

Figure 8. BMP7 suppressed fibroblastic change and maintained the functions of HCECs. (A) The elongated cell shapes of the fibroblastic phenotypes were reversed to a polygonal cell morphology in response to the presence of BMP-7 in a concentration-dependent manner. Scale bar: 50 μm. (B) BMP-7 enabled normal hexagonal cell morphology and actin cytoskeleton distribution at the cortex to be maintained. Scale bar: 100 μm. (C+D) BMP-7 maintained the subcellular localization of Na⁺/K⁺-ATPase and ZO-1at the plasma membrane. Scale bar: 100 μm. (E+F) The percentages of both Na⁺/K⁺-ATPase and ZO-1 positive cells treated with BMP-7 were significantly higher than in the control. * p<0.01, ** p<0.05. The experiment was performed in duplicate.

doi:10.1371/journal.pone.0058000.g008

that HCECs are vulnerable to undergoing massive fibroblastic change over each passage [40]. Therefore, it is essential to find means to circumvent the spontaneous transformation of the CECs in order to maintain the physiological phenotypes for the subsequent use for transplantation.

Transformation of endothelial cells to fibroblastic cells is designated as endothelial- mesenchymal transformation. Such transformation is triggered by TGF- β via the Smad2/3 pathway [16]. Endothelial-mesenchymal transformation causes the loss of the characteristic endothelial phenotypes, such as loss of the contact-inhibited monolayer and loss of the apical junctional

proteins at the plasma membrane. Furthermore, it causes induction of fibrillar proteins such as type I collagen and fibronectin. In this present study, we demonstrated that the fibroblastic phenotypes of cultivated CECs greatly lost the endothelial characteristics; expression of Na+/K+-ATPase and ZO-1 was markedly reduced and their subcellular localization was in the cytosol rather than the authentic plasma membrane location. Furthermore, fibroblastic phenotypes markedly enhance the production of fibrillar ECM proteins (type I collagen, fibronectin, and integrin $\alpha 5$) rather than basement membrane phenotypes (type IV and VIII collagens). The presence of such undesirable cells will greatly hamper the success of transplantation of cultivated cells in the clinical setting. Therefore, it is crucial to determine what causes the phenotypic changes and how to intervene in such endothelial-mesenchymal transformation processes of the cultivated CECs. The fact that phosphorylation of Smad2/3 was greatly enhanced in the fibroblastic phenotypes led us to conclude that the fibroblastic phenotypes in both primate and HCECs are mediated by TGF-β signaling. Therefore, we employed a specific inhibitor to the TGF-β receptor (SB431542) [45] to block the endothelial-mesenchymal transformation process observed in the fibroblastic phenotypes. SB431542 completely abolished the undesirable cellular changes, and when either primate or HCEC cultures were treated with SB431542, the prerequisite change of cells to fibroblastic phenotypes was completely abolished. Simultaneously, the characteristic subcellular location of ZO-1 and Na⁺/K⁺-ATPase is resumed at the plasma membrane and the expression of the two proteins is greatly increased at both mRNA and protein levels, suggesting that the barrier and pump functions in these cultures is intact. Moreover, we found that the production of fibrillar ECM proteins was greatly reduced. We further tested the effect of BMP-7, a well-known anti-EMT agent [31,34], to reverse the fibroblastic phenotypes of

References

- Melles GR, Lander F, van Dooren BT, Pels E, Beekhuis WH (2000) Preliminary clinical results of posterior lamellar keratoplasty through a sclerocorneal pocket incision. Ophthalmology 107: 1850–1856; discussion 1857.
- Price FW Jr, Price MO (2005) Descemet's stripping with endothelial keratoplasty in 50 eyes: a refractive neutral corneal transplant. J Refract Surg 21: 339–345.
- Gorovoy MS (2006) Descemet-stripping automated endothelial keratoplasty. Cornea 25: 886–889.
- Ishino Y, Sano Y, Nakamura T, Connon CJ, Rigby H, et al. (2004) Amniotic membrane as a carrier for cultivated human corneal endothelial cell transplantation. Invest Ophthalmol Vis Sci 45: 800–806.
- Mimura T, Yamagami S, Yokoo S, Usui T, Tanaka K, et al. (2004) Cultured human corneal endothelial cell transplantation with a collagen sheet in a rabbit model. Invest Ophthalmol Vis Sci 45: 2992–2997.
- Sumide T, Nishida K, Yamato M, Ide T, Hayashida Y, et al. (2006) Functional human corneal endothelial cell sheets harvested from temperature-responsive culture surfaces. FASEB J 20: 392–394.
- Koizumi N, Sakamoto Y, Okumura N, Okahara N, Tsuchiya H, et al. (2007) Cultivated corneal endothelial cell sheet transplantation in a primate model. Invest Ophthalmol Vis Sci 48: 4519–4526.
- 8. Koizumi N, Sakamoto Y, Okumura N, Tsuchiya H, Torii R, et al. (2008) Cultivated corneal endothelial transplantation in a primate: possible future clinical application in corneal endothelial regenerative medicine. Cornea 27 Suppl 1: S48–55.
- Koizumi N, Okumura N, Kinoshita S (2012) Development of new therapeutic modalities for corneal endothelial disease focused on the proliferation of corneal endothelial cells using animal models. Exp Eye Res 95: 60–67.
- Okumura N, Ueno M, Koizumi N, Sakamoto Y, Hirata K, et al. (2009) Enhancement on primate corneal endothelial cell survival in vitro by a ROCK inhibitor. Invest Ophthalmol Vis Sci 50: 3680–3687.
- Okumura N, Koizumi N, Ueno M, Sakamoto Y, Takahashi H, et al. (2012) ROCK inhibitor converts corneal endothelial cells into a phenotype capable of regenerating in vivo endothelial tissue. Am J Pathol 181: 268–277.
- Miyata K, Drake J, Osakabe Y, Hosokawa Y, Hwang D, et al. (2001) Effect of donor age on morphologic variation of cultured human corneal endothelial cells. Cornea 20: 59–63.
- 13. Joyce NC, Zhu CC (2004) Human corneal endothelial cell proliferation: potential for use in regenerative medicine. Cornea 23: S8–S19.

HCECs. BMP-7 also reversed the fibroblastic phenotypes to the normal endothelial cells with contact-inhibited monolayer and characteristic endothelial adhesion. Taken together, both SB431542 and BMP-7 can be powerful tools to maintain the normal endothelial phenotypes of the cultivated CECs, thus leading to a successful subsequent transplantation.

In conclusion, our findings showed that the use of the inhibitor to TGF-β receptor (SB431542) and/or anti-EMT molecules (BMP-7) enables HCECs to grow with maintaining normal physiological function (i.e., barrier and pump function). Although more extensive future studies would be beneficial, we have not observed any obvious adverse effects of continuous SB431542 or BMP-7 treatment on morphology and functions, even after several numbers of passages. This present study may prove to be the substantial protocol to provide the efficient *in vitro* expansion of HCECs. In addition, this novel strategy of inhibition of fibroblastic change during cultivation may ultimately provide clinicians with a new therapeutic modality in regenerative medicine, not only for the treatment of corneal endothelial dysfunctions, but also for a variety of pathological diseases in general.

Acknowledgments

The authors thank Dr. Morio Ueno, Dr. Michio Hagiya, Dr. Kiwamu Imagawa, Dr. Kenta Yamasaki, and Mr. Yuji Sakamoto for their valuable assistance with the experiments, and Mr. John Bush for reviewing the manuscript.

Author Contributions

Conceived and designed the experiments: NO EPK MN JH SK NK. Performed the experiments: NO MN. Analyzed the data: NO EPK MN JH SK NK. Contributed reagents/materials/analysis tools: NO SK NK. Wrote the paper: NO EPK NK.

- Zavadil J, Bottinger EP (2005) TGF-beta and epithelial-to-mesenchymal transitions. Oncogene 24: 5764–5774.
- Wendt MK, Allington TM, Schiemann WP (2009) Mechanisms of the epithelialmesenchymal transition by TGF-beta. Future Oncol 5: 1145–1168.
- 16. Saika S (2006) TGFbeta pathobiology in the eye. Lab Invest 86: 106–115.
- Kaimori A, Potter J, Kaimori JY, Wang C, Mezey E, et al. (2007) Transforming growth factor-beta1 induces an epithelial-to-mesenchymal transition state in mouse hepatocytes in vitro. J Biol Chem 282: 22089–22101.
- Chen KH, Harris DL, Joyce NC (1999) TGF-beta2 in aqueous humor suppresses S-phase entry in cultured corneal endothelial cells. Invest Ophthalmol Vis Sci 40: 2513–2519.
- Kim TY, Kim WI, Smith RE, Kay ED (2001) Role of p27(Kip1) in cAMP- and TGF-beta2-mediated antiproliferation in rabbit corneal endothelial cells. Invest Ophthalmol Vis Sci 42: 3142–3149.
- Naumann GO, Schlotzer-Schrehardt U (2000) Keratopathy in pseudoexfoliation syndrome as a cause of corneal endothelial decompensation: a clinicopathologic study. Ophthalmology 107: 1111–1124.
- Kawaguchi R, Saika S, Wakayama M, Ooshima A, Ohnishi Y, et al. (2001) Extracellular matrix components in a case of retrocorneal membrane associated with syphilitic interstitial keratitis. Cornea 20: 100–103.
- Koizumi N, Suzuki T, Uno T, Chihara H, Shiraishi A, et al. (2008) Cytomegalovirus as an etiologic factor in corneal endotheliitis. Ophthalmology 115: 292–297 e293.
- 23. Song JS, Lee JG, Kay EP (2010) Induction of FGF-2 synthesis by IL-1beta in aqueous humor through P13-kinase and p38 in rabbit corneal endothelium. Invest Ophthalmol Vis Sci 51: 822–829.
- Li W, Sabater AL, Chen YT, Hayashida Y, Chen SY, et al. (2007) A novel method of isolation, preservation, and expansion of human corneal endothelial cells. Invest Ophthalmol Vis Sci 48: 614

 –620.
- Zhu C, Joyce NG (2004) Proliferative response of corneal endothelial cells from young and older donors. Invest Ophthalmol Vis Sci 45: 1743–1751.
- Rheinwald JG, Green H (1975) Serial cultivation of strains of human epidermal keratinocytes: the formation of keratinizing colonies from single cells. Cell 6: 331-343
- Koizumi N, Fullwood NJ, Bairaktaris G, Inatomi T, Kinoshita S, et al. (2000) Cultivation of corneal epithelial cells on intact and denuded human amniotic membrane. Invest Ophthalmol Vis Sci 41: 2506–2513.

- 28. Ko MK, Kay EP (2001) Subcellular localization of procollagen I and prolyl 4hydroxylase in corneal endothelial cells. Exp Cell Res 264: 363-371.
- Parsons CJ, Takashima M, Rippe RA (2007) Molecular mechanisms of hepatic
- fibrogenesis. J Gastroenterol Hepatol 22 Suppl 1: S79–84.
 30. Ma FY, Sachchithananthan M, Flanc RS, Nikolic-Paterson DJ (2009) Mitogen activated protein kinases in renal fibrosis. Front Biosci (Schol Ed) 1: 171–187.
- 31. Zeisberg M, Hanai J, Sugimoto H, Mammoto T, Charytan D, et al. (2003) BMP-7 counteracts TGF-beta1-induced epithelial-to-mesenchymal transition and reverses chronic renal injury. Nat Med 9: 964–968.
- 32. Simic P, Vukicevic S (2007) Bone morphogenetic proteins: from developmental signals to tissue regeneration. Conference on bone morphogenetic proteins. EMBO Rep 8: 327-331.
- 33. Buijs JT, Rentsch CA, van der Horst G, van Overveld PG, Wetterwald A, et al. (2007) BMP7, a putative regulator of epithelial homeostasis in the human prostate, is a potent inhibitor of prostate cancer bone metastasis in vivo. Am J Pathol 171: 1047-1057.
- 34. Zeisberg M, Yang C, Martino M, Duncan MB, Rieder F, et al. (2007) Fibroblasts derive from hepatocytes in liver fibrosis via epithelial to mesenchymal transition. J Biol Chem 282: 23337–23347.
- Barry PA, Petroll WM, Andrews PM, Cavanagh HD, Jester JV (1995) The spatial organization of corneal endothelial cytoskeletal proteins and their relationship to the apical junctional complex. Invest Ophthalmol Vis Sci 36: 1115-1124.
- 36. Darlington JK, Adrean SD, Schwab IR (2006) Trends of penetrating keratoplasty in the United States from 1980 to 2004. Ophthalmology 113: 2171-2175.

- 37. Price MO, Price FW Jr. (2010) Endothelial keratoplasty a review. Clin Experiment Ophthalmol 38: 128-140.
- Khaireddin R, Wachtlin J, Hopfenmuller W, Hoffmann F (2003) HLA-A, HLA-B and HLA-DR matching reduces the rate of corneal allograft rejection. Graefes Arch Clin Exp Ophthalmol 241: 1020–1028.

 Coster DJ, Williams KA (2005) The impact of corneal allograft rejection on the
- long-term outcome of corneal transplantation. Am J Ophthalmol 140: 1112-
- 40. Engelmann K, Bednarz J, Valtink M (2004) Prospects for endothelial transplantation. Exp Eye Res 78: 573-578.
- 41. Mimura T, Shimomura N, Usui T, Noda Y, Kaji Y, et al. (2003) Magnetic attraction of iron-endocytosed corneal endothelial cells to Descemet's membrane. Exp Eye Res 76: 745–751.
- Mimura T, Yamagami S, Yokoo S, Yanagi Y, Usui T, et al. (2005) Sphere therapy for corneal endothelium deficiency in a rabbit model. Invest Ophthalmol Vis Sci 46: 3128-3135.
- Patel SV, Bachman LA, Hann CR, Bahler CK, Fautsch MP (2009) Human corneal endothelial cell transplantation in a human ex vivo model. Invest Ophthalmol Vis Sci 50: 2123–2131.
- Peh GS, Beuerman RW, Colman A, Tan DT, Mehta JS (2011) Human corneal endothelial cell expansion for corneal endothelium transplantation; an overview. Transplantation 91: 811–819.
- 45. Inman GJ, Nicolas FJ, Callahan JF, Harling JD, Gaster LM, et al. (2002) SB-431542 is a potent and specific inhibitor of transforming growth factor-beta superfamily type I activin receptor-like kinase (ALK) receptors ALK4, ALK5, and ALK7. Mol Pharmacol 62: 65-74.

Rho-associated kinase (ROCK) inhibitor eye drop treatment as a possible medical treatment for Fuchs corneal dystrophy

Noriko Koizumi, MD, PhD, a,b Naoki Okumura, MD, PhD, a,b Morio Ueno, MD, PhD, a
Hiroko Nakagawa, MD, a Junji Hamuro, PhD, a and Shigeru Kinoshita, MD, PhD

Author Affiliations: ^a Department of Ophthalmology, Kyoto Prefectural University of Medicine, Kyoto, Japan (Drs Koizumi, Okumura, Ueno, Nakagawa, Hamuro and Prof Kinoshita), ^b Department of Biomedical Engineering, Faculty of Life and Medical Sciences, Doshisha University, Kyotanabe, Japan (Dr Okumura and Prof Koizumi).

Correspondence and reprint request to: Shigeru Kinoshita, M.D., Ph.D.

Department of Ophthalmology, Kyoto Prefectural University of Medicine, 465

Kajii-cho, Hirokoji-agaru, Kawaramachi-dori, Kamigyo-ku, Kyoto 602-0841, Japan

E-mail: shigeruk@koto.kpu-m.ac.jp

Tel: +81-75-251-5578; Fax: +81-75-251-5663

Conflicts of Interest and Source of Funding: This study was supported by A-STEP FS from the Japan Science and Technology Agency (AS2314212G for Kinoshita and

Okumura) and the Funding Program for Next Generation World-Leading Researchers from the Cabinet Office in Japan (LS117 for Koizumi). None of the authors declare any conflict of interest.

ABSTRACT

Purpose: To report a case of Fuchs corneal dystrophy that was successfully treated by Rho-associated kinase (ROCK) inhibitor eye drops, subsequent to transcorneal freezing of damaged corneal endothelial cells.

Case report: A 52-yr-old Japanese male with a diagnosis of late-onset Fuchs corneal dystrophy was referred to our hospital as a candidate for keratoplasty. Best corrected vision was 20/20 in the right eye and 20/63 in the left. Multiple guttae were observed in both eyes. The right cornea was clear, but the left showed severe central oedema, with a central corneal thickness (CCT) of 703µm. We were unable to perform specular microscopy in the central cornea, but endothelial cells were observed in the mid-periphery at a density of 757 cells/mm². The patient was treated by a corneal endothelial denudation in the pre-pupillary region followed by the topical administration of a selective ROCK inhibitor, Y-27632, as eye drops for one week.

Results: Corneal clarity recovered and vision improved to 20/20 two weeks after treatment. At six months vision had improved to 20/16 and CCT measured 568μm, significantly lower than its pre-treatment value. Endothelial function and vision have been well maintained up to the most recent observation, 24 months post-treatment. The

average corneal endothelial density in the central and peripheral cornea was 1549.3 ± 89.7 and 705.0 ± 61.1 cells/mm², respectively.

Conclusions: The case highlights the possibility of medical treatments involving the use of ROCK inhibitor eye drops as an alternative to graft surgery for certain forms of corneal endothelial disease.

Keywords: corneal endothelium; medical treatment; Rho kinase inhibitor; Fuchs corneal dystrophy

INTRODUCTION

The proliferative ability of human corneal endothelial cells is severely limited in vivo. As a consequence corneal endothelial damage caused by trauma, intraocular surgery, or disease such as Fuchs corneal dystrophy often results in severe visual disturbance. Corneal transplantation, including corneal endothelial transplantation surgeries such as Descemet's stripping automated endothelial keratoplasty (DSAEK) and Descemet's membrane endothelial keratoplasty (DMEK), is a beneficial and realistic treatment for patients with endothelial dysfunction, however, patients will not be totally free from the risk of graft rejection. Moreover, corneal endothelial cell loss is a potential problem in the long term¹. Despite the value and potential of endothelial graft surgery, however, a purely pharmacological approach to endothelial recovery remains an attractive proposition.

Previously, we reported that a selective Rho-associated kinase (ROCK) inhibitor, Y-27632, promoted the proliferation of primate corneal endothelial cells *in vitro*², as well as the healing of the corneal endothelium *in vivo*³. Here, we present a case of Fuchs corneal dystrophy scheduled for DSAEK surgery, but successfully treated by ROCK inhibitor eye drop treatment subsequent to transcorneal freezing.

CASE REPORT

A 52-yr-old Japanese male with blurred vision due to corneal endothelial dysfunction was referred to the Kyoto Prefectural University of Medicine in May 2008. Visual acuity was 20/20 in the right eye and 20/63 in the left. Multiple guttae, typical of Fuchs corneal dystrophy, were observed in both eyes by slit lamp examination as well as by non-contact specular microscopy (EM-3000TM, TOMEY Corporation, Nagoya, Japan) (Figures 1A, B). The right cornea was clear, although the corneal endothelial density was 632 cells/mm². The left cornea showed severe central oedema accompanied by epithelial bullae (Figures 2A, B). The central corneal thickness was 703µm in the patient's affected left eye. We were unable to perform specular microscopy in the central cornea owing to the oedema, but endothelial cells were observed in the mid-periphery at a density of 757 cells/mm² (Figure 1B). The patient was diagnosed as late-onset Fuchs corneal dystrophy⁴. He was scheduled to have a DSAEK surgery, but in April 2010 volunteered for an investigative clinical study of a ROCK inhibitor eye drop treatment.

Treatment was initiated on 18 May 2010 according to a protocol approved by the Institutional Review Board of Kyoto Prefectural University of Medicine. First, diseased corneal endothelium in the pre-pupillary region was removed by transcorneal freezing³ by gently pressing a 2mm-diameter stainless steel rod which had been cooled

in liquid nitrogen onto the corneal surface for 15 sec. In our previous study using a rabbit model, we confirmed that this transcorneal freezing procedure could make a endothelial defect of approximately the same size as the rod-diameter in a reproducible fashion.³ After the rod was removed and after the cornea had thawed, 50µl of 10mM ROCK inhibitor, Y-27632 (Wako, Osaka, Japan), was applied topically as eye drops, repeated six times daily for seven days (18 to 24 May 2010). To prevent corneal infection, 0.3% Gatifloxacin hydrate eye drops were also applied four times daily. Epithelial erosion was detected after transcorneal freezing, but had healed by post-treatment day 3 (Figures 2C, D). No side effects, such as persistent epithelial defects or corneal stromal scars, were observed.

The patient's cornea recovered complete clarity two weeks after treatment and vision had improved to 20/20. Six months after treatment central corneal thickness was 568µm, significantly lower than its pre-treatment value. At this time vision had improved to 20/16 (Figures2E, F). Wide-field endothelial examinations 18 months after treatment using contact specular microscopy (Konan Medical, Inc. Nishinomiya, Japan; Figure 3A) showed that the average corneal endothelial densities in the central and peripheral cornea were 1549.3±89.7 and 705±61.1 cells/mm², respectively (mean ± SEM; Figure 3B). Although Fuchs corneal dystrophy is a progressive disease, in our

patient corneal clarity and good vision (20/16) have been maintained up to the most recent observation, 2-yrs after treatment (Figures 1G, H).

DISCUSSION

Rho-associated kinases (ROCKs) are protein serine/threonine kinases, which are the first identified and best characterized Rho downstream effectors. The Rho/ROCK pathway is involved in regulating the cytoskeleton, and has an influence on cell migration, apoptosis, and proliferation. ⁵⁻⁸

We previously reported that a selective ROCK inhibitor, Y-27632, promoted the proliferation of primate corneal endothelial cells *in vitro*.² In our previous experiments, Y-27632 promoted cell proliferation up to the time when cells became pre-confluent, but did not promote proliferation in confluent cells whose proliferation had been stopped by contact inhibition. Based on this, and on the results of experiments in rabbits,³ we hypothesized that the topical application of Y-27632 as an eye drop, combined with the prior partial denudation of diseased corneal endothelial cells, might be useful to promote the proliferation *in situ* of the corneal endothelium which is in the early diseased phase. We, thus, came up with the protocol reported here, which shows some potential for the new approach to treat of certain types of corneal endothelial

dysfunction.

In the post-treatment observation of the presented case, contact-specular microscopy revealed relatively small corneal endothelial cells, present at a high cell density, in the central part of cornea from where corneal endothelial cells had been removed by transcorneal freezing. The potential of topical application of ROCK inhibitor suggested by the current report clearly requires a larger comparative study to prove the effect of this new treatment, and plans are underway to conduct this. Regarding the mechanism of action of the procedure, we should also point out that spontaneous remodeling of the human corneal endothelial cells after Descement's stripping has been reported. 9, 10 Based on these reports, and also bearing in mind the existence of corneal endothelial precursors with higher proliferative ability in the peripheral cornea, 11, 12 we cannot rule out the possibility that re-establishment of patient's endothelium was not a direct result of ROCK inhibitor administration, but was the consequence of denudation of the pathologic endothelial cells. Notwithstanding the preliminary nature of the current observation, this case report suggests the possibility of a medical treatment for the early phase of diseases such as Fuchs corneal dystrophy, via the stimulation of non-affected peripheral cells with ROCK inhibitor following the destruction of diseased cells in the central endothelium by transcorneal freezing.

To the best of our knowledge, it is the first report suggesting that the *in vivo* proliferation of a patient's corneal endothelium can be stimulated by interventional medical/pharmaceutical treatment following the destruction of diseased endothelium.

We believe our new findings will contribute to the opening up of a new approach to the treatment of corneal endothelial dysfunction.

ACKNOWLEDGEMENTS

The authors thank Dr. Yoshiki Sasai (RIKEN CDB, Japan) for his invaluable advice regarding ROCK inhibitors, and Prof. Andrew Quantock (Cardiff University, UK) for useful discussions. This study was supported by A-STEP FS from the Japan Science and Technology Agency (AS2314212G for Kinoshita and Okumura) and the Funding Program for Next Generation World-Leading Researchers from the Cabinet Office in Japan (LS117 for Koizumi).

REFERENCES

- 1. Terry MA, Chen ES, Shamie N, et al. Endothelial cell loss after Descemet's stripping endothelial keratoplasty in a large prospective series. *Ophthalmology* 2008:115:488-496.
- 2. Okumura N, Ueno M, Koizumi N, et al. Enhancement on primate corneal endothelial cell survival in vitro by a ROCK inhibitor. *Invest Ophthalmol Vis Sci.* 2009;50:3680-3687.
- 3. Okumura N, Koizumi N, Ueno M, et al. Enhancement of corneal endothelium wound healing by Rho-associated kinase (ROCK) inhibitor eye drops. *Br J Ophthalmol.* 2011;95:1006-1009.
- 4. Eghrari AO, Gottsch JD. Fuchs' corneal dystrophy. *Expert Review of Ophthalmology.* 2010;5:147-159.
- 5. Coleman ML, Marshall CJ, Olson MF. RAS and RHO GTPases in G1-phase cell-cycle regulation. *Nat Rev Mol Cell Biol.* 2004;5:355-366.
- 6. Hall A. Rho GTPases and the actin cytoskeleton. *Science* 1998;279:509-514.
- 7. Olson MF, Ashworth A, Hall A. An essential role for Rho, Rac, and Cdc42
 GTPases in cell cycle progression through G1. *Science* 1995;269:1270-1272.
- 8. Watanabe K, Ueno M, Kamiya D, et al. A ROCK inhibitor permits survival of

- dissociated human embryonic stem cells. Nat Biotechnol. 2007;25:681-686.
- 9. Balachandran C, Ham L, Verschoor CA, et al. Spontaneous corneal clearance despite graft detachment in descemet membrane endothelial keratoplasty. *Am J Ophthalmol.* 2009;148:227-234.
- 10. Shah RD, Randleman JB, Grossniklaus HE. Spontaneous corneal clearing after Descemet's stripping without endothelial replacement. *Ophthalmology* 2012;119:256-60.
- Mimura T and Joyce N. Replication competence and senescence in central and peripheral human corneal endothelium. *Invest Ophthalmol Vis Sci.* 2006;47:1387-1396.
- 12. Mimura T, Yamagami S, Yokoo S, et al. Comparison of rabbit corneal endothelial cell precursors in the central and peripheral cornea. *Invest Ophthalmol Vis Sci.* 2005;46:3645-8.

FIGURE LEGENDS

Figure 1.

The corneal endothelium observed by non-contact-specular microscopy before treatment. (A) Multiple guttae were present (*), and corneal endothelial cells at a density of 632 cells/mm² were observed in the center of the right cornea. (B) We were unable to perform specular microscopy in the center of the left cornea owing to the oedema, however, endothelial cells were observed in the mid-periphery at a density of 757 cells/mm². Guttae were also observed (*).

Figure 2.

Slit-lamp photographs of our Fuchs' corneal dystrophy patient before and after transcorneal freezing and ROCK inhibitor treatment. Before treatment, central corneal oedema (A) accompanied by a lesion of epithelial bullae (B) was detected. Three days after treatment the corneal erosion created by the transcorneal freezing had already healed and mild bullae were detected (C, D). It should be noted that less corneal oedema was observed at two days compared to the pre-treatment photograph. Six months after treatment corneal oedema was significantly reduced and cornea had recovered its clarity (E). No epithelial damage was observed by fluorescein staining (F). 2-yrs after

treatment, the patient's cornea remains clear with good (20/16) vision (G, H).

Figure 3.

(A) Wide-field observation of the corneal endothelium by contact-specular microscopy 18 months after treatment. Guttae were detected mainly in the paracentral area. (B) Representative, magnified photographs from nasal peripheral, central and temporal peripheral area. Smaller cells, present at high density, were observed in the central cornea. (Scale bar = $100\mu m$)





