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Tailored immunoconjugate therapy depending on a quantity of tumor stroma

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The purpose of this study was to clarify the appropriate combination of targeting antibody and conjugate-design of anti-tumor immunoconjugate depending on a quantity of tumor stroma. Most human solid tumors including pancreatic cancer (PC) forming hypovascular and stroma-rich tumor hinders the penetration of monoclonal antibodies (mAbs) into the cells, and that leads to failure of the conventional cell-targeting immunoconjugate strategy. To overcome this drawback, SN-38 as topoisomerase 1 inhibitor was conjugated to a mAb to collagen 4, a plentiful component of the tumor stroma via ester-bond. The immunoconjugate, which was able to release SN-38 in physiological condition outside the cells, was effective to stroma-rich PC-tumor. On the other hand, anti-CD 20 mAb-PEG-SN-38 via carbamate-bond as conventional immunoconjugate, enabled SN-38 to be released by a carboxylesterase inside of the tumor cell following the internalization, showed strong anti-tumor activity against malignant lymphoma as hypervascular and stroma-poor tumor. The conjugate-design, in parallel with the choice of targeting antibodies, should be selected to maximize the therapeutic effect in each individual tumor having a distinct stromal structure. (*Cancer Sci* 2013; 104: 231–237)

Monoclonal antibody (mAb), which can target the tumor cell actively by the specific binding ability against corresponding antigen, easily extravasates from leaky tumor vessels but not from normal vessels, is long retained in the tumor by using active targeting and passive targeting based on the enhanced permeability and retention (EPR) effect.^(1–4) Therefore, numerous mAbs have been developed and conjugated with anticancer agents (ACAs) or toxins to create an “immunoconjugate strategy”.^(5–8) Recent examples of the conjugates include anti-CD33 immunoconjugate-calicheamicin and anti-CD20 radiolabeled immunoconjugate, were effective to hematological malignancy such as malignant lymphoma and leukemia.⁽⁵⁾ Heterogeneity of the tumor cells, however, prevents development of the immunoconjugate chemotherapy based on cell-specific antigen.^(9–12) Moreover, conventional immunoconjugates depend on cleavage of conjugation site with intracellular biochemical (enzymatic) process after the cell-uptake of the conjugate.^(13–16) In addition to such annoying characteristics of cancer cells themselves, most human solid tumors such as pancreatic cancer and gastric cancer, possess abundant stroma that hinders the distribution of mAbs (Fig. 1a).^(17–20) To overcome these drawbacks, we developed a unique strategy whereby the cancer-stromal targeting (CAST) therapy by cytotoxic immunoconjugate bound to the collagen 4 or fibrin network in the tumor stroma, from which the payload released gradually and distributed throughout the tumor, resulting in the arrest of tumor growth due to induced damage to tumor cells and tumor vessels.^(21,22) Besides, there

have been a few reports describing tumor stromal targeting immunoconjugates, a mAb against a cell surface antigen FAP as fibroblast targeting therapy, or a mAb against fibronectin for the targeting of tumor vascular endothelial cell in photodynamic therapy.^(23,24) However, the merits and drawbacks of anti-stromal targeting immunoconjugate therapy in relation to the conjugate-design and the amount of tumor stroma have not yet been fully elucidated.

The purpose of this study was to clarify the appropriate combination of targeting antibody and conjugate-design of anti-tumor immunoconjugate depending on the quantity of tumor stroma. Hence, we selected two types of conjugate linker: ester-bond and carbamate-bond. We hypothesized that a combination of anti-stromal targeting mAb and a linker composed of ester-bond to release ACA outside the cells would be effective against the stroma-rich cancer. Conversely, anti-cancer cell targeting via carbamate-bond to release ACA inside the cells would be effective against stroma-poor cancer. It seemed that the outcome of immunoconjugate therapy against each individual tumor having distinct stromal structure was dependent on the selection of conjugation-design, as well as targeting mAb.

Materials and Methods

Antibodies and cells. Anti-EpCAM (B8-4) and Anti-collagen 4 antibody (35-4) were prepared as previously reported.⁽²¹⁾ Anti-human CD20 antibody (rituximab) was purchased from Daiichi-Sankyo (Tokyo, Japan). Human malignant lymphoma cell line RL was purchased from the American Type Culture Collection (Rockville, MD, USA). Human PC cell line SUIT2 was purchased from the Health Science Research Resources Bank (Osaka, Japan).

In vivo imaging and immunohistochemistry. Immunohistochemistry was conducted using anti CD31 antibody (R&D Systems, Minneapolis, MN, USA), anti-collagen 4 antibody and anti-CD20 (rituximab), or anti-EpCAM antibody as first antibodies, Alexa 488-, 555- or 647-labeled anti-human, mouse, rat or goat IgG (Invitrogen, Carlsbad, CA, USA) as second antibodies.

For mouse-systemic *in vivo* imaging or tracking of antibody in the tissue, IRDye 800 (Li-Cor Biosciences, Lincoln, NE, USA) alexa-647 (Invitrogen) or Qdot 625 (Invitrogen) labeled antibodies were injected into the mice tail vein at 100 µg/body. Fluorescence images were obtained using OV110 (Olympus, Tokyo, Japan), BZ-9000 (Keyence, Osaka, Japan), LSM 710 (Carl Zeiss, Jena, Germany).

Immunoconjugate. The detailed process of chemical synthesis is shown in the Data S1. The final structure was composed of one maleimide for attachment of mAb, one PEG₁₂ (MW 865) spacer and one PEG₂₇ (MW 1422) ester-bond or

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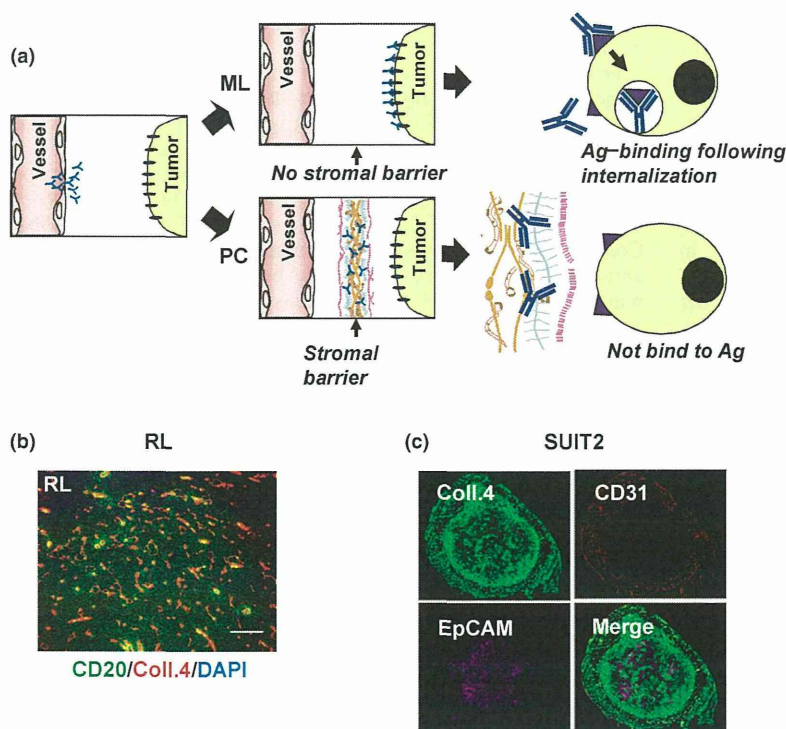


Fig. 1. The difference of tumor tissue stromal component as stromal barrier between malignant lymphoma and pancreatic cancer. (a) The schema of antibody delivery into the tumor cells. In the tumor having no stromal barrier like malignant lymphoma (ML), antibodies were delivered into the cancer cells, and can be internalized after antigen-binding. However, many human solid tumors including pancreatic cancer (PC) possess stromal barrier hindering the distribution of the immunoconjugates into cancer cells such that antigen-binding following antibody-internalization never occur. Ag, Antigen. (b) RL-tumor (ML) was stained with anti-CD20 (green), anti-collagen 4 (red) mAb and 4',6'-diamidino-2-phenylindole dihydrochloride (DAPI) (blue). Scale bar: 100 μ m. (c) SUIT2-tumor (PC) was stained with anti-EpCAM (purple), anti-collagen 4 (green) and anti-CD31 (red) mAb. Co-existence of collagen 4 and CD 31 (yellow in Merge). Coll., collagen.

carbamate-bond for attachment of one SN-38 molecule. Inter-chain disulfides of the antibodies were reduced with 10 mM DTT (Sigma).⁽¹⁴⁾ The numbers of free thiols were quantified with dinitrothiocyanobenzene (DNTB, Wako, Osaka, Japan). Reduced antibodies were reacted with maleimide-linker-SN-38 prodrugs in PBS containing 5 mM EDTA (pH 6) at room temperature for 1 h, then at 4°C overnight. The concentration of antibody-prodrug conjugates was determined using the Bradford method (Bio-Rad Protein Assay, 500-0006JA, Bio-Rad, Hercules, CA, USA). The numbers of residual thiols were quantified with dNTP. Each drug (SN-38)/antibody ratio was determined by comparing the numbers of free and residual thiols. In the characterization of the conjugates, statistical analysis was performed using Student's *t*-test.

Animal model and anti-tumor effects. Female BALB/c nude mice (5 weeks old) were purchased from SLC Japan (Shizuoka, Japan). Mice were inoculated subcutaneously in the flank with 5×10^6 cells of RL, or 2×10^6 of SUIT2. The length (L) and width (W) of tumor masses and body weight were measured every 4 days, and tumor volume was calculated using $(L \times W^2)/2$. All animal procedures were performed in compliance with the Guidelines for the Care and Use of Experimental Animals established by the Committee for Animal Experimental of the National Cancer Center. These guidelines meet the ethical standards required by law and also comply with the guidelines for the use of experimental animals in Japan. When the mean tumor volume reached approximately 140 mm³ (RL) and approximately 70 mm³ (SUIT2), mice were randomly divided into groups consisting of five mice. Immunoconjugates were administered on day 0 by the mice tail vein injection. The injection doses of antibody-SN-38 prodrug equal to an SN-38 dose of 3 mg/kg were determined by calculations based on drug (SN-38)/antibody ratio (range from 2629.08 to 3296.16 mg SN-38 per 1 mM antibody) for each drug. Statistical analysis was performed using ANOVA.

Biochemistry and hematological examination. Blood samples were taken from the healthy mice at 7 days after i.v. administration of immunoconjugates (at an equivalent SN-38

dose of 3 mg/kg). Hemograms were measured by using an auto-analyzer Celltac α MEK6358 (Nihon Kohden, Tokyo, Japan), and blood chemistry examinations were carried out by Nagahama LSL (Shiga, Japan).

Anti-collagen antibody induced arthritis. Female DBA/1J mice (5 weeks old) were purchased from SLC Japan. Anti-collagen 2 antibody (Chondrex, Redmond, WA, USA) or anti-collagen 4 antibody (clone 35-4, the same mAb in the immunoconjugate) were intraperitoneally administered on day 0 at 2 mg. Fifty micrograms of LPS (Chondrex) was intraperitoneally injected on day 3.

Results

Difference of tumor stromal component between malignant lymphoma and pancreatic cancer. We first examined the difference of the stromal component influencing the drug delivery between malignant lymphoma RL and pancreatic cancer SUIT2. Anti-CD20- or anti-EpCAM-mAb, which is specific to lymphoma or epithelial carcinoma, respectively, was used as cancer cell-specific mAb.^(5,25) Anti-collagen 4 mAb was prepared to evaluate the stromal component. RL tumor consisted of CD20-positive tumor cells and collagen-4-positive blood vessels, which was stained fine-linearly but not interspersed-fibrously like the intercellular-stroma (Fig. 1b). On the other hand, SUIT2-tumor reported as the histopathology relatively resembling original human pancreatic cancer,^(21,26) consisted of EpCAM-positive cancer-cells and collagen-4-positive extracellular component, the latter was composed of both CD31-positive blood vessel wall (yellow in Merge, Fig. 1c) and high amount of CD31-negative stroma (green in Merge, Fig. 1c).

Internalization and biodistribution of mAbs against malignant lymphoma or pancreatic cancer. Cell-uptake of fluorescent anti-CD20- or anti-EpCAM-mAb against RL cells or SUIT2 cells was evaluated, respectively. Anti-CD20 mAb was internalized and colocalized with intracellular lysosome in RL cells at 12 h after the incubation. On the other hand, anti-EpCAM mAbs, the majority of which was still retained on cell-surface

membrane of SUI2 cells, was poorly internalized at the same period (Fig. 2a). We next investigated the kinetics of entry of three mAbs into the tumors using an *in vivo* imaging system with near infrared fluorescence, which can provide deep tissue imaging with high fidelity.^(27,28) Until day 3, all fluorescent-mAbs were delivered and retained both in RL-tumor, and SUI2-tumor, indicating passive targeting (Fig. 2b). On Day 7, anti-EpCAM mAb in RL-tumor and anti-CD20 mAb in SUI2-tumor as a negative control, were almost eliminated, but anti-CD20 mAb in RL-tumor, anti-EpCAM mAb in PC-tumor and anti-collagen 4 mAb in both RL-tumor and SUI2-tumor were still retained in each tumor, indicating active targeting, that is, binding to their respective antigens within the lesion. There were also clear differences in the degree of intratumor accumulation among three mAbs. Anti-CD20 mAb accumulation was higher in RL-tumors than anti-EpCAM mAb in SUI2-tumors (Fig. 2b). On the other hand anti-collagen 4 mAb accumulation was higher in SUI2-tumors than in RL-tumors (Fig. 2b). We then examined the histological distribution of mAbs in each tumor. In RL tumor, fluorescent anti-CD20 mAb was distributed in the whole tumor area and bound to the cancer cells (Fig. 2c). Qdot-labeling system detecting lower fluorescent signals in SUI2-tumor was conducted to evaluate the biodistribution of each mAb. The quantity of anti-CD20 mAb observed in SUI2 tumor was small (Fig. 2d). Anti-EpCAM mAb was observed mainly around the tumor cell-abundant area, in which collagen 4 was negative (Fig. 2d). In contrast, anti-collagen 4 mAb was mainly observed in collagen 4-positive stroma, and rarely in the tumor cell-abundant area (Fig. 2d). Thus, we succeeded in preparing three mAbs: anti-CD 20 mAb for cell-targeting against RL tumor, anti-EpCAM mAb for cell-targeting against SUI2-tumor, and anti-collagen 4 mAb for stroma-targeting against both tumors. These three mAbs can selectively exit the vascular system through the leaky tumor vessels and

distribute within each tumor according to the nature of tissue component.

Preparation and characterization of cell-targeting or stroma-targeting Immunoconjugate-PEG-SN-38 via a carbamate-bond or ester-bond. To specify the appropriate immunoconjugate therapy against malignant lymphoma or pancreatic cancer, we prepared two types of the conjugates, one being mAb-PEG-SN-38 via a carbamate-bond⁽²⁹⁾ (Fig. 3a) and another being mAb-PEG-SN-38 via an ester-bond^(21,22) (Fig. 3b). Consequently, six types of immunoconjugates, anti-CD20, anti-EpCAM or anti-collagen 4 mAb- SN-38 via a carbamate-bond or an ester-bond, were obtained. The average number of conjugated SN-38 per one mAb (drugs/mAb), ranging from 7.0 to 8.5, was shown in Figure 3c. There was no clear loss of antigen-binding activity of each mAb after the conjugation (Fig. 3D). An *in vitro* release experiment, both bonds can be cut by a carboxylesterase localized in cytoplasm to release SN-38 inside various cells (Fig. 3e). However, in physiological conditions (non-enzymatic hydrolysis), the immunoconjugate prepared via an ester-bond can release SN-38 gradually and effectively. In contrast, the immunoconjugate via a carbamate-bond cannot release SN-38 effectively in the conditions outside the cells (Fig. 3e). We then evaluated the release profiles of SN-38 from both types of immunoconjugate in mouse blood, which contained high amounts of carboxylesterase.⁽³⁰⁾ *In vivo* analysis of mouse plasma, the concentration of unbound SN-38 or bound and unbound of SN-38 from the immunoconjugate via an ester-bond or a carbamate-bond at 72 h after the mice tail vein injection were shown. Most of the immunoconjugates in the mouse blood were protected from the enzymatic cleavage (Fig. 3f). Next, we examined the difference between carbamate-bond and ester-bond in combination with cell-targeting or stromal-targeting antibody by the cytotoxicity assay. In RL cells, anti-CD 20 immunoconjugate via carbamate-bond showed strong cytotoxicity

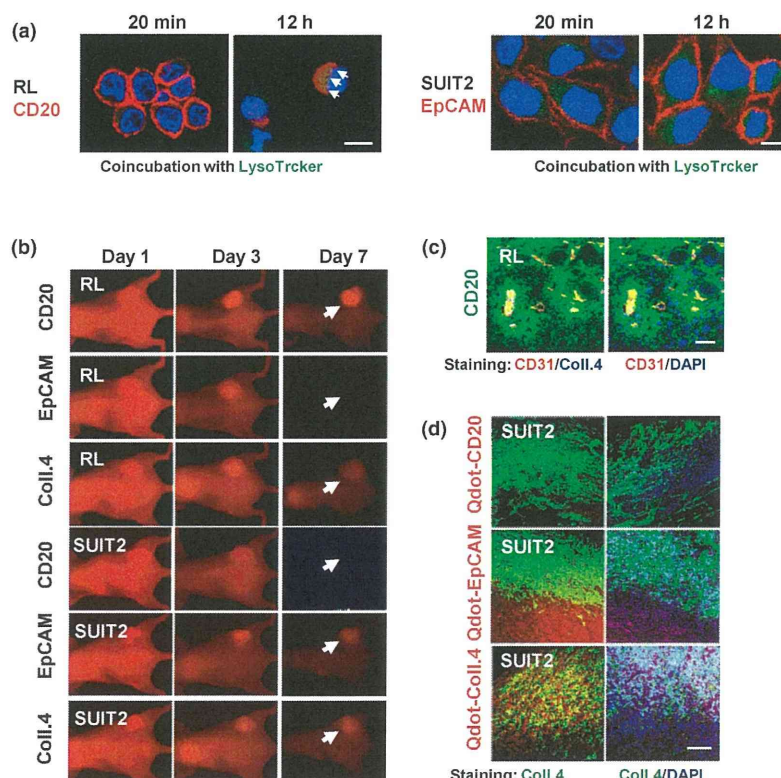


Fig. 2. Internalization and biodistribution of anti-CD20, anti-EpCAM monoclonal antibody (mAb) and anti-collagen 4 mAb. (a) Internalization of Alexafluor-633-anti-CD20 or anti-EpCAM mAb (Red) on RL or SUI2 cells was examined at 20 min or 12 h after the incubation. Arrow shows merged yellow as co-localization with lysostracker (Green). Scale bar: 10 μ m. (b) *In vivo* imaging analysis of RL-, SUI2-tumor was conducted using near-infrared labeled anti-CD20, EpCAM and collagen 4 mAbs on days 1, 3 and 7 after injection. Arrows indicate each tumor position. (c) The intra-RL-tumor distribution of Alexa 647 (green)-labeled anti-CD20 mAb was examined by confocal laser scanning microscopy at 24 h after injection. CD31 (red), Collagen 4 (blue, left panel) and nuclei (blue, right panel) were stained by immunohistochemistry or DAPI. Scale bar: 100 μ m. (d) The intra-SUI2-tumor distribution of each Qdot (red)-labeled antibody was examined by confocal laser scanning microscopy on day 7 after injection. Collagen 4 (green) or nuclei (blue) were stained by immunohistochemistry or DAPI. Scale bar: 100 μ m.

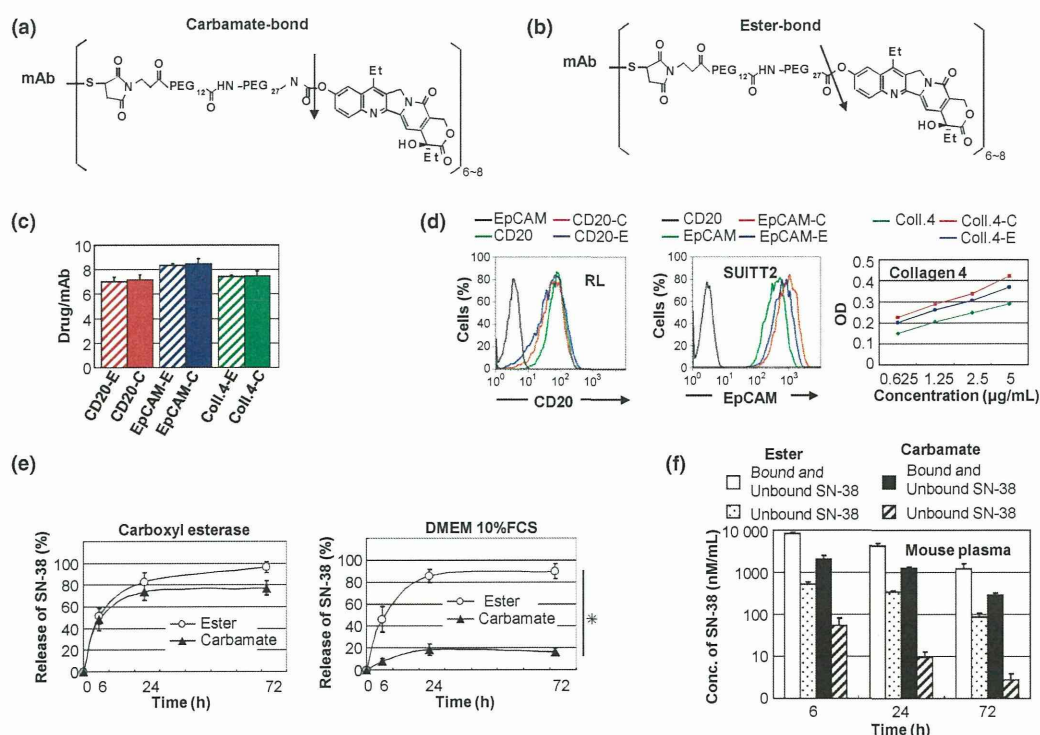


Fig. 3. Preparation and characterization of two types of immunoconjugates-PEG-SN-38 via carbamate-bond and ester-bond. (a,b) Drug design of two types of immunoconjugates; monoclonal antibody (mAb) -PEG-SN-38 via carbamate-bond (a) and mAb-PEG-SN-38 via ester-bond (b). One antibody bears six to eight molecules of SN-38. The arrow indicates the cleavage site for releasing free active SN-38. (c) The average number of conjugated SN-38 per one mAb was shown ($n = 3$). Bar = standard deviation (SD). (d) Antigen-binding activity of the mAb before and after the conjugation was shown. Anti-CD 20 and EpCAM mAb were examined by fluorescence-activated cell sorting (FACS) analysis using RL cells and SUI2 cells, respectively. Anti-collagen 4 mAb was examined by enzyme linked immunosorbent assay (ELISA) using purified protein. (e) *In vitro* release of SN-38 from two types of immunoconjugates in carboxylesterase-containing solution (left) and Dulbecco's modified eagle medium (DMEM) 10% fetal calf serum (FCS) (right) ($n = 3$). Bar standard deviation (SD), $*P < 0.05$. (f) Concentration of bound and unbound form of SN-38, and unbound form of SN-38 from two types of immunoconjugates in the mouse plasma at 6, 24, 72 h after the mice tail vein injection, were shown ($n = 3$). Concentrations of SN-38 were determined by high performance liquid chromatography (HPLC). Bar = standard deviation (SD).

compared with anti-CD 20 immunoconjugate via ester-bond significantly. In SUI2 cells, although there was no significant difference, anti-EpCAM immunoconjugate via carbamate-bond had a lower tendency in the cytotoxic effect compared to anti-EpCAM immunoconjugate via ester-bond. Anti-collagen 4 immunoconjugate via ester-bond showed higher cytotoxic activity than anti-collagen 4 immunoconjugate via carbamate-bond in both cells significantly (Table 1). These results indicated that a carbamate-bond was useful for the immunoconjugate linker to work inside of the cells and an ester-bond to work outside of the cells.

Cell-targeting or stroma-targeting immunoconjugate-PEG-SN-38 via carbamate-bond or ester-bond differs drastically in their anti-tumor effects depending on tumor stromal component in mice. Three mAbs conjugated with SN-38 via carbamate-bond or ester-bond (administered once, at an equivalent SN-38 dose of 3 mg/kg) were evaluated in order to know their anti-tumor effects in RL (CD20-positive stroma-poor human malignant lymphoma), SUI2 (EpCAM-positive stroma-rich human pancreatic tumor). In RL lymphomas, cell-targeting anti-CD20 mAb- SN-38 via carbamate-bond showed superior anti-tumor activity compared to anti-CD20 mAb- SN-38 via ester-bond after the treatment (Fig. 4a). Stroma-targeting anti-collagen 4 mAb- SN-38 via ester-bond showed significant superior anti-tumor activity as compared to saline as control, but inferior to anti-CD20 mAb- SN-38 via carbamate-bond (Fig. 4a). On the contrary to RL tumor, in SUI2 tumor, the most potent anti-tumor activity was obtained by the stroma-

Table 1. IC₅₀ of free SN-38 and SN-38 conjugated to monoclonal antibody (mAb) (immunoconjugate) for malignant lymphoma and pancreatic cancer cell lines (WST-8 assay)

Malignant lymphoma cell lines	Free SN-38	SN-38 conjugated to mAb	
		CD20	Collagen 4
		Ester vs carbamate	Ester vs carbamate
RL	4.6 ± 3.7	8.7 ± 2.9 vs 2.1 ± 1.0*	34 ± 17 vs 90 ± 30*
Pancreatic cancer cell lines	Free SN-38	SN-38 conjugated to mAb	
		EpCAM	Collagen 4
		Ester vs carbamate	Ester vs carbamate
SUI2	7.8 ± 3.6	24 ± 13 vs 15 ± 9	29 ± 15 vs 75 ± 22*

IC₅₀ (50% cell survival) (nM), Mean ± standard deviation ($n = 3$), $*P < 0.05$.

targeting anti-collagen 4 mAb- SN-38 via ester-bond (Fig. 4b). However, there was no significant difference in anti-tumor activity between anti-EpCAM mAb- SN-38 via carbamate-bond and via ester-bond, whereas the anti-tumor activity of anti-collagen 4 mAb- SN-38 via carbamate-bond was inferior to that of anti-collagen 4 mAb- SN-38 via ester-bond (Fig. 4b). These results clearly indicated that, in

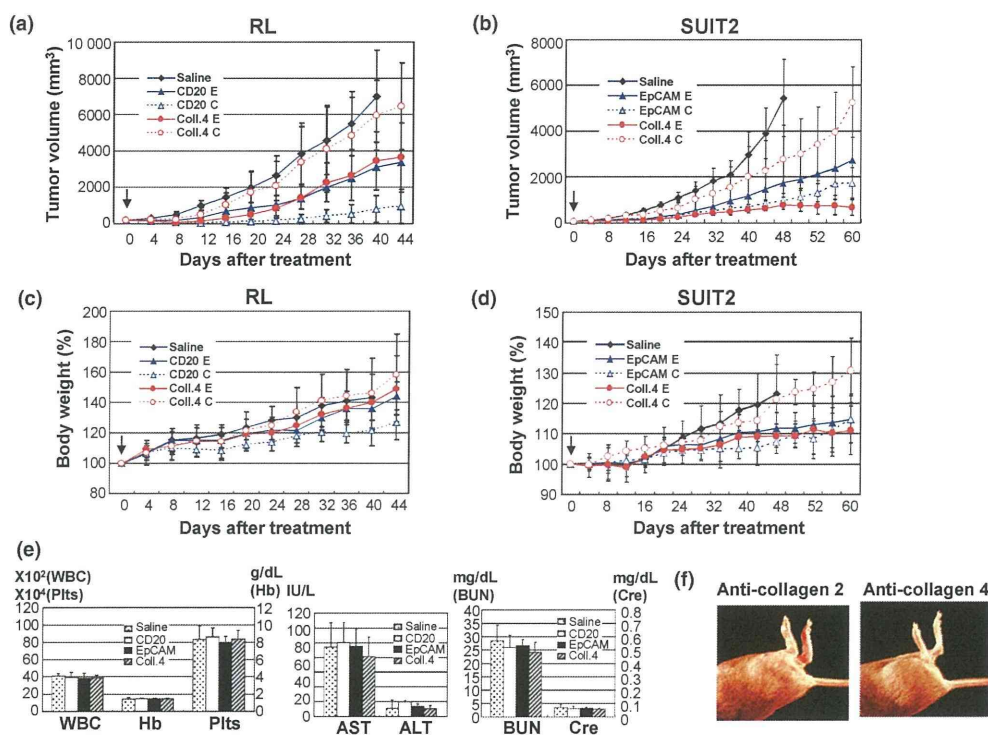


Fig. 4. Antitumor effects and toxicities of immunoconjugates-PEG-SN-38 in the combinations of anti-cell or anti-stroma targeting, carbonate-bond or ester-bond. (a,b) Anti-tumor activities and (c,d) percent changes of body weight were examined. In animal models of RL (a,c) and SUI2 (b,d), the six types of immunoconjugates (combined anti-CD20 monoclonal antibody (mAb) =CD20, anti-EpCAM mAb=EpCAM or anti-collagen 4 mAb=Coll.4 and ester-bond=E or carbamate-bond=C), or saline as control, were administered once at an equivalent SN-38 dose of 3 mg/kg to separate groups of mice ($n = 5$) by intravenous bolus injection to the mice on day 0. Arrows indicate day of administration and the curves illustrate the effect of treatment on tumor size. $P < 0.0001$ (saline versus CD20-E or CD20-C, CD20-C versus CD20-E, Coll.4-E or Coll.4-C in RL tumor; saline versus EpCAM-E, EpCAM-C or Coll.4-E, Coll.4-E versus EpCAM-C or Coll.4-C, EpCAM-C versus Coll.4-C in SUI2 tumor), $P < 0.001$ (Saline versus Coll.4-E in RL tumor; saline versus Coll.4-C, Coll.4-E versus EpCAM-E in SUI2 tumor). Bar = standard deviation (SD). (e) Hematological (WBC, Hb and Plts) and biochemical (aspartate aminotransferase [AST], alanine aminotransferase [ALT], blood urea nitrogen [BUN] and creatinine [Cre]) examination were conducted at 7 days after i.v. administration of immunoconjugates via ester-bond or saline as control. Bar = standard deviation (SD). (f) Anti-collagen antibody induced arthritis in DBA/1J mice. The arthritis was admitted on day 7 only after the administration of anti-collagen 2 (left) but not anti-collagen 4 (right) antibodies.

stroma-poor solid tumors like malignant lymphoma, cytotoxic immunoconjugate should target to the tumor cell surface and ACA should be conjugated to mAb through carbamate-bond, which can be specifically cut by a carboxylesterase inside the tumor cell after the internalization. On the other hand, in stroma-rich tumors, the immunoconjugate should target to the stroma within tumor tissue and ACA should be attached to the mAb via ester-bond, which can be cut gradually outside the tumor cell following the accumulation of the cytotoxic immunoconjugate in the tumor stroma. It is remarkable that the feature of tumor stromal component influence the outcome of the two types of immunoconjugation drugs, cell-targeting mAb-PEG-SN-38 via carbamate-bond, or stroma-targeting mAb-PEG-SN-38 via ester-bond.

Regarding normal tissue distribution and elimination of antibodies and SN-38, there was no difference among immunoconjugates on day 7 after the administration. The dose in this study did not cause significant toxicity as shown by the change of mouse body weight (Fig. 4c,d). Moreover, there was no hepatotoxicity, nephrotoxicity, or bone marrow toxicity in mice treated with all three immunoconjugates as compared to controls (Fig. 4e). In addition, no autoimmune disease-like adverse effects such as arthritis and nephritis were observed in the administration of anti-collagen 4 mAb, whereas anti-collagen 2 mAb combined with lipopolysaccharide caused severe arthritis (Fig. 4f).

Discussion

Conventional immunoconjugate is composed of cell-targeting mAb, ACA as payload and linker for the conjugation. The linker technology is an important part of the immunoconjugate strategy, and various linkers have been exploited to date. Among them, acid labile hydrazine linkage, thiol reduction of disulfide linkers, and lysosomal peptidase proteolysis of peptide linkers were favorably applied to ensure stability in blood.⁽⁶⁻⁸⁾ For these types of linkers, cell-mediated endocytosis of antibody (antibody-internalization) and intracellular biochemical (enzymatic) processing of the immunoconjugate were indispensable to make the active ACA work. Our carbamate-bond based linker, which is used in a clinically approved anticancer prodrug CPT-11 to release an active component SN-38 within the tumor cell but not in blood circulation^(29,31,32) can be classified into the conventional type mentioned above. Anti-CD 20 Immunoconjugate-PEG-SN-38 via carbamate-bond showed strong anti-tumor activity against malignant lymphoma, in which the distribution within the tumor tissue and antibody-internalization into tumor cells occur effectively. Although there were negative reports concerning the internalization of anti-CD20 mAbs, several authors, recently demonstrated internalization of anti-CD20 mAbs including rituximab in malignant lymphoma and leukemia cells.⁽³³⁻³⁶⁾ These conflicting results might reflect

the differences of cell types, mAbs or methodologies used.^(35,36) In contrast to malignant lymphoma, most human solid tumors possess abundant stroma that hinders the tissue-distribution of antibodies.^(17–22) Cell–cell interaction between malignant epithelial cells also inhibits the penetration of mAb besides tumor-stroma.^(18,37) Moreover, heterogeneity of the cells in the tumor prevents development of immunoconjugate therapy based on cancer cell-specific antigen.^(9–12) This led us to design an anti-stromal targeting immunoconjugate strategy using the tumor stroma both as a scaffold for binding and assembling immunoconjugates and as a relay base for a second attack by payload-ACA persistently released from the scaffold.^(21,22) In this drug design we selected a specially selected linker using ester-bond, which can release SN-38 in physiological condition (non-enzymatic hydrolysis) outside the cells. Both ester-bond and carbamate-bond were concerned to be cleaved by plasma carboxyl-esterase in the circulation after the injection. Cleavages of our conjugates were very low in mouse plasma, which has much higher levels of carboxyl-esterase activity than in human.⁽³⁰⁾ Recently, we conducted clinical trials of NK012, a SN-38 incorporating polymeric micelle. In this formulation, SN-38 was conjugated to poly-Glu-chain via ester-bond. From these trials, we learned that human blood also contains high amounts of carboxylesterase. Nevertheless, NK012 proved good stability in human blood circulation.^(38,39)

Anti-collagen 4 immunoconjugate exiting from vessel can bind to the outer vessel wall and cells surrounding the stroma. Anti-collagen 4 immunoconjugate via carbamate-bond was not useful because it can scarcely release SN-38 outside the cells. On the other hand, anti-collagen 4 immunoconjugate via ester-bond can release SN-38 on the stroma. Low molecular weight agent SN-38 can penetrate through stroma into the cells. SN-38 released from the scaffold of adjacent collagen-4-positive vascular wall also attack tumor endothelium.⁽²¹⁾ Anti-collagen 4 mAb-SN-38 via ester-bond exerted more potent antitumor activity compared to anti-EpCAM mAb-SN-38 via carbamate-bond or ester-bond. It is too complicated to explain these data. However, we speculate that in addition to the insufficient attainability of anti-EpCAM mAb to tumor cells by stromal barrier and its low internalization into the cells, the retention of anti-EpCAM mAb within the tumor cell lesion is lower than that of anti-collagen 4 mAb within the tumor stroma. Consequently, the amount of SN-38 released inside of the cells from anti-EpCAM mAb-SN-38 via carbamate-bond or outside of the cells from anti-EpCAM mAb-SN-38 via ester-bond, may be less than that of SN-38 released outside of the cells from anti-collagen 4 mAb-SN-38 via ester-bond.

Although there had been a concern about the influence of anti-collagen 4 immunoconjugate on normal tissues having high level of collagen 4, we observed the safety of the immunoconjugate in several mouse models. We think that cancer stromal targeting (CAST) therapy is dependent on the fundamental concept that antibodies or immunoconjugates are generally too large to pass through the normal vessel walls, whereas they can extravasate from leaky tumor vessels to achieve tumor selective targeting by using EPR effect and bind to collagen 4, a plentiful component of the tumor stroma.^(1–4,40) We also speculate that such a passive targeting effect is one of the reasons why recent anti-EGFR antibody therapies show no serious adverse effects in spite of high level EGFR expression in normal tissues including intestinal mucosa, dermis and others.^(9,10,41)

In general, human cancer is classified into three types according to the tissue component. One is hypervascular stroma-poor tumor such as malignant lymphoma, the second is hypovascular stroma-rich tumor such as pancreatic cancer and stomach cancer, and the third is intermediated tumor between

the two types such as breast cancer and colorectal cancer. We thus propose the new therapeutic strategy of immunoconjugates to the feature of individual tumor as tissue stromal component: (i) cell-targeting mAb conjugated with ACAs via carbamate-bond for hypervascular and stroma-poor tumor; (ii) stroma-targeting mAb conjugated with ACAs via ester-bond for hypovascular and stroma-rich tumor; (iii) both cell-targeting immunoconjugate via carbamate-bond and stroma-targeting via ester-bond for intermediated type of tumor (Fig. 5).

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Disclosure Statement

The authors have no conflict of interest.

Design and application of cytotoxic immunoconjugates

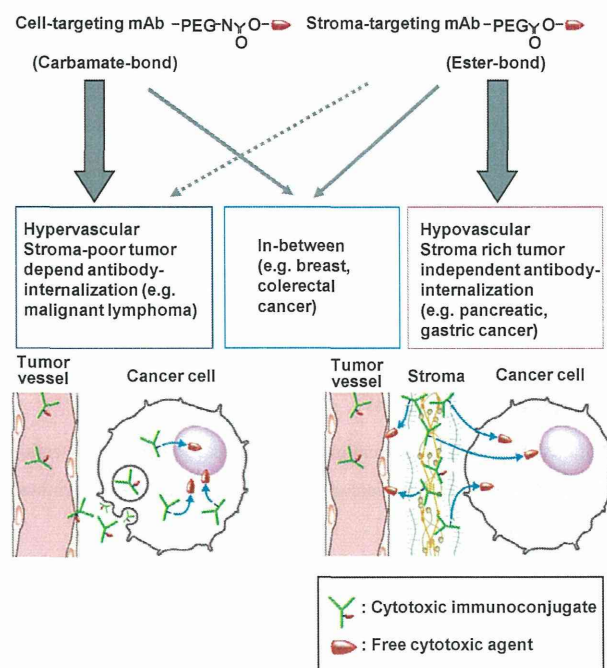


Fig. 5. Diagram of Immunoconjugate strategy to tumor tissue component and characteristic of cancer-cells. Design and application of cytotoxic immunoconjugates. SN-38 conjugated cell-targeting monoclonal antibody (mAb) via carbamate-bond is suitable for hypervascular, stroma-poor tumor dependent antibody-internalization. SN-38 conjugated stroma-targeting mAb via ester-bond is suitable for hypovascular, stroma-rich tumor independent antibody-internalization.

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Supporting Information

Additional Supporting Information may be found in the online version of this article:

Data S1. Detailed methods including linker-SN-38 derivative synthesis.



Gd-DTPA-loaded polymer–metal complex micelles with high relaxivity for MR cancer imaging

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ABSTRACT

Nanodevices for magnetic resonance imaging of cancer were self-assembled to core–shell micellar structures by metal complex formation of K_2PtCl_6 with diethylenetriaminepentaacetic acid gadolinium (III) dihydrogen (Gd-DTPA), a T_1 -contrast agent, and poly(ethylene glycol)-*b*-poly{N-[N'-(2-aminoethyl)-2-aminoethyl]aspartamide} (PEG-*b*-PAsp(DET)) copolymer in aqueous solution. Gd-DTPA-loaded polymeric micelles (Gd-DTPA/m) showed a hydrodynamic diameter of 45 nm and a core size of 22 nm. Confining Gd-DTPA inside the core of the micelles increased the relaxivity of Gd-DTPA more than 13 times ($48 \text{ mm}^{-1} \text{ s}^{-1}$). In physiological conditions Gd-DTPA/m sustainedly released Gd-DTPA, while the Pt(IV) complexes remain bound to the polymer. Gd-DTPA/m extended the circulation time in plasma and augmented the tumor accumulation of Gd-DTPA leading to successful contrast enhancement of solid tumors. μ -Synchrotron radiation-X-ray fluorescence results confirmed that Gd-DTPA was delivered to the tumor site by the micelles. Our study provides a facile strategy for incorporating contrast agents, dyes and bioactive molecules into nanodevices for developing safe and efficient drug carriers for clinical application.

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1. Introduction

Early detection of neoplastic lesions is critical for success in cancer therapy. Magnetic resonance imaging (MRI) provides a powerful diagnostic imaging modality of cancer, because of its non-invasiveness, high definition and precise three-dimensional positioning ability [1,2]. Paramagnetic compounds are widely used as MRI contrast agents (CAs) to amplify the signals of MRI tomography and improve the contrast between magnetic similar but histological dissimilar tissues [3,4]. Several low molecular weight paramagnetic complexes of gadolinium(III) are used for clinical MRI as they can decrease the longitudinal relaxation time

T_1 of surrounding water protons [5–7]. Among these complexes, Gd-DTPA, one of the most commonly used T_1 -contrast agent for clinical MRI, is featured by high thermodynamic and kinetic stabilities, which reduce the release of toxic Gd^{3+} ions, besides a longitudinal relaxivity, r_1 , of $\sim 3.5 \text{ mm}^{-1} \text{ s}^{-1}$ at 1.4 T [3,5,7]. Nevertheless, the inherent low relaxivity, short circulation time in blood and low specificity to tissues limit its applications. From this viewpoint, high relaxivity CAs with promoted tissue detection and hindered release of free Gd^{3+} ions are required to improve the performance of imaging.

Nanodevices have been recently developed to selectively deliver imaging agents to solid tumors by leaking out from circulation due to the enhanced permeability of the tumor vasculature [8–12]. In this way, macromolecules, liposomes, dendrimers and nanoparticles carrying high payloads of CAs have been developed to enhance the MR contrast of solid tumors [13–25]. In most cases, Gd-based CAs were covalently incorporated to the structures of nanocarriers to increase the bloodstream circulation, the tumor accumulation and, in some cases, the relaxivity of the loaded CAs

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[14,22,23]. Nevertheless, covalent binding of Gd-chelates to macromolecular systems may increase the risk of Gd^{3+} ion leakage from the chelates due to the prolonged half-life in the body and cumulative toxicity [26]. Thus, the design of tumor-targeted nanodevices for Gd-based CAs should consider such issues for constructing safe carriers, which could target the tumor position and release Gd-DTPA easily from nanodevices.

Among promising nanodevice systems for MRI, block copolymeric micelles, i.e. nanostructures consisting of a drug-loaded core and poly(ethylene glycol) (PEG) protective shell, present exceptional advantages including their relatively small sizes, ability to engineer drug loading mechanisms, controlled release of their cargo, prolonged life in the bloodstream and enhanced accumulation in solid tumors after intravenous administration [19–25]. Auspicious results from clinical trials of micelle formulations incorporating anticancer agents indicate the potential clinical application of polymeric micelles [27,28].

The aim of this study was to develop core-shell type micellar nanocarriers of Gd-DTPA for tumor imaging, which can incorporate Gd-DTPA via the reversible Pt-Gd-DTPA complexation, allowing sustained release of free Gd-DTPA in biological environments. In order to incorporate hydrophilic low molecular weight paramagnetic complexes Gd-DTPA into polymeric micelle, we utilized Pt(IV) ions to “cross-link” carboxyl groups of Gd-DTPA to amino groups of PEG-*b*-PAsp(DET) copolymers. Gd-DTPA/m could passively accumulate in solid tumors due to the enhanced permeability and retention (EPR) effect. Thus, the improved tumor accumulation of Gd-DTPA could enhance the T_1 weighed contrast of the malignancies in mice. Otherwise, in physiological environments such as plasma, the ligand exchange reaction of Pt(IV) from the carboxylic group of Gd-DTPA to chloride ions results in sustained release of free Gd-DTPA, which can be rapidly cleared from the body via glomerular excretion.

2. Materials and methods

2.1. Materials

α -Benzyl-L-aspartate *N*-carboxy anhydride (BLA-NCA) was obtained from Chuo Kaseihin Co., Inc. (Tokyo, Japan). Poly(ethylene glycol) (MeO-PEG-NH₂) ($M_w = 12,000$, $M_w/M_n = 1.03$) were purchased Nippon Oil and Fats Co., Ltd. (Tokyo, Japan). Benzene, *N,N*-dimethylformamide (DMF), *n*-butylamine, dichloromethane (CH₂Cl₂), diethylenetriamine (DET), potassium hexachloroplatinate(IV) (K₂PtCl₆) and *N*-methyl-2-pyrrolidone (NMP) were purchased from Wako Pure Chemical Industries (Osaka, Japan) and distilled by a general method before use. Arsenazo III and Gd-DTPA were purchased from Aldrich Chemical (Milwaukee, USA), and then Gd-DTPA was converted to sodium salt by adjusting the pH to 7 with NaOH and lyophilization. 400-mesh copper grids were obtained from Nisshin E. M. Spectra/ Por-6 membrane was purchased from Spectrum Laboratories (Rancho Dominguez, CA). Dulbecco's modified eagle's medium (DMEM) was obtained from Sigma-Aldrich Co. (St. Louis, USA). Fetal bovine serum (FBS) was purchased from MP Biomedicals, Inc. (Illkirch, France). 96-well culture plates were purchased from Becton Dickinson Labware (Franklin Lakes, USA). The Cell Counting Kit-8 was purchased from Dojindo Laboratories (Kumamoto, Japan).

Murine colon adenocarcinoma 26 (C-26) cells were kindly supplied by the National Cancer Center (Tokyo, Japan). Human umbilical vein endothelial cells (HUVEC), and the endothelial cell growth medium-2 bullet kit (EGM-2 bullet kit) were obtained from Lonza Ltd. (Basel, Switzerland). B16-F10 melanoma cells were purchased from the American Type Culture Collection (Virginia, USA). All the cells are maintained with medium in a humidified atmosphere containing 5% CO₂ at 37 °C. CDF₁ mice (female; 18–20 g body weight; 6 weeks old) were purchased from Charles River Japan (Kanagawa, Japan) and, treated following the policies of the Animal Ethics Committee of the University of Tokyo.

2.2. Synthesis of PEG-*b*-PAsp(DET)

As shown in Scheme S1, poly(ethylene glycol)-*b*-poly(β -benzyl L-aspartate) (PEG-*b*-PBLA) with different degree of polymerization (DP) (DP = 25, 35, 45) were synthesized by ring-opening polymerization of BLA-NCA initiated by MeO-PEG-NH₂ ($M_w = 12,000$) according to previously reported method with minor modification [29]. Briefly, BLA-NCA (1.1 g, 4.5 mmol) was dissolved in DMF (2 mL) and then diluted with CH₂Cl₂ (20 mL). MeO-PEG-NH₂ (1.2 g, 0.1 mmol) dissolved in CH₂Cl₂

was added to the solution of BLA-NCA. The reaction was carried out with stirring for 2 days at 35 °C. All the procedures above were under dry argon protection. PEG-*b*-PBLA was collected by precipitating in excess amount of diethyl ether, filtration and vacuumed to dry. In Figure S1, the ¹H NMR spectrum of PEG-*b*-PBLA was measured in *d*₆-DMSO at 80 °C with a JEOL EX300 spectrometer (JEOL, Tokyo, Japan) using tetramethylsilane (TMS) as an internal standard, and the degree of polymerization (DP = 25, 35, 45) was calculated from the peak intensity ratio of protons both in phenyl groups of PBLA (C₆H₅CH₂–, $\delta = 7.3$ ppm) and methylene units (–CH₂CH₂–, $\delta = 3.6$ ppm) of PEG. The polydispersity of PEG-*b*-PBLA was characterized by gel permeation chromatography (GPC) system (HLC-8220, TOSOH Co., Japan) equipped with TSK-gel columns (TOSOH Co., Tokyo, Japan) and an internal refractive index (RI) detector at 40 °C. NMP containing 50 mM LiBr was used as mobile phase with a flow rate of 0.35 mL/min and linear PEG standards were used for calibration. As shown in Figure S2, the GPC chromatogram of the prepared PEG-*b*-PBLA has a unimodal molecular weight distribution (M_w/M_n : 1.03).

PEG-*b*-PAsp(DET) with different DP (25, 35, 45) were prepared separately via aminolysis reaction of PEG-*b*-PBLA with DET following the protocol modified from our former reported method [30]. Briefly, lyophilized PEG-*b*-PBLA (200 mg, 0.01 mmol) was dissolved in NMP (2 mL) and maintained at 5 °C. DET (2.3 mL, 50-equivalent to BLA) was diluted with NMP (2.3 mL) and kept at 5 °C. PEG-*b*-PBLA was added to DET solution and reacted with stirring at 5 °C for 1 h. The reacted mixture was neutralized by 5 M HCl aqueous solution (13 mL, equivalent to the added amino groups) in an ice bath, then dialysis (MWCO: 6000–8000) against 0.01 M HCl aqueous solution, then dialyzed with Milli-Q water under 4 °C. PEG-*b*-PAsp(DET) as the chloride salt form was collected via freeze-dry of final solution. ¹H NMR (D₂O) measurement showed the disappearance of signals belonging to the benzyl group and all the signals featured with PEG-*b*-PAsp(DET) (Figure S3). Degree of polymerization (DP = 25, 35, 45) was calculated by comparing the peak intensity ratio between methylene protons of the α , β -Asp segment (peak f, $\delta = 2.8$ ppm) and methylene units (–CH₂CH₂–, $\delta = 3.6$ ppm) of PEG. GPC measurements of block copolymers were carried out utilizing a HPLC system (JASCO, Japan) equipped with Superdex 75 10/300 GL column (GE Healthcare UK, Ltd.) and UV detector set at 220 nm. The column was eluted with 10 mM AcOH buffer containing 500 mM NaCl (pH 7.4) at a flow rate of 0.5 mL/min at room temperature. The GPC chromatogram of the obtained PEG-*b*-PAsp(DET) was unimodal (Figure S4).

2.3. Preparation of Gd-DTPA-loaded micelles (Gd-DTPA/m)

K₂PtCl₆ (0, 1, 2.5, 5 and 10 mM) was dissolved in Milli-Q water adjusted to pH 7.4 using trace amount of NaOH or diluted in 5 mM PBS buffers, then mixed with Gd-DTPA (5 mM) to incubated at 37 °C for 24 h. PEG-*b*-PAsp(DET) (DP = 25, 35, 45; [DET] = 5 mM) was added to Pt/Gd-DTPA mixture ([Gd-DTPA]/[DET] = 1.0) and incubated at 4 °C for 120 h to spontaneously form Gd-DTPA loaded micelles (Gd-DTPA/m). The micelle was purified by dialysis (MWCO: 6000–8000) against Milli-Q water and ultrafiltration (MWCO: 30,000) to remove free drug and polymer in the solution. Size distribution of the Gd-DTPA/m was evaluated by dynamic light scattering (DLS) measurement at 25 °C using Zetasizer Nano ZS90 (Malvern Instruments, UK). The contents of platinum and gadolinium of micelles were determined by inductively coupled plasma-mass spectrometry (ICP-MS) (4500 ICP-MS; Hewlett Packard, USA).

2.4. Transmission electron microscopy (TEM)

The morphology of micelle was observed on a Transmission Electron Microscope (JEM-1400, JEOL, Japan) operated with 100 kv acceleration voltages and 40 uA beam current. Diluted micelle was stained by mixing with uranyl acetate solution (2%, w/v) and placed on 400-mesh copper grids. The diameters of the core of micelle and size distribution were calculated with Image J, which was designed and provided by National Institutes of Health (NIH). The elements distribution inside micelle was characterized using JEM-2100F (JEOL, Japan) under the scanning transmission electron microscopy (STEM) pattern and scanned by energy dispersive X-ray (EDX) spectra. The specimen for the elements measurements was prepared only loading micelle on 400-mesh copper grids without staining.

2.5. Arsenazo III colorimetric assay

Free Gd³⁺ was determined by the absorption of Gd-Arsenazo III complex referring one arsenazo III method [31]. Briefly, equal molar ratio of arsenazo III solution was mixed with of Gd-DTPA/K₂PtCl₆ mixtures, and then the absorbance spectra were measured using with an UV–vis spectrometer (V-570 UV/VIS/NIR Spectrophotometer, JASCO, Japan) at 660 nm. The absorbance of arsenazo III/GdCl₃, arsenazo III/Gd-DTPA, and arsenazo III were measured. All the solutions above were maintained at pH 7.4.

2.6. Release rate of Gd-DTPA from Gd-DTPA/m

The release characterization of Gd-DTPA from the micelle was evaluated by a dialysis method. 1 mL of Gd-DTPA/m solution and 1 mL PBS buffer (20 mM PBS with 300 mM NaCl) were put into dialysis bag (MWCO: 6000–8000) and