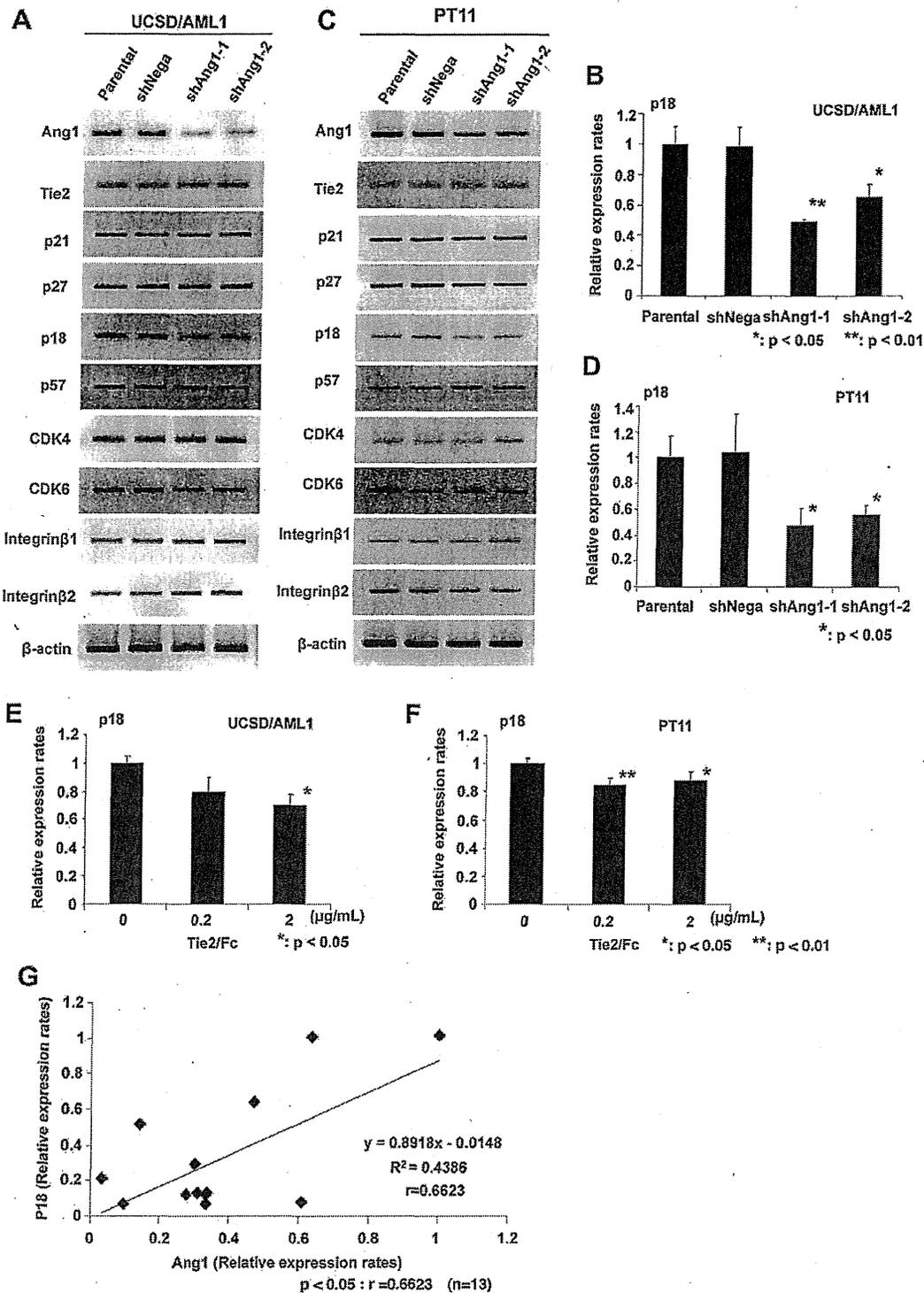


**Fig. 3.** Ang1 negatively regulates the cell cycle in EVI1<sup>high</sup> leukemia cells. (A) Effect of Ang1 knockdown on cell proliferation. shRNA against Ang1 was introduced into UCSD/AML1 cells. The proliferation of parental UCSD/AML1 cells, AML1/shNega cells, and AML1/shAng1-1 and -2 cells were examined using a Cell Counting Kit-8. The absorbance at 450 nm was measured by a microplate reader (reference wavelength: 620 nm). The data are presented as the mean  $\pm$  s.d. Student's *t*-test was used for statistical analysis. The 0 day sample was an aliquot of the plating suspension. (B) Progression through G0/G1, S, and G2/M phase after stimulation of serum-starved cells. Parental UCSD/AML1 cells, AML1/shNega cells, and AML1/shAng1-1 and -2 cells were grown in serum starvation medium with 0.5% serum for 30 h and then stimulated with 10% FCS and 1 ng/mL hGM-CSF. The cells were collected at each indicated time point after stimulation and analyzed for DNA content using PI staining. The cell cycle profiles were determined using ModFit software. The data are presented as the mean  $\pm$  s.d. Student's *t*-test was used for statistical analysis (\**p* < 0.01). (C) Western blot analysis of Rb phosphorylation at Ser780. After serum starvation with 0.5% serum for 30 h, the cells were stimulated with 10% FCS and 1 ng/mL hGM-CSF. The cells were collected at each indicated time point after stimulation and used as samples for Western blot analysis. (D) Progression of the cell population from G0 phase after serum stimulation. The cells were grown in serum starvation medium with 0.5% serum for 30 h and then stimulated with 10% FCS and 1 ng/mL hGM-CSF. The cells were stained with anti-Ki-67 antibody and 7-AAD at just before and 18 h after stimulation and then analyzed using flow cytometry. The population of Ki67-negative cells was counted as G0 phase cells. The data are presented as the mean  $\pm$  s.d. Student's *t*-test was used for statistical analysis.

expressing shRNA specific for Ang1. Ang1-knockdown cells showed enhanced proliferation compared with parental UCSD/AML1 cells or control AML1/shNega cells (Fig. 3A). Next, to investigate the influence of Ang1 knockdown on cell cycle progression, a series of UCSD/AML1 cell lines were synchronized in G0/G1 by 30 h of serum starvation, restimulated with serum and GM-CSF, and then harvested at the indicated times for cell cycle analysis (Fig. 3B). At 12 h after stimulation with FCS and GM-CSF, the percentage of cell population in G0/G1 phase was lower in the

AML1/shAng1 cells than in control cells, whereas the percentage of S phase cells was higher in the AML1/shAng1 cells at the same point. Because the tumor suppressor protein retinoblastoma (Rb) is a major regulator of the G1/S transition, we evaluated the level of phosphorylated Rb (Ser780) at just before, 12 h after, and 24 h after stimulation of starved cells with FCS and GM-CSF. Western blot analysis clearly showed that high Rb phosphorylation at Ser780 was detectable in AML1/shAng1 cells after 12 h poststimulation (Fig. 3C), whereas the level of Rb phosphorylation increased



**Fig. 4.** Ang1 regulates p18 gene expression in EVI1<sup>high</sup> leukemia cells. (A) Semi-quantitative RT-PCR analysis of Ang1, Tie2, cell cycle regulators, and cell adhesion molecules in parental UCSD/AML1 cells, AML1/shNega cells, and AML1/shAng1-1 and -2 cells. (B) Quantitative real-time RT-PCR analysis of p18 in parental UCSD/AML1 cells, AML1/shNega cells, and AML1/shAng1-1 and -2 cells. The data are presented as relative fold changes compared with the expression in parental UCSD/AML1 cells. The data are expressed as the mean ± s.d. Student's *t*-test was used for statistical analysis (\**p* < 0.05; \*\**p* < 0.01). (C) shRNA against Ang1 was introduced into PT11 cells. Semi-quantitative RT-PCR analysis of Ang1, Tie2, cell cycle regulators, and cell adhesion molecules in parental PT11 cells, PT11/shNega cells, and PT11/shAng1-1 and -2 cells. (D) Quantitative real-time RT-PCR analysis of p18 in parental PT11 cells, PT11/shNega cells, and PT11/shAng1-1 and -2 cells. The data are presented as relative fold changes compared with the expression in parental PT11 cells. The data are expressed as the mean ± s.d. Student's *t*-test was used for statistical analysis (\**p* < 0.05). (E and F) UCSD/AML1 and PT11 were incubated in medium containing Tie2/Fc at the indicated concentrations for 48 h. After incubation, p18 gene expression was analyzed by quantitative real-time RT-PCR. The data are presented as the mean ± s.d. Student's *t*-test was used for statistical analysis (\**p* < 0.05; \*\**p* < 0.01). (G) Correlation between the expression levels of Ang1 and p18. The expression levels of Ang1 and p18 in AML patient samples were examined using quantitative real-time RT-PCR analysis. To evaluate the relationship between the expression levels of Ang1 and p18, Pearson's product moment correlation coefficient (*r*) was calculated (*n* = 13, *p* < 0.05; *r* = 0.6623).

in parental UCSD/AML1 cells and AML1/Nega cells at 24 h after stimulation. These results suggest that Ang1 expression negatively regulates cell cycle progression from G0/G1 to S phase in UCSD/AML1 cells.

Because Ang1 is known to be an important factor in the maintenance of the quiescent state of HSCs in the bone marrow niche, we next investigated whether Ang1 is associated with the maintenance of G0 phase in EVI1<sup>high</sup> leukemia cells. We measured the percentages of G0 phase of cell population by double staining with Ki67 and 7-AAD at just before and 18 h after serum stimulation of starved AML1/shAng1, AML1/shNega or parental cells. At just before stimulation, the percentages of cells in G0 phase were not significantly different between AML1/shAng1 and control cells, but they were significantly lower in AML1/shAng1-1 and -2 cells (14–16%) than in parental UCSD/AML1 cells or AML1/Nega cells (20–22%) at 18 h after stimulation (Fig. 3D and Supplementary Fig. 1). These results show that knockdown of Ang1 expression did not influence entry into G0 phase during serum starvation, but it enhanced exit from G0 phase after serum stimulation, suggesting that Ang1 may inhibit cell cycle re-entry from the quiescent state.

#### 3.4. Ang1 regulates p18 gene expression through Ang1/Tie2 signaling in EVI1<sup>high</sup> leukemia cells

To clarify how Ang1 regulates the cell cycle in UCSD/AML1 cells, we assessed the gene expression levels of cyclin-dependent kinases (CDK4 and CDK6), CDK inhibitors (p21, p27, p18, and p57), and cell adhesion molecules ( $\beta$ 1 integrin and  $\beta$ 2 integrin) by semi-quantitative RT-PCR. Compared with parental UCSD/AML1 cells or AML1/shNega cells, p18 mRNA was clearly decreased in AML1/shAng1-1 and -2 cells (Fig. 4A), and the downregulation of p18 in AML1/shAng1-1 and -2 cells was also confirmed using quantitative RT-PCR (Fig. 4B). In addition, the expression of p18 was down-regulated in primary AML cells introduced with shAng1 (PT11/shAng1) (Fig. 4C and D). We next determined how Ang1 expression regulates the expression of p18 in EVI1<sup>high</sup> leukemia cells. Because Ang1 is a secreted Tie2 ligand, we hypothesized that the Ang1 protein secreted from an EVI1<sup>high</sup> leukemia cell would bind to Tie2 receptors, thereby activating a signaling pathway leading to p18 expression in the nucleus. To confirm whether the secreted Ang1 binds to Tie2 and regulates the expression of p18, recombinant human Tie2/Fc chimeric protein, which acts as a decoy receptor, was added to the medium of UCSD/AML1 and PT11 cells, and the expression levels of p18 were monitored. Quantitative RT-PCR analysis showed that p18 expression levels were decreased in the cells treated with Tie2/Fc (Fig. 4E and F). We then examined p18 gene expression levels in five leukemia cell samples from EVI1<sup>high</sup> AML patients and eight leukemia cell samples from EVI1<sup>low</sup> AML patients using quantitative RT-PCR. Significant correlations were found between the expression of Ang1 and p18 (Fig. 4G). Taken together, these results suggest that secreted Ang1 up-regulates p18 gene expression through Ang1/Tie2 signaling, resulting in a delay in cell cycle progression in EVI1<sup>high</sup> leukemia cells.

#### 4. Discussion

In this study, we demonstrated that Ang1 is highly expressed in EVI1<sup>high</sup> leukemia cells in a manner dependent on EVI1 expression. Moreover, we showed that in EVI1<sup>high</sup> leukemia cells, Ang1 suppresses cell cycle progression via maintenance of G0/G1 phase and up-regulates the expression of p18 through Ang1/Tie2 signaling. Because EVI1<sup>high</sup> AML entails a poor prognosis and is resistance to chemotherapy, our results suggest that the expression of the

negative cell cycle regulator p18 in response to active Ang1/Tie2 signaling may maintain EVI1<sup>high</sup> leukemia cells in G0/G1 phase, which may help to maintain them in the bone marrow niche, resulting in resistance to chemotherapy.

It has been reported that treatment of HSCs with Ang1 up-regulates the expression of the CDK inhibitors p18 and p57 along with that of Itgb1 and Tie2 [29], whereas expression of p57, Itgb1, and Tie2 is not influenced by knocking down Ang1 expression in UCSD/AML1 and PT11 cells, suggesting that p18 may be an important downstream factor regulating the cell cycle in response to Ang1/Tie2 signaling in EVI1<sup>high</sup> leukemia cells. Moreover, p18 has been hypothesized to preserve HSC function by limiting their cell-cycle entry [30], and Nurr1, a nuclear receptor transcription factor, maintains the quiescence of HSCs through up-regulation of p18 gene expression [31]. Therefore, p18 may also play an important role in maintaining the stem cell-like phenotypes of EVI1<sup>high</sup> leukemia cells. Ang1/Tie2 signaling between quiescent HSCs and the endosteal niche contributes to the maintenance of HSCs and protects them against various cellular stresses [17]. Because the resistance of leukemia cells to anti-cancer drugs is known to be dependent on cell quiescence in the bone marrow niche, the maintenance of stem cell-like phenotypes by p18 through the activation of Ang1/Tie2 signaling may be related to leukemia cell chemoresistance and thereby contribute to a poor prognosis.

It was reported that AML patients expressing high levels of Ang2 had significantly longer overall survival after chemotherapy than those with low Ang2 levels [32]. Because Ang2 is a Tie2 receptor antagonist, inactivation of Ang1/Tie2 signaling may lead to a favorable prognosis. This is consistent with our view that the maintenance of cell quiescence by active Ang1/Tie2 signaling is probably a main reason for the poor prognosis of EVI1<sup>high</sup> AML patients. Therefore, inhibiting Ang1/Tie2 signaling may enhance sensitivity to chemotherapy by inducing cell cycle entry from quiescence, and combining an inhibitor of Ang1/Tie2 signaling with other anti-cancer drugs may be an effective treatment for EVI1<sup>high</sup> AML. We hope that inhibitors of the Ang1/Tie2 signaling will be applied to clinical trials, which may contribute to improved patient outcomes.

#### Acknowledgment

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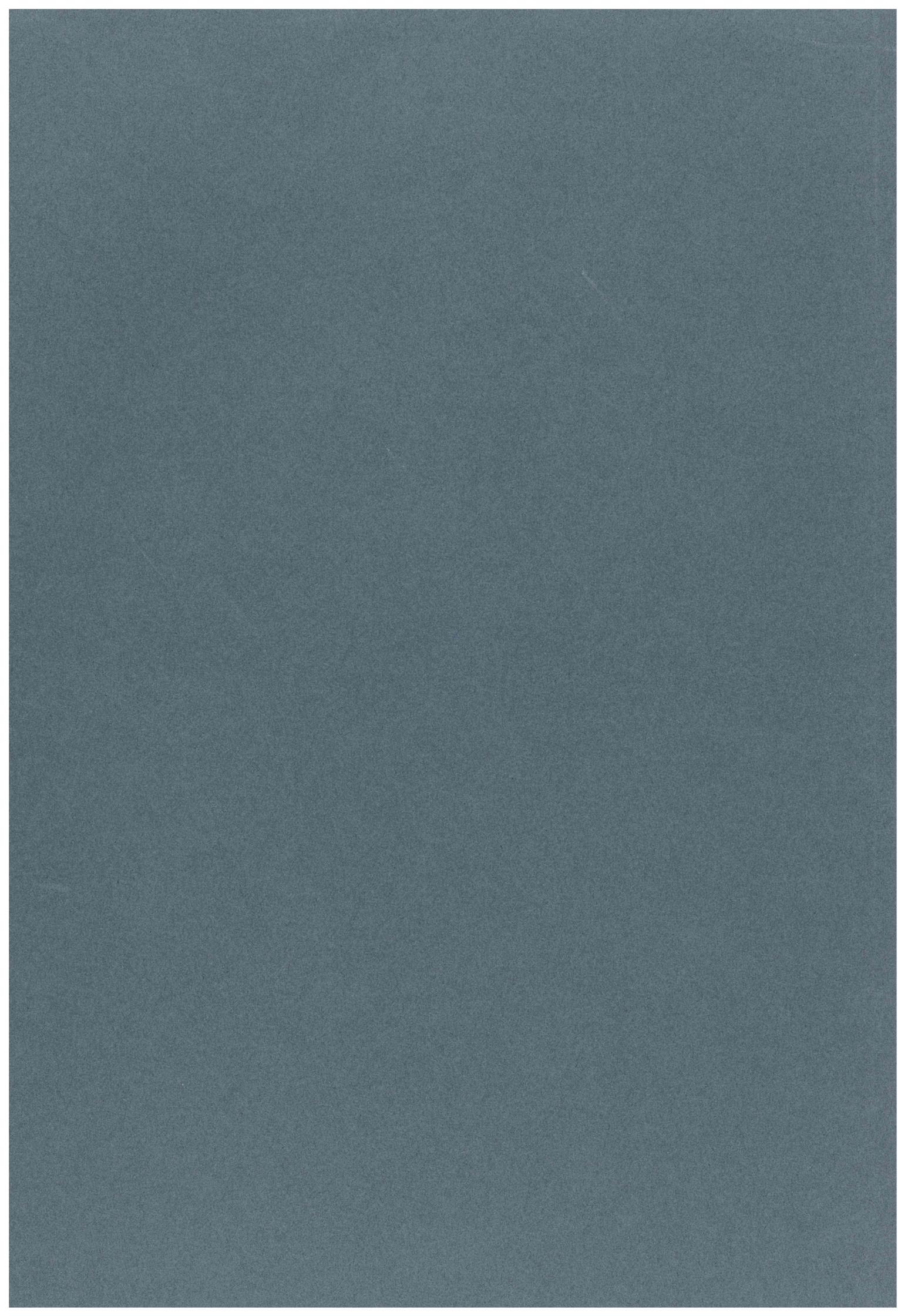
#### Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bbrc.2011.10.061.

#### References

- [1] K. Morishita, D.S. Parker, M.L. Mucenski, et al., Retroviral activation of a novel gene encoding a zinc finger protein in IL-3-dependent myeloid leukemia cell lines, *Cell* 54 (1988) 831–840.
- [2] M.L. Mucenski, B.A. Taylor, J.N. Ihle, et al., Identification of a common ecotropic viral integration site, EVI-1, in the DNA of AKXD murine myeloid tumors, *Mol. Cell. Biol.* 8 (1988) 301–308.
- [3] K. Morishita, E. Parganas, C.L. William, et al., Activation of EVI1 gene expression in human acute myelogenous leukemias by translocations spanning 300–400 kilobases on chromosome band 3q26, *Proc. Natl. Acad. Sci. USA* 89 (1992) 3937–3941.
- [4] K. Mitani, S. Ogawa, T. Tanaka, et al., Generation of the AML1-EVI-1 fusion gene in the t(3;21)(q26;q22) causes blastic crisis in chronic myelocytic leukemia, *EMBO J.* 13 (1994) 504–510.
- [5] S. Lugthart, E. van Druenen, Y. van Norden, et al., High EVI1 levels predict adverse outcome in acute myeloid leukemia: prevalence of EVI1 overexpression and chromosome 3q26 abnormalities underestimated, *Blood* 111 (2008) 4329–4337.

- [6] P.J. Valk, R.G. Verhaak, M.A. Beijten, et al., Prognostically useful gene-expression profiles in acute myeloid leukemia, *N. Engl. J. Med.* 350 (2004) 1617–1628.
- [7] S. Barjesteh van Waalwijk van Doorn-Khosrovani, C. Erpelinck, W.L. van Putten, et al., High EVI1 expression predicts poor survival in acute myeloid leukemia: a study of 319 de novo AML patients, *Blood* 101 (2003) 837–845.
- [8] R.L. Phillips, R.E. Ernst, B. Brunk, et al., The genetic program of hematopoietic stem cells, *Science* 288 (2000) 1635–1640.
- [9] I.K. Park, Y. He, F. Lin, et al., Differential gene expression profiling of adult murine hematopoietic stem cells, *Blood* 99 (2002) 488–498.
- [10] S. Shimizu, T. Nagasawa, O. Katoh, et al., EVI1 is expressed in megakaryocyte cell lineage and enforced expression of EVI1 in UT-7/GM cells induces megakaryocyte differentiation, *Biochem. Biophys. Res. Commun.* 292 (2002) 609–616.
- [11] H. Yuasa, Y. Ojke, A. Iwama, et al., Oncogenic transcription factor *evi1* regulates hematopoietic stem cell proliferation through GATA-2 expression, *EMBO J.* 24 (2005) 1976–1987.
- [12] S. Goyama, G. Yamamoto, M. Shimabe, et al., EVI-1 is a critical regulator for hematopoietic stem cells and transformed leukemic cells, *Cell Stem Cell* 3 (2008) 207–220.
- [13] S. Davis, T.H. Aldrich, P.F. Jones, et al., Isolation of angiopoietin-1, a ligand for the TIE2 receptor, by secretion-trap expression cloning, *Cell* 87 (1996) 1161–1169.
- [14] C. Suri, P.F. Jones, S. Patan, et al., Requisite role of angiopoietin-1, a ligand for the TIE2 receptor, during embryonic angiogenesis, *Cell* 87 (1996) 1171–1180.
- [15] P.C. Maisonpierre, C. Suri, P.F. Jones, et al., Angiopoietin-2, a natural antagonist for Tie2 that disrupts in vivo angiogenesis, *Science* 277 (1997) 55–60.
- [16] D.M. Valenzuela, J.A. Griffiths, J. Rojas, et al., Angiopoietins 3 and 4: diverging gene counterparts in mice and humans, *Proc. Natl. Acad. Sci. USA* 96 (1999) 1904–1909.
- [17] F. Arai, A. Hirao, M. Ohmura, et al., Tie2/angiopoietin-1 signaling regulates hematopoietic stem cell quiescence in the bone marrow niche, *Cell* 118 (2004) 149–161.
- [18] K.A. Moore, I.R. Lemischka, “Tie-ing” down the hematopoietic niche, *Cell* 118 (2004) 139–140.
- [19] J. Oval, O.W. Jones, M. Montoya, et al., Characterization of a factor-dependent acute leukemia cell line with translocation (3;3)(q21;q26), *Blood* 76 (1990) 1369–1374.
- [20] J. Oval, M. Smedsrud, R. Taetle, Expression and regulation of the EVI-1 gene in the human factor-dependent leukemia cell line, UCSD/AML1, *Leukemia* 6 (1992) 446–451.
- [21] H. Hamaguchi, K. Suzukawa, K. Nagata, et al., Establishment of a novel human myeloid leukaemia cell line (HNT-34) with t(3;3)(q21;q26), t(9;22)(q34;q11) and the expression of EVI1 gene, P210 and P190 BCR/ABL chimaeric transcripts from a patient with AML after MDS with 3q21q26 syndrome, *Br. J. Haematol.* 98 (1997) 399–407.
- [22] H. Asou, K. Suzukawa, K. Kita, et al., Establishment of an undifferentiated leukemia cell line (Kasumi-3) with t(3;7)(q27;q22) and activation of the EVI1 gene, *Jpn. J. Cancer Res.* 87 (1996) 269–274.
- [23] P. Martin, T. Papayannopoulou, HEL cells: a new human erythroleukemia cell line with spontaneous and induced globin expression, *Science* 216 (1982) 1233–1235.
- [24] R. Gallagher, S. Collins, J. Trujillo, et al., Characterization of the continuous, differentiating myeloid cell line (HL-60) from a patient with acute promyelocytic leukemia, *Blood* 54 (1979) 713–733.
- [25] C. Sundstrom, K. Nilsson, Establishment and characterization of a human histiocytic lymphoma cell line (U-937), *Int. J. Cancer* 17 (1976) 565–577.
- [26] C.B. Luzzio, B.B. Luzzio, Human chronic myelogenous leukemia cell-line with positive Philadelphia chromosome, *Blood* 45 (1975) 321–334.
- [27] Y. Matsuo, T. Adachi, T. Tsubota, et al., Establishment and characterization of a novel megakaryoblastic cell line, MOLM-1, from a patient with chronic myelogenous leukemia, *Hum. Cell* 4 (1991) 261–264.
- [28] Y. Saito, S. Nakahata, N. Yamakawa, et al., CD52 as a molecular target for immunotherapy to treat acute myeloid leukemia with high EVI1 expression, *Leukemia* 25 (2011) 921–931.
- [29] Y. Gomei, Y. Nakamura, H. Yoshihara, et al., Functional differences between two Tie2 ligands, angiopoietin-1 and -2, in regulation of adult bone marrow hematopoietic stem cells, *Exp. Hematol.* 38 (2010) 82–89.
- [30] H. Yu, Y. Yuan, H. Shen, et al., Hematopoietic stem cell exhaustion impacted by p18 INK4C and p21 Cip1/Waf1 in opposite manners, *Blood* 107 (2006) 1200–1206.
- [31] O. Sirin, G.L. Lukov, R. Mao, et al., The orphan nuclear receptor Nurr1 restricts the proliferation of hematopoietic stem cells, *Nat. Cell Biol.* 12 (2010) 1213–1219.
- [32] C. Schliemann, R. Bieker, T. Padro, et al., Expression of angiopoietins and their receptor Tie2 in the bone marrow of patients with acute myeloid leukemia, *Haematologica* 91 (2006) 1203–1211.



2013/2014B (2/2)

別紙1

総合研究報告書表紙

厚生労働科学研究費補助金

第3次対がん総合戦略研究事業

ヒトATL及びHBZトランスジェニックATL発症マウスを用いた  
比較ゲノム解析によるATL発症機構の解析に関する研究

平成23年度～25年度 総合研究報告書

研究代表者 森下 和広

平成26(2014)年 5月

# The Increased Expression of Integrin $\alpha 6$ (ITGA6) Enhances Drug Resistance in EVI1<sup>high</sup> Leukemia

Norio Yamakawa<sup>1</sup>, Kazuko Kaneda<sup>1</sup>, Yusuke Saito, Emi Ichihara, Kazuhiro Morishita\*

Division of Tumor and Cellular Biochemistry, Department of Medical Sciences, Faculty of Medicine, University of Miyazaki, Miyazaki, Japan

## Abstract

Ecotropic viral integration site-1 (EVI1) is one of the candidate oncogenes for human acute myeloid leukemia (AML) with chromosomal alterations at 3q26. High EVI1 expression (EVI1<sup>high</sup>) is a risk factor for AML with poor outcome. Using DNA microarray analysis, we previously identified that integrin  $\alpha 6$  (ITGA6) was upregulated over 10-fold in EVI1<sup>high</sup> leukemia cells. In this study, we determined whether the increased expression of ITGA6 is associated with drug-resistance and increased cell adhesion, resulting in poor prognosis. To this end, we first confirmed the expression pattern of a series of integrin genes using semi-quantitative PCR and fluorescence-activated cell sorter (FACS) analysis and determined the cell adhesion ability in EVI1<sup>high</sup> leukemia cells. We found that the adhesion ability of EVI1<sup>high</sup> leukemia cells to laminin increased with the increased expression of ITGA6 and integrin  $\beta 4$  (ITGB4). The introduction of small-hairpin RNA against EVI1 (shEVI1) into EVI1<sup>high</sup> leukemia cells reduced the cell adhesion ability and downregulated the expression of ITGA6 and ITGB4. In addition, the overexpression of EVI1 in EVI1<sup>low</sup> leukemia cells enhanced their cell adhesion ability and increased the expression of ITGA6 and ITGB4. In a subsequent experiment, the introduction of shRNA against ITGA6 or ITGB4 into EVI1<sup>high</sup> AML cells downregulated their cell adhesion ability; however, the EVI1<sup>high</sup> AML cells transfected with shRNA against ITGA6 could not be maintained in culture. Moreover, treating EVI1<sup>high</sup> leukemia cells with neutralizing antibodies against ITGA6 or ITGB4 resulted in an enhanced responsiveness to anti-cancer drugs and a reduction of their cell adhesion ability. The expression of ITGA6 is significantly elevated in cells from relapsed and EVI1<sup>high</sup> AML cases; therefore, ITGA6 might represent an important therapeutic target for both refractory and EVI1<sup>high</sup> AML.

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\* E-mail: kmorishi@medmiyazaki-u.ac.jp

These authors contributed equally to this work.

## Introduction

Ecotropic viral integration site-1 (EVI1) is an oncogenic transcription factor for murine and human myeloid leukemia [1,2]. Human EVI1 is localized on chromosome 3q26 [3]. Although only approximately 1 to 3% of acute myeloid leukemia (AML) cases result from a translocation in 3q26, the elevated expression of EVI1 has been detected in 5% to 10% of AML cases in the absence of chromosomal abnormalities at 3q26 [4]. AML with EVI1 high expression (EVI1<sup>high</sup>) is a poor prognosis subtype of AML that does not respond to currently available treatments [5].

EVI1 is a nuclear transcription factor with a DNA-binding zinc finger, an acidic amino acid cluster region and C-terminal binding protein (CtBP) motifs [6,7]. Although EVI1 has been reported to transcriptionally repress or suppress TGF $\beta$  signaling by recruiting Smad3 and the co-repressor CtBP [8–10], we showed that EVI1 is directly associated with the GATA-2 promoter and upregulates GATA-2 transcription to maintain hematopoietic stem cells (HSCs) and AML with EVI1<sup>high</sup> expression [11,12] in EVI1-deficient mice. In addition to the observed reduction in GATA-2 expression, other important factors for HSC maintenance, including Angiopoietin-1 and Tie-2, were also de-regulated in

EVI1-deficient mice [11]. These results suggest that murine Evil might de-regulate transcription factors or other signal transduction molecules necessary for HSC maintenance [11]. However, we do not know precisely how Evil is involved in the maintenance of HSCs.

Recently, there has been increased interest in understanding the regulatory interactions between osteoblasts and HSCs in the bone marrow microenvironment. Individual HSCs are typically anchored to the stroma via a network of adhesion molecules [13,14]. Recent studies have indicated the importance of these adhesion molecules (integrins and cadherins) in hematopoietic stem cell development and have shown that they function as key elements for the detection and translation of the extrinsic cues provided by the hematopoietic microenvironmental niche [15,16]. The integrins are heterodimeric complexes composed of two noncovalently associated transmembrane glycoprotein subunits: one from sixteen different alpha (a) subunits and the other from eight different beta (b) subunits [17,18].  $\alpha 4\beta 1$  integrin heterodimer, participates in both cell-cell and cell-matrix interactions with vascular cell adhesion molecule-1 (VCAM1) and fibronectin (FN). In adult mice, VLA4/VCAM1 interactions are key elements in the mobilization and homing of hematopoietic stem cells to bone marrow [19]. Moreover,

treatment with anti-VLA4 antibodies mobilizes CD34+ hematopoietic progenitor cells from the bone marrow to the peripheral blood [20]. Studies addressing the role of VLA4 in AML cell lines have described the drug resistance induced by the interaction of tumor cells with stromal cells or the extracellular matrix (ECM) as cell adhesion-mediated drug resistance (CAM-DR) [21,22]. In an analysis of 175 adult AML patients, however, VLA4 expression was not significantly associated with the response to anti-cancer drugs or with relapse-free or overall survival rates [23]. Thus, other adhesion molecules may also be important in the maintenance of HSC and leukemia stem cells.

In this study, we observed that the cell adhesion ability of the EVI1<sup>high</sup> AML cells was higher than that of the EVI1<sup>low</sup> cells. The increased adhesion of EVI1<sup>high</sup> AML cells was dependent on the expression of the integrins  $\alpha 6$  (ITGA6) and  $\beta 4$  (ITGB4) in complex with laminin, particularly laminin-332. Because the increased adhesion ability of the AML cells increased their resistance to chemotherapy, and the expression of ITGA6 was significantly higher in AML cases in relapse, ITGA6 might be a novel molecular target in EVI1<sup>high</sup> leukemia cells.

## Results

### Overexpression of ITGA6 enhances the ability of EVI1<sup>high</sup> leukemia cell lines to adhere to matrigel and laminin

To search for novel molecular targets in refractory myeloid leukemia with high EVI1 expression, we previously analyzed the gene expression profiles of 12 human myeloid cell lines using an oligonucleotide microarray (Human Genome U133 Plus 2.0 Array; Affymetrix) containing 38,500 genes [24]. We identified ITGA6 in 26 selected genes that were upregulated over ten-fold in EVI1<sup>high</sup> leukemia cells ( $p < 0.01$ ). To confirm the results of the microarray analysis, the expression levels of a series of integrin genes were determined in three EVI1<sup>low</sup> and EVI1<sup>high</sup> cell lines using semi-quantitative RT-PCR. We determined the levels of the eight different  $\beta$  integrin genes (1 through 8), fifteen different  $\alpha$  integrin genes (1 through 10, E, L, M, V, and X) and two cadherin molecules (N and VE). As shown in Fig. S1A, ITGB1, ITGB3, ITGA4, and ITGA5 were expressed in five of the six leukemia cell lines, and ITGB4 and ITGA6 were specifically expressed in the three EVI1<sup>high</sup> leukemia cell lines. To confirm the results of the expression patterns of the integrin genes, we used the six cell lines and two primary leukemia cell lines from patients with the inv(3) (q21q26) subtype (PT9 and PT11) [24] to determine the expression of ITGB1, ITGB4, ITGA4, and ITGA6. ITGB1 and ITGA4 were expressed as very late antigen-4 (VLA-4) in most of the cell lines, but ITGB4 and ITGA6 were expressed in all three leukemia cell lines and in the two primary EVI1<sup>high</sup> leukemia cell lines (Fig. 1A).

Because a number of integrin genes are involved in binding to fibronectin, laminin and matrigel [15,16], we compared the cell adhesion ability of EVI1<sup>high</sup> and EVI1<sup>low</sup> leukemia cell lines. The three human EVI1<sup>low</sup> myeloid cell lines (HL60, U937, and K562) showed a reduced binding affinity to laminin and matrigel but an increased binding affinity to fibronectin. In contrast, the three EVI1<sup>high</sup> cell lines (UCSD/AML1, MOLM1, and HNT34) exhibited an increased affinity for laminin, matrigel and fibronectin, with significant differences (Fig. 1B). Because the primary component of the matrigel used in this experiment is the laminin complex, the enhanced cell adhesion ability of EVI1<sup>high</sup> leukemia cells might depend on binding to the laminin complex.

We also investigated the ability of various leukemia cells to adhere to the murine bone marrow stromal/osteoblastic cell line MC3T3-E1. As shown in Fig. 1D, a higher percentage of cells in

the three EVI1<sup>high</sup> leukemia cell lines (UCSD/AML1, MOLM1 and HNT34) were attached to the MC3T3-E1 cells compared with the three EVI1<sup>low</sup> leukemia cell lines (HL60, K562 and U937), and this difference was statistically significant ( $p < 0.01$ ). Because the EVI1<sup>high</sup> leukemia cell lines exhibited an enhanced ability to adhere to the laminin complex (Fig. 1C), we propose that the laminin complex might be one of the main molecular targets of this increased adhesion to the MC3T3-E1 cells.

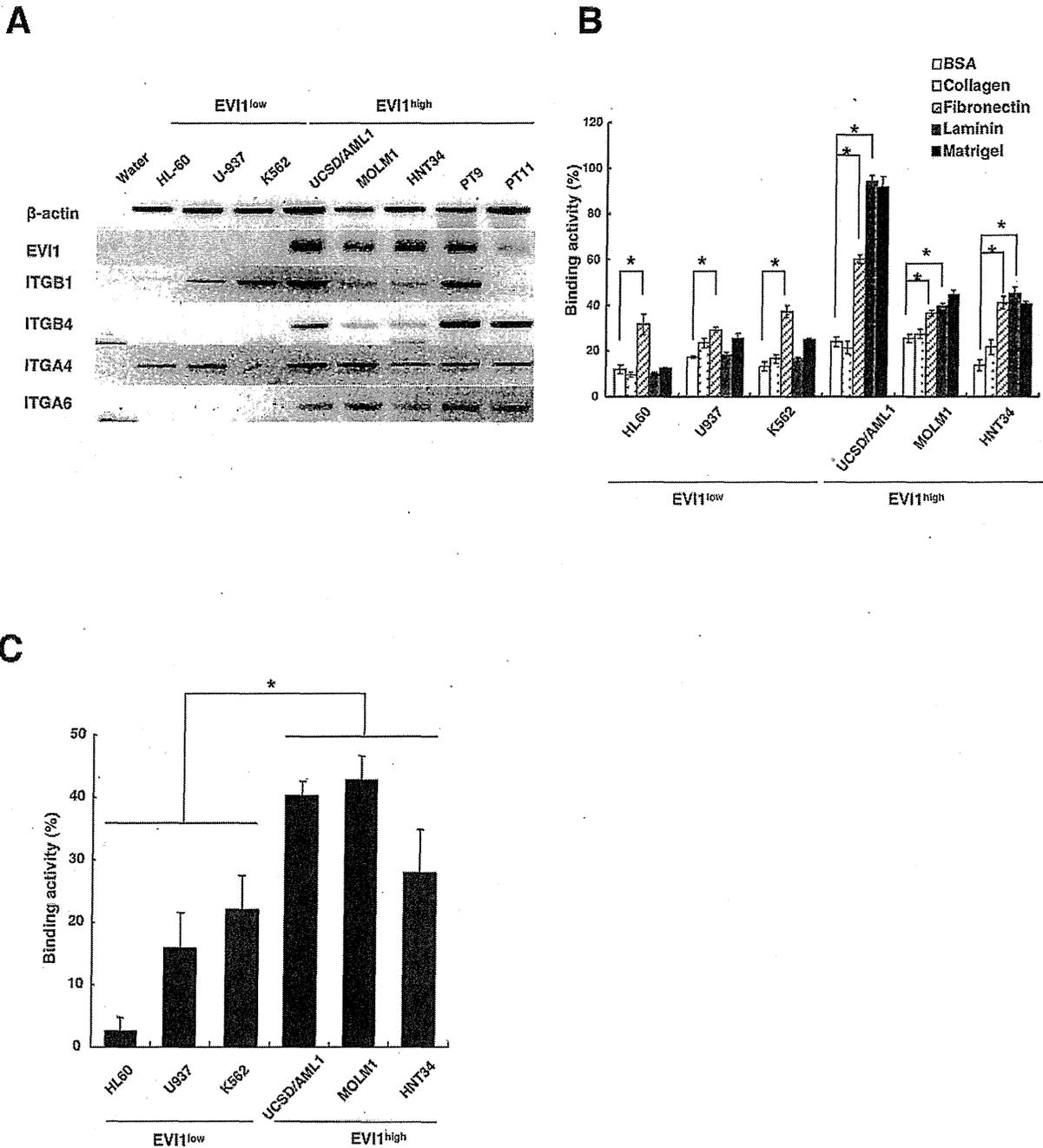
### The expression of ITGA6 and ITGB4 is dependent on the expression of EVI1

To determine which of the integrin genes were dependent on EVI1 expression, small hairpin RNA against EVI1 (shEVI1) were introduced into UCSD/AML1 cells with EVI1<sup>high</sup>. Three UCSD/AML1 cell lines expressing shEVI1 (AML1/shEVI1-1 to -3), and a control cell line expressing an shRNA for firefly luciferase (shLuc) (AML1/shLuc) were established. Because the EVI1 expression was significantly downregulated in the AML1/shEVI1-1 to 3 cell lines (Fig. 2A), we determined the expression of a series of integrin genes using semi-quantitative RT-PCR (Fig. S1B and Fig. 2A). The results showed that the expression of ITGB3, ITGB4, ITGA6, ITGA9, and VE-cadherin was downregulated along with EVI1 in the AML1/shEVI1-1 to 3 cell lines. The high expression of ITGB3, ITGB4 and ITGA6 in the UCSD/AML1 cells and the downregulation of ITGA6, ITGB4 and ITGB3 in the AML1/shEVI1-1 cells were confirmed through immunofluorescence staining and FACS analysis (Fig. 2B). Moreover, ITGB4, ITGA6, ITGA9, and VE-cadherin were also downregulated in the HNT34 cell line, and PT9 and PT11 primary AML cell lines upon shEVI1 transfection, which was associated with a reduction in matrigel cell adhesion (Figs. S1C to F). Taken together, the results demonstrated that ITGB4, ITGA6, ITGA9, and VE-cadherin are the candidate integrin genes with increased adhesion ability in EVI1<sup>high</sup> leukemia cells. Moreover, we determined the cell adhesion ability of the AML1/shEVI1 cell lines and found that the adhesion of AML1/shEVI1-1 to 3 cells to matrigel was significantly reduced compared with that of the parental UCSD/AML1 and AML1/shLuc cells (Fig. 2C). The cell adhesion of the AML1/shEVI1-1 cells to laminin, but not to fibronectin, was also significantly reduced compared with that of the AML1/shLuc cells (Fig. 2D and E).

In a subsequent experiment, we transiently introduced EVI1 and GFP expression vectors into the EVI1<sup>low</sup> U937 cells and determined the resulting levels of expression of the various integrin genes. The adhesion of U937/EVI1 cells to matrigel was increased three-fold, and the expression of ITGB3, ITGB4 and ITGA6 was also increased in U937/EVI1 cells relative to their adhesion ability and expression in the U937/parental and U937/GFP cells (Fig. 2F and Fig. S1G). Because ITGB4 and ITGA6 form a heterodimer that binds to the laminin receptor and treatment with a neutralizing antibody to ITGB3 did not suppress the cell adhesion ability of EVI1<sup>high</sup> leukemia cells (Fig. 3G), we further characterized the relationship between the expression of ITGA6/ITGB4 and the increased adhesion ability of the EVI1<sup>high</sup> leukemia cells.

### The adhesion ability of leukemia cells with EVI1<sup>high</sup> expression is specifically dependent on the expression of ITGA6 and ITGB4

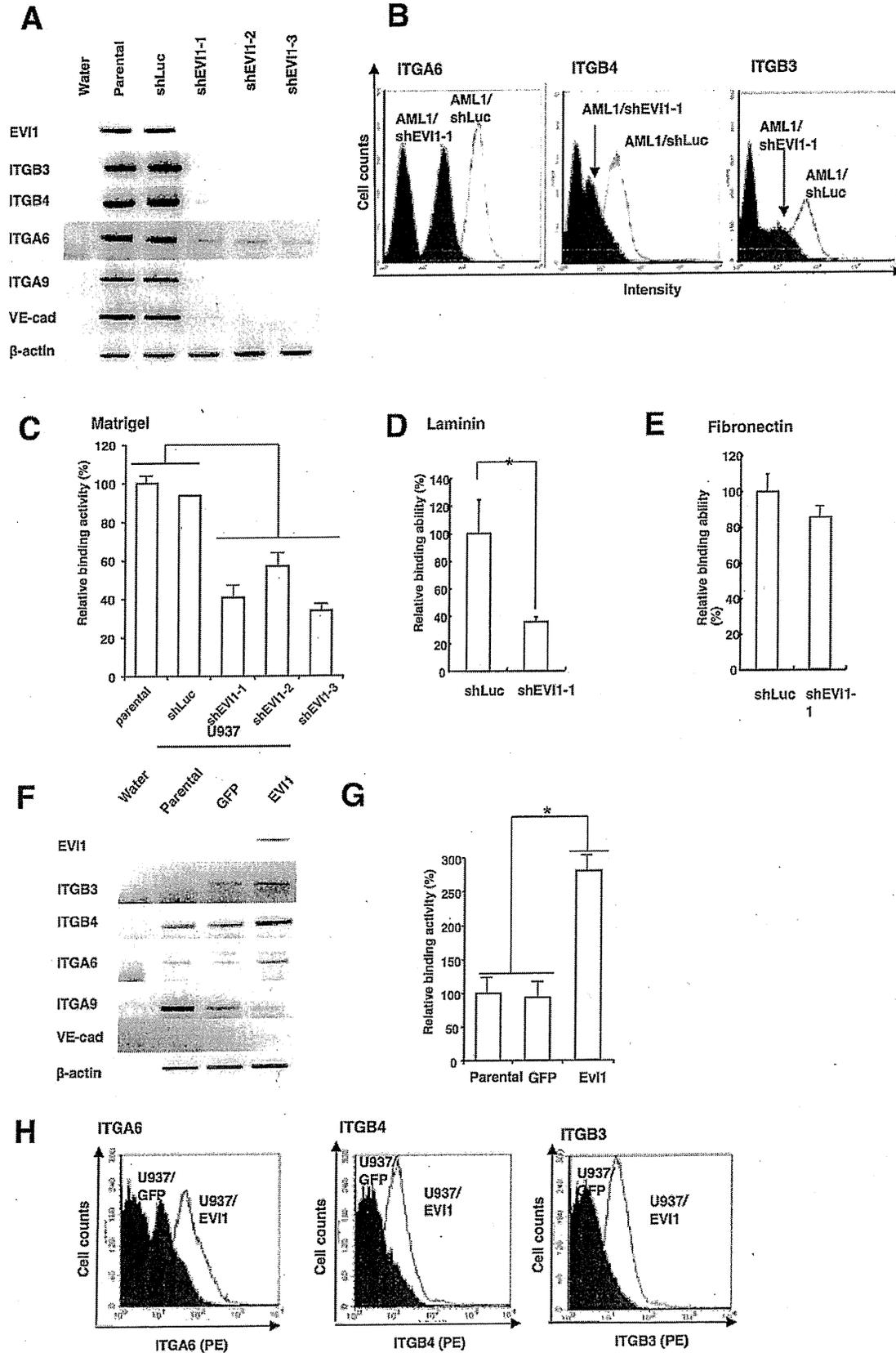
To evaluate whether the adhesion ability of the EVI1<sup>high</sup> leukemia cells was specifically dependent on the expression of ITGA6 or ITGB4, shRNA against ITGA6 or ITGB4 was introduced into UCSD/AML1 cells using an Amaxa Nucleofector,



**Figure 1. Higher cell adhesion ability in AML cell lines with EVI1<sup>high</sup> expression.** **A.** The expression of four integrin genes (ITGB1, ITGB4, ITGA4 and ITGA6), EVI1 and b-actin as a control was determined by semiquantitative RT-PCR in three different EVI1<sup>low</sup> and EVI1<sup>high</sup> AML cell lines and two primary AML cell lines with high EVI1 expression (PT9 and PT11). **B.** Six AML cell lines with low or high EVI1 expression, as indicated in the figure, were incubated in culture medium on BSA, collagen, fibronectin, laminin or matrigel-coated plates; the percentage of the total number of incubated cells that adhered to the plates was designated as the binding activity (%). Each experiment was performed in triplicate, and the experiments were independently repeated at least three times. The data are given as the mean  $\pm$  standard error (S.E). The statistical analysis was performed using the Student's *t*-test ( $*p < 0.05$ , vs. BSA-coated plate). **C.** Six AML cell lines were incubated with the murine osteoblastic cell line MC3T3-E1, and the percentage of cells that bound to MC3T3-E1 cells was determined. Each experiment was performed in triplicate, and the experiments were independently repeated at least three times. The statistical analysis was performed using the Student's *t*-test ( $*p < 0.05$ , vs. EVI1<sup>low</sup> cell lines). doi:10.1371/journal.pone.0030706.g001

and the cell adhesion ability to matrigel was determined. The expression of ITGA6 and the cell adhesion ability were significantly reduced in UCSD/AML1 cells transfected with shITGA6 (AML1/

shITGA6) (Fig. 3A and D). Similar results were obtained in UCSD/AML1 cells transfected with shITGB4 (AML1/shITGB4) (Fig. 3B and E). The growth rate of the AML1/shITGA6 cells was



**Figure 2. The expression of ITGA6 and ITGB4 is specifically dependent on the expression of EVI1.** **A.** The pattern of expression of EVI1 and five integrin genes (ITGB3, ITGB4, ITGA6, ITGA9 and VE-cadherin) in various UCSD/AML1 cell lines. Three cell lines with small hairpin RNA (shRNA) against EVI1 (shEVI1-1, -2 and -3) and the parental and UCSD/AML1 cell lines harboring an expression vector for shRNA against firefly luciferase were used to determine the levels of expression of five integrin (ITGB3, ITGB4, ITGA6, ITGA9 and VE-cadherin) and control (EVI1 and b-actin) genes using RT-PCR. **B.** The expression of ITGA6, ITGB4 and ITGB3 in AML1/shLuc and AML1/shEVI1-1 was determined using FACS analysis after staining the cells with PE-conjugated antibodies to specific integrins. **C.** The same cell lines described in (2a) were assessed for their ability to adhere to the matrigel. The relative binding activity was calculated by comparison to the basal binding activity of the parental UCSD/AML1 cell line. **D and E.** The binding activity of AML1/shLuc and AML1/EVI1-1 cells to laminin (D) and fibronectin (E) was compared. Both cell lines were cultured on laminin or fibronectin-coated plates. **F and G.** After the EVI1 expression vector was introduced into the U937 cell line, the expression of five integrin genes (ITGB3, ITGB4, ITGA6, ITGA9, VE-cadherin) and EVI1 was determined using RT-PCR. b-actin was used as a control (F). The cell adhesion ability of the U937/EVI1, U937/parental and U937/GFP cell lines was determined (G). **H.** The expression of ITGA6, ITGB4 and ITGB3 in U937/GFP and U937/EVI1 was determined using FACS analysis after staining the cells with PE-conjugated antibodies to specific integrins. Each experiment shown in Figure 2C, 2D, 2E, and 2G was performed in triplicate, and the experiments were independently repeated at least three times. The data are given as the mean  $\pm$  S.E. The statistical analysis was performed using the Student's *t*-test (\*\* $p < 0.05$ , vs. each control). doi:10.1371/journal.pone.0030706.g002

reduced, and these cells could not be maintained in culture, whereas the AML1/shITGB4 grew at rate similar to the parental UCSD/AML1 control cells, suggesting that the signal transduction of ITGA6 is essential for the maintenance of leukemia cells. Because ITGA6 forms a heterodimer with ITGB4 or ITGB1, shITGB1 was introduced into UCSD/AML1 cells, and their adhesion to matrigel was measured. The adhesion of UCSD/AML1 cells to matrigel did not change significantly upon transfection with shITGB1 (Fig. 3C and F). These results suggest that the ITGA6 and ITGB4 heterocomplex is the primary cell adhesion molecule on EVI1<sup>high</sup> leukemia cells.

To confirm the ITGA6 dependence of the increased cell adhesion of EVI1<sup>high</sup> leukemia cells, we used a series of neutralizing antibodies. Two leukemia (UCSD/AML1 and MOLM1) and two primary human AML (PT9 and PT11) cell lines were cultured on matrigel and treated with or without anti-ITGA6, ITGB2, ITGB3, or ITGB4 antibodies. Two days after treatment, the binding activity of all four cell lines was significantly reduced by the treatment with anti-ITGA6 or anti-ITGB4 antibodies; however, isotype IgG, anti-ITGB2 or anti-ITGB3 antibodies did not inhibit the binding of EVI1<sup>high</sup> leukemia cells to matrigel (Fig. 3G).

Because the EVI1<sup>high</sup> leukemia cells exhibited an increased adherence to the murine osteoblastic cell line MC3T3-E1 (Fig. 1D) we examined whether this adhesion was dependent on ITGA6 or ITGB4 expression by treating with a neutralizing antibody. Two EVI1<sup>low</sup> (K562 and U937) and three EVI1<sup>high</sup> (UCSD/AML1, PT9 and PT11) leukemia cell lines were co-cultured with MC3T3-E1 cells, and the mixed cultures were subsequently treated with anti-ITGA6 or anti-ITGB4 antibody or the control isotype anti-rabbit IgG. The ability of the three EVI1<sup>high</sup> leukemia cells to bind to MC3T3-E1 cells was significantly reduced upon treatment with anti-ITGA6 or anti-ITGB4 antibody, whereas the binding ability of the two EVI1<sup>low</sup> leukemia cells did not change (Fig. 3H). To confirm this result, we examined the adhesion of U937/parental and U937/EVI1 cells to matrigel upon treatment with anti-ITGA6 antibody. The anti-ITGA6 antibody significantly inhibited the cellular adhesion of U937/EVI1 cells to matrigel, but did not inhibit the cell adhesion of the parental U937 cells (Fig. 3I). Therefore, the expression of ITGA6 and ITGB4 was EVI1 dependent, and the increased expression of ITGA6 and ITGB4 in EVI1<sup>high</sup> leukemia cells increased their ability to adhere to the osteoblastic MC3T3-E1 cell line.

#### The downregulation of laminin-332 in osteoblastic MC3T3-E1 cells partially inhibits binding to EVI1<sup>high</sup> leukemia cells

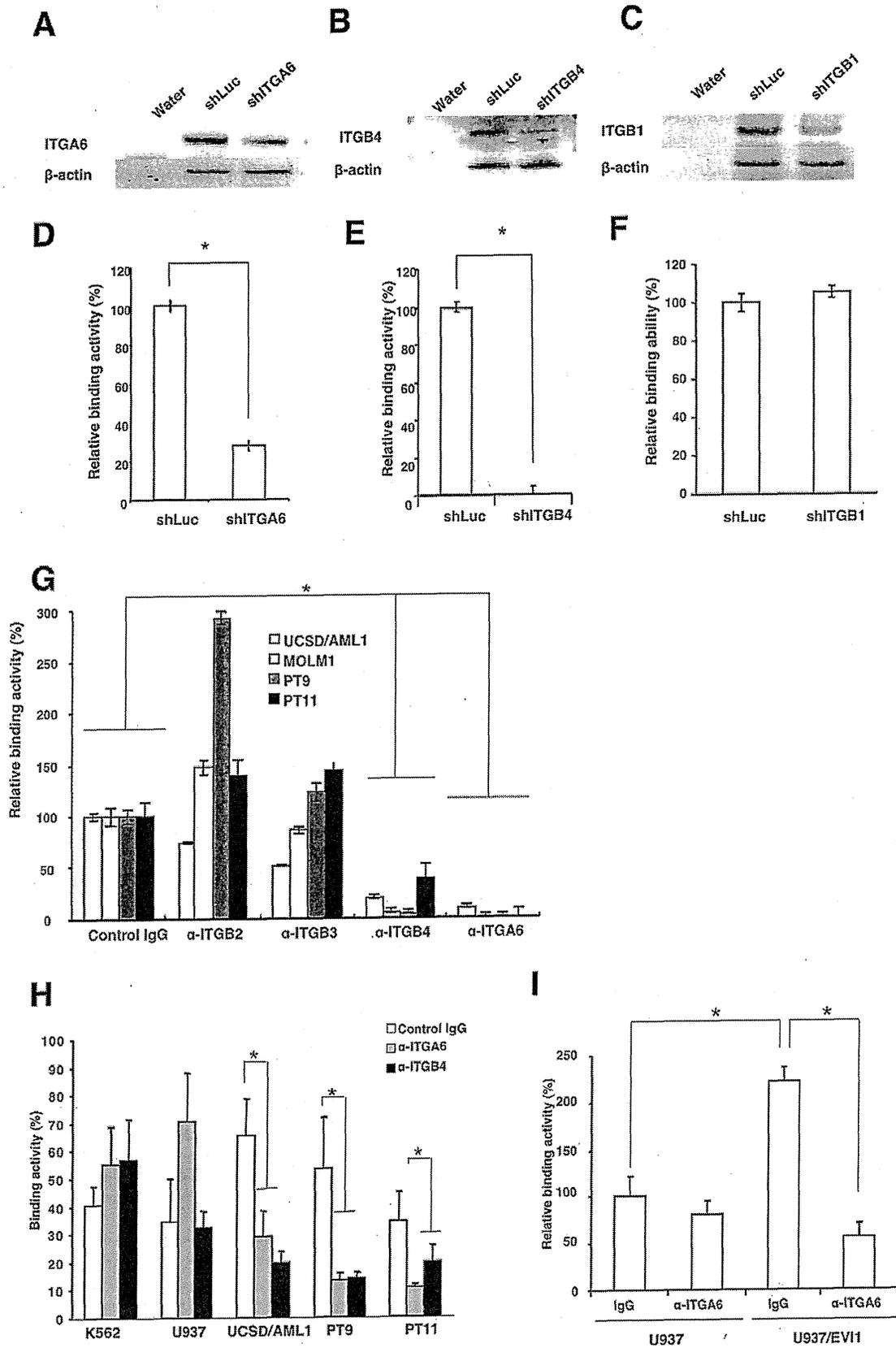
The ITGA6/ITGB4 heterocomplex has a clear specificity for laminin-332 (a3b3g2) or laminin-511 (a5b1g1) [25]. Therefore, we measured the expression of a series of laminin a, b, and g chains in

MC3T3-E1 cells; the expression of each laminin chain was clearly detected in these cells (Fig. S2). We subsequently introduced shRNA against the  $\alpha 3$  chain of laminin-332 (shLN332) into MC3T3-E1 cells and measured the ability of the MC3T3-E1 cells to adhere to various types of myeloid leukemia cells. After the introduction of shLN332 or shLuc into MC3T3-E1 cells (MC3T3-E1/shLN332 or MC3T3-E1/shLuc, respectively), the expression of the  $\alpha 3$  chain of laminin-332 was significantly reduced (Fig. 4A), but the expression of the  $\alpha 3$  chain of laminin-332 and b-actin were not changed. Compared to the MC3T3-E1/shLuc cells, the MC3T3-E1/shLN332 cells displayed significantly reduced adhesion to the EVI1<sup>high</sup> cell lines UCSD/AML1, MOLM1 and HNT34. However, their adhesion to the leukemia cell lines with EVI1<sup>low</sup> expression, such as HL60, K562, and U937, did not change (Fig. 4B). Moreover, the UCSD/AML1/shEVI1 cells displayed reduced adhesion to the MC3T3-E1/shLuc cells compared with that of the UCSD/AML1 cells, whereas the adhesion of the UCSD/AML1/shEVI1 cells to the MC3T3-E1/shLuc and MC3T3-E1/shLN332 cells did not change. These data suggest that the increased adhesion of the leukemia cell lines with EVI1<sup>high</sup> expression is mainly dependent on the laminin complex, particularly laminin-332.

#### EVI1<sup>high</sup> leukemia cells treated with ITGA6 or ITGB4-neutralizing antibodies or small hairpin RNA against EVI1 or ITGB4 recover drug sensitivity

We next determined whether treating EVI1<sup>high</sup> leukemia cells with a neutralizing antibody against integrin improves the drug sensitivity of the cells in the adhered state. Two EVI1<sup>high</sup> leukemia cell lines, UCSD/AML1 and MOLM1, and two primary leukemia cell lines, PT9 and PT11, were used in this experiment; the IC50 of Ara-C for these cells cultured on the matrigel coated plates was approximately  $10^{-6}$  M (Fig. S3). The four cell lines were seeded on matrigel-coated culture plates with or without anti-ITGA6 or anti-ITGB4 antibodies in the presence of  $10^{-6}$  M Ara-C. After four days of culture, the percentage of surviving cells in the cultures treated with anti-ITGA6 or anti-ITGB4 antibodies were significantly reduced in the two EVI1 and primary AML cell lines compared with the control cells (Figs. 5A to D). Because similar results were also obtained using the anti-cancer drug, VP-16 (Fig. S4), the treatment with anti-ITGA6 or anti-ITGB4 antibodies or anti-cancer drugs might recover the drug-sensitivity of EVI1<sup>high</sup> leukemia cells.

To determine whether the effect of Ara-C was dependent on the cell adhesion ability, AML1/shITGB4, AML1/shEVI1 and control AML1/shLuc cells were cultured on matrigel-coated or control BSA-coated plates and treated with Ara-C for three days; we could not use AML1/shITGA6 cells in this experiment. After three days of culture, the number of viable AML1/shLuc cells was



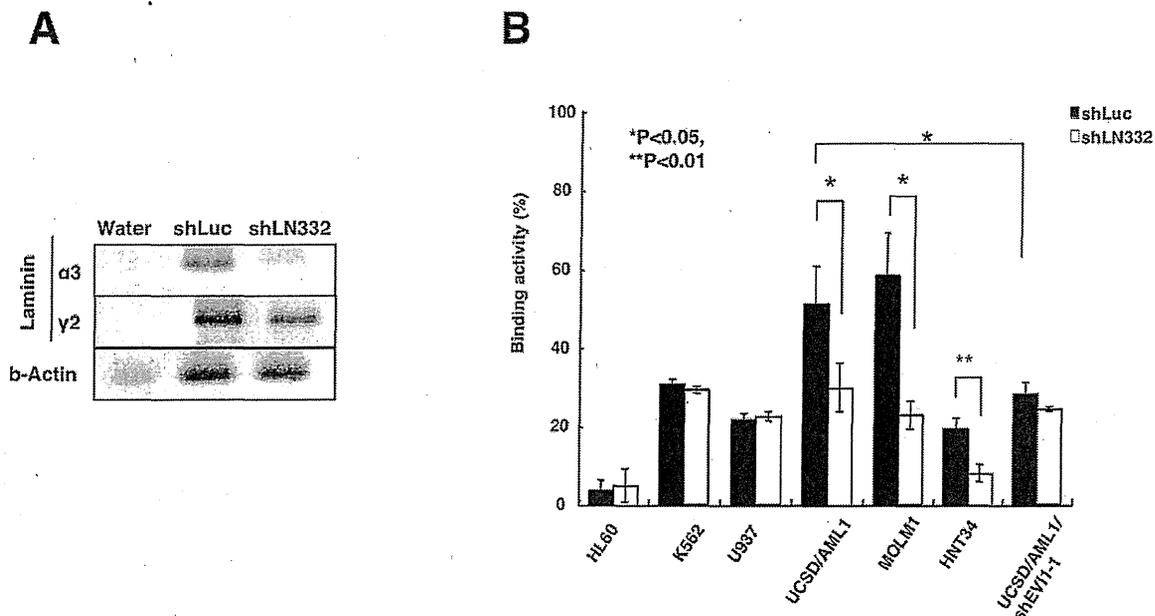
**Figure 3. The adhesion ability of EVI1<sup>high</sup> leukemia is specifically dependent on the expression of ITGA6 and ITGB4. A, B, and C.** UCSD/AML1 cells were transiently transfected with expression vectors for shRNA against ITGA6 (shITGA6), ITGB4 (shITGB4), or ITGB1 (shITGB1), and the expression of ITGA6, ITGB4 and ITGB1 was determined using RT-PCR. UCSD/AML1 cells transfected with shRNA for firefly luciferase (shLuc) were used as a control, and the expression of b-actin was used as an internal control. **D, E, and F.** The cell adhesion ability of the cells in A, B, and C was determined through co-culture with the MC3T3-E1 cell line. The relative binding activity was calculated by comparison to the basal binding activity of control AML1/shLuc cells. **G.** The effect of neutralizing antibodies to the integrins on the cell binding activity of EVI1<sup>high</sup> leukemia cells. Two leukemia (UCSD/AML1 and MOLM1) and two primary human AML (PT9 and PT11) cell lines were cultured on matrigel-coated plates in the presence or absence of anti-ITGA6, ITGB2, ITGB3, or ITGB4 antibodies. The relative binding activity was calculated by comparison to the basal binding activity of each cell line treated with the control isotype IgG. **H.** Two EVI1<sup>low</sup> (K562 and U937) and three EVI1<sup>high</sup> (UCSD/AML1, PT9 and PT11) leukemia cell lines were treated with anti-ITGA6 or ITGB4 antibodies or with the control IgG, and the binding of each cell line to the matrigel-coated plates was determined. **I.** The parental and EVI1-expressing (U937/EVI1) U937 cell lines were treated with anti-ITGA6 or control IgG, and their binding to the MC3T3-E1 cell line was determined. The relative binding activity was calculated by comparison to the basal binding activity of the parental U937 cells treated with the control isotype IgG. Each experiment was performed in triplicate, and the experiments were independently repeated at least three times. The data are given as the mean  $\pm$  S.E. The statistical analysis was performed using the Student's *t*-test ( $*p < 0.05$ , vs. each control). doi:10.1371/journal.pone.0030706.g003

significantly higher when the cells were cultured on matrigel-coated plates compared with conventional culture; however, the percentage of surviving AML1/shITGB4 cells was reduced at each time point (Fig. 5E). The survival rate of the AML1/shEVI1 cells in both the conventional and matrigel-coated cultures was reduced compared with those of the AML1/shLuc and AML1/shITGB4 cells upon Ara-C treatment, whereas there was no difference in the survival rates of the AML1/shEVI1 cells cultured under conventional and matrigel-coated methods under Ara-C treatment. These results suggest that the drug resistance of EVI1<sup>high</sup> leukemia cells is highly dependent on EVI1 expression and partially dependent on ITGB4 expression.

EVI1<sup>high</sup> leukemia cells become more quiescent when cultured on the matrigel-coated plates compared with those cultured on the BSA-coated plates

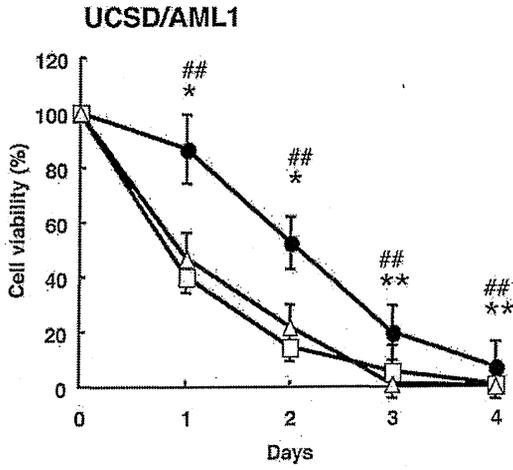
To confirm the advantages of cell adhesion in drug-responsiveness, we determined the growth and cell cycle status of AML cells

with EVI1<sup>high/low</sup>, which were cultured on matrigel or BSA-coated plates. As shown in Fig. 6A, the UCSD/AML cells with EVI1<sup>high</sup> expression (AML1/shLuc) on the matrigel-coated plates grew more faster rapidly on the matrigel-coated plates than that on the BSA-coated plates, although the growth rate of the AML1/shLuc cells was faster than that of the AML1/shEVI1 cells. The cell cycle analysis represented revealed that, relative to AML1/shLuc cells cultured on the BSA-coated plates, the population of EVI1<sup>high</sup> AML1 cells in G0/G1 and G2/M phase increased on the matrigel-coated plates, while the S-phase populations were decreased; however, each cell cycle population of AML1/shEVI1 cells did not change under either culture condition (Fig. 6B and C). Moreover, the population of AML1/shLuc cells in the G0 phase was significantly increased on the matrigel-coated plates compared with the same cells on the BSA-coated plates; however, the population of AML1/shEVI1 cells in the G0 phase was the same under both culture conditions (Fig. 6D). Because the population of U937 cells with high EVI1 expression (U937/EVI1)

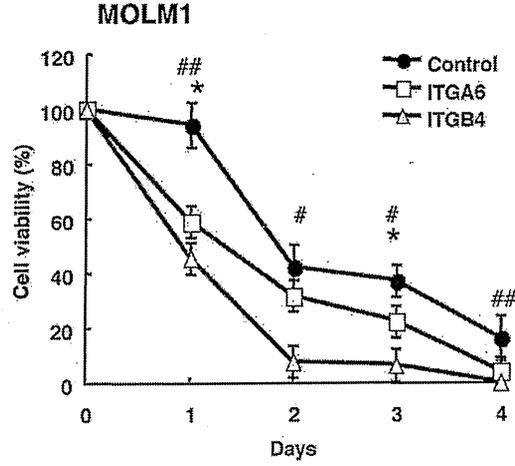


**Figure 4. Laminin-332 on MC3T3-E1 cells is a main target for binding to EVI1<sup>high</sup> leukemia cells. A.** The expression of laminin a3 and g2 in MC3T3-E1 cells transfected with shRNA for firefly luciferase (shLuc) or laminin a3 (shLN332) was determined using RT-PCR. The expression of b-actin was used as an internal expression control. **B.** The binding of various myeloid leukemia cell lines to the two cell lines MC3T3-E1/shLuc or shLN332 was determined. Each experiment was performed in triplicate, and the experiments were independently repeated at least three times. The data are given as the mean  $\pm$  S.E. The statistical analysis was performed using the Student's *t*-test ( $**p < 0.01$ ;  $*p < 0.05$ , vs. control). doi:10.1371/journal.pone.0030706.g004

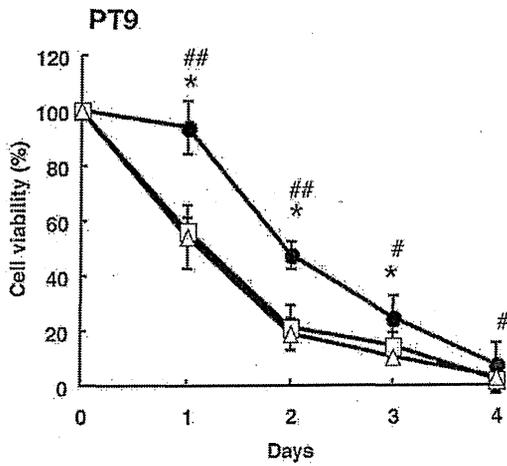
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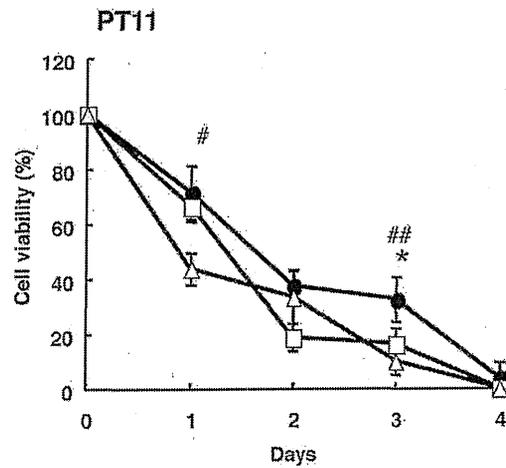
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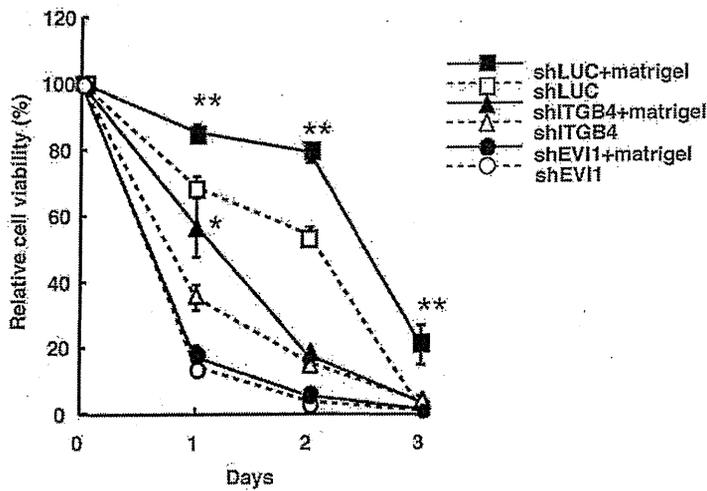
**C**



**D**



**E**



**Figure 5. Drug sensitivity of EVI1<sup>high</sup> leukemia cells is restored by treatment with neutralizing antibodies to ITGA6 or ITGB4 or by transfection with small hairpin RNAs against EVI1 or ITGB4. A–D.** Two myeloid leukemia (UCSD/AML1 and MOLM1) and two primary AML (PT9 and PT11) cell lines were treated with  $1 \times 10^{-6}$  M Ara-C and anti-ITGA6 or ITGB4 antibodies for four days, and the viable cells were counted at each indicated time point. The percent cell viability in comparison with the number of untreated cells on each indicated day is shown. **E.** The UCSD/AML1 cells expressing shRNA for firefly luciferase (shLuc), ITGB4 (shITGB4) or EVI1 (shEVI1) were treated with Ara-C on BSA or matrigel-coated plates for three days. The viable cells were counted at each time point. The relative cell viability is expressed as a percentage of the viability of the untreated cells. Each experiment was performed in triplicate, and the experiments were independently repeated at least three times. The results are given as the mean  $\pm$  S.E. The statistical analysis was performed using the Student's *t*-test (\*\* $p < 0.05$ , vs. control). doi:10.1371/journal.pone.0030706.g005

in the G0-phase was increased on the matrigel-coated plates compared with those on the BSA-coated plates (Fig. S5), the increased quiescence of adhered EVI1<sup>high</sup> AML cells might be one of the reasons for resistance to anti-cancer drugs.

## Discussion

Resistance to treatment with anti-cancer drugs is determined by a variety of factors, including somatic cell genetic differences in tumors and the phenotypes of leukemia stem cells (LSC). LSCs can infiltrate bone marrow niches, leading to enhanced self-renewal and proliferation, and enforced quiescence; the resistance to chemotherapeutic agents and adhesion molecules, such as VCAM-1 and VLA-4, has been described in the localization and retention of normal HSCs and/or leukemia cells within the bone marrow niche [19]. In this manuscript, we examined the cell adhesion ability of EVI1<sup>high</sup> leukemia cells because the increased expression of EVI1 in AML is a well-known prognostic factor for poor outcome and is associated with drug resistance [4,5]. We found that the EVI1<sup>high</sup> leukemia cells with increased ITGA6 and ITGB4 expression exhibited stronger adhesion to laminin complexes than the EVI1<sup>low</sup> leukemia cells; although, the EVI1<sup>low</sup> leukemia cells exhibited increased adhesion to fibronectin. Moreover, the interaction between EVI1<sup>high</sup> leukemia and MC3T3-E1 osteoblastic cells was partially dependent on laminin-332. The downregulation of EVI1 or ITGB4 in UCSD/AML1 leukemia cells restored chemo-sensitivity. The knockdown of ITGA6 expression in EVI1<sup>high</sup> leukemia cells greatly reduced their survival in culture; thus, we conclude that the increased cell adhesion ability of EVI1<sup>high</sup> leukemia cells is primarily dependent on the expression of ITGA6, and that signaling from ITGA6 is crucial for the survival of EVI1<sup>high</sup> leukemia cells.

The expression of the ITGA6/ITGB4 complex could promote tumor progression and the metastasis of various cancer cells, including breast, colorectal, and thyroid carcinomas. Moreover, the laminin receptor for ITGA6 is ubiquitously expressed in human and mouse hematopoietic stem and progenitor cells [26]. In combination with ITGA4, ITGA6 expression in hematopoietic stem and progenitor cells is believed to create a homing receptor for short-term stem cells [27]. ITGA6 functions during the homing of fetal liver HPCs but not during the homing and engraftment of multilineage repopulating HSCs *in vivo* [28]. Therefore, ITGA6 might function as a laminin receptor to sustain HSCs in the bone marrow niche. We are now developing anti-human ITGA6/ITGB4 complex antibodies, which is a new therapy for refractory AML. In preliminary experiments, we demonstrated that the number of EVI1<sup>high</sup> AML cells was significantly decreased in the bone marrow after the intravenous injection of anti-human ITGA6/ITGB4 complex antibodies into NOG immune deficient mice (data not shown). As a corollary, the increased expression of the ITGA6/ITGB4 complex in EVI1<sup>high</sup> leukemia might be an important factor in maintaining leukemia stem cells in the bone marrow.

The adhesion of EVI1<sup>high</sup> AML cells became more resistance to anti-cancer drug therapy, and the population of cells in the

G0-phase was increased. Because the increased population of G0-phase cells is likely dependent on the expression of EVI1, future research should focus on the characterization of the mechanism underlying the increased quiescence of cells through EVI1 expression subsequent to their adhesion to matrigel. Moreover, the induction of PI-3K/AKT/Bcl-2 signaling through the VLA-4-fibronectin interaction results in resistance to anoikis and drug-induced apoptosis [21]; we also showed the cell adhesion of EVI1<sup>high</sup> AML cells to matrigel induced expression of BCL and phosphorylation of AKT (data not shown). Therefore, the adhesion of AML cells may provide advantages to the surviving cells, and the mechanism of drug-resistance is potentially much more complicated; however, we should characterize the survival mechanism of leukemia cells in the bone marrow niche.

To support our data, we examined the expression patterns of ITGA6/ITGB4 and VLA-4 (ITGA4/ITGB1) heterodimers by comparing the gene expression profiles of EVI1<sup>high</sup> and EVI1<sup>low</sup> AML deposited in Gene Expression Omnibus (GSE6891, GEO in NCBI website) [29]. As shown in Fig. S6, the expression of ITGA6 was significantly higher in the 12 cases of EVI1<sup>high</sup> AML than in the 10 cases of EVI1<sup>low</sup> AML ( $p < 0.05$ ); however, the differences in expression of ITGB4, ITGA4 and ITGB1 did not account for the differences between the two groups. We also determined the expression patterns of four integrin genes from the expression profiles of AML cases in remission and relapse that were deposited in GEO at NCBI (GDS1059) [30]. Based on the expression profiles, the expression of ITGA6 was significantly higher in the 28 cases of AML with relapse than in the 25 cases with remission ( $p < 0.05$ ); however, the differences in expression of ITGB4, ITGA4 and ITGB1 did not account for the difference between the two groups. These results show that the increased expression of ITGA6 is an important marker in both EVI1<sup>high</sup> and relapsed AML, which suggests that ITGA6 might be an important target for the molecular-targeted treatment of refractory leukemia, including EVI1<sup>high</sup> AML.

## Materials and Methods

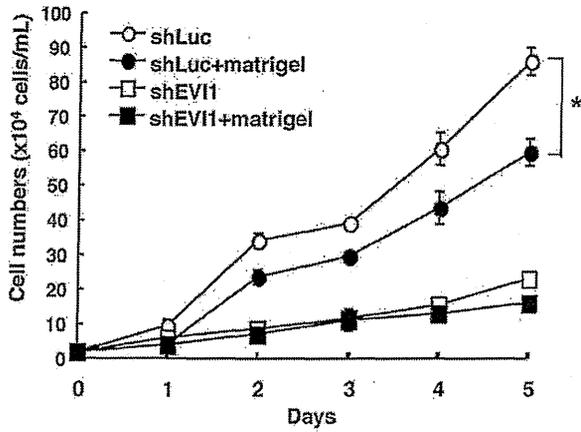
### Cell lines and culture conditions

The MOLM1, HNT34, U937, K562, and HL60 cells were cultured in RPMI1640 medium supplemented with 10% fetal calf serum at 37°C and 5% CO<sub>2</sub>. The UCSD/AML1, PT9 and PT11 cells [24] were cultured in RPMI1640 medium supplemented with 10% fetal calf serum and human granulocyte-macrophage colony stimulating factor (GM-CSF, 1 ng/ml) at 37°C and 5% CO<sub>2</sub>. The MOLM1 cells were purchased from the Hayashibara Institute, and the MC3T3-E1 and HNT34 cells were purchased from the RIKEN cell bank. The 293T cells were cultured in high-glucose DMEM supplemented with 10% fetal calf serum, and the MC3T3-E1 cells were cultured in MEM alpha supplemented with 10% fetal calf serum.

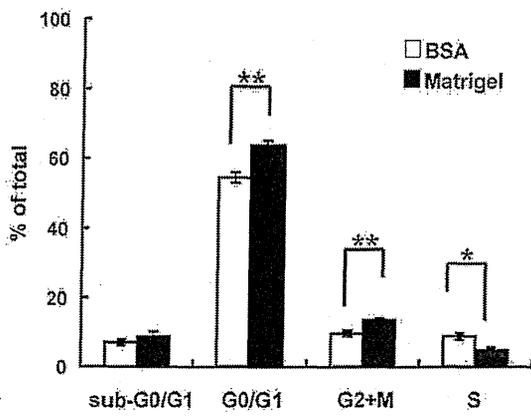
### Oligonucleotide microarray

This experimental method was previously described [24].

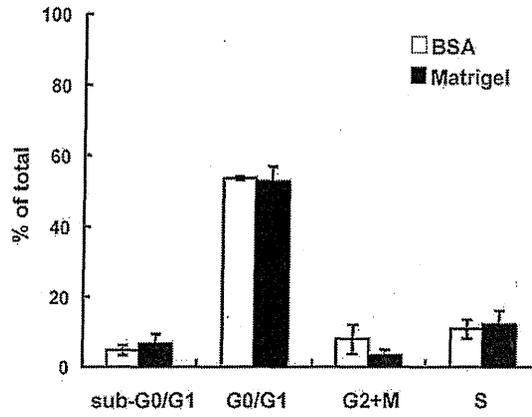
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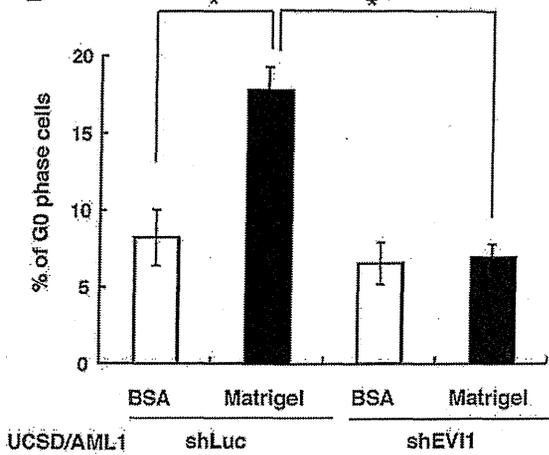
**B** AML1/shLuc



**C** AML1/shEVI1



**D**



**Figure 6. Decreased cell growth with increased cell population in G0-phase of EVI1<sup>high</sup> AML cells cultured on the matrigel-coated plates.** **A.** The UCSD/AML1 cells were transfected with shEVI1 (AML1/shEVI1) or control shLuc vector (AML1/shLuc) to determine their cell growth under normal culture conditions on BSA or matrigel-coated plates. **B and C.** The cell cycle of AML1/shLuc as a control (**B**) and AML1/shEVI1 (**C**) were analyzed using BD FACSCalibur after double staining with BrdU-APC and 7-AAD. The percentage of viable cells in each cell cycle is indicated with white (BSA-coated) and black (matrigel-coated) bars. **D.** The percentage of AML1/shEVI1 (black bars) and AML1/shLuc (white bars) cells in the G0 phase cultured on matrigel or BSA-coated plates were analyzed using BD FACSCalibur after double staining with Ki67-Alexa647 and 7-AAD. Each experiment was performed in triplicate, and the experiments were independently repeated at least three times. The results are shown as the mean  $\pm$  S.E. The statistical analysis was performed using the Student's *t*-test (\*\* $p < 0.01$ ; \*\* $p < 0.05$ , vs. control). doi:10.1371/journal.pone.0030706.g006

#### Adhesion-mediated drug resistance (CAM-DR) in AML cell lines

The UCSD/AML1, MOLM1, PT9 and PT11 cells were plated onto tissue culture plates with BSA or matrigel-coated wells and subsequently incubated with  $10^{-3}$  to  $10^{-7}$  M cytosine-arabioside (Ara-C) for 48 h. The number of viable cells was determined using trypan blue exclusion.

#### Neutralizing Antibody

The neutralizing antibodies used included rat anti-ITGA6 (Santa Cruz) and mouse anti-ITGB2, ITGB3 and ITGB4 (Millipore).

#### PCR and PCR primers

Total cellular RNA was isolated using Trizol (Invitrogen), and the cDNA was prepared using oligo(dT) primer and reverse transcriptase (SuperScript; Invitrogen). The polymerase chain reaction (PCR) was performed using Ex Taq (Takara-Bio) on a GeneAmp 2400 machine (Applied Biosystems). The primers for all integrin family members, EVI1 and b-actin were produced based on methods used in previous studies [12], [31]. The primers for N and VE-cadherin were produced based on the Quantitative PCR Primer Database (<http://lpgws.nci.nih.gov/cgi-bin/PrimerViewer>). The primers for the laminin family genes were produced based on Primer Bank (<http://pga.mgh.harvard.edu/primerbank/>).

#### Introduction of small hairpin RNAs for EVI1, ITGA6, ITGB1 and ITGB4 in UCSD/AML1 cells

The pSIREN-retroQ-ZsGreen plasmid (Takara-Bio) was used to inhibit EVI1 expression, and expression of ZsGreen (green fluorescent protein) was used as a marker. The following sequences were cloned into the BamHI-EcoRI sites of the pSIREN-retroQ-ZsGreen plasmid to create shRNAs against human EVI1 [32], 5'-GATCGTCTAAGGCTGAAGTCAAGAGACTGCTAGTTCAGCCTTAGATTTTTG-3'; human ITGA6, 5'-GATGTCCTTTTCAGGTTTCAGTAGTTATTCAAGAGAA-TAGTTACTGAATCTGAGAGGTTTTG-3'; human ITGB1, 5'-GATCTGGAGGGTTGTTTCGGGTTTCATTCAAGAGATGAAGTCCGAAGTAATCCTCCTTTTTG-3'; and human ITGB4, 5'-GATCGCCAGCGACTACACTATTGGATCTCG-AGATCCAATAGTGTAGTCGCTGGTTTTG (MISSION shRNA library, Sigma). We used the pSIREN-retroQ-ZsGreen-shLuc plasmid containing an shRNA against firefly luciferase (Takara-Bio) as a control. The retroviral particles were generated using the p10A1 packaging vector (Takara-Bio) via the transient transfection of 293T cells. The transfection was conducted using Hilymax liposome transfection reagent (Dojin). For retroviral transduction,  $1 \times 10^6$  cells were plated onto a 6-cm dish containing 5 ml retroviral supernatant containing 100 ng/ml of polybrene for 24 h. After two weeks, the ZsGreen-positive cells were sorted using a JSAN cell sorter (Bay Bioscience). The reduction of EVI1 or ITGB4 expression was confirmed using RT-PCR.

#### Establishment of stable U937 cell lines expressing EVI1

The pGCDNsam-EVI1-IRES-EGFP construct was kindly provided by Dr. A. Iwama (Chiba University, Chiba, Japan). To produce recombinant retrovirus, the plasmid DNA was transfected into 293GP cells along with the vesicular stomatitis virus G (VSV-G) expression plasmid via CaPO<sub>4</sub> precipitation. For retroviral transduction,  $1 \times 10^3$  U937 cells were plated onto 96-well flat-bottomed plates and infected for 24 h with either an EVI1 retroviral supernatant (pGCDNsam-EVI1-IRES-EGFP) or a control retroviral supernatant (pGCDNsam-IRES-EGFP) containing 100 ng/ml of polybrene. After 7 days, the green fluorescent protein (GFP)-positive U937 cells were sorted using a JSAN cell sorter (Bay Bioscience, Kobe, Japan).

#### Laminin-332 (laminin-5) knockdown in MC3T3-E1 cells

The pSIREN-retroQ-ZsGreen plasmid (Takara-Bio) was used to inhibit the expression of the  $\alpha 3$  chain of laminin-332, and the expression of ZsGreen (GFP) was used as a marker. The following sequence was cloned into the BamHI-EcoRI sites of the plasmid to create an shRNA against the  $\alpha 3$  chain of murine laminin-332: 5'-GATCTGAACTCCTTAAATGATTATGAATTCAAGAG-ATTTCATAATCATTAAAGGAGTCTTTTTTACGCGTG-3'. The target sequence for the knockdown was determined using the BLOCK-iT RNAi Designer (<http://www.invitrogen.com/rnai>). We used pSIREN-retroQ-ZsGreen-shLuc containing shRNA against firefly luciferase (Takara-Bio) as a control. We transiently introduced the laminin-332 or luciferase shRNA expression vector into MC3T3-E1 cells using an Amaxa Nucleofector (Lonza) and determined the expression levels of the  $\alpha 3$  and  $\alpha 2$  subunits of laminin-332.

#### Cell proliferation and cell viability assays

A cell proliferation assay was performed using a Cell Counting Kit-8 (Dojin) according to the manufacturer's protocol. Briefly,  $5 \times 10^3$  cells/well were plated onto a 96-well culture plate and incubated for 1–3 days. Approximately 10 ml of Cell Counting Kit-8 reagent was added to the plate, and the absorbance at 450 and 620 nm was measured using an Immunomini NJ2300 plate reader (Nunc). A cell viability assay was performed using the Cell Counting Kit-8 reagent according to the manufacturer's protocol. For this assay,  $5 \times 10^3$  cells were plated onto a 96-well culture plate containing Ara-C ( $1 \times 10^{-6}$  M), rat anti-ITGA6 (20 mg/ml) or mouse anti-ITGB4 (20 mg/ml). The cells were incubated for 1–2 days and 10 ml of Cell Counting kit-8 reagent was added. The absorbance at 450 and 620 nm was measured. The cell viability was calculated by dividing the value obtained for the treatment well by the average value obtained for the control wells. For cell viability assay, cells were seeded in  $5 \times 10^4$  cells/ml to matrigel coated culture plate. Cells were co-cultured with Ara-C ( $1 \times 10^{-6}$  M) and anti-ITGA6 antibody (20 mg/ml) or anti-ITGB4 (20 mg/ml) or isotype control IgG (20 mg/ml). After incubation for 1 to 4 days, living cell numbers were counted by trypan blue exclusion. The cell viability was calculated by dividing the value for the treatment well by the value for the control wells. Each

experiment was performed in triplicate. Data are shown as mean  $\pm$  S.E. Statistical analysis was performed using Student's *t*-test.

#### Cell adhesion assay

Four types of extracellular matrices (ECM; matrigel, fibronectin, laminin, collagen) and BSA (as a control) were used for the cell adhesion assay. A culture plate was coated overnight with a 1/40 dilution of Growth Factor Reduced-Matrigel (BD Falcon), 5 mg/cm<sup>2</sup> of fibronectin (BD Falcon), 5 mg/cm<sup>2</sup> of laminin (BD Falcon), 0.3 mg/ml of collagen (Nitta Gelatin) or 5% w/v BSA (Nacalai Tesque). After coating with ECM,  $1 \times 10^5$  cells/ml were transferred to each pre-coated well and incubated for 24 h at 37°C. After washing two times with phosphate buffered saline (PBS), the floating and adherent cells were counted using trypan blue, and the number of adherent cells was divided by the number of floating cells to calculate the cell adherence ratios. In the neutralization experiments, the cells were pre-incubated with 20 mg/ml of anti-ITGA6, ITGB2, ITGB3 or ITGB4 antibodies prior to plating on the matrigel [33].

#### FACS analysis

The intensity of ITGB3, ITGB4 and ITGA6 expression in UCSD/AML1 and U937 cells was analyzed using FACSCalibur (BD Bioscience). The cells were stained with phycoerythrin (PE)-labeled anti-human ITGA6 (Clone: GoH3), ITGB4 (Clone: 58XB4) or ITGB3 (BI-PL2) antibodies purchased from Biologend, Inc.

#### BrdU (Bromodeoxyuridine)/7-AAD (7-Aminoactinomycin D) staining

To assess the percentage of cells in sub G0/G1, G0/G1, and G2/M and S phase after 48 h of culture on matrigel or BSA-coated plates,  $1 \times 10^6$  cells from each culture were fixed in BD Cytotfix/Cytoperm buffer for 20 min., followed by a 300-mg/ml DNase treatment for 30 min. The cells were subsequently washed twice in BD Perm/Wash Buffer and stained with anti BrdU-APC (BD Biosciences/PharMingen, San Jose, CA) for 15 minutes, followed by staining with 7-AAD (BD Biosciences/PharMingen, San Jose, CA). The cells were harvested and analyzed using FACSCalibur with FL-4 (for BrdU-APC) and FL-3 (for 7-AAD) settings.

#### Ki67/7AAD staining

To assess the percentage of cells in G0 phase after 48 h of culture on matrigel or BSA-coated plates,  $1 \times 10^6$  cells from each culture were fixed in 70% ethanol for 2 h, followed by a 2.5  $\mu$ g/ml RNase treatment for 30 minutes. The cells were subsequently washed twice in PBS/1% fetal bovine serum (FBS) and stained with mouse anti-human Ki67 (BD Biosciences/PharMingen, San Jose, CA) and Alexa647-conjugated anti-mouse IgG (Invitrogen) for 30 minutes, followed by staining with 7-AAD (BD Biosciences/PharMingen, San Jose, CA). The cells were harvested and analyzed using FACSCalibur with FL-4 (for Ki67-Alexa647) and FL-3 (for 7-AAD) settings.

#### Statistical analysis

Student's *t*-test was used to compare differences between the two groups.

### Supporting Information

**Figure S1 Expression of integrin genes and cell binding ability of various myeloid leukemia cells. A.** The

expression of 25 members of the integrin family was detected using semi-quantitative RT-PCR in HL60, U937, and K562 (cell lines with EVI1<sup>low</sup> expression) and in UCSD/AML1, MOLM1, and HNT34 (cell lines with EVI1<sup>high</sup> expression). The expression of EVI1 and b-actin in these cells is also shown. **B.** The expression of 18 genes in the integrin family was detected using RT-PCR in AML1/shLuc and AML1/shEVI1 cell lines. **C and D.** The adhesion ability of various HNT-34 cell lines (Pt, parental; shLuc, transfection with small hairpin RNA for firefly luciferase; shEVI1, transfection with small hairpin RNA for EVI1) to bind to matrigel is shown in C, and the expression of integrin genes in conjunction with EVI1 and b-actin is shown in D. **E and F.** The ability of PT9-related (PT9/shLuc and PT9/shEVI1) and PT11-related (PT11/shLuc and PT11/shEVI1) cell lines to bind to matrigel was measured (E), and the expression of various integrin genes in these four cell lines was determined using RT-PCR (F). **G.** The expression patterns of integrin genes were determined in U937/GFP and U937/EVI1 cell lines in conjunction with b-actin and EVI1. (DOC)

**Figure S2 Expression of various laminin chains in MC3T3-E1 cells.** The relative expression levels of laminin a chains 1 to 5, laminin b chains 1 to 3, and laminin g chains 1 to 3 in MC3T3-E1 cells were determined using semi-quantitative RT-PCR. (DOC)

**Figure S3 Dose-response functions for AraC against four EVI1<sup>high</sup> AML cells.** UCSD/AML1, MOLM1, PT9 and PT11 cells were incubated in BSA- or matrigel-coated wells of tissue culture plates and subsequently incubated with 10-3 to 10-7 M cytosine-arabinoside (Ara-C) for 48 h. Relative cell viability is calculated as a percentage relative to standard controls. Data are shown as mean  $\pm$  S.E. Statistical analysis was performed using Student's *t*-test. A star (\*) indicates  $p < 0.05$  and double stars (\*\*\*) indicate  $p < 0.01$ . (DOC)

**Figure S4 Drug sensitivity of EVI1<sup>high</sup> and EVI1<sup>low</sup> leukemia cells cultured with or without MC3T3-E1 cells determined by treatment with VP-16 or Ara-C.** UCSD/AML1 (EVI1<sup>high</sup>) leukemia cells and HL60 (EVI1<sup>low</sup>) leukemia cells were treated with Ara-C or VP-16 for three days under plastic flasks (open diamonds) or co-cultured with MC3T3-E1 cells (closed squares); the viable cells were counted at each indicated time point. The percent cell viability compared to the number of untreated cells is shown for each indicated day. A star (\*) indicates  $p < 0.05$ . (DOC)

**Figure S5 Decreased cell growth with increased cell population in G0-phase of U937 cells with EVI1 expression cultured on the matrigel-coated plates.** A. U937 cells were introduced by EVI1 expression vector (U937/EVI1) or control vector (U937/GFP) to determine their cell growths with culture condition on BSA or matrigel-coated plates. B and C. Cell cycle of U937/GFP as a control (B) and U937/EVI1 (C) were analyzed by BD FACSCalibur after double-stained by BrdU-APC and 7-AAD. Percentages of each cell cycles were shown by white bars (BSA-coated) and black bars (matrigel-coated). D. The percentage of cells in G0 phase in U937/EVI1 (black bars) and U937/GFP cells (white bars) cultured on matrigel- or BSA-coated plates were analyzed by BD FACSCalibur after double-stained by Ki67-Alexa647 and 7-AAD. Each experiment was performed in triplicate, and experiments were independently repeated at least

three times. Results are shown as mean  $\pm$  S.E. Statistical analysis was performed using Student's t-test (\*\* $p < 0.01$ ; \*\* $p < 0.05$ , vs control).

(DOC)

**Figure S6 Expression profiles of ITGA6 and ITGB4 in AML patients.** A and B. The expression patterns of ITGA6 (A) and ITGB4 (B) are shown as gene expression profiles for ten AML patients with EVI1low and ten AML patients with EVI1high expression (<http://www.ncbi.nlm.nih.gov/geo>, accession number GSE6891 [NCBI GEO]). C and D. The expression patterns of

ITGA6 (C) and ITGB4 (D) are shown as gene expression profiles for patients in remission and for AML patients who had relapsed (<http://www.ncbi.nlm.nih.gov/geo>, accession number GDS1059 [NCBI GEO]).

(DOC)

## Author Contributions

Conceived and designed the experiments: KM NY. Performed the experiments: NY KK YS. Analyzed the data: EI YS KM. Contributed reagents/materials/analysis tools: EI YS. Wrote the paper: KM NY KK.

## References

- Morishita K, Parker DS, Mucenski ML, Jenkins NA, Copeland NG, et al. (1988) Retroviral activation of a novel gene encoding a zinc finger protein in IL-3-dependent myeloid leukemia cell lines. *Cell* 54(6): 831–840.
- Mucenski L, Taylor BA, Ihle JN, Hartley JW, Morse HC, 3rd, et al. (1988) Identification of a common ecotropic viral integration site, Evi-1, in the DNA of AKXD murine myeloid tumors. *Mol Cell Biol* 8(1): 301–308.
- Morishita K, Parganas E, Bartholomew C, Sacchi N, Valentine MB, et al. (1990) The human Evi-1 gene is located on chromosome 3q24–q28 but is not rearranged in three cases of acute nonlymphocytic leukemias containing t(3;5)(q25;q34) translocations. *Oncogene Res* 5(3): 221–231.
- Barjesteh van Waalwijk van Doorn-Khosrovani S, Erpelinck C, van Putten WL, Valk PJ, van der Poel-van de Luytgaarde S, et al. (2001) High EVI1 expression predicts poor survival in acute myeloid leukemia: a study of 319 de novo AML patients. *Blood* 101(3): 837–845.
- Lugthart S, van Drunen E, van Norden Y, van Hoven A, Erpelinck CA, et al. (2008) High EVI1 levels predict adverse outcome in acute myeloid leukemia: prevalence of EVI1 over-expression and chromosome 3q26 abnormalities underestimated. *Blood* 111(8): 4329–4337.
- Morishita K (2007) Leukemogenesis of the EVI1/MEL1 gene family. *Int J Hematol* 85(4): 279–286.
- Goyama S, Kurokawa M (2010) Evi-1 as a critical regulator of leukemic cells. *Int J Hematol* 91(5): 753–757.
- Kurokawa M, Mitani K, Irie K, Matsuyama T, Takahashi T, et al. (1998) The oncoprotein Evi-1 represses TGF- $\beta$  signalling by inhibiting Smad3. *Nature* 394(6688): 92–96.
- Nitta E, Izutsu K, Yamaguchi Y, Imai Y, Ogawa S, et al. (2005) Oligomerization of Evi-1 regulated by the PR domain contributes to recruitment of corepressor CtBP. *Oncogene* 24: 6165–6173.
- Palmer S, Brouillet JP, Kilbey A, Fulton R, Walker M, et al. (2001) Evi-1 transforming and repressor activities are mediated by CtBP co-repressor proteins. *J Biol Chem* 276(25): 834–840.
- Yuasa H, Oike Y, Iwama A, Nishikata I, Sugiyama D, et al. (2005) Oncogenic transcription factor Evi1 regulates hematopoietic stem cell proliferation through GATA-2 expression. *EMBO J* 24(11): 1976–1987.
- Shimahara A, Yamakawa N, Nishikata I, Morishita K (2010) Acetylation of lysine 564 adjacent to the C-terminal binding protein-binding motif in EVI1 is crucial for transcriptional activation of GATA2. *J Biol Chem* 285(22): 16967–16977.
- Lvesque JP, Helwani FM, Winkler IG (2010) The endosteal 'osteoblastic' niche and its role in hematopoietic stem cell homing and mobilization. *Leukemia* 24(12): 1979–1992.
- Wu JY, Scadden DT, Kronenberg HM (2009) Role of the osteoblast lineage in the bone marrow hematopoietic niches. *J Bone Miner Res* 24(5): 759–764.
- Chan JY, Watt SM (2001) Adhesion receptors on hematopoietic progenitor cells. *Br J Haematol* 112(3): 541–557.
- Zhang J, Niu C, Ye L, Huang H, He X, et al. (2003) Identification of the hematopoietic stem cell niche and control of the niche size. *Nature* 425(6960): 836–841.
- Zaidel-Bar R, Geiger B (2010) The switchable integrin adhesome. *J Cell Sci* 123(9): 1385–1388.
- Desgrosellier JS, Cheresch DA (2010) Integrins in cancer: biological implications and therapeutic opportunities. *Nat Rev Cancer* 10(1): 9–22.
- Papayannopoulou T, Craddock C, Nakamoto B, Priestley GV, Wolf NS (1995) The VLA4/VCAM-1 adhesion pathway defines contrasting mechanisms of lodgement of transplanted murine hemopoietic progenitors between bone marrow and spleen. *PNAS* 92(21): 9647–9651.
- Papayannopoulou T, Nakamoto B (1993) Peripheralization of hemopoietic progenitors in primates treated with anti-VLA4 integrin. *PNAS* 90(20): 9374–9378.
- Matsunaga T, Takemoto N, Sato T, Takimoto R, Tanaka I, et al. (2003) Interaction between leukemic-cell VLA-4 and stromal fibronectin is a decisive factor for minimal residual disease of acute myelogenous leukemia. *Nat Med* 9(9): 1158–1165.
- Matsunaga T, Fukai F, Miura S, Nakane Y, Owaki T, et al. (2008) Combination therapy of an anticancer drug with the FNIII4 peptide of fibronectin effectively overcomes cell adhesion-mediated drug resistance of acute myelogenous leukemia. *Leukemia* 22(2): 353–60.
- Becker PS, Kopecky KJ, Wilks AN, Chien S, Harlan JM, et al. (2009) Very late antigen-4 function of myeloblasts correlates with improved overall survival for patients with acute myeloid leukemia. *Blood* 113(4): 866–874.
- Saito Y, Nakahata S, Yamakawa N, Kaneda K, Ichihara E, et al. (2011) CD52 as a molecular target for immunotherapy to treat acute myeloid leukemia with high EVI1 expression. *Leukemia* 25(4): 921–31.
- Bergamini C, Sgarra C, Trerotoli P, Lupo L, Azzariti A, et al. (2007) Laminin-5 stimulates hepatocellular carcinoma growth through a different function of  $\alpha 6 \beta 4$  and  $\alpha 3 \beta 1$  integrins. *Hepatology* 46(6): 1801–1809.
- Fortunel NO, Otu HH, Ng HH, Chen J, Mu X, et al. (2003) Comment on "Stemness": transcriptional profiling of embryonic and adult stem cells" and "a stem cell molecular signature". *Science* 302(5644): 393.
- Papayannopoulou T, Priestley GV, Nakamoto B, Zafiroopoulos V, Scott LM (2001) Molecular pathways in bone marrow homing: dominant role of  $\alpha 4 \beta 1$  over  $\beta 2$ -integrins and selectins. *Blood* 98(8): 2403–2411.
- Scott LM, Priestley GV, Papayannopoulou T (2003) Deletion of  $\alpha 4$  integrins from adult hematopoietic cells reveals roles in homeostasis, regeneration, and homing. *Mol Cell Biol* 23(24): 9349–9360.
- Verhaak RG, Wouters BJ, Erpelinck CA, Abbas S, Beverloo HB, et al. (2009) Prediction of molecular subtypes in acute myeloid leukemia based on gene expression profiling. *Haematologica* 94(1): 131–134.
- Yagi T, Morimoto A, Eguchi M, Hibi S, Sako M, et al. (2003) Identification of a gene expression signature associated with pediatric AML prognosis. *Blood* 102(5): 1849–1856.
- Lin KT, Yeh SH, Chen DS, Chen PJ, Jou YS (2005) Epigenetic activation of  $\alpha 4$ ,  $\beta 2$  and  $\beta 6$  integrins involved in cell migration in trichostatin A-treated Hep3B cells. *J Biomed Sci* 12(5): 803–813.
- Liu Y, Chen L, Ko TC, Fields AP, Thompson EA (2006) Evi1 is a survival factor which conveys resistance to both TGF- $\beta$  and taxol-mediated cell death via PI3K/AKT. *Oncogene* 25(25): 3565–3575.
- Bei L, Lu Y, Bellis SL, Zhou W, Horvath E, et al. (2007) Identification of a HoxA10 Activation Domain Necessary for Transcription of the Gene Encoding beta 3 integrin during Myeloid Differentiation. *J Biol Chem* 282(23): 16846–16859.



# Cancer Research

## HTLV-1 bZIP Factor Suppresses Apoptosis by Attenuating the Function of FoxO3a and Altering Its Localization

Azusa Tanaka-Nakanishi, Jun-ichirou Yasunaga, Ken Takai, et al.

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