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defects only in terminal B-cell differentiation, but not in T cells, and represent typical patients with CVID, as originally reported.

Group C patients had a high frequency of both opportunistic infections and malignancies, suggesting that these TREC(-) patients have T-cell defects. Although group C patients had a similar TREC/KREC pattern to patients with SCID with B cells (*IL2RG* and *JAK3*; see Fig E5, A), they do not fulfill the European Society for Immunodeficiencies criteria for SCID, and no mutation was identified in the SCID genes estimated from clinical manifestation and lymphocyte subset analysis. However, from our data, they would likely benefit from undergoing similar treatment to patients with SCID or CID to prevent these complications.

Although opportunistic infections were rare in group B patients, autoimmune diseases were often observed. This is consistent with this group being TREC(+)/KREC(-) and the idea that balance between T and B cells is important to prevent autoimmune diseases in patients with CVID. Intriguingly, a group of patients with AT and NBS were also TREC(+)/KREC(-) (see Fig E4, B), which is similar to group B patients. Additionally, CD45RA+CD4+ naive T-cell numbers were reduced in most group B patients, which is similar to the phenotype exhibited by patients with AT and NBS. This finding raises the possibility that although some group B patients are also T-cell deficient, as well as B-cell deficient, and should be treated similarly to patients with CID, other patients have only B-cell deficiency and are effectively treated with IVIG substitution therapy.

By analyzing a large CVID patient cohort, the overall survival rate of patients with more than 1 complication was worse than that for patients without other complications. Our findings indicate that low TREC levels, KREC levels, or both are useful markers that correlate well with the overall survival rate in patients with CVID. Therefore we conclude that TREC and KREC are useful markers to assess the clinical severity and pathogenesis of each patient with CVID and to distinguish CID from CVID. Thus patient classification based on TREC/KREC levels would provide a helpful tool for deciding on an effective treatment plan for each patient with CVID.

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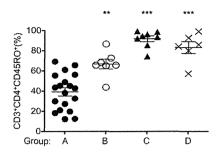


FIG E1. CD45RO $^+$ CD3 $^+$ CD4 $^+$ T-cell frequency within CD4 $^+$ CD3 $^+$ lymphocytes was analyzed among groups. CD45RO $^+$ CD3 $^+$ CD4 $^+$ lymphocyte counts were significantly higher in groups B, C, and D compared with those in group A (P < .0001). Group A: 37% \pm 16%; group B: 67% \pm 13% (**P < .01); group C: 92% \pm 8.2% (***P < .001); and group D: 83% \pm 14% (***P < .001).

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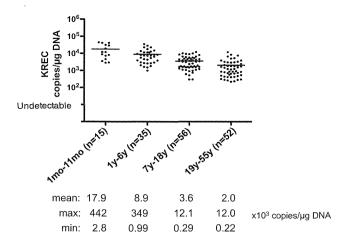


FIG E2. KREC levels were analyzed in genomic DNA samples extracted from peripheral blood of control subjects at different age groups (n = 158; age range, 1 month to 55 years). KREC levels were significantly higher in infants (17.9 \pm 3.9 \times 10³ copies/µg DNA) compared with other children's age groups (8.9 \pm 1.3 \times 10³ copies/µg DNA in the 1- to 6-year-old group and 3.6 \pm 3.8 \times 10³ copies/µg DNA in the 7- to 18-year-old group) and adults (2.0 \pm 3.3 \times 10³ copies/µg DNA; P < .0001).

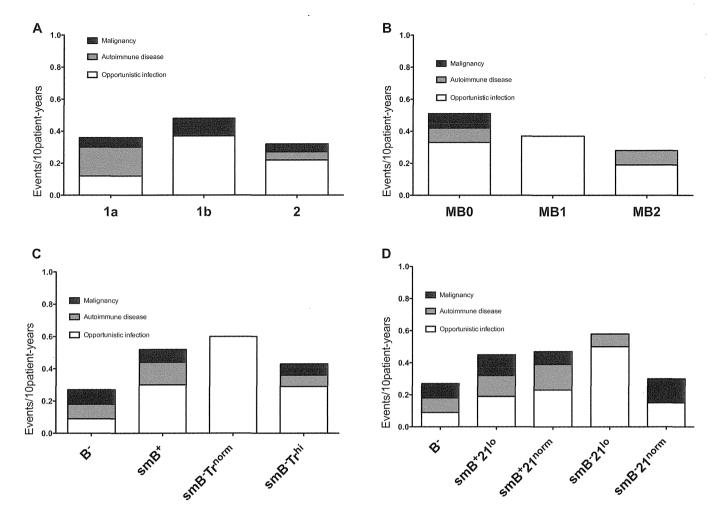


FIG E3. Patients were classified in the following way and analyzed for cumulative incidence of complications: A, Freiburg; B, Paris; and C, EUROclass classifications, according to CD38hilgMhi transitional B cells (Fig E3, A-C) or CD21^{low} B cells **(D)**. Five patients were excluded from the Freiburg and Paris classifications because of decreased B-cell numbers (<1%). Additionally, we excluded 4 patients in the Freiburg classification, 1 patient in the Paris classification, and 4 patients in the EUROclass classification for transitional B cells and 8 in the EUROclass classification for CD21 low B cells because of lack of data. The following cumulative events/10 patient-years were found. Freiburg classification: 1a, 0.36; 1b, 0.48; 2, 0.32. Paris classification: MB0, 0.50; MB1, 0.37; MB2, 0.28. EUROclass classification according to transitional B cells: B-, 0.27; smB+, 0.52; smB-Tr^{norm}, 0.60; smB-Tr^{high}, 0.43. EUROclass classification according to CD21^{lo} B cells: B-, 0.27; smB⁺21^{lo}, 0.45; smB⁺21^{norm}, 0.47; smB⁻21^{lo}, 0.58; smB⁻21^{norm}, 0.30. No classification showed any significantly increased events in any particular group according to calculated P values, as follows—Freiburg classification: 1a vs 2 = .898, 1b vs 2 = .479, 1a vs 1b = .838; Paris classification: MB0 vs MB2 = .179, MB1 vs MB2 = .654, MB0 vs MB1 = .764; EUROclass classification according to transitional B cells: B vs $smB^{+} = .298$, $smB^{-}Tr^{norm}$ vs $smB^{+} = .809$, $smB^{-}Tr^{hi}$ vs $smB^{+} = .702$, $smB^{-}Tr^{hi}$ vs $smB^{-}Tr^{norm} = .641$, $smB^{-}21^{norm} = .399, B^{-} vs smB^{+}21^{lo} = 0.474, B^{-} vs smB^{-}21^{lo} = 0.270, smB^{+}21^{lo} vs smB^{-}21^{lo} = 0.618.$

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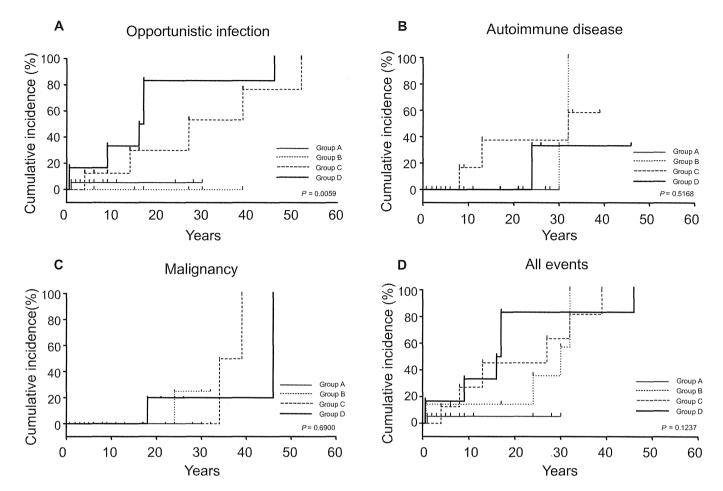


FIG E4. Comparing longitudinal cumulative incidence of complication events among groups. Cumulative incidence was estimated separately and longitudinally by using the Kaplan-Meier method and statistically compared between groups by using the log-rank test. The cumulative incidence of opportunistic infections (A), autoimmune diseases (B), malignancies (C), and all events (D) is shown.

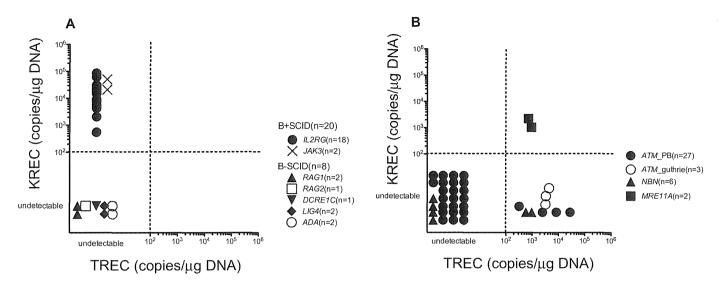


FIG E5. TREC and KREC quantification classifies patients with SCID, AT, NBS, or ataxia-telangiectasia-like disease (ATLD) into 4 groups. **A**, Patients with B⁺SCID (n=20) were classified as group C, and patients with B⁻SCID (n=8) were classified as group D; these patients were included in the previous studies. ^{5,6} **B**, Although most patients with AT (n=23) and patients with NBS (n=4) were classified as group D, TRECs were detected in peripheral blood samples (n=4 in patients with AT and n=2 in patients with NBS) and neonatal Guthrie cards (n=3) of some patients with AT, who were classified as group B. Patients with ATLD with *MRE11A* mutations were classified as group A.

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