

- SF, et al. PACS-2 controls endoplasmic reticulum-mitochondria communication and Bid-mediated apoptosis. *EMBO J* 2005;24:717-729.
37. Kerppola TK. Design and implementation of bimolecular fluorescence complementation (BiFC) assays for the visualization of protein interactions in living cells. *Nat Protoc* 2006;1:1278-1286.
38. Kerppola TK. Bimolecular fluorescence complementation (BiFC) analysis as a probe of protein interactions in living cells. *Annu Rev Biophys* 2008;37:465-487.
39. Kato H. Differential roles of MDA5 and RIG-I helicases in the recognition of RNA viruses. *Nature* 2006;441:101-105.
40. Saitoh T, Fujita N, Hayashi T, Takahara K, Satoh T, Lee H, Matsunaga K, et al. Atg9a controls dsDNA-driven dynamic translocation of STING and the innate immune response. *Proceedings of the National Academy of Sciences* 2009;106:20842-20846.
41. Castanier C, Garcin D, Vazquez A, Arnoult D. Mitochondrial dynamics regulate the RIG-I-like receptor antiviral pathway. *EMBO Rep*;11:133-138.
42. Horner SM, Liu HM, Park HS, Briley J, Gale M. Mitochondrial-associated endoplasmic reticulum membranes (MAM) form innate immune synapses and are targeted by hepatitis C virus. *Proceedings of the National Academy of Sciences*;108:14590-14595.
43. Horner SM, Park HS, Gale M. Control of innate immune signaling and membrane targeting by the hepatitis C virus NS3/4A protease are governed by the NS3 helix  $\alpha 0$ . *Journal of Virology*.
44. Egger D, Wolk B, Gosert R, Bianchi L, Blum HE, Moradpour D, Bienz K. Expression of Hepatitis C Virus Proteins Induces Distinct Membrane Alterations Including a Candidate Viral Replication Complex. *J. Virol.* 2002;76:5974-5984.
45. Gretton SN, Taylor AI, McLauchlan J. Mobility of the hepatitis C virus NS4B protein on the endoplasmic reticulum membrane and membrane-associated foci. *Journal of General*

- Virology 2005;86:1415-1421.
46. Einav S, Elazar M, Danieli T, Glenn JS. A nucleotide binding motif in hepatitis C virus (HCV) NS4B mediates HCV RNA replication. *J Virol* 2004;78:11288-11295.
  47. Elazar M, Liu P, Rice CM, Glenn JS. An N-terminal amphipathic helix in hepatitis C virus (HCV) NS4B mediates membrane association, correct localization of replication complex proteins, and HCV RNA replication. *J Virol* 2004;78:11393-11400.
  48. Moriyama M, Kato N, Otsuka M, Shao RX, Taniguchi H, Kawabe T, Omata M. Interferon-beta is activated by hepatitis C virus NS5B and inhibited by NS4A, NS4B, and NS5A. *Hepatology* 2007;1:302-310.
  49. Xu J, Liu S, Xu Y, Tien P, Gao G. Identification of the nonstructural protein 4B of hepatitis C virus as a factor that inhibits the antiviral activity of interferon-alpha. *Virus Research* 2009;141:55-62.
  50. Hofmann WP, Zeuzem S. A new standard of care for the treatment of chronic HCV infection. *Nat Rev Gastroenterol Hepatol*;advance online publication.
  51. Einav S, Gerber D, Bryson PD, Sklan EH, Elazar M, Maerkl SJ, Glenn JS, et al. Discovery of a hepatitis C target and its pharmacological inhibitors by microfluidic affinity analysis. *Nat Biotech* 2008;26:1019-1027.
  52. Rai R, Deval J. New opportunities in anti-hepatitis C virus drug discovery: Targeting NS4B. *Antiviral Research*;90:93-101.
  53. Cho NJ, Dvory-Sobol H, Lee C, Cho SJ, Bryson P, Masek M, Elazar M, et al. Identification of a class of HCV inhibitors directed against the nonstructural protein NS4B. *Sci Transl Med*;2:15ra16.
  54. Bryson PD, Cho NJ, Einav S, Lee C, Tai V, Bechtel J, Sivaraja M, et al. A small molecule inhibits HCV replication and alters NS4B's subcellular distribution. *Antiviral Research*;87:1-8.

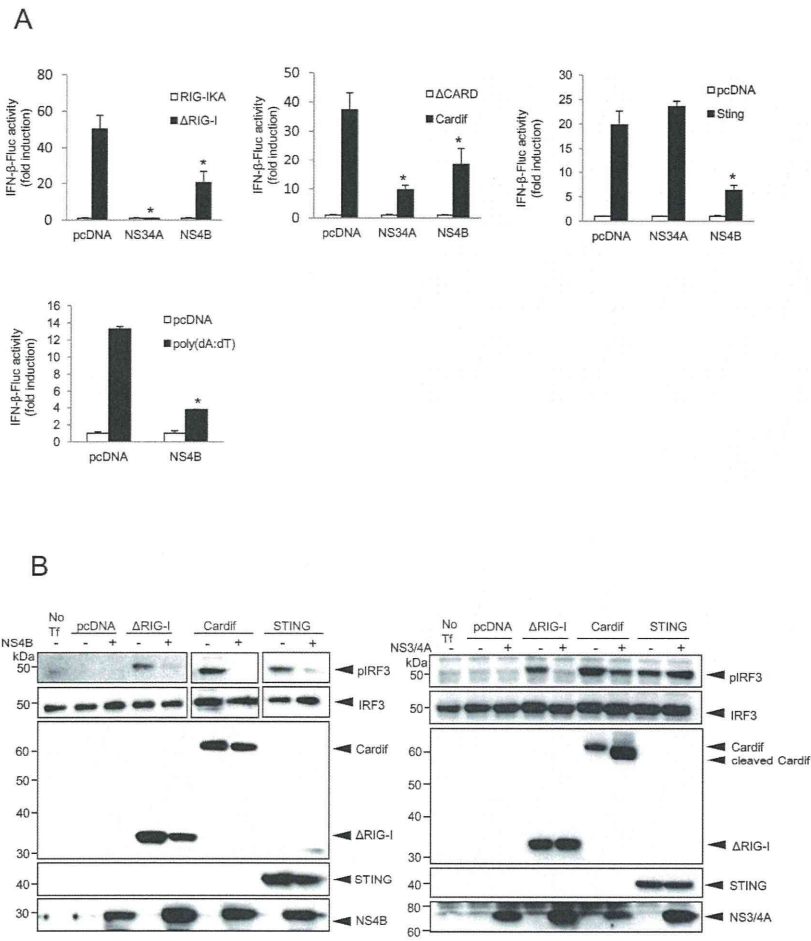


Figure 1. Nitta et al.

275x397mm (300 x 300 DPI)

AC

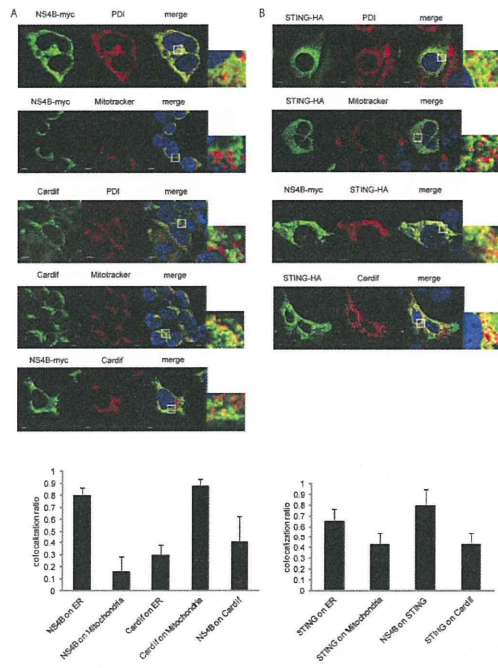


Figure 2. Nitta et al.

394x339mm (300 x 300 DPI)

Accep

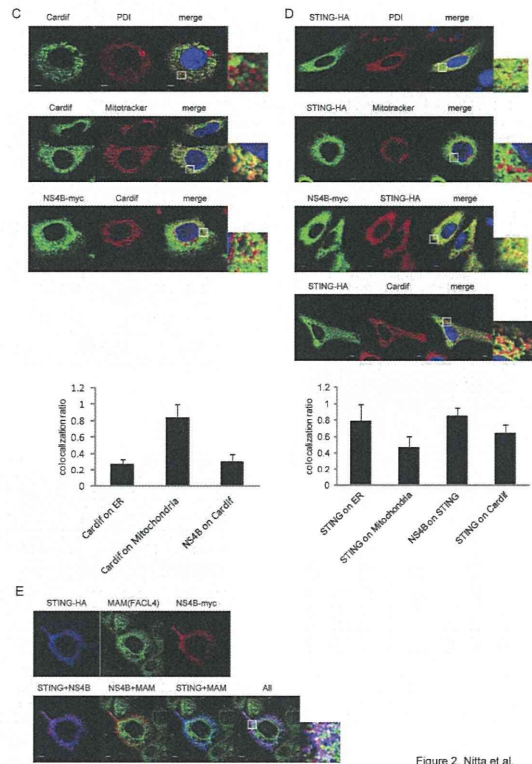


Figure 2. Nitta et al.

377x451mm (300 x 300 DPI)

AC

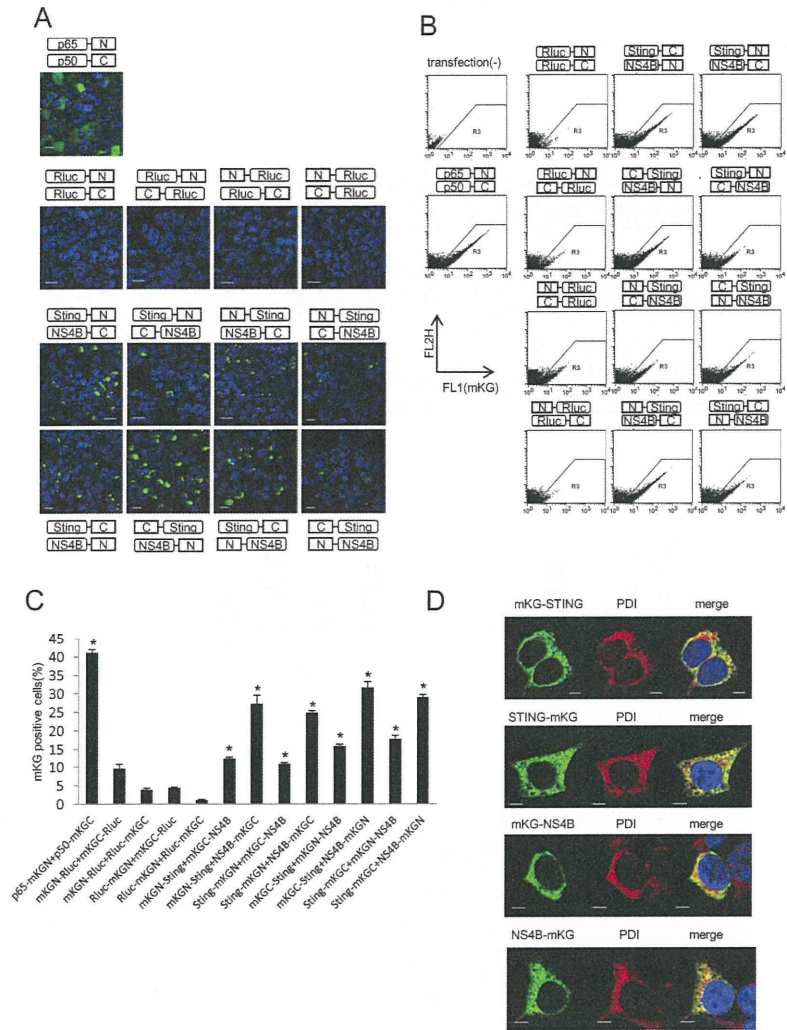


Figure 3. Nitta et al.

190x275mm (300 x 300 DPI)

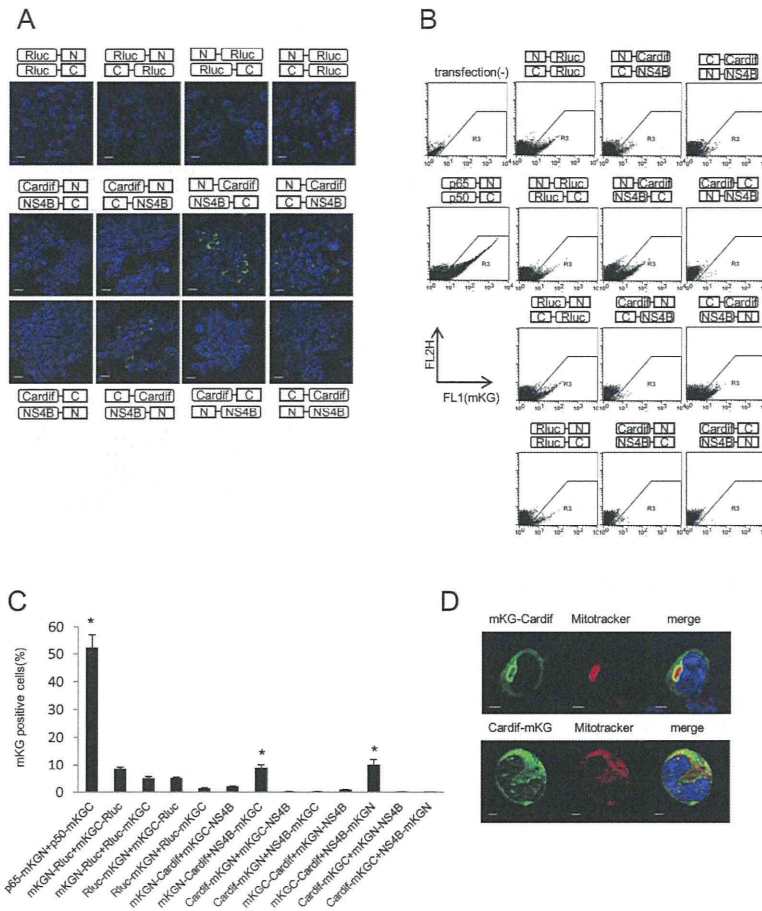


Figure 4. Nitta et al.

190x275mm (300 x 300 DPI)

AC

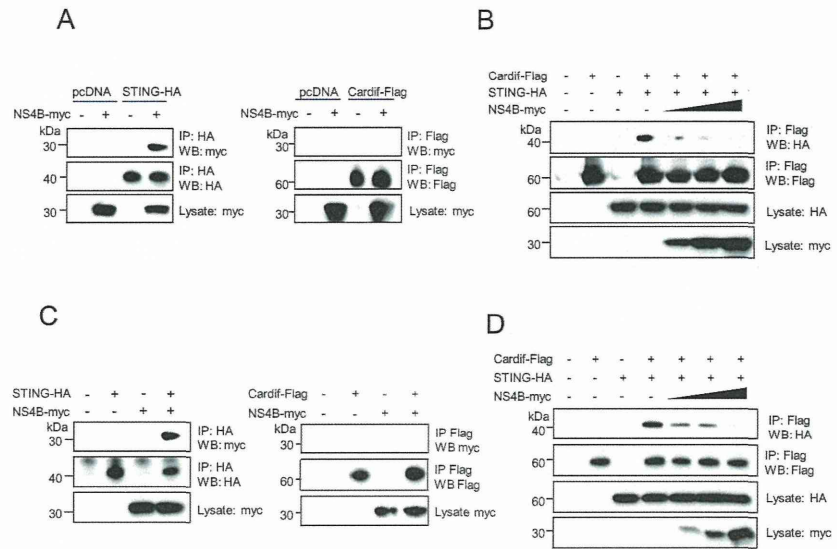


Figure 5. Nitta et al.

275x397mm (300 x 300 DPI)

AC



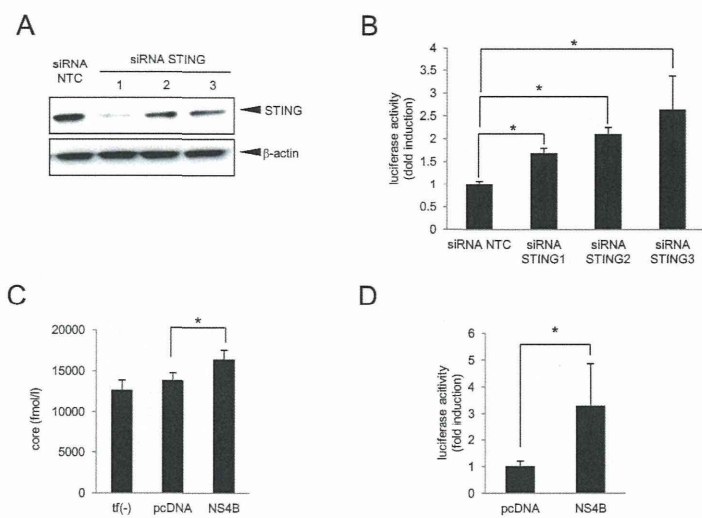


Figure 6. Nitta et al.

275x397mm (300 x 300 DPI)

AC

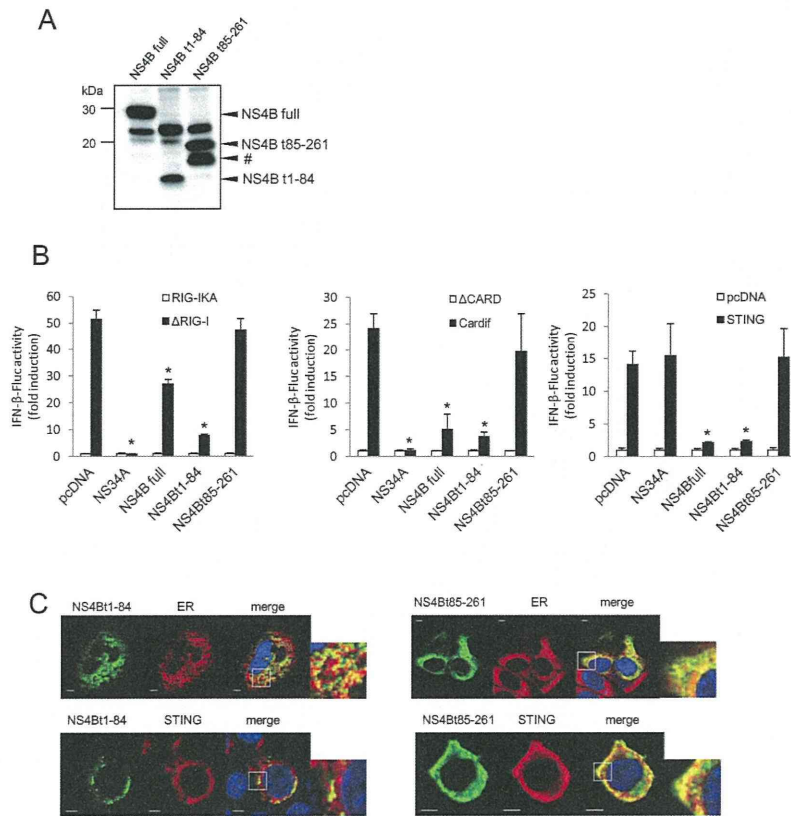


Figure 7. Nitta et al.

190x275mm (300 x 300 DPI)

AC

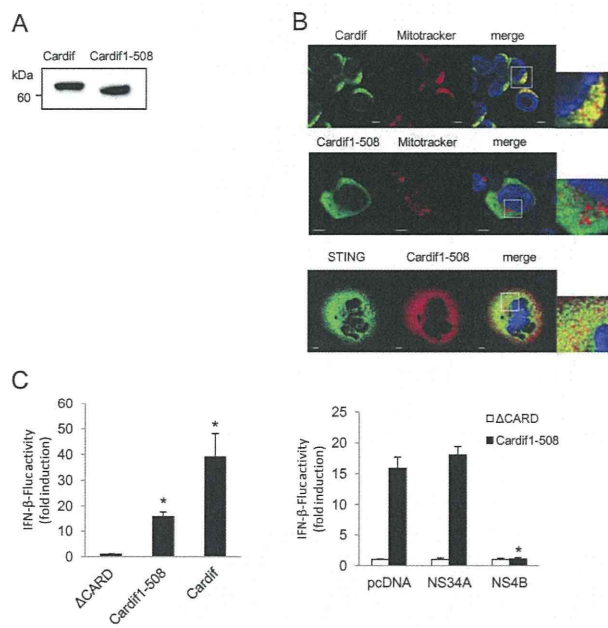


Figure 8. Nitta et al.

190x275mm (300 x 300 DPI)

AC

Figure 3D

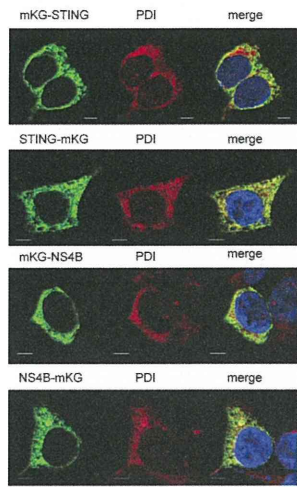


Figure 4D

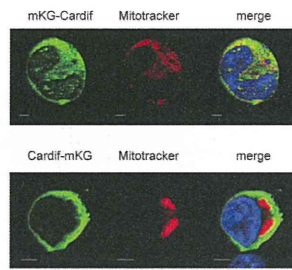


Figure 7C

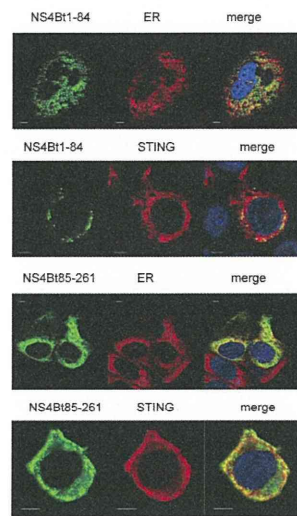
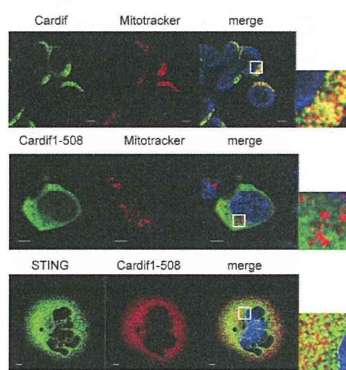


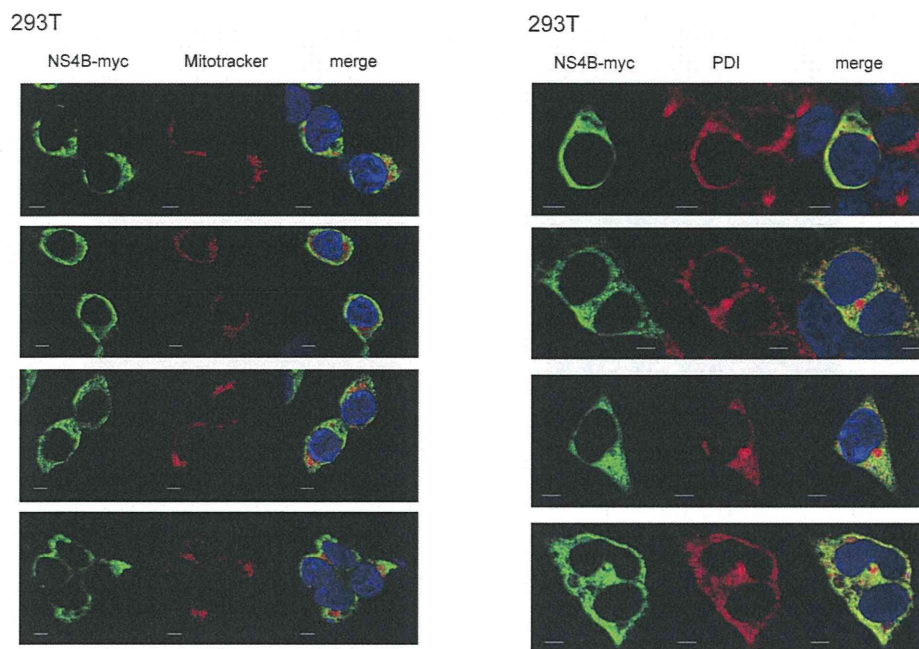
Figure 8B



Supplementary Figure for review Nitta et al.

243x311mm (300 x 300 DPI)

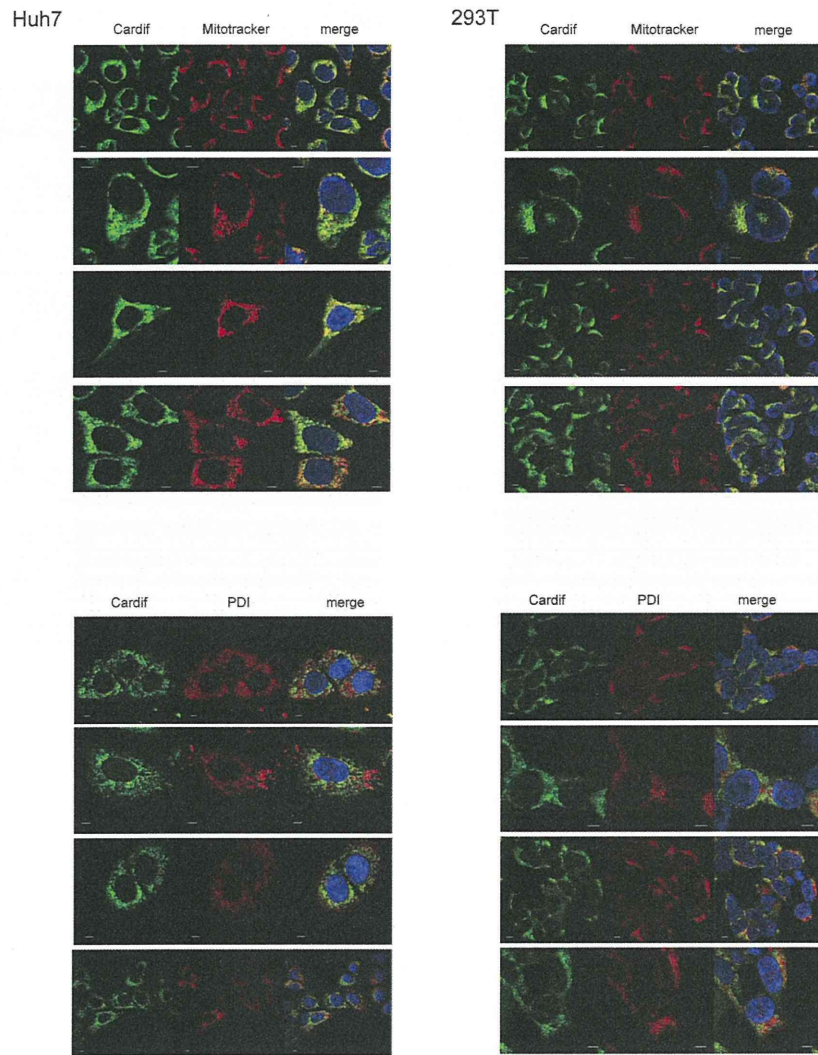
AC



Supplementary Figure for review Nitta et al.

172x225mm (300 x 300 DPI)

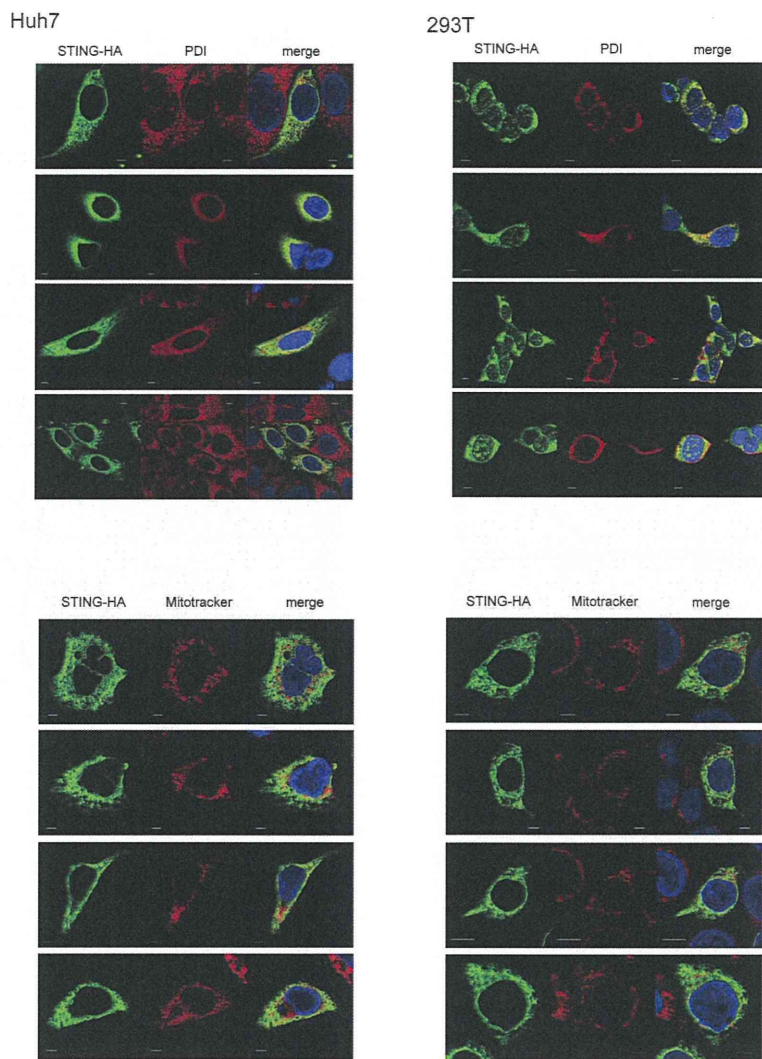
AC



Supplementary Figure for review Nitta et al.

201x281mm (300 x 300 DPI)

AC

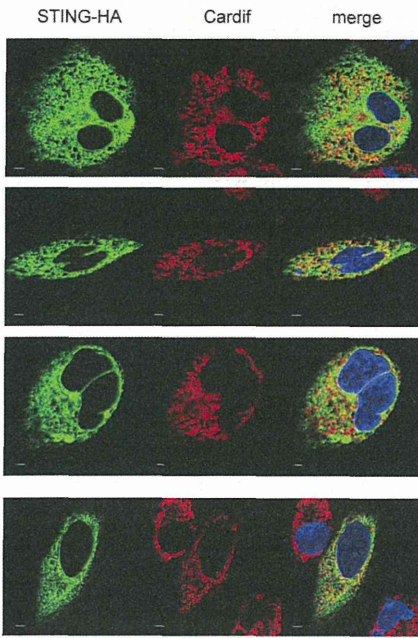


Supplementary Figure for review Nitta et al.

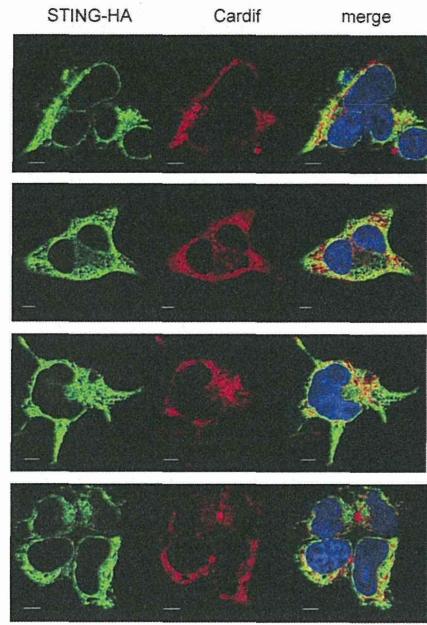
184x283mm (300 x 300 DPI)

AC

Huh7



293T

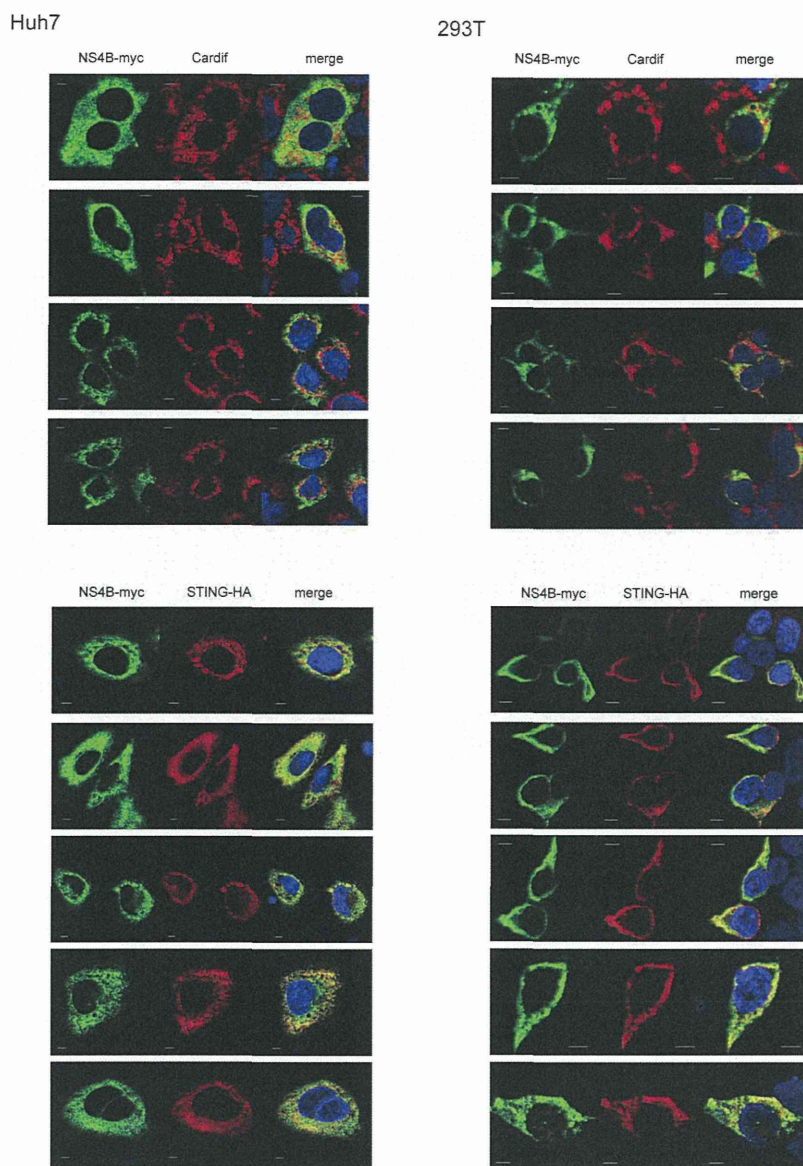


Supplementary Figure for review Nitta et al.

180x175mm (300 x 300 DPI)

Acce



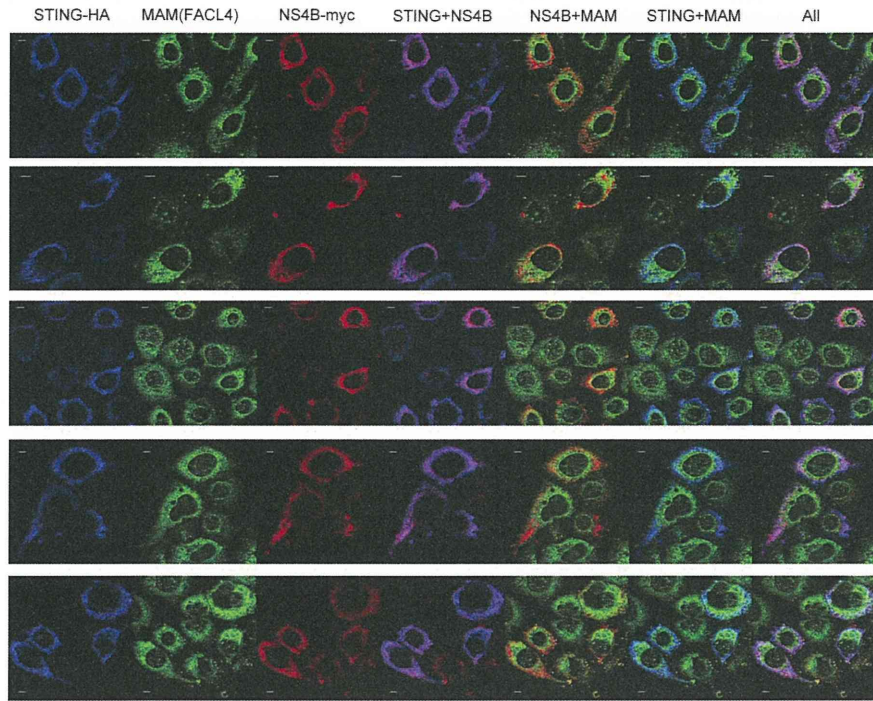


Supplementary Figure for review Nitta et al.

202x280mm (300 x 300 DPI)

AC

Huh7



Supplementary Figure for review Nitta et al.

187x240mm (300 x 300 DPI)

AC

# Add-on Therapy of Pitavastatin and Eicosapentaenoic Acid Improves Outcome of Peginterferon Plus Ribavirin Treatment for Chronic Hepatitis C

Motoyuki Kohjima,<sup>1</sup> Munechika Enjoji,<sup>2,3,4\*</sup> Tsuyoshi Yoshimoto,<sup>1</sup> Ryoko Yada,<sup>2</sup> Tatsuya Fujino,<sup>2</sup> Yoko Aoyagi,<sup>2</sup> Nobuyoshi Fukushima,<sup>1</sup> Kunitaka Fukuizumi,<sup>1</sup> Naohiko Harada,<sup>1</sup> Masayoshi Yada,<sup>5</sup> Masaki Kato,<sup>5</sup> Kazuhiro Kotoh,<sup>5</sup> Manabu Nakashima,<sup>4</sup> Naoya Sakamoto,<sup>6</sup> Yasuhito Tanaka,<sup>7</sup> and Makoto Nakamuta<sup>1,2</sup>

<sup>1</sup>Department of Gastroenterology, Kyushu Medical Center, Fukuoka, Japan

<sup>2</sup>Clinical Research Center, Kyushu Medical Center, Fukuoka, Japan

<sup>3</sup>Health Care Center, Fukuoka University, Fukuoka, Japan

<sup>4</sup>Department of Clinical Pharmacology, Fukuoka University, Fukuoka, Japan

<sup>5</sup>Department of Medicine and Bioregulatory Science, Kyushu University, Fukuoka, Japan

<sup>6</sup>Department of Gastroenterology and Hepatology, Tokyo Medical and Dental University, Tokyo, Japan

<sup>7</sup>Department of Clinical Molecular Informative Medicine, Nagoya City University Graduate School of Medical Sciences, Nagoya, Japan

Despite the use of pegylated-interferon (peg-IFN) plus ribavirin combination therapy, many patients infected with hepatitis C virus (HCV)-1b remain HCV-positive. To determine whether addition of pitavastatin and eicosapentaenoic acid (EPA) is beneficial, the “add-on” therapy option (add-on group) was compared retrospectively with unmodified peg-IFN/ribavirin therapy (standard group). Association of host- or virus-related factors with sustained virological response was assessed. In HCV replicon cells, the effects of pitavastatin and/or EPA on HCV replication and expression of innate-immunity- and lipid-metabolism-associated genes were investigated. In patients infected with HCV-1b, sustained virological response rates were significantly higher in the add-on than standard group. In both groups, sustained virological response rates were significantly higher in patients with genotype TT of IL-28B (rs8099917) than in those with non-TT genotype. Among the patients with non-TT genotype, sustained virological response rates were markedly higher in the add-on than standard group. By multivariate analysis, genome variation of IL28B but not add-on therapy remained as a predictive factor of sustained virological response. In replicon cells, pitavastatin and EPA suppressed HCV replication. Activation of innate immunity was obvious in pitavastatin-treated cells and EPA suppressed the expression of sterol regulatory element binding protein-1c and low-density lipoprotein

receptor. Addition of pitavastatin and EPA to peg-IFN/ribavirin treatment improved sustained virological response in patients infected with HCV-1b. Genotype variation of IL-28B is a strong predictive factor in add-on therapy.

**J. Med. Virol.** 85:250–260, 2013.

© 2012 Wiley Periodicals, Inc.

**KEY WORDS:** cholesterol; hepatitis C virus; IL28B; replicon system

Abbreviations: EPA, eicosapentaenoic acid; HCV, hepatitis C virus; HMGCR, HMG-CoA reductase; IRF3, IFN regulatory factor 3; ISG15, IFN-stimulated gene 15; ITPA, inosine triphosphatase; LDLR, low-density lipoprotein receptor; MAVS, mitochondrial antiviral signaling; NPC1L1, Niemann-Pick C1 like 1; OR, odds ratio; PCR, polymerase chain reaction; peg-IFN, pegylated-interferon; PUFA, polyunsaturated fatty acid; RIG-I, retinoic acid inducible gene I; SNP, single nucleotide polymorphism; SREBP, sterol regulatory element binding protein; TRAF6, TNF receptor associated factor 6.

Grant sponsor: Ministry of Health, Labor, and Welfare of Japan (Research Program of Intractable Disease); Grant sponsor: National Hospital Organization of Japan (Grant-in-Aid for Clinical Research).

Disclosure Statement: No competing financial interests exist.

\*Correspondence to: Munechika Enjoji, MD, Health Care Center, Fukuoka University, 8-19-1 Nanakuma, Jonan-ku, Fukuoka 814-0180, Japan. E-mail: enjoji@adm.fukuoka-u.ac.jp

Accepted 9 October 2012

DOI 10.1002/jmv.23464

Published online 14 November 2012 in Wiley Online Library (wileyonlinelibrary.com).

## INTRODUCTION

Nearly, 170 million people are infected with hepatitis C virus (HCV) worldwide and natural history studies show that 5–20% of patients develop cirrhosis after approximately 20 years of infection [Alter, 2005]. Currently, pegylated-interferon (peg-IFN) plus ribavirin combination therapy has become the standard care for chronic hepatitis C because it achieves high rates of sustained virological response [Aghemo et al., 2009]. However, in patients infected with genotype 1b HCV (HCV-1b), at most, 50% of individuals achieve a sustained virological response following combination therapy, and HCV-1b in high viral loads ( $>5.0$  log IU/ml) accounts for  $>70\%$  of patients with HCV infection in Japan [Kumada et al., 2006]. The response to IFN-based treatment is influenced by virus-related factors including viral load and genotypes; host-related factors, such as sex, age, insulin resistance, staging of the disease and responses to previous antiviral therapies; as well as therapeutic factors, such as dose and duration of treatment [Shiffman, 2002; Backus et al., 2007; Kanwal et al., 2007; Bortoletto et al., 2010]. In addition, as a critical genetic factor for governing the outcomes of peg-IFN plus ribavirin combination therapy, genome variation of IL28B and inosine triphosphatase (ITPA) have been identified recently. At the spot of rs8099917 in the IL28B region, patients infected with HCV-1b with the major variation type (TT) show markedly higher sustained virological response rates than those with the minor variation type (TG + GG) [Ge et al., 2009; Suppiah et al., 2009; Tanaka et al., 2009; Hayes et al., 2011]. Single nucleotide polymorphism (SNP) variation of the ITPA gene at rs1127354 is associated with anemia as an adverse effect during peg-IFN plus ribavirin combination therapy [Fellay et al., 2010; Azakami et al., 2011; Suzuki et al., 2011; Thompson et al., 2011]. In patients who have rs1127354 genotype CC (major type), ribavirin-induced anemia is more frequent and forces a reduction in dose of ribavirin, which worsens the therapeutic outcome. Alternatively, viral amino acid substitutions at core 70 and 91 are significant predictors of treatment outcome. In particular, a point mutation of core 70 from Arg to Gln is significantly associated with non-sustained virological response in patients infected with HCV-1b [Akuta et al., 2005, 2007; El-Shamy et al., 2012].

Investigation of patients treated by peg-IFN plus ribavirin combination therapy has indicated that serum cholesterol and statin use predict virological response to therapy [Harrison et al., 2010]. Recent studies have shown that virological response is improved by addition of fluvastatin or pitavastatin to peg-IFN and ribavirin treatment [Bader et al., 2008; Sezaki et al., 2009; Shimada et al., 2012]. Statins were associated with a reduced risk of hepatocellular carcinoma in a large cohort of patients with diabetes [El-Serag et al., 2009]. In other studies, it has been demonstrated that polyunsaturated fatty acids (PUFAs) inhibit HCV

replication by a mechanism that is independent of their roles in regulating lipogenesis [Leu et al., 2004; Kapadia and Chisari, 2005; Huang et al., 2007]. Takaki et al. [2007] have reported that eicosapentaenoic acid (EPA), a type of n-3 PUFA, allows maintenance of the original ribavirin dose in chronic hepatitis C patients during peg-IFN plus ribavirin combination therapy. However, the effects of these lipid modulators on chronic hepatitis C patients with intractable IL-28B allele remain unknown.

As a result of this experimental and therapeutic evidence, a new antiviral strategy to improve treatment outcome for chronic hepatitis C was designed, that is, addition of pitavastatin and EPA to peg-IFN plus ribavirin combination therapy (add-on therapy). The validity of the add-on therapy was evaluated by comparing its effect on the final outcome (i.e., sustained virological response) with that of unmodified peg-IFN plus ribavirin combination therapy (standard therapy), and pretreatment predictors of virological response were investigated. Additionally, the antiviral effect of pitavastatin and/or EPA was estimated in HCV replicon cells.

## MATERIALS AND METHODS

### Study Patients

In Kyushu Medical Center, a standard protocol in Japan (subcutaneous peg-IFN $\alpha$ 2a [180  $\mu$ g] or peg-IFN $\alpha$ 2b [median dose of 1.5  $\mu$ g/kg, range 1.3–1.7] weekly, along with oral ribavirin daily for 48 weeks) was adopted for chronic hepatitis C patients from 2005 to 2008. The dose of ribavirin was adjusted according to body weight: 600 mg for patients weighing  $<60$  kg, 800 mg for those weighing 60–80 kg, and 800 mg for those weighing  $>80$  kg. From 2008, oral pitavastatin (2 mg/day) and ethyl icosapentate (1,800 mg/day) have been added to the standard protocol (add-on protocol). It has been shown that statins contribute to improving the virological response [Bader et al., 2008; Sezaki et al., 2009]. The add-on protocol was expected to improve treatment, and was applied to all patients after 2008 in Kyushu Medical Center, but a randomized study could not be designed. In these protocols, 48- and 24-week regimens were applied to patients infected with HCV-1b and HCV-2, respectively. Patients who experienced previous therapy using peg-IFN were excluded. Patients with cirrhosis were not included. Because of the possibility that vitamin E and bile acids including ursodeoxycholic acid promote HCV replication [Chang and George, 2007; Yano et al., 2007; Scholtes et al., 2008; Nakamura et al., 2010], treatment with these agents was withdrawn at least 1 month before the initiation of antiviral treatment. The study protocol was approved by the Ethics Committee of the National Hospital Organization, and written informed consent was obtained from all patients. Finally, 238 patients (genotype 1b/2 = 176/62) who were treated with the standard protocol (standard group) and 162 patients (genotype 1b/2 = 101/61) who were treated with the add-on protocol