

FIG 2 Correlation between IRRDR[HCV-4] sequence variations and treatment outcome. (A) Average number of amino acid mutations in IRRDR[HCV-4] obtained from SVR and non-SVR patients. (B) Alignment of IRRDR[HCV-4] sequences obtained from SVR and non-SVR patients with HCV-4a, -4m, -4n, and -4o. The consensus sequence (Cons) of each subtype is shown on the top. The numbers along the sequence indicate the amino acid positions. Dots indicate residues identical to those of the Cons sequence. The numbers of the mutations in each IRRDR (4a, 4m, 4n, or 4o) are shown on the right.

predictive factors of SVR for PEG-IFN/RBV treatment outcome in HCV-4 infection, first, all available data of baseline patients' parameters and IRRDR[HCV-4] polymorphism were entered in a univariate logistic analysis. This analysis yielded 3 factors that

were correlated or nearly correlated with SVR: IRRDR[HCV-4] ≥ 4 ( $P = 0.0004$ ), patient's age (<42 years;  $P = 0.03$ ), and HCV RNA titer (<5,200 IU/ml;  $P = 0.08$ ). Subsequently, these 3 factors were entered in multivariate logistic regression analysis. This anal-

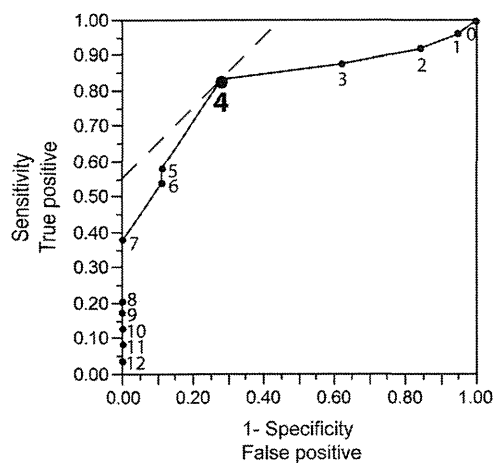


FIG 3 ROC curve analysis of IRRDR[HCV-4] sequence heterogeneity for SVR prediction. The solid line curve shows the AUC. Solid circles with numerals plotted on the curve represent different numbers of IRRDR mutations analyzed. The dashed line in the upper left corner indicates the optimal number of IRRDR[HCV-4] mutations for SVR prediction, which yields the highest sensitivity (84%) and the highest specificity (72%).

ysis revealed that the  $IRRDR[HCV-4] \geq 4$  was the only independent predictive factor for SVR in HCV-4 infection (Table 4). We then assessed SVR predictability by means of  $IRRDR[HCV-4] \geq 4$ . As shown in Table 5,  $IRRDR[HCV-4] \geq 4$  would predict SVR with a positive predictive value (PPV) of 81% ( $P = 0.002$ ) and sensitivity of 84%. On the other hand,  $IRRDR[HCV-4] \leq 3$  would predict non-SVR with a negative predictive value (NPV) of 76% ( $P = 0.02$ ) and specificity of 72%. Thus, the degree of sequence variation in IRRDR[HCV-4] would yield useful positive and negative predictive markers for PEG-IFN/RBV therapy outcome in HCV-4-infected patients.

## DISCUSSION

Both host and viral genetic factors have been implicated in influencing the clinical response to PEG-IFN/RBV therapy for HCV infection (22). It has recently been reported that host genetic polymorphisms near or within the IL28B gene on chromosome 19 show a critical impact on the treatment outcome of patients infected with HCV-1 (20, 37, 39). As for the viral factor(s), polymorphisms of NS5A and core regions of a given HCV genotype have been linked to a difference in SVR rates (3, 4, 13, 18, 30). This hypothesis was mostly inferred from studies carried out with Asian populations, in particular Japanese, with HCV-1b infection. However, whether it can be applied to non-Asian populations

infected with non-HCV-1 is still unknown. To the best of our knowledge, this is the first study that specifically examines the relationship between HCV genome heterogeneity, in particular in NS5A and core regions, and PEG-IFN/RBV treatment outcome in Egyptian patients infected with HCV-4. In analogy with our previous studies that identified IRRDR as a significant determinant for PEG-IFN/RBV treatment outcome in Japanese patients infected with HCV-1b, -2a, and -2b (12–16), we have demonstrated in the present study that sequence heterogeneity within IRRDR is closely associated with the ultimate treatment outcome in Egyptian patients infected with HCV-4. A high degree of sequence variation in IRRDR[HCV-4], i.e., more than 4 ( $IRRDR \geq 4$ ), significantly correlated with SVR, while a low degree of sequence variation in this region ( $IRRDR \leq 3$ ) correlated with non-SVR, null response, and relapse. The majority of patients with SVR (84%) had HCV with IRRDR of  $\geq 4$ . In contrast, nearly two-thirds (72%) of the patients with non-SVR had HCV with  $IRRDR \leq 3$  ( $P = 0.0004$ ) (Table 3). Notably, 21 of the 26 patients infected with HCV with  $IRRDR[HCV-4] \geq 4$  achieved SVR. Accordingly, the PPV and NPV of  $IRRDR[HCV-4] \geq 4$  for SVR and non-SVR patients were 81% ( $P = 0.002$ ) and 76% ( $P = 0.02$ ), respectively (Table 5). Our present results thus strongly suggest that the degree of sequence heterogeneity within IRRDR[HCV-4] would be a useful marker for prediction of treatment outcome in HCV-4 infection.

The molecular mechanism underlying the possible involvement of this region in IFN responsiveness of the virus is still unknown. The significant difference among IRRDR sequence patterns may suggest genetic flexibility of this region. Indeed, the C-terminal portion of NS5A was shown to tolerate sequence insertions and deletions (29). This flexibility might play an important role in modulating the interaction with various host systems, including IFN-induced antiviral machineries. It is also possible that the genetic flexibility of IRRDR is accompanied by compensatory changes elsewhere in the viral genome and that these compensatory changes affect overall viral fitness and responses to IFN-based therapy (8, 29, 41). Also, it is worth noting that IRRDR is among the most variable sequences across the different genotypes and subtypes of HCV (25) whereas its upstream and downstream sequences show a higher degree of sequence conservation (15). This may suggest that whereas the upstream and downstream sequences have a conserved function(s) across all the HCV genotypes, IRRDR sequences have a genotype-dependent or even a strain-dependent function(s).

A mutation at position 70 of the core protein of HCV-1b has been reported to be correlated with PEG-IFN/RBV treatment out-

TABLE 3 Correlation between NS5A sequence heterogeneity and virological responses in HCV-4 infection

Factor	No. of isolates/total no. (%)				P value for SVR versus:		
	SVR	Non-SVR	Null response	Relapse	Non-SVR	Null response	Relapse
$IRRDR \geq 4$	21/25 (84) <sup>a</sup>	5/18 (28)	4/13 (31)	1/5 (20)	0.0004	0.003	0.01
$IRRDR \leq 3$	4/25 (16)	13/18 (72) <sup>b</sup>	9/13 (69)	4/5 (80)			
$IRRDR \geq 5$	16/25 (64) <sup>a</sup>	2/18 (11)	1/13 (8)	1/5 (20)	0.0006	0.002	0.14
$IRRDR \leq 4$	9/25 (36)	16/18 (89) <sup>b</sup>	12/13 (92)	4/5 (80)			
$IRRDR \geq 3$	22/25 (88) <sup>a</sup>	11/18 (61)	10/13 (77)	1/5 (20)	0.066	0.39	0.006
$IRRDR \leq 2$	3/25 (12)	7/18 (39) <sup>b</sup>	3/13 (23)	4/5 (80)			

<sup>a</sup> Sensitivity (proportion of SVR patients with the favorable factor).

<sup>b</sup> Specificity (proportion of non-SVR patients with the unfavorable factor).

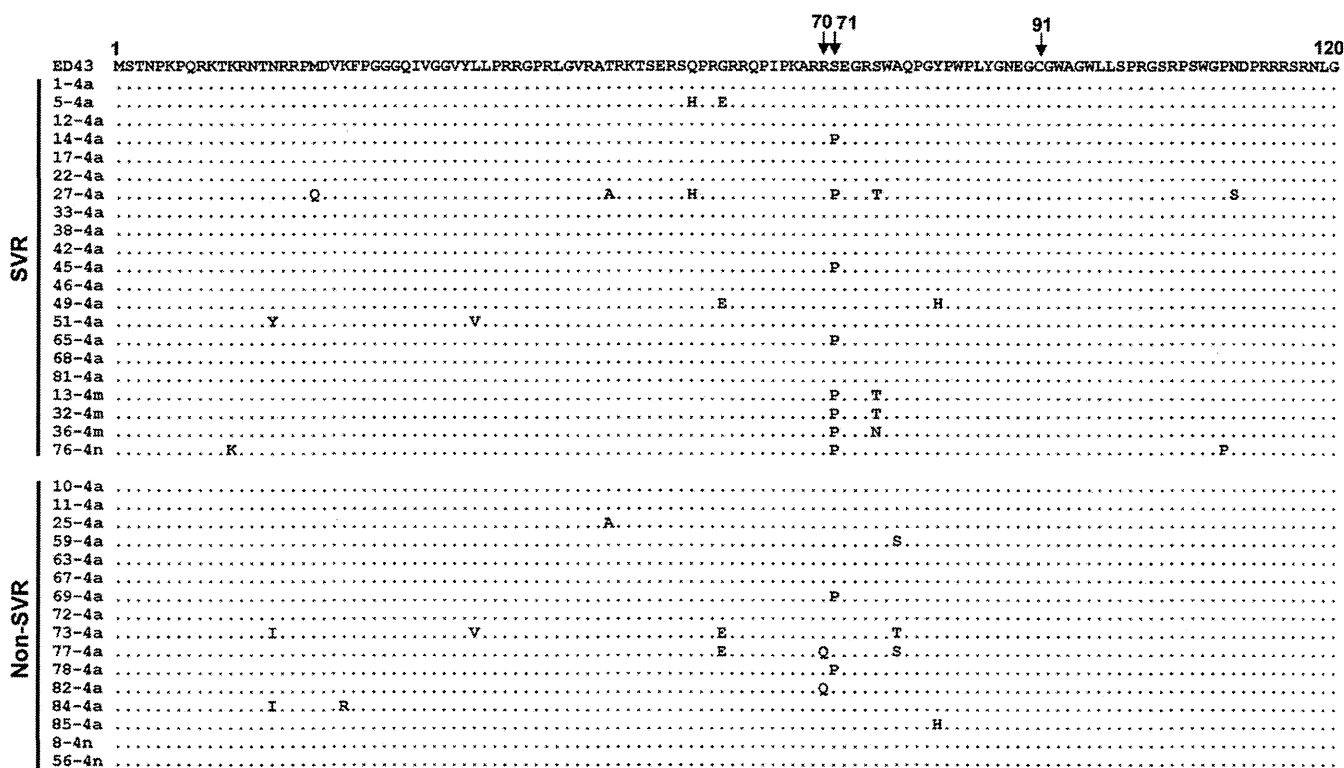


FIG 4 Sequence alignment of the core protein of HCV-4 isolates. Core protein sequences (aa 1 to 120) of HCV-4 obtained from SVR and non-SVR patients are aligned. The prototype sequence of ED43 (10) is shown on the top. The numbers along the sequence indicate the amino acid positions. Dots indicate residues identical to those of the prototype sequence.

come (4, 12). In the present study, however, we found no significant correlation between core protein polymorphism and treatment outcome in HCV-4 infection. The residue at position 70 of the core protein of all but two HCV-4 isolates analyzed in this study was Arg (Fig. 4), which is known to be associated with SVR in HCV-1b infection (4, 12). This high degree of sequence conservation at position 70 might be the reason for the lack of significant correlation between core protein polymorphism and treatment outcome in HCV-4 infection.

Single nucleotide polymorphisms (SNPs) near the IL28B region have been identified as the strongest baseline predictors of SVR to PEG-IFN/RBV in patients with HCV-1 infection. More recently, in two major studies that were carried out exclusively with HCV-4-infected patients (9, 11), the CC genotype of rs12979860 IL28B SNP was also strongly associated with SVR. It is worth noting that although the SVR rate was more than 80%

among the patients with the CC genotype, these patients represented only around 40% of total SVR cases in both studies. Furthermore, the CC genotype was found in only 34% of all Egyptian patients analyzed (9). Taken together, those observations support the idea that in addition to IL28B polymorphism, there should be an additional factor(s) that influences SVR. In this context, an interplay between IRRDR and IL28B polymorphisms might explain why some patients with undesirable IL28B genotype achieve SVR and why some patients infected with HCV isolates with IRRDR[HCV-4] ≥ 4 do not achieve SVR. Further comprehensive study is needed to validate the importance of IRRDR and IL28B polymorphisms in predicting the treatment outcome of HCV-4-infected patients.

In conclusion, the present study emphasizes the importance of IRRDR sequence heterogeneity in the prediction of PEG-IFN/RBV treatment outcome for different HCV genotype infections in

TABLE 4 Univariate and multivariate analyses for identification of independent predictive factors for SVR in HCV-4-infected patients treated with PEG-IFN/RBV therapy

Univariate analysis		Multivariate analysis	
Variable	P value	Odds ratio (95% CI)	P value
IRRDR mutations (IRRDR ≥ 4 versus IRRDR ≤ 3)	0.0004	10.5 (1.12–98.91)	0.04
Age (<42 years)	0.03		
HCV-RNA (<5,200 IU/ml)	0.08		

TABLE 5 PPV, NPV, sensitivity, and specificity of IRRDR sequence heterogeneity on the likelihood of achieving SVR and non-SVR in HCV-4 infection

Factor	PPV	NPV	Sensitivity <sup>c</sup>	Specificity <sup>d</sup>
IRRDR ≥ 4	81% (21/26) <sup>a</sup>		84% (21/25)	
IRRDR ≤ 3		76% (13/17) <sup>b</sup>		72% (13/18)

<sup>a</sup> P = 0.002.

<sup>b</sup> P = 0.02.

<sup>c</sup> Proportion of SVR patients who were infected with HCV isolates with IRRDR of ≥4.

<sup>d</sup> Proportion of non-SVR patients who were infected with HCV isolates with IRRDR of ≤3.

different ethnic groups, including Egyptian patients infected with HCV-4.

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#### REFERENCES

- Abdel-Aziz F, et al. 2000. Hepatitis C virus (HCV) infection in a community in the Nile Delta: population description and HCV prevalence. *Hepatology* 32:111–115.
- Abdel-Hamid M, et al. 2007. Genetic diversity in hepatitis C virus in Egypt and possible association with hepatocellular carcinoma. *J. Gen. Virol.* 88:1526–1531.
- Akuta N, et al. 2009. Association of amino acid substitution pattern in core protein of hepatitis C virus genotype 2a high viral load and virological response to interferon-ribavirin combination therapy. *Intervirology* 52:301–309.
- Akuta N, et al. 2007. Predictive factors of early and sustained responses to peginterferon plus ribavirin combination therapy in Japanese patients infected with hepatitis C virus genotype 1b: amino acid substitutions in the core region and low-density lipoprotein cholesterol levels. *J. Hepatol.* 46:403–410.
- Akuta N, et al. 2007. Prediction of response to pegylated interferon and ribavirin in hepatitis C by polymorphisms in the viral core protein and very early dynamics of viremia. *Intervirology* 50:361–368.
- Akuta N, et al. 2005. Association of amino acid substitution pattern in core protein of hepatitis C virus genotype 1b high viral load and non-virological response to interferon-ribavirin combination therapy. *Intervirology* 48:372–380.
- Amoroso P, et al. 1998. Correlation between virus genotype and chronicity rate in acute hepatitis C. *J. Hepatol.* 28:939–944.
- Appel N, Pietschmann T, Bartenschlager R. 2005. Mutational analysis of hepatitis C virus nonstructural protein 5A: potential role of differential phosphorylation in RNA replication and identification of a genetically flexible domain. *J. Virol.* 79:3187–3194.
- Asselah T, et al. 2012. IL28B polymorphism is associated with treatment response in patients with genotype 4 chronic hepatitis C. *J. Hepatol.* 56:527–532.
- Chamberlain RW, Adams N, Saeed AA, Simmonds P, Elliott RM. 1997. Complete nucleotide sequence of a type 4 hepatitis C virus variant, the predominant genotype in the Middle East. *J. Gen. Virol.* 78(Pt 6):1341–1347.
- De Nicola S, et al. 2012. Interleukin 28B polymorphism predicts pegylated interferon plus ribavirin treatment outcome in chronic hepatitis C genotype 4. *Hepatology* 55:336–342.
- El-Shamy A, et al. 2012. Polymorphisms of hepatitis C virus non-structural protein 5A and core protein and clinical outcome of pegylated-interferon/ribavirin combination therapy. *Intervirology* 55:1–11.
- El-Shamy A, et al. 2008. Sequence variation in hepatitis C virus nonstructural protein 5A predicts clinical outcome of pegylated interferon/ribavirin combination therapy. *Hepatology* 48:38–47.
- El-Shamy A, et al. 2007. Prediction of efficient virological response to pegylated interferon/ribavirin combination therapy by NS5A sequences of hepatitis C virus and anti-NS5A antibodies in pre-treatment sera. *Microbiol. Immunol.* 51:471–482.
- El-Shamy A, et al. 2012. Sequence heterogeneity in NS5A of hepatitis C virus genotypes 2a and 2b and clinical outcome of pegylated-interferon/ribavirin therapy. *PLoS One* 7:e30513. doi:10.1371/journal.pone.0030513.
- El-Shamy A, et al. 2011. Sequence heterogeneity of NS5A and core proteins of hepatitis C virus and virological responses to pegylated-interferon/ribavirin combination therapy. *Microbiol. Immunol.* 55:418–426.
- el-Zayadi AR, et al. 2005. Hepatocellular carcinoma in Egypt: a single center study over a decade. *World J. Gastroenterol.* 11:5193–5198.
- Enomoto N, et al. 1996. Mutations in the nonstructural protein 5A gene and response to interferon in patients with chronic hepatitis C virus 1b infection. *N. Engl. J. Med.* 334:77–81.
- Fried MW, et al. 2002. Peginterferon alfa-2a plus ribavirin for chronic hepatitis C virus infection. *N. Engl. J. Med.* 347:975–982.
- Ge D, et al. 2009. Genetic variation in IL28B predicts hepatitis C treatment-induced viral clearance. *Nature* 461:399–401.
- Hassan MM, et al. 2001. The role of hepatitis C in hepatocellular carcinoma: a case control study among Egyptian patients. *J. Clin. Gastroenterol.* 33:123–126.
- Kau A, Vermehren J, Sarrazin C. 2008. Treatment predictors of a sustained virologic response in hepatitis B and C. *J. Hepatol.* 49:634–651.
- Khatab MA, et al. 2011. Management of hepatitis C virus genotype 4: recommendations of an international expert panel. *J. Hepatol.* 54:1250–1262.
- Limaye AR, Draganov PV, Cabrera R. 2011. Boceprevir for chronic HCV genotype 1 infection. *N. Engl. J. Med.* 365:176, 177–178.
- Macdonald A, Harris M. 2004. Hepatitis C virus NS5A: tales of a promiscuous protein. *J. Gen. Virol.* 85:2485–2502.
- Maekawa S, Enomoto N. 2009. Viral factors influencing the response to the combination therapy of peginterferon plus ribavirin in chronic hepatitis C. *J. Gastroenterol.* 44:1009–1015.
- Mattsson L, Sonnerborg A, Weiland O. 1993. Outcome of acute symptomatic non-A, non-B hepatitis: a 13-year follow-up study of hepatitis C virus markers. *Liver* 13:274–278.
- Micallef JM, Kaldor JM, Dore GJ. 2006. Spontaneous viral clearance following acute hepatitis C infection: a systematic review of longitudinal studies. *J. Viral Hepat.* 13:34–41.
- Moradpour D, et al. 2004. Insertion of green fluorescent protein into nonstructural protein 5A allows direct visualization of functional hepatitis C virus replication complexes. *J. Virol.* 78:7400–7409.
- Murakami T, et al. 1999. Mutations in nonstructural protein 5A gene and response to interferon in hepatitis C virus genotype 2 infection. *Hepatology* 30:1045–1053.
- Okamoto H, et al. 1992. Typing hepatitis C virus by polymerase chain reaction with type-specific primers: application to clinical surveys and tracing infectious sources. *J. Gen. Virol.* 73(Pt 3):673–679.
- Ray SC, Arthur RR, Carella A, Bukh J, Thomas DL. 2000. Genetic epidemiology of hepatitis C virus throughout Egypt. *J. Infect. Dis.* 182:698–707.
- Sarasin-Filipowicz M. 2010. Interferon therapy of hepatitis C: molecular insights into success and failure. *Swiss Med. Wkly.* 140:3–11.
- Sherman KE, et al. 2011. Response-guided telaprevir combination treatment for hepatitis C virus infection. *N. Engl. J. Med.* 365:1014–1024.
- Simmonds P, et al. 2005. Consensus proposals for a unified system of nomenclature of hepatitis C virus genotypes. *Hepatology* 42:962–973.
- Simmonds P, et al. 1993. Classification of hepatitis C virus into six major genotypes and a series of subtypes by phylogenetic analysis of the NS-5 region. *J. Gen. Virol.* 74(Pt 11):2391–2399.
- Suppiah V, et al. 2009. IL28B is associated with response to chronic hepatitis C interferon-alpha and ribavirin therapy. *Nat. Genet.* 41:1100–1104.
- Tanaka E, Kiyosawa K. 2000. Natural history of acute hepatitis C. *J. Gastroenterol. Hepatol.* 15(Suppl):E97–E104.
- Tanaka Y, et al. 2009. Genome-wide association of IL28B with response to pegylated interferon-alpha and ribavirin therapy for chronic hepatitis C. *Nat. Genet.* 41:1105–1109.
- Timm J, et al. 2007. Characterization of full-length hepatitis C virus genotype 4 sequences. *J. Viral Hepat.* 14:330–337.
- Yuan HJ, Jain M, Snow KK, Gale M, Jr, Lee WM. 2010. Evolution of hepatitis C virus NS5A region in breakthrough patients during pegylated interferon and ribavirin therapy. *J. Viral Hepat.* 17:208–216.

