varied from 93% to 97% during 1993-2009, excluding 2007, when reported first-dose measles vaccine coverage was 83% because of a nationwide stock out. Measles vaccine first-dose is administered at 9 months of age, the second dose of measles vaccine was introduced in 2006 for children entering primary school and is administered at 6-7 years of age. Rubella vaccine was not introduced into the routine immunization.

Molecular epidemiologic data, when analyzed in conjunction with standard epidemiologic data, can help document viral transmission pathways, identify whether a virus is endemic or imported, and aid in case classification, thus enhancing control and elimination programs. Genetic analysis results showed that the H1 genotype virus was the predominant endemic measles virus in the North VietNam in 2006-2012. H1 genotype measles was also detected epidemic in Korea, China and in the centre of VietNam in 2000. In China, some studies showed that genotype H1 is the endemic genotype circulating in at least 16 years.

2B genotype of RV was circulating in some provinces in the North Viet Nam in 2008 - 2012. This genotype had a wide geographic distribution and were frequently found and were reported from 2 Middle Eastern countries, 5 European countries, 4 Southeast Asian, 4 South and Central American countries, 3 African countries and 2 Western pacific countries.

8.11. Reference

- 1. World Health Organization. Manual for the laboratory diagnosis of measles and rubella viral infection. 2nd ed. 2007
- 2, Griffin DE (2001) Measles virus. In: *Knipe DM, Howley PM, Griffin DE, Lamb RA, Martin MA, Roizman Strauss SE*. Fields virology, 4th edn. Lippincott Williams, and Wilkins, Philadelphia, pp 1401–1441).
- 3. Ono N, Tatsuo H, Hidaka Y, Aoki T, Minagawa H, Yanagi Y. Measles viruses on throat swabs from measles patients use signaling lymphocytic activation molecule (CDw150) but not CD46 as a cellular receptor. J Virol. 2001;75:4399–401. DOI: 10.1128/JVI.75.9.4399-4401.2001
- 4. Paul A. Rota, Kevin E. Brown, David W. Brown, William J. Bellini and David Feathestone. Improving Global Virologic Surveillance for Measles and Rubella. The Journal of infectious diseases 2011: 204 (Suppl 1). S506-513.
- 5. Paul A. Rota, Kevin E. Brown, David W. Brown, William J. Bellini, Youngmee Jee and David Feathestone. Global Distribution of Measles Genotypes and Measles Molecular Epidemiology. The Journal of infectious diseases 2011: 204 (Suppl 1). S514-523.
- 6. David h. Sniadack, Jorge Medoza Aldana, Dang Thi Thanh Huyen, Trieu Thi Thanh Van, Nguyen Van Cuong, Jean Marc Olive, Kohei Toda and Nguyen Tran Hien. Epidemiology of a Measles Epidemic in Vietnam 2008-2010. The Journal of infectious diseases 2011: 204 (Suppl 1). S476-482.
- 7, Xu W, Tamin A, Rota JS, Zhang L, Bellini WJ, Rota PA (1998) New genetic group of measles virus isolated in the People's Republic of China. Virus Res 54:147–156).
- 8, *Bellini, W. J., and J. P. Icenogle*. 2007. Measles and rubella viruses, p. 1378–1391. *In P. R. Murray*, E. J. Baron, M. A. Pfaller, J. H. Jorgensen, and R. H. Yolken (ed.), Manual of clinical microbiology, 9th ed. ASM Press, Washington, DC.
- 9, Caidi, H., E. S. Abernathy, A. Benjouad, S. Smit, J. Bwogi, M. Nanyunja, R.El Aouad, and J. Icenogle. 2008. Phylogenetic analysis of rubella viruses found in Morocco, Uganda, Cote d'Ivoire and South Africa from 2001 to

2007. J. Clin. Virol. 42:86-90.

10, Chen, M. H., and J. P. Icenogle. 2007. Molecular virology of rubella virus, p.1–18. In J. Banatvala and C. Peckham (ed.), Rubella virus. Elsevier, Oxford, United Kingdom.

8.12. Acknowledgement

We sincerely thank the financial supports from a grant-in-aid of Ministry of Health, Labor and Welfare, the Government of Japan (H23-Shinkou-shitei-020) and WHO Vietnam country office;

Research 9

- **6.13 Title:** Laboratory diagnosis of enteroviruses from cases with Hand, Foot, and Mouth Disease in 2011-2012 in Vietnam North
- **6.14** Name of researcher: Nguyen Thi Hien Thanh

Tran Nhu Duong

6.15 Specific objectives:

- 1. Identification of causative agents and epidemiological characteristics of HFMD
- 2. Identification of molecular characteristics of EV71,

Virological Investigation of Hand, Foot, and Mouth Disease, Northern Vietnam, 2011 6.16 Abstract

603 / 912 clinical samples from patients with hand, foot and mouth disease in 2011 collected from Northern provinces of Vietnam are positive with enteroviruses including 275 positive samples with HEV71 (45,6%) and 328 positive samples with other enteroviruses (54,4%). As the result of 328 sequenced PCR products, 177 samples were identified as Coxsackievirus A6 (53,9%), 102 as Coxsackie A16 (31,3%), 4,6% as other CoxsackieA viruses including type 3, 10, 12 and 13; 0,6% as Coxsackievirus type B; 4,3% as echo viruses; and 0,9% as Polio-Sabin virus and enteroviruse typ 96, 15 samples were identified as Rhinovirus (4,6%). Phylogenetic analysis of 49 HEV71 strains showed 2 genogroups B and C with 3 subgenogroups B5, C4 and C5 that cocirculated in norththern Vietnam in 2011. However, viruses belonging to subgenogroup C4 predominated with 73.5% and after is 20.4% as B5, only 6.1% as subgenogroup C5 that circulated during a long period before. Disease was mainly detected in children under 3 years of age

Keywords: Hand, Foot and Mouth Disease, enterovirus, human enterovirus 71, coxsackie virus A6, A16; coxsackie virusB5; Echo virus30

6.17 Introduction

Hand, foot, and mouth disease (HFMD) is a common febrile illness of early childhood, characterized by 3–4 days of fever and the development of a vesicular enanthem on the buccal mucosa, gums, and palate and a papulovesicular exanthem on the hands, feet, and buttocks (1). HFMD is caused by acute enterovirus infections, particularly by viruses belonging to the human enterovirus A (HEVA) species

The genus *Enterovirus* of the family *Picornaviridae* is divided into 9 species, 5 of which infect humans. These viruses include the prototype species poliovirus, as well as HEVA, HEVB, HEVC, and HEVD. Viruses belonging to the HEVA species include 11 serotypes of coxsackievirus A (CVA; serotypes 2–8, 10, 12, 14, and 16), and human enterovirus 71 (HEV71)

Since the discovery of HEV71 in 1969 , numerous outbreaks of this infection have occurred throughout the world, the prevalence of HEV71 infection in the Asia-Pacific region has greatly increased since 1997, concurrent with an increase in the prevalence of HFMD, HFMD outbreaks have been recorded in Japan, Malaysia, Singapore, South Korea, the People's Republic of China, and Australia, The most extensive epidemic of HEV71 occurred in Taiwan in 1998, with $\approx\!1,3\times10^5$ cases of HFMD, 405 cases related to severe neurologic syndrome , and 78 deaths, The deaths were due primarily to the development of brainstem encephalitis and neurogenic pulmonary edema

In 2003, we isolated HEV71/C4 from 01 patient with HFMD and have relation to severe neurologic syndrome, In 2008, a HFMD outbreak happened in the north of Vietnam and we isolated HEV71/C5 from 7 patients in this outbreak, Although laboratory surveillance has been shown to provide adequate warning of impending outbreaks of enteroviruses associated HFMD, laboratory surveillance for enteroviruses from patient with HFMD has not yet been established

A reverse transcription-seminested PCR (RT-snPCR) assay was developed for the detection RNA of EVs and EV71 from clinical specimens. The VP1 RT-snPCR assay was slightly more sensitive, and was used to identify EVs in clinical specimens. A product of the

expected size was successfully amplified and sequenced from clinical samples, The VP1 sequences derived from the RT-snPCR products allow rapid phylogenetic and molecular epidemiologic analysis of strains circulating during the EV season and comparison with EV sequences from past seasons or from different locations around the world,.

6.18 Materials and Methods

6.18.1 Specimen Collection

A total of 922 specimens were collected from the children with HFMD. Each child had at least 1 specimen collected from vesicle fluid, throat swab, or stool. 20% of specimens in PBS (+) were treateded with chloroform (1:10 in phosphate-buffered saline) before use for virus isolation in cell culture and molecular biology

Table 1, Primer used for research

AN32	5-GTYTGCCA		3009-3002	
AN33	5-GAYTGCCA	cDNA	3009-3002	
AN34	5-CCRTCRTA	CDNA	3111-3104	
AN35	5-RCTYTGCCA		3009-3002	
SO224-F	5-GCIATGYTIGGIACICAYRT	PCR1	2207-2226	762
SO222R	5-CICCIGGIGGIAYRWACAT	PCRI	2969-2951	
AN89F	5-CCAGCACTGACAGCAGYNGARAYNGG	PCR2/	2602-2627	375
AN88R	<i>5-TACTGGACCACCTGG</i> NGGNAYRWACAT	EVs	2977-2951	
MAS01S	5'- ATAATAGCA(C/T)T(A/G)GCGGCAGCCCA -3')	PCR2/	2352-2375	376
MAS02A	5' – AGAGGGAG(A/G)TCTATCTC(C/T)CC -3')	EV71	2709-2728	

RNA extraction, Stool suspensions were prepared by adding 5 ml of phosphate-buffered saline (+), 1 g of glass beads (Corning Inc., Corning, NY), and 0,5 ml of chloroform to 1 g of stool sample, shaking the mixture vigorously for 20 min in a mechanical shaker, and centrifuging at 1,500 _ g for 20 min at 4°C (33), the supernatant was transferred to a fresh tube, (10% stool suspensions), 140 _1 of the specimen extract was combined with an equal volume of Vertrel XF (Miller-Stephenson Chemical Co., Danbury, CT), shaken vigorously, and then centrifuged at 13,000 _ g for 1 min at room temperature, The aqueous phase was transferred to a fresh tube, Other specimen types (including blister fluid; throat swab samples) were processed without pretreatment, Twenty micrograms of proteinase K (Roche Applied Science, Indianapolis, IN) was added to 140 _1 of each liquid specimen or fecal extract, and the mixture was then incubated for 30 min at 37°C, Nucleic acid was extracted from the digested specimen with a QIAamp Viral RNA mini kit (QIAGEN, Inc., Valencia, CA), which was used according to the manufacturer's instructions, The eluted RNAs were passively dried in a benchtop desiccator under vacuum, The dried RNA was resuspended in 16 _1 of sterile nuclease-free water and stored at _20°C until use,

RT-snPCR and sequencing.

- Synthesis of cDNA was carried out in a 5 μ l reaction mixture containing 4 μ l of RNA and 1 pmol each cDNA primer (primers AN32, AN33, AN34, and AN35; Table 1), heat mix in a 70° C hear block for 5 min and immediately chill in ice water for at least 5 min. In a 20 μ l reaction mixture containing 05 μ l RNA reaction mixture, 100 μ M each deoxynucleoside triphosphate (dNTP; Invitrogen), 4 μ l of 5 x reaction buffer (GoStript, Promega), 1.5 μ l MgCl2, 20 U of RNasin (Promega Corp., Madison, WI), and 100 U of GoScript reverse transcriptase (Rocke) , incubation at 25°C for 05 min, 42°C for 60 min, and 70°C for 15 min
- Synthesis of PCR1: Following incubation, the entire 05 μ l RT reaction mixture was then used in the first PCR (final volume, 25 μ l) (PCR1), consisting of 5 μ l of 10 x PCR buffer (Roche Applied Science), 200 μ M each dNTP, 50 pmol each of primers 224 and 222 (Table 1), and 2,5 U of *Taq* DNA polymerase (Roche Applied Science), with 40 cycles of amplification (95°C for 30 s, 42°C for 30 s, 60°C for 45 s),

One microliter of the first PCR was added to a second PCR (PCR2/EV71) for seminested amplification, PCR2/EV71 contained 40 pmol each of primers MAS01S and

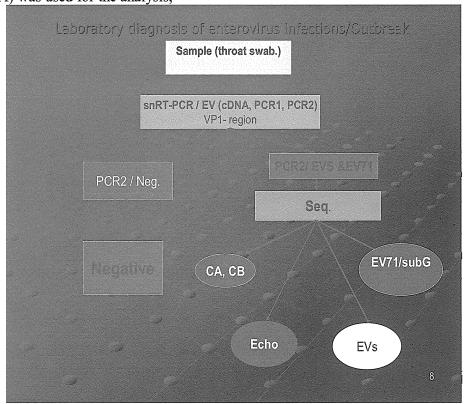
MAS02A (Table 1), 200 μM each dNTP, 5 μl of 10 x FastStart *Taq* buffer (Roche Applied Science), and 2,5 U of FastStart *Taq* DNA polymerase (Roche Applied Science) in a final volume of 50 μl, The FastStart *Taq* polymerase was activated by incubation at 95°C for 6 min prior to 40 amplification cycles of 95°C for 30 s, 60°C for 20 s, and 72°C for 15 s, The reaction products were separated and visualized on 2% agarose gels containing 0,5 μg ethidium bromide per ml

One microliter of the first PCR was added to a second PCR (PCR2/EV) for seminested amplification, PCR2 contained 40 pmol each of primers AN89 and AN88 (Table 1), 200 µM each dNTP, 5 µl of 10 x FastStart *Taq* buffer (Roche Applied Science), and 2,5 U of FastStart *Taq* DNA polymerase (Roche Applied Science) in a final volume of 50 µl, The FastStart *Taq* polymerase was activated by incubation at 95°C for 6 min prior to 40 amplification cycles of 95°C for 30 s, 60°C for 20 s, and 72°C for 15 s, The reaction products were separated and visualized on 1,2% agarose gels containing 0,5 µg ethidium bromide per ml and were purified from the gel by using a QIAquick gel extraction kit (QIAGEN), Slight variations in the sizes of the PCR products (350 to 400 bp) were observed due to VP1 gene length differences in the different serotypes, as described previously (12–14, 19), The resulting DNA templates were sequenced with a BigDye Terminator v1,1 ready reaction cycle sequencing kit on an ABI Prism 3100 automated sequencer (both from Applied Biosystems, Foster City, CA) by using primers AN89 and AN88 (Table 1),

Sequence analysis: The amplicon sequences were compared with the VP1 sequences of EV reference strains, including at least one representative of each recognized serotype, by script-driven sequential pair wise comparison with the program Gap (Wisconsin Sequence Analysis Package, version 10,2; Accelrys, Inc., San Diego, CA), as described previously (15, 18, 19), In cases where the result was not unequivocal (highest score less than 75% or second-highest score greater than 70%), the deduced amino acid sequences were compared by a similar method,

6.18.2 Statistical Methods

Differences between proportions were tested by using the χ^2 test with Yates correction or Fisher exact test, Epi Info version 6 (Centers for Disease Control and Prevention, Atlanta, GA, USA) was used for the analysis,



Flow chart for detection of all enterovirus serotypes from clinical samples

6.19 RESULTS

6.19.1 Enteroviruses were protected by snRT-PCR and Seq.

Table1. Result of Enteroviruses and EV71 by snRT-PCR in 2011

NI	N	%			(+) E	EV71	(+) Enterovirus	
N report	collected samples	tested	N (+)	% (+)	N(+)	%(+)	N(+)	%(+)
20.520	010	4.4	602	60.5	255	45.6	220	5.4.4
20.529	912	4.4	603	63.5	275	45.6	328	54.4

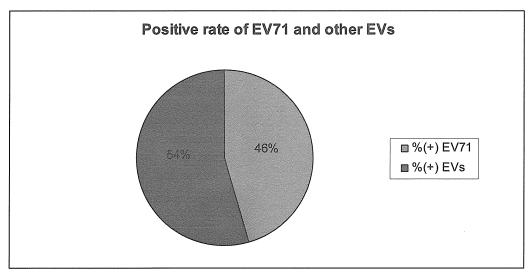


Figure 1. HEV-71 and EVs was protected by snRT-PCR

In 2011, 603 of 912 HFMD cases (63.5%) were found positive for HEV by using enterovirus general primers, and snRT-PCR method. HEV-71 and EVs occupied 46% (275) and 54% (328) respectively. (Figure 1).

Table2. Result of enterovirus serotypes by sequencing in 2011

N		Result of Enterovirus serotypes by seq.							
enterovirus	(+) Coxsackievirus - A					(+)			
by snRT- PCR	CAV- 6	CAV-16	Other CAV	(+) CB	(+) Echo	Other EV	Rhinovirus		
328	177 53,9%	102 31,3%	15 4,6%	2 0,6%	14 4,3%	3 (EV96 & Polio) 0,9%	15		
		89,6%			5,8%		4,6%		

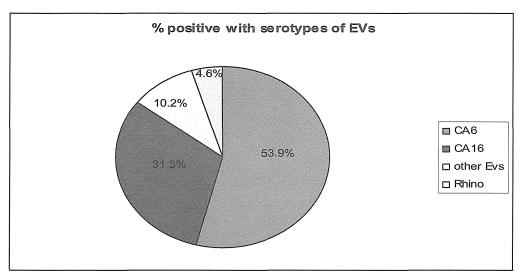


Figure 2. Enterovirus Serotypes was confirmed by Sequencing

328 of other EVs PCR-2 products were sequenced with primer pairs at position of primer F 2602-2627 or primer R 2977-2951 (375nt). The result showed with 53,9% of CoxsackieA 6; 31,3% of CoxsackieA16. The other entroviruses containing 10.2% with CV-A3(1), A10(4), A12 (8), và A13 (2); CV-B3, B4 (2);14 echovirus type 30 and 2 Polio, 1 EV96 were also detect (Figure 2)

Table 3. Enterovirus type 71's subgenogroups

TT of PCR produts/EV71	TT of EV71 PCR products sequenced	C4	C5	B5
275	49	36	3	10
% of each EV	71 subgenogroup	73.5	6.1	20.4

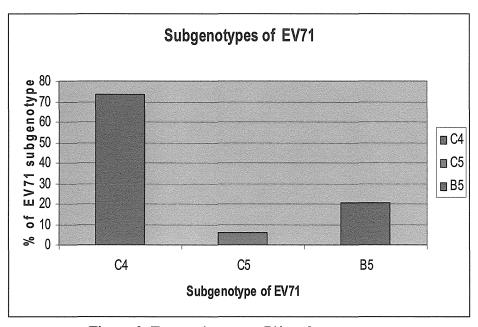


Figure 3. Enterovirus type 71's subgenogroups

49 PCR products of EV71 were amplified and sequenced. Result showed 73.5% as subgenotype C4, 6.1% as subgenotype C5 and 20.4 % as subgenotype B5

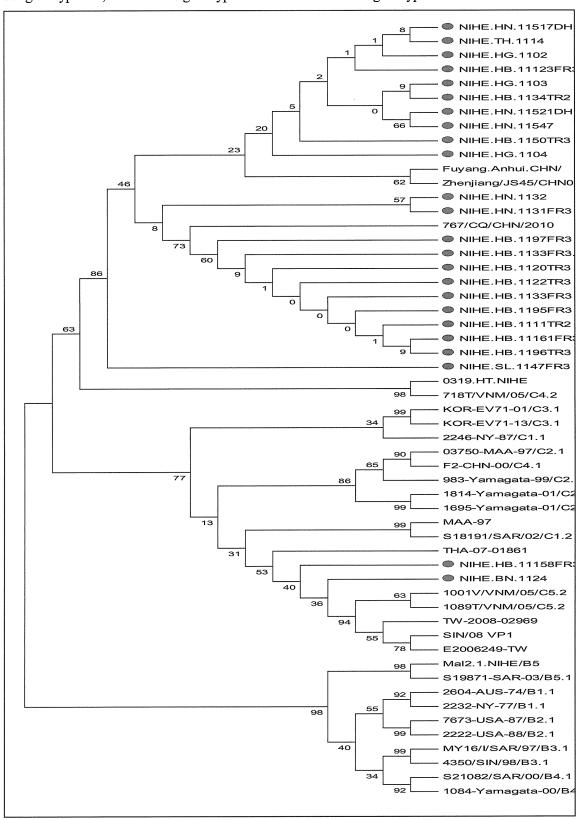


Figure 4. Phylogenetic relationships of human enterovirus 71 (HEV71) strains belonging to genogroup $\mathbb C$

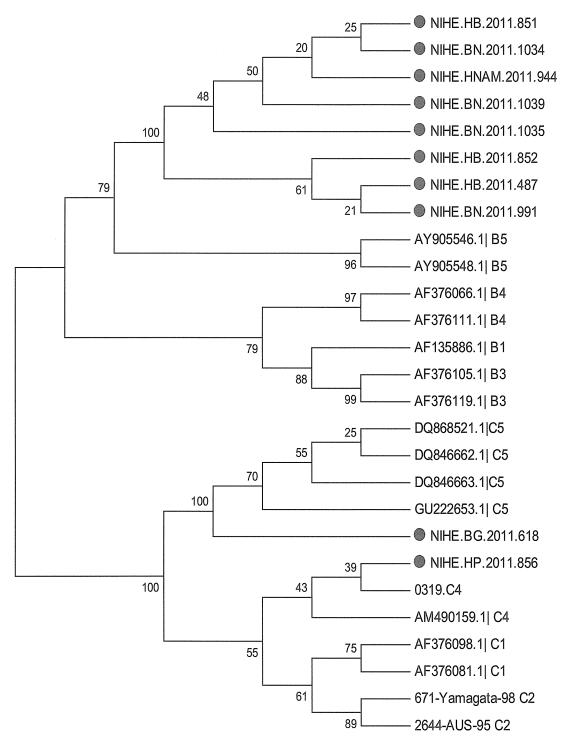


Figure 5. Phylogenetic relationships of human enterovirus 71 (HEV71) strains belonging to genogroup B and C

49 HEV-71 PCR products from 21 isolates and 28 clinical samples were sequenced. Result showed 79.6 % belonging to genogroup C (Figure 3,4) with 73.5% of subgenogroup C4 predominantly responsible for almost all HEV-71 infections in 2011 in the north of VN and 6.1% of subgenogroup C5, and 20.4% belonging to genogroupB, subgenogroup B5 (Figure4). However, the analysis of recent and previous HEV71 isolates in the Western Pacific Region showed that several subgenogroups, B1, B2, B3, B4, C1, C2, C3 and C4 were cocirculating in Australia, Malaysia, Singapore, Taiwan and Japan respectively.

Table 4. Ages distribution of 603 cases with HFMD confirmed as EV71 and other enteroviruses

Aga	N	NI(±)	N (+)	(+) EV71		(+) VRĐR khác		
Age	tested	N(+)	VRÐR	N(+)	% (+)	N(+)	%(+)	
<1	97	62	10.3%	22	8.0%	40	12.8%	
1	329	232	38.5%	97	35.3%	135	43.1%	
2	261	183	30.3%	87	31.6%	96	30.7%	
3	103	57	9.5%	33	12.0%	24	7.7%	
4	37	22	3.6%	13	4.7%	9	2.9%	
5	30	15	2.5%	7	2.5%	8	2.6%	
6 -30	39	20	3.3%	12	4.4%	8	2.6%	
No infn	16	12	2.0%	4	1.5%	8	2.6%	
TT	912	603	100%	275	100%	313	100%	

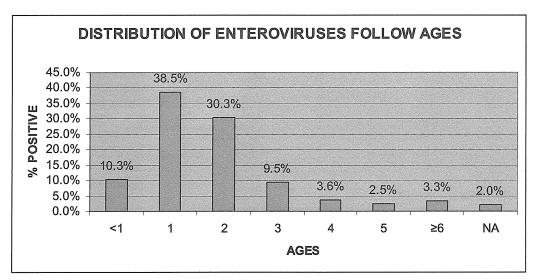


Figure 6, Ages distribution of 603 cases with HFMD confirmed as enteroviruses

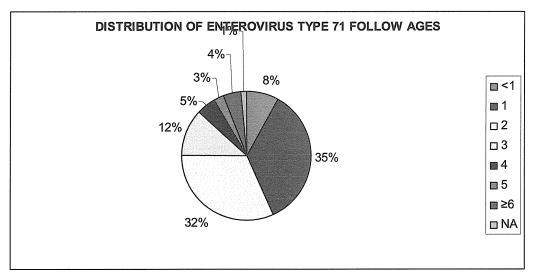


Figure 7, Ages distribution of 275 cases with HFMD confirmed as enterovirus TYPE 71

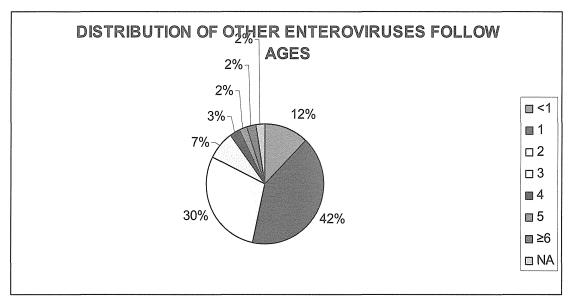


Figure 8, Ages distribution of 328 cases with HFMD confirmed as other enteroviruses

Table 4 showed the results of HFMD following age group, the disease is highly concentrated in children within 3 years, there is no difference of the incidence of diseases caused by enterovirus type 71 (46.7%) and other enterovirus (53.3%) in each age group.

Table 5. Geographic distribution of hand, foot, and mouth disease cases

		N-	N-	(1)		***************************************		(1)	
IT	Provinces		tested	(+) Enteoviruses	% (+) EVs	(+) EV71	% (+)	(+) other EVs	% (+)
		report		*****					
1	LẠNG SƠN	662	2	1	0.2%	0	0,0	1	100,0
2	HƯNG YÊN	154	3	2	0.3%	0	0,0	2	100,0
3	T, NGUYÊN	224	6	2	0.3%	0	0,0	2	100,0
4	LAI CHÂU	50	6	3	0.5%	2	66,7	1	33,3
5	NGHỆ AN	561	10	4	0.7%	1	25,0	3	75,0
6	SON LA	174	6	5	0.9%	5	100,0	0	0,0
7	ĐIỆN BIÊN	28	10	6	1.0%	5	83,3	1	16,7
8	THÁI BÌNH	814	12	6	1.0%	1	16,7	5	83,3
9	HÀ NAM	270	16	7	1.2%	2	28,6	5	71,4
10	CAO BĂNG	257	10	9	1.5%	1	11,1	8	88,9
11	HÀ TĨNH	118	13	9	1.5%	1	11,1	8	88,9
12	LÀO CAI	88	10	9	1.5%	4	44,4	5	55,6
13	YÊN BÁI	495	11	9	1.5%	2	22,2	7	77,8
14	NINH BÌNH	1,028	33	15	2.6%	1	6,7	14	93,3
15	QUẢNG NINH	473	25	16	2.7%	2	12,5	14	87,5
16	HÀ GIANG	285	33	20	3.4%	10	50,0	10	50,0
17	NAM ĐỊNH	133	25	21	3.6%	9	42,9	12	57,1
18	BẮC CẠN	380	31	22	3.7%	9	40,9	13	59,1
19	VĨNH PHÚC	872	38	23	3.9%	2	8,7	21	91,3
20	HÅI DƯƠNG	536	45	24	4.1%	1	4,2	23	95,8

21	BẮC NINH	158	42	25	4.3%	11	44,0	14	56,0
22	T, QUANG	561	24	26	4.4%	15	57,7	11	42,3
23	PHÚ THỌ	967	77	31	5.3%	1	3,2	30	96,8
24	THANH HÓA	3,744	49	33	5.6%	25	75,8	8	24,2
25	BÅC GIANG	508	60	46	7.8%	8	17,4	38	82,6
26	HẢI PHÒNG	3,046	80	63	10.7%	56	88,9	7	11,1
27	HÀ NỘI	1,579	116	70	11.9%	32	45,7	38	54,3
28	HÒA BÌNH	2,364	119	81	13.8%	69	85,2	12	14,8
	Tổng	20,529	912	588		275	46,8	313	53,2

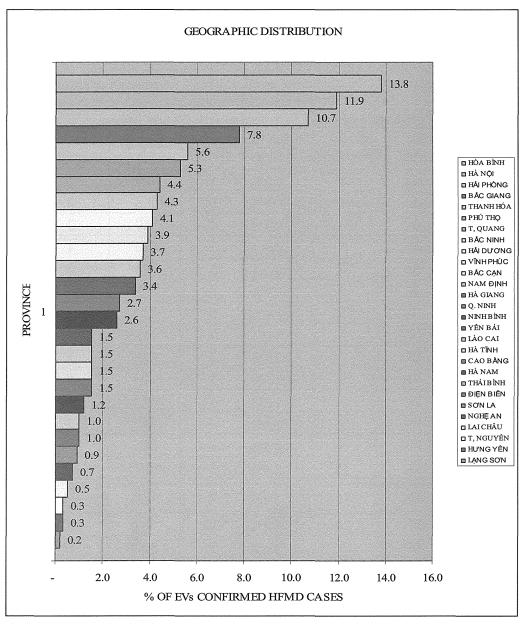
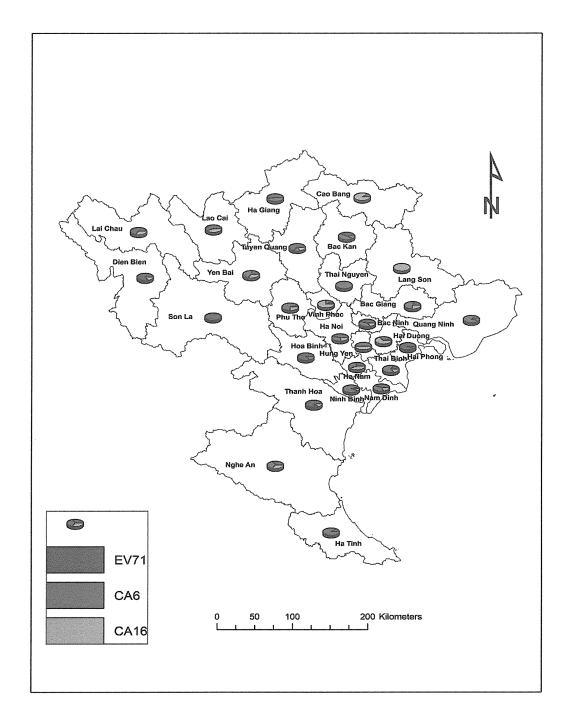


Figure 9. Geographic distribution of hand, foot, and mouth disease cases

The results showed 28/28 Northern provinces had HFMD cases. However, the HFMD cases in delta area was higher than mountain area; 24/28 provinces with the co-circulation of EV71 and other EVs. One only province (Son La province) only detected EV71 and three provinces (Hung Yen, Lang Son and Thai Nguyen province) only detected enterovirues

Table 6. Geographic distribution of hand, foot, and mouth disease cases by EV71, CAV-6 and CAV-16

CAV-Uallu CAV-	.10			
	% (+)/	% (+)/	% (+)/	% (+)/
PROVINCE	EV71	CA16	CA6	Other EVs
BẮC CẠN	40.9%	9.1%	50.0%	0.0%
BẮC GIANG	18.2%	22.7%	47.7%	9.1%
BẮC NINH	50.0%	18.2%	22.7%	4.5%
CAO BĂNG	11.1%	88.9%	0.0%	0.0%
ÐIỆN BIÊN	83.3%	16.7%	0.0%	0.0%
HÀ GIANG	55.6%	0.0%	38.9%	5.6%
HÀ NAM	28.6%	42.9%	28.6%	0.0%
HÀ NỘI	48.5%	20.6%	25.0%	2.9%
HÀ TĨNH	10.0%	0.0%	80.0%	0.0%
HẢI DƯƠNG	0.0%	56.5%	26.1%	17.4%
HẢI PHÒNG	100.0%	0.0%	0.0%	0.0%
HÒA BÌNH	93.3%	0.0%	4.0%	0.0%
HƯNG YÊN	0.0%	50.0%	50.0%	0.0%
LAI CHÂU	40.0%	20.0%	0.0%	0.0%
LẠNG SƠN	0.0%	100.0%	0.0%	0.0%
LÀO CAI	33.3%	50.0%	0.0%	16.7%
NAM ĐỊNH	42.9%	14.3%	42.9%	0.0%
NGHE AN	20.0%	20.0%	20.0%	0.0%
NINH BÌNH	7.7%	7.7%	69.2%	7.7%
PHÚ THỌ	3.3%	23.3%	46.7%	26.7%
QUẢNG NINH	13.3%	6.7%	73.3%	6.7%
SON LA	100.0%	0.0%	0.0%	0.0%
T. NGUYÊN	0.0%	0.0%	100.0%	0.0%
T. QUANG	47.6%	0.0%	38.1%	9.5%
THÁI BÌNH	14.3%	0.0%	85.7%	0.0%
THANH HÓA	64.5%	12.9%	9.7%	3.2%
VĨNH PHÚC	8.3%	79.2%	12.5%	0.0%
YÊN BÁI	33.3%	33.3%	33.3%	0.0%



<u>Figure 10.</u> Geographic distribution of hand, foot, and mouth disease cases associated with infection of human enterovirus 71, coxsackievirus A6 and coxsackievirus A6 in Northern Vietnam, 2011.

After circulation of EV71 is CAV 6 and CAV 16. There are difference for circulating of serotypes of Coxsackievirus-A in the northern provinces, some provinces with only circulation of Coxsackievirus-A6 (Ha Tinh, Thai Nguyen, Ha Giang, Hoa Binh, Tuyen Quang), and some provinces have only Coxsackievirus A16 (Cao Bang, Dien Bien, Lai Chau, Lang Son and Lao Cai), the remaining provinces have co-circulation of both Coxsackievirus-A6 and CAV16

Some provinces detected Coxsackievirus A3 (Bac Giang), Coxsackievirus A10 (Bac Giang, Lao Cai, Hai Duong, Phu Tho), Coxsackievirus A12 (Bac Giang, Phu Tho, Hanoi, Hai Duong, Quang Ninh), Coxsackievirus A13 (Tuyen Quang), echovirus type 30 in Phu Tho and

Ninh Binh, Ha Giang, Bac Ninh; especially with appearance of enterovirus type 96 in Tuyen Quang province.

Table 7. Geographic distribution of hand, foot, and mouth disease cases by EV71

subgenogroup

IT	PROVINCE	N - EV71	C4	C5	B5
1	BẮC GIANG	2		1	1
2	BẮC NINH	5	1		4
3	HÀ GIANG	4	4		
4	HÀ NAM	1			1
5	HÀ NỘI	6	6		
6	HẢI PHÒNG	6	6		
7	HOÀ BÌNH	17	14		3
8	SON LA	1	1		
9	THANH HOÁ	5	4	1	
	TT	47	36	2	9

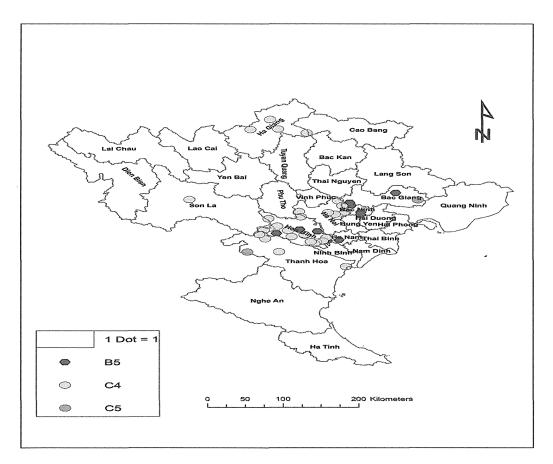


Figure 11. Geographic distribution of hand, foot, and mouth disease cases associated with infection of subgenogroup/HEV-71, in Northern Vietnam, 2011.

46 EV71 from 9 provinces in the north of Viet Nam were sequenced and analyzed. Results showed, they belonged to 2 genogroups with 3 subgenogroups B5, C4 and C5. Bac Ninh and Hoa Binh province had co-circulation of 2 genogroups B and C with subgenogroup B5 and C4; Bac Giang province had also co-circulation of 2 genogroups B and C with subgenogroup B5 and C5; Thanh Hoa province had circulation of genogroup C with 2 subgenogroups C4 and C5

6.20 DISCUSTION

6.20.1 Epidemiology of HFMD

HFMD was identified in northern Vietnam in 2011; HEV71, CVA16 and CA6 were also identified throughout the year, EV71 was the predominant virus during this time, accounting for 46% (275 cases) of HFMD compared to 54% (328 cases) for all other enteroviruses (table 1). After EV71 were CA6 and CA16, they were also the predominant viruses during this time, accounting for 53.9 % of HFMD compared to 31.3% for CVA6 and CA16 respectively (table 2).

Enteroviruses have circulated and caused HFMD outbreak in 2011 at all 28 northern provinces (table 5 and figgue 9)

6.20.2 Molecular Epidemiology of HEV71

Phylogenetic analysis based on nucleotide sequence alignment of 18 representative strains with the complete VP1 gene and 28 PCR products from clinical samples of HEV71 at 9 provinces of northern Vietnam. Result showed 2 genogroups that belonged to genogroup C with 2 subgenogroups C4, C5 and genogroup B with subgenogroup B5. Predominant EV71 strain in 2011 identified as subgenotype C4 during the HFMD outbreak of year and after subgenotype C4 was subgenogroup B5. Subgenogroup C5 was sporadic

Genogroup C with subgenogroup C4 and C5 have emerged recently in Southeast Asia, Viruses belonging to subgenogroup C4 were first identified in the People's Republic of China in 1998 and again in 2000 before their identification in northern Vietnam in 2003 (only one case), Furthermore, a new subgenogroup C5, circulated in northern Vietnam throughout 2008-2010 but decreased in 2011 and subgenogroup C4 reappeared and replaced for subgenogroup C5, and a new genogroup (genoproup B) with subgenogroup B5 appeared and caused HFMD in 2011

With evidence of the ongoing evolution of new subgenogroup and appearance of new genogroup similar to that observed for genogroup B HEV71 strains in Southeast Asia . Furthermore, the year-round detection and circulation of multiple independent genetic lineages of HEV71 suggested that this virus have circulated endemically within the human population of northern Vietnam.

平成24年度業績

*研究成果の刊行に関する一覧表

*学会発表一覧表

研究成果の刊行に関する一覧表 (平成24年度)

執筆者氏名	刊行書籍又は雑誌名 (雑誌のときは雑誌名、 巻号数、論文名)	刊行書店名	巻名	ページ	刊行年
プロジェクト1:中国					
Burns CC, Shaw J, Jorba J, Bukbuk D, Adu F, Gumede N, Pate MA, Abanida EA, Gasasira A, Iber J, Chen Q, Vincent A, Chenoweth P, Henderson E, Wannemuehler K, Naeem A, Umami RN, Nishimura Y, Shimizu H, Baba M, Adeniji A, Williams AJ, Kilpatrick DR, Oberste MS, Wassilak SG, Tomori O, Pallansch MA Kew O.	Multiple Independent Emergences of Type 2 Vaccine-Derived Polioviruses during a Large Outbreak in northern Nigeria.	J Virol		(in press)	
Fukuhara M, Iwami S, Sato K, Nishimura Y, Shimizu H, Aihara K, Koyanagi Y.	Quantification of the dynamics of enterovirus 71 infection by experimental-mathematical investigation.	J Virol	87	701-705	2012
Nishimura Y, Shimizu H.	Cellular receptors for human enterovirus species a.	Front Microbiol	3	105	2012
Nakajima N, Kitamori Y, Ohnaka S, Mitoma Y, Mizuta K, Wakita T, Shimizu H, Arita M.	Development of a transcription-reverse transcription concerted reaction method for specific detection of human enterovirus 71 from clinical specimen.	J Clin Microbiol	50	1764-1768	2012
Wong KT, Ng KY, Ong KC, Ng WF, Shankar SK, Mahadevan A, Radotra B, Su JI, Lau G, Ling AE, Chan KP, Macorelles P, Desai AS, Ravi V, Nagata N, Shimizu H, Takasaki T	Enterovirus 71 encephalomyelitis and Japanese encephalitis can be distinguished by topographic distribution of inflammation and specific intraneuronal detection of viral antigen and RNA in the central nervous system.	Neuropathology and Applied Neurobiology	38	443-453	2012
Fujimoto T, Iizuka S, Enomoto M, Abe K, Yamashita K, Hanaoka N,	Hand, Foot, and Mouth Disease Caused by Coxsackievirus A6,	Emerg Infect Dis	18	337-339	2012

Okabe N, Yoshida H, Yasui Y,	Japan, 2011.				
Kobayashi M, Fujii Y, Tanaka H,	- Capazi, 2011				
Yamamoto M, Shimizu H					
Sun Q, Zhang Y, Zhu S, Cui H, Tian H, Yan D, Huang G, Zhu Z, Wang D, Li X, Jiang H, An H Xu W.	Complete genome sequence of two coxsackievirus A1 strains that were cytotoxic to human rhabdomyosarcoma cells.	J Virol	86	10228 -10229	2012
清水博之	東アジア地域を中心とした手足 口病流行の現状	感染症		(印刷中)	2013
清水博之	手足口病、特集「感染症動向 2013」	メディカル朝 日	1	28-30	2012
清水博之	手足口病の問題点	小児科	53	751-758	2012
增本久人、南亮仁、野田日登美、 江口正宏、古川義朗、鶴田清典、 中田恵子、左近(田中)直美、山 崎謙治、高尾信一、Tao Zexin, Xu Aiqiang, Zhang Yong, Xu Wenbo, 藤本嗣人、花岡希、小長谷昌未、 吉田弘、清水博之.	国内外における手足口病流行に 関与するコクサッキーウイルス A6型の遺伝子解析.	病原微生物検 出情報	33	60-61	2012
藤本嗣人,花岡希, 小長谷昌未, 岡部信彦, 榎本美貴, 小林正明, 吉田弘, 清水博之.	2011 年に手足口病患者から検出 されたコクサッキーウイルス A6 型の遺伝子配列.	病原微生物検出情報	33	61-62	2012
武知茉莉亜,乾未来,福島若葉,中野貴司,清水博之.	手足口病・ヘルパンギーナおよび 関連合併症の入院症例に関する 全国調査(2010年分)—中間集 計結果.	病原微生物検 出情報	33	63-64	2012
清水博之	手足口病 (エンテロウイルス 71) ワクチン開発の現状.	病原微生物検 出情報	33	65-66	2012
Shimizu H (分担執筆)	A Guide to Clinical management and Public Health Response for Hand Foot Mouth Disease (HFMD)	WHO report			2011
Amemura-Maekawa J, Kikukawa K, Helbig JH, Kaneko S, Suzuki-Hashimoto A, Furuhata K, Chang B, Murai M, Ichinose M, Ohnishi M, Kura F	Working Group for <i>Legionella</i> in Japan. 2012. Distribution of monoclonal antibody subgroups and sequence-based types among <i>Legionella pneumophila</i> serogroup 1 isolates derived	Appl Environ Microbiol.	12	4263-70.	2012

	from cooling tower water,				
	bathwater, and soil in Japan.				
Tomohiro Oishi, Naruhiko Ishiwada,	the Japanese IPD Study Group.				
Kousaku Matsubara, Junichiro Nishi,	Low opsonic activity to the				
Bin Chang, Kazuyo Tamura,	infecting serotype in pediatric	Vaccine.	31	845–849	2013
Yukihiro Akeda, Toshiaki Ihara,	patients with invasive				
Moon H. Nahm, Kazunori Oishi,	pneumococcal disease.				
Sakai K, Nagata N, Ami Y, Seki F,					
Suzaki Y, Iwata-Yoshikawa N,					
Suzuki T, Fukushi S, Mizutani T,	Lethal Canine Distemper Virus				
Yoshikawa T, Otsuki N, Kurane I,	Outbreak in Cynomolgus	J Virol.	87(2)	1105-1114	2013
Komase K, Yamaguchi R,	Monkeys in Japan in 2008.				
Hasegawa H, Saijo M, Takeda M,					
Morikawa S.					
Neekun Sharma, Akitoyo Hotta,	Detection of Francisella				
Yoshie Yamamoto, Osamu Fujita,	tularensis-specific antibodies in	Clinical and			
Akihiko Uda, Shigeru Morikawa,	patients with tularemia using a	Vaccine		(in press)	2012
Akio Yamadaa, Kiyoshi	novel competitive enzyme-linked	Immunology			
Tanabayashia.	immunosorbent assay.				
Satoshi Taniguchi, Yusuke Sayama,					
Noriyo Nagata, Tetsuro Ikegami,					
Mary E Miranda, Shumpei	Analysis of the humoral immune				
Watanabe, Itoe Iizuka, Shuetsu	responses among cynomolgus	BMC	Oct 11		
Fukushi, Tetsuya Mizutani,	macaque naturally infected with	Veterinary	00011	189	2012
Yoshiyuki Ishii, Masayuki Saijo,	Reston virus during the 1996	Research,	8(1)		
Hiroomi Akashi, Yasuhiro	outbreak in the Philippines.				
Yoshikawa, Shigeru Kyuwa and					
Shigeru Morikawa.					
Shuetsu Fukushi, Hideki Tani,	Serological assays based on	Viruses			
Tomoki Yoshikawa, Masayuki	recombinant viral proteins for the	(special issue:	Oct 12	2097-114	2012
Saijo, Shigeru Morikawa.	diagnosis of arenavirus	(special issue. Arenaviruses)	4(10)	2097-114	2012
Saijo, Siligeru Worikawa.	hemorrhagic fevers.	Arenaviruses)			
Harutaka Katano, Seiichi Sato,					
Tsuyoshi Sekizuka, Akiko	Pathogenic characterization of a				
Kinumaki, Hitomi Fukumoto, Yuko	cervical lymph node derived from	Int J Clin Exp	5(8)	814-823	2012
Sato, Hideki Hasegawa, Shigeru	a patient with Kawasaki disease.	Pathol		017-023	2012
Morikawa, Masayuki Saijo,	a patient with ixawasaki disease.				
Tetsuya Mizutani, Makoto Kuroda.					

G VD VI CGV M					
Sayama Y, Demetria C, Saito M,					
Azul RR, Taniguchi S, Fukushi S,	A seroepidemiologic study of Reston ebolavirus in swine in the Philippines.	BMC Vet Res.	Jun 18 8	82	2012
Yoshikawa T, Iizuka I, Mizutani T,					
Kurane I, Malbas FF Jr, Lupisan S,					
Catbagan DP, Animas SB, Morales					
RG, Lopez EL, Dazo KR, Cruz					
MS, Olveda R, Saijo M, Oshitani					
H, Morikawa S.					
Lihoradova O, Kalveram B, Indran	The dominant-negative inhibition				
SV, Lokugamage N, Juelich TL,	of double-stranded	J Virol.	Jul 86(14)	7650-61	2012
Hill TE, Tseng CT, Gong B,	RNA-dependent protein kinase				
Fukushi S, Morikawa S, Freiberg	PKR increases the efficacy of Rift				
AN, Ikegami T.	Valley fever virus MP-12 vaccine.				
	Antigen-capture ELISA for the				
Fukushi S, Nakauchi M, Mizutani	detection of Rift Valley fever virus	J Virol	Mar	60.54	2012
T, Saijo M, Kurane I, Morikawa S.	nucleoprotein using new	Methods.	180(1-2)	68-74	2012
	monoclonal antibodies.				
Arai S, Gu SH, Baek LJ, Tabara K,	Divergent ancestral lineages of				
Bennett SN, Oh HS, Takada N,	newfound hantaviruses harbored by phylogenetically related crocidurine shrew species in	Virology.	Mar 15 424(2)	99-105	2012
Kang HJ, Tanaka-Taya K,					
Morikawa S, Okabe N, Yanagihara					
R, Song JW.	Korea.				
Kennedy JS, Gurwith M, Dekker					
CL, Frey SE, Edwards KM,	Safety and Immunogenicity of LC16m8, an Attenuated Smallpox Vaccine in Vaccinia-Naive Adults.	J Inf Dis	204(9)	1395-402	2011
Kenner J, Lock M, Empig C,					
Morikawa S, Saijo M, Yokote H,					
Karem K, Damon I, Perlroth M,					
and Greenberg RN.					
Shirato K, Maeda K, Tsuda S,					
Suzuki K, Watanabe S, Shimoda H,					
	Detection of het commerciances			·	
Ueda N, Iha K, Taniguchi S,	Detection of bat coronaviruses	Virus Genes	Feb	40-4	2012
Kyuwa S, Endoh D, Matsuyama S,	from Miniopterus fuliginosus in	virus Genes	44(1)	40-4	2012
Kurane I, Saijo M, Morikawa S,	Japan.				
Yoshikawa Y, Akashi H, Mizutani					
T.					
Nakayama, S., Tribuddharat, C.,	Molecular analyses of TEM genes	Antimicrob.			
Prombhul, S., Shimuta, K.,	and their corresponding	Agents	56	916-920	2012.
Srifuengfung, S., Unemo, M., and	penicillinase-producing Neisseria	Chemoter.			