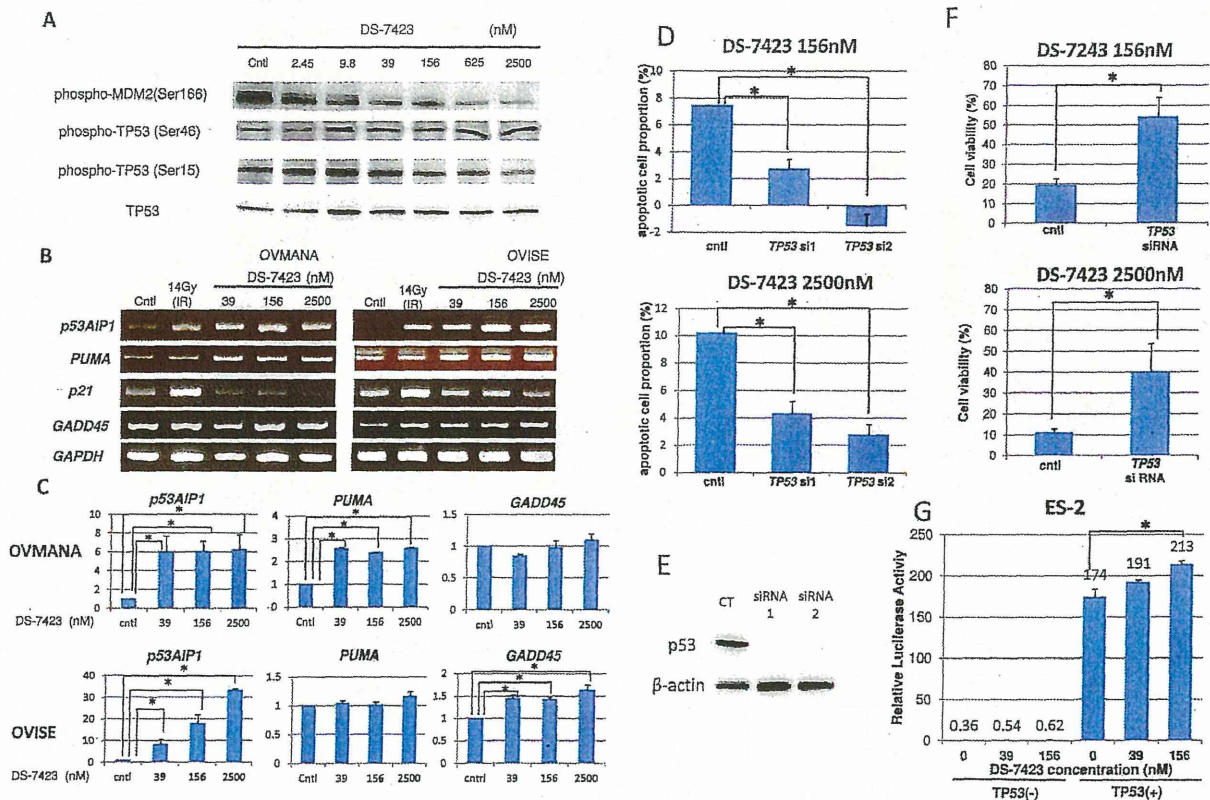


**Figure 5. DS-7423-mediated induction of apoptosis in ovarian clear cell adenocarcinoma cell lines.** (A) All nine OCCA cells were treated with DS-7423 at 156 or 2,560 nM for 48 h, and apoptotic cell proportion was evaluated using annexin-V fluorescein isothiocyanate (FITC) and propidium iodide (PI) double staining, followed by analysis using flow cytometry. The experiments were repeated 3 times, and each value is shown as the mean of 3 experiments  $\pm$  standard deviation (SD). (B) The apoptotic cells were calculated using flow cytometry by counting the cell population in the right boxes. The example shown (OVISE cells) is representative of the results obtained for all the cell lines tested. (C) The proportion of cells rendered apoptotic by exposure to DS-7423 at 156 nM and 2,560 nM was significantly higher in OCCA cells without mutations in TP53 than in OCCA cells that carry mutations in TP53. (D) Cleaved poly(ADP-ribose) polymerase (PARP) induction was evaluated by immunoblotting in OVISE cells. OVISE cells were treated with DS-7423 at 156 nM for the times indicated (left) or for 4 h at the doses indicated (right). doi:10.1371/journal.pone.0087220.g005

effect at the higher concentrations tested (>40 nM), rapamycin suppressed cell proliferation even at lower concentrations (<10 nM), and concentrations >10 nM failed to suppress the proliferation any further. This dose dependency is compatible with the phosphorylation levels of the target proteins in immunoblot-

ting data and several previous reports in other types of cancers [10,12,36]. The cell cycle profile was distinct among each cell line. For example, G1 arrest was not induced and G2/M ratio was high in OVISE cells under DS-7423 exposure. This might be partly explained by the fact that GADD45 was induced by DS-7423 in



**Figure 6. Induction of the phosphorylation of TP53 at Ser46 and the accumulation of transcripts of the genes targeted by TP53, which participate in TP53-mediated apoptosis.** (A) Immunoblotting in OVMANA cells treated with DS-7423 at the indicated doses. Phosphorylation levels of MDM2 were inversely associated with p-TP53 at Ser46, but not with p-TP53 at Ser15. (B) Semi-quantitative RT-PCR in OVMANA and OVICE cells treated with DS-7423 at the indicated doses. Both *p53AIP1* and *PUMA* were induced by DS-7423. CT, untreated (negative) control; IR, irradiation at 14 Gy (positive control). *GADD45* was induced in OVICE, but not in OVMANA cells. (C) Quantification of the semi-quantitative RT-PCR in (B). Each experiment was repeated 3 times, and each value is shown as the mean of 3 experiments  $\pm$  SD. \* $p < 0.05$  (D) Effect of TP53 knockdown on apoptosis induction by DS-7423. TP53 was knocked down by two independent siRNAs specific to TP53 (siRNA1 and 2) in OVICE cells, which do not carry any mutation in TP53. The apoptotic cell population was evaluated using annexin-V staining, as described in Figure 5. The experiments were repeated 3 times, and each value is shown as the mean of 3 experiments  $\pm$  SD. \* $p < 0.05$  (E) Suppression of TP53 expression by siRNAs was confirmed by immunoblotting. (F) Effect of TP53 knockdown on cell proliferation by DS-7423 in MTT assay of OVICE cells. TP53 was knocked down by a siRNA1 specific to TP53 and MTT assay was subsequently performed as in Figure 2. Knockdown of TP53 diminished the anti-proliferative effect caused by DS-7423 on OVICE cells. The experiments were repeated 3 times, and each value is shown as the mean of 3 experiments  $\pm$  SD. \* $p < 0.05$  (G) TP53 expression plasmid (0.1  $\mu\text{g}/\mu\text{L}$ ) was cotransfected with pp53 TA Luc (0.25  $\mu\text{g}/\text{mL}$ ) plasmid into ES-2 cells mutated in TP53. The addition of DS-7423 increased the relative luciferase activity of TP53 in a dose-dependent manner. The experiments were repeated 3 times, and each value is shown as the mean of 3 experiments  $\pm$  SD. \* $p < 0.05$ . doi:10.1371/journal.pone.0087220.g006

these cells. Thus, the action mechanism of DS-7423 might be distinct in each type of cells, regardless of the TP53 status. Resistance to mTOR (mTORC1) inhibitors might be induced by several mechanisms, including increased activity of another mTOR complex, mTORC2, or upregulation of receptor tyrosine kinases such as insulin-like growth factor-1 receptor (IGF-1R) [37,38]. The use of mTORC1 inhibitors to treat OCCAs is currently being investigated in phase 2 clinical trials. The currently ongoing GOG (Gynecologic Oncology Group)-0268 (NCT01196429) trial recruits OCCA patients and treats the subjects with carboplatin and paclitaxel, followed by temsirolimus (CCI-779). A report on six cases with weekly administration of temsirolimus in recurrent OCCA patients showed partial response in one patient and stable disease in another patient [39]. However, given that our data suggest that dual PI3K/mTOR inhibitors, such as DS-7423, might be more promising than single mTORC1

inhibitors, clinical trials that involve a dual PI3K/mTOR inhibitor, such as DS-7423, seem warranted for OCCA.

DS-7423 induced significantly higher levels of apoptotic cell death in OCCA cells without mutations in TP53 than in OCCA cells with TP53 mutations. This result suggests both that the mutational status of TP53 might be a good biomarker to predict apoptosis induction by DS-7423, and that apoptosis depends on TP53 function. TP53 is degraded by MDM2, a ubiquitin ligase for TP53, and the MDM2 function is augmented by the kinase activity of Akt. Akt-mediated phosphorylation of MDM2 blocks its binding to p19ARF, increasing the degradation of TP53 [40,41]. DS-7423 increased the level of p-TP53 at Ser46, which results in induction of *p53AIP1* and *PUMA* (genes involved in TP53-mediated apoptosis) [29,42–44]. This data suggests that the apoptotic effect of DS-7423 depends, at least in part, on TP53 activity. The reasons for p-TP53 (Ser46), not p-TP53 (Ser15), being clearly induced and for apoptosis being preferentially



induced by high doses of DS-7423 should be further clarified. In addition, other non-apoptotic genes were not significantly induced by DS-7423, except for GADD45 in OVISe cells. Further analyses are warranted whether TP53 function is more involved in apoptosis rather than in cell cycle arrest and/or DNA repair process by DS-7423. Another possibility is that other proteins (such as FOXOs) which act downstream of Akt might also play a role in the induction [45]. Dephosphorylation of FOXOs at their Akt sites induces their nuclear translocation and triggers apoptosis by induction of pro-survival genes of the BCL2 family [46,47]. The observation that the phosphorylation of FOXO1/3a was suppressed by DS-7423, regardless of TP53 status, suggests that the pro-apoptotic effect of DS-7423 cannot be explained exclusively by the phosphorylation of FOXOs. The use of siRNA to knockdown TP53 rescued OCCA cells from apoptosis caused by DS-7423. We also confirmed by MTT assay that the anti-proliferative effect of DS-7423 was significantly diminished by knocking down TP53, suggesting that intact TP53 function might enhance the anti-tumor effect of DS-7423. Recently, it was reported that cell death caused by a PI3K inhibitor, BKM-120, was associated with TP53 status in glioma cells [48], and that PI3K/AKT inhibition was suggested to induce TP53-dependent apoptosis in HTLV-1-transformed cells [49]. These data also support the importance of wild-type TP53 in the induction of the cytotoxic effect of PI3K pathway inhibitors.

The involvement of multiple molecules in the activation of the PI3K/mTOR pathway underscores the critical need to develop predictive biomarkers that might also serve as therapeutic targets. Mutations of *PIK3CA* and amplification of *HER2* have been proposed to be useful biomarkers in breast cancer [50,51], whereas mutant Ras has been suggested to be a biomarker of resistance in several solid tumor cells [52]. All these biomarkers (*PIK3CA*, *HER2* and Ras) are focused on the RTK/Ras/PI3K pathway itself, and not on the cytotoxic effects associated with PI3K/mTOR inhibitors. Our data suggest that the presence of *PIK3CA* mutation and any other PI3K-activating alteration alone might not predict the sensitivity of OCCA cells to DS-7423. ES-2 cells, with no mutations in the RTK/Ras/PI3K pathway genes examined, showed low level of p-Akt, and the effect of DS-7423 in ES-2 xenografts was less robust, suggesting that the level of PI3K pathway activation would still be important for the sensitivity. However, the mutational status of TP53 might represent a better biomarker for the selection of tumors that could be killed by DS-7423 treatment. The frequency of mutations in *TP53* in OCCA was much less frequent than for ovarian cancers with other histology types [15,53]. These results indicate that OCCAs would be good candidates for clinical studies on the dual PI3K/mTOR inhibitor, DS-7423.

Our study has several limitations. First, cytostatic effect is still essential to suppress cell proliferation, regardless of TP53 status. Second, the ratio of apoptotic cells is low (less than 20%) even at high concentrations of DS-7423. Third, the mechanism of cytostatic effect by DS-7423 in OCCA is cell type dependent (i.e. G1 arrest was not induced in OVISe and ES-2 cells). Thus, careful consideration is required to evaluate the TP53-dependent cytotoxic effect of DS-7423. Further studies are warranted to elucidate the mechanism of action of DS-7423, and more efficient

induction of apoptosis might be needed for clinical application of this drug in OCCA.

## Supporting Information

**Figure S1 Immunoblotting of OCCA cells (ES-2 and JHOC-9), treated with DS-7423 at concentrations ranging from 0 to 2,500 nmol/L.** As shown in Figure 3, phosphorylation of AKT and its target proteins were downregulated by DS-7423. In ES-2 cells, basal level of p-AKT at Thr 308 was very low (as shown in Fig. 1), but p-AKT at Ser473 was clearly suppressed by DS-7423. (PPTX)

**Figure S2 *In vivo* effect of DS-7423 in nude mice.** (A) Western blot of total lysates from the TOV-21G and RMG-1 xenografts, total lysates were harvested 2 and 6 h after the last drug administration of DS-7423. The levels of p-Akt (Thr-308) and p-S6 (Ser-240/244) were assessed. (B) Subcutaneous xenograft tumors in athymic BALB/c mice were established after injection of ES-2 cells. Mice were treated daily at the indicated doses (1.5, 3 or 6 mg/kg/day, totally 8 times) of DS-7423 or non-treated control. Estimated tumor volumes were smaller in mice treated daily with 6 mg/kg of DS-7423, compared to the control. Western blot of total lysates from the ES-2 xenografts (treated with 6 mg/kg of DS-7423) was also shown below. (PPTX)

**Figure S3 The size of apoptotic cell population was compared between DS-7423 and rapamycin in OVISe cells, using annexin-V FITC and PI double staining (as shown in Fig. 5A–5B).** The percentage of apoptotic cells was significantly higher in cells treated with DS-7423, compared with those with rapamycin. (PPTX)

**Figure S4 Semi-quantitative RT-PCR in OVMANA and OVISe cells treated with DS-7423 at the indicated doses.** Each expression level of p53R2, TIGAR, GLS2, GADD45, 14-3-3 sigma and PAI-1 was not enhanced by DS-7423. Each experiment was repeated 3 times, and each value is shown as the mean of 3 experiments  $\pm$  SD. (PPTX)

**Table S1 Phosphorylation and mutational status in 9 OCCA cell lines.** Elevated phosphorylation of cMET, HER2 and HER3, and mutations of *PIK3CA*, *PTEN*, *KRAS* and *TP53* were listed in each cell line. (XLSX)

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## Author Contributions

Conceived and designed the experiments: T Kashiyama KO YS YH KM. Performed the experiments: T Kashiyama YI YS YH KI CM RK. Analyzed the data: T Kashiyama KO YS YH. Contributed reagents/materials/analysis tools: YI AM T Koso T Fukuda MT K Shoji K Sone TA OW-H KK SN KM FM HA TY YO T Fujii. Wrote the paper: T Kashiyama KO.

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# Matrix Metalloproteinase (MMP)-9 in Cancer-Associated Fibroblasts (CAFs) Is Suppressed by Omega-3 Polyunsaturated Fatty Acids *In Vitro* and *In Vivo*

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## Abstract

Cancer associated fibroblasts (CAFs) are responsible for tumor growth, angiogenesis, invasion, and metastasis. Matrix metalloproteinase (MMP)-9 secreted from cancer stroma populated by CAFs is a prerequisite for cancer angiogenesis and metastasis. Omega-3 polyunsaturated fatty acids (omega-3 PUFA) have been reported to have anti-tumor effects on diverse types of malignancies. Fat-1 mice, which can convert omega-6 to omega-3 PUFA independent of diet, are useful to investigate the functions of endogenous omega-3 PUFA. To examine the effect of omega-3 PUFA on tumorigenesis, TC-1 cells, a murine epithelial cell line immortalized by human papillomavirus (HPV) oncogenes, were injected subcutaneously into fat-1 or wild type mice. Tumor growth and angiogenesis of the TC-1 tumor were significantly suppressed in fat-1 compared to wild type mice. cDNA microarray of the tumors derived from fat-1 and wild type mice revealed that MMP-9 is downregulated in fat-1 mice. Immunohistochemical study demonstrated immunoreactivity for MMP-9 in the tumor stromal fibroblasts was diffusely positive in wild type whereas focal in fat-1 mice. MMP-9 was expressed in primary cultured fibroblasts isolated from fat-1 and wild type mice but was not expressed in TC-1 cells. Co-culture of fibroblasts with TC-1 cells enhanced the expression and the proteinase activity of MMP-9, although the protease activity of MMP-9 in fat-1-derived fibroblasts was lower than that in wild type fibroblasts. Our data suggests that omega-3 PUFAs suppress MMP-9 induction and tumor angiogenesis. These findings may provide insight into mechanisms by which omega-3 PUFAs exert anti-tumor effects by modulating tumor microenvironment.

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## Introduction

The tumor microenvironment is comprised of microvascular endothelial cells, adjacent normal epithelial cells and cancer-associated fibroblasts (CAFs), and is reported to be an important regulator of tumorigenesis [1,2]. As the most common cellular population found in the tumor microenvironment, CAFs are responsible for the synthesis of proteins involved in the remodeling of the extracellular matrix (ECM), and for the secretion of growth factors and cytokines that regulate tumor cell proliferation and invasion [3,4]. In murine ovarian cancer xenograft models, the p53/NF- $\kappa$ B pathway in CAFs significantly increased in vivo tumor growth [5]. In colon cancer, Zhu Y et al. report that IL-1 $\beta$  increased colon cancer cell proliferation and invasion by up-regulating COX-2 signaling in CAFs [6].

Matrix-metalloproteinases (MMPs) are synthesized as proenzymes and typically activated by proteolytic removal of a propeptide [7]. MMPs are reported to influence tumor progression by facilitating events pivotal for neovascularization and establishment of distant metastasis including proliferation, survival and migration of endothelial, tumor and stromal cells [8,9]. MMP-2 and MMP-9 are implicated as prerequisites for angiogenesis and metastasis in the carcinogenic process. MMP-2 is expressed in the various cancer cell lines [10]. In contrast, MMP-9 has very limited or no expression in these cancer cells. Instead, MMP-9 is well-known to be secreted from cancer stromal fibroblasts and endothelial cells [11,12]. MMP-9 is a member of a family of zinc containing endoproteinases that is involved in degradation of extracellular matrix (ECM) and in vascular remodeling [13].

Docosahexaenoic acid (DHA, 22:6n-3) and eicosapentaenoic acid (EPA, 20:5n-3) are representative mediators of omega-3

polyunsaturated fatty acids (omega-3 PUFAs) and exert anti-inflammatory effects in acute and chronic pathological inflammatory reactions by counteracting inflammation [14]. Omega-3 PUFAs are also reported to have anti-cancer effects based on in vitro and vivo studies [15–17]. Several mechanisms have been proposed to explain the anti-cancer effects of omega-3 PUFAs. Omega-3 PUFAs alter the growth of tumor cells by modulating cell replication, by interfering with components of the cell cycle or by increasing cell death via necrosis or apoptosis [18,19]. Omega-3 PUFAs are also known to exert anti-angiogenic effects by inhibiting the production of many angiogenic mediators including: vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), and prostaglandin E2 (PGE2) [20–24].

Dietary supplementation is a traditional approach to modify tissue nutrient composition in animal studies of nutrition. Feeding animals diets that alter specific nutritional and non-nutritional components can help to differentiate experimental groups; however, it can be exceedingly difficult to provide diets that are identical in all but a single or a small but controlled number of components. Kang et al. recently engineered a transgenic mouse that carries the fat-1 gene from the roundworm *Caenorhabditis elegans* [25]. This gene encodes an omega-3 fatty acid desaturase that catalyzes the conversion of omega-6 to omega-3 PUFAs and that is absent in most animals, including mammals. There is a remarkable difference in the tissue omega-6/omega-3 PUFA ratio between wild type and fat-1 transgenic mice [26]. Fat-1 mice, which typically exhibit a balanced ratio of omega-6 to omega-3 PUFAs in their tissues and organs independent of diet, allow carefully controlled studies to be performed in the absence of potential confounding factors of diet. This makes them a useful model to investigate the biological properties of endogenous omega-3 PUFAs [25]. Several reports using fat-1 mice have demonstrated anti-cancer effects of omega-3 PUFAs [27–30]. In these investigations, omega-3 PUFAs exerted anti-cancer effects by suppressing inflammatory reactions and PGE2 secretion from cancer cells. To date, there are few studies that investigate the involvement of omega-3 PUFAs in the biology of CAFs.

In this study, we hypothesized that omega-3 PUFAs may alter tumor microenvironments by influencing CAF activity. To examine this hypothesis, fibroblasts derived from fat-1 and wild type mice were assessed both in vitro and in vivo under conditions allowing interaction with TC-1 cancer cells. TC-1 cells were derived from the epithelium of C56BL/6 mice and immortalized by human papillomavirus (HPV) type 16 E6 and E7 oncoproteins. They are commonly used in vitro and in vivo in murine models of HPV-related cancer [31]. Here, we investigated the involvement of omega-3 PUFAs in TC-1 tumorigenesis by comparing fat-1 and wild type mice. Our specific focus involved the study of tumor-associated fibroblasts. These models are useful in the study of CAFs because the cancer cells originate in wild type murine epithelium while the cancer stromal components, including CAFs, come from fat-1 (omega-3 PUFAs-rich) or wild type (normal PUFAs) mice.

## Materials and Methods

### Animals and diet

Fat-1 mice were created on a C57BL/6 background as described [26] and subsequently backcrossed (at least four times) onto a C57BL/6 background. Animals were fed a special diet (AIN-76A+10% safflower oil; CLEA Japan, Inc.) that contained 10.3% total fat with fatty acid composition of C16:0 (7.6%), C18:0 (2.7%), C18:1n-9 (14.1%), C18:2n-6 (73.2%), C18:3n-3 (0.3%), C20:4n-6 (<0.1%), C20:5n-3 (<0.1%), C22:6n-3 (<0.1%), high

in n-6 and low in n-3 fatty acids, until the desired age (6–8 weeks) for experiments. To prevent the oxidation of lipids in the diet, all foods were stored in the refrigerator with antioxidants (AGELESS; Mitsubishi Gas Chemical Inc.), and prepared newly every two days. Animal studies were approved by the University of Tokyo Animal Committee.

### Tumor growth assay in mice

TC-1 cells are derived from a primary lung epithelial cell from C56BL/6 mice and immortalized using HPV 16 E6/E7 plus c-Ha-ras (kind gift from Dr. T. C. Wu, Johns-Hopkins University, Baltimore MD USA) [32]. TC-1 cells were cultured in DMEM (Gibco, NY, USA) containing 10% FBS, 100 U/ml penicillin, 0.1 mg/ml streptomycin, and 0.25 g/ml amphotericin B. Eight-week-old female mice were injected with  $5 \times 10^6$  murine TC-1 cells suspended in 100  $\mu$ l of DMEM. Tumor volume, based on caliper measurements, was calculated at 7 and 14 days after injection according to the following formula: (tumor volume) =  $1/2 \times (\text{the shortest diameter})^2 \times (\text{the largest diameter})$ . Mice were sacrificed 14 days after inoculation, tumors were excised and stored at  $-80^\circ\text{C}$  for future analyses.

### cDNA microarray

Total RNA from TC-1 tumors (above) was extracted using an RNeasy minikit (QIAGEN, Hilden, Germany). For the cDNA microarray analysis, 0.5  $\mu$ g of pooled total RNA was amplified and labeled using an Amino Allyl MessageAmp™ II mRNA Amplification kit (Applied Biosystems, Foster City, CA, USA) according to the manufacturer's instructions. Each sample of mRNA labeled with Cy3 and reference mRNA labeled with Cy5 was cohybridized to the Gene™ Mouse Oligo chip 24 k (Toray Industries Inc., Tokyo, Japan) at  $37^\circ\text{C}$  for 16 h. After hybridization, each DNA chip was washed and dried. Hybridization signals derived from Cy3 and Cy5 were scanned using Scan Array Express (PerkinElmer, Waltham, MA, USA). The scanned image was analyzed using GenePix Pro (MDS Analytical Technologies, Sunnyvale, CA, USA). All analyzed data were scaled by global normalization. GEO accession number is GSE54079.

### Immunohistochemistry

Paraffin sections (4  $\mu$ m) of TC-1 tumors were dewaxed in xylene and rehydrated through graded ethanol to water. Antigens were retrieved by boiling in 10 mM citrate buffer (pH 6.0) for 30 min. The cooled sections were incubated in DAKO REAL Peroxidase-Blocking solution (DAKO, Carpinteria, CA, USA) for 10 min to quench endogenous peroxidase. To block nonspecific binding, sections were incubated in DAKO Protein Blocking solution (DAKO) for 10 min at room temperature. Sections were then incubated with a rabbit polyclonal antibody against mouse MMP-9 (PAB12714, Abnova, 1:100 dilution) in DAKO REAL Antibody Diluent (DAKO) overnight at  $4^\circ\text{C}$ . The slides were incubated for 1 hour at room temperature with peroxidase-conjugated secondary antibodies, washed, incubated with DAB, counterstained with hematoxylin, dehydrated through an ethanol series and xylene, and mounted. To evaluate tumor microvessel formation, tumor sections were stained for CD-31 using a rat monoclonal antibody against mouse CD-31 (ab56299, Abcam, Tokyo, Japan, 1:100 dilution).

### RT-quantitative PCR (RT-qPCR)

Total RNA was extracted from TC-1 tumors and cultured fibroblasts using an RNeasy minikit (QIAGEN, Hilden, Germany), followed by reverse transcription. cDNA was amplified for

40 cycles in a Light Cycler 480 (Roche, Basel, Switzerland) using a Universal Probe Master Mix and the following primers and Universal Probe Library (UPL) probes (Roche). The primer pairs and the universal probes corresponding to the each primer that were used in amplifications were as follows: mouse  $\beta$ -actin, 5'-ATTGAAAGATCAGCCAAGACC-3' and 5'-CCGAATCTCACGGACTAGTGT-3' probe88; mouse MMP-9, 5'-ACGACATAGACGGCATCCA-3' and 5'-GCTGTGGTTTCAGTTGTG-GTG-3' probe19. Expression of MMP-9 was normalized using  $\beta$ -actin mRNA as an internal standard. Expression levels were calculated by the comparative Ct method using  $\beta$ -actin as an endogenous reference gene.

#### Primary fibroblast culture and co-culture with TC-1 cells

Lungs were isolated from fat-1 transgenic and wild type mice and washed with saline to remove blood cells. Isolation and culture of pulmonary fibroblasts were performed using methods described previously [33]. Lung tissues were minced into small pieces and incubated in DMEM (Gibco, NY, USA) containing type I collagenase (0.25%; Sigma-Aldrich, St Louis, MO, USA) and deoxynuclease I (15 U/ml; TaKaRa, Tokyo, Japan) for 120 min at 37°C. The resultant dispersed cells were separated by filtration through nylon cell strainers (70  $\mu$ m, BD, Franklin Lakes, NJ, USA). Fibroblasts in the filtrate were collected, placed into 10 cm dishes in DMEM containing 10% FBS, 100 U/ml penicillin, 0.1 mg/ml streptomycin, and 0.25 mg/ml amphotericin B, and incubated for 7–10 days. Fibroblasts were purified from other cell population by differential adhesion and serial passage.

Confluent fibroblasts and TC-1 cells were trypsinized and resuspended in DMEM containing 10% FBS, 100 U/ml penicillin, 0.1 mg/ml streptomycin, and 0.25 g/ml amphotericin B and  $1 \times 10^5$  cells/ml cells of each cell type were plated together in 12 well culture plates. Co-cultures were incubated at 37°C 5% CO<sub>2</sub> in a humidified atmosphere for 24 hours. Homotypic cultures served as controls.

#### Gelatin zymography

Gelatin zymography assays were performed using a Gelatin zymography kit (Cosmobio, Sapporo, Japan) according to the manufacturer's instructions. Cell culture supernatants were collected and centrifuged at 1,500 rpm for 5 minutes. The cell free supernatant was mixed with 2 $\times$  sample buffer and electrophoresed using precast gels (10% polyacrylamide, 0.1% gelatin) at 4°C for 1 hour. Subsequent enzymatic reactions were performed at 37°C overnight. Gelatinase activities were visualized using specific staining solutions and destained in acetic acid-methanol-dH<sub>2</sub>O (1:3:6). For semi-quantitative analyses, gelatin zymography bands were analyzed using image analysis software (ImageJ).

#### Statistical analysis

Data are presented as means  $\pm$  SEM. Statistical analyses were carried out by Student's *t*-test, or Wilcoxon analysis using JMP software. A value of  $P < 0.05$  was considered significant. In the figure legends, asterisks indicate those comparisons with statistical significance ( $p < 0.05$ ).

## Results

### Tumor growth and angiogenesis of TC-1 tumors is suppressed in fat-1 mice

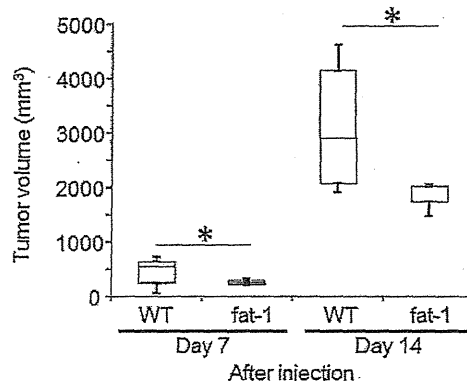
To investigate the effect of omega-3 PUFAs on cervical cancer tumorigenesis, we injected TC-1 cells subcutaneously into fat-1 and the litter-mate wild type C57/BL6 mice. TC-1 tumor formation rates and tumor growth were assessed by the number

of mice forming tumors and three-dimension tumor sizes, respectively. There was no difference in tumor formation rates between fat-1 and wild type mice. TC-1 tumor sizes were plotted for 14 days in fat-1 and wild-type controls (Fig. 1). Tumor growth rates were consistently lower in fat-1 mice when compared to wild type mice at all time points. In fat-1 mice, tumor size at 14 days after injection was significantly smaller than in wild-type controls (Fig. 1). Although cell growth of TC-1 cells is dependent on HPV16 E6/E7 expression, E6 and E7 expression in TC-1 tumors was not downregulated in fat-1 mice (Fig. S1).

To further delineate mechanisms behind these differences in tumor growth in this model, particularly in terms of the possible role of host-derived cancer-associated stromal components including CAFs, we next examined whether omega-3 PUFAs modified angiogenesis in TC-1 tumor. We assessed semi-quantitatively tumor microvessel density in TC-1 tumors derived from fat-1 and wild type mice by counting the number of CD31-positive microvessels in immunohistochemical assay (Fig. 2A and 2B). CD31 immunostaining of TC-1 tumors derived from fat-1 and wild type mice (Fig. 2A) demonstrated hypovascularity of the fat-1-derived TC-1 tumors when compared with wild type-derived tumors. The number of CD31-positive microvessels per high-power field in fat-1 mice was significantly lower than that in wild type mice (Fig. 2B). These *in vivo* data indicated that TC-1 tumor growth and angiogenesis were at least suppressed in fat-1 mice when compared with wild type counterparts although it was difficult to accurately estimate TC-1 cell growth in fat-1 mice.

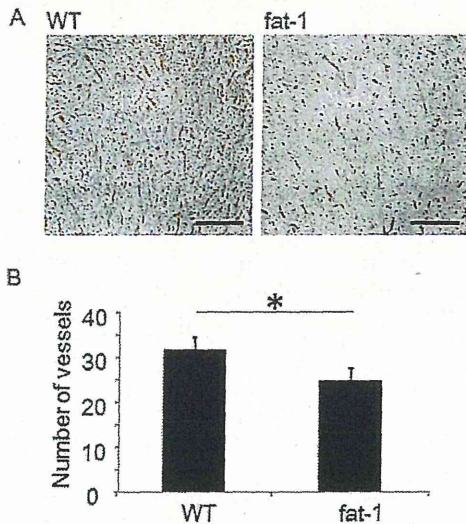
### MMP-9 is downregulated in fat-1 mice-derived TC-1 tumors

To investigate potential differences in gene expression profiles of TC-1 tumors growing in the skin of fat-1 and wild type mice, TC-1 tumor tissues obtained from fat-1 and wild type mice were analyzed by cDNA microarray. Since omega-3 PUFAs are reported to influence cell proliferation and inflammation, Table 1 lists relative gene expression levels for representative genes associated with the tumor growth and inflammation (Table 1). With the exception of EGF, expression levels of almost all inflammatory cytokines/chemokines and growth factors in TC-1



**Figure 1. Tumor growth rates in fat-1 and wild type (WT) mice.**  $5 \times 10^6$  murine TC-1 cells suspended in 100  $\mu$ l of DMEM were injected s.c. into each of 10 fat-1 and wild type mice. Tumor volume, based on caliper measurements, was calculated at 7 and 14 days after injection according to the following formula: (tumor volume) =  $1/2 \times (\text{the shortest diameter})^2 \times (\text{the largest diameter})$ . Mean values with standard deviations are presented. Asterisks indicate those comparisons (fat-1 vs. wild type mice) with statistical significance ( $p < 0.05$ ). doi:10.1371/journal.pone.0089605.g001





**Figure 2. Omega-3 PUFAs suppress tumor vasculogenesis.** CD31 immunostaining of the TC-1 tumor derived from wild type (WT) mice (A) and fat-1 (B). Bars indicate 200  $\mu$ m. (C) Microvessel densities in TC-1 tumors are expressed as the representative number of labeled vessels in 4 fields ( $n=5$ ). Mean values with standard deviations are presented. Asterisks indicate those comparisons (fat-1 vs. wild type mice) with statistical significance ( $p<0.05$ ). doi:10.1371/journal.pone.0089605.g002

tumors from fat-1 mice were higher than those, in tumors from wild-type controls. This suggested that anti-inflammatory and anti-cell growth effects of omega-3 PUFAs are unlikely to be central to their anti-tumor activities, at least in this model. We next examined the expression of MMPs in these TC-1 tumors to further address stroma-related angiogenesis in the tumor micro-environments (Table 1). cDNA microarray demonstrated that the expression of MMP-2 and -9 were suppressed in fat-1 mice-derived TC-1 tumor while those of other MMPs were tended to be upregulated when compared to controls. To confirm these effects at the RNA level, RT-qPCR for MMP-9 was performed. MMP-9 RNA levels in fat-1 mice were approximately 60% lower than those in wild type controls (Fig. 3A). TC-1 tumor immunohistochemistry confirmed these results at the protein level. MMP-9 immunoreactivity in wild type mouse-derived TC-1 tumors was clearly stronger than fat-1 mouse-derived tumors (Fig. 3B). High power histochemical analysis of MMP-9 immunoreactivity in TC-1 cells (Fig. 3B, inserts) revealed negligible expression, while that of the stromal components including CAFs and endothelial cells was strongly positive only in the wild type-derived TC-1 tumors. These data indicated that the production of MMP-9 in CAFs and endothelial cells was clearly suppressed in fat-1 mice.

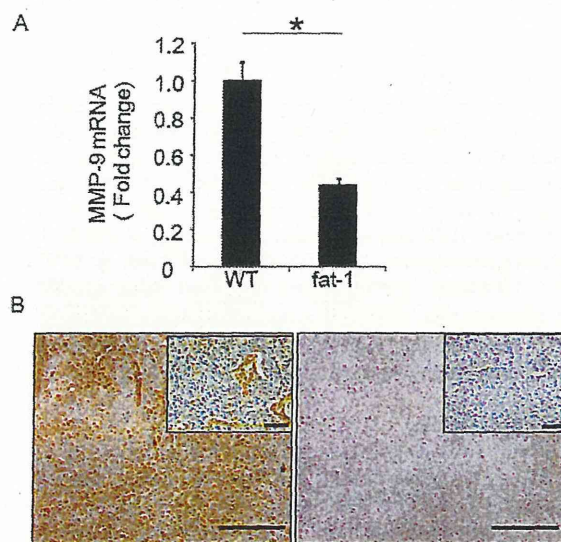
MMP-9 expression and gelatinase activity were suppressed in cultured primary fibroblasts derived from fat-1 mice

To mimic the cancer stromal microenvironment in vitro, we cultured fibroblasts isolated from fat-1 and wild type mice with TC-1 cells. We used lung tissues from fat-1 and wild type mice as the sources for fibroblasts, and differential adhesion methodology for their isolation. All fibroblasts were passaged 3–4 times prior to use in experiments. Baseline MMP-9 expression levels in primary fibroblasts from fat-1 mice were approximately 60% lower than those from wild type-derived fibroblasts (Fig. 4A). Culture

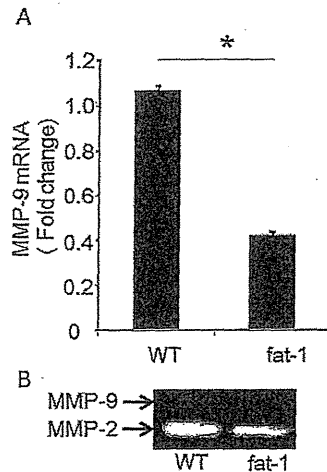
**Table 1.** Cytokine, growth factor, and MMP gene expression comparisons in TC-1 tumors from fat-1 vs wild type mice.

Genes	fat-1/WT ratio
EGF	0.4
CXCL-12	1.2
HGF	1.3
TGF- $\alpha$	1.4
IL-6	2.0
IFN- $\gamma$	2.4
TNF- $\alpha$	3.3
VEGF	3.7
IL-1 $\beta$	12.0
<b>MMP-9</b>	<b>0.4</b>
<b>MMP-2</b>	<b>0.6</b>
MMP-7	1.9
MMP-1b	2.9
MMP-3	2.9
MMP-13	2.9
MMP-1a	3.6
MMP-16	4.1
MMP-10	11.6

doi:10.1371/journal.pone.0089605.t001



**Figure 3. MMP-9 expression is downregulated in TC-1 tumors from fat-1 mice.** Total RNA was extracted from TC-1 tumors, followed by reverse transcription. MMP-9 mRNA levels were measured by qRT-PCR. Expression levels of MMP-9 were normalized to  $\beta$ -actin as an internal standard ( $n=4$  in each group). Asterisks indicate those comparisons (fat-1 vs. wild type (WT) mice) with statistical significance ( $p<0.05$ ). MMP-9 immunostaining of TC-1 tumors derived from wild type (WT) and fat-1 mice. Bars indicate 200  $\mu$ m in low-power fields, 50  $\mu$ m in high-power fields. doi:10.1371/journal.pone.0089605.g003



**Figure 4. MMP-9 expression and enzymatic activity in primary-cultured fibroblasts.** (A) Primary fibroblasts isolated from murine lungs were cultured. Total RNA from fibroblasts was reverse transcribed and MMP-9 mRNA levels were measured by qRT-PCR. Expression levels of MMP-9 were normalized to  $\beta$ -actin as an internal standard. Data are the representative of three independent experiments. The data were analyzed by using the Student's *t*-test. Asterisks indicate those comparisons (fat-1 vs. wild type (WT) mice) with statistical significance ( $p < 0.05$ ). (B) Gelatin zymography: Supernatant from primary fibroblast cultures were collected and separated by electrophoresis. Gelatinase activities were visualized by standard staining techniques. doi:10.1371/journal.pone.0089605.g004

supernatants from each primary fibroblast subtype were collected and subjected to polyacrylamide gel electrophoresis to examine differences in MMP gelatinase activities (Fig. 4B). Using this method, MMP-2 was detected but MMP-9 was not in both fat-1 and wild type fibroblast

Next, these primary fibroblasts were co-cultured with TC-1 cells to examine fibroblasts activation upon in vitro exposure to cancer cells<sup>11</sup>. Fibroblasts and TC-1 cells were co-cultured for 24 hours and MMP-9 transcription was measured by RT-qPCR. Fibroblasts from fat-1 and from wild type mice increased demonstrated MMP-9 expression upon exposure to TC-1 cells (Fig. 5A). In fibroblasts derived from fat-1 mice, the extent of MMP-9 induction was lower than that in fibroblasts from wild type mice (Fig. 5A). MMP-9 was not expressed in homotypic TC-1 cells, consistent with immunohistochemical data from our in vivo model. Furthermore, we confirmed that MMP-9 was not expressed in the TC-1 cells by using a transwell co-culture model (Fig. S2). Therefore, MMP-9 in the co-culture condition was derived not from TC-1 cells but from fibroblasts. MMP-2 gelatinase activity was detected in all cell culture conditions, including homotypic TC-1 cell cultures, and was not altered by co-culture conditions (Fig 5B, 5D). MMP-9 gelatinase activities were, however, detectable in fibroblast-TC-1 cell co-cultures, although MMP-9 gelatinase activity involving fat-1 fibroblasts was clearly suppressed when compared with wild type fibroblasts (Fig 5B, 5C), again supporting the concept that MMP-9 expression and gelatinase activity are suppressed by endogenous omega-3 PUFAs in CAFs derived from fat-1 mice.

## Discussion

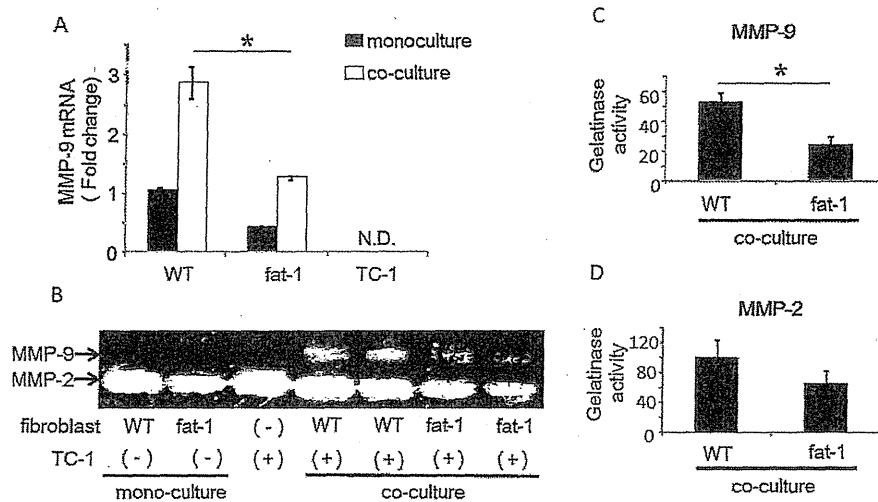
In this study, we examined the involvement of defined dietary factors (omega-3 and omega-6 PUFAs) in tumorigenesis using

HPV-positive TC-1 cells and proposed a novel anti-tumor mechanism for omega-3 PUFAs that depends on the activities of CAFs. Omega-3 PUFAs have been shown to suppress cancer incidence and growth in various types of cancers [18,19,27]. Many studies have demonstrated that omega-3 PUFAs have these effects through anti-inflammatory responses directly and/or indirectly [25–27] as well as through inducing tumor cell apoptosis and/or suppressing tumor cell proliferation [18,19]. However, there have been few reports about the effects of omega-3 PUFAs on CAFs. CAFs have been implicated in facilitating the growth of several tumors by directly stimulating tumor cell proliferation and by enhancing angiogenesis [34,35]. Targeting genes and signaling pathways mediating interaction of CAFs and tumor-microenvironment are considered to be essential for development of new and effective cancer therapies [36,37]. In this study, using fat-1 mice and TC-1 tumor cells, we were able to clarify the effect of omega-3 PUFA-rich CAFs on tumorigenesis.

Many molecules, including growth factors, cytokines, and MMPs, play stimulatory and inhibitory roles in promoting angiogenesis [21]. We investigated gene expression of angiogenesis-related cytokines, growth factors and MMPs in the TC-1 tumors of fat-1 and wild type mice by cDNA microarray. The expression levels of almost all inflammatory cytokines/chemokines and growth factors in TC-1 tumors from fat-1 mice were higher than those in tumors from wild-type controls. In contrast, EGF and MMP-2 and -9 expression levels in fat-1 mice were lower than those in wild type mice. Since down-regulation of EGF in TC-1 tumors from fat-1 mice was predicted to promote TC-1 cell proliferation but tumor size in fat-1 mice was significantly smaller than in wild-type controls, we hypothesize that differences in MMP production between fat-1 and wild type mice may be responsible for the suppression of TC-1 tumor growth. Our in vitro data verified that MMP-9 derived from CAFs activated by TC-1 cell exposure was downregulated in the fat-1 mice. MMPs are reported to influence tumor progression by facilitating events pivotal for neovascularization and for the establishment of distant metastasis including proliferation, survival and migration of endothelial, tumor and stromal cells [8,9]. MMP inhibitors reduce angiogenesis, tumor number, and tumor growth, as does genetic ablation of MMP-9 [38]. In contrast to MMP-2, which is constitutively expressed, MMP-9 levels are usually low and enzyme expression is induced by cytokines that stimulate NF- $\kappa$ B [8,39]. Furthermore, elevated serum and tissue levels of MMP-9 are reported to be associated with cancer invasion and metastasis [40]. In breast cancer, MMP-9 activity has been localized around CAFs and CAFs co-cultured with cancer cells which secrete TGF- $\beta$ , TNF- $\alpha$ , and other cytokines, increase production of MMP-9 [11], consistent with our in vitro data. In our data, suppression of MMP-9 would contribute to accompany hypo-angiogenesis in the tumor stromal component in fat-1 tumor.

Several transcription factors, including activator protein-1 (AP-1), Sp-1, and NF- $\kappa$ B, are reported to be involved in alterations in MMP-9 expression following exposure to various cytokines [41–43]. Conversely, several reports have demonstrated that omega-3 PUFAs have an inhibitory effect on the NF- $\kappa$ B pathway when activated by various stimuli [43–47]. Our own data suggest that activation of the NF- $\kappa$ B pathway upon co-culture with TC-1 cells may be suppressed by elevated level of omega-3 PUFAs in fibroblast obtained from fat-1 mice.

In our experiments, we consistently used primary fibroblasts that had been passaged 3–4 times only. Nevertheless, lipid mediator analysis of the fibroblasts revealed that those derived from fat-1 mice produced larger amounts of omega-3 PUFAs including EPA-derived metabolites when compared with those



**Figure 5. The increased MMP-9 expression and activity in TC-1/fibroblast co-cultures is inhibited in fat-1 mice.** (A) Isolated fibroblasts were co-cultured with TC-1 cells for 24 hours and expression levels of MMP-9 in the fibroblasts were measured by RT-qPCR. Expression levels of MMP-9 were normalized to  $\beta$ -actin as an internal standard. The data are representative of three independent experiments. The data were analyzed using the Student's *t*-test. Asterisks indicate those comparisons (fat-1 vs. wild type (WT) mice) with statistical significance ( $p < 0.05$ ). "N.D." indicates 'not detected'. (B) Gelatin zymography: Supernatants from fibroblast homotypic cultures and fibroblast/TC-1 co-cultures were collected and separated by electrophoresis. Gelatinase activities were visualized by standard staining techniques. (C, D) For semi-quantitative analyses, gelatin zymography bands were analyzed using image analysis software. Results are represented as mean  $\pm$  SEM of three independent experiments. The data were analyzed using the Student's *t*-test. Asterisks indicate those comparisons (fat-1 vs. wild type (WT) mice) with statistical significance ( $p < 0.05$ ). doi:10.1371/journal.pone.0089605.g005

derived from wild type controls (Fig.S3), confirming that the cultured fibroblasts retained the characteristic to make omega-3 PUFA rich environment.

TC-1 tumor microarray data showed that several inflammatory cytokines/chemokines and growth factors were upregulated in fat-1 mice. However, the data indicated gene expression levels in both TC-1 cells and stromal components, including fibroblasts, endothelial and immune cells. Therefore, expression levels of each cytokine/chemokine were dependent on their primary sources of production. MMP-9 was produced by the fibroblasts derived from fat-1 mice but not by TC-1 cells. Therefore, suppression of the NF- $\kappa$ B pathway by elevated levels of omega-3 PUFAs in fat-1-derived stromal components may have a specific and potent effect on MMP-9 expression levels when compared with the other inflammatory cytokines/chemokines. On the other hand, a previous study demonstrates that omega-3 PUFAs activate NK cells and increase proportions of activated CD8+ cells; this is followed by enhanced anti-tumor effects [48]. Therefore, omega-3 PUFAs may exert both anti-inflammatory and pro-inflammatory effects on immune cells.

In this study, we have demonstrated that an omega-3 PUFAs-rich microenvironment can suppress MMP-9 secretion from CAFs and that this is associated with subsequent tumor hypo-angiogenesis. This study proposes a novel anti-tumor effect of omega-3 PUFAs by modulating tumor microenvironment especially on CAFs.

## Supporting Information

**Figure S1 Expression levels of E6 and E7 mRNA in TC-1 tumor.** Total RNA was extracted from TC-1 tumors, followed by reverse transcription. E6 and E7 mRNA levels were measured by qRT-PCR. Expression levels of E6 and E7 mRNA were normalized to  $\beta$ -actin as an internal standard. The E6 primers were forward, 5'- TGCACAGAGCTGCAAACAAC -3', and

reverse, 5'- AGCATATGGATTCCCATCTC -3'. The E7 primers were forward, 5'- TTTGCAACCAGAGACAACACTGA -3', and reverse, 5'- GCCCATTAAACAGGTCTTCCA -3'. (TIF)

**Figure S2 MMP-9 mRNA was not induced from TC-1 cells by co-culture with fibroblasts.** 150  $\mu$ L of suspensions ( $2 \times 10^6$ /mL) of TC-1 cells or fibroblasts were added to the upper chamber, 500  $\mu$ L of suspensions ( $1 \times 10^5$ /mL) of TC-1 cells or fibroblasts were added to the lower chamber of the 24 well Transwell plates (1  $\mu$ m pore) and placed in an incubator with 5% CO<sub>2</sub> at 37°C for 24 h and expression levels of MMP-9 in the TC-1 or fibroblasts in the lower chamber were measured by RT-qPCR. N.D. indicates 'not detected'. (TIF)

**Figure S3 Lipid mediator analysis of fibroblast/TC-1 co-cultured medium.** Supernatants from fibroblast/TC-1 co-cultures were collected and LC-MS/MS-based mediator lipidomics was performed on Acquity UPLC BEH C<sub>18</sub> column (1.0 mm  $\times$  150 mm  $\times$  1.7  $\mu$ m) using Acquity UltraPerformance LC system (Waters Co.) coupled to an electrospray (ESI) triple quadrupole mass spectrometer (QTRAP5500; AB SCIEX). The MS/MS analyses were performed in negative ion mode, and the eicosanoids and docosanoids were identified and quantified by multiple reaction monitoring. Calibration curves between 1 and 1000 pg and the LC retention times for each compounds were constructed with synthetic standards. (TIF)

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## Author Contributions

Conceived and designed the experiments: AT K. Kawana KT AY MA K. Koga. Performed the experiments: AT K. Kawana KT AY YI KN TI HN

SK YM. Analyzed the data: AT K. Kawana KT AY K. Koga TA KA OW KO HA MA JK TF YO. Contributed reagents/materials/analysis tools: AT KT AY. Wrote the paper: AT K. Kawana MA.

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# Poor prognosis of ovarian cancer with large cell neuroendocrine carcinoma: Case report and review of published works

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## Abstract

Large cell neuroendocrine carcinoma (LCNEC) is well-reported to result in unfavorable prognoses in many organ cancers while being rarely reported in gynecologic cancer, especially ovarian and endometrial cancers. Here we report a case of ovarian cancer with LCNEC which spread to distant organs within 1 year of primary surgery despite the fact that the post-surgical stage was Ia. The case received platinum-based chemotherapy as an adjuvant therapy after her curative surgery. However, LCNEC in the case was resistant to the chemotherapy. In our review of published works, ovarian cancer cases with LCNEC show poor prognoses regardless of adjuvant chemotherapy following complete resection. Median overall survival was 10 months in stage I cases. Development of chemotherapy sensitive for LCNEC is needed.

**Key words:** chromogranin A, gynecologic cancer, large cell neuroendocrine carcinoma, ovarian cancer, platinum-based chemotherapy.

## Introduction

Large cell neuroendocrine carcinoma (LCNEC) is synonymous with 'undifferentiated carcinoma of non-small cell neuroendocrine type' and defined as 'a malignant tumor composed of large cells that show neuroendocrine differentiation'.<sup>1</sup> However, there exist no generally accepted criteria for neuroendocrine tumor differentiation, which usually depends on a combination of typical structural, immunohistochemical and ultrastructural findings. World Health Organization (WHO) criteria describe that LCNEC of the lung is characterized by positive immunostaining for chromogranin A, synaptophysin or CD56

(N-CAM) and at least one of them is enough if the staining is clear cut.<sup>2</sup> LCNEC of the ovary is very rare and only 35 cases have been reported previously worldwide.<sup>2–17</sup> Its prognosis is generally very poor, even when the diagnosis is made at an early stage. We experienced a case of LCNEC of the ovary who died of disseminated disease within 7 months after the primary surgery despite extensive surgery and adjuvant chemotherapy.

## Case Report

A 50-year-old woman, gravida 3 para 2, presented with abdominal distension for 1 month. Physical

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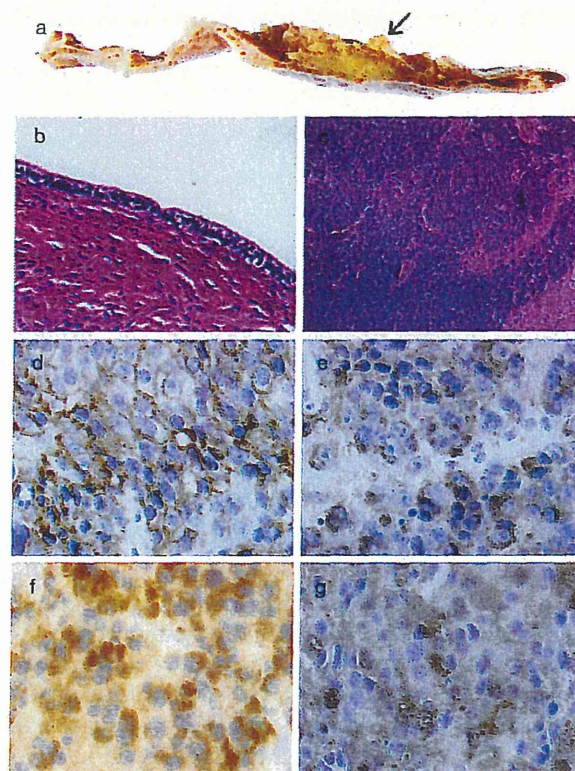
Reprint request to: Professor Kei Kawana, Department of Obstetrics and Gynecology, Graduate School of Medicine, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8655, Japan. Email: kkawana-tky@umin.org

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examination revealed a firm mass in the lower abdomen that was equivalent to the head of a newborn infant. Ultrasonographic tomography, magnetic resonance imaging and computed tomography (CT) demonstrated a monocular cyst measuring 15 cm × 12 cm × 10 cm in diameter in the abdominal cavity. Abdominal organs were otherwise normal and the lungs were normal. Preoperative serum level of carbohydrate antigen (CA)125, CA19-9, carcinoembryonic antigen,  $\alpha$ -fetoprotein and lactate dehydrogenase were within normal ranges. A smooth-surfaced right ovarian tumor was found at laparotomy. The uterus, tubes, left ovary and omentum were normal and no other tumor was evident in the abdominal cavity. Total abdominal hysterectomy, bilateral salpingo-oophorectomy, omentectomy and pelvic lymphadenectomy were performed. A smooth round tumor 15 cm in diameter was resected without rupture. In the tumor wall, several thickened lesions were seen (Fig. 1a, arrowhead). Hematoxylin-eosin staining of the tumor revealed microscopically mucinous epithelium lining the inside of the tumor wall and several parts relevant to the thickened lesions were composed of poorly differentiated large cells forming a front of the normal mucinous epithelium (Fig. 1b,c). To assess features of the large cells, immunohistochemistry was performed. Positivity for CD56, chromogranin A, synaptophysin and neuron-specific enolase (NSE) demonstrated the large cells were neuroendocrine components (Fig. 1d–g). Taken together, the ovarian tumor was classified as LCNEC associated with mucinous adenoma. Remarkable vascular invasion was observed in the tumor wall, especially around the thickened lesions.

She was diagnosed as having ovarian cancer of stage Ia (pT1aN0M0) of LCNEC associated with mucinous adenoma and received EP (cisplatin 75 mg/m<sup>2</sup> and etoposide 100 mg/m<sup>2</sup> once a day for 5 days) as an adjuvant chemotherapy. However, multiple liver metastases were detected by CT at 4 months after her primary operation when she received three rounds of EP. Then, her chemotherapy regimen was changed to TC (paclitaxel 175 mg/m<sup>2</sup> and carboplatin AUC 6) or CPT-11 (90 mg/m<sup>2</sup>) alone as second- or third-line chemotherapy, respectively. Clinical responses to any regimen were not observed in this case. Her cancer was thought to be refractory for any regimen and progressed rapidly. Unfortunately, she died of progressive disease only 7 months after her primary operation.



**Figure 1** Pathological and immunohistochemical findings of this case. (a) Macroscopic finding: the cut surface of the right ovary was smooth. Several thickened lesions were seen in the tumor wall (arrow). (b) Microscopic finding (hematoxylin–eosin [HE] staining): mucinous epithelium lined inside of the tumor wall (original magnification, ×100). (c) Microscopic finding (HE staining): poorly-differentiated large cells in several parts relevant to the thickened lesions (×40). (d–g) Immunohistochemical studies of this case for representative neuroendocrine markers: CD56 (d: ×400), chromogranin A (e: ×400), synaptophysin (f: ×400) and neuron-specific enolase (g: ×400).

## Discussion

Large cell neuroendocrine carcinoma of the ovary is included in the WHO tumor classification. To date, 35 cases of ovarian LCNEC have been reported in the published work. Thirty-three cases received an operation and been diagnosed postoperatively as ovarian LCNEC. These included 16 cases of stage I, three cases of stage II, eight cases of stage III and six cases of stage IV. Most of these cases seem to be associated with benign ovarian epithelial neoplasms. Of 33 cases, pure neuroendocrine carcinoma accounted for only four



cases while the remaining neuroendocrine carcinomas coexisted with ovarian epithelial tumors or germ cell tumors.

The tumor in our case was lined with benign epithelium mucinous adenoma. On the other hand, most of the tumor consisted of a poorly differentiated component of large tumor cells exhibiting proliferation in the thickened wall. Nuclei were large and had prominent nucleoli. Vascular invasion was prominent. Immunohistochemistry revealed that the cells of the poorly differentiated component were diffusely positive for CD56, chromogranin A, synaptophysin and NSE. Taken together, the tumor fulfilled the histopathological criteria for a neuroendocrine carcinoma.<sup>2</sup>

A high frequency of vascular invasion has been reported in neuroendocrine carcinomas of the lung and other sites.<sup>18</sup> In terms of gynecologic cancer, vascular invasion in LCNEC of the cervix has been reported to be much more common than in other types of cervical carcinomas.<sup>19</sup> Our case showed a definite vascular invasion at the lesions of LCNEC, which might have resulted in the distant metastasis in spite of the early stage.

Large cell neuroendocrine carcinoma in any organs is thought to be an aggressive tumor with high mortality despite extensive surgery and adjuvant chemotherapy. In our case, LCNEC spread to distant organs within 1 year after primary surgery despite the fact that the post-surgical stage was Ia and adjuvant chemotherapy was performed. However, one report mentions that cases of ovarian LCNEC, particularly those of stage I and/or those who have received platinum-based therapy, may have a favorable prognosis.<sup>13</sup> Then, we addressed the overall survival rate with ovarian LCNEC of stage I by Kaplan–Meier curve based on the published works (Fig. 2). Overall survival was available in 15 previously reported cases, with a median follow-up period of 9 months (including our study). Among these 16 cases, nine died of the disease within 3–19 months after primary operation and the remaining seven patients were alive at follow-up periods ranging 6–120 months. The median overall survival was 10 months and 1-year overall survival rate was 47.1% on the Kaplan–Meier curve. This suggests that the LCNEC of the ovary has a very poor prognosis even at stage I.

Veronesi *et al.* reported a series of 144 cases of lung LCNEC who received debulking surgery. Of them, 21 and 24 cases received neoadjuvant or postoperative adjuvant chemotherapy, respectively, and response rate of the chemotherapy was 80% in 15 cases with data

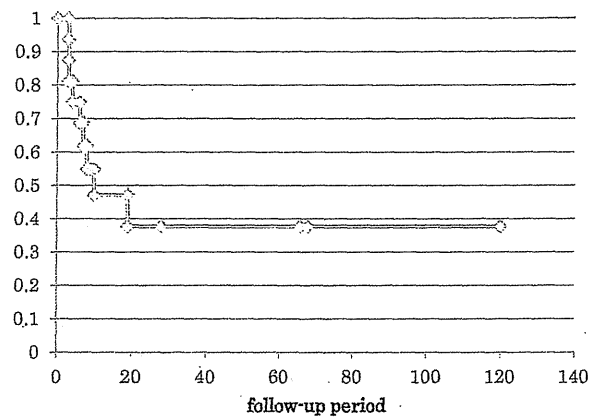


Figure 2 Kaplan–Meier curve of overall survival for ovarian large cell neuroendocrine carcinoma stage I. This case and 15 cases reported as stage I in the published work are summarized. One-year overall survival of these cases was 47.1% by Kaplan–Meier method.

available. They demonstrated that combination of chemotherapy and surgery improved overall survival of the lung LCNEC stage I with marginal significance when compared with surgery alone.<sup>20</sup> Another group also reported that two cases of lung stage I LCNEC who received cisplatin and etoposide combination chemotherapy survived for 2 and 5 years after complete pulmonary resection, respectively.<sup>21</sup> These data allowed for a possibility that platinum-based adjuvant chemotherapy after curative surgery may result in relatively long survival of the lung LCNEC cases of early stage. These rationales encouraged us to add combination adjuvant chemotherapy of cisplatin and etoposide to our case. However, our case died of disseminated disease within 7 months of the primary surgery. Chemoresponsiveness of the ovarian LCNEC to platinum-based chemotherapy may be lower than that of the lung LCNEC. Additional cases of this carcinoma will have to be collected to establish optimal adjuvant chemotherapy.

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# High-risk human papillomavirus correlates with recurrence after laser ablation for treatment of patients with cervical intraepithelial neoplasia 3: A long-term follow-up retrospective study

Kanako Inaba, Kazunori Nagasaka, Kei Kawana, Takahide Arimoto, Yoko Matsumoto, Tetsushi Tsuruga, Mayuyo Mori-Uchino, Shiho Miura, Kenbun Sone, Katsutoshi Oda, Shunsuke Nakagawa, Tetsu Yano, Shiro Kozuma and Tomoyuki Fujii

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## Abstract

**Aim:** The purpose of our study was to evaluate the efficacy of laser ablation as a conservative treatment for cervical intraepithelial neoplasia 3 (CIN3) and assess whether the human papillomavirus (HPV) test is useful to predict recurrence after treatment.

**Materials and Methods:** A total of 134 patients who received laser ablation for treatment of CIN3 were enrolled in this study. During the follow-up period, patients were followed with cytological and colposcopic evaluations. Recurrence of CIN3 was regarded as the primary end-point. HPV genotype was tested before and after treatment. Post-treatment cumulative recurrence rates were estimated and comparisons by both patient age and HPV genotype were performed.

**Results:** Overall cumulative recurrence rate of CIN3 in the first year after treatment was 22.6% for all patients. No significant correlation was shown between patient age and recurrence. Patients infected by specific genotypes (16, 18, 31, 33, 52, and 58) frequently failed to clear the infection after treatment. The 1-year recurrence-free survival in those positive after treatment for eight high-risk genotypes (16, 18, 31, 33, 35, 45, 52, and 58) was significantly lower (66.7%), compared to that in those positive for other high-risk types (78.6%). The recurrence-free survival of those who remained HPV-positive after treatment was significantly lower than those who turned negative.

**Conclusion:** Laser ablation should be performed prudently with appropriate patient counseling about recurrence rate. Considering its minimal invasiveness, laser ablation is effective, especially for young patients who are negative for eight high-risk genotypes. With regard to HPV testing, although genotyping has significant value for predicting recurrence, screening for all genotypes warrants further evaluation.

**Key words:** cervical intraepithelial neoplasia 3, human papillomavirus testing, laser ablation, recurrence, treatment efficacy.

## Introduction

The spread of systematic screening programs has detected more cervical intraepithelial neoplasia

(CIN) and has succeeded in producing marked declines in cervical carcinoma incidence and mortality in the developed countries where screening programs and treatment for pre-invasive lesions are

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### Introduction

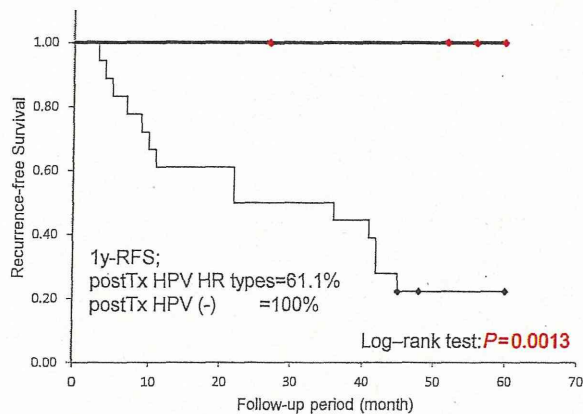
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**Figure 5** Recurrence-free survival (RFS) between post-treatment human papillomavirus (HPV) genotypes. Post-treatment 1-year RFS for those positive for 13 high-risk HPV was 61.1%, versus 100% for those who were HPV-negative. It is unsurprising that no recurrence was seen in any HPV-negative cases after laser ablation. —, negative; —, high-risk types.

combined], and cryotherapy), reporting that overall cumulative rate of CIN2/3 recurrence in the first 6 years after treatment of 14.0% for women originally treated for CIN3. Estimated rates of CIN2/3 after treatment were lowest for cold-knife conization, followed by LEEP and laser, and highest for cryotherapy. This result corroborates the invasiveness of the respective procedure.

Further evidences are warranted for the treatments of CIN3 though; considering many women with high-grade CIN are of reproductive age, treatment should not only be effective but also have minimum adverse effects on future fertility and obstetric outcomes. Several previous studies have clearly demonstrated that excisional treatments result in long-term adverse obstetric outcomes, including preterm delivery, low birthweight, or premature rupture of membranes; thus, an increasing number of patients are likely to choose less invasive treatment like laser ablation.<sup>5</sup>

The data in our study suggest that laser ablation is an optional conservative treatment for CIN3 with minimal invasiveness. Regarding obstetric outcomes, it is difficult to follow patients after CIN treatment in terms of pregnancy because of the premalignant nature of the lesion, so no data were available in this study. Nevertheless, as laser ablation has thus far been shown to be free of any adverse effect on obstetric outcomes, it should be taken into consideration especially for young patients of child-bearing age. Even for lesions, previ-

sional use of laser ablation may provide a 'grace period' for young women with desire to bear children before more invasive interventions are performed. Therefore, many women can postpone, or even entirely avoid, the unnecessary adverse outcomes associated with invasive intervention.

With regard to follow-up surveillance after treatment of high-grade CIN, the role of HPV-DNA testing has been an area of interest due to its high sensitivity and negative predictive value (NPV) for detecting recurrent disease. In a meta-analysis of 11 studies by Zielinski *et al.*,<sup>12</sup> sensitivity, NPV, and specificity of high-risk HPV testing for recurrence were 91% (95% CI = 86–95), 98% (97–99), and 79% (76–82), respectively. That study demonstrated that combining cytology with high-risk HPV testing increases rates even further: sensitivity was 96% (89–99), NPV was 99% (98–100), and specificity was 81% (77–84). Specificity is indeed relatively low, but considering that this is not a screening test but rather a follow-up for a potentially lethal disease, NPV and sensitivity should be valued higher than specificity.

We demonstrated in this study that HPV infection is not likely to disappear, but persists after laser ablation in women positive for eight 'higher-risk' HPV. Considering that cervical cancer is caused by persistent infection with a subset of carcinogenic HPV, this failure of viral clearance should lead to the increase in cervical cancer in future. Indeed, we confirmed the significant difference in RFS between those who were positive for the eight higher-risk types of post-treatment versus those who were positive for other types or altogether negative. This result supports the efficacy of HPV testing and is consistent with several previous studies.

However, several critical problems with HPV-PCR testing remain. First, while PCR testing can distinguish individual genotype, detection sensitivities differ greatly among HPV genotypes. Furthermore, routine PCR-based HPV DNA testing is not suitable for daily clinical use due to its high cost and complex technique using specific primers. For the purposes of this study context, therefore, we recommend commercially available HPV testing using the hybrid capture method, Hybrid Capture II (HC2), which targets a group of 13 high-risk HPV genotypes but does not distinguish which genotypes are present. This intensive test also represents well the eight higher-risk genotypes; identification of the existence of high-risk HPV by HC2 may sufficiently inform the likelihood of recurrence. In Japan, HC2 has been covered by insurance since 2010, but the target of this test is only limited for patients

Table 2 Detection rate of HPV after treatment for each HPV type before treatment

HPV type before treatment	Detection of HPV after treatment†	
	Positive (%)	Negative (%)
16/18	66.6	33.3
31/33/52/58	52.4	47.6
Others	25.0	75.0

†Cochran–Armitage test  $P < 0.0001$ . Detection rates – i.e. whether the same HPV type was detected before and after laser ablation, which can be interpreted as a good indicator of disease clearance. HPV was persistently detected after treatment more often in the higher-risk types, especially in types 16 and 18, than in other HPV types (16/18 vs 31/33/52/58 vs others; 66.6% vs 52.4% vs 25.0%;  $P < 0.0001$  by Cochran–Armitage test). HPV, human papillomavirus.

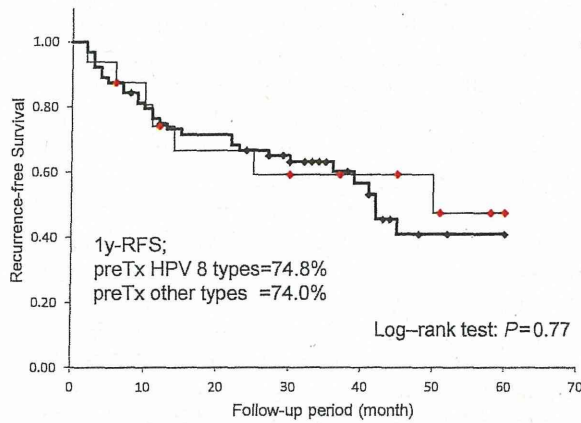


Figure 3 Recurrence-free survival (RFS) by pre-treatment human papillomavirus (HPV) genotype. We compared RFS between those who were positive for the 8 higher-risk types versus those positive for other HPV types prior to ablation; no significant difference between the two groups was identified. —, 8 types; ---, Others.

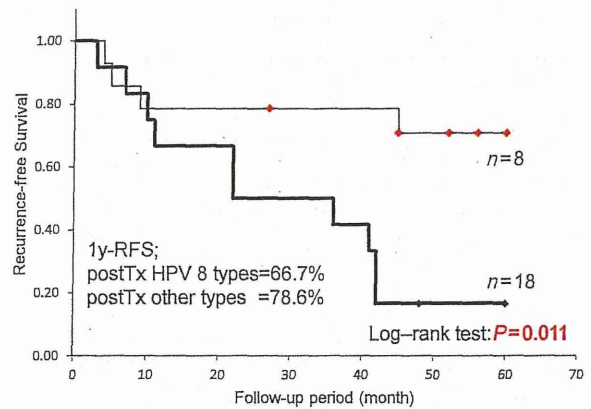


Figure 4 Recurrence-free survival (RFS) by post-treatment human papillomavirus (HPV) genotype. After ablation, 1-year RFS was 66.7% for those who were positive for the 8 higher-risk HPV types, which was significantly lower than the 78.6% observed in those positive for other HPV types ( $P = 0.011$  by log-rank test). —, 8 types; ---, Others.

compared RFS by post-treatment HPV genotypes. As shown in Figure 4, 1-year RFS was 66.7% for those who were positive for the 8 higher-risk types after ablation, which was significantly lower than the 78.6% observed in those positive for other HPV types ( $P = 0.011$  by log-rank test). Furthermore, Figure 5 shows that the 1-year RFS for those positive for all high-risk 13 types of HPV after treatment was 61.1%, versus 100% in HPV-negative subjects ( $P = 0.0013$  by log-rank test).

All these data suggest that the persistence of high-risk HPV genotypes, especially of the eight types, might play a key role in the development of recurrence.

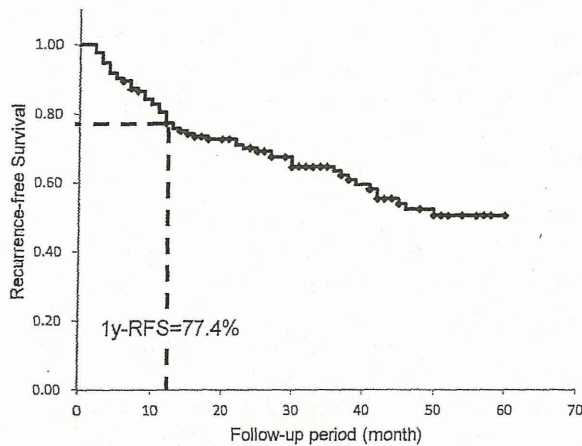
### Discussion

In 2011, a large Dutch multi-cohort study, one of the representative prospective studies on long-term

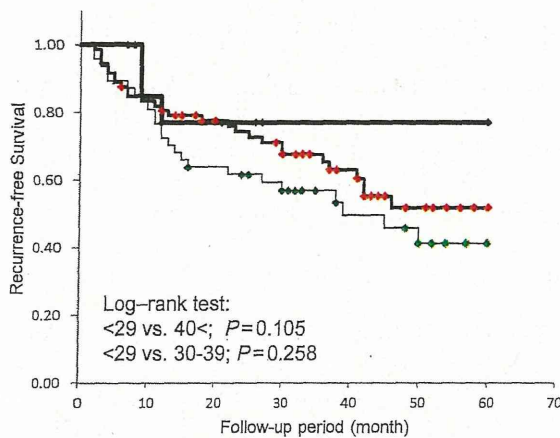
efficacy of CIN treatment, reported on recurrence risk after treatment for CIN2/3.<sup>16</sup> According to that study, the recurrence risk of CIN was quite low after highly successful treatment followed by normal cytology or negative HPV testing. However, all participants were treated by excisional methods, such as cold-knife conization or LEEP, so the data did not present any information on the prognostic outcomes of conservative treatments, including laser ablation.

As mentioned, many studies have demonstrated that conservative methods, including cold-knife conization, LEEP, cryotherapy, and laser ablation, show similarly low morbidity and are equally successful at preventing invasive cervical cancer.<sup>4,17,18</sup> However, in 2009, Melnikow *et al.*<sup>10</sup> compared the long-term follow-up of 37 142 patients with CIN by treatment modality (cold-knife conization, LEEP, laser [conization and ablation





**Figure 1** Overall cumulative recurrence rate after treatment. The overall cumulative recurrence rate of cervical intraepithelial neoplasia 3 in the first 12 months after treatment was 22.6%. RFS, recurrence-free survival.



**Figure 2** Recurrence rates compared by age. No significant elevated risk of recurrence was seen between younger age (<29) and older age (<29 vs >40;  $P = 0.105$ , <29 vs 30-39;  $P = 0.258$ , by log-rank test). —, <29; —, 30-39; —, 40+.

(Fig. 1). Figure 2 shows recurrence rates compared by age. It is worth noting that no significant elevated risk of recurrence was seen between younger age (<29) and older age (<29 vs >40,  $P = 0.105$ ; <29 vs 30-39,  $P = 0.258$ , by log-rank test). Among recurrence cases, 27 patients underwent second laser ablation, nine patients underwent cervical conization, and one patient underwent total hysterectomy. Other patients were followed up with intensive cytological and colposcopic evaluations

**Table 1** Distribution of HPV type before treatment

HPV type before treatment	Rate (%)
16	34
18	2
31	2
33	5
52	20
58	13
Others	24

The HPV genotypes of 83 patients were identified by polymerase chain reaction-based HPV DNA testing both before and after ablation. Seventy-six women were positive on HPV-DNA testing (91.6%). The distribution of HPV genotypes was classified according to the Matsumoto criteria of 8 'higher-risk' types. Two of the higher-risk types (35, 45) were not detected in this study. All cervical intraepithelial neoplasia 3 cases in this study had mono-infection. HPV, human papillomavirus.

with informed consent. The re-recurrence rate of CIN3 after second ablation was 11.1%.

Next, we evaluated the association between HPV genotype and efficacy of laser ablation, assessing whether HPV genotyping might predict failure after laser ablation. Of the 134 patients, we examined the HPV genotype of 83 patients, identified by PCR-based HPV DNA testing both before and after ablation. Single infection was observed in all CIN3 cases in this study, although there is a possibility that some patients might have multiple infections, especially in the CIN1-2 population. The median follow-up period was 17 months, with a minimum follow-up of 6 months. The median age of these 83 patients was 36 years; 76 women were positive on HPV-DNA testing (91.6%). The distribution of HPV genotypes, classified according to the Matsumoto criteria of eight 'higher-risk' types, before laser ablation is shown in Table 1.

Table 2 shows detection rates – that is, whether the same HPV type was detected before and after laser ablation, which can be interpreted as a good indicator of disease clearance. HPV was persistently detected after treatment more often in the higher-risk types, especially in type 16 and 18, than in other HPV types (16/18 vs 31/33/52/58 vs others; 66.6% vs 52.4% vs 25.0%;  $P < 0.0001$  by Cochran-Armitage test).

In addition, we compared post-treatment RFS by HPV genotypes. As shown in Figure 3, we first compared RFS by pre-treatment HPV genotypes, that is, between those who were positive for the 8 higher-risk types and those positive for other HPV types before ablation; no statistically significant difference was seen between the two ( $P = 0.77$  by log-rank test). We then