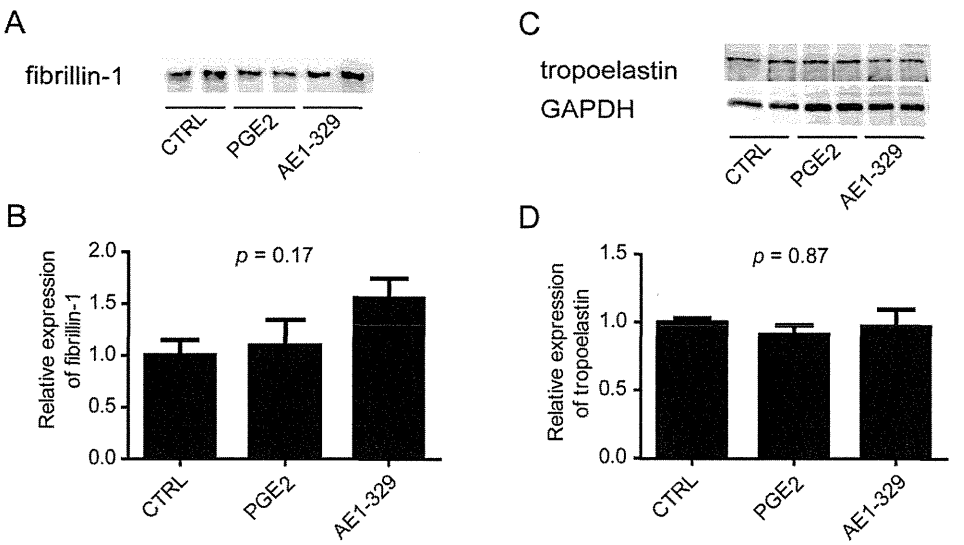
Supplemental Figure 1



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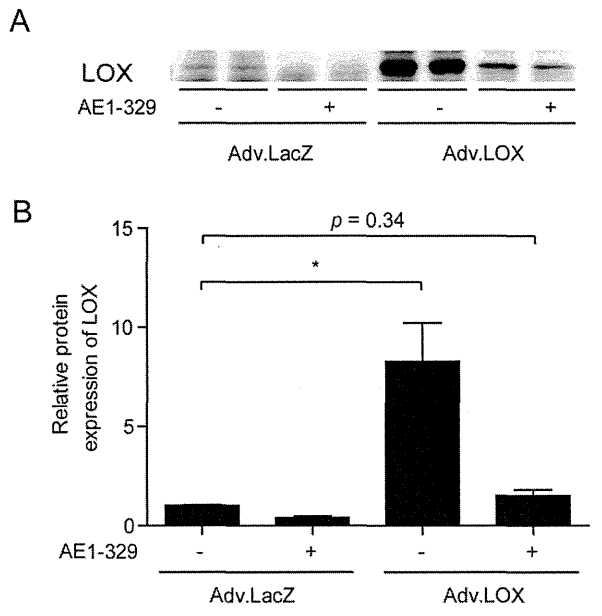

Supplemental Figure 3

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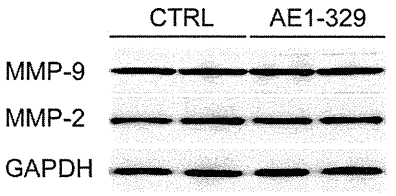

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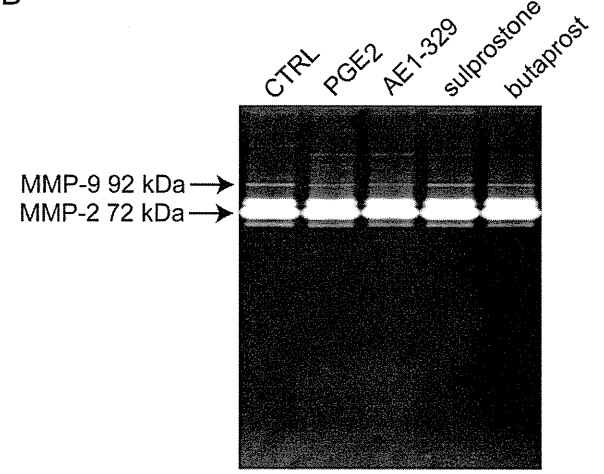

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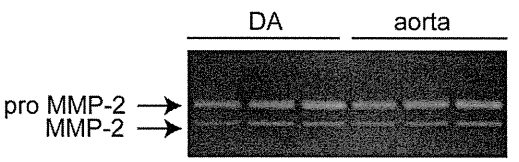

Supplemental Figure 4

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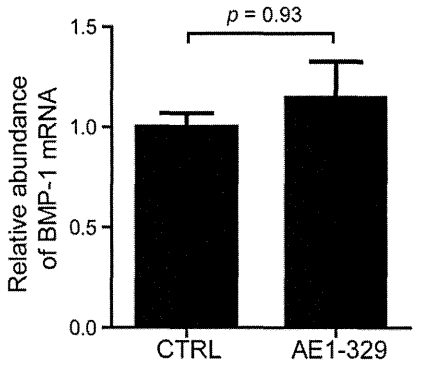

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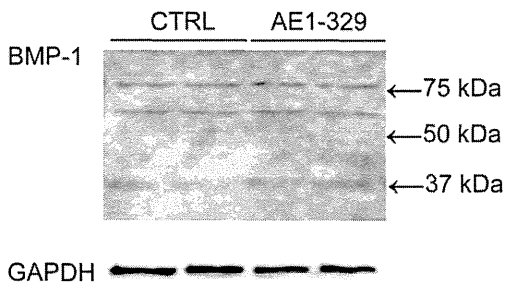
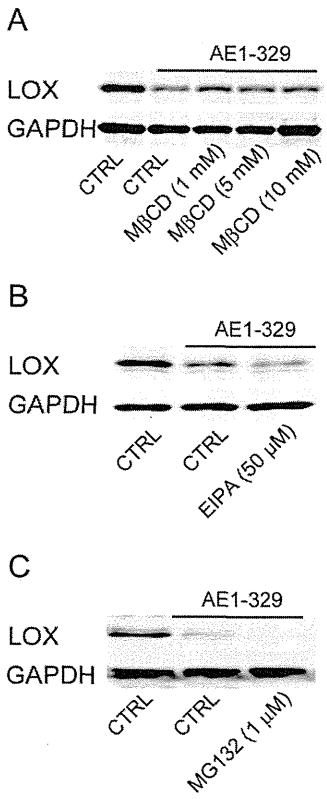


Supplemental Figure 5



Supplemental Figure Legends

Supplemental Figure 1

EP4 signaling did not affect protein expression of tropoelastin and fibrillin-1.

(A) Protein expression of fibrillin-1 in culture medium of DASMCs treated with either PBS, PGE_2 (1 μM), or AE-329 (1 μM) for 72 h. (B) Quantification of (A), n = 4. (C) Protein expression of tropoelastin in whole cell lysate of DASMCs treated with either PBS, PGE_2 (1 μM), or AE-329 (1 μM) for 72 h. (D) Quantification of (C), n = 4.

Supplemental Figure 2

Overexpression of LOX protein in DASMCs transfected with Adv.LOX.

(A) Protein expression of LOX in culture medium of DASMCs transfected Adv.LacZ or Adv.LOX in the presence or absence of AE-329 (1 μM). The time-course of transfection and drug administration was same as Figure 4I. (B) Quantification of (A), n = 4, $*p < 0.05$.

Supplemental Figure 3

EP4 signaling did not affect expression or activation of MMP-2 or -9 in DASMCs.

(A) Protein expression of MMP-2 and-9 in DASMCs treated with or without AE1-329 (1 μM) for 72 h. (B) Gelatin zymography of DASMCs treated with 1 μM of PGE2 or each EP agonist. (C) Gelatin zymography of the rat DA and aorta on the 21st day of gestation.

Supplemental Figure 4

EP4 signaling did not change BMP-1 expression in DASMCs.

(A) Expression of BMP-1 mRNA in DASMCs treated with or without AE1-329 (1 μM) for 24 h. n = 4. (B) Representative image of protein expression of BMP-1 in DASMCs treated with or without AE1-329 (1 μM) for 72 h.

Supplemental Figure 5

LOX degradation was associated with caveolar endocytosis, macropinocytosis, and proteosome.

(A) Representative figures of protein expression of LOX in whole cell lysate of DASMCs treated with MbCD, EIPA, or MG132 in the presence of AE1-329 (1 μM).

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Three-dimensional multilayers of smooth muscle cells as a new experimental model for vascular elastic fiber formation studies

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ABSTRACT

Objective: Elastic fiber formation is disrupted with age and by health conditions including aneurysms and atherosclerosis. Despite considerable progress in the understanding of elastogenesis using the planar culture system and genetically modified animals, it remains difficult to restore elastic fibers in diseased vessels. To further study the molecular mechanisms, in vitro three-dimensional vascular constructs need to be established. We previously fabricated vascular smooth muscle cells (SMCs) into three-dimensional cellular multilayers (3DCMs) using a hierarchical cell manipulation technique, in which cells were coated with fibronectin-gelatin nanofilms to provide adhesive nano-scaffolds. Since fibronectin is known to assemble and activate elastic fiber-related molecules, we further optimized culture conditions.
Methods and results: Elastica stain, immunofluorescence, and electron microscopic analysis demonstrated that 3DCMs, which consisted of seven layers of neonatal rat aortic SMCs cultured in 1% fetal bovine serum (FBS) in Dulbecco's modified Eagle's medium, exhibited layered elastic fibers within seven days of being in a static culture condition. In contrast, the application of adult SMCs, 10% FBS, ε-poly(-lysine) as an alternative adhesive for fibronectin, or four-layered SMCs, failed to generate layered elastic fiber formation. Radioimmunoassay using [^{3}H]valine further confirmed the greater amount of cross-linked elastic fibers in 3DCMs than in monolayered SMCs. Layered elastic fiber formation in 3DCMs was inhibited by the lysyl oxidase inhibitor β-aminopropionitrile, or prostaglandin E_2. Furthermore, infiltration of THP-1-derived macrophages decreased the surrounding elastic fiber formation in 3DCMs.
Conclusion: 3DCMs may offer a new experimental vascular model to explore pharmacological therapeutic strategies for disordered elastic fiber homeostasis.

1. Introduction

Arterial walls have a highly organized layer structure that consists of various cells and extracellular matrix (ECM) components. In particular, the vascular media is composed of a dense population of concentrically organized smooth muscle cells (SMCs) and elastic fibers, and plays a pivotal role in maintaining sufficient blood pressure, even during variations in hemodynamic stress. In physiological conditions, SMCs synthesize elastin and other specific molecules, which are incorporated into elastic fibers and arranged into concentric rings of elastic lamellae around the arterial media [1]. In contrast, arterial compliance and distensibility are impaired in the presence of cardiovascular disease and risk factors such as aortic aneurysm, atherosclerosis, ischemic heart disease, aging, hypertension, cigarette smoking, and diabetes [2]. Hence, impaired elastic properties are associated with arterial dysfunction and pathophysiology [2,3].

Changes in arterial elastic properties are the result of alterations in the intrinsic structural properties of the artery, including the fracturing and thinning of elastic fibers. Current approaches to

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examining the elastogenesis and degradation of elastic fibers rely heavily on the use of the planar culture of vascular SMCs and genetically modified animals. These approaches have been instrumental in numerous discoveries and have been modified to create very elegant experimental designs [1,4–7]. Currently, however, no pharmacological strategy to promote elastogenesis and prevent the degradation of elastic fiber formation is available, and the molecular mechanisms of the regulation of elastic fiber formation remain to be studied. The two-dimensional (2D) monolayer culture system is a useful method for isolating specific factors and their effects on specific cell types [5–7], but it lacks the native-like layered structure of elastic fibers. Therefore, changes in the spatial arrangement of elastic fibers induced by various stimuli and the infiltration of immune cells cannot be observed. In-vivo analysis, on the other hand, often fails to discriminate among the various and complex factors. In this context, in vitro reconstruction vessel models overcoming these limitations are considered potential platforms of vascular biology that can provide further insights into the spatiotemporal molecular mechanisms of elastic fiber formation.

We previously developed a novel three-dimensional (3D) cell construction method [8] and created 3D-layered blood vessel constructs consisting of human umbilical arterial SMCs and human umbilical vascular endothelial cells [9]. To develop the 3D-cellular multilayers (3DCMs), we fabricated nanometer-sized cell adhesives like ECM scaffolds onto the surface of a cell membrane, which enables another cellular layer to adhere to the coated cell surface. We employed a layer-by-layer (LbL) technique to fabricate fibronectin–gelatin nanofilms onto living cell membranes, because the LbL technique produces nanometer-sized polymer films with a controllable nanometer thickness through the alternate immersion into interactive polymer solutions. We found that approximately 6 nm of fibronectin-based nanofilms were suitable for developing stable adhesive scaffolds and for creating allogeneic or xenogeneic multiple cell layers.

In addition to the cell adhesion effect, fibronectin has been known to orchestrate the assembly of the ECM [10–15]. In particular, recent reports suggest that fibronectin fiber assembles pericellularly into fibrillin microfibrils that have a complex structural organization and are widespread in elastic tissues [10,16]. Furthermore, fibronectin binds to lysyl oxidase (LOX), a cross-linking enzyme for elastic fibers, and acts as a scaffold for enzymatically active 30 kDa LOX [14]. Using scanning electron microscopy and transmission electron microscopy, we observed that fibronectin formed extracellular fibrils in the abovementioned 3DCMs within 24 h cultures [9], suggesting that fibronectin-coated SMCs have the potential to produce elastic fiber assemblies. In this context, we aimed to create the first scaffold-free 3D cellular multilayers (3DCMs) that are specifically designed for investigating the spatial regulation of vascular elastic fibers by employing this LbL assembly technique. The present study demonstrates that the optimized culture conditions provided layered elastic fiber formation in the 3DCMs consisting of neonatal rat SMCs within seven days of static culture conditions. In the 3DCMs, it was observed in the vertical view that macrophage infiltration or prostaglandin E_2 (PGE_2) changed the spatial arrangement of elastic fibers.

2. Materials and methods

Expanded materials and methods are described in Supplemental data.

2.1. Animals

Neonate (day 1) and adult Wistar rats (4–5months old) were obtained from Japan SLC, Inc. (Shizuoka, Japan). All animal studies were approved by the institutional animal care and use committees of Yokohama City University.

2.2. Cell culture

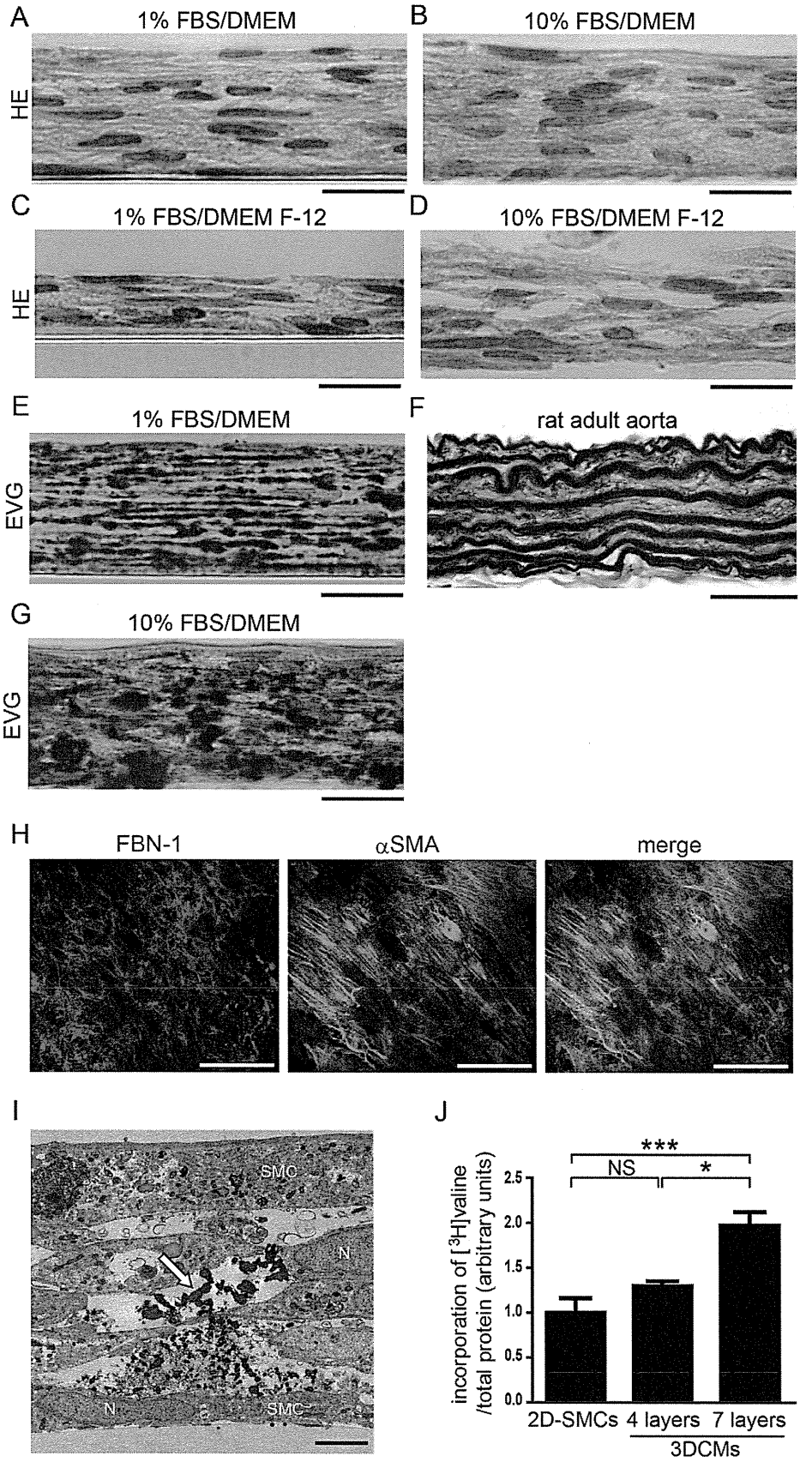
Vascular SMCs in primary culture were obtained from the aorta of rat neonates (day 1) as previously described [17–19]. Briefly, the minced tissues were digested with a collagenase-dispase enzyme mixture at 37 °C for 20 min. The cell suspensions were then centrifuged, and the medium was changed to a collagenase II enzyme mixture. After 12 min of incubation at 37 °C, cell suspensions were plated on 35 mm poly-L-lysine-coated dishes The growth medium contained Dulbecco's modified Eagle medium (DMEM) with 10% fetal bovine serum (FBS), 100 U/ml penicillin, and 100 mg/ml streptomycin (Invitrogen, Carlsbad, CA). Human adult aortic SMCs were obtained from Lonza (Walkersville, MD, USA). The confluent SMCs were used at passages 5–7. THP-1 cells were obtained from the Health Science Research Resources Bank (Osaka, Japan) and were maintained in RPMI 1640 medium (Wako, Osaka, Japan) supplemented with 10% FBS. All cells were cultured in a moist tissue culture incubator at 37 °C in 5% CO_2-95% ambient mixed air.

2.3. Construction of 3DCMs

Construction of 3DCMs was performed as previously described [8]. Briefly, a cell disk LF (Sumitomo Bakelite, Tochigi, Japan) was rinsed with 50 mM Tris–HCl buffer solution (pH 7.4) for 15 min and coated with fibronectin (0.2 mg/ml) for 30 min at 37 °C. SMCs were cultured on the cell disk (11×10^4 cells/cm^2) and incubated for 12 h in 10% FBS/DMEM. The monolayered SMCs were then immersed alternatively in a solution of fibronectin (0.2 mg/ml) and gelatin (0.2 mg/ml). After nine steps of LbL assembly with fibronectin and gelatin, a second cell layer was seeded on the first cell layer (11×10^4 cells/cm^2) and incubated for 6–12 h at 37 °C. The cycles of LbL nanofilm assembly and subsequent cell seeding were repeated six times in four days to construct seven-layered 3DCMs. During the first four days, the medium was refreshed daily. Twelve hours after the seeding of the last layer, the culture media was changed to DMEM or DMEM/F-12 (Gibco, Carlsbad, CA) containing 1% FBS or 10% FBS. The 3DCMs were incubated for an additional 48 h and either fixed in buffered 10% formalin or harvested in TRIzol (Invitrogen, Carlsbad, CA). The time-course of the 3DCM experiments is shown in Supplemental Fig. 1A. Stimulation by β-aminoproprionitrile (BAPN) or PGE_2 (1 μM) was performed simultaneously with the medium change on day 5. For macrophage infiltration assay, seven-layered 3DCMs at day 5 were put on a 24-well plate and applied with 500 μl of RPMI 1640 medium containing THP-1 cells (2.0×10^5 cells) with or without phorbol 12-myristate 13-acetate (PMA, 0.1 μM). The 3DCMs were incubated for 1 h at 37 °C. Next, unattached THP-1 cells were washed out with PBS and 3DCMs were incubated in 1% FBS/DMEM for 72 h until fixation. Control monolayered neonatal rat aortic SMCs (2D-SMCs) were plated on day 4 in the same density as a single layer in 3DCMs (11×10^4 cells/cm^2), and were incubated for 48 h. Four-layered 3DCMs were constructed from day 3 following the same time-course. The proportional increase in the thickness of 3DCMs with the number of seeding events was shown in Supplemental Fig. 1B. To confirm the effect of fibronectin nanofilms, ε-poly(lysine) (0.2 mg/ml) was used as an alternative adhesive polymer.

2.4. Quantitative measurement of insoluble elastin

Newly synthesized insoluble elastin was measured as previously described [20,21]. Briefly, after the seven-layered 3DCMs and 2D-


A
1% FBS/DMEM
B
10% FBS/DMEM
HE
C
1% FBS/DMEM F-12
D
10% FBS/DMEM F-12
HE
E
1% FBS/DMEM
F
rat adult aorta
EVG
G
10% FBS/DMEM
EVG
H
FBN-1
αSMA
merge
I
SMC
N
N
SMC
J
***
NS
*
incorporation of [3H]valine /total protein (arbitrary units)
2.5
2.0
1.5
1.0
0.5
0.0
2D-SMCs
4 layers
7 layers
3DCMs

SMCs were constructed for four days and one day, respectively, as mentioned above, they were incubated in 2 ml of 1% FBS/DMEM in the presence of 5 μCi [^{3}H]valine (ART 0466, American Radiolabeled Chemicals, Inc., St. Louis, MO) for an additional three days. One day after the monolayered SMCs were plated, the cells were harvested in 0.1 M acetic acid on ice. After centrifugation, the pellets were boiled in 0.1 N NaOH for 1 h. The NaOH-insoluble fractions were collected by centrifugation and were boiled in 5.7 N HCl for 1 h. The samples mixed with scintillation fluid were measured for radioactivity using a scintillation counter (Beckman Coulter).

2.5. Statistical analyses

Data are presented as the mean ± standard error of the mean (SEM) of independent experiments. Data were analyzed by the unpaired Student's *t*-test or one-way analysis of variance (ANOVA) followed by Newman–Keuls multiple comparison test. $p < 0.05$ was considered significant.

3. Results

3.1. 3DCMs exhibited layered elastic fiber formation

Most of the elastic fiber proteins are expressed in the second half of development and increase throughout the early postnatal period [22]. This is followed by a decrease in expression to low levels that persist through adulthood [23]. Consistent with this in vivo evidence, it has been suggested that neonatal rat SMCs produce a greater amount of elastin and elastic fibers compared to adult rat SMCs [24,25]. We confirmed that mRNA expression levels of elastic fiber–related genes, such as tropoelastin, fibrilin-1, fibulin-4, fibulin-5, LOX, and LOX-like 1, are significantly greater in the neonatal rat aorta (1 day old) than in the adult rat aorta (Supplemental Fig. 2).

On the basis of these data, we used neonatal rat SMCs to fabricate 3DCMs as in vitro vascular constructs. First, we cultured 3DCMs with four different culture media, including DMEM and DMEM F-12, because DMEM F-12 includes proline, one of the major amino acids that constitutes elastin protein, and copper, which enhances LOX activity [26]. Contrary to expectations, nucleus counting using HE stain revealed that viability was better in the 3DCMs cultured in DMEM (Fig. 1A–B, Supplemental Fig. 3) than in DMEM F-12 containing either 10% or 1% FBS (Fig. 1C–D). Next, we compared elastic fiber formation using Elastica van Gieson (EVG) stain in 3DCMs cultured in DMEM containing 10% or 1% FBS. We found that 3DCMs cultured in 1% FBS/DMEM exhibited layered elastic fiber formation (Fig. 1E), although the elastic fiber formation did not reach that observed in adult rat aorta (Fig. 1F) The 3DCMs culture in 10% FBS/DMEM exhibited a certain amount of positive stain of elastic fibers, but no layered elastic fiber formation was clearly detected (Fig. 1G). In the 3DCMs culture in 1% FBS/DMEM, co-immunofluorescence for fibrillin-1 and α-smooth muscle actin (αSMA) demonstrated a fiber meshwork of fibrillin-1 on the SMC surface (Fig. 1H). Electron microscopic analysis also confirmed the presence of elastic laminae between SMC layers (Fig. 1I).

To investigate whether the use of fibronectin nanofilms is required for elastic fiber formation in the 3DCMs, we applied ε-poly(lysine), a synthesized polymer that electrically binds to gelatin, as an alternative to fibronectin in the 3DCMs. We found that ε-poly(lysine) did not provide layered elastic fiber formation (Supplemental Fig. 4A). As expected, the 3DCMs composed of adult human aortic SMCs or adult rat aortic SMCs also did not exhibit elastic fiber formation (Supplemental Fig. 4B–C). Furthermore, we compared elastic fiber formation between the 3DCMs and another 3D model, i.e., the spheroid culture system. We constructed spheroids of neonatal rat SMCs with the same culture conditions of 3DCM construction (Supplemental Fig. 5A) and found that a few elastic laminae were formed in the outer shell of rat SMC spheroids, but not in the porous core (Supplemental Fig. 5B–C).

To quantify the amount of mature (i.e., cross-linked) elastic fibers in 3DCMs, we metabolically labeled newly synthesized elastin with [^{3}H]valine, and measured the incorporation of [^{3}H]valine in the NaOH-insoluble fraction of these cells, which reflects the amount of newly synthesized mature elastic fibers [27]. As shown in Fig. 1J, in 3DCMs cultured with 1% FBS/DMEM, we detected a significant increase in the incorporation of [^{3}H]valine into the insoluble fraction compared to 2D-SMCs cultured in the same culture medium. Interestingly, seven-layered 3DCMs produced a larger amount of cross-linked elastic fibers than four-layered 3DCMs even if the values were compared in the same amount of protein (Fig. 1J and Supplemental Fig. 4D). These data suggest that seven-layered 3DCMs consisting of fibronectin coating neonatal rat aortic SMCs cultured in 1% FBS/DMEM produced layered elastic fibers in vitro.

3.2. Expression of elastic fiber-related genes in 3DCMs

Elastic fibers are complex structures that contain elastin as well as microfibrils and fibulins [1]. Elastin is the major component of mature elastic fibers, and microfibrils, such as fibrillin-1 and fibrillin-2, are known to facilitate elastin assembly and provide overall structure to the growing elastic fiber [1]. Tropoelastin's lysine residues are, in turn, modified to form covalent cross-links between and within elastin molecules by LOX, resulting in the functional form of elastic fibers [1,26]. The deposition of fibulin-5 on microfibrils also promotes the coacervation and alignment of tropoelastins on microfibrils, and facilitates the cross-linking of tropoelastin by tethering LOX–like 1, 2, and 4 enzymes [20]. Fibulin-4 interacts with LOX and recruits LOX to elastin monomers as well [28].

We examined the mRNA expression levels of these genes in the 3DCMs cultured in DMEM containing 1% or 10% FBS and 2D-SMCs. The expression levels of tropoelastin and fibrillin-2 mRNAs were greater in the 3DCMs cultured in 1% FBS/DMEM than in 10% FBS/DMEM (Fig. 2A–B). The expression levels of other genes, including fibrillin-1, fibulin-4, fibulin-5, LOX, and LOX-like 1, were similar among different serum concentrations (Fig. 2C–G). Similar serum withdrawal effects on tropoelastin and fibrillin-2 mRNA were also observed in 2D-SMCs (Supplemental Fig. 6A–B). In 3DCMs cultured in 1% FBS/DMEM, abundant protein expressions of tropoelastin and LOX were also observed (Fig. 2H–I). However, there was no difference in the expression of elastic fiber-related genes between 2D-SMCs and 3DCMs (Fig. 2A–G, J–K). These data

Fig. 1. 3DCMs cultured in 1% FBS/DMEM exhibited layered elastic fibers. (A–B) Hematoxylin eosin (HE) stain of 3DCMs cultured in DMEM containing either 1% (A) or 10% FBS (B). HE stain of 3DCMs cultured in DMEM F-12 containing either 1% (C) or 10% FBS (D). (E) Elastica van Gieson (EVG) stain of 3DCMs cultured in 1% FBS/DMEM. Scale bars: 10 μm. (F) EVG stain of rat adult aorta. Scale bar: 50 μm. (G) EVG stain of 3DCMs cultured in 10% FBS/DMEM. Scale bar: 10 μm. (H) Confocal images of the fibrillin-1 and αSMA immunofluorescence in the 3DCMs cultured in 1% FBS/DMEM. red: fibrillin-1. Green: αSMA. Scale bar: 50 μm. (I) Electron microscopic image of 3DCMs. White arrow indicates elastic fibers. N: nucleus. Scale bar: 2.0 μm. (J) Radioimmunoassay of elastogenesis in 2D-SMCs, four-layered 3DCMs, and seven-layered 3DCMs cultured in 1% FBS/DMEM. $n = 4–8$. $^*p < 0.05$; $^{***}p < 0.001$; NS: not significant.

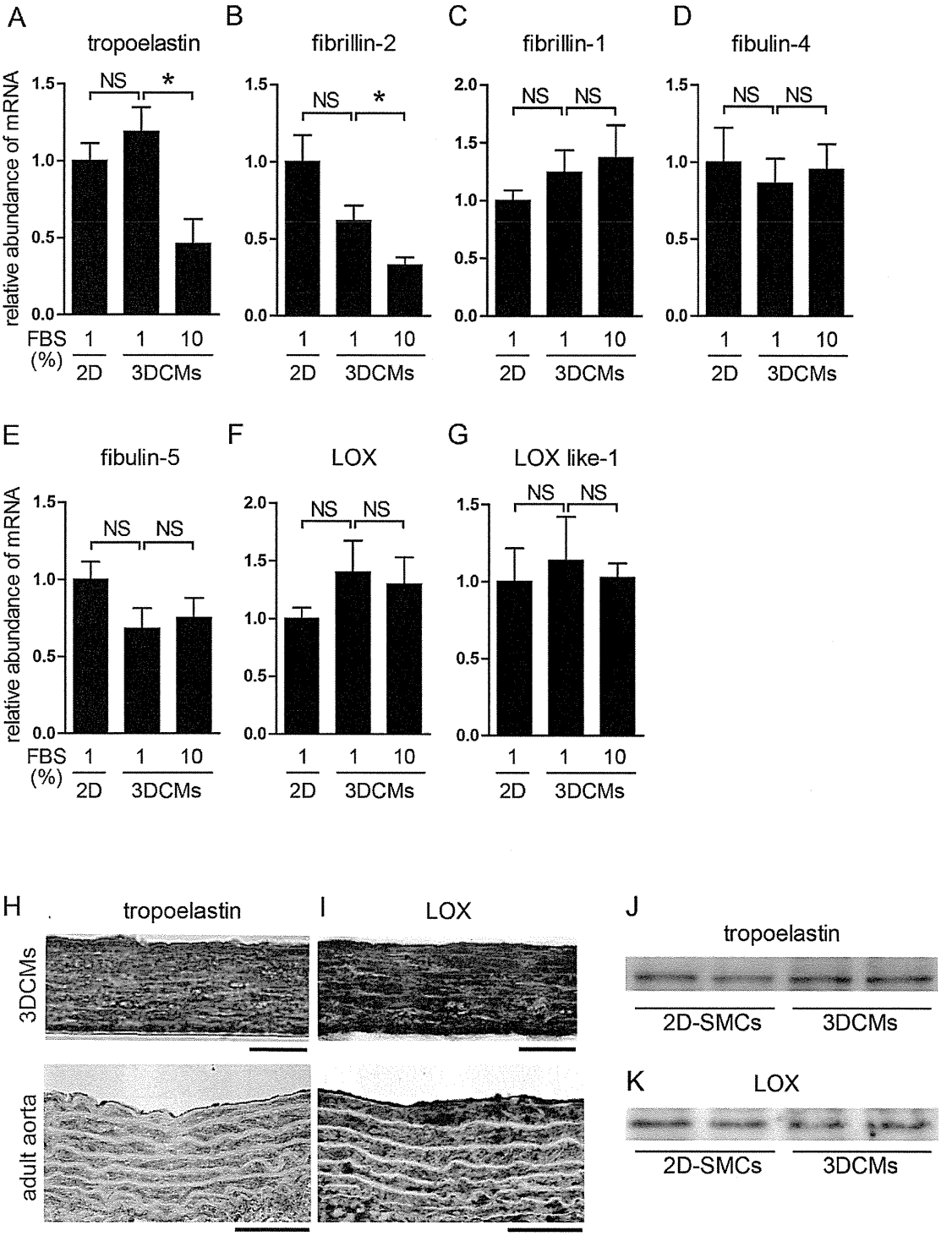


Fig. 2. Expressions of elastic fiber–related genes in 3DCMs. (A–G) Expressions of tropoelastin (A), fibrillin-2 (B), fibrillin-1 (C), fibulin-4 (D), fibulin-5 (E), LOX (F), and LOX–like 1 (G) mRNA were compared between 2D-SMCs (2D) cultured in 1% FBS/DMEM and 3DCMs cultured either in 1% FBS/DMEM or 10% FBS/DMEM. $n = 6–8$. $^*p < 0.05$. NS: not significant. (H–I) Immunohistochemistry for tropoelastin (H) and LOX (I). Brown color indicates positive staining. top panels: 3DCMs. Scale bars: 10 μm lower panels: rat adult aorta. Scale bars: 50 μm. (J–K) Immunoblot analysis for elastin (J), and LOX (K) in 2D-SMCs and 3DCMs.

suggest that the 3D-SMC arrangement per se did not affect elastic fiber–related gene expressions, and that the serum withdrawal likely induced an enhancement of the expression of tropoelastin and fibrillin-2, resulting in elastic fiber formation in the 3DCMs cultured in 1% FBS/DMEM.

3.3. Characterization of the SMC phenotype in 3DCMs

In contrast to the highly differentiated SMCs in mature adult organs, SMCs readily undergo phenotypic changes after isolation and subsequent conventional planar culture [29]. We examined

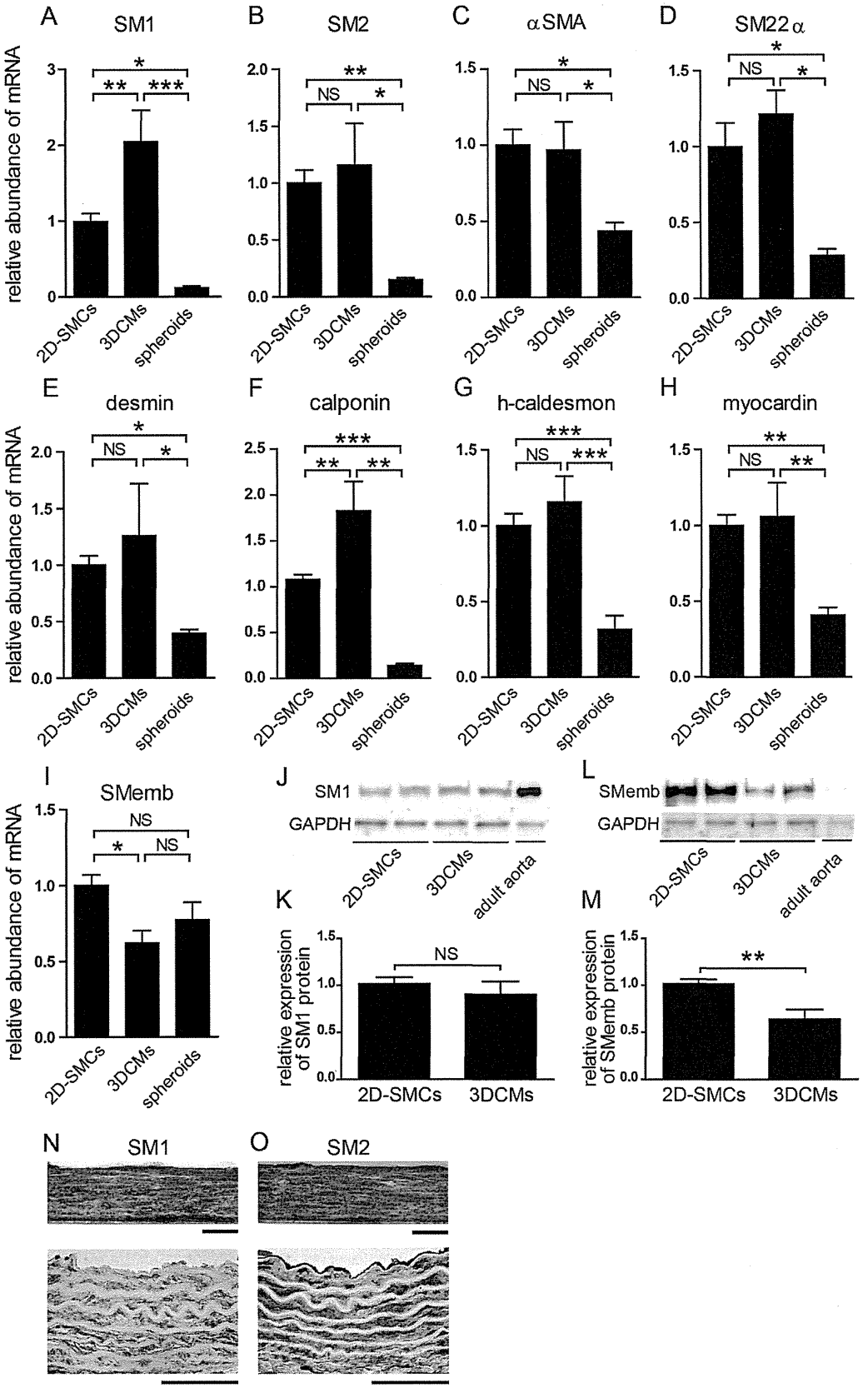


Fig. 3. Characterization of SMC phenotype in 3DCMs. (A–H) Messenger RNA expressions of differentiation SMC markers, SM1 (A), SM2 (B), αSMA (C), SM22α (D), desmin (E), calponin (F), h-caldesmon (G), myocardin (H), and a dedifferentiation SMC marker, SMemb (I) in 2D-SMCs, 3DCMs, and SMC spheroids. $n = 4–8$. (J) Immunoblot analysis for SM1 in 2D-SMCs and 3DCMs. Rat adult aorta was used as positive control. (K) Quantification of (J). $n = 10$. (L) Immunoblot analysis for SMemb in 2D-SMCs and 3DCMs. (M) Quantification of (L). $n = 10$. *, $p < 0.05$; **, $p < 0.01$; ***, $p < 0.001$. NS: not significant. Immunohistochemistry for SM1 (N) and SM2 (O). Brown color indicates positive staining. top panels: 3DCMs. Scale bars: 10 μm lower panels: rat adult aorta. Scale bars: 50 μm.

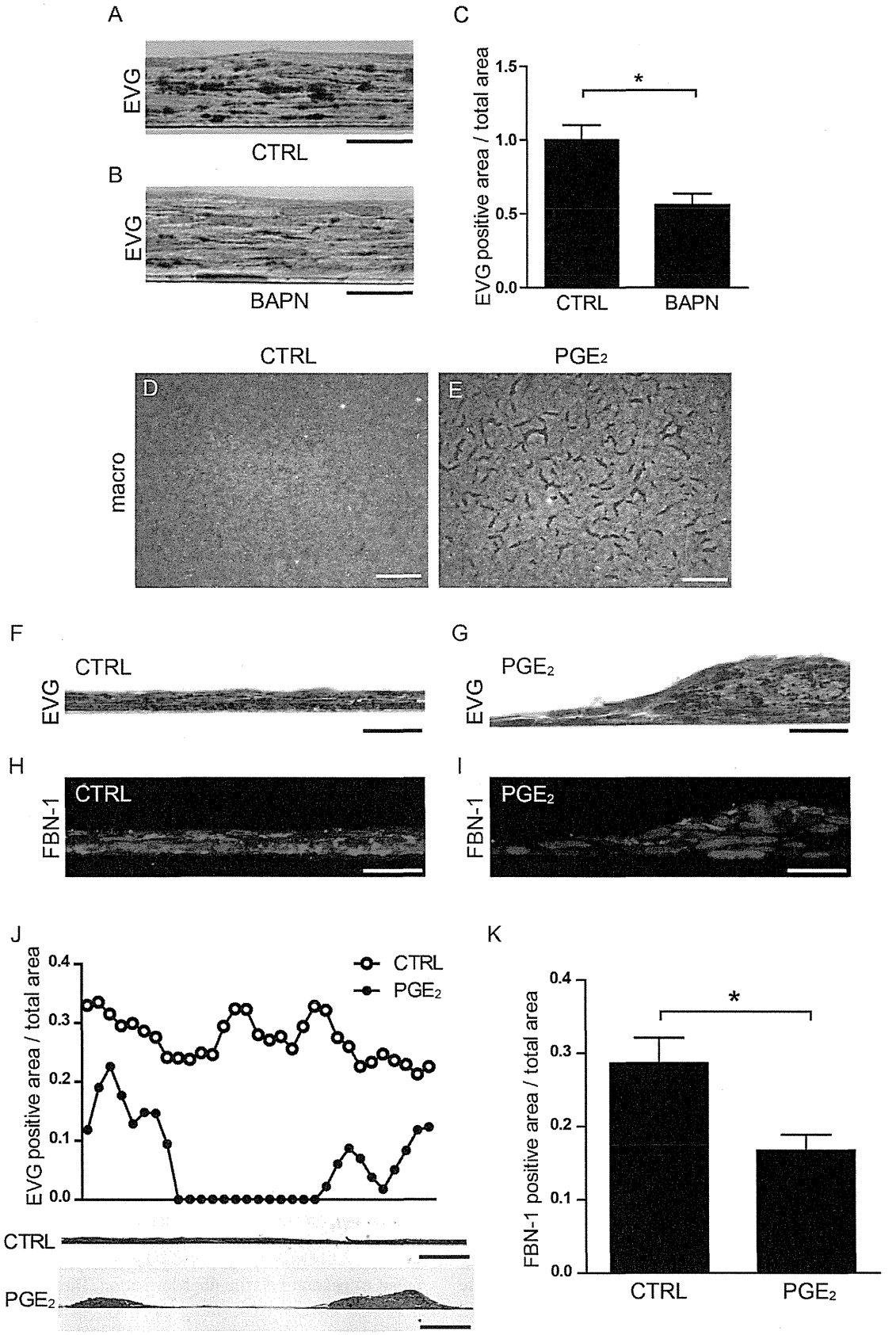


Fig. 4. Pharmacological manipulation of elastic fiber formation in 3DCMs. (A–B) EVG stain of 3DCMs. CTRL (A), and BAPN (1 μM) stimulation (B). Scale bars: 10 μm. (C) Quantification of the density of elastic fibers in (A) and (B). $n = 5$. (D–E) Macroscopic images of 3DCMs treated with or without PGE_2 (1 μM) (PGE_2 and CTRL, respectively). Scale bars:

SMC differentiation in this culture system in comparison to 2D-SMCs and SMC spheroids by monitoring the expression of various SMC markers.

It has been reported that the expression levels of differentiation SMC markers, such as SM1, SM2, αSMA, SM22α, and desmin, were highly expressed in the medial layer of SMCs but reduced in human subendotherial intima of atherosclerotic lesions [30–33] and human and rabbit aortic aneurysmal lesions [34,35]. Differentiation SMC markers such as the high molecular weight isoform of caldesmon (h-caldesmon) and calcium-binding protein calponin are also suggested to be downregulated in human atheromatous intima [31,32]. The serum response factor coactivator gene myocardin is required for the expression of many SMC differentiation marker genes and for the initial differentiation of SMC [29]. SMemb is known to be a dedifferentiation SMC marker that is abundantly expressed in human fetal arteries and immature adult cells that are present in vascular injury-induced intima [29]. Expression analysis of mRNAs demonstrated that most differentiation markers were similarly expressed between 2D-SMCs and 3DCMs (Fig. 3A–H). Although the expression levels of SM1 mRNA were higher in 3DCMs than in 2D-SMCs, the SM1 protein expression did not differ between these two culture systems (Fig. 3J–K). Among the SMC phenotypic markers examined, only SMemb was reduced in the 3DCMs compared to the 2D-SMCs in both mRNA and protein (Fig. 3I, L,M).

We also examined the SMC phenotypes of SMC spheroids in comparison with 3DCMs. mRNA expressions of all differentiation SMC markers examined in the whole spheroids were lower than in 3DCMs and even 2D-SMCs (Fig. 3A–H). The expression level of SMemb mRNA in the spheroids was similar to 2D-SMCs and 3DCMs.

Immunohistochemistry showed that SM1 and SM2 proteins were homogenously expressed in 3DCMs (Fig. 3N,O). In SMC spheroids, SM1 and SM2 were expressed mainly in the outer shell, while the dedifferentiation marker SMemb was expressed in the porous core (Supplemental Fig. 5D,F).

These data suggested that the 3DCMs exhibited similar SMC phenotypes to 2D-SMCs, although a differentiated phenotype was maintained compared to the other types of 3D culture, i.e., SMC spheroids.

3.4. Pharmacological manipulation of elastic fiber formation in 3DCMs

To examine the molecular mechanisms of the elastogenesis and degradation of elastic fibers, and to explore new therapeutic strategies against elastic fiber formation disorders, pharmacological manipulations of 3DCMs are essential. In this context, we examined whether elastic fiber formation in 3DCMs was regulated by the drug that inhibits the cross-linking of elastin monomers to form insoluble elastic fibers [1,26]. When BAPN, an inhibitor of LOX, was applied to 3DCMs for the last two days of culture, the elastic fiber formation was significantly inhibited (Fig. 4A–C).

The biosynthesis of PGE_2 is increased in human atherosclerotic plaques [36] and has been implicated in atherosclerotic plaque rupture [37]. PGE_2 is abundantly produced in aortic aneurysmal lesions and it decreases elastic fiber formation [38,39]. Therefore, we stimulated the 3DCMs with PGE_2 and found that PGE_2 stimulation induced a partial disruption of the layer structure of SMCs (Fig. 4D–E) and decreased elastic fiber formation (Fig. 4F–K). These data suggest the potential use of 3DCMs for pharmacological manipulation of elastic fiber formation.

3.5. Infiltration of macrophages decreased elastic fibers in 3DCMs

The infiltration of macrophages into the vascular wall is a critical step in the progression of atherosclerosis and aortic aneurysm, and macrophages are the main source of PGE_2. We applied THP-1 (human monocyte cell line) to the 3DCMs with or without PMA, which induces the differentiation of THP-1, into activated macrophages. Non-activated THP-1 (CTRL) did not infiltrate 3DCMs or affect the SMC layer structure or elastic fiber formation (Fig. 5A, C, E, and G). On the other hand, activated macrophages infiltrated the 3DCMs (Fig. 5D, H) and decreased the surrounding elastic fibers and disrupted the layer structure of SMCs (Fig. 5)B, F, H, and I.

4. Discussion

Despite numerous discoveries related to the molecular mechanisms of elastic fiber formation, circulatory diseases associated with elastic fiber disorder, such as aneurysms and atherosclerosis, remain the predominant cause of mortality and morbidity in the developed world [40]. No pharmacological strategy to restore the elastic fiber assembly in diseased vessels or to inhibit the progression of elastin-related diseases is currently available. To obtain further mechanistic insights into elastic fiber formation, in the current study, we sought to create a new experimental vascular model designed specifically for vascular elastic fiber research. We optimized culture conditions and found that 3DCMs consisting of fibronectin-gelatin-coated neonatal rat aortic SMCs cultured in 1% FBS/DMEM exhibited layered elastic fiber formation within seven days, and that the amount of newly synthesized insoluble elastic fibers was significantly greater in 3DCMs than in 2D-SMCs cultured in the same medium.

Recent advances in tissue-engineered blood vessels that are free of synthetic scaffolds enabled the assembly of multiple layers of SMCs and the production of elastic fiber deposition in vitro [41]. These techniques provide excellent mechanical properties as implantable vascular constructs but require an excessively long culturing time and/or an in vivo bioreactor, neither of which are practical for ex vivo experimental vascular models. In this study, fibronectin nanofilms prepared on the surface of SMCs, which has not been used in other tissue-engineered blood vessels, were adopted in 3DCMs for hierarchical cell manipulation and rapid induction of elastic fiber formation. The recent proteomics approach enabled comprehensive molecular interactions in elastic fiber assembly and revealed that fibronectin and microfibril fibrillin play a central role [42]. Kinsey et al. and Sabatier et al. demonstrated that, in addition to expression, fibronectin assembly is a prerequisite for fibrillin assembly [10,16]. It is noted that fibronectin assembly and subsequent fibrillin assembly in vitro were observed within a couple of days in the static culture condition [10,16]. Other lines of in vitro studies demonstrated that fibronectin is critical for the deposition of latent transforming growth factor β-binding protein-1 (LTBP-1), which is associated with microfibrils [11], and that fibronectin assembly promotes LOX activation [14]. Our previous report demonstrated that pericellular fibronectin fibers were observed in 3DCMs within 24 h [9]. In addition, our results demonstrated that the use of ε-poly(lysine) instead of fibronectin failed to generate elastic fiber formation. These data support our

1 mm. (F–G) EVG stain of 3DCMs. (H–I) Immunofluorescent images of fibrillin-1 in 3DCMs red: fibrillin-1. Scale bars: 50 μm. CTRL (D,F,H) and PGE_2 stimulation (E,G,I). (J) top panel: Histograms showing the density of EVG positive area/total area. lower panels: EVG stain of CTRL 3DCMs and PGE_2 stimulated 3DCMs. Scale bars: 100 μm. (K) Quantification of density of fibrillin-1 in (H) and (I). $n = 11$. $^*p < 0.05$.

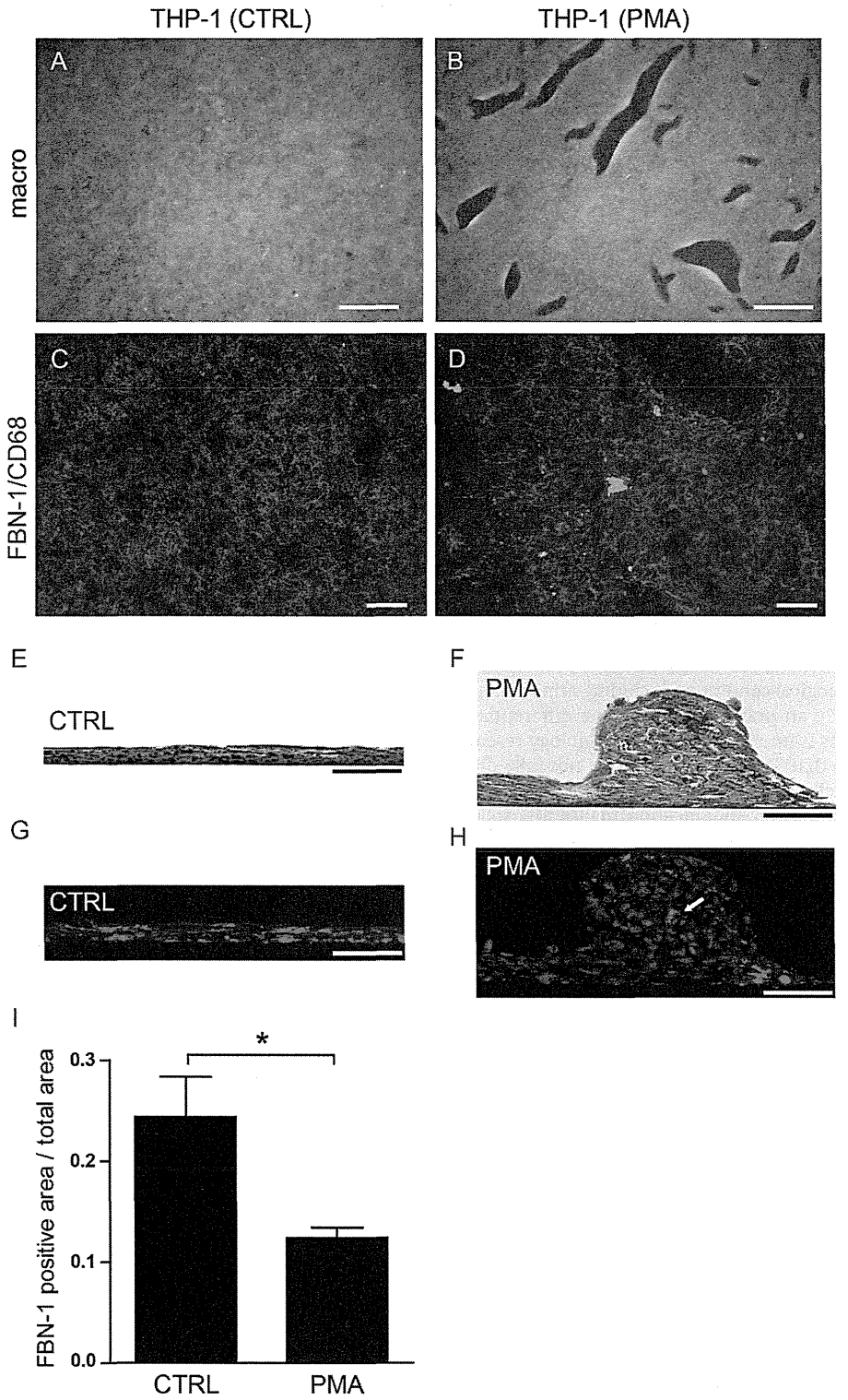


Fig. 5. Infiltration of macrophages decreased elastic fibers in 3DCMs (A–B) Macroscopic images of 3DCMs treated with non-activated THP-1 (CTRL), or activated macrophages (PMA). Scale bars: 1 mm. (C–D) Immunofluorescent images of fibrillin-1 (red) and CD68 (green) in 3DCMs. (E–F) EVG stain of 3DCMs. (G–H) Immunofluorescent image of fibrillin-1 (red) and CD 68 (green) in 3DCMs. White arrow indicates infiltrated macrophage. Scale bars: 50 μm. (I) Quantification of the density of fibrillin-1 in (G) and (H). $n = 11$. $^*p < 0.05$.

concept that the pericellular fibronectin meshwork on SMCs contributes to elastic fiber assembly in 3DCMs.

Elastin molecules play a crucial role in elastic fiber formation [1]. It has been reported that elastin is expressed maximally at the G_0 and minimally at the G_2/M phase during the cell cycle [43,44]. In accordance with previous reports, serum withdrawal increased tropoelastin mRNA expression in both 3DCMs and 2D-SMCs. Our data suggest that serum withdrawal–induced cell cycle regulation

has a favorable effect on elastic fiber formation. Fibrillin-2 which contributes to the initial assembly of elastic fibers as well as fibrillin-1 was also induced by serum-starved conditions in both 3DCMs and 2D-SMCs. Cell cycle regulation potentially contributes to the formation of elastic fibers in 3DCMs via the increase in elastin and fibrillin-2.

In addition to the fibronectin nanofilm and culture media component, the use of neonatal vascular SMCs may also contribute to elastic fiber formation in 3DCMs, because it has been well documented that adult vascular SMCs lose the ability to produce components related to elastic fiber assembly, including tropoelastin and fibrillins [1,22,24]. In our study, 3DCMs using adult human aortic SMCs did not produce elastic fibers, even if the same fibronectin coating and culture conditions were applied. In addition to these factors, our data suggest that a certain number of SMC layers are required for elastic fiber formation. Although the exact mechanisms of the rapid induction of elastic fibers in 3DCMs are not known, the multiple factors in 3DCMs, such as the type of SMC, the ECM coated on the cells, the culture media components, and the number of SMC layers are considered to allow layered elastic fibers even in short periods in a static culture condition.

Under physiological conditions, the majority of SMCs in the medial layer of the artery are considered to be in a differentiated (contractile) state that is hardly retained in planar cell culture [29]. Unexpectedly, our results suggest that SMC phenotypes in 3DCMs are similar to those in 2D-SMCs except SMemb expression. Since the differentiated status in SMCs varies through developmental stages and under pathological conditions, including atherosclerosis and vascular injury [45], an understanding of the differentiation status of SMCs in 3DCMs is important in vascular biology research. One study limitation is that SMCs in 3DCMs were not fully differentiated and the elasticity seemed significantly low compared to native vascular tissues. Further modification of 3DCMs, such as modulating SMC phenotypes before creating 3D construction by means of pharmacological stimuli or mechanical stress, is required in future studies.

In the present study, we demonstrated that elastic fiber formation in 3DCMs was disturbed by a LOX inhibitor and PGE_2, suggesting that pharmacological manipulation could be applied in 3DCMs. The relatively small reduction in elastic fiber formation by BAPN may be due to the 2-day administration out of the 7 days of culture. In conventional in vivo study using systemic drug administration, drug metabolism profoundly affects it efficacy, and its indirect effects cannot be discriminated. Although emerging evidence has demonstrated advances in drug delivery systems such as nanoparticles and antibody-drug conjugates that enable tissue-specific targeting strategy, they remain impractical for initial drug screening [46]. 3DCMs may offer a new strategy to examine the direct effects of various drugs on vascular elastic fiber formation and phenotypic changes of SMCs.

There have been excellent in vitro systems using 2D culture to examine molecular mechanisms of elastic fiber formation [5–7], because insoluble mature elastic fibers can be formed in certain conditions of 2D culture systems. However, elastic fiber formation in a vertical sectional view cannot be observed in 2D culture systems. In this study, it is suggested that the infiltration of macrophages and its effects on the spatial arrangement of elastic fibers can be observed in 3DCMs.

Spheroid culture system has been established as a 3D culture of SMCs. For example, the spheroids with vascular SMCs retain electrophysiological and contractile properties [47,48]. Paracrine effects of SMCs on gap junction formation of endothelial cells were also shown using the spheroids with human vascular cells [49]. To the best of our knowledge, however, elastic fiber formation and the effect of macrophage infiltration on SMCs have not been reported in SMC spheroids. Our study revealed that, in SMC spheroids, a few numbers of elastic laminae were formed between the outer shell and porous core at least under the same culture conditions as 3DCMs. Since a single spheroid has around 100 μm in diameter and consists of heterogeneous SMC phenotypes (Supplemental Fig. 5), it seems difficult to construct larger numbers of elastic laminae. On the other hand, the 3DCMs exhibited multiple layered elastic laminae spread over an area of at least 5 mm^2 (Fig. 4D). Furthermore, it was visible with a low magnification that macrophage infiltration induced disruption of elastic fiber assembly in the 3DCMs. In comparison with spheroid culture system, 3DCMs would be suitable for studies of elastic fiber formation in pathophysiological setting.

In conclusion, 3DCMs have the potential to be a new vascular experimental platform in which the spatial and temporal regulation of vascular elastic fibers can be observed. Since the regulation of vascular elastic fiber formation can be examined in 3DCMs, they may be useful for exploring pharmacological therapeutic strategies against disordered elastic fiber homeostasis, such as atherosclerotic vascular disease for which no pharmacological treatment is currently available.

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Disclosures

No disclosure.

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Appendix A. Supplementary data

Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.atherosclerosis.2014.01.045.

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Store-Operated Ca^{2+} Entry (SOCE) Regulates Melanoma Proliferation and Cell Migration

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Abstract

Store-operated Ca^{2+} entry (SOCE) is a major mechanism of Ca^{2+} import from extracellular to intracellular space, involving detection of Ca^{2+} store depletion in endoplasmic reticulum (ER) by stromal interaction molecule (STIM) proteins, which then translocate to plasma membrane and activate Orai Ca^{2+} channels there. We found that STIM1 and Orai1 isoforms were abundantly expressed in human melanoma tissues and multiple melanoma/melanocyte cell lines. We confirmed that these cell lines exhibited SOCE, which was inhibited by knockdown of STIM1 or Orai1, or by a pharmacological SOCE inhibitor. Inhibition of SOCE suppressed melanoma cell proliferation and migration/metastasis. Induction of SOCE was associated with activation of extracellular-signal-regulated kinase (ERK), and was inhibited by inhibitors of calmodulin kinase II (CaMKII) or Raf-1, suggesting that SOCE-mediated cellular functions are controlled via the CaMKII/Raf-1/ERK signaling pathway. Our findings indicate that SOCE contributes to melanoma progression, and therefore may be a new potential target for treatment of melanoma, irrespective of whether or not Braf mutation is present.

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Introduction

Melanoma has the poorest prognosis among skin cancers, although drugs targeting aberrant ERK signaling, i.e., mutated serine/threonine-protein kinase Braf, have improved both overall and progression-free survival times [1]. However, this therapy is not effective in patients without Braf mutation, and some patients with Braf mutation rapidly acquire resistance to Braf inhibitors [2]. Accordingly, a different approach to target ERK signaling regardless of Braf mutation is needed.

Intracellular Ca^{2+} signaling regulates diverse cellular functions including proliferation and cell migration [3]. Store-operated Ca^{2+} entry (SOCE) is a major mechanism of Ca^{2+} import from extracellular to intracellular space, especially in non-excitable cells [4]. In general, activation of inositol 1,4,5-trisphosphate (IP_3) receptors on the endoplasmic reticulum (ER) evokes a rapid and transient release of Ca^{2+} from the ER store. The resulting decrease of Ca^{2+} concentration in the ER is sensed by the EF-hand motif of stromal interaction molecules (STIM), which then translocate to the plasma membrane, where they interact with Orai Ca^{2+} channel subunits [5], leading to Ca^{2+} influx from extracellular space to restore the Ca^{2+} concentration in ER [6].

The physiological functions of STIM and Orai have been studied mainly in connection with the immune system [7–10]. Orai channels control Ca^{2+} release-activated Ca^{2+} (CRAC) currents in lymphocytes [4], and also contribute to SOCE currents in other types of cells, such as endothelial cells [11]. STIM1 and Orai1, but not STIM2, Orai2 or Orai3, have roles in cell migration of smooth muscle cells [12,13]. Examination of a library of randomized ribozymes indicated that STIM1 is a metastasis-related gene [14]. SOCE is involved in proliferation, cell migration, and angiogenesis in cervical cancer [15], and cell migration in breast cancer [16]. However, the role of SOCE in melanoma has been little investigated, except for a recent paper demonstrating Akt signaling activation in mouse melanoma cells, especially in lipid rafts [17]. In the present study, we show that SOCE promotes melanoma progression by enhancing cell proliferation, migration, and metastasis through activation of ERK signaling via the CaMKII/Raf-1/ERK pathway.