N, et al. An in vivo model for monitoring the transdifferentiation of bone marrow cells into functional hepatocytes. *J Biochem*. 2003; 134(4): 551-8.
- 10) Terai S, Sakaida I, Nishina H, et al. Lesson from the GFP/CC14 model-translational research project: the development of cell therapy using autologous bone marrow cells in patients with liver cirrhosis. *J Hepatobiliary Pancreat Surg*. 2005; 12(3): 203-7.
- 11) Sakaida I, Terai S, Yamamoto N, et al. Transplantation of bone marrow cells reduces CC14-induced liver fibrosis in mice. *Hepatology*. 2004; 40(6): 1304-11.
- 12) Ishikawa T, Terai S, Urata Y, et al. Fibroblast growth factor 2 facilitates the differentiation of transplanted bone marrow cells into hepatocytes. *Cell Tissue Res*. 2006; 323(2): 221-31.
- 13) Maeda M, Takami T, Terai S, et al. Autologous bone marrow cell infusions suppress tumor initiation in hepatocarcinogenic mice with liver cirrhosis. *J Gastroenterol Hepatol*. 2012; 27(Suppl 2): 104-11.
- 14) Terai S, Ishikawa T, Omori K, et al. Improved liver function in patients with liver cirrhosis after autologous bone marrow cell infusion therapy. *Stem Cells*. 2006; 24(10): 2292-8.
- 15) Terai S, Sakaida I. Current status of autologous bone marrow cell infusion therapy for liver cirrhosis patients. *Hepatol Res*. 2008; 38(The 6 Japan Society of Hepatology Single Topic Conference: Liver Failure: Recent Progress and Pathogenesis to Management. 28-29 September 2007, Iwate, Japan): S72-5.
- 16) Terai S, Sakaida I. Autologous bone marrow cell infusion therapy for liver cirrhosis patients. *J Hepatobiliary Pancreat Sci*. 2011; 18(1): 23-25.

- 17) Kim JK, Park YN, Kim JS, et al. Autologous bone marrow infusion activates the progenitor cell compartment in patients with advanced liver cirrhosis. *Cell Transplant*. 2010; 19(10): 1237-46.
- 18) Saito T, Okumoto K, Haga H, et al. Potential therapeutic application of intravenous autologous bone marrow infusion in patients with alcoholic liver cirrhosis. *Stem Cells Dev*. 2011; 20(9): 1503-10.
- 19) Gordon MY, Levicar N, Pai M, et al. Characterization and clinical application of human CD34+ stem/progenitor cell populations mobilized into the blood by granulocyte colony-stimulating factor. *Stem Cells*. 2006; 24(7): 1822-30.
- 20) Spahr L, Lambert JF, Rubbia-Brandt L, et al. Granulocyte-colony stimulating factor induces proliferation of hepatic progenitors in alcoholic steatohepatitis: a randomized trial. *Hepatology*. 2008; 48(1): 221-9.
- 21) Pai M, Zacharoulis D, Milicevic MN, et al. Autologous infusion of expanded mobilized adult bone marrow-derived CD34+ cells into patients with alcoholic liver cirrhosis. *Am J Gastroenterol*. 2008; 103(8): 1952-8.
- 22) Han Y, Yan L, Han G, et al. Controlled trials in hepatitis B virus-related decompensate liver cirrhosis: peripheral blood monocyte transplant versus granulocyte-colony-stimulating factor mobilization therapy. *Cytotherapy*. 2008; 10(4): 390-6.
- 23) Falzetti F, Aversa F, Minelli O, et al. Spontaneous rupture of spleen during peripheral blood stem-cell mobilisation in a healthy donor. *Lancet*. 1999; 353(9152): 555.
- 24) Spahr L, Chalandon Y, Terraz S, et al. Autologous bone marrow mononuclear cell transplantation in patients with decompensated alcoholic liver disease: a randomized controlled trial. 2013; 8(1): e53719. doi: 10.1371/journal.pone.0053719. Epub 2013 Jan 14.
- 25) Mohamadnejad M, Namiri M, Bagheri M, et al. Phase I human trial of autologous bone marrow-hematopoietic stem cell transplantation in patients with decompensated cirrhosis. *World J Gastroenterol*. 2007; 13(24): 3359-63.
- 26) am Esch JS 2nd, Knoefel WT, Klein M, et al. Portal application of autologous CD133+ bone marrow cells to the liver: a novel concept to support hepatic regeneration. *Stem Cells*. 2005; 23(4): 463-70.
- 27) Furst G, Schulte am Esch J, Poll LW, et al. Portal vein embolization and autologous CD133+ bone marrow stem cells for liver regeneration: initial experience. *Radiology*. 2007; 243(1): 171-9.
- 28) Peng L, Xie DY, Lin BL, et al. Autologous bone marrow mesenchymal stem cell transplantation in liver failure patients caused by hepatitis B: Short-term and long-term outcomes. *Hepatology*. 2011; 54(3): 820-8.
- 29) Lyra AC, Soares MB, da Silva LF, et al. Feasibility and safety of autologous bone marrow mononuclear cell transplantation in patients with advanced chronic liver disease. *World J Gastroenterol*. 2007; 13(7): 1067-73.
- 30) Lyra AC, Soares MB, da Silva LF, et al. Infusion of autologous bone marrow mononuclear cells through hepatic artery results in a short-term improvement of liver function in patients with chronic liver disease: a pilot randomized controlled study. *Eur J Gastroenterol Hepatol*. 2010; 22(1): 33-42.
- 31) Mohamadnejad M, Alimoghaddam K, Mohyeddin-Bonab M, et al. Phase I trial of autologous bone marrow mesenchymal stem cell transplantation in patients with decompensated liver cirrhosis. *Arch Iran Med*. 2007; 10(4): 459-66.
- 32) Kharaziha P, Hellstrom PM, Noorinayer B, et al. Improvement of liver function in liver cirrhosis patients after autologous mesenchymal stem cell injection: a phase I-II clinical trial. *Eur J Gastroenterol Hepatol*. 2009; 21(10): 1199-205.
- 33) Amer ME, El-Sayed SZ, El-Kheir WA, et al. Clinical and laboratory evaluation of patients with end-stage liver cell failure injected with bone marrow-derived hepatocyte-like cells. *Eur J Gastroenterol Hepatol*. 2011; 23(10): 936-41.

Bone marrow cell-based regenerative therapy for liver cirrhosis

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Although the efficacy of this treatment modality needs to be evaluated in more detail in a large number of patients, regenerative therapy using bone marrow cells for advanced liver diseases has considerable potential.

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Abstract

Bone marrow cells are capable of differentiation into liver cells. Therefore, transplantation of bone marrow cells has considerable potential as a future therapy for regeneration of damaged liver tissue. Autologous bone marrow infusion therapy has been applied to patients with liver cirrhosis, and improvement of liver function parameters has been demonstrated. In this review, we summarize clinical trials of regenerative therapy using bone marrow cells for advanced liver diseases including cirrhosis, as well as topics pertaining to basic *in vitro* or *in vivo* approaches in order to outline the essentials of this novel treatment modality.

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Key words: Bone marrow; Liver regeneration; Cirrhosis; Stem cell; Transplantation

Core tip: Bone marrow cells, which include multipotent progenitor cells, are capable of differentiation into liver cells. Autologous bone marrow infusion therapy has been applied to cirrhotic patients, and improvement of liver function parameters has been demonstrated.

INTRODUCTION

Bone marrow cells (BMCs) are capable of differentiating into liver cells^[1-4] because they include stem cells known as multipotent adult progenitor cells^[5,6]. These cells have been shown to produce albumin when cultured with hepatocyte growth factor (HGF)^[7] and various liver-specific proteins, including albumin, when cultured with mature hepatocytes^[8]. Using cells obtained with a negatively selective magnetic cell separation system for efficient sorting of rat BMCs enriched with stem cells, we have shown that BMCs differentiate into cells expressing liver-specific genes when cultured with mature hepatocytes or HGF^[9]. As there is now much evidence indicating that BMCs can differentiate into cells resembling liver cells *in vitro*^[6-11], the characteristics of such BMCs are of great interest in the context of liver-regenerative medicine^[12-14].

Liver cirrhosis is the end stage of chronic liver disease, and is associated with many serious systemic complications resulting from both liver failure and portal hypertension. This condition has a poor prognosis and is difficult to treat. Therefore, development of an effective liver-regenerative therapy for liver cirrhosis is an urgent priority. Liver transplantation is the only curative remedy for cirrhotic patients, but is associated with many problems such as donor shortage, surgical complications,

rejection and high cost. As an alternative approach, regenerative cell therapy using stem cells is now attracting attention. Multipotent stem cells present in bone marrow are a particularly promising candidate for this purpose. In this review, we summarize clinical trials of liver-regenerative therapy using BMCs for advanced liver diseases including cirrhosis, as well as topics pertaining to basic *in vitro* or *in vivo* approaches in order to outline the essentials of this novel treatment modality.

MIGRATION AND ENGRAFTMENT OF TRANSPLANTED BMCs TO THE INJURED LIVER IN STUDIES USING ANIMAL MODELS

Although BMCs can show liver cell lineage differentiation *in vitro*, an understanding of the dynamics of transplanted BMCs *in vivo* is essential for the development of BMC-based regenerative therapy. In this context, two important issues need to be clarified: (1) How do transplanted BMCs migrate to and engraft in the liver? and (2) Is there a relationship between the degree of liver damage and the extent of migration of transplanted cells? A previous study using model rats with carbon tetrachloride (CCl₄)-induced liver injury has demonstrated that transplanted BMCs derived from transgenic rats expressing green fluorescent protein^[15] in the spleen migrated to and remained in the periportal area of the recipient's damaged liver^[16]. These transplanted cells expressed liver cell markers such as alpha-fetoprotein as well as Notch signaling markers for stem cells, suggesting that the BMCs retained in the recipient liver possess the potential to differentiate into liver cells.

Migration of transplanted BMCs to the liver after injection into the spleen has been compared in two models of liver injury induced by administration of CCl₄ and 2-acetylaminofluorene (2-AAF)^[17], respectively, focusing particularly on differences in levels of liver mRNA for growth factors such as HGF and fibroblast growth factor (FGF), which have been shown to be responsible for efficient liver cell lineage differentiation of BMCs^[9,18,19]. Interestingly, transplanted BMCs were found to engraft into CCl₄-induced injured liver characterized by submassive hepatic necrosis and induction of high levels of HGF and FGF, but not into liver damaged by 2-AAF^[20]. A higher degree of HGF induction is characteristic of more severe liver damage^[21,22]. These findings suggest that transplanted BMCs migrate more effectively to a liver with greater damage, and that this transplantation approach would be clinically promising for treatment of advanced liver diseases. However, further studies are needed to clarify the factors produced by both BMCs and hepatocytes that contribute to better differentiation of BMCs into liver cells *in vivo*, thus improving the effectiveness of BMC transplantation.

HUMORAL FACTORS BENEFICIAL FOR LIVER REGENERATION AFTER BMC TRANSPLANTATION

The degree of liver function and fibrosis, as well as survival rate, have been shown to improve significantly after BMC transplantation in animal models of severe liver injury^[23,24]. With regard to the mechanisms of liver regeneration resulting from BMC transplantation, many of the physiological and regenerative roles of transplanted BMCs remain unclear. However, it can be said with certainty that humoral factors produced in the liver during the regenerative process after BMC transplantation have a crucial role in both improvement of liver fibrosis and liver cell lineage differentiation of stem cells originating from BMCs and hepatic epithelial stem cells.

Improvement of liver fibrosis results from fibrolysis through the proteolytic action of BMC-induced factors. In this context, matrix metalloproteinase (MMP) activity is particularly noteworthy^[25]. Sakaida *et al.*^[23] showed that BMC transplantation ameliorated liver fibrosis in the CCl₄-induced liver-injury model, and that the fibrolytic change was attributable to MMP-9 secreted by BMCs that had migrated to fibrotic areas of the liver.

The liver cell lineage differentiation of BMCs occurs through the cooperative action of a variety of growth factors such as HGF or FGF induced in the injured liver^[11,20,26]. Such differentiation may be accompanied by early elevation of the apolipoprotein A1 level in serum and liver^[27]. Administration of FGF2 in combination with BMC transplantation synergistically ameliorates liver fibrosis in models of liver injury induced by CCl₄^[28]. In addition, in severe liver injury where hepatocyte proliferation is strongly inhibited, hepatic stem cells such as oval cells are induced and show differentiation toward a liver cell lineage, thus leading to liver regeneration^[29,30].

As BMC transplantation is successfully adaptable to cases of severe liver injury, it has been hypothesized that transplanted BMCs interact with hepatic epithelial stem cells and influence the subsequent proliferation and differentiation of stem cells. Studies of the interaction between BMCs and hepatic stem cells can provide new insight into the mechanisms of recovery from severe liver damage through liver regeneration after BMC transplantation. In this context, *in vitro* analysis using a system for co-culture of BMCs and an established epithelial hepatic stem cell line has been conducted. Haga *et al.*^[31] demonstrated that the expression of *FGF2* mRNA was upregulated in BMCs co-cultured with hepatic stem cells, and that expression of mRNAs for both albumin and tyrosine aminotransferase, representative of mature hepatic cells, became detectable in hepatic stem cells after culture with FGF2 protein. Thus, BMCs stimulate both proliferation and differentiation of hepatic stem cells into the hepatocyte lineage, and FGF2 is one of the factors produced by interaction with BMCs, which stimulates

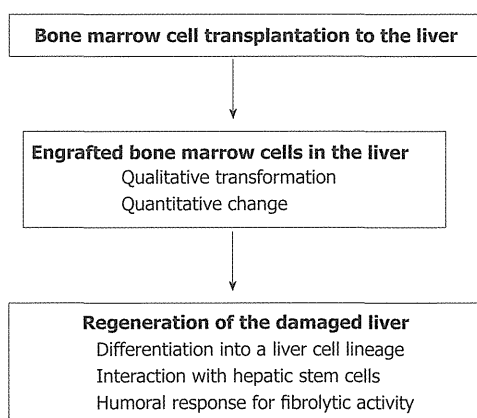


Figure 1 Putative action of transplanted bone marrow cells that include multipotent stem cells for regeneration of damaged liver.

such differentiation. Cross-talk between bone marrow stem cells and hepatic epithelial stem cells may underlie the process of liver regeneration, and this is an area of interest for future investigation. Figure 1 shows an overall representation of the putative action of transplanted BMCs in the regeneration of damaged liver.

CLINICAL TRIALS OF BMC TRANSPLANTATION FOR ADVANCED LIVER DISEASES

BMC transplantation has received increasing attention as a promising therapy for advanced and severe liver diseases such as cirrhosis. Clinical trials of BMC administration to patients with advanced liver diseases have been performed, and improvement of liver function parameters such as the serum level of albumin, Child-Pugh score or Model for Endstage Liver Disease score have been reported^[32-40]. Another study has shown that intraportal administration of autologous CD133⁺ BMCs and subsequent portal venous embolization of right liver segments resulted in a 2.5-fold increase in the mean proliferation rate of the left lateral segment, in comparison with controls not receiving BM transfusion^[41]. These findings suggest that transplanted BMCs have a potential role in liver regeneration and proliferate in the recipient liver. Recently, autologous BMC transplantation - a technique named autologous BMC infusion (ABMi) therapy - has been applied to multi-center patients with liver cirrhosis due to hepatitis C^[42], hepatitis B^[43] and excess alcohol intake^[44] using almost the same protocol, and a series of studies have demonstrated improvement of the serum albumin level, leading to improvement of the Child-Pugh score.

Although BMC administration for advanced liver diseases including cirrhosis is an attractive strategy in the field of cell therapy for liver regeneration, many concerns need to be addressed^[45-47]. As *in vitro* and *in vivo* experiments have clearly shown, BMCs induce fibrolysis and show hepatocyte differentiation, and they may interact

with hepatic epithelial stem cells to aid their differentiation into the hepatocyte lineage. However, it is still unclear how infused BMCs work to improve liver function in humans. A clinical trial of ABMi for patients with cirrhosis demonstrated that the number of AFP-positive cells increased significantly in the liver relative to the situation before ABMi^[42]. In addition, ABMi appeared to induce hepatocyte proliferation in the liver, as expression of proliferating cell nuclear antigen, a marker of hepatocyte proliferation, was significantly increased after ABMi in comparison with the pretreatment situation. Although these findings suggest that transplanted BMCs have a potential role in liver regeneration and proliferate in the recipient liver, it remains unknown whether fully functional hepatocytes are induced by ABMi. The characteristics of stem cells present among BMCs that show hepatocyte differentiation require further elucidation.

The factors that determine the difference between effectiveness and non-effectiveness of ABMi are unclear. Collateral circulation resulting from the portal vein disorganization that characterizes liver cirrhosis may affect the flow and effective migration of infused BMCs to the liver, and thus migration of infused cells to the liver may partly depend on the portal venous pressure. In addition, the expression levels of cellular adhesion molecules associated with the attachment of infused cells to liver tissue may vary a great deal among patients. The long-term effectiveness of this therapy in terms of survival rate has not been demonstrated. These issues should be evaluated by a randomized controlled trial involving a large number of patients. Additionally, other issues that impact the efficacy of this therapy, *i.e.*, the long-term culture conditions optimal for stocking BMCs for repeated infusion, the optimal cell population to employ, the optimal number of cells to infuse, the effectiveness of repeated infusion and the optimal route for cell delivery need to be investigated further.

In conclusion, regenerative therapy using BMCs for advanced liver diseases including cirrhosis has considerable potential. Further studies are needed to develop a better method of BMC transplantation that can contribute to improvement of liver function and to clarify the long-term effectiveness of this therapy.

REFERENCES

- 1 Petersen BE, Bowen WC, Patrene KD, Mars WM, Sullivan AK, Murase N, Boggs SS, Greenberger JS, Goff JP. Bone marrow as a potential source of hepatic oval cells. *Science* 1999; **284**: 1168-1170 [PMID: 10325227 DOI: 10.1126/science.284.5417.1168]
- 2 Alison MR, Poulson R, Jeffery R, Dhillion AP, Quaglia A, Jacob J, Novelli M, Prentice G, Williamson J, Wright NA. Hepatocytes from non-hepatic adult stem cells. *Nature* 2000; **406**: 257 [PMID: 10917519 DOI: 10.1038/35018642]
- 3 Theise ND, Badve S, Saxena R, Henegariu O, Sell S, Crawford JM, Krause DS. Derivation of hepatocytes from bone marrow cells in mice after radiation-induced myeloablation. *Hepatology* 2000; **31**: 235-240 [PMID: 10613752 DOI: 10.1002/hep.510310135]
- 4 Theise ND, Nimmakayalu M, Gardner R, Illei PB, Morgan G,

- Teperman L, Henegariu O, Krause DS. Liver from bone marrow in humans. *Hepatology* 2000; **32**: 11-16 [PMID: 10869283 DOI: 10.1053/jhep.2000.9124]
- 5 **Jiang Y**, Jahagirdar BN, Reinhardt RL, Schwartz RE, Keene CD, Ortiz-Gonzalez XR, Reyes M, Lenvik T, Lund T, Blackstad M, Du J, Aldrich S, Lisberg A, Low WC, Largaespada DA, Verfaillie CM. Pluripotency of mesenchymal stem cells derived from adult marrow. *Nature* 2002; **418**: 41-49 [PMID: 12077603 DOI: 10.1038/nature00870]
 - 6 **Schwartz RE**, Reyes M, Koodie L, Jiang Y, Blackstad M, Lund T, Lenvik T, Johnson S, Hu WS, Verfaillie CM. Multipotent adult progenitor cells from bone marrow differentiate into functional hepatocyte-like cells. *J Clin Invest* 2002; **109**: 1291-1302 [PMID: 12021244]
 - 7 **Oh SH**, Miyazaki M, Kouchi H, Inoue Y, Sakaguchi M, Tsuji T, Shima N, Higashio K, Namba M. Hepatocyte growth factor induces differentiation of adult rat bone marrow cells into a hepatocyte lineage in vitro. *Biochem Biophys Res Commun* 2000; **279**: 500-504 [PMID: 11118315 DOI: 10.1006/bbrc.2000.3985]
 - 8 **Avital I**, Inderbitzin D, Aoki T, Tyan DB, Cohen AH, Ferrareso C, Rozga J, Arnaout WS, Demetriou AA. Isolation, characterization, and transplantation of bone marrow-derived hepatocyte stem cells. *Biochem Biophys Res Commun* 2001; **288**: 156-164 [PMID: 11594767 DOI: 10.1006/bbrc.2001.5712]
 - 9 **Okumoto K**, Saito T, Hattori E, Ito JI, Adachi T, Takeda T, Sugahara K, Watanabe H, Saito K, Togashi H, Kawata S. Differentiation of bone marrow cells into cells that express liver-specific genes in vitro: implication of the Notch signals in differentiation. *Biochem Biophys Res Commun* 2003; **304**: 691-695 [PMID: 12727209 DOI: 10.1016/S0006-291X(03)00637-5]
 - 10 **Miyazaki M**, Akiyama I, Sakaguchi M, Nakashima E, Okada M, Kataoka K, Huh NH. Improved conditions to induce hepatocytes from rat bone marrow cells in culture. *Biochem Biophys Res Commun* 2002; **298**: 24-30 [PMID: 12379214 DOI: 10.1016/S0006-291X(02)02340-9]
 - 11 **Okumoto K**, Saito T, Hattori E, Ito JI, Suzuki A, Misawa K, Ishii R, Karasawa T, Haga H, Sanjo M, Takeda T, Sugahara K, Saito K, Togashi H, Kawata S. Differentiation of rat bone marrow cells cultured on artificial basement membrane containing extracellular matrix into a liver cell lineage. *J Hepatol* 2005; **43**: 110-116 [PMID: 15893847 DOI: 10.1016/j.jhep.2005.01.037]
 - 12 **Mitaka T**. Hepatic stem cells: from bone marrow cells to hepatocytes. *Biochem Biophys Res Commun* 2001; **281**: 1-5 [PMID: 11178951 DOI: 10.1006/bbrc.2001.4270]
 - 13 **Faris RA**, Konkin T, Halpert G. Liver stem cells: a potential source of hepatocytes for the treatment of human liver disease. *Artif Organs* 2001; **25**: 513-521 [PMID: 11493271 DOI: 10.1046/j.1525-1594.2001.025007513.x]
 - 14 **Forbes S**, Vig P, Poulsom R, Thomas H, Alison M. Hepatic stem cells. *J Pathol* 2002; **197**: 510-518 [PMID: 12115866 DOI: 10.1002/path.1163]
 - 15 **Hakamata Y**, Tahara K, Uchida H, Sakuma Y, Nakamura M, Kume A, Murakami T, Takahashi M, Takahashi R, Hirabayashi M, Ueda M, Miyoshi I, Kasai N, Kobayashi E. Green fluorescent protein-transgenic rat: a tool for organ transplantation research. *Biochem Biophys Res Commun* 2001; **286**: 779-785 [PMID: 11520065 DOI: 10.1006/bbrc.2001.5452]
 - 16 **Okumoto K**, Saito T, Hattori E, Ito JI, Suzuki A, Misawa K, Sanjo M, Takeda T, Sugahara K, Saito K, Togashi H, Kawata S. Expression of Notch signalling markers in bone marrow cells that differentiate into a liver cell lineage in a rat transplant model. *Hepatol Res* 2005; **31**: 7-12 [PMID: 15652464 DOI: 10.1016/j.hepres.2004.11.005]
 - 17 **Petersen BE**, Zajac VF, Michalopoulos GK. Hepatic oval cell activation in response to injury following chemically induced periportal or pericentral damage in rats. *Hepatology* 1998; **27**: 1030-1038 [PMID: 9537443 DOI: 10.1002/hep.510270419]
 - 18 **Sekhon SS**, Tan X, Micsenyi A, Bowen WC, Monga SP. Fibroblast growth factor enriches the embryonic liver cultures for hepatic progenitors. *Am J Pathol* 2004; **164**: 2229-2240 [PMID: 15161655 DOI: 10.1016/S0002-9440(10)63779-0]
 - 19 **Lange C**, Bassler P, Lioznov MV, Bruns H, Kluth D, Zander AR, Fiegel HC. Hepatocytic gene expression in cultured rat mesenchymal stem cells. *Transplant Proc* 2005; **37**: 276-279 [PMID: 15808618 DOI: 10.1016/j.transproceed.2004.11.087]
 - 20 **Okumoto K**, Saito T, Haga H, Hattori E, Ishii R, Karasawa T, Suzuki A, Misawa K, Sanjo M, Ito JI, Sugahara K, Saito K, Togashi H, Kawata S. Characteristics of rat bone marrow cells differentiated into a liver cell lineage and dynamics of the transplanted cells in the injured liver. *J Gastroenterol* 2006; **41**: 62-69 [PMID: 16501859 DOI: 10.1007/s00535-005-1723-8]
 - 21 **Tsubouchi H**, Kawakami S, Hirono S, Miyazaki H, Kimoto M, Arima T, Sekiyama K, Yoshida M, Arakaki N, Daikuhara Y. Prediction of outcome in fulminant hepatic failure by serum human hepatocyte growth factor. *Lancet* 1992; **340**: 307 [PMID: 1353217 DOI: 10.1016/0140-6736(92)92396-W]
 - 22 **Maher JJ**. Cell-specific expression of hepatocyte growth factor in liver. Upregulation in sinusoidal endothelial cells after carbon tetrachloride. *J Clin Invest* 1993; **91**: 2244-2252 [PMID: 7683700 DOI: 10.1172/JCI116451]
 - 23 **Sakaida I**, Terai S, Yamamoto N, Aoyama K, Ishikawa T, Nishina H, Okita K. Transplantation of bone marrow cells reduces CCl4-induced liver fibrosis in mice. *Hepatology* 2004; **40**: 1304-1311 [PMID: 15565662 DOI: 10.1002/hep.20452]
 - 24 **Terai S**, Sakaida I, Yamamoto N, Omori K, Watanabe T, Ohata S, Katada T, Miyamoto K, Shinoda K, Nishina H, Okita K. An in vivo model for monitoring trans-differentiation of bone marrow cells into functional hepatocytes. *J Biochem* 2003; **134**: 551-558 [PMID: 14607982 DOI: 10.1093/jb/mvg173]
 - 25 **Haraguchi T**, Tani K, Koga M, Oda Y, Itamoto K, Yamamoto N, Terai S, Sakaida I, Nakazawa H, Taura Y. Matrix metalloproteinases (MMPs) activity in cultured canine bone marrow stromal cells (BMSCs). *J Vet Med Sci* 2012; **74**: 633-636 [PMID: 22167104 DOI: 10.1292/jvms.11-0395]
 - 26 **Ishikawa T**, Terai S, Urata Y, Marumoto Y, Aoyama K, Sakaida I, Murata T, Nishina H, Shinoda K, Uchimura S, Hamamoto Y, Okita K. Fibroblast growth factor 2 facilitates the differentiation of transplanted bone marrow cells into hepatocytes. *Cell Tissue Res* 2006; **323**: 221-231 [PMID: 16228231 DOI: 10.1007/s00441-005-0077-0]
 - 27 **Yokoyama Y**, Terai S, Ishikawa T, Aoyama K, Urata Y, Marumoto Y, Nishina H, Nakamura K, Okita K, Sakaida I. Proteomic analysis of serum marker proteins in recipient mice with liver cirrhosis after bone marrow cell transplantation. *Proteomics* 2006; **6**: 2564-2570 [PMID: 16548057 DOI: 10.1002/pmic.200500018]
 - 28 **Ishikawa T**, Terai S, Urata Y, Marumoto Y, Aoyama K, Murata T, Mizunaga Y, Yamamoto N, Nishina H, Shinoda K, Sakaida I. Administration of fibroblast growth factor 2 in combination with bone marrow transplantation synergistically improves carbon-tetrachloride-induced liver fibrosis in mice. *Cell Tissue Res* 2007; **327**: 463-470 [PMID: 17093919 DOI: 10.1007/s00441-006-0334-x]
 - 29 **Shiota G**, Kunisada T, Oyama K, Udagawa A, Nomi T, Tanaka K, Tsutsumi A, Isono M, Nakamura T, Hamada H, Sakatani T, Sell S, Sato K, Ito H, Kawasaki H. In vivo transfer of hepatocyte growth factor gene accelerates proliferation of hepatic oval cells in a 2-acetylaminofluorene/partial hepatectomy model in rats. *FEBS Lett* 2000; **470**: 325-330 [PMID: 10745090 DOI: 10.1016/S0014-5793(00)01337-5]
 - 30 **Hu Z**, Evarts RP, Fujio K, Marsden ER, Thorgerisson SS. Expression of hepatocyte growth factor and c-met genes during hepatic differentiation and liver development in the rat. *Am J Pathol* 1993; **142**: 1823-1830 [PMID: 8506951]
 - 31 **Haga H**, Saito T, Okumoto K, Ugajin S, Sato C, Ishii R, Nishise Y, Ito J, Watanabe H, Saito K, Togashi H, Kawata S.

- Enhanced expression of fibroblast growth factor 2 in bone marrow cells and its potential role in the differentiation of hepatic epithelial stem-like cells into the hepatocyte lineage. *Cell Tissue Res* 2011; **343**: 371-378 [PMID: 21152936 DOI: 10.1007/s00441-010-1093-2]
- 32 **Lyra AC**, Soares MB, da Silva LF, Fortes MF, Silva AG, Mota AC, Oliveira SA, Braga EL, de Carvalho WA, Genser B, dos Santos RR, Lyra LG. Feasibility and safety of autologous bone marrow mononuclear cell transplantation in patients with advanced chronic liver disease. *World J Gastroenterol* 2007; **13**: 1067-1073 [PMID: 17373741]
- 33 **Mohamadnejad M**, Namiri M, Bagheri M, Hashemi SM, Ghanaati H, Zare Mehrjardi N, Kazemi Ashtiani S, Malekzadeh R, Baharvand H. Phase 1 human trial of autologous bone marrow-hematopoietic stem cell transplantation in patients with decompensated cirrhosis. *World J Gastroenterol* 2007; **13**: 3359-3363 [PMID: 17659676]
- 34 **Gordon MY**, Levicar N, Pai M, Bachellier P, Dimarakis I, Al-Allaf F, M'Hamdi H, Thalji T, Welsh JP, Marley SB, Davies J, Dazzi F, Marelli-Berg F, Tait P, Playford R, Jiao L, Jensen S, Nicholls JP, Ayav A, Nohandani M, Farzaneh F, Gaken J, Dodge R, Alison M, Apperley JF, Lechler R, Habib NA. Characterization and clinical application of human CD34+ stem/progenitor cell populations mobilized into the blood by granulocyte colony-stimulating factor. *Stem Cells* 2006; **24**: 1822-1830 [PMID: 16556705]
- 35 **Pai M**, Zacharoulis D, Milicevic MN, Helmy S, Jiao LR, Levicar N, Tait P, Scott M, Marley SB, Jestice K, Glibetic M, Bansi D, Khan SA, Kyriakou D, Rountas C, Thillainayagam A, Nicholls JP, Jensen S, Apperley JF, Gordon MY, Habib NA. Autologous infusion of expanded mobilized adult bone marrow-derived CD34+ cells into patients with alcoholic liver cirrhosis. *Am J Gastroenterol* 2008; **103**: 1952-1958 [PMID: 18637092 DOI: 10.1111/j.1572-0241.2008.01993.x]
- 36 **Han Y**, Yan L, Han G, Zhou X, Hong L, Yin Z, Zhang X, Wang S, Wang J, Sun A, Liu Z, Xie H, Wu K, Ding J, Fan D. Controlled trials in hepatitis B virus-related decompensate liver cirrhosis: peripheral blood monocyte transplant versus granulocyte-colony-stimulating factor mobilization therapy. *Cytotherapy* 2008; **10**: 390-396 [PMID: 18574771 DOI: 10.1080/14653240802129901]
- 37 **Peng L**, Xie DY, Lin BL, Liu J, Zhu HP, Xie C, Zheng YB, Gao ZL. Autologous bone marrow mesenchymal stem cell transplantation in liver failure patients caused by hepatitis B: short-term and long-term outcomes. *Hepatology* 2011; **54**: 820-828 [PMID: 21608000 DOI: 10.1002/hep.24434]
- 38 **Mohamadnejad M**, Alimoghaddam K, Mohyeddin-Bonab M, Bagheri M, Bashtar M, Ghanaati H, Baharvand H, Ghavamzadeh A, Malekzadeh R. Phase 1 trial of autologous bone marrow mesenchymal stem cell transplantation in patients with decompensated liver cirrhosis. *Arch Iran Med* 2007; **10**: 459-466 [PMID: 17903050]
- 39 **Kharaziha P**, Hellström PM, Noorinayer B, Farzaneh F, Aghajani K, Jafari F, Telkabadi M, Atashi A, Honardoost M, Zali MR, Soleimani M. Improvement of liver function in liver cirrhosis patients after autologous mesenchymal stem cell injection: a phase I-II clinical trial. *Eur J Gastroenterol Hepatol* 2009; **21**: 1199-1205 [PMID: 19455046 DOI: 10.1097/MEG.0b013e32832a1f6c]
- 40 **Amer ME**, El-Sayed SZ, El-Kheir WA, Gabr H, Gomaa AA, El-Noomani N, Hegazy M. Clinical and laboratory evaluation of patients with end-stage liver cell failure injected with bone marrow-derived hepatocyte-like cells. *Eur J Gastroenterol Hepatol* 2011; **23**: 936-941 [PMID: 21900788 DOI: 10.1097/MEG.0b013e3283488b00]
- 41 **am Esch JS**, Knoefel WT, Klein M, Ghodsizad A, Fuerst G, Poll LW, Piechaczek C, Burchardt ER, Feifel N, Stoldt V, Stockschlader M, Stoecklein N, Tustas RY, Eisenberger CF, Peiper M, Häussinger D, Hosch SB. Portal application of autologous CD133+ bone marrow cells to the liver: a novel concept to support hepatic regeneration. *Stem Cells* 2005; **23**: 463-470 [PMID: 15790766]
- 42 **Terai S**, Ishikawa T, Omori K, Aoyama K, Marumoto Y, Urata Y, Yokoyama Y, Uchida K, Yamasaki T, Fujii Y, Okita K, Sakaida I. Improved liver function in patients with liver cirrhosis after autologous bone marrow cell infusion therapy. *Stem Cells* 2006; **24**: 2292-2298 [PMID: 16778155]
- 43 **Kim JK**, Park YN, Kim JS, Park MS, Paik YH, Seok JY, Chung YE, Kim HO, Kim KS, Ahn SH, Kim do Y, Kim MJ, Lee KS, Chon CY, Kim SJ, Terai S, Sakaida I, Han KH. Autologous bone marrow infusion activates the progenitor cell compartment in patients with advanced liver cirrhosis. *Cell Transplant* 2010; **19**: 1237-1246 [PMID: 20525430 DOI: 10.3727/096368910X506863]
- 44 **Saito T**, Okumoto K, Haga H, Nishise Y, Ishii R, Sato C, Watanabe H, Okada A, Ikeda M, Togashi H, Ishikawa T, Terai S, Sakaida I, Kawata S. Potential therapeutic application of intravenous autologous bone marrow infusion in patients with alcoholic liver cirrhosis. *Stem Cells Dev* 2011; **20**: 1503-1510 [PMID: 21417817 DOI: 10.1089/scd.2011.0074]
- 45 **Kallis YN**, Alison MR, Forbes SJ. Bone marrow stem cells and liver disease. *Gut* 2007; **56**: 716-724 [PMID: 17145739]
- 46 **Lorenzini S**, Andreone P. Stem cell therapy for human liver cirrhosis: a cautious analysis of the results. *Stem Cells* 2007; **25**: 2383-2384 [PMID: 17540855]
- 47 **Terai S**, Tanimoto H, Maeda M, Zaito J, Hisanaga T, Iwamoto T, Fujisawa K, Mizunaga Y, Matsumoto T, Urata Y, Marumoto Y, Hidaka I, Ishikawa T, Yokoyama Y, Aoyama K, Tsuchiya M, Takami T, Omori K, Yamamoto N, Segawa M, Uchida K, Yamasaki T, Okita K, Sakaida I. Timeline for development of autologous bone marrow infusion (ABMi) therapy and perspective for future stem cell therapy. *J Gastroenterol* 2012; **47**: 491-497 [PMID: 22488349 DOI: 10.1007/s00535-012-0580-5]

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WJG 20th Anniversary Special Issues (2): Hepatitis C virus

Transmission of hepatitis C virus: Self-limiting hepatitis or chronic hepatitis?

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Abstract

It has been suggested that hepatitis C virus (HCV) is selectively transmitted to a new host as an infectious clone from multiple HCV variants (quasispecies) in the donor. Most individuals with HCV infection develop chronic hepatitis, but approximately 15%-40% of them clear the virus spontaneously and the hepatitis is resolved in a self-limiting manner in the acute phase of infection. This difference in the outcome of acute hepatitis C is attributable to both viral characteristics and genetic regulation of infection. In particular, the evolutionary dynamics of the infecting virus and host genetic polymorphisms pertaining mainly to the immune system, including polymorphisms in the region of the Interleukin 28B gene encoding interferon- λ -3, are associated with susceptibility to HCV infection.

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Key words: Hepatitis C; Spontaneous clearance; Interleukin 28B; Single nucleotide polymorphism; Interferon- λ

Core tip: Most individuals with hepatitis C virus (HCV)

infection develop chronic hepatitis, but in some the hepatitis is resolved in a self-limiting manner in the acute phase of infection. What factors are responsible for this difference in the outcome of hepatitis C? The evolutionary dynamics of the infecting virus and host genetic polymorphisms pertaining mainly to the immune system, including the Interleukin 28B gene, as well as susceptibility to HCV infection, are important in determining the outcome of infection.

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INTRODUCTION

Hepatitis C virus (HCV) infection is a major threat to public health, and about 170 million people are estimated to be infected worldwide with a potential risk of progression to cirrhosis and hepatocellular carcinoma^[1,2]. This review summarizes the two current topics of HCV study: the transmission mode of HCV with multiple variants (quasispecies) and the factors associated with susceptibility to HCV infection, with special reference to viral characteristics and host genetic variation.

MODE OF HCV TRANSMISSION: HOW IS HCV WITH MULTIPLE VARIANTS TRANSMITTED?

HCV shows significant genetic heterogeneity among isolates, and the degree of variability is unevenly distributed throughout the viral genome: some regions are conserved and some are highly variable^[3]. In particular, the hyper-

variable region 1 (HVR1) of the *HCV E2* gene encoding a putative envelope glycoprotein mutates at a high rate, resulting in a wide spectrum of mutants referred to as “quasispecies” during infection^[4,5]. Some virions may contain defective RNA genomes, which also affect the infectivity and replicability of the virus^[6]. The mixture of clones present determines the biological and immunological properties of the virus.

How is HCV with multiple variants (quasispecies) transmitted to the new host? Does the status of transmitted HCV consist of multiple clones or a selected single clone? The transmission mode of HCV has been investigated by sequencing of the recovered viral genome from both donor and recipient^[7,8]. HCV infection in human communities has occurred sporadically because no effective neutralizing vaccine against HCV has been developed. In particular, HCV infection in health-care workers through exposure to patient’s blood due to a needle stick accident or accidental droplet transmission is a serious problem^[9-12]. We previously reported a case of HCV infection resulting from a needle stick accident, and had an opportunity to investigate how HCV variants from the donor are transmitted to the recipient by comparing the HCV HVR1 genome encoding the envelope E2 protein recovered from the serum of both the donor and recipient^[7]. In this case, we had observed the recipient before the onset of hepatitis and collected serum samples after obtaining informed consent. Thus, we were able to compare the HCV HVR1 genome between the donor’s HCV at inoculation and the recipient’s HCV just after onset of viremia. Interestingly, a minor subset of the donor’s HCV clones was selectively transmitted to the recipient, and this selection determined the predominant clone in the new host. Several clones that appeared to stem from the recipient’s predominant clone had one amino acid change within the HVR1 region during this short period. This particular case progressed to chronic hepatitis, and the same phenomenon has been demonstrated in the case of acute, self-limiting hepatitis^[8]. These data suggest that a minor clone of the donor’s HCV is transmitted and adapts to the new host. The precise mechanism of this viral selection in the initial phase of transmission has not been elucidated.

The simplicity of the transmitted viral strain in the initial phase of infection may explain some of the important clinical manifestations. Anti-viral therapy using interferon elicits a favorable response in the acute phase of HCV infection^[13-16]. In addition, if a single strain is transmitted selectively in the initial phase of infection, this specific strain may be one of the factors determining disease activity. In fact, a study using a model of HCV transmission has demonstrated that a specific HCV strain recovered from a patient with fulminant hepatitis caused unusually severe hepatitis in a chimpanzee to which it was transmitted^[17]. At present, the specific strain of HCV responsible for progressive liver disease cannot be discriminated from viral quasispecies in contaminated blood. Further investigation would be useful for clarifying the

specific viral strain responsible for the disease, and such efforts would be important for planning future strategies for the development of an effective therapeutic vaccine.

SELF-LIMITING HEPATITIS OR CHRONIC HEPATITIS? HOW IS SUSCEPTIBILITY TO HCV DETERMINED?

The spontaneous clearance rate of HCV in the acute phase of infection

Most individuals with HCV infection fail to clear the virus and develop chronic hepatitis with a risk of progression to cirrhosis and hepatocellular carcinoma. However, a small proportion of individuals are known to show resolution of the infection in a self-limiting manner. The rate of spontaneous viral clearance in acute HCV infection is reported to be approximately 15%-40% of all HCV-infected individuals^[18-20]. Although differences in study populations such as race may influence the clearance rate in each cohort, a systematic review of 31 studies has estimated this rate to be 26%^[20]. We have previously reported a Japanese population-based cohort study of the natural history of HCV infection in an area where community-acquired acute hepatitis C is endemic; here, the spontaneous viral clearance rate was estimated to be approximately 20%^[21,22]. What is the difference between self-limiting resolution of hepatitis and progression to chronic hepatitis? Comparative studies of this issue have focused on both viral characteristics and genetic regulation.

Viral characteristics influencing the outcome of acute hepatitis C

After the establishment of HCV infection, the viral genome mutates at a high rate, especially in the HVR1 of the HCV E2 region. The evolutionary dynamics of the infected virus are associated with the outcome of acute hepatitis C; genetic stasis and a high rate of evolution of HCV HVR1 are associated with resolution of infection in self-limiting hepatitis and progression to chronic infection, respectively^[23]. The case we experienced progressed to chronic infection and 8 of 30 homogeneously predominant HCV HVR1 clones recovered from the recipient developed one amino acid mutation within this region during a short period of only 6 wk after infection^[7]. As for the relationship between the viral load at the time of infection and the outcome of acute HCV infection, a recent study has shown that a high viral load in the initial phase of infection is associated with spontaneous viral clearance, leading to self-limiting resolution of hepatitis^[24]. A high viral load may trigger strong innate immunity in the acute phase. However, it has also been reported that viral clearance may occur after a low infectious dose of HCV has been transmitted^[25]. In addition, spontaneous viral clearance rarely occurs in the chronic phase of HCV infection where a low viral load is associated with spontaneous clearance^[26]. The spontaneous clearance of HCV may thus depend on the immune system of indi-

viduals rather than the viral load. Further studies using a greater number of cohorts are needed to clarify the relationship between spontaneous viral clearance and the initial viral load, as well as the degree of induction of the innate immune response.

Genetic regulation of HCV infection

HCV-specific humoral and cellular immune responses are detectable in infected individuals, and a strong immune response against HCV favors viral clearance^[18,27]. Genetic variation in host genes involved in immune response is likely to account for the difference in outcome. In particular, induction of natural killer (NK) cells in the innate immune response during the acute phase of infection plays a crucial role in resolving HCV infection. We have previously reported differences in genetic variations between HCV-infected individuals with and without viremia in the Japanese population^[22], where a single nucleotide polymorphism (SNP) of transforming growth factor (TGF)- β 1, which suppresses the proliferation and cytotoxicity of NK cells (the -509CC genotype or -509C allele), was associated with high HCV clearance rates and low transcriptional activity of TGF- β 1^[28]. The killer cell immunoglobulin-like receptor (KIR) and its human leukocyte antigen (HLA) have been reported to influence the outcome of HCV infection. Combinations of genotypes involving genes encoding the inhibitory NK cell receptor KIR2DL3 and HLA-C1 ligand directly influence HCV clearance in Caucasians and African Americans with an expected low infectious dose of HCV^[25]. These data suggest that a diminished inhibitory effect of NK cells resulting from such gene regulation confers protection against HCV.

In a recent genome-wide association study, SNPs in the region of the Interleukin 28B (*IL28B*) gene encoding interferon- λ -3 were shown to be closely associated with the virologic response of HCV to antiviral therapy^[29-31]. Patients carrying an *IL28B* homozygote for the major alleles of rs12979860 (CC genotype)^[29] or rs8099917 (TT genotype)^[30] show a greater propensity to achieve a sustained virologic response to pegylated interferon- α and ribavirin therapy than those carrying an *IL28B* heterozygote or homozygote for its minor allele. This SNP (rs12979860) also influences the outcome of HCV infection in the context of natural history; the CC genotype enhances resolution of HCV infection with spontaneous clearance among individuals of European and African ancestry^[32]. This CC genotype has also been reported to be associated with a higher rate of spontaneous clearance in Asian populations^[33]. In addition, a recent study has demonstrated that SNPs in the region of *IL28B* (rs12979860) and HLA class II (rs4273729) are independently associated with spontaneous resolution of HCV infection in individuals of European and African ancestry^[34]. A prospective follow-up study of patients who developed acute hepatitis C also revealed a strong correlation between the *IL28B* C allele at rs12979860 and clearance^[24]. Taken together, the SNP of *IL28B* (rs12979860) can be a marker

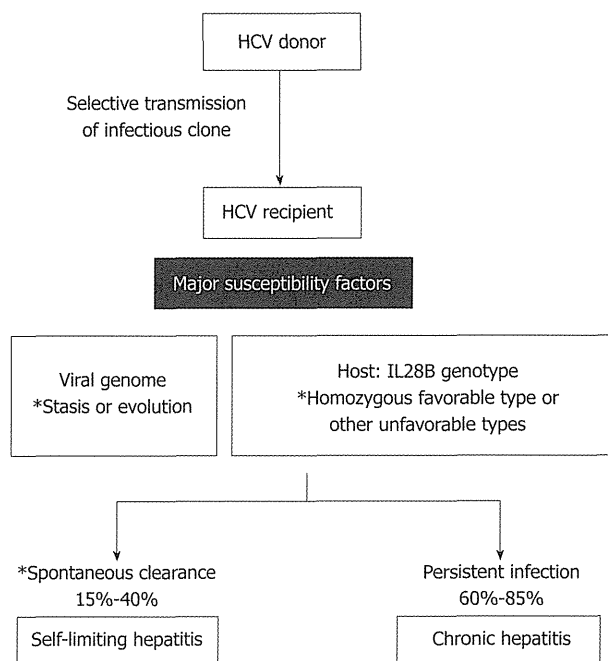


Figure 1 Transmission of hepatitis C virus, and the significance of viral and host factors for predicting the outcome of infection. HCV: Hepatitis C virus; IL28B: Interleukin 28B.

for indicating whether immediate antiviral treatment needs to be started in patients with acute hepatitis C^[35]. Recently, upstream of the *IL28B* gene, a dinucleotide variant ss469415590 (TT or Δ G), in which ss469415590 (Δ G) activates the *IFNL4* gene encoding interferon- λ -4 protein through a genome frameshift, has been reported to be more strongly associated with HCV clearance in individuals of African ancestry than the SNP of *IL28B* (rs12979860), but comparable to that in Europeans and Asians^[36]. This variant is in high linkage disequilibrium with rs12979860, and further investigations are expected to elucidate the functional role of ss469415590 (Δ G) that activates the *IFNL4* gene in association with the innate immune response to HCV.

CONCLUSION

Both the viral characteristics of an infecting clone and genetic regulation of infection by the host determine differences in the outcome of acute HCV infection (Figure 1). The evolutionary dynamics of the virus and genetic polymorphisms in the host pertaining mainly to the immune system influence susceptibility to HCV. In particular, the discovery of SNPs in the region of the *IL28B* gene has led to the characterization of a novel genetic marker of hepatitis C that is able to predict self-limiting viral clearance in the acute phase of infection as well as the response to antiviral therapy.

REFERENCES

- 1 Cohen J. The scientific challenge of hepatitis C. *Science* 1999; 285: 26-30 [PMID: 10428695]

- 2 **Kiyosawa K**, Sodeyama T, Tanaka E, Gibo Y, Yoshizawa K, Nakano Y, Furuta S, Akahane Y, Nishioka K, Purcell RH. Interrelationship of blood transfusion, non-A, non-B hepatitis and hepatocellular carcinoma: analysis by detection of antibody to hepatitis C virus. *Hepatology* 1990; **12**: 671-675 [PMID: 2170265]
- 3 **Major ME**, Feinstone SM. The molecular virology of hepatitis C. *Hepatology* 1997; **25**: 1527-1538 [PMID: 9185778]
- 4 **Ogata N**, Alter HJ, Miller RH, Purcell RH. Nucleotide sequence and mutation rate of the H strain of hepatitis C virus. *Proc Natl Acad Sci USA* 1991; **88**: 3392-3396 [PMID: 1849654]
- 5 **Kurosaki M**, Enomoto N, Marumo F, Sato C. Rapid sequence variation of the hypervariable region of hepatitis C virus during the course of chronic infection. *Hepatology* 1993; **18**: 1293-1299 [PMID: 8244252]
- 6 **Martell M**, Esteban JI, Quer J, Genescà J, Weiner A, Esteban R, Guardia J, Gómez J. Hepatitis C virus (HCV) circulates as a population of different but closely related genomes: quasi-species nature of HCV genome distribution. *J Virol* 1992; **66**: 3225-3229 [PMID: 1313927]
- 7 **Saito T**, Watanabe H, Shao L, Okumoto K, Hattori E, Sanjo M, Misawa K, Suzuki A, Takeda T, Sugahara K, Ito JI, Saito K, Togashi H, Kawata S. Transmission of hepatitis C virus quasispecies between human adults. *Hepatol Res* 2004; **30**: 57-62 [PMID: 15519268]
- 8 **Liu CH**, Chen BF, Chen SC, Lai MY, Kao JH, Chen DS. Selective transmission of hepatitis C virus quasi species through a needlestick accident in acute resolving hepatitis. *Clin Infect Dis* 2006; **42**: 1254-1259 [PMID: 16586384]
- 9 **Sulkowski MS**, Ray SC, Thomas DL. Needlestick transmission of hepatitis C. *JAMA* 2002; **287**: 2406-2413 [PMID: 11988061]
- 10 **Mizuno Y**, Suzuki K, Mori M, Hayashi K, Owaki T, Hayashi H, Kumada K, Ohba K, Mizokami M. Study of needlestick accidents and hepatitis C virus infection in healthcare workers by molecular evolutionary analysis. *J Hosp Infect* 1997; **35**: 149-154 [PMID: 9049819]
- 11 **Frijstein G**, Hortensius J, Zaaijer HL. Needlestick injuries and infectious patients in a major academic medical centre from 2003 to 2010. *Neth J Med* 2011; **69**: 465-468 [PMID: 22058270]
- 12 **Suzuki K**, Mizokami M, Lau JY, Mizoguchi N, Kato K, Mizuno Y, Sodeyama T, Kiyosawa K, Gojbori T. Confirmation of hepatitis C virus transmission through needlestick accidents by molecular evolutionary analysis. *J Infect Dis* 1994; **170**: 1575-1578 [PMID: 7527827]
- 13 **Omata M**, Yokosuka O, Takano S, Kato N, Hosoda K, Imazeki F, Tada M, Ito Y, Ohto M. Resolution of acute hepatitis C after therapy with natural beta interferon. *Lancet* 1991; **338**: 914-915 [PMID: 1681268]
- 14 **Sharland M**, Patton MA, Hill L. Ectrodactyly of hands and feet in a child with a complex translocation including 7q21.2. *Am J Med Genet* 1991; **39**: 413-414 [PMID: 1877619 DOI: 10.1093/jac/dkn346]
- 15 **Fabrizi F**, Dixit V, Messa P, Martin P. Interferon therapy of acute hepatitis C in dialysis patients: meta-analysis. *J Viral Hepat* 2012; **19**: 784-791 [PMID: 23043385 DOI: 10.1111/j.1365-2893.2012.01607.x]
- 16 **Nunnari G**, Montineri A, Portelli V, Savalli F, Fatuzzo F, Cacopardo B. The use of peginterferon in monotherapy or in combination with ribavirin for the treatment of acute hepatitis C. *Eur Rev Med Pharmacol Sci* 2012; **16**: 1013-1016 [PMID: 22913149]
- 17 **Farci P**, Munoz SJ, Shimoda A, Govindarajan S, Wong DC, Coiana A, Peddis G, Rubin R, Purcell RH. Experimental transmission of hepatitis C virus-associated fulminant hepatitis to a chimpanzee. *J Infect Dis* 1999; **179**: 1007-1011 [PMID: 10068599]
- 18 **Di Bisceglie AM**. Natural history of hepatitis C: its impact on clinical management. *Hepatology* 2000; **31**: 1014-1018 [PMID: 10733560]
- 19 **Gerlach JT**, Diepolder HM, Zchoval R, Gruener NH, Jung MC, Ulsenheimer A, Schraut WW, Schirren CA, Waechter M, Backmund M, Pape GR. Acute hepatitis C: high rate of both spontaneous and treatment-induced viral clearance. *Gastroenterology* 2003; **125**: 80-88 [PMID: 12851873]
- 20 **Micallef JM**, Kaldor JM, Dore GJ. Spontaneous viral clearance following acute hepatitis C infection: a systematic review of longitudinal studies. *J Viral Hepat* 2006; **13**: 34-41 [PMID: 16364080]
- 21 **Ishibashi M**, Shinzawa H, Kuboki M, Tsuchida H, Takahashi T. Prevalence of inhabitants with anti-hepatitis C virus antibody in an area following an acute hepatitis C epidemic: age-and area-related features. *J Epidemiol* 1996; **6**: 1-7 [PMID: 8795951]
- 22 **Saito T**, Ji G, Shinzawa H, Okumoto K, Hattori E, Adachi T, Takeda T, Sugahara K, Ito JI, Watanabe H, Saito K, Togashi H, Ishii K, Matsuura T, Inageda K, Muramatsu M, Kawata S. Genetic variations in humans associated with differences in the course of hepatitis C. *Biochem Biophys Res Commun* 2004; **317**: 335-341 [PMID: 15063762]
- 23 **Farci P**, Shimoda A, Coiana A, Diaz G, Peddis G, Melpolder JC, Strazzer A, Chien DY, Munoz SJ, Balestrieri A, Purcell RH, Alter HJ. The outcome of acute hepatitis C predicted by the evolution of the viral quasispecies. *Science* 2000; **288**: 339-344 [PMID: 10764648]
- 24 **Liu L**, Fisher BE, Thomas DL, Cox AL, Ray SC. Spontaneous clearance of primary acute hepatitis C virus infection correlated with high initial viral RNA level and rapid HVR1 evolution. *Hepatology* 2012; **55**: 1684-1691 [PMID: 22234804 DOI: 10.1002/hep.25575]
- 25 **Khakoo SI**, Thio CL, Martin MP, Brooks CR, Gao X, Astemborski J, Cheng J, Goedert JJ, Vlahov D, Hilgartner M, Cox S, Little AM, Alexander GJ, Cramp ME, O'Brien SJ, Rosenberg WM, Thomas DL, Carrington M. HLA and NK cell inhibitory receptor genes in resolving hepatitis C virus infection. *Science* 2004; **305**: 872-874 [PMID: 15297676]
- 26 **Watanabe H**, Saito T, Shinzawa H, Okumoto K, Hattori E, Adachi T, Takeda T, Sugahara K, Ito JI, Saito K, Togashi H, Suzuki R, Hayashi M, Miyamura T, Matsuura Y, Kawata S. Spontaneous elimination of serum hepatitis C virus (HCV) RNA in chronic HCV carriers: a population-based cohort study. *J Med Virol* 2003; **71**: 56-61 [PMID: 12858409]
- 27 **Rehermann B**, Nascimbeni M. Immunology of hepatitis B virus and hepatitis C virus infection. *Nat Rev Immunol* 2005; **5**: 215-229 [PMID: 15738952]
- 28 **Kimura T**, Saito T, Yoshimura M, Yixuan S, Baba M, Ji G, Muramatsu M, Kawata S. Association of transforming growth factor-beta 1 functional polymorphisms with natural clearance of hepatitis C virus. *J Infect Dis* 2006; **193**: 1371-1374 [PMID: 16619184]
- 29 **Ge D**, Fellay J, Thompson AJ, Simon JS, Shianna KV, Urban TJ, Heinzen EL, Qiu P, Bertelsen AH, Muir AJ, Sulkowski M, McHutchison JG, Goldstein DB. Genetic variation in IL28B predicts hepatitis C treatment-induced viral clearance. *Nature* 2009; **461**: 399-401 [PMID: 19684573 DOI: 10.1038/nature08309]
- 30 **Tanaka Y**, Nishida N, Sugiyama M, Kurosaki M, Matsuura K, Sakamoto N, Nakagawa M, Korenaga M, Hino K, Hige S, Ito Y, Mita E, Tanaka E, Mochida S, Murawaki Y, Honda M, Sakai A, Hiasa Y, Nishiguchi S, Koike A, Sakaida I, Imamura M, Ito K, Yano K, Masaki N, Sugauchi F, Izumi N, Tokunaga K, Mizokami M. Genome-wide association of IL28B with response to pegylated interferon-alpha and ribavirin therapy for chronic hepatitis C. *Nat Genet* 2009; **41**: 1105-1109 [PMID: 19749757 DOI: 10.1038/ng.449]
- 31 **Balagopal A**, Thomas DL, Thio CL. IL28B and the control of hepatitis C virus infection. *Gastroenterology* 2010; **139**: 1865-1876 [PMID: 20950615 DOI: 10.1053/j.gastro.2010.10.004]

- 32 **Thomas DL**, Thio CL, Martin MP, Qi Y, Ge D, O'Huigin C, Kidd J, Kidd K, Khakoo SI, Alexander G, Goedert JJ, Kirk GD, Donfield SM, Rosen HR, Tobler LH, Busch MP, McHutchison JG, Goldstein DB, Carrington M. Genetic variation in IL28B and spontaneous clearance of hepatitis C virus. *Nature* 2009; **461**: 798-801 [PMID: 19759533 DOI: 10.1038/nature08463]
- 33 **Hung CH**, Chang KC, Lu SN, Wang JH, Chen CH, Lee CM, Hu TH. Spontaneous clearance of hepatitis C virus in an interleukin 28B favorable genotype highly prevalent area. *Hepatology* 2013; **57**: 2089-2090 [PMID: 22886694 DOI: 10.1002/hep.26002]
- 34 **Duggal P**, Thio CL, Wojcik GL, Goedert JJ, Mangia A, Lantich R, Kim AY, Lauer GM, Chung RT, Peters MG, Kirk GD, Mehta SH, Cox AL, Khakoo SI, Alric L, Cramp ME, Donfield SM, Edlin BR, Tobler LH, Busch MP, Alexander G, Rosen HR, Gao X, Abdel-Hamid M, Apps R, Carrington M, Thomas DL. Genome-wide association study of spontaneous resolution of hepatitis C virus infection: data from multiple cohorts. *Ann Intern Med* 2013; **158**: 235-245 [PMID: 23420232 DOI: 10.7326/0003-4819-158-4-201302190-00003]
- 35 **Mangia A**, Santoro R, Copetti M, Massari M, Piazzolla V, Spada E, Cappucci G, Missale G, Mottola L, Agostinacchio E, Mauro Ld, Zuccaro O, Maio P, Pellegrini F, Folgori A, Ferrari C. Treatment optimization and prediction of HCV clearance in patients with acute HCV infection. *J Hepatol* 2013; **59**: 221-228 [PMID: 23587473 DOI: 10.1016/j.jhep.2013.04.007]
- 36 **Prokunina-Olsson L**, Muchmore B, Tang W, Pfeiffer RM, Park H, Dickensheets H, Hergott D, Porter-Gill P, Mumy A, Kohaar I, Chen S, Brand N, Tarway M, Liu L, Sheikh F, Astemborski J, Bonkovsky HL, Edlin BR, Howell CD, Morgan TR, Thomas DL, Rehermann B, Donnelly RP, O'Brien TR. A variant upstream of IFNL3 (IL28B) creating a new interferon gene IFNL4 is associated with impaired clearance of hepatitis C virus. *Nat Genet* 2013; **45**: 164-171 [PMID: 23291588 DOI: 10.1038/ng.2521]

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Possible autoimmune hepatitis induced after chronic active Epstein–Barr virus infection

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Abstract Chronic active Epstein–Barr virus infection (CAEBV) can be manifested in a variety of systemic conditions, including interstitial pneumonia, malignant lymphoma, and coronary aneurysm. Sometimes it may be associated with hepatic failure, although the mechanism underlying CAEBV-related hepatotoxicity remains unclear. We encountered a case of autoimmune hepatitis (AIH) associated with CAEBV. A 61-year-old male was referred to our hospital because of abnormal liver enzyme levels after initial diagnosis of CAEBV had been made by laboratory tests and liver biopsy. On admission, positivity for anti-nuclear antibody was evident, and examination of the liver biopsy specimen showed findings compatible with AIH. Steroid administration was initiated, and the liver function parameters subsequently improved. Although phenotypic changes in liver biopsy specimens are rare in this condition, the present case could provide clues to the possible pathogenesis of AIH.

Keywords Autoimmune hepatitis · Steroids · Anti-nuclear antibody · Epstein–Barr virus

Abbreviations

CAEBV Chronic active Epstein–Barr virus infection
AIH Autoimmune hepatitis
ANA Anti-nuclear antibody

Introduction

Chronic active Epstein–Barr virus infection (CAEBV) is a rare condition producing chronic or repeated symptoms mimicking infectious mononucleosis (IM), and an abnormal pattern of anti-EBV antibodies. CAEBV is also known to demonstrate a variety of clinical symptoms, possibly due to an abnormal lymphoproliferative reaction [1]. Accumulation of reported cases has suggested that the prognosis of CAEBV is worse than was originally considered [2, 3]. Although the pathogenesis of CAEBV remains mostly unclear, the source of the EBV-infected cells, and whether they are T cells or natural killer cells appear to be determinants of outcome [4]. CAEBV can cause various forms of liver injury, ranging from simple liver enzyme abnormality to hepatic failure [5].

Autoimmune hepatitis (AIH) is a classical autoimmune liver disease with characteristic clinical manifestations including a female predominance, presence of autoantibodies (especially anti-nuclear antibody; ANA) and hyperimmunoglobulinemia. AIH sometimes presents as an acute form, in which ANA is frequently negative and serum immunoglobulin (Ig) G levels are normal. As is the case for other autoimmune liver diseases, the pathogenesis of AIH is unknown. However, there are a number of possible factors involved in initiation of the abnormal autoimmune reaction in AIH, including a genetic predisposition to exogenous infections.

Here we report a case of AIH that was possibly induced by CAEBV.

Case report

A 61-year-old male was referred to our hospital because of persistent liver enzyme abnormalities and general fatigue.

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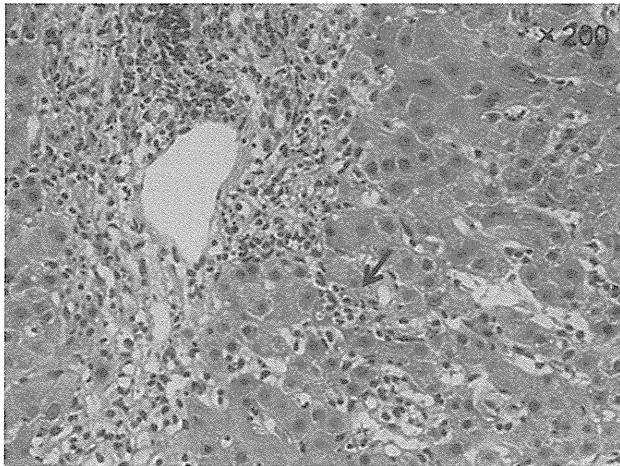


Fig. 1 Initial liver biopsy revealed mononuclear cell infiltration in the sinusoids, suggesting changes after EBV infection

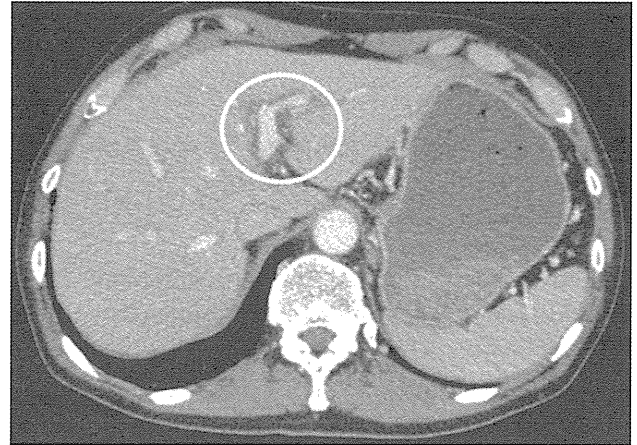


Fig. 2 Hypo-enhanced area reflecting periportal edema and presence of mild hepatosplenomegaly were noted by enhanced abdominal CT scan

Table 1 Laboratory tests on admission to our hospital

T-Bil	2.7 mg/dL	WBC	$3.50 \times 10^3/\mu\text{L}$
D-Bil	1.7 mg/dL	Neut	58.3 %
AST	696 IU/L	Lymph	30.5 %
ALT	634 IU/L	Mono	6.0 %
LDH	400 IU/L	Eosino	3.3 %
GGTP	312 IU/L	Baso	1.9 %
ALP	366 IU/L	RBC	$3.80 \times 10^6/\mu\text{L}$
TP	7.9 g/dL	Hb	12.4 g/dL
Alb	3.8 g/dL	Platelets	128,000/ μL
BUN	11 mg/dL	PT (%)	93 %
Crea	0.54 mg/dL	T-cho	172 mg/dL
Na	140 mEq/L	TG	283 mg/dL
K	4.2 mEq/L	HbA1c	4.6 %
Cl	104 mEq/L	HBsAg	Negative
CRP	0.84 mg/dL	HBsAb	Negative
IgG	2,440 mg/dL	HBcAb	Negative
IgA	331 mg/dL	IgM HAAb	Negative
IgM	107 mg/dL	IgM HBcAb	Negative
AFP	50.6 ng/mL	HCV Ab	Negative
		HIV Ab	Negative
(Urine)		ANA Ab	$\times 320$
pH	6.0	Anti-LKM Ab	Negative
Sugar	–	Anti M2 Ab	<5.0 Negative
Protein	–	EBV VCA IgM	Negative
Blood	–	EBV VCA IgG	$\times 2,560$
Keton	–	EBNA	$\times 160$
Bilirubin	1+	EB EA-DR IgG	$\times 20$
		EB EA-DR IgA	$< \times 10$
Bone marrow: no atypical cell, no hemophagocytosis		EBV DNA (PCR) ^a	1.3×10^3 copy
		sIL-2 receptor	1,300 IU/mL

^a EBV monoclonality was negative

His medical history was unremarkable except for rhinosinusitis at 20 years of age and hyperuricemia at 54 years of age. The clinical history of allergy for insect bite was negative. His family history included colon cancer in the father and essential hypertension in the mother. He had been a non-smoker and a moderate drinker (500 mL of beer per day). In December 2010, he had felt general fatigue and visited his family practitioner. Symptoms such as lymphadenopathy and fever were negative except for persistent general fatigue. Clinical laboratory tests revealed liver enzyme abnormalities, and the patient was referred to the community hospital, where CAEBV was initially suspected on the basis of positive peripheral blood EBV DNA with 1.8×10^2 copies/ 10^6 cells (normal limit $< 2.0 \times 10^1$ copies/ 10^6 cells, by real-time polymerase chain reaction; PCR) [6]. Liver biopsy was performed, and the findings were consistent with liver injury associated with CAEBV (Fig. 1), mainly non-specific mononuclear cell infiltration of the hepatic sinusoids. However, the patient's liver enzymes persistently fluctuated, and therefore he was referred to our hospital in May 2011. Physical examination on admission showed a height of 170.0 cm, body weight 65 kg, body temperature 36.3 °C, blood pressure 124/79 mmHg, clear consciousness, icteric conjunctiva, and hepatomegaly palpable for 2 finger breadths without splenomegaly. No systemic lymph node swelling was evident. The results of laboratory tests on admission are shown in Table 1—these included an elevated transaminase level, a high IgG level, elevated ANA titers ($80\times$ to $320\times$), and a high titer of EBV viral capsid antigen (VCA) IgG. We could not perform HLA-DR genotyping assay with the patient. Imaging (contrast-enhanced computed tomography; CT) revealed a hypo-enhanced area possibly reflecting periportal edema, and mild hepatosplenomegaly (Fig. 2). Positive EBV DNA (1.3×10^3 copies/ 10^6 cells) in

peripheral blood on admission strongly suggested the presence of persistent CAEBV. However, in view of the increased titer of ANA, we performed a second liver biopsy, which revealed fibrous enlargement of the portal area, presence of interfacial hepatitis, and plasma cell-predominant infiltration, thus supporting a diagnosis of AIH (Fig. 3). With regard to the diagnosis of AIH, application of two scoring systems—(1) revised scoring system formulated in 1999 [7] scored 12 points, and (2) simplified criteria for the international AIH guidelines [8] scored 6 points—suggested probable AIH with the present case. The

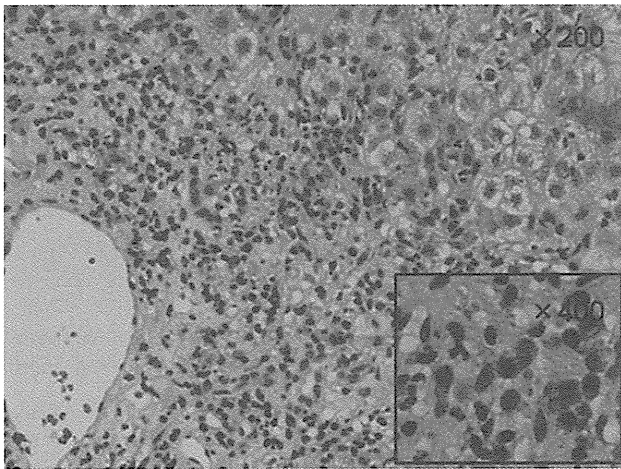
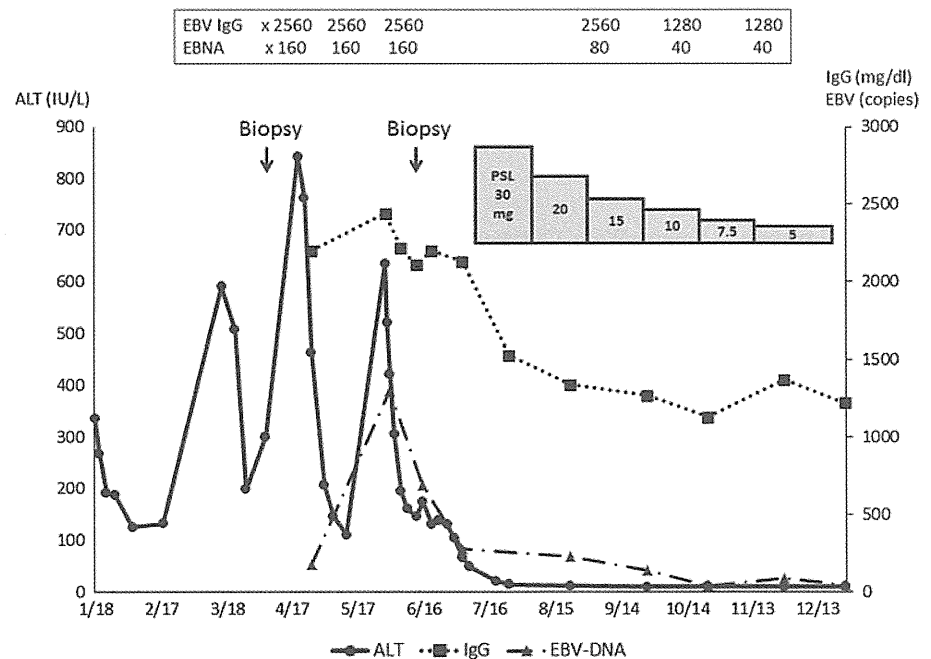


Fig. 3 A second liver biopsy at our hospital before steroid therapy demonstrated fibrotic periportal enlargement, interface hepatitis, and plasma cell infiltration. EBV DNA was positive in the tissue by qualitative PCR assay. $\times 200$ (lower panel $\times 400$)

Fig. 4 Clinical course of the present case. Levels of liver enzymes and serum IgG, EBV viral load, EBV IgG, EBNA were ameliorated after steroid administration



tissue was positive for EBV DNA by PCR qualitative assay (detection limit 2.0×10^1 copies/ 1.0×10^6 cells equivalent DNA), whereas both the latent membrane protein 1 and EBV-encoded RNA in situ hybridization were negative. Considering the findings of liver biopsy and serological changes, we started steroid administration at 30 mg/day. The dose of steroid was gradually tapered, and the results of serum laboratory tests (alanine aminotransferase [ALT] and IgG levels) improved accordingly. Moreover, the amount of peripheral blood EBV DNA levels, the titers of EBV-VCA IgG and Epstein–Barr nuclear antigen (EBNA) decreased. After steroid administration for 6 months, the patient has remained free of signs and symptoms (Fig. 4).

Discussion

The present case had been initially diagnosed as CAEBV. However, the persistent liver enzyme abnormalities prompted us to perform further examinations. Elevation of the ANA titer from $80\times$ (at the initial community hospital) to $320\times$ (at our hospital), along with the pathological changes seen in both of the liver biopsy samples, suggested a diagnosis of AIH, probably resulting from EBV infection.

The pathogenesis of AIH is still unknown, although genetic factors, drug-induced autoimmunity, viral infections, and environmental factors have been thought to trigger AIH [9, 10]. Among these triggering factors, hepatitis A viral infection has been reported [11, 12], although EBV infection appears to be rare in this context. Proposed

mechanisms responsible for inducing autoimmune reactions have included the molecular mimic theory, involving possible similarities between virus-derived exogenous amino acid sequences and host autoantigens [13], although the precise details remain unclear.

On the other hand, the diagnostic criteria for CAEBV formulated by the EBV infection study group in 2003 include (1) persistent or relapsing symptoms of infectious mononucleosis, (2) abnormal antibody reactions to EBV, associated with an elevated titer of VCA or early antigen (EA) antibody, or an increased EBV viral load in the affected organ (including peripheral blood), and (3) a chronic disease course distinct from other known diseases [14]. In the present case, we confirmed that persistent liver enzyme abnormalities were present for >3 months, together with an increased EBV viral load in blood and positive EBV-DNA in hepatic tissue. Compared to the reported case [5], our case seems to be atypical since our case lacked traditional high fever, lymphadenopathy, and allergy to insect bite for CAEBV. However, our patient was older (61 years) than the previous case (14 years) [5], and this may be related to the reason why typical symptoms were missing in the current case. Our patient was treated with prednisolone, and this led to amelioration of the serum ALT and IgG levels, and a decline of the EBV viral load. Antiviral drugs, cytokine therapy, immunosuppressive therapy, cellular transplantation, and allogeneic bone marrow transplantation therapy are reportedly effective for CAEBV, although none of them are established forms of treatment [14]. In our case, immunosuppressive therapy using steroids seems to have been effective so far, and there has been no disease flare-up for 2 years.

In conclusion, we have reported a case of AIH possibly resulting from EBV infection. A possible diagnosis of AIH should be considered in patients with prolonged liver dysfunction subsequent to EBV infection.

Disclosures

Conflict of Interest: Yoshiko Wada, Chikako Sato, Kyoko Tomita, Rika Ishii-Aso, Hiroaki Haga, Kazuo Okumoto, Yuko Nishise, Hisayoshi Watanabe, Tadasih Togashi, Yoshiyuki Ueno declare that they have no conflict of interest.

Human/Animal Rights: All procedures followed were in accordance with the ethical standards of the responsible committee on human experimentation (institutional and national) and with the Helsinki Declaration of 1975, as revised in 2008(5).

Informed Consent: Informed consent was obtained from all patients for being included in the study.

References

1. Rickinson AB. Chronic, symptomatic Epstein–Barr virus-infections. *Immunol Today*. 1986;7:13–4.
2. Ohshima K, Suzumiya J, Sugihara M, et al. Clinicopathological study of severe chronic active Epstein–Barr virus infection that developed in association with lymphoproliferative disorder and/or hemophagocytic syndrome. *Pathol Int*. 1998;48:934–43.
3. Okano M, Matsumoto S, Osato T, et al. Severe chronic active Epstein–Barr-virus infection syndrome. *Clin Microbiol Rev*. 1991;4:129–35.
4. Kimura H, Morishima T, Kanegane H, et al. Prognostic factors for chronic active Epstein–Barr virus infection. *J Infect Dis*. 2003;187:527–33.
5. Kimura H, Hoshino Y, Kanegane H, et al. Clinical and virologic characteristics of chronic active Epstein–Barr virus infection. *Blood*. 2001;98:280–6.
6. Kimura H, Morita M, Yabuta Y, et al. Quantitative analysis of Epstein–Barr virus load by using a real-time PCR assay. *J Clin Microbiol*. 1999;37:132–6.
7. Alvarez E, Berg PA, Bianchi FB, et al. International autoimmune hepatitis group report: review of criteria for diagnosis of autoimmune hepatitis. *J Hepatol*. 1999;31:929–38.
8. Hennes EM, Zeniya M, Czaja AJ, et al. Simplified criteria for the diagnosis of autoimmune hepatitis. *Hepatology*. 2008;48:169–76.
9. Krawitt EL. Medical progress: autoimmune hepatitis. *N Engl J Med*. 2006;354:54–66.
10. Vento S, Guella L, Mirandola F, et al. Epstein–Barr-virus as a trigger for autoimmune hepatitis in susceptible individuals. *Lancet*. 1995;346:608–9.
11. Vento S, Garofano T, Diperrì G, et al. Identification of hepatitis-a virus as a trigger for autoimmune chronic hepatitis type-1 in susceptible individuals. *Lancet*. 1991;337:1183–7.
12. Huppertz HI, Treichel U, Gassel AM, et al. Autoimmune hepatitis following hepatitis-a virus-infection. *J Hepatol*. 1995;23:204–8.
13. Wucherpfennig KW. Mechanisms for the induction of autoimmunity by infectious agents. *J Clin Invest*. 2001;108:1097–104.
14. Okano M, Kawa K, Kimura H, et al. Proposed guidelines for diagnosing chronic active Epstein–Barr virus infection. *Am J Hematol*. 2005;80:64–9.