4 weeks), 26 weeks (270 patients; daily for 4 weeks followed by 2 or 3 times a week), 52 weeks (103 patients; 2 or 3 times a week), 104 weeks (80 patients; 2 or 3 times a week), and more than 104 weeks (73 patients; 2 or 3 times a week). The median duration of treatment was 26 weeks (range 4–981).

The numbers of responders were evaluated at 6 months and 1, 3, 5, and 10 years after the completion of IFN therapy. In the baseline HBeAg-positive patients, responders were defined as patients who showed normalization of serum ALT level (normal level 6–30 IU/L), HBeAg clearance, and low HBV DNA level (<5 log copies/mL) at 6 months after completion of IFN therapy. In addition, baseline HBeAg-positive patients who showed continuous normalization of ALT levels, HBeAg clearance, and low HBV DNA level for more than 6 months until each test point at 1, 3, 5, and 10 years after completion of IFN therapy were also classified as "responders." In the baseline HBeAg-negative patients, responders were defined as those who showed sustained normalization of ALT level and low HBV DNA level (<4 log copies/mL) for more than 6 months until each test point after completion of IFN therapy.

All patients not considered to be responders were termed "non-responders." Patients receiving other therapies (IFN or nucleoside/nucleotide analogues) after the completion of IFN therapy were also termed non-responders.

Blood tests and serum viral markers

Routine biochemical tests were performed monthly via standard procedures during and for the first 12 months following the completion of IFN treatment and at least every 2 months thereafter. Levels of HBsAg, HBeAg, and anti-HBe were determined using radioimmunoassay kits (Abbott Diagnostics, Chicago, IL, USA) or a chemiluminescent enzyme immunoassay (CLEIA; Lumipulse System; Fujirebio, Tokyo, Japan). HBV DNA levels were measured using a branched-chain DNA probe assay (bDNA) (Chiron Laboratory Service, Van Nuys, CA, USA), a transcription-mediated amplification and hybridization protection assay (TMA-HPA) (Chugai Diagnostics Science, Tokyo, Japan), or a polymerase chain reaction (PCR)-based assay (COBAS Amplicor HBV Monitor Test or COBAS TaqMan HBV Test; Roche Diagnostics, Indianapolis, IN, USA).

HBV genotype

The major genotypes of HBV were determined using an enzyme-linked immunosorbent assay (ELISA; Institute of Immunology, Tokyo, Japan) or a PCR-invader assay (BML, Tokyo, Japan) according to the methods described by Usuda et al. [16] or Tadokoro et al. [17].

Statistical analysis

Differences between groups were examined for statistical significance using the χ^2 or Fisher's exact test and Mann–Whitney U-test where appropriate. Independent predictive factors associated with response to IFN treatment were determined using multivariate multiple logistic regression. The following 14 potential predictors of response to IFN treatment were assessed in this study: age, sex, pretreatment with IFN, duration of IFN treatment, severity of liver disease (CH or liver cirrhosis), HBV genotype, and levels of aspartate transaminase (AST), ALT, bilirubin, albumin, platelets, α fetoprotein (AFP), HBeAg, and HBV DNA. All factors found to be at least marginally associated with response to IFN therapy ($P < 0.10$) were entered into the multivariate multiple logistic regression analysis. The above calculations were performed using the Windows SPSS software package version 11.0.1 J (SPSS, Chicago, IL, USA).

The odds ratios (ORs) and 95% confidence intervals (CIs) were calculated to assess the relative risk. Independent risk factors predicting the achievement of HBsAg seroclearance were studied using stepwise Cox regression analysis. Potential factors predicting the achievement of HBsAg seroclearance assessed here were the above 14 variables, each transformed into categorical data consisting of two simple ordinal numbers for univariate and multivariate analyses. All factors found to be at least marginally associated with HBsAg seroclearance ($P < 0.10$) were tested in the multivariate Cox proportional hazard model. A Kaplan–Meier estimate was performed using the SPSS software, and P values were calculated using the Cox-Mantel log-rank test. A two-tailed P value of <0.05 was considered statistically significant.

Results

Study population

Twenty-four (4%), 37 (6%), 504 (82%), 1 (0.2%), 1 (0.2%), and 1 (0.2%) patients were infected with HBV genotypes A, B, C, D, H, and B + C, respectively. Genotype could not be measured in the remaining 47 patients. The baseline characteristics of the patients are shown in Table 1. Although few patients had genotypes A and B, the distribution of HBV genotype was similar to that in patients with CHB who had received care in our hospital, with a follow-up period of more than 2 years [18]. Twenty-two of 24 patients with genotype A, 14 of 37 with

genotype B, 342 of 504 with genotype C, 1 of 1 with genotype H, and 34 of 47 with unknown genotype were HBeAg-positive at the commencement of treatment. While we were able to measure HBV DNA levels in 254 patients at the commencement of IFN therapy, levels in the remaining 361 could not be measured owing to a lack of commercial kits before the bDNA assay was available. The numbers of patients receiving other additional therapies after the completion of IFN therapy were 111 (HBeAg-positive/-negative, 90/21), 92 (67/25), 34 (25/9), and 61 (39/22) at the 1-, 3-, 5-, and 10-year time points, respectively.

Response to interferon therapy in all patients

The IFN response rates in all patients were 21% (105/497), 18% (86/491), 21% (90/428), 23% (82/359), and 25% (59/235) at 6 months and 1, 3, 5, and 10 years, respectively, after completion of the IFN therapy (Fig. 1). In patients with genotype A, the response rate was highest at 6 months post-treatment and gradually decreased at subsequent time points from 1 to 10 years thence. In patients with genotype B, response rates were over 20% at all time points except for 6 months post-treatment, whereas rates in patients with genotype C were under 25% at all time points (Fig. 2a).

Evaluation of efficacy of IFN in relation to clinical factors in all patients

The data of all patients were subjected to univariate analyses to determine the clinical factors contributing to the efficacy of IFN at each time point. We then investigated the significance of response to IFN therapy using multivariate logistic regression analysis.
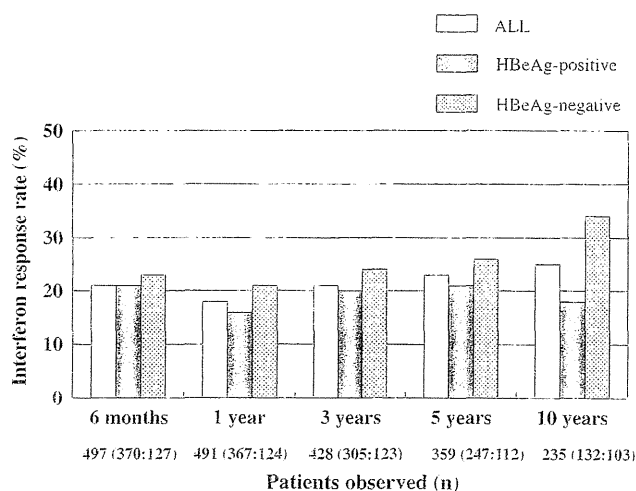

Fig. 1 Interferon response rates of all patients and hepatitis B e antigen (HBeAg)-positive and HBeAg-negative patients at 6 months and 1, 3, 5, and 10 years

Multivariate analyses including the variables noted above revealed several parameters that independently influenced the outcome of IFN therapy; namely, at 6 months: age ($P = 0.013$), HBV DNA level ($P = 0.019$), and duration of treatment ($P = 0.034$); at 1 year: HBV DNA level ($P < 0.001$) and age ($P = 0.001$); at 3 years: duration of treatment ($P < 0.001$), age ($P = 0.013$) and albumin level ($P = 0.013$); at 5 years: albumin level ($P = 0.004$), sex ($P = 0.005$), and pretreatment with IFN ($P = 0.039$); and at 10 years: HBeAg ($P < 0.001$) (Table 2).

Response to interferon therapy and evaluation of efficacy of IFN in relation to clinical factors in HBeAg-positive patients

Response rates in baseline HBeAg-positive patients were 21% (76/370), 16% (60/367), 20% (61/305), 21% (53/247), and 18% (24/132) at 6 months and 1, 3, 5, and 10 years, respectively (Fig. 1). In patients with genotype A, the response rate was highest at 6 months post-treatment and the rate was roughly equivalent to the 6 months post-treatment rate at subsequent time points from 1 to 10 years. Response rates in patients with genotype B in particular were above 40% at all time points except at 6 months, although few patients had genotype B. On the other hand, response rates in patients with genotype C were under 20% at all time points (Fig. 2a).

In addition, multivariate analyses in HBeAg-positive patients also revealed several parameters that independently influenced the outcome of IFN therapy–at 6 months: duration of treatment ($P = 0.001$) and age ($P = 0.014$); at 1 year: age ($P = 0.011$) and HBV DNA level ($P = 0.027$); at 3 years: sex ($P = 0.008$), duration of treatment ($P = 0.019$), age ($P = 0.020$), pretreatment with IFN ($P = 0.029$), and albumin level ($P = 0.043$); at 5 years: sex ($P = 0.002$) and pretreatment with IFN ($P = 0.005$); and at 10 years, genotype ($P = 0.019$) and AST ($P = 0.035$) (Table 3).

Response to interferon therapy and evaluation of efficacy of IFN in relation to clinical factors in HBeAg-negative patients

Response rates in baseline HBeAg-negative patients were 23% (29/127), 21% (26/124), 24% (29/123), 26% (29/112), and 34% (35/103) at 6 months and 1, 3, 5, and 10 years, respectively (Fig. 1). Rates in patients with genotype C were gradually increased at subsequent time points, whereas those in patients with genotype B remained under 30% at all time points (Fig. 2b).

In addition, univariate and multivariate analyses in HBeAg-negative patients revealed that duration of treatment (≥ 1 year) independently influenced the outcome of

A

☐ Genotype A
▨ Genotype B
▨ Genotype C

B

☐ Genotype A
▨ Genotype B
▨ Genotype C

C

☐ Genotype A
▨ Genotype B
▨ Genotype C
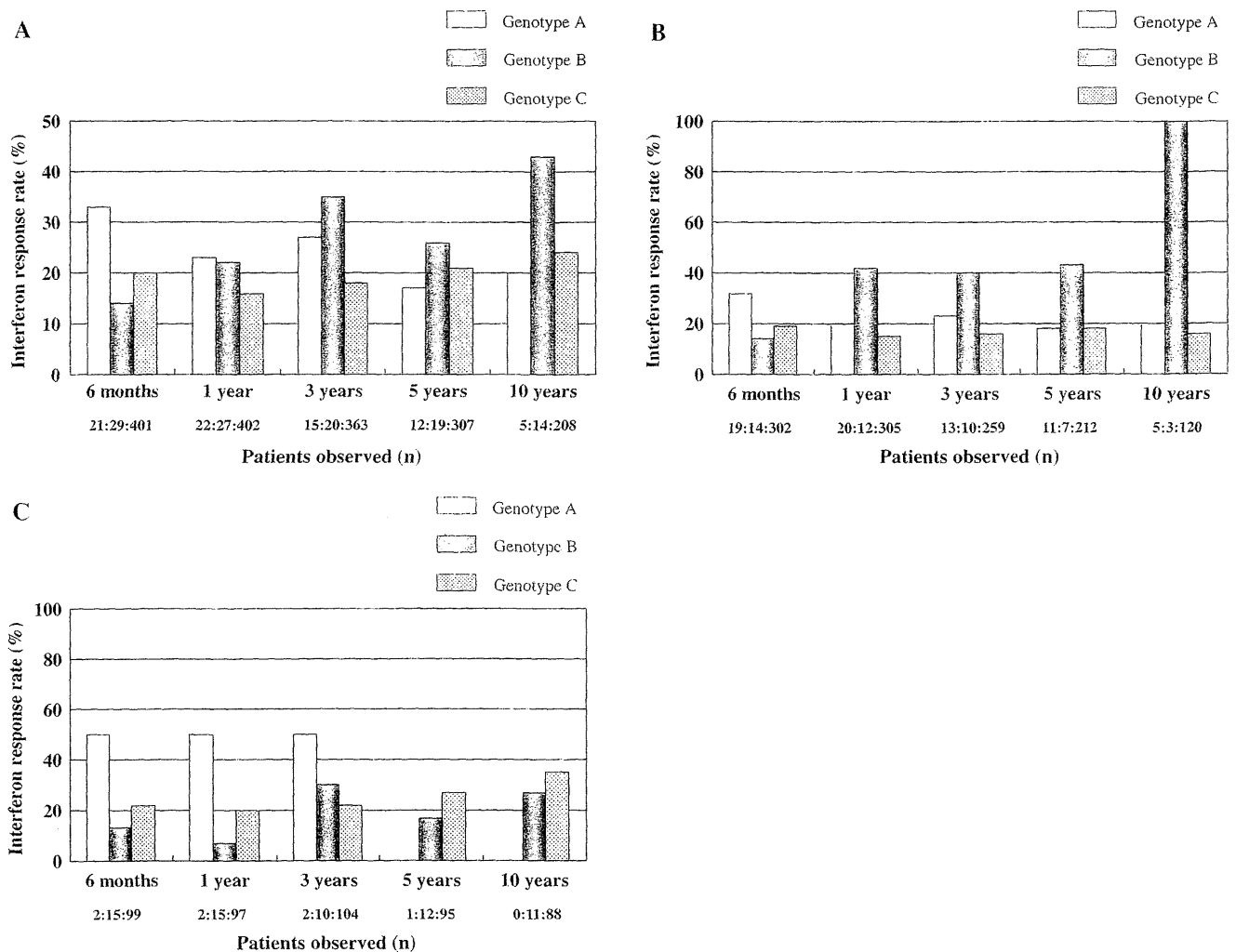

Fig. 2 Interferon response rates of patients with genotypes A, B, and C at 6 months and 1, 3, 5, and 10 years. a All patients, b HBeAg-positive patients, c HBeAg-negative patients

IFN therapy at 6 months, and at 1 and 3 years. No parameters independently influenced the outcome of IFN therapy at 5 or 10 years.

Evaluation of efficacy of IFN in relation to HBs antigen seroclearance

The HBsAg seroclearance rate in this study was obtained from patients who received IFN therapy alone: 69 of 615 patients (11%) achieved seroclearance of HBsAg. The cumulative HBsAg seroclearance rates in all patients from the commencement date of IFN therapy were 6.5% at 5 years, 15% at 10 years, 35% at 15 years, and 44% at 20 years (Kaplan–Meier method; Fig. 3a). No patients experienced the reappearance of HBsAg after seroclearance. Five factors found to be associated with achievement of HBsAg seroclearance on univariate analysis were: male sex $(P = 0.002)$, age ≥ 30 years $(P = 0.011)$, genotype A $(P = 0.038)$, HBeAg-negativity $(P = 0.045)$, and bilirubin

≤ 1.0 mg/dL $(P = 0.064)$. On multivariate analysis, independent factors predicting the achievement of HBsAg seroclearance were: age ≥ 30 years, genotype A, and male sex (Table 4). The cumulative HBsAg seroclearance rate for genotype A patients was significantly higher than the rate for those with genotypes B or C $(P = 0.0116)$ (Fig. 3b).

Relationship between the response to IFN and the development of hepatocellular carcinoma

Twenty-nine patients developed hepatocellular carcinoma (HCC) during the observation period, excluding 17 patients who received other additional therapies after the completion of IFN therapy and developed HCC thereafter. IFN response rates in the 29 patients who developed HCC were 5% (1/22), 5% (1/20), 10% (2/20), 13% (2/15), and 13% (2/16), respectively, at 6 months and 1, 3, 5, and 10 years after the completion of IFN. No patient developed HCC after HBsAg seroclearance.

Table 2 Factors associated with response to interferon therapy for all patients at 6 months and 1, 3, 5, and 10 years

Parameter	Univariate analysis		Multivariate analysis	
	OR (95% CI)	P	OR (95% CI)	P
6 Months after completion of IFN therapy (n = 229)				
Duration of treatment (≥1 year)	2.680 (1.724–4.166)	<0.001	2.107 (1.058–4.198)	0.034
HBV DNA level (≤7.0 log copies/mL)	2.165 (1.107–4.219)	0.026	2.309 (1.148–4.630)	0.019
Age (<30 years)		0.057	2.451 (1.209–4.950)	0.013
1 year after completion of IFN therapy (n = 231)				
Duration of treatment (≥1 year)	2.553 (1.588–4.104)	<0.001		
HBV DNA level (≤7.0 log copies/mL)	3.268 (1.597–6.667)	0.001	4.464 (2.058–9.709)	<0.001
Age (<35 years)	1.799 (1.125–2.874)	0.014	3.831 (1.718–8.547)	0.001
3 years after completion of IFN therapy (n = 397)				
Duration of treatment (≥1 year)	2.410 (1.495–3.885)	<0.001	2.739 (1.618–4.634)	<0.001
Age (<30 years)	2.070 (1.215–3.521)	0.009	2.110 (1.171–3.802)	0.013
Albumin (≥3.9 g/dL)	1.697 (1.045–2.757)	0.030	2.009 (1.158–3.486)	0.013
Genotype (non-C)	2.155 (1.033–4.504)	0.041		
5 years after completion of IFN therapy (n = 356)				
Albumin (≥3.9 g/dL)	1.869 (1.108–3.153)	0.017	2.321 (1.316–4.093)	0.004
Pretreatment with IFN (positive)	1.770 (1.016–3.084)	0.048	1.821 (1.029–3.222)	0.039
Sex (female)		0.060	2.381 (1.297–4.367)	0.005
Duration of treatment (≥1 year)		0.080		
10 years after completion of IFN therapy (n = 234)				
HBeAg (negative)	2.315 (1.269–4.219)	0.006	2.252 (1.230–4.115)	0.009
ALT (≥100 IU/L)	1.972 (1.053–3.690)	0.036		
Pretreatment with IFN (positive)		0.058		

ALT alanine transaminase, *IFN* interferon, *HBV* hepatitis B virus, *HBeAg* hepatitis B e antigen, *CI* confidence interval, *OR* odds ratio, *n* number submitted to multivariate analysis, including all factors found to be associated with response to IFN therapy

Discussion

Although IFN has been reported to exert beneficial effects in CHB patients, the response rate is not high. A meta-analysis published in 1993 reviewed 15 randomized controlled studies involving 837 adult patients who received IFN-α for 4–6 months, and elimination of HBeAg occurred in 33% of the treated patients [8]. In previous studies, we found the response rates among HBeAg-positive patients at 6 months after the completion of therapy to be 20 and 31% for 6 months and 1 year of IFN therapy, respectively [13, 15]. Although a recent meta-analysis reported that IFN increased the incidence of HBeAg and HBsAg seroclearance after long-term follow up of 3–7 years [12], the factors that influenced the clinical outcome were unclear.

In Japan, from 1988, 4-week IFN treatment was reimbursed by the healthcare system, and since 2002, 24-week IFN treatment has been conducted. In the present study, these two regimens were the major ones, and other regimens were used in clinical studies at our hospital (including previously reported studies [14, 15]). Although the durations of treatment differed, we analyzed the factors

associated with long-term response to IFN therapy, including the factor of duration of treatment.

In the present study, response rates were low among HBeAg-positive patients and relatively high among HBeAg-negative patients at all time points. Approximately 20% of the HBeAg-positive patients had sustained a response at 6 months to 10 years of follow up. Long-term follow-up studies after a four- to six-month course of IFN therapy in HBeAg-positive patients in European and Taiwanese studies showed higher (33–75%) response rates (HBeAg loss) than our study [7, 19, 20]. The difference in response rates between our present study and previous studies in other countries may be due to differences in ethnicity or HBV genotype (mainly genotype C in Japan). Moreover, the low IFN response rates at 1, 3, 5, and 10 years in the HBeAg-positive patients in our study were likely due to the change in treatments (IFN or nucleoside/nucleotide analogues). On the other hand, the response rates of HBeAg-negative patients in the present study were about 20% at 6 months and gradually increased thereafter. The sustained response rate in HBeAg-negative patients was usually <30% in European studies [21–23]. The response

Table 3 Factors associated with response to interferon therapy for HBeAg-positive patients at 6 months and 1, 3, 5, and 10 years

Parameter	Univariate analysis		Multivariate analysis	
	OR (95% CI)	P	OR (95% CI)	P
6 months after completion of IFN therapy ($n = 279$)				
Duration of treatment (≥ 1 year)	2.449 (1.457–4.114)	0.001	2.801 (1.540–5.096)	0.001
Age (<35 years)	1.855 (1.112–3.096)	0.017	2.128 (1.164–3.891)	0.014
1 year after completion of IFN therapy ($n = 172$)				
Duration of treatment (≥ 1 year)	2.483 (1.407–4.380)	0.002		
HBV DNA level (≤ 7.0 log copies/mL)	3.509 (1.495–8.264)	0.005	3.003 (1.130–7.937)	0.027
Age (<35 years)	1.996 (1.133–3.521)	0.015	3.610 (1.351–9.615)	0.011
3 years after completion of IFN therapy ($n = 283$)				
Age (<35 years)	2.041 (1.155–3.597)	0.013	2.083 (1.122–3.861)	0.020
Duration of treatment (≥ 1 year)	2.055 (1.153–3.661)	0.016	2.130 (1.132–4.008)	0.019
Pretreatment with IFN (positive)	2.054 (1.050–4.019)	0.041	2.336 (1.091–4.998)	0.029
Albumin (≥ 3.9 g/dL)		0.055	1.974 (1.020–3.820)	0.043
Sex (female)		0.089	2.646 (1.284–5.464)	0.008
5 years after completion of IFN therapy ($n = 247$)				
Sex (female)	2.571 (1.328–4.975)	0.006	2.924 (1.477–5.814)	0.002
Pretreatment with IFN (positive)	2.460 (1.213–4.988)	0.015	2.870 (1.377–5.980)	0.005
10 years after completion of IFN therapy ($n = 122$)				
Genotype (non-C)	5.319 (1.222–23.26)	0.032	6.410 (1.364–30.30)	0.019
AST (≥ 100 IU/L)		0.081	2.932 (1.078–7.972)	0.035

AST aspartate transaminase, *IFN* interferon, *HBV* hepatitis B virus, *HBeAg* hepatitis B e antigen, *CI* confidence interval, *OR* odds ratio, *n* number submitted to multivariate analysis, including all factors found to be associated with response to IFN therapy

rates of HBeAg-negative patients in our present study and the studies in other countries [21–23] were similar.

Few reports have identified the factors associated with long-term virological response to IFN therapy. In our present study, HBeAg-negativity was the most important factor for predicting a long-term response (10 years). While the HBV DNA level was important for predicting the response at 6 months and 1 year for all patients and the response at 1 year for HBeAg-positive patients, other factors (age, sex, albumin level, AST, IFN pretreatment, and duration of treatment) were found to be important at some time points for all patients and for HBeAg-positive patients. The HBV DNA level may not have been associated with long-term response to IFN therapy because the follow-up period (median 5.7 years) in patients with an HBV DNA level measurable with commercial kits was significantly shorter than that in the other patients (median 11.2 years: $P < 0.001$).

Previous studies have reported that high ALT levels, low HBV DNA level, female sex, and elevated liver activity and level of fibrosis on liver biopsy were major pretreatment factors correlated with a response to IFN [8–11, 24]. However, in these studies the follow-up times for judging the response were short (typically 6 months to 1 year). Our present study has clarified that HBeAg, HBV DNA level, age, sex, IFN pretreatment, duration of treatment, and levels of albumin and AST are important factors in the

long-term response to IFN. Further, non-C genotype was an important factor for long-term response in HBeAg-positive patients. Kao et al. [25] and Lin et al. [20] reported that HBV genotype B was associated with a higher response rate to IFN-α therapy than genotype C among CHB patients positive for HBeAg. Similarly, response rates among HBeAg-positive patients with genotype B in the present study were also higher than the response rates in those with genotype C in terms of long-term response (Fig. 2b). The long-term response rate among HBeAg-negative patients was relatively higher than that in HBeAg-positive patients. Previous reports have shown that response rates to a 6- to 12-month course of IFN-α in HBeAg-negative CHB patients range from 10 to 47% (average 24%) [26–29]. In addition, our previous report showed that 9 of 12 (75%) patients who received IFN-β twice per week for 24 weeks responded to the therapy [14]. However, the follow-up periods of these studies were short, and the long-term efficacy has not been clarified. While the efficacy of IFN in HBeAg-negative patients was high in the present study, the factors that might be useful in predicting a sustained response were less well-defined than those in HBeAg-positive patients, as previously reported [5].

A meta-analysis of IFN therapy published in 2010 reviewed 6 clinical controlled studies including 828 patients who received IFN [12]. The duration of follow-up
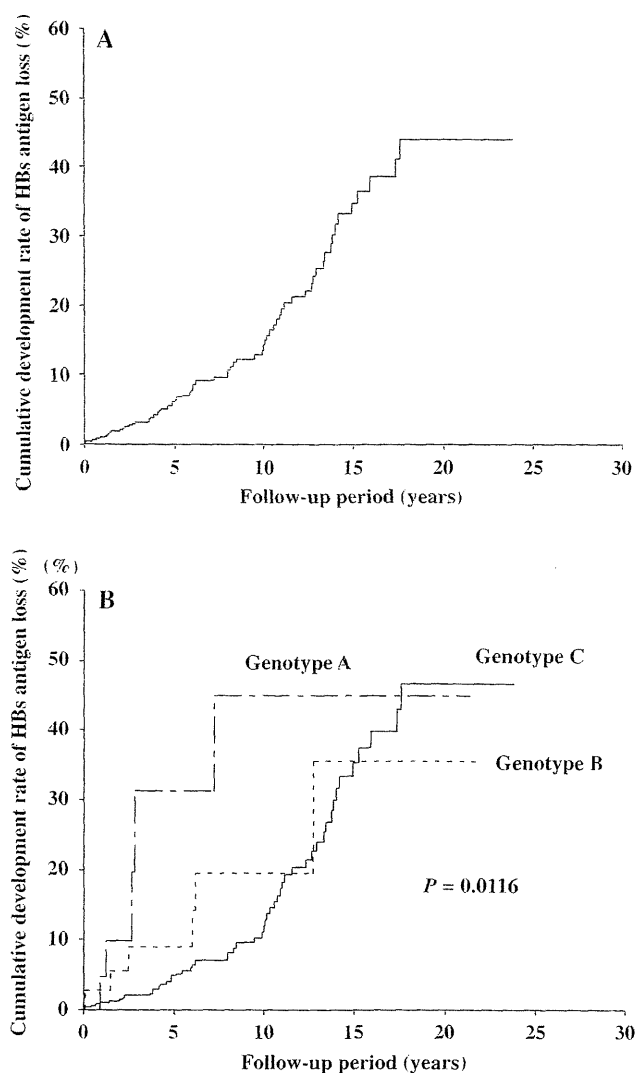
Fig. 3 Cumulative clearance of hepatitis B surface (*HBs*) antigen in patients treated with interferon (Kaplan–Meier method). **a** All patients. **b** patients stratified by genotypes A, B, and C

Table 4 Factors associated with HBsAg seroclearance by interferon therapy. determined by multivariate analysis

Parameter	Category	Hazard ratio	95% CI	P
Age	<30 years	1		0.002
	≥30 years	4.433	1.703–11.538	
Genotype	A	1		0.004
	B	0.296	0.087–1.005	
	C	0.199	0.075–0.528	
Sex	Female	1		0.005
	Male	2.962	1.387–6.327	

HBsAg hepatitis B surface antigen. *CI* confidence interval

ranged from 35.8 months to 7 years, and HBsAg seroclearance occurred in 9.5% (79/828). In the present study, we observed HBsAg seroclearance in 69 of 615 (11%)

patients, with a median follow-up duration of 8.1 years. However. few reports have investigated factors predicting the achievement of HBsAg seroclearance. In our study. important factors for achieving HBsAg seroclearance were age ≥30 years, genotype A, and male sex. Patients with genotype A had primarily been infected during adulthood via sexual contact, and the average duration of infection was relatively short. In contrast, most Japanese carriers are infected perinatally and possess HBV genotype C. and therefore the efficacy of IFN therapy for patients with genotype C may be low. Male sex was also an important factor in determining potential to achieve HBsAg seroclearance. although female sex was an important factor in determining long-term response to IFN therapy. In our previous study of HBsAg seroclearance (mainly spontaneous seroclearance), we found that response rates were low among females (19%; 45/231) [30]. These present and previous findings indicate that male patients tended to achieve HBsAg seroclearance more frequently than females, although the reason is unclear. We previously reported that Kaplan–Meier analysis in 486 patients who received lamivudine therapy for 5 and 10 years showed an estimated loss of HBsAg in 3 and 13% of the patients, respectively, [31]. The cumulative clearance rates of HBsAg, also determined by Kaplan–Meier analysis, in patients treated with IFN were higher than those in the patients treated with lamivudine, albeit that there were differences in the baseline characteristics of the patients at the commencement of the respective therapies. The effects of IFN therapy in modulating the host immune response might induce HBsAg clearance.

In conclusion, we investigated the long-term efficacy of IFN therapy in Japanese CHB patients. Response rates were low among HBeAg-positive patients and relatively high among HBeAg-negative patients at all time points examined. HBeAg-negative status, HBV DNA level, age, sex, pretreatment with IFN, duration of treatment, and levels of albumin and AST were important factors in predicting long-term response for all patients and for HBeAg-positive patients. Age, genotype, and sex were important factors in predicting ability to achieve HBsAg seroclearance. Further studies exploring the efficacy of therapy over a longer duration may be necessary to confirm these findings and establish true response rates to IFN therapy, including treatment with pegylated IFN.

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References

1. Beasley RP, Hwang LW, Lin CC, Chien CS. Hepatocellular carcinoma and hepatitis B virus. A prospective study of 22,707 men in Taiwan. Lancet. 1981;2:1129–233.
2. Dienstag JL, Perrillo RP, Schiff ER, Bartholomew M, Vicary C, Rubin M. A preliminary trial of lamivudine for chronic hepatitis B infection. N Engl J Med. 1995;333:1657–61.
3. Marcellin P, Chang TT, Lim SG, Tong MJ, Sievert W, Shiffman ML, et al. Adefovir dipivoxil for the treatment of hepatitis B e antigen-positive chronic hepatitis B. N Engl J Med. 2003; 348:808–16.
4. Chang TT, Gish RG, de Man R, Gadano A, Sollano J, Chao YC, et al. A comparison of entecavir and lamivudine for HBeAg-positive chronic hepatitis B. N Engl J Med. 2006;354:1001–10.
5. Lok ASF, Heathcote EJ, Hoofnagel JH. Management of hepatitis B: 2000-summary of a workshop. Gastroenterology. 2001;120: 1828–53.
6. Liaw YF, Sung JJ, Chow WC, Farrell G, Lee CZ, Yuen H, et al. Lamivudine for patients with chronic hepatitis B and advanced liver disease. N Engl J Med. 2004;351:1521–31.
7. van Zonneveld M, Honkoop P, Hansen BE, Niesters HG, Darwish Murad S, de Man RA, et al. Long-term follow-up of alpha-interferon treatment of patients with chronic hepatitis B. Hepatology. 2004;39:804–10.
8. Wong DHK, Cheung AM, O'Rourke K, Naylor CD, Detsky AS, Heathcote J. Effect of alpha-interferon treatment in patients with hepatitis B e antigen-positive chronic hepatitis B. Ann Intern Med. 1993;119:312–23.
9. Brook MG, Karayiannis P, Thomas HC. Which patients with chronic hepatitis B virus infection will respond to alpha-interferon therapy? Hepatology. 1989;10:761–3.
10. Perrillo RP, Schiff ER, Davis GL, Bodenheimer HC Jr, Lindsay K, Payne J, et al. A randomized, controlled trial of interferon alfa-2b alone and after prednisolone withdrawal for the treatment of chronic hepatitis B. N Engl J Med. 1990;323:295–301.
11. Perrillo RP, Lai CL, Liaw YF, Dienstag JL, Schiff ER, Schalm SW, et al. Predictors of HBeAg loss after lamivudine treatment for chronic hepatitis B. Hepatology. 2002;36:186–94.
12. Yang YF, Zhao W, Xia HM, Zhong YD, Huang P, Wen J. Long-term efficacy of interferon alpha therapy on hepatitis B viral replication in patients with chronic hepatitis B: a meta-analysis. Antiviral Res. 2010;85:361–5.
13. Suzuki F, Arase Y, Akuta N, Tsubota A, Suzuki Y, Sezaki H, et al. Efficacy of 6-month interferon therapy in chronic hepatitis B virus infection in Japan. J Gastroenterol. 2004;39:969–74.
14. Arase Y, Chayama K, Tsubota A, Murashima N, Suzuki Y, Koida I, et al. A randomized, double-blind, controlled trial of natural interferon-α therapy for e-antigen-negative chronic hepatitis B patients with abnormal transaminase levels. J Gastroenterol. 1996;31:559–64.
15. Arase Y, Tsubota A, Saitoh S, Suzuki Y, Kobayashi M, Suzuki F, et al. Randomized, controlled trial of natural interferon-α therapy for e-antigen-positive chronic hepatitis B patients. Hepatol Res. 2002;23:98–104.
16. Usuda S, Okamoto H, Imawari H, Baba K, Tsuda F, Miyakawa Y, et al. Serological detection of hepatitis B virus genotypes by ELISA with monoclonal antibodies to type-specific epitopes in preS2-region product. J Virol Method. 1999;80:97–112.
17. Tadokoro K, Kobayashi M, Yamaguchi T, Suzuki F, Miyauchi S, Egashira T, et al. Classification of hepatitis B virus genotypes by the PCR-Invader method with genotype-specific probes. J Virol Method. 2006;138:30–9.
18. Kobayashi M, Arase Y, Ikeda K, Tsubota A, Suzuki Y, Saitoh S, et al. Clinical characteristics of patients infected with hepatitis B virus genotypes A, B and C. J Gastroenterol. 2002;37:35–9.
19. Niederau C, Heintges T, Lange S, Goldmann G, Niederau CM, Mohr L, et al. Long-term follow-up of HBeAg-positive patients treated with interferon alfa for chronic hepatitis B. N Engl J M. 1996;334:1422–7.
20. Lin SM, Yu KL, Lee CM, Chien RN, Sheen IS, Chu CM, et al. Interferon therapy in HBeAg positive chronic hepatitis reduces progression to cirrhosis and hepatocellular carcinoma. J Hepatol. 2007;46:45–52.
21. Papatheodoridis GV, Manesis E, Hadziyannis SJ. The long-term outcome of interferon-α treated and untreated patients with HBeAg-negative chronic hepatitis B. J Hepatol. 2001;34:306–13.
22. Brunetto MR, Oliveri F, Coco B, Leandro G, Colombatto P, Gorin JM, et al. Outcome of anti-HBe positive chronic hepatitis B in alpha-interferon treated and untreated patients: a long term cohort study. J Hepatol. 2002;36:263–70.
23. Lampertico P, Ninno ED, Vigano M, Romeo R, Donato MF, Sablon E, et al. Long-term suppression of hepatitis B e antigen-negative chronic hepatitis B by 24-month interferon therapy. Hepatology. 2003;37:756–63.
24. Lau DTY, Everhart J, Kleiner DE, Park Y, Vergalla J, Schmid P, et al. Long-term follow-up of patients with chronic hepatitis B treated with interferon alfa. Gastroenterology. 1997;113:1660–7.
25. Kao JH, Wu NH, Chen PJ, Lai MY, Chen DS. Hepatitis B genotypes and the response to interferon therapy. J Hepatol. 2000;33:998–1002.
26. Hadziyannis S, Bramou T, Makris A, Moussoulis G, Zignego L, Papaioannou C. Interferon alfa-2b treatment of HBeAg negative/serum HBV DNA positive chronic active hepatitis B. J Hepatol. 1990;11:S133–6.
27. Pastore G, Santantonio T, Milella A, Monno L, Mariano N, Moschetta R, et al. Anti-HBe-positive chronic hepatitis B with HBV-DNA in serum: response to a 6-month course of lymphoblastoid interferon. J Hepatol. 1992;20:221–5.
28. Fattovich G, Farci P, Rugge M, Brollo G, Mandas A, Pontisso P, et al. A randomized, controlled trial of lymphoblastoid interferon-alfa in patients with chronic hepatitis B lacking HBeAg. Hepatology. 1992;15:584–9.
29. Lampertico P, Del Ninno E, Manzin A, Donato MF, Rumi MG, Lunghi G, et al. A randomized, controlled trial of a 24-month course of interferon alfa 2b in patients with chronic hepatitis B who had hepatitis B virus DNA without hepatitis B e antigen in serum. Hepatology. 1997;26:1621–5.
30. Arase Y, Ikeda K, Suzuki F, Suzuki Y, Saitoh S, Kobayashi M, et al. Long-term outcome after hepatitis B surface antigen seroclearance in patients with chronic hepatitis B. Am J Med. 2006;119:71 e9–e16.
31. Kobayashi M, Suzuki F, Akuta N, Hosaka T, Sezaki H, Yatsuji H et al. Loss of hepatitis B surface antigen from the serum o patients with chronic hepatitis treated with lamivudine. J Med Virol. 2007;79:1472–7.
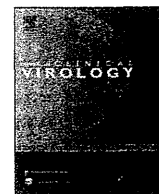
Short Communication

Prevalence of hepatitis C virus variants resistant to NS3 protease inhibitors or the NS5A inhibitor (BMS-790052) in hepatitis patients with genotype 1b

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ABSTRACT

Background: Hepatitis C virus (HCV) of genotype 1b is the most prevalent worldwide, and the least responsive to interferon-based treatments. A combination therapy with two direct-acting antivirals has shown promising results in patients with HCV-1b, but the prevalence of drug-resistant variants before treatment is not known in the Japanese population.
Objectives: To detect HCV variants resistant to NS3 protease inhibitors or the NS5A inhibitor (BMS-790052) in hepatitis patients infected with HCV-1b.
Study design: Drug-resistant mutations were determined in the 362 hepatitis patients infected with HCV-1b who had not received direct-acting antivirals before.
Results: Amino-acid substitutions resistant to NS3 inhibitors (V36A, T54S, Q80H and D168E) were detected in 15 of the 307 (4.9%) patients, who had been examined, and T54S (3.3%) predominated over V36A (0.3%), Q80R (0.7%) and D168E (0.7%) in them. Amino-acid substitutions resistant to BMS-790052 (L31M and/or Y93H) were detected in 33 of the 294 (11.2%) patients, and Y93H (8.2%) predominated over L31M (2.7%). One of the 239 (0.4%) patients, who had been examined for amino-acid substitutions in both NS3 and NS5A regions, possessed HCV-1b variants resistant to NS3 inhibitors (T54S) and BMS-790052 (L31M).
Conclusions: Mutations conferring resistance to NS3 inhibitors or BMS-790052 were frequent in our treatment-naive study population, but double mutants with possible resistance to both drugs were rare. Since single mutations did not result in treatment failure in a previous pilot trial combining BMS-790052 and an NS3 inhibitor, larger trials of this drug regimen appear warranted in the Japanese population.

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1. Background

Worldwide, an estimated 170 million people are infected with hepatitis C virus (HCV) persistently,[1] and approximately one-third of them develop life-threatening liver diseases, such as decompensated cirrhosis and hepatocellular carcinoma.[2] The triple therapy with an NS3 protease inhibitor, telaprevir or boceprevir, in combination with pegylated (PEG)-interferon (IFN) and ribavirin (RBV), has increased sustained virological response (SVR) to about 70% in the patients with HCV of genotype 1b (HCV-1b).[3-7] Still, approximately 30% of them fail to clear HCV by the triple therapy, and, in addition, many more cannot receive it because of contraindications, such as advanced ages, anaemia and co-morbid conditions.

Recently, a combination therapy with two direct-acting antivirals (DAAs), which are targeted to different regions in the viral genome, was introduced to treatment of patients with HCV-1b, and has gained promising results. Thus, a second-generation NS3 protease inhibitor (BMS-650032 [asunaprevir]) combined with an NS5A inhibitor (BMS-790052 [daclatasvir]) for 24 weeks induced SVR in two of the two,[8] as well as in 10 of the 10,[9] patients with HCV-1b with excellent safety profiles.

2. Objectives

For extending the combination treatment with BMS-650032 and BMS-790052 to many more patients with HCV-1b, it is necessary to examine how frequently viral variants, which have resistance to NS3 protease inhibitors or NS5A inhibitors,[10-13] occur in patients with HCV-1b.

3. Study design

3.1. Patients

During 2000 through 2010, sera were obtained from the 362 patients with HCV-1b at the Department of Hepatology in Tora-nomon Hospital in Tokyo, and had been stored frozen at −80 °C. They all were treatment-naive to NS3 protease inhibitors or the NS5A inhibitor (BMS-790052); 134 of them (37.0%) had received IFN-based treatments previously. The nucleotide sequence of the NS3 region in HCV RNA was determined in 307 patients and that of NS5A region in 294, and sequences of both NS3 and NS5A were determined in 239.

3.2. Sequencing NS3 and NS5A regions

HCV RNA was amplified by polymerase chain reaction with appropriate nested primers in NS3[14] or NS5A[15] region, and sequences of the N-terminal 609 nucleotides in the NS3 region and those of the N-terminal 600 nucleotides in the NS5A region were determined by the direct sequencing method. The major sequences were adopted, which would represent the consensus sequence. They have been deposited in the Genbank under the accession numbers AB693834–AB693872 and AB709241–AB709802.

3.3. Amino-acid substitutions for the resistance to NS3 protease inhibitors or the NS5A inhibitor (BMS-790052)

V36A/M/L/G, T54A/S, V55A, Q80K/R/H/G, R155K/T/I/M/G/L/S/Q, A156V/T/S/I/G, D168A/V/E/G/N/T/Y/H/I and V170A have been identified as amino-acid substitutions resistant to NS3 protease inhibitors, including linear ketoamids (telaprevir, boceprevir, SCH900518 and BI201335) and macrocyclic compounds (MK7009, TMC435350, ITMN191, GS-9256, ABT450 and BMS-791325).[16,17] L31M and Y93H have been recognised as the most powerful substitutions in HCV-1b for the resistance to BMS-790052.[18-21]

4. Results

4.1. Baseline characteristics of patients with HCV-1b who were naive to DAAs

Table 1 lists the baseline characteristics of the 362 patients infected with HCV-1b. Of them, 134 (37.0%) had received IFN-based treatments previously, including 78 (21.6%) with IFN monotherapy and 56 (15.4%) given combination therapy with IFN or PEG-IFN and RBV. Liver biopsies had been performed on 201 of the 362 (55.5%) patients. The majority of them (47.5%) had fibrosis stages ≤F2, by the classification of Desmet et al.,[22] and none had cirrhosis.

4.2. Amino-acid substitutions for the resistance to NS3 inhibitors or the NS5 inhibitor (BMS-790052)

Table 2 shows frequencies of amino-acid substitutions for the resistance to NS3 inhibitors in 307 patients. Of them, 15 (4.9%) were infected with HCV-1b variants having V36A, T54S, Q80R or D168E, and T54S predominated over Q80R, V36A and D168E. Resistance

Table 1
Baseline characteristic of the patients infected with HCV of genotype 1b who were naive to direct-acting antivirals.

Demographic data	(n = 362)
Male (%)	213 (58.8%)
Age (years)	55 (18–75)
IFN-based treatments	
Treatment-naive	228 (63.0%)
IFN monotherapy	78 (21.6%)
IFN (or PEG-IFN) plus ribavirin	56 (15.4%)
Laboratory data	
Alanine aminotransferase (IU/L)	54 (12–348)
Asparatate aninotransferase (IU/L)	41 (17–350)
Platelets (×10³/mm³/μL)	174 (64–366)
HCV RNA (log IU/mL)	6.7 (<1.2 to >7.6)
Stage of liver fibrosis[a]	(n = 201)
F1	117 (58.2%)
F2	55 (27.4%)
F3	29 (14.4%)
F4	0

Values are the number with percentage in parentheses or the mean with range in parentheses.
[a] Classified by the criteria of Desmet et al.[22]

Table 2
Substitutions of amino acids in the NS3 protease region for the resistance to NS3 inhibitors in Japanese patients in the present study and in European or American patients with HCV-1b retrieved from the Genbank.

Substitutions	This study (n = 307) n (%)	Database[a] (n = 400) n (%)
V36A	1 (0.3%)	1 (0.3%)
T54A	0	1 (0.3%)
T54S	10 (3.3%)	5 (1.2%)
V55A	0	1 (0.3%)
Q80R	2 (0.7%)	16 (4.0%)
A156T	0	1 (0.3%)
D168E	2 (0.7%)	2 (0.5%)
V170A	0	2 (0.5%)
Total	15 (4.9%)	29 (7.3%)

[a] HCV-1b sequences were retrieved from the Genbank. There were 400 sequences in total, exclusive of repetitive sequences, including 307 from France, 53 from Spain, 6 from Germany and 34 from USA.

profiles are comparable between Japanese patients in this study and 366 European and 34 American patients (total: 400 patients) retrieved from the Genbank.

Table 3 shows frequencies of amino-acid substitutions for the resistance to the NS5 inhibitor (BMS-790052) in the 294 patients. Y93H predominated over L31M, and one patient had both Y93H and L31M. Overall, 33 (11.2%) of them were infected with HCV-1b variants with L31M or Y93H, or both. One of the 239 (0.4%) patients, for whom both NS3 and NS5A sequences had been examined, was infected with HCV-1b variants with resistance to NS3 inhibitors (T54S) and NS5A inhibitor (L31M).

Table 3
Substitutions of amino acids in the NS5A region for the resistance to BMS-790052 in Japanese patients in the present study and in patients with HCV-1b retrieved from the European HCV database.

Substitutions	This study (n = 294) n (%)	Database[a] (n = 1796) n (%)
L31M	8 (2.7%)	68 (3.8%)
L31V	0	38 (2.1%)
Y93H	24 (8.2%)	149 (8.3%)
Y93H/L31M	1 (0.3%)	Unknown
Total	33 (11.2%)	255 (14.2%)

[a] The sequences of HCV-1b were retrieved from the European HCV database and reported by Fridell et al.[18]

Factors influencing HCV-1b variants resistant to NS3 inhibitors or BMS-790052 were evaluated by univariate analysis with use of the Statistical Package for Social Sciences (SPSSII v.11.0, IBM Co., Chicago, IL, USA). None of age, sex, transaminase levels, platelet counts, HCV RNA loads and histological stages increased the prevalence of HCV-1b variants resistant to either of these two kinds of DAAs.

5. Discussion

DAAs have different antiviral targets and distinct resistance profiles that are dependent on HCV genotypes/subtypes.[16,21,23] For treatment of patients with HCV-1b, a combination of a second-generation NS3 protease inhibitor (BMS-650032) and an NS5A inhibitor (BMS-790052) has gained SVR in two of the two, as well as 10 of the 10, patients with HCV-1b.[8,9] By contrast, the combination therapy was less effective in the nine patients with HCV-1a, and viral breakthroughs occurred in six (67%) of them.[8] In HCV-1a, only one nucleotide mutation gives rise to amino-acid substitutions resistant to NS3 protease inhibitors (R155K/T/S/M/I), instead of two required in HCV-1b,[23] which would be responsible, at least in part, for poor responses to the combination therapy in patients with HCV-1a.

There is a possibility that HCV-1b variants resistant to both BMS-650032 and BMS-790052 may be selected during the combination therapy, and result in viral breakthroughs during treatment. Of the 307 patients, who had been examined, 15 (4.9%) were infected with HCV-1b with amino-acid substitutions for the resistance to NS3 protease inhibitors. Of the NS3 resistance mutations detected, only D168E is relevant to the second-generation protease inhibitors,[16,17] and, therefore, only 0.7% of the treatment-naive patients carried relevant resistance mutations when focussing on a possible combination of BMS-650032 with other DAAs. It needs to be pointed out that a possibility remains for the presence of minor HCV populations with resistance to DAAs that might have escaped the detection by direct sequencing.

HCV-1b variants with L31M or Y93H, which confers strong resistance to the NS5A inhibitor (BMS-790052),[20] were detected in 33 of the 294 (11.2%) patients with HCV-1b; one of them was infected with variants with both L31M and Y93H. Such a frequency is comparable to those in 1796 patients from the European HCV database (L31M, 5.9%; Y93C/H, 8.4%).[18] Variants with Y93H were detected in 3 of the 10 (30%) patients receiving the combination therapy with BMS-650032 and BMS-790052.[9] Since they all gained SVR, variants with Y93H alone, in the absence of those resistant to macrocyclic NS3 protease inhibitors, would not cause treatment failure in the patients who receive the combination therapy. Co-occurrence of variants resistant to NS3 protease inhibitors and those to the NS5A inhibitor was observed in only one of the 239 (0.4%) patients for whom both of them were examined. They may or may not exist on the same virion, because they were detected by direct sequencing. Therefore, results suggest that most patients with HCV-1b in our geographic area can be good candidates to succeed in resolving infection after combination therapy with NS3 inhibitors and BMS-790052.

Competing interest: Dr. Kumada reports having received investigator, lecture and consulting fees from Bristol-Myers KK. No other potential conflicts of interest relevant to this article were reported.

Ethical approval: Informed consent was obtained from each patient.

References

1. http://www.who.int/mediacentre/factsheets/fs164/en/. Accessed August 2011.
2. Seeff LB. Natural history of chronic hepatitis C. *Hepatology* 2002;**36**: S35–46.
3. Hezode C, Forestier N, Dusheiko G, Ferenci P, Pol S, Goeser T, et al. Telaprevir and peginterferon with or without ribavirin for chronic HCV infection. *N Engl J Med* 2009;**360**:1839–50.
4. Kumada H, Toyota J, Okanoue T, Chayama K, Tsubouchi H, Hayashi N. Telaprevir with peginterferon and ribavirin for treatment-naive patients chronically infected with HCV of genotype 1 in Japan. *J Hepatol* 2012;**56**: 78–84.
5. McHutchison JG, Everson GT, Gordon SC, Jacobson IM, Sulkowski M, Kauffman R, et al. Telaprevir with peginterferon and ribavirin for chronic HCV genotype 1 infection. *N Engl J Med* 2009;**360**:1827–38.
6. Bacon BR, Gordon SC, Lawitz E, Marcellin P, Vierling JM, Zeuzem S, et al. Boceprevir for previously treated chronic HCV genotype 1 infection. *N Engl J Med* 2011;**364**:1207–17.
7. Poordad F, McCone Jr J, Bacon BR, Bruno S, Manns MP, Sulkowski MS, et al. Boceprevir for untreated chronic HCV genotype 1 infection. *N Engl J Med* 2011;**364**:1195–206.
8. Lok AS, Gardiner DF, Lawitz E, Martorell C, Everson GT, Ghalib R, et al. Preliminary study of two antiviral agents for hepatitis C genotype 1. *N Engl J Med* 2012;**366**:216–24.
9. Chayama K, Takahashi S, Toyota J, Karino T, Ikeda K, Ishikawa H, et al. Dual therapy with the NS5A inhibitor BMS-790052 and the NS3 protease inhibitor BMS-650032 in HCV genotype 1b-infected null responders. *Hepatology* 2012;**55**:742–8.
10. Bartels DJ, Zhou Y, Zhang EZ, Marcial M, Byrn RA, Pfeiffer T, et al. Natural prevalence of hepatitis C virus variants with decreased sensitivity to NS3.4A protease inhibitors in treatment-naive subjects. *J Infect Dis* 2008;**198**: 800–7.
11. Cubero M, Esteban JI, Otero T, Sauleda S, Bes M, Esteban R, et al. Naturally occurring NS3-protease-inhibitor resistant mutant A156T in the liver of an untreated chronic hepatitis C patient. *Virology* 2008;**370**:237–45.
12. Gaudieri S, Rauch A, Pfafferott K, Barnes E, Cheng W, McCaughan G, et al. Hepatitis C virus drug resistance and immune-driven adaptations: relevance to new antiviral therapy. *Hepatology* 2009;**49**:1069–82.
13. Kuntzen T, Timm J, Berical A, Lennon N, Berlin AM, Young SK, et al. Naturally occurring dominant resistance mutations to hepatitis C virus protease and polymerase inhibitors in treatment-naive patients. *Hepatology* 2008;**48**: 1769–78.
14. Suzuki F, Suzuki Y, Akuta N, Sezaki H, Yatsuji H, Arase Y, et al. Sustained virological response in a patient with chronic hepatitis C treated by monotherapy with the NS3-4A protease inhibitor telaprevir. *J Clin Virol* 2010;**47**: 76–8.
15. El-Shamy A, Nagano-Fujii M, Sasase N, Imoto S, Kim SR, Hotta H. Sequence variation in hepatitis C virus nonstructural protein 5A predicts clinical outcome of pegylated interferon/ribavirin combination therapy. *Hepatology* 2008;**48**:38–47.
16. Halfon P, Locarnini S. Hepatitis C virus resistance to protease inhibitors. *J Hepatol* 2011;**55**:192–206.
17. Romano KP, Ali A, Royer WE, Schiffer CA. Drug resistance against HCV NS3/4A inhibitors is defined by the balance of substrate recognition versus inhibitor binding. *Proc Natl Acad Sci U S A* 2010;**107**:20986–91.
18. Fridell RA, Qiu D, Wang C, Valera L, Gao M. Resistance analysis of the hepatitis C virus NS5A inhibitor BMS-790052 in an in vitro replicon system. *Antimicrob Agents Chemother* 2010;**54**:3641–50.
19. Fridell RA, Wang C, Sun JH, O'Boyle 2nd DR, Nower P, Valera L, et al. Genotypic and phenotypic analysis of variants resistant to hepatitis C virus nonstructural protein 5A replication complex inhibitor BMS-790052 in humans: in vitro and in vivo correlations. *Hepatology* 2011;**54**:1924–35.
20. Gao M, Nettles RE, Belema M, Snyder LB, Nguyen VN, Fridell RA, et al. Chemical genetics strategy identifies an HCV NS5A inhibitor with a potent clinical effect. *Nature* 2010;**465**:96–100.
21. Scheel TK, Gottwein JM, Mikkelsen LS, Jensen TB, Bukh J. Recombinant HCV variants with NS5A from genotypes 1-7 have different sensitivities to an NS5A inhibitor but not interferon-alpha. *Gastroenterology* 2011;**140**: 1032–42.
22. Desmet VJ, Gerber M, Hoofnagle JH, Manns M, Scheuer PJ. Classification of chronic hepatitis: diagnosis, grading and staging. *Hepatology* 1994;**19**: 1513–20.
23. Sarrazin C, Zeuzem S. Resistance to direct antiviral agents in patients with hepatitis C virus infection. *Gastroenterology* 2010;**138**:447–62.

Research Article

Long-term continuous entecavir therapy in nucleos(t)ide-naïve chronic hepatitis B patients

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Background & Aims: We determined the antiviral potency and viral resistance rate after 4 years of continuous entecavir treatment in patients with chronic hepatitis B (CHB) infection.
Methods: The cumulative rates of undetectable hepatitis B virus DNA (HBV DNA; $<2.6 \log_{10}$ copies/ml), hepatitis B e antigen (HBeAg) seronegativity, seroconversion, alanine aminotransferase (ALT) normalization, and entecavir signature mutations were calculated in 474 nucleos(t)ide-naïve CHB patients (HBeAg-positive: 47%) on continuous entecavir treatment for 4 years.
Results: Median age was 47 years and follow-up period was 2.4 years, with 403, 281, 165, and 73 patients followed-up for at least 1, 2, 3, and 4 years, respectively. Incremental increases were observed in the rates of undetectable HBV DNA, HBeAg seroclearance and seroconversion, and ALT normalization, reaching 96%, 42%, 38% and 93%, respectively, by the fourth year. In all, 100% and 93% of patients negative and positive for HBeAg, respectively, had undetectable HBV DNA at year 4. Of 165 patients, HBV DNA was detectable in nine patients after 3 years. Multivariate analysis identified HBV DNA level ($\leqslant 7.6 \log_{10}$ copies/ml, OR = 15.8; 95% CI = 43.1–79.9, P = 0.001) as an independent predictor of undetectable HBV DNA at year 3. Five patients experienced virological breakthrough including two (0.4%) who developed entecavir-resistance mutations.
Conclusions: Continuous treatment of nucleos(t)ide-naïve CHB patients with entecavir over 4 years was associated with 96% chance of undetectable HBV DNA and only 0.4% chance of emerging entecavir-resistant mutations.

Introduction

Approximately 350–400 million people worldwide have chronic hepatitis B (CHB) infection, the majority of whom live in the Asia–Pacific region [1,2]. CHB patients with elevated viral load are at risk of developing cirrhosis, liver failure, and hepatocellular carcinoma.

Recent investigations have shown that entecavir suppressed HBV DNA replication to undetectable levels and normalized alanine aminotransferase (ALT) levels in nucleos(t)ide-naive CHB patients in Japan and other countries [3–10]. In addition, genotypic resistance to long-term entecavir monotherapy remained rare [5,6,9,10]. To date, there are two 5-year studies [6,8] and two 3-year studies [7,9] of entecavir therapy for nucleos(t)ide-naïve patients. Both studies stemmed from extension studies with the original cohorts from two large-scale phase III trials of treatment-naïve patients [3,4]. In these trials, patients were administered 0.5 mg entecavir for 1 year and later divided into three categories: (i) complete responders, defined as patients with HBV DNA $<7 \times 10^5$ copies/ml and ALT level <1.25 times the upper limit of normal (ULN) for hepatitis B e antigen (HBeAg)-negative patients and an additional loss of HBeAg for HBeAg-positive patients; (ii) non-responders, defined as HBV DNA $\geqslant 7 \times 10^5$ copies/ml; and, (iii) virological responders, defined as HBV DNA $<7 \times 10^5$ copies/ml and ALT >1.25× ULN regardless of HBeAg status or persistent HBeAg for HBeAg-positive patients. Treatment was terminated in the complete responders but continued in virological responders. Non-responders were provided additional therapy in a rollover study in which some patients were initially treated with a combination of 1 mg entecavir and lamivudine for several months before receiving 1 mg entecavir as monotherapy. Furthermore, a substantial proportion of complete responders relapsed after various intervals following cessation of therapy and they were also assigned to a rollover study receiving 1 mg entecavir monotherapy. Because of these strict protocols, the precise viral-suppression and drug-resistance data for treatment-naïve patients who were treated continuously with 0.5 mg entecavir daily (the recommended dosage) remain unavailable.

The aims of this cohort study were (1) to investigate the efficacy of entecavir in clinical practice beyond 4 years for nucleos(t)ide-naïve CHB and cirrhosis patients, (2) to explore baseline factors associated with virological response to entecavir,

and (3) to investigate virological breakthrough during long-term entecavir treatment.

Patients and methods

Study population

We performed a retrospective analysis of 474 CHB and cirrhosis patients who received entecavir treatment at the Department of Hepatology, Toranomon Hospital, Tokyo, from March 2004 to May 2011, and adhered to the treatment for more than 6 months (Table 1). All patients were negative for hepatitis C serological markers, but all had detectable HBV surface antigen (HBsAg) for at least 6 months prior to the start of entecavir therapy. Two patients received 0.01 mg entecavir and one patient received 0.1 mg entecavir for 24 weeks, prior to 0.5 mg/day from a phase II study ETV-047 in Japan [11]. The other patients received 0.5 mg entecavir. None had received other nucleos(t)ide analogs. The diagnosis of chronic hepatitis and cirrhosis was established by needle biopsy, peritoneoscopy, or clinically before treatment. The clinical criteria for chronic hepatitis included elevated ALT levels over 6 months and absence of clinical evidence of portal hypertension, such as esophageal varices, ascites, hepatic encephalopathy, and features suggestive of cirrhosis on ultrasonography. Chronic hepatitis and cirrhosis were diagnosed in 374 and 102 patients, respectively. Twenty-eight patients were lost to follow-up, including 10 patients who moved to other locations, seven who never visited the hospital again, two who became pregnant, four who died, four who had virological breakthrough (VBT), and one who showed disappearance of HBsAg. Moreover, 18 patients developed HCC during treatment and their data until loss to follow-up or diagnosis of HCC were analyzed. Informed consent was obtained from each patient enrolled in the study and the study protocol conformed to the ethical guidelines of the Declaration of Helsinki and was approved by the Toranomon Hospital Ethical Committee.

Analysis of treatment efficacy

The clinical efficacy of entecavir was assessed as the proportion of patients who achieved HBV DNA suppression to undetectable levels (<2.6 $\log_{10}$ copies/ml), and those who achieved ALT normalization (<1 × ULN). HBV DNA was measured using

the polymerase chain reaction (PCR)-based Amplicor HBV Monitor assay (Roche Diagnostics, Indianapolis, IN, lower limit of detection of 2.6 $\log_{10}$ copies/ml) [12]. HBeAg seroclearance and seroconversion were also analyzed. Measurements were made on stored samples taken at baseline and every year after that since entecavir treatment initiation.

Statistical analysis

Differences between groups were examined for statistical significance using the χ^2 test where appropriate. Spearman correlation coefficient (two-tailed) was used to evaluate the correlation between albumin and other factors. Independent predictive factors associated with response to entecavir treatment were determined using multivariate multiple logistic regression. The following 12 potential predictors of response to entecavir treatment were assessed in this study: age, sex, severity of liver disease (CH or cirrhosis), HBV genotype, as well as levels of aspartate transaminase (AST), ALT, bilirubin, albumin, platelets, α fetoprotein (AFP), HBeAg, and HBV DNA. All factors found to be at least marginally associated with undetectable levels of HBV DNA after 1-4 years (p <0.10) were entered into the multivariate multiple logistic regression analysis. The above calculations were performed using The Statistical Package for Social Sciences version 11.0.1J (SPSS Inc., Chicago, IL).

The odds ratios (ORs) and 95% confidence intervals (CIs) were calculated to assess the relative risk confidence. Independent risk factors predicting achievement of HBeAg seroclearance and seroconversion were analyzed using stepwise Cox regression analysis. Potential factors that could predict achievement of HBeAg seroclearance assessed here were the above 11 variables, each transformed into categorical data consisting of two simple ordinal numbers for univariate and multivariate analyses. All factors found to be at least marginally associated with HBeAg seroclearance and seroconversion (p <0.10) were tested in the multivariate Cox proportional hazard model. A Kaplan-Meier estimate was performed using the SPSS software, and p values were calculated using the Cox-Mantel log-rank test. The Mann-Whitney U test was used for comparison of HBV DNA levels in patients with seroconversion to those with seroclearance. A two-tailed p value <0.05 was considered statistically significant.

Table 1. Characteristics of patients at the start of entecavir therapy. Table data are number of patients or median (range).

Demography	
n	474
Sex, male/female	321/153
Age, yr	47 (17-82)
Family history of HBV	291 (61%)
Cirrhosis	102 (22%)
Median duration of treatment, yr (range)	2.37 (0.5-7.2)
Laboratory data	
AST, IU/L	52 (14-1595)
ALT, IU/L	70 (8-2121)
Bilirubin, mg/dl	0.7 (0.2-3.9)
γ-GTP, IU/L	38 (9-679)
Albumin, g/dl	3.9 (1.9-5.1)
Alpha fetoprotein, ng/ml	5 (1-379)
Viral load, $\log_{10}$ copies/ml	6.7 (<2.6->9.0)
HBeAg-positive	222 (47%)
HBV genotypes, A/B/C/H/unknown	12/67/336/2/57

A

n = 402 184 217 281 132 148 165 84 80 73 41 32
Unknown 1   1   1

Total
HBeAg+
HBeAg-

B

n = 9 56 288 8 39 217 4 17 132 3 4 65

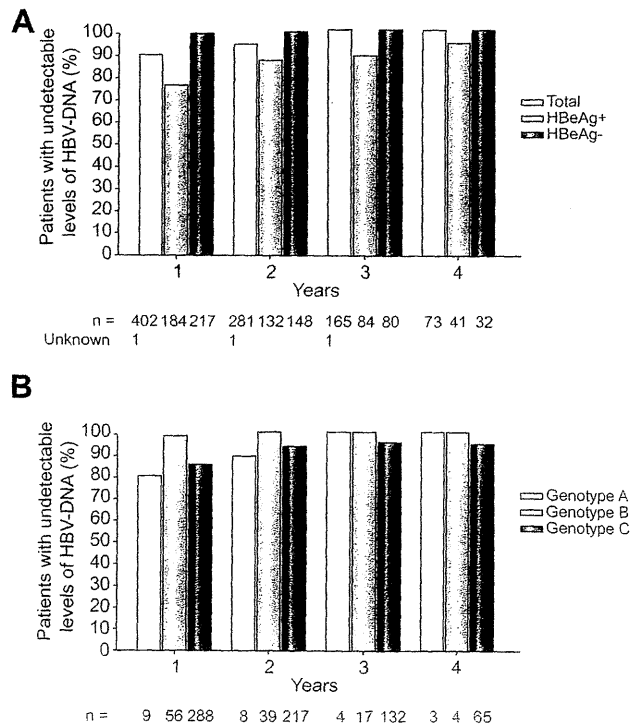Genotype A
Genotype B
Genotype C

Fig. 1. **Percentages of patients who had undetectable levels of HBV DNA between years 1 through 4.** (A) HBeAg-positive and negative patients and (B) patients with genotype A, B, or C.

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Table 2. Univariate and multivariate analyses of host and viral factors associated with undetectable levels of HBV DNA at year 1.

Parameter	Univariate analysis		Multivariate analysis	
	OR (95% CI)	p value	OR (95% CI)	p value
Sex (female)	1.06 (0.56-2.02)	0.842		
Age (>40 yr)	1.85 (1.0-3.4)	0.047		
Cirrhosis (present)	2.39 (0.98-5.81)	0.048		
Albumin (>4 g/dl)	1.28 (0.63-2.62)	0.494		
Bilirubin (>1.2 g/dl)	1.63 (0.56-4.76)	0.366		
ALT (>5 x IU/L)	4.57 (1.38-15.1)	0.007	11.9 (3.3-41.7)	<0.001
AST (>5 x IU/L)	2.25 (0.67-7.53)	0.178		
γ-GTP (≤20 IU/L)	1.75 (0.60-5.08)	0.300		
AFP (>10 ng/ml)	1.63 (0.61-4.37)	0.328		
Platelets (≤10/mm^3)	2.39 (0.56-10.3)	0.288		
Genotype (B)	9.57 (1.29-70.92)	0.007		
HBeAg (negative)	23.78 (7.25-77.95)	<0.001	8.5 (2.3-31.2)	0.001
HBV DNA (≤7.6 $\log_{10}$ copies/ml)	16.5 (8.0-34.2)	<0.001	10.0 (4.3-23.1)	<0.001

Table 3. Univariate and multivariate analyses of host and viral factors associated with undetectable levels of HBV DNA at year 2.

Parameter	Univariate analysis		Multivariate analysis	
	OR (95% CI)	p value	OR (95% CI)	p value
Sex (male)	0.524 (0.169-1.627)	0.257		
Age (>40 yr)	2.825 (1.1-7.25)	0.025		
Cirrhosis (present)	3.06 (0.69-13.5)	0.173		
Albumin (≤3.5 g/dl)	4.64 (0.603-35.73)	0.134		
Bilirubin (≤0.5 g/dl)	2.80 (0.79-9.93)	0.126		
ALT (>5 x IU/L)	5.35 (0.7-40.9)	0.054	16.7 (2.0-136.8)	0.009
AST (>5 x IU/L)	2.62 (0.34-20.3)	0.298		
γ-GTP (≤100 IU/L)	1.79 (0.557-5.73)	0.304		
AFP (>15 ng/ml)	2.12 (0.27-16.95)	0.699		
Platelets (≤12/mm^3)	4.74 (0.619-36.31)	0.136		
Genotype (B)	1.082 (1.042-1.123)	0.076		
HBeAg (negative)	23.21 (3.05-176.46)	<0.001		
HBV DNA (≤7.6 $\log_{10}$ copies/ml)	39.91 (8.912-178.76)	<0.001	121.7 (15.3-965.9)	<0.001

Results

Study population

Of the 474 subjects in this study, 68% were males, and the mean age was 47 years. The mean HBV DNA level was $6.7 \log_{10}$ copies/ml, mean ALT level was 70 IU/L, and 47% of patients were HBeAg-positive. At baseline, there were 12, 67, and 336 patients of genotype A, B, and C, respectively, and among the patients belonging to these genotypes, 4, 11, and 188, respectively, were HBeAg-positive.

Virological response

Undetectable levels of HBV DNA were identified at years 1 through 4 in 88% (353/402), 93% (262/281), 95% (156/165), and

96% (70/73) of patients, respectively (Fig. 1A). Among the HBeAg-positive patients at baseline, 75% (138/184), 86% (114/132), 89% (75/84), and 93% (38/41), and among the HBeAg-negative patients at baseline, 99% (214/217), 99% (147/148), 100% (80/80), and 100% (32/32) had undetectable levels of HBV DNA at years 1 through 4, respectively.

Among the patients with genotype A, 78% (7/9), 88% (7/8), 100% (4/4), and 100% (3/3) of patients had undetectable levels of HBV DNA at years 1 through 4, respectively (Fig. 1B). Among the HBeAg-positive patients with genotype A at baseline, 50% (2/4), 67% (2/3), 100% (2/2), and 100% (2/2) had undetectable levels of HBV DNA at years 1 through 4, respectively. Among patients with genotype B, 98% (55/56), 100% (39/39), 100% (17/17), and 100% (4/4) had undetectable levels at years 1 through 4, respectively (Fig. 1B). Among the HBeAg-positive patients with genotype B at baseline, 88% (7/8), 100% (5/5), and 100% (3/3) had
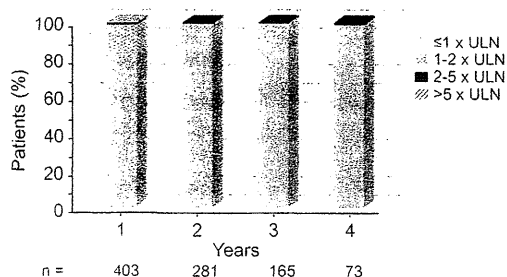
Fig. 2. Percentages of patients with ALT level <1× upper limit of normal level (ULN), <1–2× ULN, 2–5× ULN, and 5× ULN.

undetectable levels of HBV DNA at years 1 through 3, respectively. None of the patients with genotype B at baseline were HBeAg-positive at year 4. Among the patients with genotype C, 85% (246/288), 93% (201/217), 95% (125/132), and 95% (62/65) had undetectable levels of HBV DNA at years 1 through 4, respectively (Fig. 1B), and of these, 74% (116/156), 87% (102/117), 91% (67/74), and 92% (35/38), respectively, were HBeAg-positive.

Factors associated with detectable levels of HBV DNA at years 1, 2, and 3

Of the 402 patients, 353 had no detectable HBV DNA after 1 year. At the start of treatment, factors associated with undetectable levels of HBV DNA in the first year were age (>40 years, $p = 0.047$), cirrhosis (present, $p = 0.048$), ALT (>5× ULN, $p = 0.007$), genotype (B, $p = 0.007$), HBeAg (negative, $p < 0.001$), and HBV DNA level (<7.6 $\log_{10}$ copies/ml, $p < 0.001$), by univariate analysis (Table 2). Multivariate analysis identified three parame-

ters, namely ALT (>5× ULN, OR = 11.9; 95% CI = 3.3–41.7, $p < 0.001$), HBeAg (negative, OR = 8.5; 95% CI = 2.3–31.2, $p = 0.001$), and HBV DNA level (<7.6 $\log_{10}$ copies/ml, OR = 10.0; 95% CI = 4.3–23.1, $p < 0.001$).

Of 281 patients, HBV DNA was undetectable in 262 patients in the second year, with univariate analysis identifying the following associated factors: age (>40 years, $p = 0.025$), ALT (>5× ULN, $p = 0.054$), HBeAg (negative, $p < 0.001$), and HBV DNA level (≤7.6 log copies/ml, $p < 0.001$). Of these, multivariate analysis identified ALT (>5× ULN, OR = 16.7; 95% CI = 2.0–136.8, $p = 0.009$) and HBV DNA level (≤7.6 $\log_{10}$ copies/ml, OR = 121.7; 95% CI = 15.3–965.9, $p < 0.001$) as significant factors (Table 3).

Of 165 patients, HBV DNA was undetectable in 156 patients in the third year, with univariate analysis identifying the following associated factors at the start of treatment: Gender (male, $p = 0.04$), HBeAg (negative, $p = 0.002$) and HBV DNA level (≤7.6 log copies/ml, $p < 0.001$). Multivariate analysis identified only HBV DNA level as significant (≤7.6 $\log_{10}$ copies/ml, OR = 15.8; 95% CI = 43.1–79.9, $p = 0.001$).

Biochemical response

The percentages of patients with normal ALT levels (<1x ULN) at years 1, 2, 3, 4 were 83% (336/403), 89% (251/281), 92% (151/165), and 93% (68/73), respectively (Fig. 2). In HBeAg-positive patients at baseline, those who achieved normal ALT levels at years 1, 2, 3, 4 were 81% (148/183), 88% (116/132), 90% (76/84), and 95% (39/41), respectively. The respective data for HBeAg-negative patients at baseline were 85% (187/219), 91% (134/148), 93% (74/80), and 91% (29/32).

HBeAg seroclearance and seroconversion

HBeAg positivity at baseline was detected in 222 patients (47%) (Table 1), and Fig. 3A shows the cumulative clearance of HBeAg calculated with the Kaplan–Meier method. The percentages of patients with seroclearance were 16%, 24%, 37%, and 42% at years 1 through 4, respectively. Univariate analysis identified the following HBeAg seroclearance-associated factors at the start of treatment: age (>40 years, $p = 0.052$), platelet count (<12 × 10⁴/mm³, $p = 0.028$), and HBV DNA (<7.0 log copies/ml, $p = 0.006$). Multivariate analysis identified HBV DNA (<7 $\log_{10}$ copies/ml, RR = 1.9; 95% CI = 1.2–3.1, $p = 0.007$) as the only significant determinant of seroclearance. Of 70 patients who achieved anti-HBe seroclearance, 52 patients achieved anti-HBe seroconversion. Fig. 3B shows the cumulative seroconversion rate of HBeAg calculated by the Kaplan–Meier test. The proportions of patients who showed seroconversion were 12%, 18%, 29%, and 38% at years 1 through 4, respectively. Univariate analysis demonstrated that age (>40 years, $p = 0.020$), albumin (<3.5 g/dl, $p = 0.021$) and platelet count (<20 × 10⁴/mm³, $p = 0.067$) correlated with HBeAg seroconversion at the start of treatment. Multivariate analysis that included the above factors identified serum albumin as the only significant determinant of seroconversion (<3.5 g/dl, RR = 2.0; 95% CI = 1.1–3.6, $p = 0.019$). One patient achieved anti-HBe seroconversion at 25 months but became positive again at 28 months. Other patients who achieved anti-HBe seroconversion did not show HBeAg reversion. One patient achieved anti-HBe seroconversion but remained HBV DNA positive (Table 4, Patient 5). Another patient remained positive for HBV
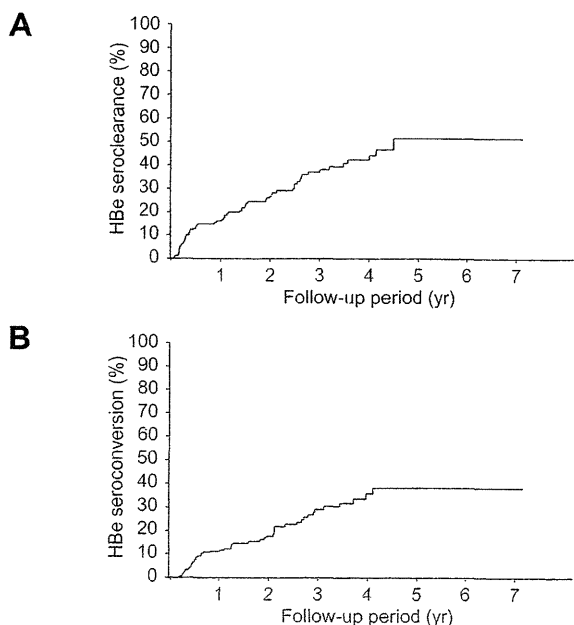

Fig. 3. Change of the HBeAg status during follow-up. Cumulative rates of (A) HBe seroclearance and (B) HBe seroconversion in HBeAg-positive patients, analyzed with the Kaplan–Meier test.
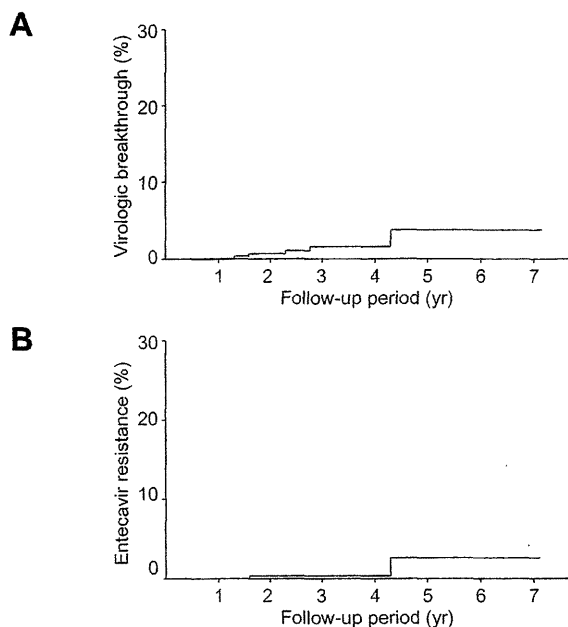
Research Article

A

B

Fig. 4. Cumulative rates of patients who showed resistance to therapy analyzed with the Kaplan–Meier test. (A) Virologic breakthrough (VBT) and (B) entecavir-resistance.

DNA after anti-HBe seroconversion. One patient became negative for HBsAg at week 28.

Monitoring resistance to treatment

Five patients showed VBT during the treatment period, including two patients (Patient 1 had been reported previously [13]) who developed entecavir-resistant mutations. None of the five patients had mutation(s) for entecavir at baseline. VBT was defined as any increase in serum HBV DNA by $>1 \log_{10}$ copies/ml from nadir or redetection of serum HBV DNA at levels 10-fold the lower limit of detection of the HBV DNA assay after having an undetectable result. Table 4 shows the patient baseline demo-

graphics, HBV DNA levels, and viral resistance profiles. All patients were positive for HBeAg and had serum levels of HBV DNA $>6 \log_{10}$ copies/ml at baseline. The median period until the appearance of the mutation was 120 (68–224) weeks. Two of the 49 (4%) patients who had detectable HBV DNA at the end of the first year subsequently developed resistance to entecavir. Furthermore, 3 of 49 (6%) patients who had detectable HBV DNA at the end of the first year developed VBT. Fig. 4A and B show the cumulative percentages of VBT and entecavir-resistance cases analyzed by the Kaplan–Meier test.

Discussion

Long-term data are rare for nucleoside-naïve patients treated continuously for more than 4 years with entecavir at the recommended dose of 0.5 mg daily. The only available data [6,8] were generated from follow-up studies of two phase III registration trials [3,4] in which patients showing complete response and non-responders were taken off entecavir. In the rollover studies, entecavir was administered to these patients at 1-mg dose at varying periods after cessation of the initial treatment. This double dose of entecavir was also given to patients showing a partial virological response after 48–96 weeks of entecavir at 0.5 mg daily. The present study has several unique features addressing specific and unanswered questions about entecavir treatment. It provided long-term results with respect to antiviral potency, viral resistance, and clinical safety for treatment-naïve patients who were treated continuously with entecavir at 0.5 mg daily for 4 years. Specifically, we found excellent viral suppression with 96% of patients achieving undetectable HBV DNA levels, only 1.1% (5/475) chance of viral breakthrough, and no clinically serious side effects after 4 years of treatment.

Genotype B was a significant factor associated with undetectable HBV DNA after the first year, although there were no significant differences after subsequent years. Previous studies showed conflicting results on the effect of HBV genotype on the response to lamivudine, with genotypes A, B, and C not affecting the antiviral response to lamivudine [14–16]. However, we have previously found that 47%, 84%, and 76% of patients had undetectable HBV DNA after the third year among patients of genotype

Table 4. Characteristics of patients with virologic breakthrough.

Patient No.	1	2	3	4	5
Age (yr)/gender	40/M	28/M	39/M	51/F	64/M
At start of entecavir therapy					
HBeAg status	+	+	+	+	+
HBV DNA ($\log_{10}$ copies/ml)	>7.6	>7.6	7.2	7.2	6.2
HBV genotype	H	A	C	C	C
Viral load at maximum suppression ($\log_{10}$ copies/ml)	<2.6	<2.6	<2.6	3.1	<2.6
Time of detection of mutation (wk)	83	224	120	68	145
HBV DNA ($\log_{10}$ copies/ml), maximum	6.8	7.2	7.1	7.6	7.8
Mutational pattern	L180M+/S202G+/M204V	L180M+/T184I+/S202G+/M204V	L180M+/M204V, L180M+/M204I	A181T	A181S+/T184A+/M204I

A, B, and C, respectively [17,18]. The difference among these groups was probably due to the younger age of patients of genotype A and that they were often positive for HBeAg compared to those of genotype B or C. However, the genotype was not a significant predictor of HBV DNA loss after >2 years of entecavir therapy in the present study. There was also no difference in HBeAg seronegativity with entecavir among patients infected with genotype A, B, or C virus. These results were consistent with studies on lamivudine therapy [14,18].

In this study, HBeAg positivity was a significant factor associated with detectable HBV DNA at years 1 through 3, and these results were consistent with those reported by Zoutendijk et al. [10]. In addition, lower HBV DNA and HBeAg negativity at baseline were associated with enhanced response to lamivudine therapy [18–20]. We have also previously reported that lamivudine induced a better response in HBeAg-negative patients with higher levels of serum ALT [17]. The most important factor of long-term entecavir therapy therefore was low HBV DNA level.

Low HBV DNA level at baseline correlated significantly with HBeAg seroclearance, but not with seroconversion. One of the reasons was that patients who showed HBeAg seroclearance but no seroconversion had lower HBV DNA (median; $6.7 \log_{10}$ copies/ml) at baseline compared to patients with seroconversion (median; $7.5 \log_{10}$ copies/ml, $p = 0.005$).

Univariate analysis showed that age (>40 years), serum albumin level (<3.5 g/dl), and platelet count ($<20 \times 10^4/mm^3$) correlated with HBeAg seroconversion rate. We also investigated the correlation between serum albumin and other factors. Serum albumin level correlated significantly with age ($r = -0.378$, $n = 216$, $p < 0.001$), platelet count ($r = 0.262$, $n = 215$, $p < 0.001$), AFP ($r = -0.372$, $n = 161$, $p < 0.001$), cirrhosis ($P < 0.001$) and male sex ($p = 0.004$). Multivariate analysis identified low serum albumin level (<3.5) as the only significant determinant of HBeAg seroconversion. In this regard, Chien et al. [21] reported that pre-treatment ALT was the only significant determinant of HBeAg seroconversion during lamivudine therapy. The reasons for the different findings are probably related to the study design. In our study, the age of patients at baseline was higher (47 vs. 32 years) and the duration of treatment was longer (2.4 [median] vs. 1 year) than in the study of Chien et al. [21]. Furthermore, differences in the pharmacodynamics of lamivudine and entecavir could also contribute to the observed differences between the two studies.

On the other hand, resistant mutants and breakthrough hepatitis seemed to be less frequent during long-term therapy with entecavir than with lamivudine [16–19], indicating that entecavir is better than lamivudine for long-term treatment of CHB and cirrhosis patients. Tenney et al. [6] reported that 9 out of 663 (1.4%) patients had baseline lamivudine-resistant mutations, and other studies also found only small numbers of preexisting lamivudine-resistant mutations in treatment-naïve patients [22–24]. It is known that the HBV rtM204V (usually with concomitant rt180M) mutation often acquires one of the entecavir signature mutations at rt184, rt202, or rt250 over long-term treatments and patients develop clinical HBV DNA breakthroughs. Although in vitro studies showed that rt204I mutations with or without rt180M conferred 3- to 21-fold decrease in entecavir susceptibility [25], in clinical practice, patients with rt204I mutations, even with the entecavir signature mutations, have lower levels of phenotypic resistance to entecavir and can often achieve undetectable HBV DNA levels [6,9,26]. Interestingly, there were three

patients in the present study with VBT who had no HBV DNA mutations at rt184, rt202, or rt250 with rt180M and rt204V (entecavir-resistance). The rtM204V/I mutation, lamivudine's signature mutation, is necessary but not sufficient for entecavir-resistance, causing an 8- to 10-fold decrease in susceptibility to entecavir compared with wild-type HBV. Other mutations at positions rtT184, rtS202, and rtM250 confer additional decreases in entecavir susceptibility [25,27,28]. In the present study, two patients (Patients #3 and 5) with mutations at position rtM204V/I, without rtT184, rtS202, or rtM250 mutations, showed emergence of VBT, as did one patient (Patient #4) with an rtA181T mutation, which was first reported in a LAM-treated patient [29]. Although the rtA181T mutation is related to resistance to adefovir dipivoxil, this mutation has not been linked to additional decreases in entecavir susceptibility. Future in vitro analyses using replication-competent HBV clones in patients with rtA181T mutations are therefore necessary.

In conclusion, long-term treatment of treatment-naïve CHB patients with 0.5 mg/day entecavir for 4 years suppressed HBV DNA to undetectable levels in more than 90% of patients, regardless of HBeAg status and genotype. Moreover, the drug was very safe and rarely induced resistance mutations. Further studies exploring the therapeutic efficacy over longer durations may be necessary to confirm these findings.

Conflict of interest

Financial support

References

[1] Lavanchy D, Hepatitis B. Virus epidemiology, disease burden, treatment, and current and emerging prevention and control measures. J Viral Hepat 2004;11:97–107.

[2] For the Chronic Hepatitis B Guideline Working Party of the Asian-Pacific Association for the Study of the Liver, Liaw YF, Leung N, Kao JH, Piratvisuth T, Gane E, Han KH, et al. Asian-Pacific consensus statement on the management of chronic hepatitis B. Hepatol Int 2008;2:263–283.

[3] For BEHoLD AI463022 Study Group, Chang TT, Gish RG, de Man R, Gadano A, Sollano J, Chao YC, et al. A comparison of entecavir and lamivudine for HBeAg-positive chronic hepatitis B. N Engl J Med 2006;354:1001–1010.

[4] For BEHoLD AI463027 Study Group, Lai CL, Shouval D, Lok AS, Chang TT, Cheinquer H, Goodman Z, et al. Entecavir versus lamivudine for patients with HBeAg-negative chronic hepatitis B. N Engl J Med 2006;354:1011–1020.

[5] Colonno RJ, Rose R, Baldick CJ, Levine S, Pokornowski K, Yu CF, et al. Entecavir resistance is rare in nucleoside naive patients with hepatitis B. Hepatology 2006;44:1656–1665.

[6] Tenney DJ, Rose RE, Baldick CJ, Pokornowski KA, Eggers BJ, Fang J, et al. Long-term monitoring shows hepatitis B virus resistance to entecavir in nucleoside-naive patients is rare through 5 years of therapy. Hepatology 2009;49:1503–1514.

[7] Yokosuka O, Takaguchi K, Fujioka S, Shindo M, Chayama K, Kobashi H, et al. Long-term use of entecavir in nucleoside-naive Japanese patients with chronic hepatitis B infection. J Hepatol 2010;52:791–799.

[8] Chang TT, Lai CL, Kew Yoon S, Lee SS, Coelho HS, Carrilho FJ, et al. Entecavir treatment for up to 5 years in patients with hepatitis B e antigen-positive chronic hepatitis B. Hepatology 2010;51:422–430.

[9] Yuen M-F, Seto W-K, Fung J, Wong DK-H, Yuen JC-H, Lai C-L. Three years of continuous entecavir therapy in treatment-naïve chronic hepatitis B

patients: Viral suppression, viral resistance, and clinical safety. Am J Gastroenterol 2011;106:1264–1271.

[10] For the VIRGIL Surveillance Study Group, Zoutendijk R, Reijnders JGP, Brown A, Zoulim F, Mutimer D, Deterding K, et al. Entecavir treatment for chronic hepatitis B: adaptation is not needed for the majority of naïve patients with a partial virological response. Hepatology 2011;54:443–451.

[11] Furusyo N, Nakashima H, Kashiwagi K, Kubo N, Hayashida K, Usuda S, et al. Clinical outcomes of hepatitis B virus (HBV) genotypes B and C in Japanese patients with chronic HBV infection. Am J Trop Med Hyg 2002;67:151–157.

[12] Matsuyama K, Hayashi K, Miura T. The quantitative assay for HBV DNA and the detection of HBV DNA point mutation by polymerase chain reaction— 'AMPLICOR HBV MONITOR test' and 'HBV pre core/core promoter mutation detection kit. Kan Tan Sui 2000;41:59–71.

[13] Suzuki F, Akuta N, Suzuki Y, Yatsuji H, Sezaki H, Arase Y, et al. Selection of a virus strain resistant to entecavir in a nucleoside-naïve patient with hepatitis B of genotype H. J Clin Virol 2007;39:149–152.

[14] Chan HL, Wong ML, Hui AY, Chim AM, Tse AM, Hung LC, et al. Hepatitis B virus genotype has no impact on hepatitis B e antigen seroconversion after lamivudine treatment. World J Gastroenterol 2003;9:2695–2697.

[15] Yuen MF, Wong DK, Sablon E, Yuan HJ, Sum SM, Hui CK, et al. Hepatitis B virus genotypes B and C do not affect the antiviral response to lamivudine. Antivir Ther 2003;8:531–534.

[16] Moskovitz DN, Osiowy C, Giles E, Tomlinson G, Heathcote EJ. Response to long-term lamivudine treatment (up to 5 years) in patients with severe chronic hepatitis B, role of genotype and drug resistance. J Viral Hepat 2005;12:398–404.

[17] Suzuki F, Tsubota A, Arase Y, Suzuki Y, Akuta N, Hosaka T, et al. Efficacy of lamivudine therapy and factors associated with emergence of resistance in chronic hepatitis B virus infection in Japan. Intervirology 2003;46:182–189.

[18] Kobayashi M, Suzuki F, Akuta N, Suzuki Y, Arase Y, Ikeda K, et al. Response to Long-term lamivudine treatment in patients infected with hepatitis b virus genotypes A, B, and C. J Med Virol 2006;78:1276–1283.

[19] Lai CL, Chien RN, Leung NW, Chang TT, Guan R, Tai DI, et al. A one-year trial of lamivudine for chronic hepatitis B. Asia Hepatitis Lamivudine Study Group. N Engl J Med 1998;339:61–68.

[20] Liaw YF. Therapy of chronic hepatitis B: current challenges and opportunities. J Viral Hepat 2002;9:393–399.

[21] Chien R-N, Liaw Y-F, Atkins M. Pretherapy alanine transaminase level as a determinant for hepatitis b e antigen seroconversion during lamivudine therapy in patients with chronic hepatitis B. Hepatology 1999;30:770–774.

[22] Kobayashi S, Ide T, Sata M. Detection of YMDD motif mutations in some lamivudine-untreated asymptomatic hepatitis B virus carriers. J Hepatol 2001;34:584–586.

[23] Kirishima T, Okanoue T, Daimon Y, Itoh Y, Nakamura H, Morita A, et al. Detection of YMDD mutant using a novel sensitive method in chronic liver disease type B patients before and during lamivudine treatment. J Hepatol 2002;37:259–265.

[24] Matsuda M, Suzuki F, Suzuki Y, Tsubota A, Akuta N, Hosaka T, et al. YMDD mutants in patients with chronic hepatitis B before treatment are not selected by lamivudine. J Med Virol 2004;74:361–366.

[25] Baldick CJ, Eggers BJ, Fang J, Levine SM, Pokornowski KA, Rose RE, et al. Hepatitis B virus quasispecies susceptibility to entecavir confirms the relationship between genotypic resistance and patient virologic response. J Hepatol 2008;48:895–902.

[26] Baldick CJ, Tenney DJ, Mazzucco CE, Eggers BJ, Rose RE, Pokornowski KA, et al. Comprehensive evaluation of hepatitis B virus reverse transcriptase substitutions associated with entecavir resistance. Hepatology 2008;47:1473–1482.

[27] Tenney DJ, Levine SM, Rose RE, Walsh AW, Weinheimer SP, Discotto L, et al. Clinical emergence of entecavir-resistant hepatitis B virus requires additional substitutions in virus already resistant to lamivudine. Antimicrob Agents Chemother 2004;48:3498–3507.

[28] Osborn M. Safety and efficacy of entecavir for the treatment of chronic hepatitis B. Infect Drug Resist 2011;4:55–64.

[29] Yeh CT, Chien RN, Chu CM, Liaw YF. Clearance of the original hepatitis B virus YMDD-motif mutants with emergence of distinct lamivudine-resistant mutants during prolonged lamivudine therapy. Hepatology 2000; 31:1318–1326.

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Determinants of the clinical outcome of patients with severe acute exacerbation of chronic hepatitis B virus infection

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Abstract

Background Severe acute exacerbation of chronic hepatitis B can sometimes occur and lead to hepatic failure and death. The objective of this study was to elucidate the predictors of progression to hepatic decompensation during severe acute exacerbation.

Methods We prospectively analyzed 37 consecutive patients with acute exacerbation of chronic hepatitis B (accompanied by jaundice and coagulopathy) for clinical outcome and factors that influenced the development of severe acute exacerbation, including viral kinetics.

Results Fourteen (37.8%) patients progressed to severe acute exacerbation (accompanied by encephalopathy). Multivariate analysis identified serum bilirubin (>5 mg/dl, $P = 0.002$) as a significant determinant of progression to hepatic failure and prothrombin activity (<45%, $P = 0.028$) and as a determinant of liver-related death. The hepatitis B virus (HBV) DNA level before therapy was measured in 25 patients. HBV DNA levels increased or did not change from before commencement of treatment in all 11 patients who progressed to severe acute exacerbation. On the other hand, HBV DNA levels did not change or increased in 8 of 14 patients (57%) with acute exacerbation ($P = 0.02$).

Conclusions Serum bilirubin and prothrombin activities were significant predictors of clinical outcome in patients with severe acute exacerbation of chronic hepatitis B. Viral kinetics until commencement of therapy can predict the severity of acute exacerbation of chronic hepatitis B.

Keywords Hepatitis B · Acute exacerbation · HBV DNA · Genotype · Encephalopathy

Abbreviations

AE	Acute exacerbation
ALT	Alanine aminotransferase
BCP	Basal core promoter
CS	Corticosteroid
HBV	Hepatitis B virus
IFN	Interferon
LMV	Lamivudine
NA	Nucleos(t)ide analogue
PC	Pre-core
PT	Prothrombin activity
SAE	Severe acute exacerbation

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Introduction

More than 3 billion people worldwide and approximately 1.5 million people in Japan are chronically infected with hepatitis B virus (HBV), and chronic HBV infection is one of the most common causes of chronic hepatic failure and hepatocellular carcinoma (HCC) [1, 2]. Other complications of HBV infection include fulminant hepatitis and acute liver failure [3, 4]. Acute exacerbation (AE) in HBV carriers occurs either through a natural course [5, 6] or following intensive chemotherapy or immunosuppressive

therapy [7, 8]. Some abrupt flares may be so severe that decompensation or even fulminant hepatic failure may occur [9–11]. Previous studies have identified pre-existing cirrhosis, high serum bilirubin levels, prolonged prothrombin time, pre-core/core promoter mutants, and high HBV DNA levels as factors associated with hepatic decompensation during AE in HBV carriers, though little is known about the predictive factors [9, 12, 13].

Liver transplantation is suitable therapy for acute hepatic failure, but the rate of liver transplantation has remained about 20% in Japan, where living donor liver transplantation is dominant [14, 15]. Thus, it is necessary to establish other effective therapies for patients with AE apart from liver transplantation. Steroids can rapidly inhibit excessive immune response and inflammatory reactions, and have been reported to be effective in cases of severe and potentially life-threatening exacerbation of chronic HBV (CHB) infection [16]. With the advent of oral nucleos(t)ide analogues (NAs), most guidelines recommend NAs for patients with AE of CHB infection [17–19], and several observational studies reported the use of NAs [9–11, 20, 21]. Timely use of potent anti-HBV agents, such as NAs, interferon (IFN), and steroids [22], during and/or after the development of hepatic decompensation could be potentially effective against various host- and virus-related factors.

The aim of the present study was to investigate the factor(s) that influence the rapid development of hepatic decompensation during AE of CHB.

Materials and Methods

Patients

The study subjects were patients with AE admitted to the Department of Hepatology, Toranomon Hospital, Tokyo, between 1984 and 2010. All patients were either followed up at our hospital with clinicopathologically proven CHB infection or were new patients with sudden-onset hepatic flares who visited our hospital outpatient clinic or were referred to our hospital from other clinics/hospitals. The diagnosis of CHB carrier state was established based on either positivity for hepatitis B surface antigen (HBsAg) for at least 6 months prior to the development of AE, or the presence of a high titer of anti-hepatitis B core antibodies (anti-HBcAb), together with negativity or a low titer of IgM anti-HBcAb. Chronic hepatitis and cirrhosis were confirmed by laparoscopy, needle biopsy, or ultrasonography, or treatment for these conditions for 1 year before the development of AE. AE of CHB infection was diagnosed by the following criteria: (1) an abrupt increase in serum alanine aminotransferase (ALT) levels to >300 IU/l

in patients with original ALT levels of less than 5× the upper limit of normal or an abrupt two-fold increase in the serum ALT level to greater than 5× the upper limit of normal, (2) hyperbilirubinemia [serum bilirubin (Bil) >3.0 mg/dl], (3) evidence of coagulopathy with plasma prothrombin activity (PT) of <60% during the clinical course, and (4) lack of encephalopathy at admission. We also applied the following exclusion criteria: (1) the presence of viral markers other than HBV (hepatitis A, C, D, E, Epstein-Barr virus, cytomegalovirus, herpes simplex virus), (2) HBV reactivation induced by immunomodulators or chemo-/immunosuppressive therapy, (3) asymptomatic HBV carriers, (4) recent exposure to drugs and chemical agents as well as recent heavy alcohol intake, (5) breakthrough hepatitis caused by NAs, (6) evidence of decompensated liver disease before the onset of exacerbation as characterized previously, (7) HCC diagnosed by ultrasonography or computed tomography, and (8) coexistence of other serious medical conditions and other liver diseases, or metabolic diseases. Progression to severe acute exacerbation (SAE) was diagnosed by the development of hepatic encephalopathy of more than grade 2 within 8 weeks of onset associated with coagulopathy (PT <40%).

HBV DNA levels were measured serially to investigate the effects of HBV kinetics on the prognosis of patients with severe AE. HBV DNA levels were measured before treatment in 25 patients. "Before treatment" represented 1–8 weeks before commencement of treatment. HBV DNA levels were also measured after treatment in 27 patients. "After treatment" was defined as 2 weeks after commencement of therapy. Viral kinetics was assessed using the same assay in all individuals. The Local Ethics Committee of Toranomon Hospital approved the study, and informed consent was obtained from all patients.

Virological markers

Serial blood samples were obtained during the clinical course of AE and stored at −80°C until used for HBV molecular analysis. Serological tests for HBsAg, HBsAb, hepatitis e antigen (HBeAg), IgM anti-HBcAb, total anti-HBcAb, and anti-HBeAb were conducted using radioimmunoassay kits (Abbot Diagnostics, Chicago, IL, USA) according to the instructions provided by the manufacturer. Precore (PC) mutations were analyzed by PCR enzyme-linked mini-sequence assay (Roche Diagnostics, Tokyo, Japan), and basal core promoter (BCP) mutations were analyzed by PCR specific probe assay (Roche Diagnostics, Tokyo, Japan). HBV DNA was measured by Amplicor monitor assay (dynamic range 2.6–7.6 log copies/ml, Roche Diagnostics, Tokyo, Japan), COBAS TaqMan v.2.0 (dynamic range 2.1–9.0 log copies/ml, Roche Diagnostics), transcription-mediated amplification and hybridization

protect assay (TMA-HPA) (dynamic range 3.7–8.7 LGE/ml, Chugai Diagnostics Science Co., Tokyo) or sandwich hybridization assay with signal amplification using branched DNA (bDNA, dynamic range 0.7–3800 Meq/ml). The major genotype of HBV was determined using enzyme-linked immunosorbent assay (ELISA, Institute of Immunology, Tokyo, Japan) or PCR-invader assay (BML, Inc, Tokyo, Japan) based on the methods described previously [23, 24]. HBVDNA levels assessed by bDNA were re-measured by TaqMan PCR assay using stored serum samples.

Statistical analysis

Continuous variables were expressed as median (range), and compared by Mann–Whitney U test. Categorical variables were compared by χ^2 test or Fisher's exact test, as appropriate. Univariate analysis was applied to determine the relationship between SAE and each of the following factors: sex, age, presence of compensated cirrhosis, and various biological and virological markers as measured at baseline (bilirubin, PT, ALT, albumin, HBeAg, HBV DNA, and HBV genotype, PC and BCP mutations). Each continuous variable was transformed into two categories based on the value with the largest capacity to discriminate between patients for univariate and multivariate analyses. Factors that correlated significantly with SAE were entered into multiple logistic regression analysis, and the odds ratio (OR) with 95% confidence intervals (95% CI) were determined. All analyses were performed using The Statistical Package for Social Sciences (SPSS II v. 11.0, Chicago, IL, USA), and statistical significance was taken as a two-sided P value <0.05.

Results

Clinical features of severe acute exacerbation

A total of 37 patients (30 men and 7 women) fulfilled the criteria of AE and were included in this study. The baseline characteristics at the commencement of therapy of these 37 patients are shown in Table 1. Twenty-two patients were observed at our hospital, and 15 patients were referred from another hospital after the onset of hepatic flares. The majority of patients had genotype C, and 27 patients (72.9%) were HBeAg positive. The PC and BCP mutations were determined in 27 patients; 22 patients had mutations in the PC region, 16 patients had mutations in the BCP region, and 12 patients had mutations in both the PC and BCP regions. During the clinical course, the peak median values were: ALT 713 IU/l (range 307–2857), bilirubin 8.4 mg/dl (3.0–51.4), and PT 47.6% (12.0–60.0).

Table 1 Baseline characteristics of the 37 patients infected with HBV who developed severe acute exacerbation at the commencement of therapy

Number	37
Sex (male/female)	30/7
Age (years)	45 (23–63)
Family history (yes/no)	21/16
Cirrhosis (present/absent)	7/30
Albumin (g/dl)	3.4 (2.5–4.6)
Bilirubin (mg/dl)	4.7 (1.0–30.7)
AST (IU/l)	601 (64–2593)
ALT (IU/l)	657 (124–2142)
LDH (IU/l)	297 (106–594)
Platelets ($\times 10^4$/mm^3)	12.3 (6.2–32.0)
α-Fetoprotein (μg/ml)	62.0 (3.0–1600)
Prothrombin activity (%)	53 (26–80)
Genotype (A/B/C)	0/5/32
HBeAg (positive/negative)	27/10
HBV-DNA ($\log_{10}$ copies/ml)	8.5 (6.8–8.9)
PC (wild/mutant/ND)	5/22/10
BCP (wild/mutant/ND)	11/16/10

Data are median values (range) or number of patients

AST aspartate aminotransferase, *ALT* alanine aminotransferase, *LDH* lactate dehydrogenase, *HBeAg* hepatitis B envelope antigen, *PC* pre core, *BCP* basal core promoter, *ND* not done

Treatment

NAs were used in 19 patients, IFN in 8, and corticosteroids (CS) in 20 patients. In addition, 7 patients were treated with a combination of NAs and CS; 2 patients were treated with three drugs (NAs, IFN, and CS). At the time of the study, lamivudine (LMV) was not yet available for the treatment of chronic hepatitis B, and thus IFN was used; 6 patients were treated with both IFN and CS. None of the patients underwent liver transplantation.

Prognosis of severe acute exacerbation and factors associated with progression to hepatic failure

Of the 37 patients admitted with CHB infection and AE, 23 (62.2%) did not develop SAE. The remaining 14 (37.8%) patients developed SAE; 9 (24.3%) patients died of liver-related death, but 5 (13.5%) survived. Further analysis showed that 8 (36.4%) of 22 patients who were observed in our hospital developed AE, and 6 (27.3%) of these patients died, whereas 6 (40.0%) of 15 patients who were referred from other hospitals after the onset of exacerbation developed AE, and 3 (20.0%) of these patients died. There was no significant difference in prognosis by treatment facility before AE. Ten of 37 patients experienced AE before 2000 when LMV was available in Japan, and 19