



Figure 8. Changes in the value of power spectrum of heart rate variability in response to 0.5 mg/kg T-2 toxin with blockades. (A) low-frequency (LF) power; (B) high-frequency (HF) power; (C) LF/HF ratio; (D) total power. Asterisks: significant difference (* $p < 0.05$, ** $p < 0.01$) from the T-2 toxin (0.5 mg/kg) group. Data are based on mean values obtained for each 2 or 3 days and expressed as means \pm SEM.

immediately after T-2 toxin administration. Instead, they predominantly appeared after 8 h, and the arrhythmias lasted for the three days of observation after the T-2 toxin administration. This finding indicates that T-2 toxin administration takes time to produce an arrhythmogenic condition in the rat's heart and that the cardiac influence lasted for at least three days. Similarly, in our previous study, DON, which is one of mycotoxins in the trichothecene group, also induced arrhythmia that was exhibited mostly at 10–20 h after toxin administration (Ngampongsa et al. 2011). Although it is feasible to consider that direct toxic effects (Bunner and Morris 1988) on myocytes were associated with the occurrence of arrhythmia, most of the second-degree AV blocks and sinus bradycardia in the present study seemed to be induced or augmented by the vagal activity since such a conduction disturbance was largely diminished by the atropine treatment.

If T-2 toxin induced hypotension for a few days following administration of T-2 toxin, it should provoke a reflex inhibition of the vagal activity and augmentation of the sympathetic nerve activity. From the results of an acute experiment in the dog, the reduction in arterial blood pressure and increase in heart rate were suggested to be mediated by the sympathetic nervous system (Bubien and Woods 1987). In the present study, it was assumed that the tonus of the parasympathetic nervous activity was enhanced intermittently to regulate heart rate and systemic blood pressure by means of antagonistic neural mechanisms during basically high tension of the sympathetic nervous activity for a longer period after

the administration of T-2 toxin. Such events of parasympathetic activation might cause a momentary decrease in firing rate at the sinoatrial node and also produce second-degree AV blocks, which lead to a slowing of atrioventricular conduction.

The causal relationship between arrhythmias and the decrease in LF, HF and total powers observed at 0.5 mg/kg of T-2 toxin in the present study was unclear. Although the arrhythmias such as sinus bradycardia and atrioventricular blocks are considered to be associated with momentary alteration of balance between the sympathetic and parasympathetic nervous activities, such nervous activities may be not necessarily reflected the HRV analysis.

A significant increase in occurrence of ventricular extrasystoles was identified with the combination of 0.5 mg/kg-T-2 toxin and atropine. The increase in heart rate results in an elevation in workload of the heart and might produce cardiac conditions tending to cause ventricular extrasystoles. It is known that tachycardia produces an accumulation of superoxide anion released from the mitochondrial electron transport chain in myocytes (Han et al. 2001). In addition, T-2 toxin itself has been clarified to be a substance producing oxidative stress in tissues involving the brain (Chaudhari et al. 2009; Chaudhary and Rao 2010; Doi and Uetsuka 2011). Although the detailed mechanism of ventricular extrasystoles observed in the present study was not elucidated, the oxidative stress, to some extent, may be attributed to this arrhythmia, as it has been reported that the number of cases of ventricular extrasystole in cases

of myocardial reperfusion was significantly decreased by treatment with superoxide dismutase (Kónya et al. 1992).

In conclusion, the results of the present study demonstrated and confirmed that T-2 toxin has a property of potent cardiac toxicity, the appearance of which was largely influenced by the autonomic nervous activity, lasting for at least three days in conscious rats with a concentration of 0.1 mg/kg or more administered subcutaneously.

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Declaration of interest

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References

- Abe H, Hanada H, Kohshi K, Nakashima Y. 1999. Treatment of advanced atrioventricular block with beta-adrenergic blockade therapy. *Pacing Clin Electrophysiol* 22:1097-1099.
- Ballantyne B, Marrs TC, Syversen T, eds. (2009). *General and Applied Toxicology*. Volume 3, 3rd edition. New York: MacMillan Reference (UK)/Grove's Dictionaries (US), 2149.
- Borison HL, Goodheart ML. 1989. Neural factors in acute emetic, cardiovascular, and respiratory effects of T-2 Toxin in cats. *Br J Exp Pathol* 64:570-575.
- Brady WJ, Swart G, DeBehnke DJ, Ma OJ, Aufderheide TP. 1999. The efficacy of atropine in the treatment of hemodynamically unstable bradycardia and atrioventricular block: prehospital and emergency department considerations. *Resuscitation* 41:47-55.
- Bubien JK, Woods WT Jr. 1986. Differential effects of trichothecenes on the canine cardiac action potential. *Toxicol* 24:467-472.
- Bubien JK, Woods WT Jr. 1987. Direct and reflex cardiovascular effects of trichothecene mycotoxins. *Toxicol* 25:325-331.
- Bunner DL, Morris ER. 1988. Alteration of multiple cell membrane functions in L-6 myoblasts by T-2 toxin: an important mechanism of action. *Toxicol Appl Pharmacol* 92:113-121.
- Carré F, Maison-Blanche P, Ollivier L, Mansier P, Chevalier B, Vicuna R, Lessard Y, Coumel P, Swynghedauw B. 1994. Heart rate variability in two models of cardiac hypertrophy in rats in relation to the new molecular phenotype. *Am J Physiol* 266:H1872-H1878.
- Chaudhari M, Jayaraj R, Bhaskar AS, Lakshmana Rao PV. 2009. Oxidative stress induction by T-2 toxin causes DNA damage and triggers apoptosis via caspase pathway in human cervical cancer cells. *Toxicology* 262:153-161.
- Chaudhary M, Rao PV. 2010. Brain oxidative stress after dermal and subcutaneous exposure of T-2 toxin in mice. *Food Chem Toxicol* 48:3436-3442.
- Cooley JW, Turkey JW. 1965. An algorithm for the machine calculation of complex Fourier series. *Math Comp* 19:297-301.
- Doi K, Uetsuka K. 2011. Mechanisms of Mycotoxin-Induced Neurotoxicity through Oxidative Stress-Associated Pathways. *Int J Mol Sci* 12:5213-5237.
- Feuerstein G, Goldstein DS, Ramwell PW, Zerbe RL, Lux WE Jr, Faden AI, Bayorh MA. 1985. Cardiorespiratory, sympathetic and biochemical responses to T-2 toxin in the guinea pig and rat. *J Pharmacol Exp Ther* 232:786-794.
- Han D, Williams E, Cadenas E. 2001. Mitochondrial respiratory chain-dependent generation of superoxide anion and its release into the intermembrane space. *Biochem J* 353:411-416.
- Kónya L, Kékesi V, Juhász-Nagy S, Fehér J. 1992. The effect of superoxide dismutase in the myocardium during reperfusion in the dog. *Free Radic Biol Med* 13:527-532.
- Kuwahara M, Yayou K, Ishii K, Hashimoto S, Tsubone H, Sugano S. 1994. Power spectral analysis of heart rate variability as a new method for assessing autonomic activity in the rat. *J Electrocardiol* 27:333-337.
- Love JN, Enlow B, Howell JM, Klein-Schwartz W, Litovitz TL. 2002. Electrocardiographic changes associated with beta-blocker toxicity. *Ann Emerg Med* 40:603-610.
- MacKenzie R. 2005. Short PR interval. *J Insur Med* 37:145-152.
- Magnuson BA, Schiefer HB, Hancock DS, Bhatti AR. 1987. Cardiovascular effects of mycotoxin T-2 after topical application in rats. *Can J Physiol Pharmacol* 65:799-802.
- Ngampongsa S, Ito K, Kuwahara M, Kumagai S, Tsubone H. 2011. Arrhythmias and alterations in autonomic nervous function induced by deoxynivalenol (DON) in unrestrained rats. *J Toxicol Sci* 36:453-460.
- Pang VF, Lorenzana RM, Beasley VR, Buck WB, Haschek WM. 1987. Experimental T-2 toxicosis in swine. III. Morphologic changes following intravascular administration of T-2 toxin. *Fundam Appl Toxicol* 8:298-309.
- Pomeranz B, Macaulay RJ, Caudill MA, Kutz I, Adam D, Gordon D, Kilborn KM, Barger AC, Shannon DC, Cohen RJ. 1985. Assessment of autonomic function in humans by heart rate spectral analysis. *Am J Physiol* 248:H151-H153.
- Sato N, Ueno Y, Enomoto M. 1975. Toxicological approaches to the toxic metabolites of Fusaria. VIII. Acute and subacute toxicities of T-2 toxin in cats. *Jpn J Pharmacol* 25:263-270.
- Schoental R, Joffe AZ, Yagen B. 1979. Cardiovascular lesions and various tumors found in rats given T-2 toxin, a trichothecene metabolite of Fusarium. *Cancer Res* 39:2179-2189.
- Sherman Y, More R, Yagen B, Yarom R. 1987. Cardiovascular pathology induced by passive transfer of splenic cells from syngeneic rats treated with T-2 toxin. *Toxicol Lett* 36:15-22.
- Sirén AL, Feuerstein G. 1986. Effect of T-2 toxin on regional blood flow and vascular resistance in the conscious rat. *Toxicol Appl Pharmacol* 83:438-444.
- Ueno Y. (1977). Trichothecenes: an overview. In: Rodrickes JV, Hesseltine CW, Mehlmann MA, eds. *Mycotoxins in Human and Animal Health*. IL: Pathotox Pub, 189-207.
- Ueno Y. 1985. The toxicology of mycotoxins. *Crit Rev Toxicol* 14:99-132.
- Ueno Y. (1986). Trichothecenes as environmental toxicants. In: Hodgson E. *Reviews in Environmental Toxicology* 2. Amsterdam/NY/Oxford: Elsevier, 303-341.
- Wannemacher RW, Bunner DL, Neufeld HA. (1991). Toxicity of trichothecenes and other related mycotoxins in laboratory animals. In: Smith JE, Anderson RA. *Mycotoxins and Animal Foods*. Boca Raton, FL: CRC Press, 499-552.
- Weaver GA, Kurtz HJ, Bates FY, Chi MS, Mirocha CJ, Behrens JC, Robison TS. 1978. Acute and chronic toxicity of T-2 mycotoxin in swine. *Vet Rec* 103:531-535.
- Yarom R, Yagen B. 1986. T-2 toxin effect on the ultrastructure of myocardial microvasculature. *Br J Exp Pathol* 67:55-63.

