

Ito A, Mima T, Yamamoto Y-S-Z, Hagiya M, Nakanishi J, Ito M, Hosokawa Y, Okada M, <u>Murakami Y</u> , Kondo T.	Novel application for pseudopodia proteomics using excimer laser ablation and two-dimensional difference gel electrophoresis.	<i>Lab Investigation</i>	92	1374-1385	2012
Nakata H, Wakayama T, Adthapanyawanich K, Nishiuchi T, <u>Murakami Y</u> , Takai Y, Iseki S.	Compensatory upregulation of myelin protein zero-like 2 expression in spermatogenic cells in cell adhesion molecule-1-deficient mice.	<i>Acta Histochem Cytochem</i>	45	47-56	2012
Kikuchi S, Iwai M, Sakurai-Yageta M, Tsuboi Y, Ito T, Masuda T, Tsuda H, Kanai Y, Onizuka M, Sato Y, and <u>Murakami Y</u> .	Expression of a splicing variant of the CADM1 specific to small cell lung cancer.	<i>Cancer Science</i>	103	1051-1057	2012
Mimae T, Okada M, Hagiya M, Miyata Y, Tsutani Y, Inoue T, <u>Murakami Y</u> , Ito A.	Notch2 and Six1 are up-regulated during progression of early-stage lung adenocarcinoma and define its unfavorable subset at advanced stages.	<i>Clinical Cancer Research,</i>	18	945-948	2012
Nagara Y, Hagiya M, Hatano, N, Futai, E, Suo S, Takaoka Y, <u>Murakami Y</u> , Ishiura S, and Ito A.	Tumor suppressor cell adhesion molecule 1 (CADM1) is cleaved by A disintegrin and metalloprotease 10 (ADAM10) and subsequently cleaved by gamma-secretase complex.	<i>Biochem Biophys Res Commun,</i>	417	462-467	2012

Takahashi Y, Iwai M, Kawai T, Arakawa A, Ito T, Sakurai-Yageta M, Ito A, Goto A, Saito M, Kasumi F, and <u>Murakami Y.</u>	Aberrant expression of tumor suppressors, CADM1 and 4.1B, in invasive lesions of primary breast cancer.	<i>Breast Cancer</i>	19	242-252	2012
Nagata M, Sakurai-Yageta M, Yamada D, Goto A, Ito A, Fukuhara H, Kume H, Morikawa T, Fukayama M, Homma Y, and <u>Murakami Y.</u>	Aberrations of a cell adhesion molecule CADM4 in renal clear cell carcinoma.	<i>International Journal of Cancer</i>	130	1329-1337	2012
Watanabe K, Emoto N, Hamano E, Sunohara M, Kawakami M, Kage H, Kitano K, Nakajima J, <u>Goto A,</u> Fukayama M, Nagase T, Yatomi Y, Ohishi N, Takai D.	Genome structure-based screening identified epigenetically silenced microRNA associated with invasiveness in non-small-cell lung cancer.	<i>Int J Cancer.</i>	130	2580-2590	2012
Kitagawa H, Watanabe K, Kage H, Inoh S, <u>Goto A,</u> Fukayama M, Nagase T, Ohishi N, Takai D.	Pulmonary Venous Invasion, Determined by Chest Computed Tomographic Scan, as a Potential Early Indicator of Zygomycosis Infection: A Case Series.	<i>J Thorac Imaging</i>	27	W97-99	2012

Ota, S., Ishikawa, S., Takazawa, Y., Goto, A., Fujii, T., Ohashi, K., Fukayama, M.	Quantitative analysis of viral load per haploid genome revealed the different biological features of merkel cell polyomavirus infection in skin tumor.	<i>PLOS ONE</i> .	7	e39954	2012
Sohn J, Schetter A, Yfantis H, Ridnour L, Horikawa I, Khan M, Robles A, Hussain S, Goto, A., Bowman E, Hofseth L, Bartkova J, Bartek J, Wogan G, Wink D, Harris CC.	Macrophages, nitric oxide and microRNAs are associated with DNA damage response pathway and senescence in inflammatory bowel disease.	<i>PLOS ONE</i>	7	e44156	2012
Abe J, Ueha S, Yoneyama H, Shono Y, Kurachi M, Goto A, Fukayama M, Tomura M, Kakimi K, Matsushima K.	B cells regulate antibody responses through the medullary remodeling of inflamed lymph nodes.	<i>Int Immunol</i>	24	17-27	2012
Makoto Yamagishi ^{1,2} , Kazumi Nakano ¹ , Ariko Miyake ¹ , Tadanori Yamochi ¹ , Yayoi Kagami ¹ , Akihisa, Tsutsumi ¹ , Yuka Matsuda ¹ , Aiko Sato-Otsubo ³ , Satsuki Muto ^{1,3} , Ate Utsunomiya ⁴ , Kazunari Yamaguchi ⁵ , Kaoru Uchimar ⁶ , Seishi Ogawa ³ , and Toshiki Watanabe.	Polycomb-Mediated Loss of miR-31 Activates NIK-dependent NF-κB Pathway in Adult T-cell Leukemia and Other Cancers.	<i>Cancer cell</i>	17	121-135	2012

Ito T, Williams-Nate Y, Iwai M, Tsuboi M, Hagiyaama M, Ito A, Sakurai-Yageta M, and <u>Murakami Y.</u>	Transcriptional regulation of the <i>CADMI</i> gene by retinoic acid during the neural differentiation of murine embryonal carcinoma P19 cells.	<i>Genes to Cells</i>	16	791-802	2011
Hagiyaama M, Furuno T, Hosokawa Y, Iino T, Ito T, Inoue T, Kakanishi M, <u>Murakami Y</u> and Ito A	Enhanced Nerve-Mast Cell Interaction by a Neuronal Short Isoform of Cell Adhesion Molecule-1, CADM1.	<i>Journal of Immunology</i>	186	5983-5992	2011
Mimae T, Tsuta K, Takahashi F, Yoshida A, Kondo T, <u>Murakami Y</u> , Okada M, Takeuchi M, Asamura H, Tsuda H.	Steroid Receptor Expression in Thymomas and Thymic Carcinomas	<i>Cancer</i>	117	4396-4405	2011
Hosokawa Y, Hagiyaama M, Iino T, <u>Murakami Y</u> , Ito A.	Non-contact estimation of intercellular breaking force using a femtosecond laser impulse quantified by atomic force microscopy.	<i>Proc. Natl. Acad. Sci. USA</i>	108	1777-1782	2011

Tsuda M, Ebihara Y, Mochizuki S, Uchimaru K, Tojo A, Tsuji K.	Reduced dose chemotherapy for acute promyelocytic leukemia with adult Down syndrome.	British Journal of Haematology	155	122-132	2011
Yamin Tian, Seiichro Kobayashi, Nobuhiro Ohno, Masamichi Isobe, Mayuko Tsuda, Yuji Zaike, Nobukazu Watanabe, Kenzaburo Tani, Arinobu Tojo and <u>Kaoru Uchimaru</u>	Leukemic T cells are specifically enriched in a unique CD3 ^{dim} CD7 ^{low} subpopulation of CD4 ⁺ T cells in acute-type adult T-cell leukemia.	<i>Cancer Sci.</i>	102	569-577	2011
Kitano K, Watanabe K, Emoto N, Kage H, Hamano E, Nagase T, Sano A, Murakawa T, Nakajima J, <u>Goto A</u> , Fukayama M, Yatomi Y, Ohishi N, Takai D.	CpG island methylation of microRNAs is associated with tumor size and recurrence of non-small-cell lung cancer.	<i>Cancer Sci.</i>	102	2126-2131	2011
<u>Goto A</u> , Li CP, Ota S, Niki T, Ohtsuki Y, Kitajima S, Yonezawa S, Koriyama C, Akiba S, Uchima H, Lin YM, Yeh KT, Koh JS, Kim CW, Kwon KY, Nga ME, Fukayama M.	Human papillomavirus infection in lung and esophageal cancers: analysis of 485 Asian cases.	<i>J Med Virol.</i>	83	1383-1390	2011

Morita S, <u>Goto A</u> , Sakatani T, Ota S, Murakawa T, Nakajima J, Maeda E, Fukayama M:	Multicystic mesothelioma of the pericardium.	<i>Pathol Int.</i>	61	319-321	2011
Miyazaki H, <u>Goto A</u> , Hino R, Ota S, Okudaira R, Murakawa T, Nakajima J, Fukayama M	Pleural cavity angiosarcoma arising in chronic expanding hematoma after pneumonectomy.	<i>Hum Pathol.</i>	42	1576-1579	2011

III. 研究成果の刊行物・別刷り



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Original contribution

Expression of PRMT5 in lung adenocarcinoma and its significance in epithelial-mesenchymal transition ^{☆,☆☆}

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Summary Although protein arginine methyltransferase 5 (PRMT5) has been implicated in various cancers, its expression pattern in lung adenocarcinoma cell lines and tissues has not been elucidated enough. In this study, microarray analysis of 40 non-small-cell lung carcinoma cell lines showed that *PRMT5* was a candidate histone methyltransferase gene that correlated with epithelial-mesenchymal transition. Immunocytochemical analysis of these cell lines indicated that the expression of PRMT5 was localized to the cytoplasm of E-cadherin-low and vimentin-high cell lines, whereas it was predominant in the nucleus and faint in the cytoplasm of E-cadherin-high and vimentin-low cell lines. Immunohistochemical analysis of lung adenocarcinoma cases (n = 130) revealed that the expression of PRMT5 was high in the cytoplasm of 47 cases (36%) and the nuclei of 34 cases (26%). The marked cytoplasmic expression of PRMT5 was frequently observed in high-grade subtypes (1 of 17 low grade, 21 of 81 intermediate grade, and 25 of 32 high grade; $P < .0001$) such as solid adenocarcinoma with the low expression of thyroid transcription factor 1 (the master regulator of lung) and low expression of cytokeratin 7 and E-cadherin (2 markers for bronchial epithelial differentiation), whereas the high nuclear expression of PRMT5 was frequently noted in adenocarcinoma in situ, a low-grade subtype (6 of 17 low grade, 25 of 81 intermediate grade, and 3 of 32 high grade; $P = .0444$). The cytoplasmic expression of PRMT5 correlated with a poor prognosis ($P = .0089$). We herein highlighted the importance of PRMT5 expression, especially its cytoplasmic expression, in the process of epithelial-mesenchymal transition and loss of the bronchial epithelial phenotype of lung adenocarcinoma.

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1. Introduction

The loss of E-cadherin, as a criterion standard of epithelial-mesenchymal transition (EMT), has been reported in approximately 10% of resected primary lung adenocarcinoma cases [1], and no effective therapeutic method has yet been established for lung cancer with the features of EMT [2].

We recently demonstrated that lung adenocarcinoma could be classified into 2 groups: a bronchial epithelial phenotype and mesenchymal-like phenotype [3]. The “bronchial epithelial phenotype” represents a group of lung adenocarcinomas that highly expresses bronchial epithelial markers and includes the thyroid transcription factor 1 (TTF-1)-positive terminal respiratory unit type [4] in addition to TTF-1-negative tumors that highly express bronchial epithelial markers such as cytokeratin 7 (CK7) and mucin 1 (MUC1) [3]. Mutations or amplifications in *epidermal growth factor receptor (EGFR)*, *c-met proto-oncogene (MET)*, and *human epidermal growth factor receptor 2 (HER2)* are frequent in the bronchial epithelial phenotype. In contrast, the “mesenchymal-like phenotype” is characterized by the absence of bronchial epithelial markers (TTF-1, MUC1, and CK7), no *EGFR*, *MET*, or *HER2* mutations or amplifications, and presence of the features of EMT, such as the low expression of E-cadherin and high expression of fibroblast growth factor receptor 1 (FGFR1), vimentin, and Zinc finger E-box-binding homeobox 1 (ZEB1) [3]. The absence of *EGFR*, *MET*, or *HER2* mutations or amplifications in the mesenchymal-like phenotype suggests that other genetic or epigenetic abnormalities may play a role in this group of tumors.

The identification of epigenetic regulatory factor mutations including *MLL*, *EZH2*, *ARID1A*, and *DNMT3A* in various tumors has recently been attracting a lot of interest [5-8]. Tumors with these mutations have an undifferentiated, stem cell-like, and EMT phenotype, which suggests that epigenetic mechanisms through histone modifications may be correlated with EMT in tumors. In this study, we focused on protein arginine methyltransferase 5 (PRMT5). PRMT5 catalyzes the symmetrical dimethylation of arginine residues on histone and nonhistone substrates and plays multiple roles in cellular processes, including differentiation, proliferation, apoptosis, and ribosome biogenesis [9-15]. Although the overexpression of PRMT5 has been reported in various cancers including lung cancer [13,16-21], its expression pattern in terms of cytoplasmic and nuclear localization in each histologic subtype of lung adenocarcinoma and its relation to bronchial epithelial markers, *EGFR* status, clinicopathological factors, and prognosis have not yet been elucidated in detail. We herein described the distinct expression pattern of PRMT5 and its significance in malignant progression, especially in EMT.

2. Materials and methods

2.1. Gene expression profile of 40 non-small-cell lung carcinoma (NSCLC) cell lines

We used the microarray analysis data of 40 cell lines. Detailed information is available in the Supplementary methods and our previous studies [3,22].

2.2. Cell lines and medium

We used 6 cell lines (HCC4006, H1650, PC3, A549, H522, and H1651) for Western blotting and immunocytochemistry. HCC4006, H1650, and PC3, harboring *EGFR* mutations, were used as representatives of the bronchial epithelial phenotype, whereas A549, H522, and H1651 with wild-type *EGFR* were used as representatives of the mesenchymal-like phenotype in our previous study [3]. All cell lines were maintained in RPMI 1640 media supplemented with 10% fetal bovine serum and 1% antibiotics in a humidified atmosphere with 5% carbon dioxide and 95% air.

2.3. Antibodies

The antibodies used in this study are summarized in Supplementary Table S1.

2.4. Protein analysis and immunocytochemistry of cell lines

Experimental details of Western blotting and immunocytochemistry are given in the Supplementary methods.

2.5. Tissue microarray sections

We used tissue microarrays (TMAs) that were produced to accommodate primary lung adenocarcinoma tissue core sections collected from patients who had undergone surgical resection at the University of Tokyo Hospital between June 2005, and September 2008. Informed consent was obtained from all patients, and the study was approved by the institutional ethics review committee. The demographic and clinicopathological details of patients and tumors are provided in the Supplementary methods.

2.6. Histopathologic grading of each subtype of lung adenocarcinoma

Each case was classified according to the predominant histopathologic subtype in the invasive lesions on TMA

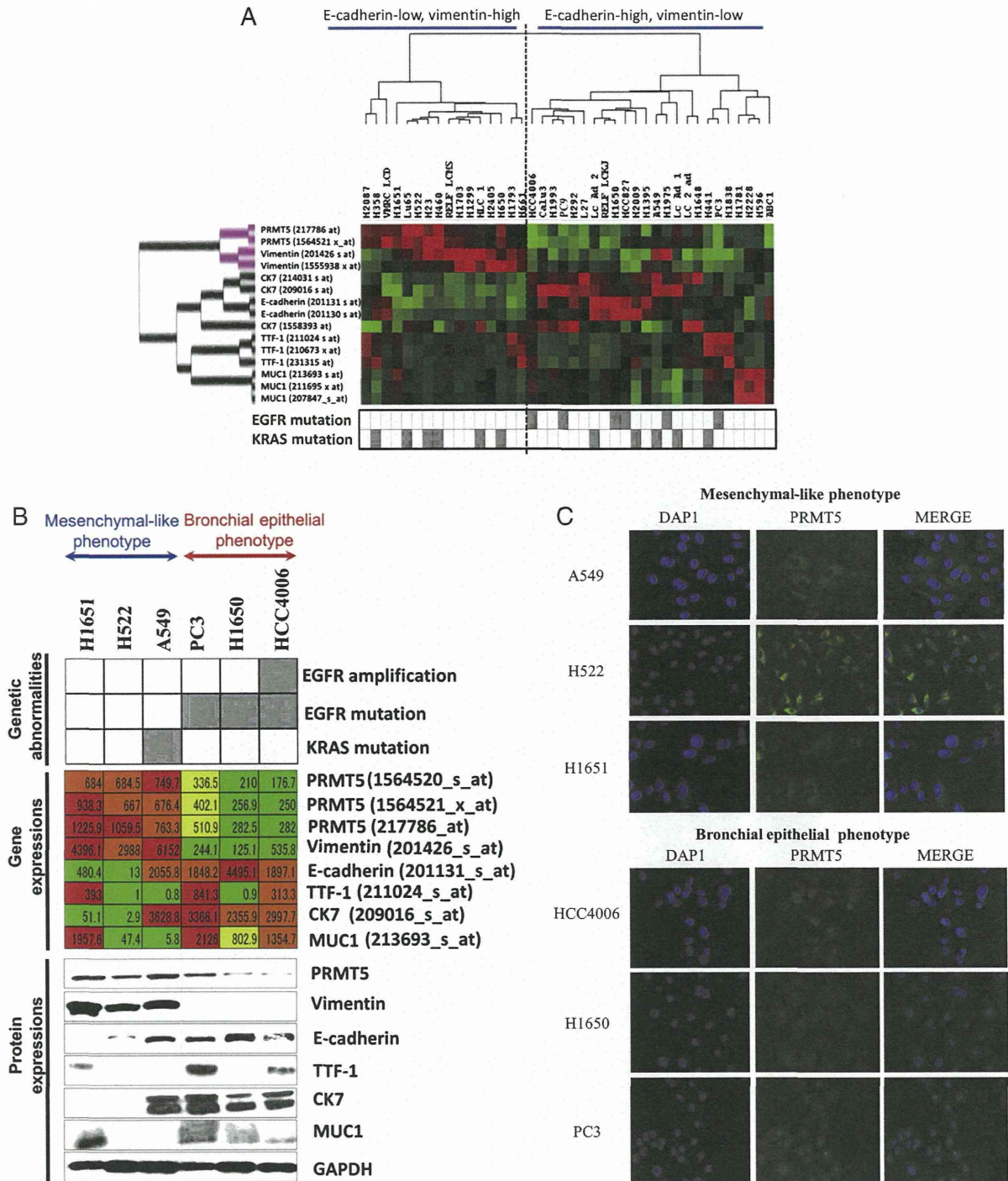


Fig. 1 A, Hierarchical cluster analysis of 40 lung cancer cell lines using *PRMT5*, *vimentin*, *CK7*, *TTF-1*, *E-cadherin*, and *MUC1* gene expression. The genetic statuses of *EGFR* and *KRAS* are shown in the lower panel (the gray box indicates the presence of genetic abnormalities, and the white box shows the absence of genetic abnormalities). B, Genetic statuses of *EGFR* and *KRAS* (upper panel) and gene expression levels (middle panel) and protein expression levels (lower panel) of *PRMT5*, *vimentin*, *E-cadherin*, *TTF-1*, *CK7*, and *MUC1* in the 6 cell lines. Color indications in the middle lane are as follows: red means more than or equal to 1.5 times the average of each gene expression level; orange, less than 1.5 times the average and more than or equal to the average; yellow, less than the average and more than or equal to half the average; and green, less than half the average. C, *PRMT5* immunocytochemical expression in mesenchymal-like phenotype cell lines (A549, H522, and H1651) and bronchial epithelial phenotype cell lines (PC3, H1650, and HCC4006). (Original magnification $\times 400$) (blue, 4',6-diamidino-2-phenylindole (DAPI) for nuclear staining; green, *PRMT5*; merge, mixed nuclear and *PRMT5* view). Gene and protein expression of *PRMT5* in cell lines.