cules based on these lead compounds and to perform appropriate structure–activity relationship studies.

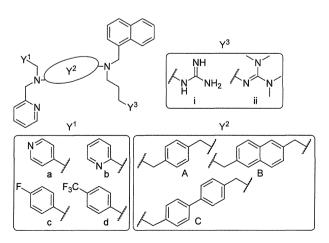
## Results and Discussion

#### Design

We initially designed compounds that contain 1,4-phenylenedimethanamine, one amino group of which is linked to guanidine and naphthalene moieties, and the other to 2-pyridylmethyl and naphthalene analogues, as shown in Figure 2. The

Figure 2. New compounds containing the 1,4-phenylenedimethanamine structure.

adoption of these functional moieties is based on structures of compound **3**, which contains 4-fluorobenzyl and 2-pyridylmethyl amino groups, and compound **4**, which contains two naphthalene moieties. Thus, 2-methylquinoline, 2-methylnaphthalene, 2-methoxy-6-methylnaphthalene, 2-bromo-6-methylnaphthalene, and 2-fluoro-6-methylnaphthalene (X-CH<sub>2</sub>) moieties were introduced on a nitrogen atom of the 1,4-phenylene-dimethanamino group in compounds **19 a-c** and **23 d,e**. Furthermore, compounds with 1,4-phenylenedimethanamine, naphthalene-2,6-diyldimethanamine, and [1,1'-biphenyl]-4,4'-diyldimethanamine structures as spacer templates (H<sub>2</sub>N-Y<sup>2</sup>-NH<sub>2</sub>) were designed as shown in Figure 3 to refine the spacers. Monocyclic aromatic groups, 4- or 2-pyridylmethyl, 4-fluorobenzyl, and 4-trifluoromethylbenzyl groups (Y¹-CH<sub>2</sub>) were intro-



**Figure 3.** New compounds containing the 1,4-phenylenedimethanamine, naphthalene-2,6-diyldimethanamine, and [1,1'-biphenyl]-4,4'-diyldimethanamine structures.

duced on a nitrogen atom of the above spacer templates, and guanidino and tetramethylguanidino groups were used as substituents for Y<sup>3</sup> in compounds **37** a–**42** d.

## Chemistry

The synthesis of compounds 19a-c is shown in Scheme 1. Condensation of *N*-Boc-3-aminopropylbromide (6) and *N*-Ns-aminonaphthalen-1-yl-methane (9; Ns=2-nitrobenzenesulfonyl) followed by removal of the Ns group produced the amine 11. The *N*-Ns-4-aminomethylbenzoic acid derived Weinreb

amide 14 was treated with DIBAL to afford the corresponding aldehyde, the reductive amination of which was performed by treatment with amine 11 to afford the tertiary amine 15. Introduction of a 2-pyridinylmethyl group into 15 by means of Mitsunobu reaction followed by removal of the Ns group yielded amine 17. Introduction of 2-methylquinoline, 2-methylnaphthalene, and 2-methoxy-6-meth-

ylnaphthalene groups by reductive amination of 17 produced amines 18a-c, respectively, and subsequent removal of the Boc group followed by N-guanylation yielded the desired compounds 19a-c.

As shown in Scheme 2, introduction of 2-bromo-6-methylnaphthalene and 2-fluoro-6-methylnaphthalene moieties into 15 by Mitsunobu reaction followed by removal of the Ns group yielded amines 21 d and 21 e, respectively. Introduction of a 2-pyridinylmethyl group by reductive amination of 21 d and 21 e produced amines 22 d and 22 e, respectively, and subsequent removal of the Boc group followed by N-guanylation yielded the desired compounds 23 d and 23 e.

Scheme 3 shows the synthesis of 37a–39d and 40a–42d. Introduction of 4-pyridylmethyl, 2-pyridylmethyl, or 4-fluorobenzyl and 4-trifluoromethylbenzyl groups into *N*-Ns-(pyridin-2-ylmethyl)amide 25 by Mitsunobu reaction followed by removal of the Ns group yielded amines 27a–d, respectively. Treatment of 1,4-phenylenedimethane, naphthalene-2,6-diyldimethane, and [1,1'-biphenyl]-4,4'-diyldimethane-derived dibromides 28–30 with amine 11 afforded the tertiary amines 31–33, respectively. Subsequent treatment of 31–33 with amines 27a–d yielded amines 34a–36d. Subsequent removal of the Boc group followed by N-guanylation and N-tetramethylguanylation yielded the desired compounds 37a–39d and 40a–42d, respectively.

## **Biological studies**

The CXCR4 binding affinity of the synthesized compounds was assessed through inhibition of [125]CXCL12 binding to Jurkat cells, which express CXCR4. The activity was evaluated for compounds **19a-c** containing 2-methylquinoline, 2-methylnaphthalene, 2-methoxy-6-methylnaphthalene, and **23 d**,e,

Scheme 1. Reagents and conditions: a) Boc<sub>2</sub>O, Et<sub>3</sub>N, MeOH/MeCN (1:1), 98%; b) LiAlH<sub>4</sub>, THF, 0°C, 89%; c) NsCl, Et<sub>3</sub>N, THF, 78%; d) K<sub>2</sub>CO<sub>3</sub>, DMF, 60°C, 96%; e) PhSH, K<sub>2</sub>CO<sub>3</sub>, DMF, 95%; f) NsCl, Et<sub>3</sub>N, THF, 88%; q) EDCI·HCl, HOBt·H<sub>2</sub>O, NHCH<sub>3</sub>(OCH<sub>3</sub>)·HCl, Et<sub>3</sub>N, DMF, 88%; h) DIBAL/n-hexane, CH<sub>2</sub>Cl<sub>2</sub>, -78°C; i) NaBH(OAc)<sub>3</sub>, AcOH, amine 11, 1,2-dichloroethane, 43% (two steps); j) PPh<sub>3</sub>, DEAD, 2-pyridinemethanol, THF, 76%; k) PhSH, K<sub>2</sub>CO<sub>3</sub>, DMF, 87%; l) NaBH-(OAc)<sub>3</sub>, AcOH, X-CHO, 1,2-dichloroethane, 70% (18a), 69% (18b), 49% (18c); m) 4 M HCl/dioxane; n) N,N-diisopropylethylamine, 1-amidinopyrazole-HCl, DMF, 28% (19 a), 58% (19 b), 46% (19 c) (two steps). Ns = 2-nitrobenzenesulfonyl.

with 2-bromo-6-methylnaphthalene and 2-fluoro-6-methylnaphthalene moieties, respectively (X-CH<sub>2</sub>), introduced on a nitrogen atom of the 1,4-phenylenedimethanamino group. The percent inhibition data for all compounds at 10 µm are listed in Table 1. With the exception of 19c, which contains a 2-me-

Table 1. CXCR4 binding affinities of compounds 19a-c and 23 d,e.						
Compd	X <sup>[a]</sup>	Inhibition [%] <sup>[b]</sup>				
19a	a	14.4 ± 1.0				
19 b	b	$7.0 \pm 0.6$				
19 c	С	0				
23 d	d	$9.0\pm2.2$				
23 e	е	$9.5 \pm 1.3$				
FC131	-	100				

[a] The structures of X (a-e) are shown in Figure 2. [b] CXCR4 binding affinity was assessed based on inhibition of [1251]CXCL12 binding to Jurkat cells; percent inhibition values for all compounds at 10 µм were calculated relative to that of FC131 (100%).

thoxynaphthalene group, the compounds showed significant but very weak binding affinity. With an electron-donating methoxy group, the 2-methoxynaphthalene moiety is an electron-rich aromatic group. The quinoline, 2-bromonaphthalene, and 2-fluoronaphthalene moieties are electron-deficient aromatic groups because of the electron-deficient pyridine ring and electron-withdrawing fluorine and bromine atoms. It is suggested that when X represents bicyclic or electron-rich aromatic groups, the compounds are unlikely to be potent ligands.

Because some compounds containing bicyclic or electronrich aromatic groups at the group X position in Figure 2 do not have high binding affinity CXCR4, compounds in Figure 3 in which Y1 is a monocyclic and electron-deficient aromatic group were designed: 4pyridylmethyl, 2-pyridylmethyl, 4-fluorobenzyl, and 4-trifluoromethylbenzyl groups (Y¹-CH<sub>2</sub>) were introduced onto the nitrogen atom. In addition, as spacer templates (H<sub>2</sub>N-Y<sup>2</sup>-NH<sub>2</sub>) 1,4-phenylenedimethanamine, naphthalene-2,6-diyldimethanamine, and [1,1'-biphenyl]-4,4'-diyldimethanamine structures were introduced to refine the spacer struc-

tures, and quanidino and tetramethylguanidino groups were used as Y<sup>3</sup> substituents. The CXCR4 binding affinities of compounds 37a-42d were evaluated (Table 2). None of these compounds showed more than 50% inhibition at 10 µм. In general, 4-trifluoromethylbenzyl, [1,1'-biphenyl]-4,4'-diyldimethanamine, and tetramethylguanidino moieties seem to be more suitable as candidates for Y1-CH2, H2N-Y2-NH2, and Y3, respectively. Among these synthetic compounds, 40b, containing 2-pyridylmethyl, 1,4-phenylenedimethanamine and tetramethylguanidino groups, and 42d containing 4-trifluoromethylbenzyl, [1,1'-biphenyl]-4,4'-diyldimethanamine and tetramethylguanidino groups, have the highest binding affinity for CXCR4.

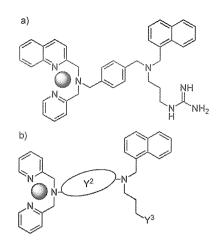
As described above in the Introduction, aza-macrocyclic compounds such as the Dpa-Zn complex 1[35] and the Dpacyclam compound 2<sup>(36)</sup> have high binding affinities toward CXCR4. The zinc complex of 2 also has a higher CXCR4 binding affinity. Thus, the CXCR4 binding affinities of the zinc complexes of 19a, containing 2-pyridylmethyl and 2-methylquino-

Scheme 2. Reagents and conditions: a) PPh<sub>3</sub>, DEAD, X-CH<sub>2</sub>OH, THF, RT, 97% (20 d), 59% (20 e); b) PhSH, K<sub>2</sub>CO<sub>3</sub>, DMF, RT, 42% (21 d), 64% (21 e); c) NaBH-(OAC)<sub>3</sub>, AcOH, 2-pyridinecarbaldehyde, 1,2-dichloroethane, RT, 78% (22 d), 85% (22 e); d) 4 M HCl/dioxane, RT; e) DIPEA, 1-amidinopyrazole·HCl, DMF, RT, 24% (23 d), 18% (23 e) (two steps).

Compd	$Y^{1[a]}$	Y <sup>2[b]</sup>	Y <sup>3[c]</sup>	Inhibition [%] <sup>[d]</sup>	Compd	Y <sup>1[a]</sup>	Y <sup>2[b]</sup>	$Y^{3[c]}$	Inhibition [%] <sup>[d</sup>
37 a	a	Α	i	9.6 ± 1.9	40 a	a	Α	ii	0
37 b	b	Α	i	$21.4 \pm 2.8$	40 b	b	Α	ii	$41.5 \pm 4.8$
37 c	c	Α	i	$8.5 \pm 1.8$	40 c	c	Α	ii	$12.7 \pm 4.0$
37 d	d	Α	i	$22.3\pm1.4$	40 d	d	Α	ii	$23.8\pm6.0$
38 a	a	В	i	0	41 a	a	В	ii	$\textbf{3.2} \pm \textbf{2.2}$
38 b	b	В	i	$4.7 \pm 1.3$	41 b	b	В	ii	$21.6 \pm 2.6$
38 c	c	В	i	$4.2 \pm 6.0$	41 c	c	В	ii	$13.2\pm1.5$
38 d	d	В	i	$4.1 \pm 4.1$	41 d	d	В	ii	$18.4 \pm 1.2$
39 a	a	C	i	$8.1\pm1.1$	42 a	a	C	ii	$8.8\pm1.0$
39 b	b	C	i	$18.0 \pm 1.1$	42 b	b	C	ii	0
39 c	c	C	i	$26.0\pm3.0$	42 c	c	C	ii	$26.6 \pm 4.4$
39 d	d	C	i	$27.9 \pm 5.2$	42 d	d	C	ii	$45.0 \pm 3.0$

[a–c] The structures of  $Y^1$ ,  $Y^2$ , and  $Y^3$  are shown in Figure 3 as a–d, A–C, and i–ii, respectively. [d] CXCR4 binding affinity was assessed based on the inhibition of [ $^{125}$ ]CXCL12 binding to Jurkat cells; percent inhibition values for all compounds at 10  $\mu$ m were calculated relative to that of FC131 (100%).

line groups, and **37 b**, **38 b**, **39 b**, **40 b**, **41 b**, and **42 b**, containing the Dpa group, were evaluated (Figure 4). ZnCl<sub>2</sub> (10 equiv relative to each compound) was added to phosphate-buffered saline (PBS) solutions of these compounds to form zinc(II) complexes. Chelation of the nitrogen atoms of **37 b** and **40 b** with the zinc(II) ion has been demonstrated by changes in NMR chemical shifts upon ZnCl<sub>2</sub> titration as zinc chelates as described in our previous studies. [35,36] The percent inhibition of the zinc complexes at 5 µm is listed in Table 3. A remarkable increase in CXCR4 binding affinity of all the zinc complexes



**Figure 4.** Zinc complexes of a) **19a** and b) **37b**, **38b**, **39b**, **40b**, **41b**, and **42b**. The shaded circle represents the position of the zinc cation in the chelate. The structures of  $Y^2$  and  $Y^3$  are shown in Figure 3 as A–C and i–ii, respectively.

except **39 b** is observed if the inhibitory activities of the zinc complexes at  $5 \,\mu\text{M}$  (Table 3) are compared with those of the corresponding metal-free compounds at  $10 \,\mu\text{M}$  (Tables 1 and 2). The high activity of the zinc complexes is consistent with results reported in our previous work, <sup>[35,36]</sup> and suggests that the formation of chelates of the nitrogen atoms in the compounds with the zinc(II) ion might enhance their interaction with CXCR4. Fixation of the functional moieties by zinc(II) che-

lation, progression of electron deficiency of the aromatic moieties, interaction of the zinc(II) ion with residues on CXCR4, etc., might be considered as reasons for the enhanced CXCR4 binding affinity of the zinc complexes. to previous reports,[39,40] in the case of chelation of the zinc complexes of AMD3100, a divalent metal ion such as zinc(II) in one of the bicyclam rings increased this compound's affinity for CXCR4 through a specific interaction with the carboxylate of Asp262 of CXCR4. A similar phenomenon could be occurring in the zinc complexes of the present compounds. The IC<sub>so</sub> values of the

zinc complexes of  $37\,b$  and  $40\,b$  containing 1,4-phenylenedimethanamine were evaluated to be 2.1  $\mu M$ . In comparing the CXCR4 binding affinity of the zinc complexes of  $37\,b$ ,  $38\,b$ ,  $39\,b$ ,  $40\,b$ ,  $41\,b$ , and  $42\,b$ , 1,4-phenylenedimethanamine is the most suitable spacer template  $(H_2N-Y^2-NH_2)$ , and naphthalene-2,6-diyldimethanamine is the second most effective. As substituents for  $Y^3$ , the tetramethylguanidino group is more appropriate than guanidine. The reason for this property has not been clarified yet; however, the tetramethyl group might stabilize a positively charged nitrogen atom, or might enhance a hy-

Scheme 3. Reagents and conditions: a) NsCl, Et<sub>3</sub>N, THF, 84%; b) Y¹-CH<sub>2</sub>OH, DEAD, PPh<sub>3</sub>, THF, 53% (26 a), 92% (26 b), 70% (26 c), 97% (26 d); c) PhSH,  $K_2CO_3$ , DMF, 97% (27 a), 74% (27 b), 91% (27 c), 91% (27 d); d) Kl,  $K_2CO_3$ , 11, MeCN, 78% (31), 53% (32), 71% (33); e) Kl,  $K_2CO_3$ , amine 27 a–d, MeCN, 25% (34 a), 78% (34 b), 80% (34 c), 90% (34 d), 38% (35 a), 75% (35 b), 67% (35 c), 55% (35 d), 23% (36 a), 59% (36 b), 80% (36 c), 80% (36 d); f) 4 M HCl/dioxane; g) DIPEA, 1-amidinopyrazole·HCl, DMF, 19% (37 a), 49% (37 b), 52% (37 c), 30% (37 d), 42% (38 a), 56% (38 b), 62% (38 c), 44% (38 d), 39% (39 a), 48% (39 b), 87% (39 c), 50% (39 d) (two steps); h) 4 M HCl/dioxane; j) DIPEA, 2-(1*H*-benzotriazole-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate, DMF, 24% (40 a), 36% (40 b), 31% (40 c), 32% (40 d), 31% (41 a), 14% (41 b), 47% (41 c), 27% (41 d), 37% (42 a), 25% (42 b), 27% (42 c), 44% (42 d) (two steps).

drophobic interaction with residues on CXCR4. Comparison of the CXCR4 binding affinity of the zinc complexes of **19a** and **37b** shows that the 2-pyridylmethyl group is more suitable than the 2-methylquinoline group as X-CH<sub>2</sub> or Y<sup>1</sup>-CH<sub>2</sub> introduced on the nitrogen atom.

## Conclusions

New low-molecular-weight CXCR4 ligands were designed synthesized. The most potent compounds are 37b and 40b, zinc complexes with a Dpa group on the 1,4-phenylenedimethanamine spacer template. The distances between all the functional moieties of the compounds linked by the 1,4-phenylenedimethanamine spacer might be appropriate for interaction with CXCR4. These compounds exhibited IC50 values at micromolar levels in CXCR4 binding affinity. Zinc complexation of Dpa-containing compounds resulted in a remarkable increase in CXCR4 binding affinity relative to the corresponding zinc-free compounds. The results reported herein might provide useful insight into the design of novel CXCR4 ligands, complementing information from other compounds such as T140, FC131, KRH-1636. These compounds will be useful for the development of future therapeutic strategies for CXCR4-relevant diseases.

## **Experimental Section**

#### Chemistry

Synthetic strategies of compounds reported in the present study are described in Results and Discussion above, and details are provided in the Supporting Information. Zn<sup>II</sup> complex formation was performed by treatment of the compounds with 10 equiv ZnCl<sub>2</sub> in PBS. The Zn<sup>II</sup> complexes were characterized by the chemical shifts of their methylene protons in <sup>1</sup>H NMR spectroscopic analysis. The DpaZn<sup>II</sup> complex was characterized

previously.[35] Detailed data are provided in the Supporting Information

7-1-1-	3 CVC	DA laims	1: £	Charles and			1		1 1 1 1 1 1 1 1 1 1	201	201
1able	3. LAL	R4 bind	mic ai	mmues	D: CO	moour	HUS I	3 d. :	M 10.	580.	530.
ann	ill in or	id 42b i	n zinci	III cami	1 DV						

Compd	Inhibition [%] <sup>[a]</sup>	IC <sub>50</sub> [пм] <sup>[b]</sup>
19a	34.5 ± 6.5	ND
37 b	$93.4 \pm 6.4$	2100
38b	$25.6 \pm 2.4$	ND
39 b	0	ND
40 b	$98.0 \pm 1.0$	2100
41 b	$80.7 \pm 0.8$	ND
42 b	$35.9 \pm 0.9$	ND
FC131 <sup>[c]</sup>	100	15.9

[a] CXCR4 binding affinity was assessed based on the inhibition of  $[^{125}]$ CXCL12 binding to Jurkat cells; percent inhibition values for all zinc complexes at 5  $\mu$ M were calculated relative to that of FC131 (100%). [b] IC<sub>50</sub>: zinc complex concentration required for 50% inhibition of  $[^{125}]$ CXCL12 binding to Jurkat cells; all data are the mean values from at least three independent experiments; ND: not determined. [c] Metal free.

#### Biological assays

CXCR4 binding assays of compounds based on the inhibition of [<sup>125</sup>I]CXCL12 binding to Jurkat cells were performed as reported by Tanaka et al.<sup>[38]</sup>

# **Acknowledgements**

T.T., C.H., and N.O. are supported by JSPS research fellowships for young scientists. This work was supported in part by a Grant-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science, and Technology of Japan, and Health and Labour Sciences Research Grants from the Japanese Ministry of Health, Labor, and Welfare.

**Keywords:** aza-macrocycles · chemokine receptors · CXCR4 · low-molecular-weight ligands · zinc complexes

- [1] T. Nagasawa, H. Kikutani, T. Kishimoto, Proc. Natl. Acad. Sci. USA 1994, 91, 2305–2309.
- [2] C. C. Bleul, M. Farzan, H. Choe, C. Parolin, I. Clark-Lewis, J. Sodroski, T. A. Springer, *Nature* 1996, 382, 829–833.
- [3] E. Oberlin, A. Amara, F. Bachelerie, C. Bessia, J. L. Virelizier, F. Arenzana-Seisdedos, O. Schwartz, J. M. Heard, I. Clark-Lewis, D. L. Legler, M. Loetscher, M. Baggiolini, B. Moser, *Nature* 1996, 382, 833 835.
- [4] K. Tashiro, H. Tada, R. Heilker, M. Shirozu, T. Nakano, T. Honjo, Science 1993, 261, 600–603.
- [5] J. Wang, L. He, C. A. Combs, G. Roderiquez, M. A. Norcross, Mol. Cancer Ther. 2006, 5, 2474 – 2483.
- [6] T. Tanaka, W. Nomura, T. Narumi, A. Masuda, H. Tamamura, J. Am. Chem. Soc. 2010, 132, 15899–15901.
- [7] C. C. Bleul, R. C. Fuhlbrigge, J. M. Casanovas, A. Aiuti, T. A. Springer, J. Exp. Med. 1996, 184, 1101 1109.
- [8] K. Tachibana, S. Hirota, H. Iizasa, H. Yoshida, K. Kawabata, Y. Kataoka, Y. Kitamura, K. Matsushima, N. Yoshida, S. Nishikawa, T. Kishimoto, T. Nagasawa. *Nature* 1998, 393, 591 594.
- [9] T. Nagasawa, S. Hirota, K. Tachibana, N. Takakura, S. Nishikawa, Y. Kitamura, N. Yoshida, H. Kikutani, T. Kishimoto, *Nature* 1996, 382, 635-638.
- [10] Y. Zhu, T. Yu, X.-C. Zhang, T. Nagasawa, J. Y. Wu, Y. Rao, *Nat. Neurosci.* 2002, 5, 719 – 720.
- [11] R. K. Stumm, C. Zhou, T. Ara, F. Lazarini, M. Dubois-Dalcq, T. Nagasawa, V. Hollt, S. Schulz, J. Neurosci. 2003, 23, 5123 – 5130.

- [12] T. Koshiba, R. Hosotani, Y. Miyamoto, J. Ida, S. Tsuji, S. Nakajima, M. Kawaguchi, H. Kobayashi, R. Doi, T. Hori, N. Fujii, M. Imamura, Clin. Cancer Res. 2000, 6, 3530–3535.
- [13] A. Müller, B. Homey, H. Soto, N. Ge, D. Catron, M. E. Buchanan, T. McClanahan, E. Murphy, W. Yuan, S. N. Wagner, J. L. Barrera, A. Mohar, E. Verastegui, A. Zlotnik, *Nature* 2001, 410, 50–56.
- [14] H. Tamamura, A. Hori, N. Kanzaki, K. Hiramatsu, M. Mizumoto, H. Naka-shima, N. Yamamoto, A. Otaka, N. Fujii, FEBS Lett. 2003, 550, 79 83.
- [15] N. Tsukada, J. A. Burger, N. J. Zvaifler, T. J. Kipps, *Blood* 2002, 99, 1030 1037
- [16] J. Juarez, K. F. Bradstock, D. J. Gottlieb, L. J. Bendall, *Leukemia* 2003, 17, 1294 1300.
- [17] H. K. Deng, R. Liu, W. Ellmeier, S. Choe, D. Unutmaz, M. Burkhart, P. D. Marzio, S. Marmon, R. E. Sutton, C. M. Hill, C. B. Davis, S. C. Peiper, T. J. Schall, D. R. Littman, N. R. Landau, *Nature* 1996, 381, 661–666.
- [18] Y. Feng, C. C. Broder, P. E. Kennedy, E. A. Berger, Science 1996, 272, 872–877.
- [19] T. Nanki, K. Hayashida, H. S. El-Gabalawy, S. Suson, K. Shi, H. J. Girschick, S. Yayuz, P. E. Lipsky, J. Immunol. 2000, 165, 6590 – 6598.
- [20] H. Tamamura, M. Fujisawa, K. Hiramatsu, M. Mizumoto, H. Nakashima, N. Yamamoto, A. Otaka, N. Fujii, FEBS Lett. 2004, 569, 99 – 104.
- [21] T. Murakami, T. Nakajima, Y. Koyanagi, K. Tachibana, N. Fujii, H. Tamamura, N. Toshida, M. Waki, A. Matsumoto, O. Yoshie, T. Kishimoto, N. Yamamoto, T. Nagasawa, J. Exp. Med. 1997, 186, 1389 1393.
- [22] H. Tamamura, Y. Xu, T. Hattori, X. Zhang, R. Arakaki, K. Kanbara, A. Omagari, A. Otaka, T. Ibuka, N. Yamamoto, H. Nakashima, N. Fujii, *Biochem. Biophys. Res. Commun.* 1998, 253, 877 882.
- [23] H. Tamamura, A. Omagari, S. Oishi, T. Kanamoto, N. Yamamoto, S. C. Peiper, H. Nakashima, A. Otaka, N. Fujii, *Bioorg. Med. Chem. Lett.* 2000, 10, 2633–2637.
- [24] N. Fujii, S. Oishi, K. Hiramatsu, T. Araki, S. Ueda, H. Tamamura, A. Otaka, S. Kusano, S. Terakubo, H. Nakashima, J. A. Broach, J. O. Trent, Z. Wang, S. C. Peiper, *Angew. Chem.* 2003, 115, 3373–3375; *Angew. Chem. Int. Ed.* 2003, 42, 3251–3253.
- [25] H. Tamamura, K. Hiramatsu, S. Ueda, Z. Wang, S. Kusano, S. Terakubo, J. O. Trent, S. C. Peiper, N. Yamamoto, H. Nakashima, A. Otaka, N. Fujii, J. Med. Chem. 2005, 48, 380–391.
- [26] H. Tamamura, T. Araki, S. Ueda, Z. Wang, S. Oishi, A. Esaka, J. O. Trent, H. Nakashima, N. Yamamoto, S. C. Peiper, A. Otaka, N. Fujii, J. Med. Chem. 2005, 48, 3280 3289.
- [27] C. Hashimoto, T. Tanaka, T. Narumi, W. Nomura, H. Tamamura, Expert Opin. Drug Discovery 2011, 6, 1067 – 1090.
- [28] H. Tamamura, H. Tsutsumi, H. Masuno, S. Mizokami, K. Hiramatsu, Z. Wang, J. O. Trent, H. Nakashima, N. Yamamoto, S. C. Peiper, N. Fujii, Org. Biomol. Chem. 2006, 4, 2354–2357.
- [29] K. Ichiyama, S. Yokoyama-Kumakura, Y. Tanaka, R. Tanaka, K. Hirose, K. Bannai, T. Edamatsu, M. Yanaka, Y. Niitani, N. Miyano-Kurosaki, H. Takaku, Y. Koyanagi, N. Yamamoto, *Proc. Natl. Acad. Sci. USA* 2003, 100, 4185 4190.
- [30] E. De Clercq, N. Yamamoto, R. Pauwels, J. Balzarini, M. Witvrouw, K. De Vreese, Z. Debyser, B. Rosenwirth, P. Peichl, R. Datema, Antimicrob. Agents Chemother. 1994, 38, 668–674.
- [31] D. Schols, S. Struyf, J. Van Damme, J. A. Esté, G. Henson, E. De Clercq, J. Exp. Med. 1997, 186, 1383 1388.
- [32] S. Hatse, K. Princen, E. De Clercq, M. M. Rosenkilde, T. W. Schwartz, P. E. Hernandez-Abad, R. T. Skerlj, G. J. Bridger, D. Schols, *Biochem. Pharma-col.* 2005, 70, 752 – 761.
- [33] Z. Liang, W. Zhan, A. Zhu, Y. Yoon, S. Lin, M. Sasaki, J. M. A. Klapproth, H. Yang, H. E. Grossniklaus, J. Xu, M. Rojas, R. J. Voll, M. M. Goodman, R. F. Arrendale, J. Liu, C. C. Yun, J. P. Snyder, D. C. Liotta, H. Shim, *PLoS One* 2012, 7, e34038.
- [34] S. Pettersson, V. I. Pérez-Nueno, L. Ros-Blanco, R. Puig de La Bellacasa, M. O. Rabal, X. Batllori, B. Clotet, I. Clotet-Codina, M. Armand-Ugón, J. A. Esté, J. I. Borrell, J. Teixidó, ChemMedChem 2008, 3, 1549 – 1557.
- [35] H. Tamamura, A. Ojida, T. Ogawa, H. Tsutsumi, H. Masuno, H. Nakashima, N. Yamamoto, I. Hamachi, N. Fujii, J. Med. Chem. 2006, 49, 3412 – 3415.
- [36] T. Tanaka, T. Narumi, T. Ozaki, A. Sohma, N. Ohashi, C. Hashimoto, K. Itotani, W. Nomura, T. Murakami, N. Yamamoto, H. Tamamura, ChemMed-Chem 2011, 6, 834–839.

- [37] T. Narumi, T. Tanaka, C. Hashimoto, W. Nomura, H. Aikawa, A. Sohma, K. Itotani, M. Kawamata, T. Murakami, N. Yamamoto, H. Tamamura, *Bioorg. Med. Chem. Lett.* 2012, 22, 4169–4172.
- [38] T. Tanaka, H. Tsutsumi, W. Nomura, Y. Tanabe, N. Ohashi, A. Esaka, C. Ochiai, J. Sato, K. Itotani, T. Murakami, K. Ohba, N. Yamamoto, N. Fujii, H. Tamamura, Org. Biomol. Chem. 2008, 6, 4374 4377.
- [39] L. Ole Gerlach, J. S. Jakobsen, K. P. Jensen, M. R. Rosenkilde, R. T. Skerlj, U. Ryde, G. J. Bridger, T. W. Schwartz, *Biochemistry* 2003, 42, 710-717.

[40] M. M. Rosenkilde, L.-O. Gerlach, J. S. Jakobsen, R. T. Skerlj, G. J. Bridger, T. W. Schwartz, J. Biol. Chem. 2004, 279, 3033 – 3041.

Received: August 24, 2012 Published online on October 19, 2012

