

始と感受性試験結果が得られるまでにタイムラグがあるため、薬剤耐性遺伝子を検査することで迅速に感受性を判定することは、適切な薬剤を選択するうえ、ならびに迅速な感染対策を行ううえで重要な情報となりうる。

喀痰塗抹検査にて陽性であった場合に、結核症を念頭におき隔離入院が行われる。抗酸菌排菌NTM症患者に対する念のための感染対策上の結核隔離入院は、患者個人の貴重な時間を束縛するのみならず医療経済上も大きな支出である。このNTM/MDR-TBストリップにより迅速に結核症を否定し非結核性抗酸菌症と診断することで、不必要な結核隔離入院を回避でき、医療経済上、患者の人権上において恩恵を得られる可能性が十分に考えられる。

### 謝 辞

喀痰検体の保存ならびに薬剤耐性遺伝子の検出は厚生労働科学研究費補助金（グラント番号：H21-新興-一般-008およびH24-新興-一般010）の一部で実施した。

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Original Article

CLINICAL EVALUATION OF A LINE PROBE ASSAY KIT FOR THE IDENTIFICATION OF *MYCOBACTERIUM* SPECIES AND DETECTION OF DRUG-RESISTANT *MYCOBACTERIUM TUBERCULOSIS*

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**Abstract** [Background] Multidrug resistance (MDR) involves resistance to both isoniazid and rifampicin, which makes the treatment of tuberculosis very difficult. Extensive drug resistance (XDR) occurs when, in addition to isoniazid and rifampicin resistance, the microorganisms are resistant to a fluoroquinolone and an injectable agent (e.g., kanamycin, amikacin, or capreomycin).

Generally, drug susceptibility testing takes more than 3–4 weeks after the initial cultivation. There is an urgent need to identify methods that can rapidly detect both the presence of *Mycobacterium tuberculosis* and the status of drug resistance.

[Purpose] This study was aimed at evaluating the line probe assay (LiPA; Nipro Co.), for the identification of *Mycobacterium* species and detection of mutations associated with anti-tuberculous drugs.

[Results] We found that LiPA enabled the rapid identification of *M. tuberculosis*, *M. avium*, *M. intracellulare*, and *M. kansasii*. When the results of the LiPA and conventional drug susceptibility tests were compared, there was no difference in the susceptibility to rifampicin, pyrazinamide, and levofloxacin; however, there was a difference in the susceptibility to isoniazid.

[Conclusion] Thus, LiPA can be used for the rapid identification of *Mycobacterium* species and the determination of susceptibility to drugs, which can help in the early initiation of appropriate treatment, leading to a reduction in infectiousness.

**Key words:** Multidrug-resistant *Mycobacterium tuberculosis*, Rapid diagnosis, Line probe assay

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# Methylation of 23S rRNA Nucleotide G748 by RlmA<sup>II</sup> Methyltransferase Renders *Streptococcus pneumoniae* Telithromycin Susceptible

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Several posttranscriptional modifications of bacterial rRNAs are important in determining antibiotic resistance or sensitivity. In all Gram-positive bacteria, dimethylation of nucleotide A2058, located in domain V of 23S rRNA, by the dimethyltransferase Erm(B) results in low susceptibility and resistance to telithromycin (TEL). However, this is insufficient to produce high-level resistance to TEL in *Streptococcus pneumoniae*. Inactivation of the methyltransferase RlmA<sup>II</sup>, which methylates the N-1 position of nucleotide G748, located in hairpin 35 of domain II of 23S rRNA, results in increased resistance to TEL in erm(B)-carrying *S. pneumoniae*. Sixteen TEL-resistant mutants (MICs, 16 to 32  $\mu\text{g/ml}$ ) were obtained from a clinically isolated *S. pneumoniae* strain showing low TEL susceptibility (MIC, 2  $\mu\text{g/ml}$ ), with mutation resulting in constitutive dimethylation of A2058 because of nucleotide differences in the regulatory region of erm(B) mRNA. Primer extension analysis showed that the degree of methylation at G748 in all TEL-resistant mutants was significantly reduced by a mutation in the gene encoding RlmA<sup>II</sup> to create a stop codon or change an amino acid residue. Furthermore, RNA footprinting with dimethyl sulfate and a molecular modeling study suggested that methylation of G748 may contribute to the stable interaction of TEL with domain II of 23S rRNA, even after dimethylation of A2058 by Erm(B). This novel finding shows that methylation of G748 by RlmA<sup>II</sup> renders *S. pneumoniae* TEL susceptible.

In prokaryotic cells, rRNAs and ribosomal proteins undergo specific posttranscriptional modifications by a wide variety of enzymes during maturation (1). Although the functions of most of these modifications are currently unknown, many are believed to modulate rRNA maturation and to affect its stability. In *Escherichia coli*, the 23S rRNA contains 25 known modifications, of which 14 are methylations, 9 are pseudouridines, 1 is a methylated pseudouridine, and 1 is unknown (1, 2). Approximately one-third of modified residues of the 23S rRNA are clustered around the nascent peptide exit tunnel (NPET), which starts at the peptidyltransferase center (PTC) and spans the body of the large ribosomal subunit (3). Macrolide antibiotics bind in the NPET, close to the PTC, hindering the passage of the newly synthesized polypeptides through the NPET (3).

However, some modifications play important roles in determining antibiotic resistance or sensitivity (4). Among the proteins involved, the Erm family of proteins are known as methyltransferases that methylate the N-6 position of nucleotide A2058 in 23S rRNA domain V (5). Methylation of A2058, which forms an interaction site for the MLS<sub>B</sub> (macrolide, lincosamide, and streptogramin B) antibiotics, results in MLS<sub>B</sub> resistance (6). Erm methyltransferases have been classified into 2 types according to whether they add one or two methyl groups to this nucleotide position. Dimethylation confers the MLS<sub>B</sub> type II phenotype, with high resistance to all MLS<sub>B</sub> antibiotics, whereas monomethylation confers the MLS<sub>B</sub> type I phenotype, with high resistance to lincosamides and low to moderate resistance to macrolides and streptogramin B antibiotics (4, 6). Production of Erm dimethyltransferases in many pathogenic bacteria is the most common mechanism of macrolide resistance.

Telithromycin (TEL) is a semisynthetic derivative of the 14-member ring macrolide erythromycin (ERY), which was the first ketolide approved for clinical use. Its high efficacy has been dem-

onstrated against *Streptococcus pneumoniae* isolates that cause community-acquired respiratory tract disease (7–9). TEL binds more strongly to ribosomes than ERY does, largely because of the alkyl-aryl substituent extending from positions 11 and 12 of the macrolactone ring (10, 11). The alkyl-aryl arm stacks on nucleotides A752 and U2609 to form a base pair that bridges domains II and V in 23S rRNA (12). Chemical probing and primer extension analyses have shown that nucleotide A752 is also involved in TEL binding in *E. coli* and *Staphylococcus aureus* (12). This may allow TEL to overcome common macrolide resistance mechanisms, including target modifications directed by Erm monomethyltransferases and mutations in the 23S rRNA and ribosomal proteins that interrupt macrolide binding (13, 14). However, the production of Erm dimethyltransferases causes TEL resistance in some Gram-positive bacteria, e.g., *Streptococcus pyogenes* (15).

*S. pneumoniae* resistance to macrolides has increased worldwide. PROTEKT year 5 data (2003 to 2004) show that the most common resistance mechanism globally has been the erm(B)-encoded Erm dimethyltransferase, which is present in 55% of macrolide-resistant isolates (16). Furthermore, the PneumoWorld study in Europe and the Asian Network for Surveillance of Resistant Pathogens (ANSRP) have shown that among erythromycin-resistant *S. pneumoniae* isolates, ~80% carry the erm(B) gene (17, 18). Although high-level TEL resistance can easily be created from

Received 24 January 2013 Returned for modification 15 February 2013

Accepted 20 May 2013

Published ahead of print 28 May 2013

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doi:10.1128/AAC.00164-13

TABLE 1 Bacterial strains used in this study

| Strain                                  | Relevant characteristics  | Reference or source                 |
|---|---|-------------------------------------|
| <i>Escherichia coli</i> strains         |   |                                     |
| DH5 $\alpha$                            | F <sup>-</sup> <i>recA endA gyrA thi hsdR supE relA</i> $\Delta$ ( <i>lacZYA-argF</i> ) <i>deoR</i> $\phi$ 80d <i>lacZ</i> $\Delta$ M15 | Laboratory collection               |
| DH5 $\alpha$ Z1                         | Same as DH5 $\alpha$ , but <i>tetR lacI<sup>q</sup> Spec<sup>r</sup></i>  | Laboratory collection               |
| <i>Streptococcus pneumoniae</i> strains |   |                                     |
| TIGR4                                   | Prototroph  | ATCC                                |
| S1                                      | TEL-resistant clinical isolate  | Miroku Medical Laboratory Co., Ltd. |
| Sp32 to Sp41                            | TEL-resistant mutants of S1 isolated from 8- $\mu$ g/ml TEL-containing BHI-Y agar plates  | This study                          |
| Sp42 to Sp47                            | TEL-resistant mutants of S1 isolated from 16- $\mu$ g/ml TEL-containing BHI-Y agar plates   | This study                          |
| Sp182                                   | S1 <i>erm(B)::aad(9)</i>  | This study                          |
| Sp274                                   | S1 <i>tlrB::aad(9)</i>  | This study                          |
| Sp303                                   | TIGR4 <i>tlrB::aad(9)</i>   | This study                          |

an *erm(B)*-carrying strain in a laboratory (19), many *erm(B)*-carrying strains of *S. pneumoniae* remain clinically sensitive to TEL (15). A few TEL-resistant strains of *erm(B)*-carrying *S. pneumoniae* have been isolated (20, 21). A large deletion upstream of the *erm(B)* gene resulted in constitutively high dimethylation at A2058, leading to TEL resistance (MICs, 8 to 16  $\mu$ g/ml) (21). On the other hand, a clinical isolate of *S. pneumoniae* that is highly resistant to TEL (MIC, >256  $\mu$ g/ml) carries an *erm(B)* gene encoding a truncated leader peptide and has mutations in the ribosomal protein L4 (20). Therefore, it remains unclear which determinant is essential for the high-level TEL resistance in *erm(B)*-carrying *S. pneumoniae*.

We have investigated the strong antibacterial activity of TEL that is effective even against *erm(B)*-carrying *S. pneumoniae*. TEL-resistant mutants were initially generated *in vitro* from a clinically isolated strain showing a low TEL susceptibility by introducing mutations into the regulatory region of *erm(B)* mRNA, and the whole-genome sequences of mutants were analyzed by next-generation sequencing. We thereby identified a mutation in the gene encoding RlmA<sup>II</sup>, which methylates position N-1 of nucleotide G748 in 23S rRNA domain II (22). Further analyses showed that methylation of G748 by RlmA<sup>II</sup> renders *erm(B)*-carrying *S. pneumoniae* TEL susceptible.

## MATERIALS AND METHODS

**Bacterial strains, plasmids, and media.** Bacterial strains and plasmids used in this study are shown in Tables 1 and 2, respectively. *S. pneumoniae* strain S1 revealing reduced TEL susceptibility (MIC, 2  $\mu$ g/ml) was clinically isolated in Japan in 2010 and provided by Miroku Medical Laboratory Co., Ltd. (Nagano, Japan). Pneumococci were routinely cultured at 37°C and 5% CO<sub>2</sub> in brain heart infusion plus 0.5% yeast extract (BHI-Y) broth and BHI-Y agar supplemented with 5% horse blood. *E. coli* was grown in L broth (1% Bacto tryptone, 0.5% Bacto yeast extract, 0.5%

TABLE 2 Plasmids used in this study

| Plasmid   | Relevant characteristics   | Reference or source   |
|-----------|--|-----------------------|
| pUC18     | Cloning vector   | Laboratory collection |
| pT7Blue   | Cloning vector   | Novagen               |
| pLZ12-Km2 | Shuttle vector   | 55                    |
| pTKY858   | pT7Blue with 618-bp <i>erm(B)</i> fragment   | 31                    |
| pTKY862   | pLZ12-Km2 with <i>aad(9)</i> Sp resistance cassette  | 31                    |
| pTKY1041  | pLZ12-Km2 with 1,401-bp <i>erm(B)</i> fragment from S1   | This study            |
| pTKY1042  | pT7Blue with disrupted <i>erm(B)</i> fragment by insertion of <i>aad(9)</i> Sp resistance cassette             | This study            |
| pTKY1109  | pUC18 with 328-bp <i>tlrB</i> fragment   | This study            |
| pTKY1110  | pUC18 with disrupted <i>rlmA<sup>II</sup></i> fragment by insertion of an <i>aad(9)</i> Sp resistance cassette | This study            |
| pTKY1111  | pLZ12-Km2 with 1,065-bp <i>tlrB</i> fragment from S1   | This study            |

sodium chloride, pH 7.4) and L agar. When necessary, the medium was supplemented with kanamycin (25 to 500  $\mu$ g/ml), tetracycline (10  $\mu$ g/ml), erythromycin (25  $\mu$ g/ml), spectinomycin (100  $\mu$ g/ml), and ampicillin (25  $\mu$ g/ml). For inducible conditions for *erm(B)*, 25  $\mu$ g/ml ERY was added during growth.

**Antimicrobial susceptibility testing.** Susceptibility to antibiotics was determined by the serial 2-fold dilution method, using Mueller-Hinton agar plates supplemented with 5% lysed horse blood. Susceptibility or resistance of pneumococci to TEL and ERY was assessed in accordance with the recommendations of the Clinical and Laboratory Standards Institute (23).

**Transformation.** Synthetic competence-stimulating peptide (CSP) 1 or 2 and the method of Iannelli and Pozzi (24) were used to transform *S. pneumoniae* S1 or TIGR4, respectively, into transformation-competent states.

**Construction of *erm(B)* and *tlrB* disruption mutants.** Disruption of *erm(B)* was performed as follows. The pTKY858 plasmid, carrying part of the *erm(B)* gene, was cleaved with StyI. The overhanging ends were blunted with T4 polymerase and ligated to a fragment containing the spectinomycin resistance gene (*Sp*), which was generated from pTKY862 after digestion with BamHI followed by blunting with T4 DNA polymerase. The resulting plasmid, pTKY1042, was used to replace  $\Delta$ *erm(B)::Sp* in *S. pneumoniae* strain S1.

Disruption of the *tlrB* gene, which encodes RlmA<sup>II</sup>, was performed as follows. A fragment including part of the *tlrB* gene was amplified from chromosomal DNA prepared from *S. pneumoniae* strain S1 by a PCR using the forward primer 5'-CTAGAATTCAAATCTCAAGCCCAACTTC-3' and the reverse primer 5'-CCAATGCATGCCAGTTGGGTTCA C-3'. The PCR product was digested with EcoRI-SphI, and the fragment was cloned into pUC18. The resultant plasmid, pTKY1109, was cleaved with HincII and ligated to the *Sp* fragment. The resulting plasmid, pTKY1110, was used to replace  $\Delta$ *tlrB::Sp* in *S. pneumoniae*. Double-cross-over events in all constructed mutants were assessed by PCR and Southern hybridization.

**Construction of plasmids.** To construct the pTKY1041 plasmid, carrying *erm(B)*, the gene was amplified from the chromosome of S1 by using the primers *erm(B)*LP3-EcoRI-F (5'-CGGAATCCTTGCTTCGTTG-3') and *erm(B)*-R5-BamHI (5'-CGCGGATCCTTAGCTCCTTGGA-3'). The generated fragment was cleaved with EcoRI at the 5' end and BamHI at the 3' end and then cloned into the pLZ12-Km2 vector. To construct the pTKY1111 plasmid, carrying *tlrB*, the *tlrB* gene was amplified from the chromosome of S1 by using the primers *rlmA<sup>II</sup>*-XhoI-F

(5'-CTGTACTCGAGTACGGCAAGGCGACG-3') and *rlmA*<sup>II</sup>-ApaI-R (5'-GGTTTGGGGCCCTGTTCTTATGCGTTTG-3'). The generated fragment was cleaved with XhoI at the 5' end and ApaI at the 3' end and then cloned into the pLZ12-Km2 vector.

**Next-generation sequencing and data analysis.** Relative mutations in the bacterial genome were estimated by sequencing the genomic DNA samples extracted from each strain. Total DNA was extracted by the method of Blue and Mitchell (25). For preparation of the library DNA, 500 ng of each total DNA was sheared to 800 bp by Covaris (M&S). The sheared DNA products were purified using a MiniElute PCR purification kit (Qiagen) and then were used as templates for pyrosequencing with a GS Junior platform (Roche). The data were treated by a data analysis pipeline. Both reads from S1 and Sp36 were assembled into contigs by the *de novo* assembler. The nucleotide sequence polymorphisms among these strains were listed by mapping sequence reads on the assembled contigs with a reference mapper program. We collected nucleotide substitutions meeting the following 2 conditions: (i) the substitution was supported by >5 sequencing reads, and (ii) >90% of the reads supported the substitution. To find substitutions affecting protein function, we predicted open reading frames (ORFs) on the assembled contigs by using the RAST Web server, version 3.0 (<http://rast.nmpdr.org/>), and annotated the relationship between substitutions and ORFs. Correspondence between substitutions and ORFs was assigned by a custom Perl script implemented with Bioperl 1.6 (26).

**DMS modification of ribosomes *in vivo*.** Dimethyl sulfate (DMS) modification of ribosomes *in vivo* was based on a previously published protocol (27, 28). Bacterial cells of strain S1 were grown in BHI-Y broth to the exponential growth phase. TEL (25  $\mu$ M) was added to one aliquot of each culture for 1 h, followed by collection of the cells by centrifugation. Each pellet was resuspended in 80  $\mu$ l DMS buffer (50 mM sodium cacodylate, pH 7.0, 1 mM EDTA), with or without TEL. DMS solution was added (final concentration, ~50 mM) to each culture, and the cultures were incubated for 45 min. Reactions were stopped by the addition of 20  $\mu$ l ice-cold stop buffer (1.5 M sodium acetate, pH 7.0, 1 M  $\beta$ -mercaptoethanol). Cells were harvested at 4°C for extraction of total RNAs and primer extension as described below.

**Primer extension to detect methylated G748 and dimethylated A2058 in 23S rRNA.** The degree of methylation of each RNA was assayed by a primer extension method initially described by Morgan and colleagues (29). Briefly, bacterial cultures were grown in BHI-Y broth to the exponential growth phase. Total RNAs were extracted from bacterial cultures by use of RNeasy minikits (Qiagen). Two micrograms of total RNA, a 400 nM concentration of an oligonucleotide primer containing 5'-linked fluorescein isothiocyanate (FITC), and 500  $\mu$ M (each) dATP, dCTP, dTTP, and ddGTP for G748 or dGTP, dCTP, dTTP, and ddATP for A2058 were mixed, heated to 95°C for 1 min, and cooled to allow annealing at 53°C. A mixture containing SuperScript III reverse transcriptase (Invitrogen) was added, and the reaction was continued at 53°C for an additional 20 min. The primers were complementary to 23S rRNA bases 756 to 790 (5'-TTTGGAAATTTCTCCGCTACCCACAAGTCATCCAAG-3') for m<sup>1</sup>G748 and bases 2066 to 2101 (5'-CAGACACTCAATATCAAACTGACAGTAAAGCTCCATG-3') for m<sup>2</sup>A2058. Reactions were terminated by the addition of Novex Tris-borate-EDTA (TBE)-urea sample buffer (Invitrogen) after incubation at 70°C for 15 min. Extension products were resolved in a 10% TBE-urea gel and visualized using a TyphoonFLA9000 photoimager (GE Healthcare).

**Molecular modeling and molecular mechanics.** The three-dimensional (3-D) crystal structure of the 50S subunit of the 70S ribosome with TEL bound from *E. coli* (Protein Data Bank entry 3OAT) was used as a template for the initial structure. The methylated form of G748 was generated by the Leap module of the AMBER 11.0 program package. Each model was placed in a rectangular box filled with ~775,000 TIP3P water molecules. The molecular mechanics calculations for methylated and unmethylated forms of G748 in 23S rRNA were carried out using the AMBER 11.0 program package, together with the AMBER parm10 force

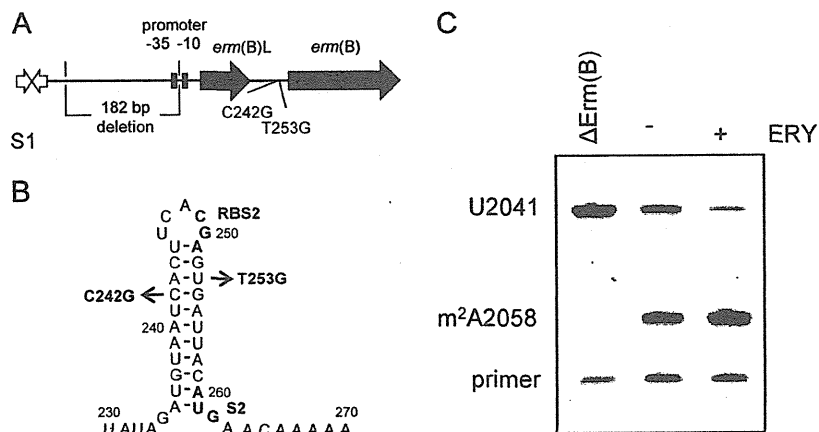
field and all\_modrna08 force field. Partial charges for the TEL atoms were calculated using the quantum chemical calculation in GAUSSIAN 03 (Gaussian, Inc., Carnegie, PA) followed by reduced electrostatic potential fitting, implemented in the antechamber module of AMBER. The 2 systems were energetically minimized for 10,000 steps (5,000 steps of the steepest-descent method followed by 5,000 steps of the conjugate gradient method). A cutoff distance of 10 Å for nonbonded interactions was used. The  $\pi$ -CH interaction energy was estimated by the formula proposed by Yuki et al. (30).

## RESULTS

**Mutations in the regulatory region of *erm*(B) mRNA result in highly increased dimethylation of A2058 in 23S rRNA in an *S. pneumoniae* clinical isolate.** The *S. pneumoniae* isolate S1, which has reduced TEL susceptibility (MIC, 2  $\mu$ g/ml), has mutations (A138G, T389C, C724T, and A1745T) in the 4 alleles encoding 23S rRNA and a mutation (S20N) in ribosomal protein L4, which have often been detected in many macrolide-resistant *S. pneumoniae* strains (31–33). Strain S1 also has an *erm*(B) gene in the Tn3872-like structure (Fig. 1A). Tn3872 is a Tn916-like conjugative transposon with an *erm*(B) gene, which is carried in a Tn917-like element (34). Tn917 carries an *erm*(B) gene (accession no. M11180) but does not mediate resistance to other antimicrobial agents, such as tetracycline (35). A 5' leader sequence of 259 nucleotides that has the potential to encode a leader peptide [*erm*(B)L] of 27 amino acids is the regulatory region of *erm*(B) mRNA involved in mRNA stabilization and translational attenuation, through the formation of alternative mRNA structures (36). Since mutation in the regulatory region of *erm*(B) results in TEL resistance (21, 36), we sequenced the region carrying *erm*(B). Two nucleotide differences [C242G and T253G; nucleotide numbers in *erm*(B) mRNA (36)] were found (Fig. 1A and B). A 182-bp deletion in the region upstream of *erm*(B) [positions –87 to –269 relative to the ATG start codon of *erm*(B)L] that included the –35 region of the putative promoter was found (Fig. 1A).

To examine the contribution of the *erm*(B) region to the reduced TEL sensitivity of strain S1, we constructed an *erm*(B)-disrupted mutant by inserting the spectinomycin resistance gene *aad*(9) into *erm*(B) of strain S1 and determined the TEL MIC of the resultant mutant, Sp182. Disruption of *erm*(B) resulted in a significant decrease in the TEL MIC, to the level for strain ATCC 49619 (<0.03  $\mu$ g/ml), which was used as a standard drug-susceptible strain. TEL sensitivity was complemented when the *erm*(B) gene was provided by plasmid pTKY1041. To define the essential nature of *erm*(B) for the low TEL susceptibility of strain S1, the laboratory strain TIGR4, which is susceptible to all antibiotics, was also transformed with S1 *erm*(B). Susceptibility testing of the transformants showed that TIGR4 derivatives with pTKY1041 had low TEL sensitivity (MIC, 1  $\mu$ g/ml), suggesting that an *erm*(B) gene with nucleotide alterations (C247G and T253G) and a 182-bp deletion determines the low TEL susceptibility of strain S1.

The increased level of A2058 dimethylated by Erm in 23S rRNA is related to an increase of TEL resistance (37, 38). The methyltransferase encoded by *erm*(B) is constitutively or inducibly synthesized (6). It is necessary to constitutively synthesize Erm(B) for TEL resistance, because TEL does not induce the synthesis of methyltransferases (39). To show whether A2058 in the 23S rRNA of strain S1 is constitutively dimethylated by Erm(B), the levels of dimethylation at A2058 in strain S1 cultured with and without ERY, which acts as an inducer of *erm*(B) (36), were assayed by a primer extension assay. Dimethylation at A2058 arrests the prog-



**FIG 1** Effects of mutations in the *erm(B)* region on dimethylation at A2058 in 23S rRNA. (A) Diagram representing the *erm(B)* region in Tn3872 and the mutant *erm(B)* region of S1. (B) Secondary structure of translationally inactive form of the regulatory region of *erm(B)* mRNA and mutations detected in the parental strain S1. RBS2, ribosome-binding site; S2, start codon of the methyltransferase structural gene. (C) Levels of dimethylation at A2058 in 23S rRNAs extracted from strain S1 cultured with and without ERY. Dimethylated A2058 was detected by reverse transcriptase extension from bases 2066 to 2101. Dimethylation at A2058 aborts the extension.

ress of reverse transcription at position 2059, whereas transcription on unmethylated or monomethylated A2058 proceeds until it is terminated by incorporation of ddATP at U2041. A large percentage (71.0% ± 8.4%) of the A2058 in strain S1 was dimethylated even in the absence of ERY (Fig. 1C), suggesting that there was constitutive expression of *erm(B)*. However, the *erm(B)* gene seems to continue to be inducible, because ERY exposure increased the level of dimethylation to 76.7% ± 9.7%. This slight increase in dimethylation by ERY exposure did not increase the TEL MIC (2 μg/ml). In strain TIGR4 harboring the pTKY1041 plasmid, the level of constitutive dimethylation at A2058 was also similar to that in strain S1 (data not shown). These results suggest that the highly increased dimethylation at A2058 in 23S rRNA due to mutations in the regulatory region of *erm(B)* reduces TEL sensitivity in *S. pneumoniae* strain S1.

**Isolation of TEL-resistant mutants of clinical isolate S1.** TEL-resistant mutants were generated from strain S1. A total of 16

mutants were isolated from strain S1 (MIC, 2 μg/ml) by selection with 8 μg/ml (10 mutants; Sp32 to Sp41) and 16 μg/ml (6 mutants; Sp42 to Sp47) of TEL. All the mutants had increased MICs (16 to 32 μg/ml) (Table 3). ERY exposure did not increase the TEL MIC of any of the mutants. To examine whether the increased resistance to TEL was stable, the mutants were subcultured in drug-free BHI-Y broth at 37°C and 5% CO<sub>2</sub> in air for 10 generations. The mutants were stable, such that subculturing them without the selective pressure of TEL did not generate derivatives with decreased TEL MICs. The primer extension assay showed that the level of dimethylated A2058 in each TEL-resistant mutant was similar to that in strain S1 under conditions both with and without ERY (Table 3), suggesting that a mutation unrelated to *erm(B)* function results in high-level TEL resistance.

To determine mutations involved in increased TEL resistance, whole-genome sequencing of strains S1 and Sp36 was performed by next-generation sequencing technology, and the sequences of

**TABLE 3** Mutations in RlmA<sup>II</sup> proteins of TEL-resistant *S. pneumoniae* strains

| Mutation in RlmA <sup>II</sup>                          | Strain           | MIC <sup>a</sup> (μg/ml) |      | Level (%) of dimethylation at A2058 <sup>b</sup> |             |
|---|------------------|--------------------------|------|--|-------------|
|   |                  | -ERY                     | +ERY | -ERY   | +ERY        |
| Wild type   | S1               | 2                        | 2    | 71.0 ± 8.4                                       | 76.7 ± 9.7  |
| Cys23Arg (67tGT <sub>69</sub> → cGT)                    | Sp44             | 32                       | 32   | 87.5 ± 5.6                                       | 81.9 ± 13.2 |
| Cys36Arg (106tGC <sub>108</sub> → cGC)                  | Sp47             | 32                       | 32   | 77.3 ± 2.2                                       | 66.7 ± 29.7 |
| Leu113Pro (337CtA <sub>339</sub> → CcA)                 | Sp42, Sp43       | 32                       | 32   | 90.4 ± 7.7                                       | 82.6 ± 16.4 |
| Ala135Thr (403gCT <sub>405</sub> → aCT)                 | Sp39             | 32                       | 32   | 89.8 ± 7.9                                       | 75.7 ± 17.2 |
| Leu164Pro (490CtG <sub>491</sub> → CcG)                 | Sp33             | 16                       | 16   | 69.2 ± 6.3                                       | 94.2 ± 9.4  |
| Leu165Pro (492CtT <sub>494</sub> → CcT)                 | Sp37             | 32                       | 32   | 79.8 ± 17.5                                      | 70.8 ± 7.2  |
| Arg200Cys (598cGT <sub>600</sub> → tGT)                 | Sp36, Sp46       | 32                       | 32   | 82.5 ± 9.0                                       | 80.8 ± 14.8 |
| Tyr212Cys (634TaT <sub>636</sub> → TgT)                 | Sp32, Sp34       | 16                       | 32   | 91.2 ± 5.7                                       | 81.7 ± 11.4 |
| Leu248Pro (739CtA <sub>741</sub> → CcA)                 | Sp45             | 32                       | 32   | 77.1 ± 7.8                                       | 80.27 ± 7.5 |
| Trp142stop codon (424TgG <sub>426</sub> → TaG)          | Sp38             | 16                       | 16   | 75.8 ± 11.4                                      | 80.9 ± 10.0 |
| Frame shift by insertion of an A between nt 355 and 356 | Sp35, Sp40, Sp41 | 32                       | 32   | 87.9 ± 7.9                                       | 82.6 ± 8.8  |
| ΔRlmA <sup>II</sup>                                     | Sp274            | 16                       | 16   | 73.7 ± 10.0                                      | 85.7 ± 11.5 |

<sup>a</sup> Values for at least 3 independent experiments are given.

<sup>b</sup> Quantification of the band corresponding to dimethylation at A2058 relative to the total band. Mean values and standard deviations for at least 3 independent experiments are given.

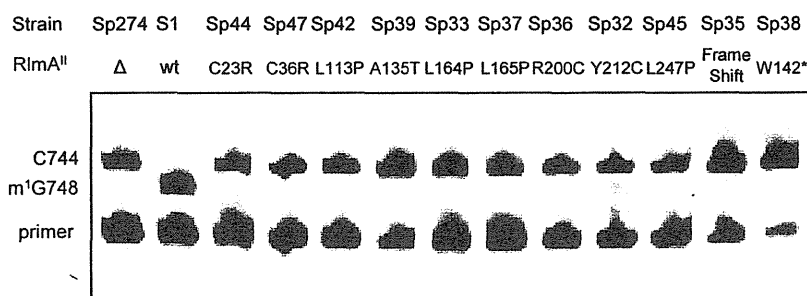


FIG 2 Levels of methylation at G748 in 23S rRNAs extracted from TEL-resistant *S. pneumoniae* strains. rRNAs were sequenced by reverse transcriptase extension from bases 756 to 790. Methylation at G748 N-1 aborts the extension.

ORFs were analyzed for the two strains. As a result, 2 mutations were found in Sp36. One mutation was in a gene encoding a glycosyltransferase, leading to an Ala21Val alteration. The genomic sequences that have been reported show that the gene encoding the glycosyltransferase is not conserved among *S. pneumoniae* strains, suggesting that this mutation may be unrelated to the common mechanism of TEL resistance. The other was in the *tlrB* gene, encoding RlmA<sup>II</sup>, which is a methyltransferase; this mutation results in an Arg200Cys alteration. To detect alterations in RlmA<sup>II</sup> proteins from other TEL-resistant mutants, DNA fragments containing the *tlrB* genes from TEL-resistant mutants were generated by PCR amplification and sequenced. Each TEL-resistant mutant had one nucleotide difference in the *tlrB* gene (Table 3). The *tlrB* gene consists of 849 nucleotides (from ATG to TAA) which are translated to a polypeptide of 282 amino acid residues with a molecular mass of 35.1 kDa. In strain Sp38, a G752A difference changed Trp142 to a stop codon. In strains Sp35, Sp40, and Sp41, one adenine base insertion between nucleotides 355 and 356 generated a frameshift mutation creating amino acid changes in residues 121 to 125, with a stop codon at residue 126. In the other strains, each point mutation caused one amino acid change (Table 3). RlmA<sup>II</sup> methylates the N-1 position of nucleotide G748 in 23S rRNA domain II of *S. pneumoniae* (40). To reveal the level of N-1 methylation at G748 in TEL-resistant mutants, total RNAs were extracted from the parental strain S1, TEL-resistant mutants, and an RlmA<sup>II</sup>-deficient mutant, Sp274, which was constructed by inserting *aad(9)* into the *tlrB* gene of strain S1. The synthesis of a cDNA strand was initiated by annealing an oligonucleotide to positions 756 to 790 of the 23S rRNA. The reverse transcriptase stops synthesizing the DNA strand when it reaches the nucleotide preceding an m<sup>1</sup>G nucleotide, since the N-1 of guanosine required for proper Watson-Crick base pairing to a cytosine is blocked by a methyl group (29). In the absence of methylation at position 748, the reverse transcriptase stops synthesizing the DNA strand when it reaches the cytosine at nucleotide 744, because of the presence of ddGTP in the mixture. The top band in the strain S1 lane in Fig. 2 corresponds to m<sup>1</sup>G748, whereas the top band in the RlmA<sup>II</sup>-deficient mutant Sp274 lane corresponds to C744. For all TEL-resistant strains, bands corresponding to C744 appeared (Fig. 2). These results indicate that the function of RlmA<sup>II</sup> disappeared in TEL-resistant mutants.

To confirm that the inactivation of RlmA<sup>II</sup> confers TEL resistance, we determined the TEL MIC of the RlmA<sup>II</sup>-deficient mutant Sp274 derived from strain S1 (Table 3). Susceptibility testing showed that disruption of RlmA<sup>II</sup> resulted in resistance of *S. pneu-*

*moniae* to TEL (MIC, 16 μg/ml). TEL resistance due to RlmA<sup>II</sup> disruption was complemented (MIC, 1 μg/ml) in strain Sp274 when the pTKY1111 plasmid, carrying *tlrB* of strain S1, was provided, suggesting that RlmA<sup>II</sup> is involved in the TEL susceptibility of strain S1. On the other hand, the RlmA<sup>II</sup> disruption in the laboratory strain TIGR4 did not cause TEL resistance, i.e., both mutant strain Sp303 and parental strain TIGR4 showed TEL MICs of <0.03 μg/ml. From these results, we conclude that RlmA<sup>II</sup> functioning is involved in low TEL susceptibility of *S. pneumoniae* and that the loss of function in combination with the constitutive dimethylation of A2058 by Erm(B) confers high-level TEL resistance in *S. pneumoniae*.

**The methyl group at the N-1 position of G748 contributes to stable interaction of TEL with the ribosome.** The alkyl-aryl side chain of TEL increases the affinity of the ketolide scaffold for the ribosome by a factor of several hundred (10). A structural study of TEL binding to the 70S ribosome from *E. coli* showed that this side chain closely approaches the loop of hairpin 35 in domain II of 23S rRNA and establishes a stacking interaction with the A752-U2609 base pair (12). However, the orientation of this side chain is dependent on the nucleotide sequence of 23S rRNA (12, 41). In *S. pneumoniae* 23S rRNA, the sequence of the A752-U2609 base pair is conserved. A deletion of A752 leads to resistance to TEL (42). This suggests that the A752-U2609 base pair contributes to the proper interaction of the alkyl-aryl side chain with the *S. pneumoniae* 23S rRNA. To examine the interaction of TEL with the A752-U2609 base pair after dimethylation of A2058 by Erm(B), we probed the S1 ribosome in solution by RNA footprinting for TEL protection of A752, because binding of the antibiotic to *E. coli* and *S. aureus* ribosomes protects A752 from chemical modification with DMS (12). Nucleotide A752 was protected from chemical modification in the S1 ribosome treated with TEL (Fig. 3), indicating that A752 is involved in the specific binding of TEL, even after dimethylation of A2058 by Erm(B).

The effect of methylation at G748 was investigated theoretically by comparing two energetically optimized conformations of TEL-binding regions, with and without a methyl group at G748 (Fig. 4). In the energetically minimized structure of the unmethylated-G748 model, the distances from the center of the 5-member ring and that of the 6-member ring of A752 to the center of the aromatic ring of the alkyl-aryl group of TEL were both 3.9 Å (Fig. 4A). In the methylated-G748 model, these distances were estimated to be 3.9 Å and 3.7 Å, respectively (Fig. 4B). The methyl group of G748 could push the alkyl-aryl group of TEL toward the aromatic rings of A752. As a result, tighter π-π packing between

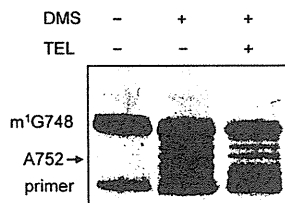


FIG 3 RNA footprinting for TEL protection of A752 in 23S rRNA containing dimethylated A2058. DMS was added to bacterial cultures of RlmA<sup>II</sup>-positive strain S1 after treatment with and without TEL. Reactions were quenched, and total RNAs were purified and used in primer extension assays to detect base modifications near the TEL interaction site in 23S rRNA. Unmodified RNA was used as a control (DMS -, TEL -). The gel shows the primer extension products from bases 756 to 790.

the alkyl-aryl ring of TEL and the aromatic rings of A752 was formed in the methylated-G748 model than in the unmethylated-G748 model. In contrast, 2 hydrogen bonds, formed by one of the hydrogen atoms on the amino group at the N-6 position of A2058 bonding with the oxygen atom of a hydroxyl group and the nitrogen atom of the N(CH<sub>3</sub>)<sub>2</sub> group on the cladinose sugar of TEL, were not affected by methylation of G748 (Fig. 4). This suggests that the affinity of TEL for the ribosome decreases when G748 is not methylated due to the inactivation of RlmA<sup>II</sup>. Furthermore, the methyl group of G748 pointing toward the alkyl-aryl group of TEL also formed a  $\pi$ -CH interaction with the aromatic ring of the alkyl-aryl group. The distance between the carbon atom of the methyl group and the center of the aromatic ring of the alkyl-aryl group was estimated to be 4.0 Å, corresponding to an interaction energy of -0.61 kcal/mol (Fig. 4B). This direct interaction between the methyl group of G748 and TEL may also contribute to stabilization of TEL binding to the ribosome.

## DISCUSSION

Inactivation of the methyltransferase RlmA<sup>II</sup>, which methylates the N-1 position of nucleotide G748, located in hairpin 35 of domain II of 23S rRNA, results in increased resistance to TEL when A2058 is constitutively dimethylated by Erm(B) in *S. pneumoniae*. Constitutive dimethylation of A2058 by Erm(B) causes low sus-

ceptibility to TEL in *S. pneumoniae* (Fig. 1). The central macrolactone ring of TEL establishes hydrophobic interactions with nucleotides 2057, 2058, and 2611 in domain V, which partially form the tunnel wall on the side of the PTC (3). Both the oxygen and nitrogen atoms of the cladinose sugar of TEL form hydrogen bonds with one of the hydrogen atoms of the amino group on A2058 (Fig. 4) (12). Each hydrogen atom of the amino group of A2058 may contribute to these interactions. In the case of monomethylation, one hydrogen atom maintains the interactions with TEL. Therefore, constitutive dimethylation is necessary for resistance and low susceptibility to TEL.

We found 2 nucleotide differences (C242G and T253G) in the regulatory region of the *erm(B)* gene of strain S1 (Fig. 1B). Two stem-loop structures are formed in the regulatory region of *erm(B)* mRNA and are altered due to ERY binding to the ribosome (36). A stem-loop downstream of these structures sequesters the translational initiation region of the methylase structural gene. Thus, it is necessary for the translational initiation of the methylase to collapse this stem-loop. Both nucleotide differences are located in the downstream stem-loop (Fig. 1B). A T253A nucleotide difference has been identified in laboratory-derived mutants of *S. pneumoniae* showing resistance to TEL (19) and in clinical isolates of TEL-resistant enterococci (36). Min et al. (36) showed that the T253A mutation increases the minimum free energy of the stem-loop structure from -13.2 to -8.5 kcal/mol, which results in destabilization of the structure. The minimum free energy of the structure with the C242G and T253G changes is increased further. Therefore, these differences result in stable expression of Erm(B), leading to an increase in the dimethylation level of A2058 and in low susceptibility to TEL. Furthermore, a 182-bp deletion was found in the upstream region of *erm(B)* compared with the sequence of Tn3872 (Fig. 1A). The -35 region of the *erm(B)* promoter was included in the deleted region (35). Transcription of the *erm(B)* gene of plasmid pIP1527 starts from 3 different sites following 3 overlapping promoters, which are recognized by both *E. coli* and *Bacillus subtilis* RNA polymerases (43). Two promoters are also located within the upstream region of *erm(B)* of strain S1. Thus, the *erm(B)* gene of strain S1 could be transcribed from these promoters.

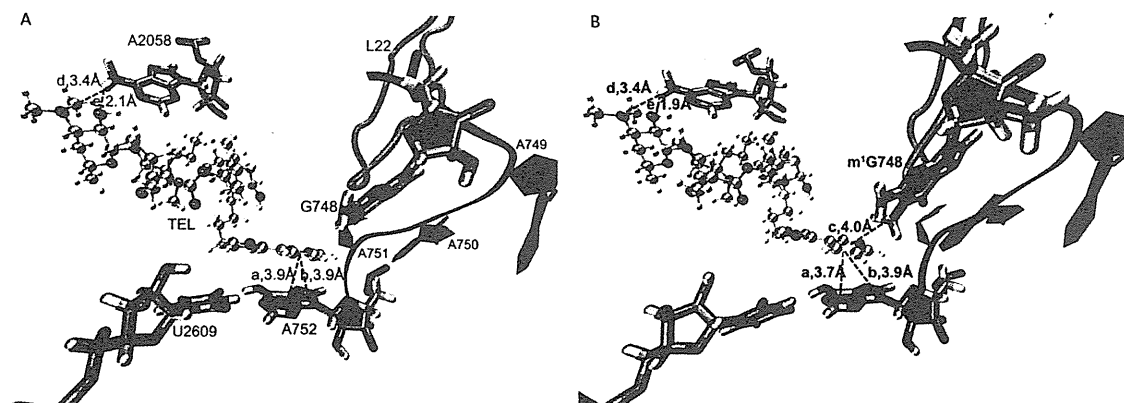


FIG 4 Energetically minimized structures of TEL binding to the 50S subunit of the ribosome containing unmethylated (A) and methylated (B) G748. Residues 748 to 752 in hairpin 35 are represented by a ribbon model (blue). Ribosomal protein L22 is shown by a tube model (green). TEL is represented by a ball-and-stick model. Dashed lines show the distances from the center of the aromatic ring of the alkyl-aryl chain of TEL to the center of each aromatic ring of A752 (a and b) and to the carbon atom of the methyl group at the N-1 position of G748 (c) and from a hydrogen atom of the amino group at the N-6 position of A2058 to the nitrogen atom (d) and to the oxygen atom of the hydroxyl oxygen group (e) on the cladinose sugar of TEL.



The loop in hairpin 35 in domain II of 23S rRNA, which consists of 8 nucleotides (<sub>745</sub>GUUGAAAA<sub>752</sub>), is close to the NPET construction formed by ribosomal proteins L4 and L22 and portions of domain V of 23S rRNA (Fig. 4) (3). The side chains of TEL and a 16-member ring macrolide, tylosin, can also interact with hairpin 35 in addition to interacting with domain V. Thus, the interaction of both TEL and tylosin with the ribosome persists even after monomethylation of A2058 by Erm (11, 44). However, interaction of TEL with hairpin 35 differs from that of tylosin (45). The C<sub>14</sub>-linked mycinose of tylosin interacts with 2 nucleotides, G748 and A751, in hairpin 35 (46). Thus, methylation of G748 by RlmA<sup>II</sup>, which was first identified in *Streptomyces fradiae*, an organism that naturally produces tylosin (47), inhibits the binding of tylosin to the ribosome when A2058 is monomethylated by Erm (44). Similarly, in *S. pneumoniae*, methylation of G748 by RlmA<sup>II</sup> is assumed to inhibit the binding of tylosin, because disruption of RlmA<sup>II</sup> increases sensitivity of the *S. pneumoniae* strain to tylosin when nucleotide A2058 is methylated by Erm (our unpublished data). In the interaction with TEL, the methylation of G748 by RlmA<sup>II</sup> may increase the proper interaction of the alkyl-aryl chain with hairpin 35, even after dimethylation of A2058 (Fig. 3). The alkyl-aryl ring of TEL is locked in its proper place because of the interaction with the aromatic rings of A752 by  $\pi$ - $\pi$  packing (Fig. 4) (12). In the *S. pneumoniae* ribosome, the  $\pi$ -CH interaction can be formed between the alkyl-aryl ring and the methyl group of G748 (Fig. 4B). Moreover,  $\pi$ - $\pi$  interactions within A752 are more stable in the methylated-G748 ribosome model than in the unmethylated-G748 ribosome model (Fig. 4). Thus, constitutive dimethylation of A2058 by Erm alone is insufficient to confer TEL resistance in *S. pneumoniae*.

Hairpin 35 may be involved in the interaction of ERY (11, 48, 49). However, mutations and a deletion of A752 do not affect the ERY MIC in *E. coli* (45). In contrast, A752 deletion in *S. pneumoniae* 23S rRNA results in a significant increase in MICs of the other macrolides, in addition to TEL (42). Increased MICs of clindamycin and pristinamycin are moderate compared with those of the other macrolides (42). This result suggests that hairpin 35 in *S. pneumoniae* 23S rRNA is involved in the direct interaction of the central macrolactone ring.

Comparison of the amino acid sequences of RlmA<sup>I</sup> and RlmA<sup>II</sup> indicates that these enzyme classes are homologous; ~29% of the residues are conserved across species (40, 50). In this study, several mutants generating an amino acid change in RlmA<sup>II</sup> were isolated, all of which reduced RlmA<sup>II</sup> activity (Fig. 2). Each amino acid function was considered according to the results of a structural study of RlmA<sup>I</sup> (50). Residues Cys23 and Cys36 are conserved among RlmA classes. They are located within a Zn-binding domain and coordinate with a single Zn ion (50). Residues Leu113, Ala135, Arg200, and Leu247 are located in  $\alpha$ -helices  $\alpha$ 3,  $\alpha$ 4,  $\alpha$ 6, and  $\alpha$ 7. Residues Leu164 and Leu165 are located in the  $\beta$ 8  $\beta$ -strand. RlmA<sup>I</sup> works as a dimer, because it contains a wide, W-shaped cleft—a putative binding site for the rRNA substrate. Helices  $\alpha$ 3 and  $\alpha$ 4 and the  $\beta$ 8 strand may be involved in the maintenance of this cleft. Helices  $\alpha$ 6 and  $\alpha$ 7 may be involved in the interaction within the dimer. Therefore, amino acid changes within these regions reduce RlmA<sup>II</sup> activity because of weaker binding to rRNA. Despite functional similarity, the target nucleotide of RlmA<sup>II</sup> is different from that of RlmA<sup>I</sup> (40). In hairpin 35 in *E. coli* 23S rRNA, other modifications are known in addition to G745 methylation by RlmA<sup>I</sup> (3). Nucleotides U746 and U747 are

modified to pseudouridine by RluA and via C-5 methylation by RlmC, respectively (51, 52). In some bacterial species, hairpin 35 is unmodified (53). BLAST searches show that both RluA and RlmC homologs are identified in *S. pneumoniae*. Alignments of these proteins between *S. pneumoniae* and *E. coli* show few similarities. YefA, a homolog of RlmC in *Bacillus subtilis*, methylates U747 and U1939 (54). Therefore, hairpin 35 and other regions of *S. pneumoniae* 23S rRNA may be modified in a different pattern from those of other bacterial species, which might affect the binding of macrolides and ketolides in *S. pneumoniae*.

#### ACKNOWLEDGMENTS

We thank N. Kitagawa, Y. Kuroe, K. Sone, K. Nagamatsu, and A. Kimura for technical assistance. Theoretical calculations were partly performed using the Research Centre for Computational Science, Okazaki, Japan.

This work was supported by a grant (grant H24-Shinkou-Ippan-010) from the Ministry of Health, Labor and Welfare of Japan.

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