

10. スタチンのアミロイドβ蛋白低下作用とそのメカニズム：培養神経細胞を用いた検討 12:58
 玉岡晃¹⁾、○保坂愛^{1,2)}、織田彰子¹⁾、富所康志¹⁾、荒木亘²⁾
 筑波大学医学医療系神経内科学¹⁾、国立精神・神経医療研究センター神経研究所²⁾
11. 造血幹細胞を利用した脳アミロイドーシス治療戦略の開発 13:11
 芦原英司、○高田和幸¹⁾、北村佳久²⁾
 京都薬科大学病態生理学分野
12. フェノール化合物は特異的結合によって Aβ オリゴマー形成及びシナプス毒性を抑制する 13:24
 山田正仁¹⁾、○小野賢二郎¹⁾、Lei Li²⁾、高村雄策³⁾、吉池裕二⁴⁾、池田篤平¹⁾、
 西条寿夫³⁾、高島明彦⁴⁾、David B. Teplow⁵⁾、Michael G. Zagorski²⁾
 金沢大学大学院脳老化・神経病態学（神経内科学）¹⁾、
 Department of Chemistry、Case Western Reserve University²⁾、
 富山大学大学院医学薬学研究部システム情動科学³⁾、国立長寿医療研究センター研究所⁴⁾、
 Department of Neurology、David Geffen School of Medicine at UCLA⁵⁾

国際シンポジウム (14:00~16:40)

International Symposium on Intra and Extracellular

Amyloid Formation Mechanism in Amyloidosis

IV. 透析アミロイドーシス (16:50~17:42)

座長 内木宏延

13. 透析患者の手根管症候群手術既往歴の疫学調査 16:50
西慎一¹⁾、山縣邦弘²⁾、中井滋³⁾、星野純一⁴⁾、椿原美治⁵⁾
神戸大学大学院腎臓内科 腎血液浄化センター¹⁾、筑波大学医学医療系腎臓内科学²⁾、
藤田保健衛生大学臨床工学科³⁾、虎ノ門病院分院内科⁴⁾、
大阪大学大学院医学系研究科腎疾患統合医療学寄付講座⁵⁾
14. アミロイドの沈着部位・程度が透析患者の QOL に与える影響について 17:03
○星野純一¹⁾、澤直樹²⁾、三瀬広記³⁾、住田圭一⁴⁾、平松里佳子⁵⁾、長谷川詠子⁶⁾、山内真之⁷⁾、
早見典子⁸⁾、諏訪部達也⁹⁾、乳原善文¹⁰⁾、高市憲明¹¹⁾
虎の門病院腎センター
15. C-末端 unfolded β_2 -microglobulin のモデル蛋白としての Δ N₆ β_2 -m の検証 17:16
安東由喜雄¹⁾、○本宮善恢²⁾、城野博史³⁾、宇治義則⁴⁾
熊本大学大学院生命科学研究部神経内科学分野¹⁾、医療法人翠悠会²⁾
富山大学付属病院検査部³⁾
16. β_2 -ミクログロブリンアミロイド線維の細胞毒性に関する検討 17:29
内木宏延、○大越忠和¹⁾、長谷川一浩²⁾、小澤大作³⁾
福井大学医学部病因病態医学講座分子病理学領域

18:00~意見交換会
(11階 朱鷺の間)

1月25日(金)

V. FAP (9:00~9:39)

座長 安東由喜雄

17. 肝移植後のFAP患者における組織沈着アミロイドの病理組織学的特徴と
生化学的検討 9:00
安東由喜雄¹⁾、○大嶋俊範¹⁾、植田光晴¹⁾、山下太郎¹⁾、三隅洋平¹⁾、田崎雅義¹⁾、
神力悟²⁾、大林光念²⁾、大矢雄希³⁾、阿曾沼克弘³⁾、猪股裕紀洋³⁾
熊本大学大学院生命科学研究部神経内科学分野¹⁾、
熊本大学医学部附属病院アミロイドーシス診療体制構築事業²⁾、
熊本大学大学院生命科学研究部小児移植外科学分野³⁾

18. FAP患者肝を用いたドミノ移植患者剖検例におけるアミロイド沈着様式の検討 9:13
○山下太郎¹⁾、小池春樹²⁾、大嶋俊範¹⁾、田崎雅義¹⁾、三隅洋平¹⁾、神力悟³⁾、植田光晴¹⁾、
城野博史⁴⁾、大林光念²⁾、祖父江元²⁾、安東由喜雄¹⁾
熊本大学大学院生命科学研究部神経内科学分野¹⁾、名古屋大学大学院神経内科²⁾、熊本大学医
学部附属病院アミロイドーシス診療体制構築事業³⁾、熊本大学大学院生命科学研究部臨床薬物
動態学分野⁴⁾

19. 霊長類トランスサイレチンの立体構造解析 9:26
○水口峰之¹⁾、植田光晴²⁾、安東由喜雄³⁾
富山大学大学院医学薬学研究部¹⁾、熊本大学医学部附属病院中央検査部²⁾、
熊本大学大学院生命科学研究部神経内科学分野³⁾

VI. FAP (9:39~10:05)

座長 池田修一

20. FAPにおける肝移植後の末梢神経障害の進行に対する加齢の影響 9:39
○小池春樹¹⁾、大山健¹⁾、橋本里奈¹⁾、川頭祐一¹⁾、飯島正博¹⁾、亀井秀弥²⁾、
木内哲也²⁾、祖父江元¹⁾
名古屋大学神経内科¹⁾、名古屋大学移植外科²⁾
21. 高齢発症ATTR Val30Met FAP患者の末梢神経機能に対するジフルニサルの
有効性の検討 9:52
池田修一¹⁾、○関島良樹¹⁾、東城加奈¹⁾、森田洋¹⁾、小池春樹²⁾、祖父江元²⁾
信州大学脳神経内科 リウマチ・膠原病内科¹⁾、名古屋大学医学部附属病院神経内科²⁾

VII. その他のアミロイドーシス (10:05~10:31)

座長 樋口京一

22. アミロイドーシス特有の病理像を認識するプローブの探索(7) 10:05

○^{くどうゆきつか}工藤幸司¹⁾、小熊幸恵¹⁾、岡村信行^{1,2)}、古川勝敏³⁾、荒井啓行³⁾、星井嘉信⁴⁾、宇田裕史⁵⁾、佐伯修⁵⁾、奥田恭章⁶⁾、中村正⁷⁾、坂井勇仁⁸⁾、和田庸子⁸⁾、中野正明⁸⁾、佐藤弘恵⁹⁾、小関由美¹⁰⁾、田村裕昭¹¹⁾、神谷百合香¹²⁾、寺井千尋¹²⁾、亀田智宏¹³⁾、谷口義典¹⁴⁾、公文義雄¹⁴⁾、松下正人¹⁵⁾、江原重幸¹⁶⁾、檜崎雅司¹⁷⁾、岩崎由恵¹⁸⁾、川上純¹⁹⁾、吉崎和幸²⁰⁾

東北大学病院臨床研究推進センター¹⁾、東北大学医学系研究科機能薬理学分野²⁾、東北大学加齢医学研究所³⁾、山口大学医学系研究科情報解析医学系学域病理形態分野(病理学第一講座)⁴⁾、堺温心会病院内科⁵⁾、道後温泉病院リウマチセンター内科⁶⁾、熊本リウマチセンターリウマチ膠原病内科⁷⁾、新潟大学大学院医歯学総合研究科内部環境医学講座(第2内科)⁸⁾、新潟県立リウマチセンター⁹⁾、東京女子医大付属膠原病リウマチ痛風センター¹⁰⁾、勤医協中央病院内科(リウマチ・膠原病)¹¹⁾、自治医科大学付属さいたま医療センターアレルギーリウマチ科¹²⁾、香川大学医学部内分泌代謝・血液・免疫・呼吸器内科¹³⁾、高知大学医学部内分泌代謝・腎臓内科¹⁴⁾、大阪南医療センター¹⁵⁾、神戸徳洲会病院¹⁶⁾、大阪大学大学院医学系研究科免疫アレルギー内科¹⁷⁾、八尾徳洲会総合病院¹⁸⁾、長崎大学病院第一内科¹⁹⁾、大阪大学先端科学イノベーションセンター²⁰⁾

23. マウス AApoAII アミロイド線維の形成・伸長は apoA-II タンパク質の 62 番アミノ酸組成に影響される 10:18

○^{さわしたじんこ}澤下仁子¹⁾、田耕¹⁾、羅宏敏¹⁾、李琳¹⁾、森政之¹⁾、亀谷富由樹²⁾、樋口京一¹⁾
信州大学医学系研究科疾患予防医科学系加齢生物学講座¹⁾、
(財)東京都医学総合研究所 認知症・高次脳機能研究分野²⁾

Coffee Break (10:31~10:46)

VIII. AL アミロイドーシス Part 1 (10:46~11:38)

座長 今井裕一

24. LC-MS/MS でのみ確定診断できた AL アミロイドーシスの 4 症例 10:46
○畑裕之^{1,5)}、大林光念²⁾、田崎雅義³⁾、内場光⁴⁾、安東由喜雄³⁾、満屋裕明¹⁾
熊本大学医学部附属病院血液内科¹⁾、熊本大学医学部附属病院中央検査部²⁾、熊本大学大学院生命科学研究部神経内科学分³⁾、熊本大学医学部附属病院輸血細胞治療部⁴⁾、熊本大学大学院生命科学研究部生体情報解析学⁵⁾
25. AL と TTR アミロイド心の LC-MS/MS 解析 10:59
○畑裕之^{1,5)}、大林光念²⁾、田崎雅義³⁾、高潮征爾⁶⁾、内場光浩⁴⁾、安東由喜雄³⁾、満屋裕明¹⁾
熊本大学医学部附属病院血液内科¹⁾、熊本大学医学部附属病院中央検査部²⁾、熊本大学大学院生命科学研究部神経内科学分³⁾、熊本大学医学部附属病院輸血細胞治療部⁴⁾、熊本大学大学院生命科学研究部生体情報解析学⁵⁾、熊本大学医学部附属病院循環器内科⁶⁾
26. シェーグレン症候群に合併した多源性結節性 AL アミロイド症 3 例の検討 11:12
○池田修一¹⁾、日根野晃代¹⁾
信州大学医学部脳神経内科 リウマチ・膠原病内科
27. 多彩なニューロパチを呈した IgMλ 型 AL アミロイドーシスの 1 例 11:25
○野畑宏信¹⁾、山田祐一郎¹⁾、鈴木啓介¹⁾、菅憲広¹⁾、北川渡¹⁾、三浦直人¹⁾、今井裕一¹⁾
愛知医科大学腎臓・リウマチ膠原病内科

IX. AL アミロイドーシス Part 2 (11:38~12:30)

座長 島崎千尋

28. 原発性全身性 AL アミロイドーシスにおける大量メルファラン療法後非寛解例 および再発例に対する治療の検討 11:38
○加藤修明¹⁾、松田正之²⁾、池田修一¹⁾
信州大学医学部脳神経内科、リウマチ・膠原病内科¹⁾、
信州大学医学部附属病院難病診療センター²⁾
29. AL amyloidosis 患者に対する Bortezomib を使用した維持療法 11:51
麻奥英毅¹⁾、○片山雄太²⁾、板垣充弘²⁾、大地哲朗²⁾、岡谷健史²⁾、今中亮太²⁾、許鴻平²⁾、岩戸康治³⁾、辰元為仁⁴⁾、岡田武規⁵⁾、許泰一²⁾
広島赤十字・原爆病院検査部¹⁾、広島赤十字・原爆病院血液内科部²⁾、広島赤十字・原爆病院輸血部³⁾、広島赤十字・原爆病院腎臓内科部⁴⁾、広島赤十字・原爆病院循環器内科部⁵⁾

30. 原発性 AL アミロイドーシスに対するボルテゾミブ・メルファラン・デキサメタゾン療法
の安全性と有用性に関する研究：臨床第 I/II 相試験の進捗状況 (II) 12:04
○島崎千尋¹⁾、淵田真一¹⁾、石田禎夫²⁾、澤村守夫³⁾、鈴木憲史⁴⁾、小谷岳春⁵⁾、今井裕一⁶⁾、
麻奥英毅⁷⁾、安倍正博⁸⁾、宮本敏浩⁹⁾、畑 裕之¹⁰⁾、飯田真介¹¹⁾、村上博和¹²⁾、安東由喜雄¹³⁾
社会保険京都病院血液内科¹⁾、札幌医科大学第一内科²⁾、国立病院機構西群馬病院血液内科³⁾、
日本赤十字医療センター血液内科⁴⁾、金沢大学血液内科⁵⁾、愛知医科大学腎臓・膠原病内科⁶⁾、
広島赤十字原爆病院血液内科⁷⁾、徳島大学血液内科⁸⁾、九州大学血液腫瘍内科⁹⁾、熊本大学血
液内科¹⁰⁾、名古屋市立大学血液・化学療法内科¹¹⁾、群馬大学保健学科¹²⁾、熊本大学大学院生
命科学研究部神経内科学分野¹³⁾
31. 当院で行った全身性 AL アミロイドーシスに対する自家末梢血幹細胞移植 12:17
35 例の検討
○塚田信弘^{つかたのぶひろ}、宮崎寛至、阿部有、関根理恵子、中川靖章、鈴木憲史
日本赤十字社医療センター 血液内科

12:30~13:30 昼食

X. AA アミロイドーシス Part 1 (13:30~14:09) 座長 山田俊幸

32. チーターは複数回の遺伝子重複により形成された 4 個の serum amyloid A 遺伝子 13:30
をもつ
陳磊¹⁾、宇根有美²⁾、○樋口京一^{ひぐちけいいち}¹⁾、劉穎業¹⁾、丁欣¹⁾、澤下仁子¹⁾、森政之¹⁾
信州大学医学系研究科疾患予防医科学系加齢生物学講座¹⁾、麻布大学獣医学部病理学研究室²⁾
33. アミロイド特異成分 AA76 の検出 13:43
○山田俊幸^{やまだとしゆき}¹⁾、佐藤純司¹⁾、奥田恭章²⁾
自治医科大学臨床検査医学¹⁾、道後温泉病院リウマチセンター²⁾
34. SAA のアミロイド線維形成におけるアイソフォームの影響 13:56
山田俊幸¹⁾、○田中将史^{たなかまさみ}²⁾、高瀬ひろか²⁾、向高弘²⁾
自治医科大学臨床検査医学¹⁾、神戸薬科大学薬品物理化学研究室²⁾

XI. AA アミロイドーシス Part 2 (14:09~14:48) 座長 奥田恭章

35. Caplan 症候群に合併した AA アミロイドーシス 14:09
中村正¹⁾、○太良史郎²⁾、高岡宏和¹⁾、貞松智貴²⁾、大林光念³⁾、安東 由喜雄³⁾
くまもと森都総合病院リウマチ膠原病内科¹⁾、くまもと森都総合病院血液内科²⁾、
熊本大学アミロイドーシス診療体制構築事業³⁾
36. 尿検査による腎アミロイドーシスの診断法および腎臓内のアミロイドの動態について 14:22
西慎一¹⁾、○黒田毅²⁾、伊藤由美³⁾、今井直史³⁾、中枝武司³⁾、和田庸子³⁾、
中野正明⁴⁾、山田俊幸⁵⁾
神戸大学大学院腎臓内科・腎血液浄化センター¹⁾、新潟大学保健管理センター²⁾
新潟大学大学院医歯学総合研究科腎・膠原病内科学分野³⁾、新潟大学医学部保健学科⁴⁾
自治医科大学臨床検査医学講座⁵⁾
37. リウマチ性疾患に合併する反応性 AA アミロイドーシスに対する抗 IL-6 14:35
レセプター抗体療法と抗 TNF 療法の臨床的有用性の比較・検討 - 第二報
○奥田恭章^{おくだやすあき}
道後温泉病院リウマチセンター

Coffee Break (14:48~15:03)

XII. AA アミロイドーシス Part 3 (15:03~15:55) 座長 吉崎和幸

38. AA アミロイドーシスにおけるエタネルセプトのアミロイド沈着に与える影響 15:03
—腎と消化管との相違について—
○佐伯修^{さいいきおさむ}、宇田裕史、三上有子、水本綾、原田環、高間俊郎
東大阪市立総合病院内科
39. 低分子化合物による血清アミロイド A (SAA) 発現の制御 (RA 滑膜細胞での検討) 15:16
○石田清志^{いしだきよし}¹⁾、中村正²⁾
独立行政法人国立病院機構長崎医療センター臨床研究センター¹⁾、
くまもと森都総合病院リウマチ膠原病内科²⁾

40. T細胞活性化調整剤による関節リウマチ合併AAアミロイドーシス治療： 15:29
抗fPRL1抗体を用いた検討
○中村正¹⁾、公文義雄²⁾、平田真哉³⁾、高岡宏和¹⁾、下村泰三⁴⁾、
鈴木仁⁴⁾、飯干明⁵⁾、伊勢紘平⁵⁾
くまもと森都総合病院リウマチ膠原病内科¹⁾、高知大学医学部病態情報診断学²⁾、熊本大学附
属病院血液・膠原病・感染免疫診療部³⁾、くまもと森都総合病院血液内科⁴⁾、くまもと森都総
合病院整形外科⁵⁾
41. AAアミロイドーシスのトシリズマブによる治療研究 15:42
7. AAアミロイドーシス臨床研究会による治療経過終了
○吉崎和幸¹⁾、山田正仁²⁾、池田修一³⁾、安東由喜雄⁴⁾、今井裕一⁵⁾、奥田恭章⁶⁾、河野裕夫⁷⁾、
工藤幸司⁸⁾、黒田毅⁹⁾、高市憲明¹⁰⁾、中里雅光¹¹⁾、山田俊幸¹²⁾、江口勝美¹³⁾、寺井千尋¹⁴⁾、
中村正¹⁵⁾、葦田清次¹⁶⁾、田中敏郎¹⁷⁾、稲田進一¹⁸⁾、公文義雄¹⁹⁾、小関由美²⁰⁾、佐伯修²¹⁾、田
村裕昭²²⁾、土橋浩章²³⁾、中野正明²⁴⁾、松原司²⁵⁾、山名征三²⁶⁾、佐伯行彦²⁷⁾
大阪大学大学院工学研究科応用化学専攻免疫医科学¹⁾、金沢大学²⁾、信州大学³⁾、熊本大学⁴⁾、
愛知医科大学⁵⁾、道後温泉病院⁶⁾、山口大学⁷⁾、東北大学⁸⁾、新潟大学⁹⁾、虎ノ門病院¹⁰⁾、宮
崎大学¹¹⁾、自治医科大学¹²⁾、長崎大学¹³⁾、自治医科大学¹⁴⁾、熊本整形外科病院¹⁵⁾、自治医科
大学¹⁶⁾、大阪大学¹⁷⁾、都立多摩総合医療センター¹⁸⁾、高知大学¹⁹⁾、東京女子医科大学²⁰⁾、堺
温心会病院²¹⁾、勤医協中央病院²²⁾、香川大学²³⁾、新潟大学²⁴⁾、松原メイフラワー病院²⁵⁾、東
広島記念病院²⁶⁾、国立病院機構大阪南医療センター²⁷⁾

16:00 終了挨拶 研究代表者 安東由喜雄

International Symposium on Intra and Extracellular Amyloid Formation Mechanism in Amyloidosis

PROGRAM & ABSTRACTS

Date: January 24 (Thu) 2013

14 : 00 ~ 16 : 40

Place: KKR Hotel Tokyo, 1-4-1 Otemachi Chiyoda-ku Tokyo, Japan

Speakers:

Satoshi Yamashita (Kumamoto University, Japan)

Seung-Jae Lee (Konkuk University, Korea)

Yoshitaka Nagai (National Center of Neurology and Psychiatry, Japan)

Maria João Saraiva (University of Port, Portugal)

**Organized by the Amyloidosis Research Committee, Research on
Intractable Diseases, Health and Labour Sciences Research Grants,
The Ministry of Health, Labour and Welfare, Japan**

Chairperson: Yukio Ando, MD, PhD

**Kumamoto University Graduate School of Medical Science,
Department of Neurology**

International Symposium on Intra and Extracellular Amyloid Formation Mechanism in Amyloidosis

General information

Date: January 24, 2013

14:00~16:40 (Registration Fee: Free)

Place: KKR Hotel Tokyo, 11th Floor Banquet Room KUJAKU

1-4-1 Otemachi Chiyoda-ku Tokyo, Japan Phone: 03-3287-2921 FAX: 03-3287-2913

Reception Party: January 24, 2013

18:00~ (Registration Fee: 6,000Yen)

KKR Hotel Tokyo, 11th Floor Banquet Room TOKI

Neighboring the Imperial Palace, KKR Hotel Tokyo is conveniently located 5 minutes by car from Tokyo Station, 40 minutes from Haneda Airport and 80 minutes from Narita Airport by car. The Hotel is also directly connected to a subway station. Nearby attractions include Ginza, Akihabara, Tsukiji, Asakusa, Ueno, Roppongi and Odaiba.

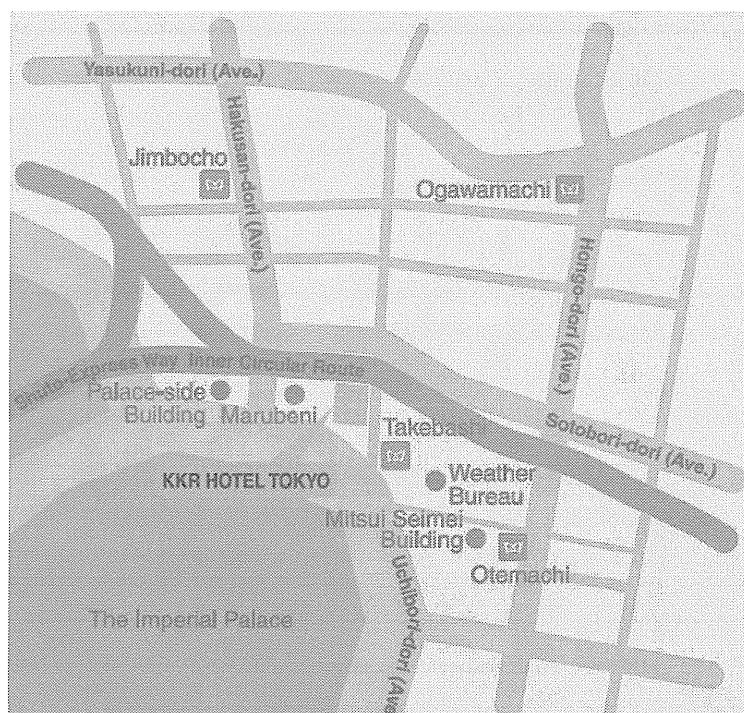
Accessing from nearby train stations

20 minutes on foot, 5 minutes by car from the Marunouchi north exit of Tokyo Station

Directly connected from Exit 3b of Takebashi Station on Tozai Line

5 minutes on foot from Exit C2 of Otemachi Station on Chiyoda Line

5 minutes on foot from Exit A9 of Jinbocho Station on Metro Subway



Contact: Secretariat

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Program

January 24, 2013,

KKR Hotel Tokyo, 11th Floor Banquet Room KUJAKU

Opening Remarks

14:00-14:10 Yukio Ando (Kumamoto University, Japan)

Lecture 1

Chairpersons: Shu-ichi Ikeda (Shinshu University, Japan)

14:10-14:40 Satoshi Yamashita (Kumamoto University, Japan)

Lecture 2

Chairpersons: Keiichi Higuchi (Shinshu University, Japan)

14:40-15:10 Seung-Jae Lee (Konkuk University, Korea)

Mechanism of disease progression through cell-to-cell amyloid propagation in neurodegenerative diseases

Coffee Break

15:10-15:30

Lecture 3

Chairpersons: Hironobu Naiki (University of Fukui, Japan)

15:30-16:00 Yoshitaka Nagai (National Center of Neurology and Psychiatry)

Toxic protein conformational transition and amyloid fibril formation in the polyglutamine diseases.

Lecture 4

Chairpersons: Yukio Ando (Kumamoto University, Japan)

16:00-16:30 Maria João Saraiva (IBMC, University of Port, Portugal)

Clearance of extracellular misfolded proteins in systemic amyloidosis: experience with transthyretin

Closing Remarks

16:30-16:40 Yukio Ando (Kumamoto University, Japan)

Role of TDP-43 aggregation in neurodegenerative disease

Satoshi Yamashita, M.D., Ph.D.

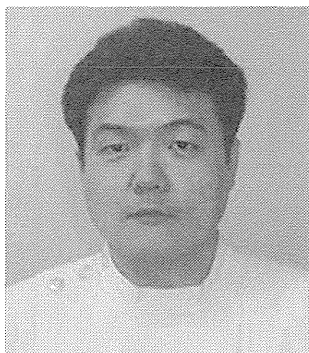
Department of Neurology, Graduate School of Medical Sciences, Kumamoto University

TAR-DNA-binding protein of 43kDa (TDP-43) was found to be one of the major disease proteins in the pathological inclusions of amyotrophic lateral sclerosis (ALS) and frontotemporal lobar degeneration (FTLD). Recently, several studies have shown the abnormal accumulation of TDP-43 in skeletal muscles of patients with sporadic inclusion body myositis (sIBM).

sIBM is a progressive myopathy characterized by muscle weakness and atrophy and onset of symptoms after 50 years of age. Although the disease is one of the most common myopathies in Caucasian people, a successful treatment for this disease is unavailable. sIBM belongs to the category of inflammatory myopathies and can be considered a conformational disorder, because it is associated with abnormal intracellular accumulation of multiple unfolded/misfolded proteins, including amyloid-beta (A β), phosphorylated tau (p-tau) in the form of paired helical filaments, and others, with ubiquitin immunoreactivity.

Increasing evidences suggest a similarity in the pathophysiological mechanisms of neuronal cell death in ALS and myofiber degeneration in sIBM. We here demonstrate that TDP-43, optineurin (OPTN), and to a lesser extent fused in sarcoma/translocated in liposarcoma (FUS/TLS) were more frequently accumulated in the cytoplasm in patients with sIBM and oculopharyngeal muscular dystrophy (OPMD) than in patients with polymyositis (PM), dermatomyositis (DM), or neurogenic muscular atrophy. Cu/Zn superoxide dismutase (SOD1) was accumulated in a small percentage of myofibers in patients with sIBM and OPMD, and to a very small extent in patients with PM and DM. Interestingly, confocal microscopy imaging showed that TDP-43 proteins more often colocalized with OPTN than with FUS/TLS, p62, and phosphorylated Tau. These findings suggest that OPTN in cooperation with TDP-43 might be involved in the pathophysiological mechanisms of skeletal muscular degeneration in myopathy with rimmed vacuoles.

To confirm the primary toxicity of TDP-43 to myofibers, we are generating transgenic mice having wild-type TDP-43 gene that is driven by muscle creatine kinase promoter. In the preliminary data at 8 week-old, the transgenic mice showed TDP-43-immunoreactive cytoplasmic aggregation in the quadriceps muscles. Interestingly, the mean diameter of myofibers with TDP-43 aggregates was significantly smaller than that of myofibers without aggregates or myofibers on non-transgenic littermates. For the better understanding of role of TDP-43 aggregation in neuromuscular degenerative diseases, the analyses of TDP-43 aggregation both in the sIBM patients and muscle-specific TDP-43 transgenic mice would be a useful tool.



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EDUCATION:

| | | | |
|----------------------------|-------|------|-----------------|
| Kumamoto University, Japan | M.D. | 1995 | Medicine |
| Kumamoto University, Japan | Ph.D. | 2003 | Medical Science |

POSITIONS:

2009-present: Assistant Professor, Dept. of Neurology, Kumamoto University Hospital

2007-2009: Neurologist and Research scientist, Dept. of Neurology, Kumamoto University Hospital

2004-2007: Postdoctoral research scientist, Dept. of Neurology, Columbia University, NY, USA

2003-2004: Neurologist (Dept. of Neurology, Kumamoto University Hospital, and Dept. of Neurology, National Kumamoto Hospital)

1999-2003: Ph.D. program, Graduate School of Medical Sciences, Kumamoto University

1995-1999: Medical Residency (Dept. of Internal Medicine, Kumamoto University Hospital, Dept. of Internal Medicine, Kumamoto City Hospital, and Dept. of Neurology, Kumamoto Saishunso Hospital)

OTHER EXPERIENCE AND PROFESSIONAL MEMBERSHIPS

1995: Japanese National License to practice medicine
2003: Diplomat, Japanese Board of Neurology
Member: Society for Neuroscience, Japanese Society of Neurology, The Japanese Society of Internal Medicine, The Japan Neuroscience Society

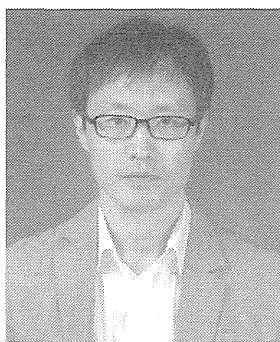
Mechanism of disease progression through cell-to-cell amyloid propagation in neurodegenerative diseases

Seung-Jae Lee, Ph.D.

Department of Biomedical Science and Technology,

Konkuk University, Seoul, Korea

Progressive accumulation of specific protein aggregates is a defining feature of many major neurodegenerative diseases, including Alzheimer's disease, Parkinson's disease, fronto-temporal dementia, Huntington's disease, and Creutzfeldt-Jakob disease (CJD). Findings from several recent studies have suggested that aggregation-prone proteins, such as tau, α -synuclein, polyglutamine-containing proteins, and amyloid- β , can spread to other cells and brain regions, a phenomenon considered unique to prion disorders, such as CJD and bovine spongiform encephalopathy. Cell-to-cell propagation of protein aggregates may be the general underlying principle for progressive deterioration of neurodegenerative diseases. This may also have significant implications in cell replacement therapies, as evidenced by the propagation of α -synuclein aggregates from host to grafted cells in long-term transplants in Parkinson's patients. In this talk, I will review recent progress in protein aggregate propagation in experimental model systems and discuss outstanding questions and future perspectives. Understanding the mechanisms of this pathological spreading may open the way to unique opportunities for development of diagnostic techniques and novel therapies for protein misfolding-associated neurodegenerative diseases.



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POSITIONS:

- 2011-present Professor, Department of Biomedical Science and Technology, Konkuk University, Seoul, Korea
- 2006-2011 Associate Professor, Department of Biomedical Science and Technology, Konkuk University, Seoul, Korea
- 2000-2006 Assistant Professor, The Parkinson's Institute, Sunnyvale, CA
- 1998-2000 Instructor in Neurology, Center for Neurologic Diseases, Harvard Medical School and Brigham and Women's Hospital, Boston, MA
- 1996-1998 Postdoctoral Fellow, Center for Neurologic Diseases, Harvard Medical School and Brigham and Women's Hospital, Boston, MA
- 1995-1996 Postdoctoral Fellow, Laboratory of Molecular Cardiology, NHLBI, NIH, Bethesda, MD

EDUCATION:

- 1992-1995 PhD Department of Life Science, Pohang University of Science and Technology, Pohang, Korea
- 1990-1992 M.S. Department of Life Science, Pohang University of Science and Technology, Pohang, Korea
- 1985-1989 B.S. Department of Biology Education, Seoul National University, Seoul, Korea

AWARDS:

- 1995-1997 NIH Postdoctoral Fellowship from Fogarty International Center
- 2000 International Parkinson's Research Award (Parkinson's disease foundation)
- 2010 Excellence in Basic Research (Ministry of Education, Science, and Technology)

PROFESSIONAL SOCIETIES:

- 1999- member, Society for Neuroscience
- 2002- member, American Society for Biochemistry and Molecular Biology
- 2009- member, Korean Society for Molecular and Cellular Biology
- 2007- member, Korean Society for Brain and Neural Science
- 2007- member, Korean Society for Neurodegenerative disease

Toxic protein conformational transition and amyloid fibril formation in the polyglutamine diseases.

Yoshitaka Nagai, M.D., Ph.D.

Department of Degenerative Neurological Diseases, National Institute of Neuroscience, National Center of Neurology and Psychiatry

Abnormal aggregation and deposition of misfolded proteins in the brain have been recognized as a common molecular pathogenesis of various neurodegenerative diseases including Alzheimer's disease, Parkinson's disease, and the polyglutamine (polyQ) diseases, which are classified as the conformational diseases. The polyQ diseases are a group of at least nine inherited neurodegenerative diseases including Huntington's disease and various spinocerebellar ataxias, which are caused by an abnormal expansion of the polyQ stretch (>35-40) within each unrelated disease-causative protein. The expansion of the polyQ stretch is thought to trigger misfolding and aggregation of the disease-causative proteins, leading to their deposition as inclusion bodies inside affected neurons, and eventually resulting in neurodegeneration. To elucidate the structural alterations of the expanded polyQ protein during the aggregation process, we performed structural analyses of the polyQ protein. We found that the expanded polyQ protein undergoes a conformational transition to a β -sheet dominant structure in the monomeric state, which precedes its assembly into insoluble amyloid-like fibrillar aggregates. Most importantly, we further revealed that the soluble β -sheet monomer of the expanded polyQ protein triggers cytotoxicity by microinjection experiments. From a therapeutic point of view, we identified QBP1 (SNWKWWPGIFD), a peptide sequence that preferentially binds to the expanded polyQ stretch by phage display screening. We showed that QBP1 prevents the toxic β -sheet transition and amyloid-like aggregation of the expanded polyQ protein *in vitro*, and further that QBP1 suppresses polyQ-induced neurodegeneration in *Drosophila*. From high-throughput screening of a chemical compound library (46,000), we have identified approximately 100 polyQ aggregate inhibitors as therapeutic candidates so far. We also utilized molecular chaperones, which belong to the quality control system against protein misfolding, and demonstrated that genetic expression and pharmacological induction of molecular chaperones suppress polyQ-induced neurodegeneration in *Drosophila* and mouse models. Finally, we demonstrated therapeutic effects of selective degradation of the expanded polyQ protein on a polyQ disease mouse model. We therefore conclude that protein misfolding and aggregation are promising therapeutic targets not only for the polyQ diseases, but also for other conformational neurodegenerative diseases.



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BIOGRAPHY:

- 2008-present Section Chief, Department of Degenerative Neurological Diseases, National Institute of Neuroscience, National Center of Neurology and Psychiatry
- 2007-2008 Associate Professor, Div of Clinical Genetics, Dept of Medical Genetics, Osaka University Graduate School of Medicine, Japan
- 2001-2007 Assistant Professor, same as the above
- 2000-2001 JSPS Postdoctoral fellow, Dept of Molecular Medical Science, Osaka Bioscience Institute, Japan
- 1997-2000 Research Associate, Div of Neurology, Dept of Medicine, Duke University Medical Center, NC, USA
- 1995 Ph.D. (Dr. of Medical Science), Osaka University Graduate School of Medicine, Japan
M.D., Osaka University Medical School, Japan

AWARDS:

The Award for Young Investigator of Japanese Society for Neurochemistry

SELECTED PUBLICATIONS:

- Nagai Y, Popiel HA. *Curr Pharm Des* 14: 3267-3279 (2008)
- Fujikake N, et al. *J Biol Chem* 283: 26188-26197 (2008)
- Takahashi Y, et al. *J Biol Chem* 282: 24039-24048 (2007)
- Nagai Y, et al. *Nat Struct Mol Biol* 14: 332-340 (2007)
- Popiel HA, et al. *Mol Ther* 15: 303-309 (2007)
- Nagai Y, et al. *Hum Mol Genet* 12: 1253-1260 (2003)
- Nagai Y, et al. *J Biol Chem* 275: 10437-10442 (2000)

Clearance of extracellular misfolded proteins in systemic amyloidosis: experience with transthyretin

Maria João Saraiva, Ph.D.

Institute for Molecular and Cellular Biology (IBMC) and Institute for Biomedical Sciences (ICBAS), University of Porto, Portugal

Extracellular protein misfolding and aggregation occurring in systemic amyloidosis triggers inflammation, oxidative stress, matrix remodeling, the unfolded-protein-response and ER pathways that resemble in many aspects, including common molecular players and scenarios, to those described in local amyloidoses affecting for example the central nervous system (CNS), such as Alzheimer Disease. Thus, similarities and dissimilarities in toxicity found between the CNS and the periphery are very useful to pinpoint and guide us to the treatment of aging-associated neurodegenerative disorders. Understanding the two-way crosstalk between the extracellular milieu and the cell is a major trend in diseases related to protein aggregation. In particular, mechanisms involved in the clearance of protein aggregates, both extra and intracellular are pivotal and need detailed analyses for the development of therapeutic strategies. Studies with small compounds or molecules, such as antibodies, known to recognize and disrupt amyloidogenic structures, have proven efficient in removing and promoting clearance of protein aggregates in studies with experimental models of misfolding disorders. However, the mechanisms and key players in these processes are largely unknown. Extracellular molecular chaperones are capable to repair or target damaged proteins to degradation by binding to extracellular misfolded proteins and promoting their disposal either by endocytosis for intracellular degradation or degradation by the extracellular matrix. It is clear that more than a single approach to treat FAP effectively is mandatory; in this regard, the experience achieved so far with clearance based approaches is very promising and will be discussed.



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EDUCATION:

- 1971-1976 B.Sc. (Biology), University of Porto (grade:16/20)
1976-1978 M.Sc. (Biochemistry), University of London
1980-1984 Ph.D. with Honors (Biochemistry), University of Porto
1991- Qualified as Professor in Biochemistry ("agregação"), University of Porto

RESEARCH AND PROFESSIONAL EXPERIENCE

- 1977-1984 Research Assistant, (leading to a PhD), Instituto de Ciências Biomédicas
1981-1983 Staff Associate, Medicine, Columbia, Univ. College of P&S, P&S, New York
1984-1986 Visiting Scientist, Medicine, Columbia University, College of P&S, New York (6 mo./year)
1984-1989 Assistant Professor (Biochemistry), Instituto de Ciências Biomédicas
1989 Visiting Scientist, Neurogenetics Unit, Massachusetts Gen Hospital, Boston, (Feb-July)
1989-1993 Associate Professor (Biochemistry), Instituto de Ciências Biomédicas
1990-1994 Member of the Board of Directors, Centro de Estudos de Paramiloidose
1992-1994 Visiting Scientist in the Institute of Cancer Research, Columbia University (3 months/year)
1984-1996 Research Associate, Centro de Estudos de Paramiloidose, Porto
1994-1995 Chairman of the Department of Molecular Biology, Instituto de Ciências Biomédicas
1997-2001 Director of the Amyloid Unit of Institute for Molecular and Celular Biology, Universidade do Porto
2007-2009 Coordinator of the Basic and Clinical Neurobiology Division at Institute for Molecular and Celular Biology - IBMC, Universidade do Porto
1999-present Member of the Board of Directors of the Center for Predictive and Preventive Medicine at the Institute for Mol and Celular Biology, Universidade do Porto
1994-present Full Professor (Biochemistry), Instituto de Ciências Biomédicas, Universidade do Porto, Portugal
2002-present Director of the Molecular Neurobiology Group at Institute for Molecular and Celular Biology - IBMC, Universidade do Porto
2010-present Vice-Director of Institute for Molecular and Celular Biology - IBMC, Universidade do Porto

