

Figure 1. Association between the genetic variant of *AKT1* and memory performance measured by the Wechsler Memory Scale-Revised (WMS-R). The memory performance was measured using five domain scores, i.e. attention/concentration, verbal memory, visual memory, delayed recall, and general memory, of the WMS-R. Significant effects of diagnosis and genotype were observed. No diagnosis-genotype interaction was found. Memory performance in patients with schizophrenia was lower than in healthy subjects. A carriers showed lower memory performance compared to individuals with the G/G genotype. Error bars represent the standard error.

attention/concentration performance, as compared with the homozygous G subjects.

Association between the genetic variant of *AKT1* and attentional performance measured by the CPT

The attention/concentration index of the WMS-R measures a complex attentional function comprising three subscales of mental control, digit span and visual memory span (Wechsler 1987), while the CPT-IP assesses vigilance/sustained attention in the attentional function. To evaluate the association between SNP5 and attentional function using a different assessment tool for attention, we examined the association using CPT-IP. There was no difference in demographic variables between *AKT1* genotype groups (Table II). The attention/concentration index of the WMS-R had significantly positive correlations with the attentional indices measured by the CPT in healthy sub-samples with an overlap ($D'2$: $r=0.24$, $df=119$, $P=0.0068$, $D'3$: $r=0.35$, $df=119$, $P=8.23 \times 10^{-5}$, and $D'4$: $r=0.35$, $df=119$, $P=1.02 \times 10^{-4}$). Because $D'2$, $D'3$ and $D'4$ of the CPT cannot simply be combined and were correlated

with each other, each D' was included in the MANCOVA analysis as dependent variables, and diagnosis and genotype statuses were included in the analysis as independent variables. Two-way MANCOVA revealed significant effects of diagnosis ($F_{1,172} = 11.44$, $P=7.11 \times 10^{-7}$), genotype ($F_{1,172} = 3.71$, $P=0.013$) and their interaction ($F_{1,172} = 4.01$, $P=0.0086$) on attentional performance (Table V and Figure 2). As a genotype–diagnosis interaction was found, we analyzed the genotype effect on attentional performance in patients and controls separately. The attentional performance of the A carriers was significantly lower than that of homozygous G subjects in patients with schizophrenia ($F_{1,53} = 4.49$, $P=0.0070$), while there was no significant genotype effect in the healthy controls ($P=0.38$). Post hoc two-way ANCOVA revealed significant effects of diagnosis ($D'2$: $F_{1,174} = 15.35$, $P=1.28 \times 10^{-4}$, $D'3$: $F_{1,174} = 28.01$, $P=3.60 \times 10^{-7}$, and $D'4$: $F_{1,174} = 29.23$, $P=2.10 \times 10^{-7}$), genotype ($D'2$: $F_{1,174} = 8.99$, $P=0.0031$ and $D'3$: $F_{1,174} = 4.76$, $P=0.030$) and their interaction ($D'2$: $F_{1,174} = 11.14$, $P=0.0010$) (Figure 2). All three scores in patients with schizophrenia were significantly lower than those in healthy controls. A carriers had lower

Table V. Association of a genetic variant of *AKT1* with the scores of CPT.

	Schizophrenia (<i>N</i> = 60)		Control (<i>N</i> = 121)		<i>P</i> values ($F_{1,174}$ values)		
	G/G (<i>N</i> = 35)	A carriers (<i>N</i> = 25)	G/G (<i>N</i> = 57)	A carriers (<i>N</i> = 64)	Diagnosis effect	Genotype effect	Interaction
MANCOVA					7.11×10^{-7} (11.44)	0.013 (3.71)	0.0086 (4.01)
<i>D</i> '2	3.54 ± 0.67	3.00 ± 0.74	3.67 ± 0.47	3.70 ± 0.47	1.28×10^{-4} (15.35)	0.0031 (8.99)	0.0010 (11.14)
<i>D</i> '3	2.87 ± 0.85	2.51 ± 0.82	3.46 ± 0.67	3.31 ± 0.65	3.60×10^{-7} (28.01)	0.030 (4.76)	0.36 (0.86)
<i>D</i> '4	1.56 ± 0.96	1.33 ± 0.76	2.31 ± 0.90	2.30 ± 0.87	2.10×10^{-7} (29.23)	0.47 (0.53)	0.47 (0.53)

MANCOVA: multivariate analysis of covariance. Means ± SD and *P* values are shown. Significant *P* values are indicated in bold.

scores for *D*'2 and *D*'3 than homozygous G subjects (Figure 2). As a genotype–diagnosis interaction was found only in *D*'2, we analyzed the effects of genotype on *D*'2 score in patients and controls separately. The *D*'2 score of the A carriers was significantly lower than that of homozygous G subjects in patients with schizophrenia ($F_{1,55} = 13.51$, $P = 5.39 \times 10^{-4}$); by contrast, there was no significant genotype effect in the healthy controls ($F_{1,116} = 0.24$, $P = 0.63$) (Figure 2). No interaction was found for *D*'3 or *D*'4 ($P = 0.36$, 0.47, respectively). The *D*'2 score of the A carriers was still significantly lower than the score of homozygous G subjects in patients with schizophrenia even after adjusting scores of positive and negative symptoms in PANSS and the chlorpromazine equivalent of total antipsychotics as covariates ($F_{1,52} = 14.09$, $P = 4.41 \times 10^{-4}$). These results suggest that variation in the *AKT1* gene could be related to deficits of performance in sustained attention and vigilance of attention in patients with schizophrenia.

Effect of the *AKT1* polymorphism on brain structure

We first performed an exploratory whole brain analysis to investigate the effects of diagnosis, genotype

and their interaction on gray matter volumes. The only difference in demographic variables between the *AKT1* genotype groups was for years of education in patients (Table III). We found significant effects of diagnosis and diagnosis–genotype interaction (uncorrected $P < 0.001$), while we did not find any genotype effect for all subjects (uncorrected $P > 0.001$). Patients with schizophrenia showed smaller gray matter volumes compared with controls, mainly in the frontal lobe and the temporal lobe (data not shown), which was consistent with previous studies (Chan et al. 2009; Ellison-Wright and Bullmore 2010). Significant diagnosis–genotype interactions were gray matter volumes in the right inferior parietal lobule, the right superior frontal gyrus, the left superior temporal gyrus and the left caudate (Table VI).

It has been suggested that three regions, the right inferior parietal lobule, the thalamus and the anterior cingulate gyrus, are involved in attentional processes (Salgado-Pineda et al. 2004). These three regions have been smaller volumes in patients with schizophrenia compared with controls and have been associated with functional deficits of attention during CPT-IP. In addition, *AKT1* SNP4 has been associated with reduced gray matter volumes in the bilateral

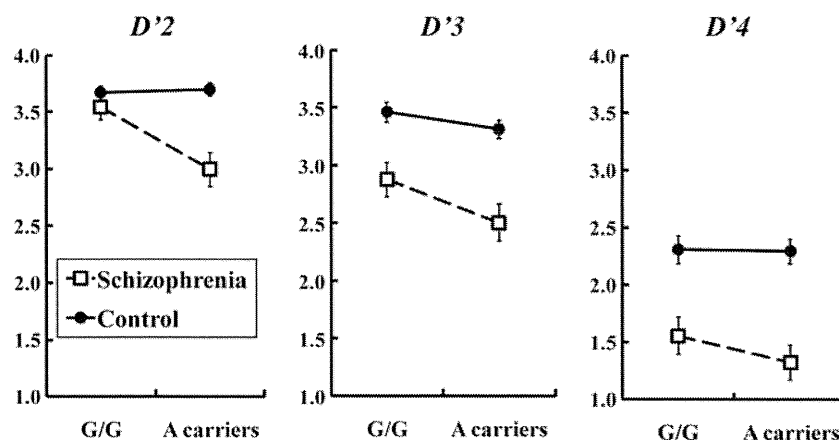


Figure 2. Association between the genetic variant of *AKT1* and attentional performance measured by the continuous performance test (CPT). Attentional performance was measured by three different conditions, i.e. *D*'2, *D*'3, and *D*'4, of the CPT. Significant effects of diagnosis, genotype and their interaction were observed. The genotype effect was observed in patients with schizophrenia, but not in healthy subjects. Patients with schizophrenia showed lower attentional performance than healthy subjects. Patients with A allele showed lower attentional performance than patients with G/G genotype. Error bars represent the standard error.

Table VI. Regions of the diagnosis-*AKT1* genotype interaction on brain morphology in an exploratory whole brain analysis (uncorrected $P < 0.001$, cluster size > 180).

Brain regions	R/L	BA	Cluster Size	T_{206} values	Talairach coordinates		
					x	y	z
<i>AKT1</i> genotype–diagnosis interaction							
Frontal lobe							
Superior frontal gyrus	R	11	249	4.27	11	49	–18
Temporal lobe							
Superior temporal gyrus	L	41	390	4.00	–51	–20	3
Superior temporal gyrus	L	22	408	3.64	–54	–37	13
Parietal lobe							
Inferior parietal lobule	R	40	193	4.06	55	–27	35
Subcortical							
Caudate	L	–	211	3.71	–11	3	5

R, right; L, left; BA, Brodmann area.

caudate and right prefrontal cortex (Tan et al. 2008b) and SNP2 has been associated with decreased gray matter density in the medial, dorsolateral, and inferior prefrontal cortex (Pietilainen et al. 2009). Thus, we hypothesized that the variation in *AKT1* could be associated three regions associated with attentional function or/and the caudate and prefrontal cortex. Based on the priori hypothesis, we applied *FWE* correction to gray matter regions of interest in the right inferior parietal lobule (*FWE*-corrected $P = 0.006$, Figure 3A), the right superior frontal gyrus (*FWE*-corrected $P = 0.003$, Figure 3B), and the left caudate (*FWE*-corrected $P = 0.017$, Figure 3C).

We next focused on the three regions of diagnosis–genotype interaction based on the priori hypothesis. Two-way ANOVA revealed significant diagnosis–genotype interactions in the extracted region from the center of the right inferior parietal lobule (55, –27, 35; $F_{1,210} = 14.63$, $P = 1.72 \times 10^{-4}$), the right superior frontal gyrus (11, 49, –18; $F_{1,210} = 16.40$, $P = 7.21 \times 10^{-5}$) and the left caudate (–11, 3, 5; $F_{1,210} = 11.44$, $P = 8.56 \times 10^{-4}$). As the genotype–diagnosis interactions were found, we analyzed the effects of genotype on these regions in patients and controls separately (Figure 3D–F). In patients with schizophrenia, A carriers had smaller gray matter volumes in these regions than homozygous G subjects, respectively (the right inferior parietal lobule; $F_{1,53} = 12.26$, $P = 9.46 \times 10^{-4}$, the right superior frontal gyrus; $F_{1,53} = 16.19$, $P = 1.83 \times 10^{-4}$ and the left caudate; $F_{1,53} = 8.69$, $P = 0.0048$). In contrast, in controls, homozygous G subjects had marginally smaller gray matter volumes in these regions than A carriers (the right inferior parietal lobule; $F_{1,157} = 5.45$, $P = 0.021$, the right superior frontal gyrus; $F_{1,157} = 3.88$, $P = 0.051$ and the left caudate; $F_{1,157} = 3.49$, $P = 0.064$). Even if handedness, which was a confounding factor, was co-varied in these VOI analyses, these effects of the variation in *AKT1*

on the extracted VOI remained significant. These data suggest that *AKT1* SNP5 might be associated with several brain morphological vulnerabilities in patients with schizophrenia, and that the smaller volume of the right inferior parietal lobule might be associated with structural vulnerability for attentional performance in schizophrenia, as the volume of the inferior parietal lobule has been positively correlated with attentional performance in patients with schizophrenia (Salgado-Pineda et al. 2004).

Discussion

We evaluated the association between SNP5 in *AKT1* and attentional performance and brain structure in a Japanese population. We examined this particular polymorphism after it was associated with schizophrenia in Asian populations in a recent meta-analysis (Shi et al. 2008) and after it was shown to predict treatment response to risperidone in Japanese patients with schizophrenia (Ikeda et al. 2008). We found that A carriers of SNP5, which is the risk allele for schizophrenia in Asian populations, performed worse on attentional performance, a capacity that is impaired in patients with schizophrenia compared with healthy subjects, than subjects with the G/G genotype as determined by WMS-R and CPT. In the CPT, the poorer performance in A carriers was found in the $D'2$ and the $D'3$. In the $D'2$ condition, the poorer attentional performance in A carriers was found only in the patients with schizophrenia, but not in controls. There was no association between the genotype and performance in the $D'4$ condition. The difficulty of the three processing-load conditions ($D'2$, $D'3$, and $D'4$) is different, with the $D'2$ condition being simpler than the others, suggesting that the lower CPT load ($D'2$) might be associated with a simpler attentional function than the higher CPT loads ($D'3$ and $D'4$). It is expected to be worsening

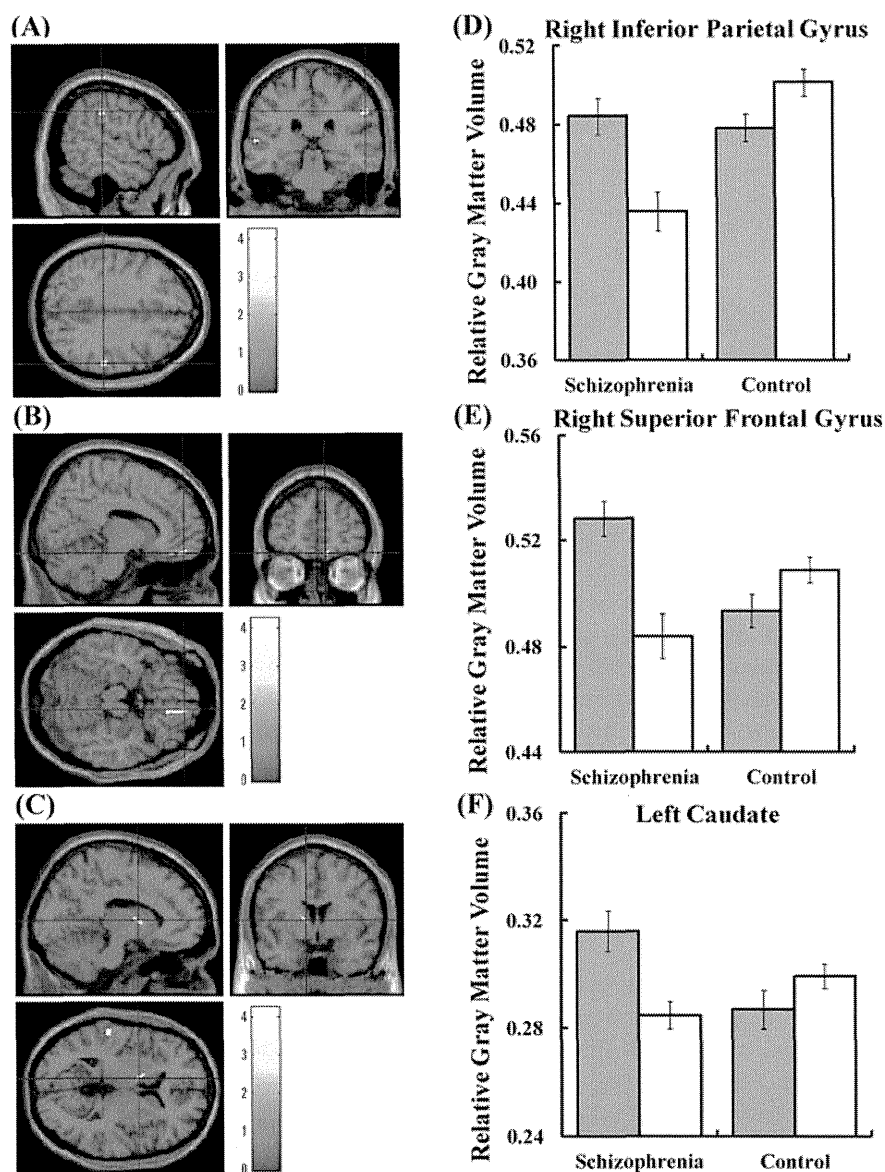


Figure 3. Impact of diagnosis-*AKT1* genotype interaction on gray matter volumes. (A–C) Anatomical localizations are displayed on coronal, sagittal, and axial sections of a normal MRI spatially normalized into the Montreal Neurological Institute template (uncorrected $P < 0.001$, cluster size > 180). Significant clusters of diagnosis-genotype interaction was in the right inferior parietal lobule (Talairach coordinates; 55, -27, 35, **A**), in the right superior frontal gyrus (11, 49, -18, **B**) and in the left caudate (-11, 3, 5, **C**). The regions are shown as cross-hairline. The colour bars show t values corresponding to the colour in the figures. (D–F) Each column shows relative gray matter volumes extracted from the right inferior parietal lobule (55, -27, 35, **D**), right superior frontal gyrus (11, 49, -18, **E**) and left caudate (-11, 3, 5, **F**). Gray bars represent individuals with G/G genotype, while white bars represent A carriers. Error bars represent the standard error.

performance with increasing task difficulty and to increase the difference of the performance between cases and controls. The reason why the *AKT1* genotype was not associated with the D^4 condition might be considered a floor effect in the CPT condition because of difficulty to achieve the D^4 condition in patients and controls. On the other hand, the reason why the genotype was not associated with the D^2 in controls might be considered a ceiling effect in the CPT condition because of ease to achieve the D^2 condition in healthy subjects. The attention/concentration index of the WMS-R assesses a com-

plex attentional function comprising three subscales of mental control, digit span and visual memory span (Wechsler 1987). The correlations of the attention/concentration of the WMS-R with D^2 ($r = 0.24$), D^3 ($r = 0.35$) or D^4 ($r = 0.35$) were modest. These results require a careful interpretation because attentional functions in these measures may imply different cognitive processes. These results suggest that A carrier might have deficits in attentional performance and that the attentional deficits in A carrier might be detected by easier tasks in patients with schizophrenia.

We found that the gray matter volume of the right inferior parietal lobule in carriers of the A allele was significantly smaller than that in homozygous G subjects with schizophrenia. There was no genotype effect on the gray matter volumes in two other regions, the thalamus and the anterior cingulate gyrus, in patients with schizophrenia (data not shown), although it has been reported that these regions are also related to attentional function (Salgado-Pineda et al. 2004). The right inferior parietal lobule have been smaller volumes in patients with schizophrenia compared with controls and have been associated with functional deficits of attention during CPT-IP (Salgado-Pineda et al. 2004). The previous study measured stimuli of one-digit numbers as CPT-IP performance (*D'1*). The *D'1* is simpler condition than the *D'2*, although we did not measure stimuli of one-digit numbers. These data suggest that variation of the *AKT1* gene might be associated with the gray matter volume of the right inferior parietal lobule with structural vulnerability of simple attentional performance in schizophrenia. To reinforce assumption of relevance of inferior parietal lobule for attentional processing and to investigate potential genotype effects on such relationship, we performed a preliminary investigation for gray matter volume-behavioural correlations in the overlapping sub-samples. Although the gray matter volume of the right inferior parietal lobule had marginal positive correlations with attentional performances, the correlations did not reach statistical significance due to the small sample size (data not shown). Since the analyses of the associations between *AKT1* and memory, attention and gray matter volume were performed on different sub-samples with insufficient overlap, our results could not be clearly interpreted as the findings linking *AKT1*, the inferior parietal lobule and attentional dysfunction into one multi-model gene-morphology-cognition vulnerability factor. It would be desirable to perform all of the association analyses between *AKT1*, neurocognitive measures and brain images on the same subjects of age and sex matched case-control. Further studies are needed to elucidate the link on such subjects.

We also found relationships between *AKT1* SNP5 and gray matter volumes in the right superior frontal gyrus and the left caudate. These regions have been associated with different SNPs in *AKT1* gene (Pietilainen et al. 2009; Tan et al. 2008b). Interestingly, three independent studies showed evidence for a role of *AKT1* in the brain morphology. Frontostriatal circuits are related with cortical dopaminergic function and implicated in schizophrenia (Pantelis et al. 1997). Consistent with the preclinical evidence that couples *AKT1* to dopaminergic frontostriatal function and cellular structure (Beaulieu et al. 2004, 2005; Emamian et al.

2004), human genetic variation in *AKT1* has been associated with cognitive performances referable to the frontostriatal dopaminergic system (Tan et al. 2008b). Brain networks between frontal cortex and striatum are implicated in the cognitive functions reflecting executive function, IQ, and processing speed (Alexander et al. 1986; Pantelis et al. 1997; Tan et al. 2008b). Although we did not evaluate these cognitive functions in this study, *AKT1* SNP5 might also be associated with these cognitive functions.

In these VOI approaches, A carriers had smaller gray matter volumes than homozygous G subjects in patients, but there was a trend in the opposite direction in healthy subjects. This finding can be interpreted in two ways. First, it is possible that the genetic variant is related in a different way to dysregulated levels of *AKT1* protein or dopamine between patients and controls. Second, because we applied three multiple comparisons in the VOI analyses, there is the possibility of type 1 errors in healthy controls (the right inferior parietal lobule; corrected $P=0.063$, the right superior frontal gyrus; corrected $P=0.15$ and the left caudate; corrected $P=0.19$). Both these explanations are speculative and have to be treated with caution.

Since *Akt1*-knockout mice showed deficits in hippocampal neurogenesis and behavioural effects associated with the hippocampus (Balu et al. 2011), hippocampus might also be a structure associated with attentional performance and memory. However, we did not find genotype effect on the hippocampus in the present study. It remains unclear whether *AKT1* is related to hippocampal volume in patients with schizophrenia, and thus further investigation is needed.

The present study is the first report demonstrating an association between SNP5 in the *AKT1* gene and neuropsychological parameters and gray matter volume in an Asian population. Four studies have investigated an association between SNPs in *AKT1* and learning, memory or attention in Caucasian populations (Pinheiro et al. 2007; Tan et al. 2008b; Pietilainen et al. 2009; Blasi et al. 2011). In subjects with European ancestry, there was an association between haplotypes comprising SNP3, SNP4 and SNP5 as well as the single SNP4 in *AKT1* and a factor comprised of IQ and processing speed; however, associations of the same SNP with six other cognitive factors, including verbal memory, working memory, visual memory, Wisconsin card sort, attention or digit span, were not found (Tan et al. 2008b). In healthy subjects, there was an association between SNP4 in *AKT1* and correct responses in the CPT (Blasi et al. 2011). On the other hand, there was no significant association between any of the five SNPs (rs3803300 (SNP1), SNP2, SNP3, SNP4 and SNP5) in *AKT1* and five domain scores (processing

speed, reasoning, verbal memory, working memory, and vigilance) in patients with schizophrenia (Pinheiro et al. 2007). Moreover, an association of SNP2 with verbal learning and memory was found in Finnish twins (Pietilainen et al. 2009). However, no association of SNP4 or SNP5 with any neuropsychological parameters has been shown. These inconsistencies might be effects of mixed populations or mixed subjects, since Pinheiro et al. analyzed the associations in subjects of African, European, and other ethnic populations and Pietilainen et al. analyzed the associations in combined samples of twin pairs of healthy controls, patients with schizophrenia, and patients with bipolar disorder. Taken together, these data suggest that SNPs in the *AKT1* gene, which is associated with schizophrenia, might exert different genotype effects on neuropsychological parameters in different ethnic populations.

Previous reports have indicated that lithium or valproate target AKT1 (Gupta et al. 2011). In the present study, there were only three patients taking lithium (G/G $N=2$, mean dose \pm SD (mg/day): 600.0 ± 0.0 , A carriers $N=1$, 400.0) and six patients taking valproate (G/G $N=4$, 800.0 ± 163.3 , A carriers $N=2$, 400.0 ± 282.3). There was no significant difference of memory, attention or gray matter volume between patients with and without these drugs treatment (data not shown), suggesting that these drugs might not affect our findings.

The D2/AKT1/GSK-3 β signalling pathway has been involved in the downstream intracellular effects of dopamine. Dopaminergic dysregulation plays an important role in the pathophysiology of schizophrenia. Different molecular pathways as downstream of dopamine D2 receptors have been identified: the classic cAMP-PKA pathway and the cAMP-independent D2 signalling cascade pathway that includes the AKT1, which phosphorylates to inhibit another protein kinase, GSK3 β (Freyberg et al. 2010). D2 stimulation by dopamine inhibits AKT1 signalling through dephosphorylation via the β -arrestin2/phosphatase PP2A complex (Beaulieu et al. 2007a,b), indicating the specific relationship between D2 receptor signalling and AKT1. In the *Akt1*-deficient mice, impaired working memory and prepulse inhibition of startle were caused by D2 agonists (Emamian et al. 2004; Lai et al. 2006). D2 receptors are essential for the Akt inhibition by dopamine (Beaulieu et al. 2007b). Importantly, reduced protein expression of AKT1 and phosphorylation of GSK3 β have been indicated in postmortem brains (prefrontal cortex) and lymphocytes of patients with schizophrenia (Emamian et al. 2004; Thiselton et al. 2008). Furthermore, a D2 antagonist antipsychotic increases AKT1 and GSK-3 β phosphorylation (Kang et al. 2004). Combined with the earlier

involvement of D2 signaling in attentional processing (Zhang et al. 2007) and the relationship between *AKT1* and attentional function in this study, the D2/AKT1/GSK-3 β signalling pathway might be involved in the pathophysiology of attentional deficits in schizophrenia.

There were several limitations to this study. After confirmed that the patients' symptoms were stable and doses of their antipsychotic treatments were maintained over 4 weeks, we performed all tests in patients with schizophrenia. However, we did not exclude the possibility that treatment status affect subtle genetic effects in patients with schizophrenia. To reduce the type I error due to small clusters, we applied only clusters of more than 180 contiguous voxels in the neuroimaging analysis. Although we have empirically selected the cluster size threshold of 180 contiguous voxels, it should be noted that the extent to which the type I error is reduced is unknown. A whole brain FWE correction was not used in the analysis. SVC for a radius of 10 mm around the peak applied in this study was liberal. These could lead to a type I error in this study. It is unclear whether SNP5 is associated with biological activity on AKT signalling, such as AKT1 protein levels and phosphorylation of GSK3 β . The lack of such association makes it unclear if our results are directly linked with *AKT1* rs2494732 or with other polymorphisms in linkage disequilibrium with this genetic variant. Further study to elucidate functional molecular effects of SNP5 is required.

In conclusion, the *AKT1* polymorphism could be related to the deficits in simple performance of the attentional tasks and the gray matter volumes of the right inferior parietal lobule and the frontostriatal region in Japanese patients with schizophrenia. Further studies are needed to elucidate an underlying genetic vulnerability to neurobiological traits in schizophrenia.

Acknowledgements

We thank all individuals who participated in this study. We also thank Louise Verrall for the English proofreading of the manuscript. This work was supported by Grants-in-Aid from the Japanese Ministry of Health, Labor and Welfare (H19-kokoro-002, Comprehensive Research on Disability Health and Welfare, and the Research Committee of System Development for Clinical Trials in Psychiatry and Neurology), the Japanese Ministry of Education, Culture, Sports, Science and Technology (18689030 and 22390225), the Core Research for Evolutionary Science and Technology of Japan Science and Technology Agency, Grant-aid for Scientific Research on Priority Areas -Research on Pathomechanisms of Brain

Disorders from the MEXT (18023045) and Japan Foundation for Neuroscience and Mental Health.

Statement of interest

None to declare.

References

- Alexander GE, DeLong MR, Strick PL. 1986. Parallel organization of functionally segregated circuits linking basal ganglia and cortex. *Annu Rev Neurosci* 9:357–381.
- Ashburner J, and Friston KJ. 2000. Voxel-based morphometry – the methods. *Neuroimage* 11:805–821.
- Bajestan SN, Sabouri AH, Nakamura M, Takashima H, Keikhae MR, Behdani F, et al. 2006. Association of AKT1 haplotype with the risk of schizophrenia in Iranian population. *Am J Med Genet B Neuropsychiatr Genet* 141B:383–386.
- Balu DT, Carlson GC, Talbot K, Kazi H, Hill-Smith TE, Easton RM, et al. 2011. Akt1 deficiency in schizophrenia and impairment of hippocampal plasticity and function. *Hippocampus*.
- Beaulieu JM, Sotnikova TD, Yao WD, Kockeritz L, Woodgett JR, Gainetdinov RR, et al. 2004. Lithium antagonizes dopamine-dependent behaviors mediated by an AKT/glycogen synthase kinase 3 signaling cascade. *Proc Natl Acad Sci USA* 101: 5099–5104.
- Beaulieu JM, Sotnikova TD, Marion S, Lefkowitz RJ, Gainetdinov RR, Caron MG. 2005. An Akt/beta-arrestin 2/PP2A signaling complex mediates dopaminergic neurotransmission and behavior. *Cell* 122:261–273.
- Beaulieu JM, Gainetdinov RR, Caron MG. 2007a. The Akt-GSK-3 signaling cascade in the actions of dopamine. *Trends Pharmacol Sci* 28:166–172.
- Beaulieu JM, Tirotta E, Sotnikova TD, Masri B, Salahpour A, Gainetdinov RR, et al. 2007b. Regulation of Akt signaling by D2 and D3 dopamine receptors in vivo. *J Neurosci* 27: 881–885.
- Betcheva ET, Mushiroda T, Takahashi A, Kubo M, Karachanak SK, Zaharieva IT, et al. 2009. Case-control association study of 59 candidate genes reveals the DRD2 SNP rs6277 (C957T) as the only susceptibility factor for schizophrenia in the Bulgarian population. *J Hum Genet* 54:98–107.
- Blasi G, Napolitano F, Ursini G, Taurisano P, Romano R, Caforio G, et al. 2011. DRD2/AKT1 interaction on D2 c-AMP independent signaling, attentional processing, and response to olanzapine treatment in schizophrenia. *Proc Natl Acad Sci USA* 108:1158–1163.
- Cardno AG, and Gottesman, II. 2000. Twin studies of schizophrenia: from bow-and-arrow concordances to star wars Mx and functional genomics. *Am J Med Genet* 97:12–17.
- Chan RC, Di X, McAlonan GM, Gong QY. 2011. Brain anatomical abnormalities in high-risk individuals, first-episode, and chronic schizophrenia: An activation likelihood estimation meta-analysis of illness progression. *Schizophr Bull* 37:177–188.
- Chen WJ, Faraone SV. 2000. Sustained attention deficits as markers of genetic susceptibility to schizophrenia. *Am J Med Genet* 97:52–57.
- Cornblatt BA, Keilp JG. 1994. Impaired attention, genetics, and the pathophysiology of schizophrenia. *Schizophr Bull* 20: 31–46.
- Cornblatt BA, Risch NJ, Faris G, Friedman D, Erlenmeyer-Kimling L. 1988. The Continuous Performance Test, identical pairs version (CPT-IP): I. New findings about sustained attention in normal families. *Psychiatry Res* 26:223–238.
- Cornblatt BA, Lenzenweger MF, Erlenmeyer-Kimling L. 1989. The continuous performance test, identical pairs version: II. Contrasting attentional profiles in schizophrenic and depressed patients. *Psychiatry Res* 29:65–85.
- Ellison-Wright I, Bullmore E. 2010. Anatomy of bipolar disorder and schizophrenia: a meta-analysis. *Schizophr Res* 117:1–12.
- Emamian ES, Hall D, Birnbaum MJ, Karayiorgou M, Gogos JA. 2004. Convergent evidence for impaired AKT1-GSK3beta signaling in schizophrenia. *Nat Genet* 36:131–137.
- Freyberg Z, Ferrando SJ, Javitch JA. 2010. Roles of the Akt/GSK-3 and Wnt signaling pathways in schizophrenia and antipsychotic drug action. *Am J Psychiatry* 167:388–396.
- Good CD, Johnsrude IS, Ashburner J, Henson RN, Friston KJ, Frackowiak RS. 2001. A voxel-based morphometric study of ageing in 465 normal adult human brains. *Neuroimage* 14: 21–36.
- Green MF. 2006. Cognitive impairment and functional outcome in schizophrenia and bipolar disorder. *J Clin Psychiatry* 67(Suppl 9):3–8; discussion 36–42.
- Grimes CA, Jope RS. 2001. The multifaceted roles of glycogen synthase kinase 3beta in cellular signaling. *Prog Neurobiol* 65:391–426.
- Gupta A, Schulze TG, Nagarajan V, Akula N, Corona W, Jiang XY, et al. 2011. Interaction networks of lithium and valproate molecular targets reveal a striking enrichment of apoptosis functional clusters and neurotrophin signaling. *Pharmacogenom J*, in press.
- Harris SL, Gil G, Robins H, Hu W, Hirshfield K, Bond E, et al. 2005. Detection of functional single-nucleotide polymorphisms that affect apoptosis. *Proc Natl Acad Sci USA* 102: 16297–16302.
- Hashimoto R, Numakawa T, Ohnishi T, Kumamaru E, Yagasaki Y, Ishimoto T, et al. 2006. Impact of the DISC1 Ser704Cys polymorphism on risk for major depression, brain morphology and ERK signaling. *Hum Mol Genet* 15:3024–3033.
- Hashimoto R, Hashimoto H, Shintani N, Chiba S, Hattori S, Okada T, et al. 2007. Pituitary adenylate cyclase-activating polypeptide is associated with schizophrenia. *Mol Psychiatry* 12:1026–1032.
- Horwood JM, Dufour F, Laroche S, Davis S. 2006. Signalling mechanisms mediated by the phosphoinositide 3-kinase/Akt cascade in synaptic plasticity and memory in the rat. *Eur J Neurosci* 23:3375–3384.
- Ide M, Ohnishi T, Murayama M, Matsumoto I, Yamada K, Iwayama Y, et al. 2006. Failure to support a genetic contribution of AKT1 polymorphisms and altered AKT signaling in schizophrenia. *J Neurochem* 99:277–287.
- Ikeda M, Iwata N, Suzuki T, Kitajima T, Yamanouchi Y, Kinoshita Y, et al. 2004. Association of AKT1 with schizophrenia confirmed in a Japanese population. *Biol Psychiatry* 56:698–700.
- Ikeda M, Yamanouchi Y, Kinoshita Y, Kitajima T, Yoshimura R, Hashimoto S, et al. 2008. Variants of dopamine and serotonin candidate genes as predictors of response to risperidone treatment in first-episode schizophrenia. *Pharmacogenomics* 9:1437–1443.
- Kang UG, Seo MS, Roh MS, Kim Y, Yoon SC, Kim YS. 2004. The effects of clozapine on the GSK-3-mediated signaling pathway. *FEBS Lett* 560:115–119.
- Karege F, Perroud N, Schurhoff F, Meary A, Marillier G, Burkhardt S, et al. 2010. Association of AKT1 gene variants and protein expression in both schizophrenia and bipolar disorder. *Genes Brain Behav* 9:503–511.
- Lai WS, Xu B, Westphal KG, Paterlini M, Olivier B, Pavlidis P, et al. 2006. Akt1 deficiency affects neuronal morphology and predisposes to abnormalities in prefrontal cortex functioning. *Proc Natl Acad Sci USA* 103:16906–16911.
- Lee KY, Joo EJ, Jeong SH, Kang UG, Roh MS, Kim SH, et al. 2010. No association between AKT1 polymorphism and

- schizophrenia: a case-control study in a Korean population and a meta-analysis. *Neurosci Res* 66:238–245.
- Lin CH, Yeh SH, Lu KT, Leu TH, Chang WC, Gean PW. 2001. A role for the PI-3 kinase signaling pathway in fear conditioning and synaptic plasticity in the amygdala. *Neuron* 31:841–851.
- Liu YL, Fann CS, Liu CM, Wu JY, Hung SI, Chan HY, et al. 2006. Absence of significant associations between four AKT1 SNP markers and schizophrenia in the Taiwanese population. *Psychiatr Genet* 16:39–41.
- Mathur A, Law MH, Megson IL, Shaw DJ, Wei J. 2010. Genetic association of the AKT1 gene with schizophrenia in a British population. *Psychiatr Genet* 20:118–122.
- Meyer-Lindenberg A, Weinberger DR. 2006. Intermediate phenotypes and genetic mechanisms of psychiatric disorders. *Nat Rev Neurosci* 7:818–827.
- Mizuno M, Yamada K, Takei N, Tran MH, He J, Nakajima A, et al. 2003. Phosphatidylinositol 3-kinase: a molecule mediating BDNF-dependent spatial memory formation. *Mol Psychiatry* 8:217–224.
- Norton N, Williams HJ, Dwyer S, Carroll L, Peirce T, Moskvina V, et al. 2007. Association analysis of AKT1 and schizophrenia in a UK case control sample. *Schizophr Res* 93:58–65.
- Ohtsuki T, Inada T, Arinami T. 2004. Failure to confirm association between AKT1 haplotype and schizophrenia in a Japanese case-control population. *Mol Psychiatry* 9:981–983.
- Pantelis C, Barnes TR, Nelson HE, Tanner S, Weatherley L, Owen AM, et al. 1997. Frontal-striatal cognitive deficits in patients with chronic schizophrenia. *Brain* 120(Pt 10): 1823–1843.
- Pietilainen OP, Paunio T, Loukola A, Tuulio-Henriksson A, Kieseppa T, Thompson P, et al. 2009. Association of AKT1 with verbal learning, verbal memory, and regional cortical gray matter density in twins. *Am J Med Genet B Neuropsychiatr Genet* 150B:683–692.
- Pinheiro AP, Keefe RS, Skelly T, Ollarte M, Leviel K, Lange LA, et al. 2007. AKT1 and neurocognition in schizophrenia. *Aust NZ J Psychiatry* 41:169–177.
- Salgado-Pineda P, Junque C, Vendrell P, Baeza I, Bargallo N, Falcon C, et al. 2004. Decreased cerebral activation during CPT performance: structural and functional deficits in schizophrenic patients. *Neuroimage* 21:840–847.
- Sanders AR, Duan J, Levinson DF, Shi J, He D, Hou C, et al. 2008. No significant association of 14 candidate genes with schizophrenia in a large European ancestry sample: implications for psychiatric genetics. *Am J Psychiatry* 165:497–506.
- Schwab SG, Hoefgen B, Hanses C, Hassenbach MB, Albus M, Lerer B, et al. 2005. Further evidence for association of variants in the AKT1 gene with schizophrenia in a sample of European sib-pair families. *Biol Psychiatry* 58:446–450.
- Sei Y, Li Z, Song J, Ren-Patterson R, Tunbridge EM, Iizuka Y, et al. 2010. Epistatic and functional interactions of catechol-O-methyltransferase (COMT) and AKT1 on neuregulin1-ErbB signaling in cell models. *PLoS One* 5:e10789.
- Shi J, Gershon ES, Liu C. 2008. Genetic associations with schizophrenia: meta-analyses of 12 candidate genes. *Schizophr Res* 104:96–107.
- Skellley SL, Goldberg TE, Egan MF, Weinberger DR, Gold JM. 2008. Verbal and visual memory: characterizing the clinical and intermediate phenotype in schizophrenia. *Schizophr Res* 105:78–85.
- Snitz BE, Macdonald AW, 3rd, Carter CS. 2006. Cognitive deficits in unaffected first-degree relatives of schizophrenia patients: a meta-analytic review of putative endophenotypes. *Schizophr Bull* 32:179–194.
- Sugishita M. 2001. Japanese Wechsler Memory Scale-Revised. Tokyo: Nihonbunkakagakusha.
- Sui L, Wang J, Li BM. 2008. Role of the phosphoinositide 3-kinase-Akt-mammalian target of the rapamycin signaling pathway in long-term potentiation and trace fear conditioning memory in rat medial prefrontal cortex. *Learn Mem* 15:762–776.
- Sun J, Kuo PH, Riley BP, Kendler KS, Zhao Z. 2008. Candidate genes for schizophrenia: a survey of association studies and gene ranking. *Am J Med Genet B Neuropsychiatr Genet* 147B:1173–1181.
- Tan HY, Callicott JH, Weinberger DR. 2008a. Intermediate phenotypes in schizophrenia genetics redux: is it a no brainer? *Mol Psychiatry* 13:233–238.
- Tan HY, Nicodemus KK, Chen Q, Li Z, Brooke JK, Honea R, et al. 2008b. Genetic variation in AKT1 is linked to dopamine-associated prefrontal cortical structure and function in humans. *J Clin Invest* 118:2200–2208.
- Thiselton DL, Vladimirov VI, Kuo PH, McClay J, Wormley B, Fanous A, et al. 2008. AKT1 is associated with schizophrenia across multiple symptom dimensions in the Irish study of high density schizophrenia families. *Biol Psychiatry* 63:449–457.
- Tsuang M. 2000. Schizophrenia: genes and environment. *Biol Psychiatry* 47:210–220.
- Turunen JA, Peltonen JO, Pietilainen OP, Hennah W, Loukola A, Paunio T, et al. 2007. The role of DTNBP1, NRG1, and AKT1 in the genetics of schizophrenia in Finland. *Schizophr Res* 91:27–36.
- Wechsler D. 1987. Wechsler Memory Scale Manual, Revised. San Antonio, TX: Psychological Corporation.
- Weinberger DR, Egan MF, Bertolino A, Callicott JH, Mattay VS, Lipska BK, et al. 2001a. Prefrontal neurons and the genetics of schizophrenia. *Biol Psychiatry* 50:825–844.
- Weinberger DR, Egan MF, Bertolino A, Callicott JH, Mattay VS, Lipska BK, et al. 2001b. Prefrontal neurons and the genetics of schizophrenia. *Biol Psychiatry* 50:825–844.
- Xu MQ, Xing QH, Zheng YL, Li S, Gao JJ, He G, et al. 2007. Association of AKT1 gene polymorphisms with risk of schizophrenia and with response to antipsychotics in the Chinese population. *J Clin Psychiatry* 68:1358–1367.
- Zhang Y, Bertolino A, Fazio L, Blasi G, Rampino A, Romano R, et al. 2007. Polymorphisms in human dopamine D2 receptor gene affect gene expression, splicing, and neuronal activity during working memory. *Proc Natl Acad Sci USA* 104:20552–20557.



ORIGINAL INVESTIGATION

The *KCNH2* gene is associated with neurocognition and the risk of schizophrenia

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Abstract

Objectives. A genetic variant (rs3800779; M30) in the *KCNH2* gene has been associated with schizophrenia, a lower intelligence quotient (IQ) and processing speed scores, altered brain functions and increased *KCNH2-3.1* mRNA levels in the hippocampus. The aims of this study were to investigate whether the *KCNH2* polymorphism is associated with schizophrenia-related neurocognitive deficits and to confirm the association between the variant and schizophrenia. **Methods.** The effects of the risk genotype on IQ and seven neurocognitive batteries were examined by the analysis of covariance in 191 healthy subjects. We performed a meta-analysis of the association between M30 and schizophrenia using five independent ethnic groups (1,720 cases; 2,418 controls). **Results.** Consistent with the previous study, we provided evidence that subjects with the risk T carriers had significantly lower IQ scores than those with the G/G genotype ($P=0.048$). Of the seven neurocognitive batteries, subjects with the risk genotype demonstrated lower performances on attention/vigilance ($P=0.0079$) and working memory ($P=0.0066$) relative to subjects with the G/G genotype. Meta-analysis demonstrated evidence for an association between M30 and schizophrenia without showing heterogeneity across studies (odds ratio = 1.18; $P=0.0017$). **Conclusions.** These data suggest that the *KCNH2* polymorphism could be associated with schizophrenia-related neuropsychological deficits and the risk of developing schizophrenia.

Key words: schizophrenia, *KCNH2* (potassium channel, voltage-gated subfamily H, member 2), intelligence quotient (IQ), single nucleotide polymorphism (SNP), meta-analysis, neurocognition

Introduction

Schizophrenia is a common, complex psychiatric disease characterized by both clinical and genetic heterogeneity. There are strong genetic components of the disease with an estimated heritability of approximately 80% (Cardno and Gottesman 2000; Tsuang 2000). Attempts have been made to minimize this heterogeneity and to clarify the genetic architecture. One strategy for gene discovery proposes using quantitative neurobiological traits as

intermediate phenotypes instead of relying on the diagnosis of schizophrenia alone to identify cases for investigation (Meyer-Lindenberg and Weinberger 2006; Tan et al. 2008a). This strategy has the potential to reduce clinical and genetic heterogeneity by applying alternative phenotypes that better reflect the underlying genetic vulnerability than does diagnostic categorization. Neurocognitive deficits, a core component of schizophrenia (Green 2006), are considered promising intermediate phenotypes for gene

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(Received 22 February 2011; accepted 7 July 2011)

ISSN 1562-2975 print/ISSN 1814-1412 online © 2011 Informa Healthcare
DOI: 10.3109/15622975.2011.604350



discovery in schizophrenia (Snitz et al. 2006; Husted et al. 2009). There is substantial evidence suggesting that most cognitive abilities have a genetic basis (Chen et al. 1998; Posthuma et al. 2001; Berrettini 2005; Husted et al. 2009). The estimated heritabilities of processing speed, attention/vigilance, verbal intelligence quotient (IQ) and performance IQ are 33–48, 48–62, 85 and 69%, respectively.

Recently, Huffaker et al. identified a potential schizophrenia susceptibility (human ether-a-go-go-related) gene, *KCNH2*, which encodes a voltage-activated potassium channel (Huffaker et al. 2009). The *KCNH2* gene contains 15 exons spanning 33 kb on chromosome 7q35–q36. A genetic variant (rs3800779; M30) in the *KCNH2* gene predicts lower IQ and processing speed scores, decreased hippocampal volume, altered memory-linked hippocampal functions and working memory-linked prefrontal functions. It also predicts increased expression levels of a primate- and brain-specific *KCNH2*-3.1 isoform in the hippocampus (Huffaker et al. 2009). Expression of *KCNH2*-3.1 in rodent cortical neurons causes a marked alteration in *KCNH2* channel physiology resulting in high-frequency, nonadapting neuronal firing patterns (Huffaker et al. 2009). In this study, we examined the effects of the M30 genotype on IQ and seven neurocognitive functions shown to be associated with genetic liability in schizophrenia. We then conducted a meta-analysis of M30 in previously reported samples added to a Japanese sample to establish further evidence for an association between the *KCNH2* gene and schizophrenia.

Methods and materials

Subjects

Neurocognitive test data were available for 191 Japanese healthy individuals (49.2% males (94/97); mean age \pm SD: 36.0 ± 11.5 years; years of education \pm SD: 15.5 ± 2.4 years). Data from different number of subjects were available in each test (general IQ 143 subjects, speed of processing 188, attention/vigilance 191, working memory 190, Verbal Learning and Memory 190, Visual Learning and Memory 190, Reasoning and problem solving 150, and Social cognition 86). Demographic variables for subjects included in each cognitive test are shown in Supplementary Table I (available online). Although we attempted to examine all neurocognitive tests from all subjects as much as we could, all tests data were available for 83 subjects. Because an association between an SNP in the *KCNH2* gene and cognitive function was observed in healthy controls, we attempted to replicate the previous association

finding in healthy controls (Huffaker et al. 2009). The use of healthy subjects to investigate an association between a genetic variant and neurocognitive function avoids the potential confounders related to the duration of illness and medical treatment. Healthy controls were recruited by local advertisements in Osaka, Japan. Psychiatrically, medically and neurologically healthy controls were evaluated using the Structured Clinical Interview for DSM-IV-Non-Patient Edition (SCID-I/NP) to exclude individuals who had received psychiatric medications. Subjects were also excluded from this study if they had neurological or medical conditions that could potentially affect the central nervous system, such as atypical headaches, head trauma with loss of consciousness, chronic lung disease, kidney disease, chronic hepatic disease, thyroid disease, active cancer, cerebrovascular disease, epilepsy, seizures, substance-related disorders or mental retardation. We excluded any control subjects with neurological disorders or first- or second-degree relatives with psychiatric disorders using an unstructured interview. All subjects were biologically unrelated Japanese individuals.

The subjects for the genetic association study consisted of 478 unrelated patients with schizophrenia (48.3% males (231/247); mean age \pm SD: 48.4 ± 15.7 years) and 640 unrelated healthy controls (46.3% males (296/344); mean age \pm SD: 58.9 ± 21.4 years). All subjects used in this analysis are unrelated Japanese, as described previously (Ohi et al. 2009b, 2010). Cases were recruited from both outpatients and inpatients at Osaka University Hospital and the psychiatric hospitals. Each subject with schizophrenia had been diagnosed by at least two trained psychiatrists based on an unstructured clinical interview; diagnoses were made based on the criteria of the DSM-IV. Controls were recruited through local advertisements. Psychiatrically healthy controls were evaluated using unstructured interviews to exclude individuals who had current or past contact with psychiatric services. Written informed consent was obtained for all subjects after the procedures had been fully explained. This study was carried out in accordance with the World Medical Association's Declaration of Helsinki and was approved by the Research Ethical Committee of Osaka University.

SNP selection and SNP genotyping

We selected rs3800779 (M30) in the *KCNH2* gene because this SNP has been associated with schizophrenia, as described in the introduction (Huffaker et al. 2009). Venous blood was collected from the subjects, and genomic DNA was extracted from whole blood according to standard procedures. The SNP was genotyped using the custom-designed

TaqMan 5'-exonuclease allelic discrimination assay (Applied Biosystems, Foster City, CA, USA), as described previously (Hashimoto et al. 2007). No deviation from Hardy–Weinberg equilibrium in the examined SNP was detected in patients with schizophrenia or in controls ($P > 0.05$).

Neurocognitive testing

General intellectual function was derived from the Full Scale IQ portion of the Wechsler Adult Intelligence Scale-Third Edition (WAIS-III) (Wechsler 1997). The Measurement and Treatment Research to Improve Cognition in Schizophrenia (MATRICS) Neurocognition Committee selected seven neurocognitive domains from all available factor-analytic studies of cognitive performance in schizophrenia patients (Green et al. 2004; Nuechterlein et al. 2004). Seven neurocognitive batteries were selected based upon previous studies to assess the following seven domains (Nuechterlein et al. 2004, 2008): (1) speed of processing, (2) attention/vigilance, (3) working memory, (4) verbal learning and memory, (5) visual learning and memory, (6) reasoning and problem solving, and (7) social cognition. The speed of processing was assessed using the Category Fluency Test (total number of animals named in 60 s) (Sumiyoshi et al. 2004). Attention/vigilance was evaluated using the Continuous Performance Test-Identical Pairs version (d') (Cornblatt et al. 1988). Working memory was measured using the Wechsler Memory Scale-Revised (WMS-R) digit span subtest (number of correct trials) (Sugishita 2001). Verbal learning and memory was assessed using the immediate recall portion of the Rey Auditory Verbal Learning Test (Lezak 1995) in which the participants were asked to recall a list of 15 words spoken by a tester. The procedure was repeated five times (sessions 1–5), and the sum of the recalled words from sessions 1 to 5 was used for the analysis. If the participants scored 15/15, we treated the scores of the participant as 15 after the session; possible scores range from 0 to 75. Visual learning and memory was evaluated using the visual reproduction I subtest of the WMS-R (number of correct trials) (Sugishita 2001). Reasoning and problem solving was measured using the tower of Hanoi task (number of correct trials) (Ohi et al. 2009a). Social cognition was assessed using the Emotion Recognition test (correct rate of the Facial Emotion Labeling Test (FELT)) (Sekiya et al. 2008). The subjects included in this analysis were assessed by trained clinical psychologists to obtain scores on the WAIS-III Full Scale IQ and the seven schizophrenia-related neurocognitive batteries.

Meta-analysis

The studies included in the meta-analysis were obtained using PubMed using the search terms “*KCNH2*” and “schizophrenia”. The analyzed data encompassed all publications up to October 2010. Additionally, references cited in the publications obtained were examined to identify additional potentially relevant studies that might not be listed in PubMed. Studies were included in the meta-analysis if they met the following criteria: (1) published in a peer-reviewed journal in English and (2) included a genetic association study between the *KCNH2* gene and schizophrenia. Our meta-analysis included allele frequency data from all available case–control studies only and did not include the original family-based dataset that provided strong evidence for the positive association in the original report by Huffaker et al. (Huffaker et al. 2009). We calculated each number of alleles from the allele frequency and the odds ratio data for each study.

Statistical analyses

Statistical analyses were performed using the PASW Statistics 18.0 software (SPSS Japan Inc., Tokyo, Japan). Differences in clinical characteristics between patients and controls or between genotypes were analyzed using χ^2 -tests for categorical variables and the Mann–Whitney U -test for continuous variables. Based on the assumption that demographic variables such as age and education years might not be fitted to a normal distribution, we used the nonparametric Mann–Whitney test arbitrary to assess the demographic variables. The presence of Hardy–Weinberg equilibrium was examined using the χ^2 -test for goodness of fit. To control for confounding factors such as age, sex and years of education, we used a one-way analysis of covariance (ANCOVA) for neurocognitive tests, based on the assumption that the neurocognitive variables could be fitted to a normal distribution. The effect of the *KCNH2* genotype on IQ was analyzed by a one-way ANCOVA with sex and years of education as covariates because the IQ scores were already corrected for age. The effects on the seven neurocognitive domains were analyzed by a one-way ANCOVA with age, sex and years of education as covariates. Bonferroni correction was applied for multiple testing on seven domains to avoid type I errors. Standardized effect sizes were indicated using Cohen's d and η^2 .

The meta-analysis was performed using the Comprehensive Meta-Analysis software (Version 2.0, BIOSTAT, Englewood Cliffs, NJ, USA). Cochran's χ^2 -based Q -statistical test was performed to assess possible heterogeneity among studies. The fixed-effect

Table I. Impact of M30 in the *KCNH2* gene on IQ and on seven cognitive batteries.

Variables	Healthy subjects					ANCOVA			
	<i>n</i>	<i>n</i>	T carrier	<i>n</i>	G/G	Cohen's <i>d</i>	<i>F</i>	<i>P</i> values	η^2
General intellectual function	143	29	107.5 ± 15.4	114	110.3 ± 10.9	-0.21	3.98	0.048	0.028
Speed of processing	188	35	22.4 ± 5.4	153	21.2 ± 4.7	0.24	0.76	0.39	0.004
Attention/vigilance	191	36	3.63 ± 0.51	155	3.75 ± 0.47	-0.24	7.20	0.0079	0.037
Working memory	190	36	15.3 ± 3.9	154	16.8 ± 3.7	-0.39	7.55	0.0066	0.039
Verbal learning and memory	190	36	56.4 ± 9.0	154	57.9 ± 7.9	-0.17	2.02	0.16	0.011
Visual learning and memory	190	36	38.8 ± 2.9	154	39.3 ± 2.1	-0.20	2.49	0.12	0.013
Reasoning and problem solving	150	31	13.2 ± 7.5	119	13.8 ± 7.1	-0.08	0.95	0.33	0.006
Social cognition	86	19	58.8 ± 13.1	67	62.3 ± 13.1	-0.27	2.49	0.12	0.030

IQ, intelligence quotient; ANCOVA, analysis of covariance. Means ± SD are shown. The effect sizes are typically categorized as small ($d = 0.20$, $\eta^2 = 0.01$), medium ($d = 0.50$, $\eta^2 = 0.06$) or large ($d = 0.80$, $\eta^2 = 0.14$). To control for confounding factors, the effect of the *KCNH2* genotype on IQ was analyzed by one-way ANCOVA with sex and years of education as covariates because the IQ scores were already corrected for age. The effects on seven neurocognitive domains were analyzed by one-way ANCOVA with age, sex and years of education as covariates.

model described by Mantel-Haenszel was applied in the absence of heterogeneity ($p > 0.05$). The significance of the pooled odds ratio (OR) was assessed using a z -test. The significance level for all statistical tests was set at two-tailed $P < 0.05$.

Results

The effect of the KCNH2 risk polymorphism on IQ and on seven neurocognitive batteries

There were no differences in demographic variables – age, sex, or years of education – between genotype groups in each cognitive test (Supplementary Table I available online). As shown in Table I, we found a significant genotype effect on general intellectual function ($F_{1,139} = 3.98$, $P = 0.048$). Additionally, we found significant genotype effects on attention/vigilance ($F_{1,186} = 7.20$, $P = 0.0079$) and working memory

($F_{1,185} = 7.55$, $P = 0.0066$) from the seven batteries. The effect sizes (η^2) of IQ, attention/vigilance and working memory were 0.028, 0.037 and 0.039, respectively. Subjects with the risk T carriers had lower performances on these tests than did those with the G/G genotype (Figure 1). The genotype effect on working memory remained positive after the correction for multiple tests (corrected $P = 0.046$), while the genotype effect on attention/vigilance did not reach statistical significance after the correction (corrected $P = 0.055$). No significant genotype effect was found in any other cognitive batteries ($P > 0.05$).

Association between a genetic variant in the KCNH2 gene and schizophrenia by meta-analysis

The frequency of the T allele of M30 was higher in patients (11.0%) than in controls (9.7%) in the Japanese population used in this study. The direction

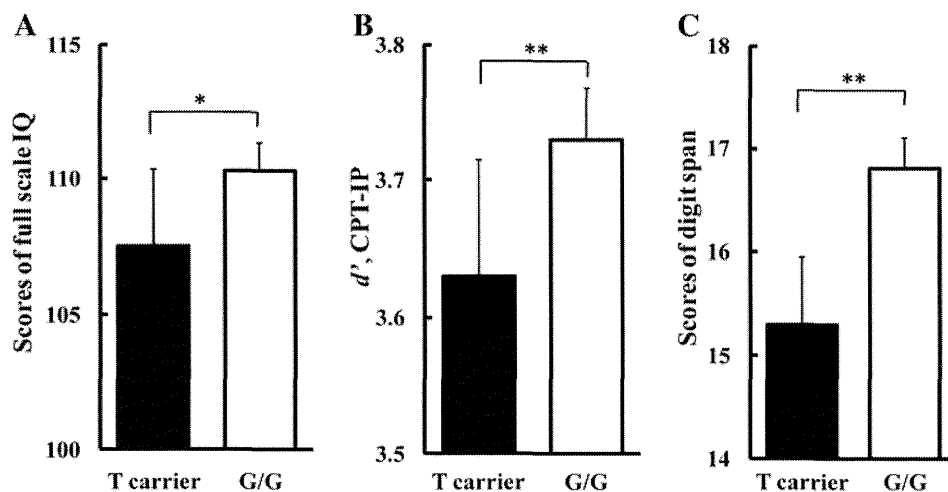


Figure 1. Association between the *KCNH2* risk genotype and IQ, attention/vigilance (B) and working memory (C). The x-axis represents T carriers and individuals with the G/G genotype. The y-axis represents scores of each test. Error bars represent the standard error of the mean. * $P < 0.05$, ** $P < 0.01$.

Table II. Demographics of the combined studies.

	Authors	Ethnicities	Patients	Controls	Diagnostic criteria
<i>Case-control studies</i>					
1	Huffaker et al. (2009)	German	905	1323	DSM-IV and ICD-10
2	Huffaker et al. (2009)	Armenian	161	161	ICD-10
3	Huffaker et al. (2009)	Italian	92	220	DSM-IV
4	Atalar et al. (2010)	Turkish	84	74	DSM-IV
5	Hashimoto et al. (present study)	Japanese	478	640	DSM-IV
<i>Family-based studies</i>					
6	Huffaker et al. (2009)	USA (CBDB)	296 Caucasian families		DSM-IV
7	Huffaker et al. (2009)	USA (NIMH-GI)	71 Caucasian families		DSM-III-R

CBDB, Clinical Brain Disorders Branch; NIMHGI, National Institute of Mental Health-Genetics Initiative.

We selected five independent case-control and two family-based data sets from previous and present studies. Two family-based samples (studies 6 and 7) were excluded from the present study because the published genotype data (affecteds and unaffecteds) were not available and the family-based samples with a family history of schizophrenia were not representative of the general population. Because we simply examined the association in the case-control samples, we included four independent case-control samples (studies 1, 2, 3, 4 and 5) (1,720 cases; 2,418 controls).

of the difference in allele frequency between patients and controls is consistent with previous studies (Huffaker et al. 2009; Atalar et al. 2010); however, the results did not represent a statistically significant difference between the groups ($z = 0.99$, $P = 0.32$, OR (95% confidence interval) = 1.15 (0.87–1.51)]. Our study size of 478 cases and 640 controls in a Japanese population had insufficient power (< 0.80) to detect as small an effect as an OR of 1.12, as described in the previous genome-wide association study (O'Donovan et al. 2008). Thus, we performed a meta-analysis to provide enough power to detect such a small effect. We included five independent case-control samples, as described in Table II (1,720 cases, 2,418 controls) (Huffaker et al. 2009; Atalar et al. 2010). The meta-analysis of M30 in all available schizophrenia data sets provided evidence for an association with schizophrenia ($z = 3.14$, $P = 0.0017$, OR (95% confidence interval) = 1.18 (1.06–1.31)] and no evidence for heterogeneity across studies ($Q = 3.55$, $P = 0.47$) (Table III, Figure 2). A sensitivity analysis revealed that the evidence for the association was not dependent upon the inclusion of any one

data set (Supplementary Figure 1 available online).

Discussion

In this study, we replicated the association between the risk genotype *KCNH2* and IQ, and we further demonstrated the associations of the genotype with attention/vigilance and working memory in healthy Japanese subjects. We provided evidence that subjects with the risk T carriers had lower performances on these cognitive tests than did those with the G/G genotype. The effect sizes of the differences in these tests between individuals with T carriers and those with the G/G genotype were small to medium. Huffaker et al. reported a significant association between the M30 genotype and performance on IQ testing and on processing speed, which was extracted as a factor in healthy subjects (Huffaker et al. 2009). We did not find an association between processing speed and the risk genotype; however, we found associations between attention/vigilance and working memory and the risk genotype. To assess genotype

Table III. Comparison of allele frequencies of the *KCNH2* polymorphism (M30) in the combined samples.

M30 (rs3800779)	SCZ, Number of alleles (%)			CON, Number of alleles (%)			Statistics for each study		
	T	G	Sum	T	G	Sum	P value (z)	OR (95% CI)	Weight (fixed)
German	615 (34.0)	1195 (66.0)	1810	820 (31.0)	1826 (69.0)	2646	0.035 (2.10)	1.15 (1.01–1.30)	65.4
Armenian	105 (32.5)	217 (67.5)	322	87 (27.0)	235 (73.0)	322	0.13 (1.51)	1.30 (0.93–1.82)	9.2
Italian	51 (27.9)	133 (72.1)	184	114 (26.0)	326 (74.0)	440	0.63 (0.48)	1.10 (0.75–1.62)	7.1
Turkish	55 (32.7)	113 (67.3)	168	31 (20.9)	117 (79.1)	148	0.020 (2.33)	1.84 (1.10–3.06)	4.1
Japanese	105 (11.0)	851 (89.0)	956	124 (9.7)	1156 (90.3)	1280	0.32 (0.99)	1.15 (0.87–1.51)	14.1
Pool	931 (27.1)	2509 (72.9)	3440	1176 (24.3)	3660 (75.7)	4836	0.0017 (3.14)^a	1.18 (1.06–1.31)	

SCZ, patients with schizophrenia; CON, healthy controls.

^aTest of heterogeneity: $Q = 3.55$, $df(Q) = 4$, $P(Q) = 0.47$, $I^2 = 0$.

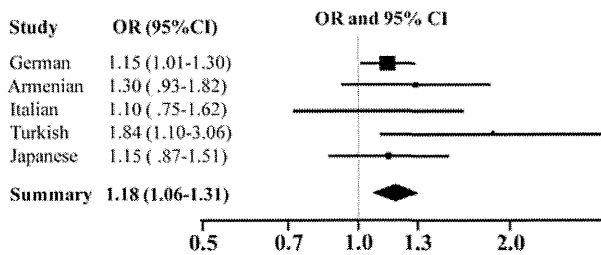


Figure 2. Forest plot of M30 results in the *KCNH2* gene based upon all combined populations. Solid squares and horizontal lines indicate the weighted odds ratios and 95% confidence intervals. The overall results are shown by the diamond. The results of the meta-analysis shown here are under the fixed-effects model.

effects on cognitive function, we measured seven domains based on MATRICS, a method different from the factor analyses-derived cognitive dimensions used in the previous study. Thus, the discrepancy between studies might be due to the differences in the cognitive dimensions, the methodology of the cognitive data analysis (direct measurements versus factors calculated by several measurements) and/or ethnic difference between European and Japanese individuals.

Second, we provide further evidence for an association between M30 and schizophrenia in combined case-control samples having now added a Japanese population. The allele frequencies of M30 in Japanese and European were different (-0.11 vs. -0.27) based on previous and present genome dataset. However, there was no evidence for heterogeneity across studies in the meta-analysis, suggesting that there was no obvious population stratification in the combined case-control samples. It is important to note that our meta-analysis did not include the family-based sample that showed strong evidence for association in the original report. A leave-one-out sensitivity analysis revealed that the significant meta-analysis results were not being driven by a single data set. Removal of any one data set did not negate the significance of the association from the meta-analysis. As expected, the effect size observed in this study was quite small (1.16), consistent with the results from a GWAS report (O'Donovan et al. 2008). Our data are consistent with the concept that many susceptibility risk alleles for schizophrenia come from common variants of small effect. Our data also suggest that a common allele could have a stronger influence on intermediate phenotypes than on the diagnosis of schizophrenia. Despite the importance of cognitive deficits in schizophrenia, no drug has been approved for the treatment of this aspect of schizophrenia. Some antipsychotics bind and inhibit *KCNH2* with affinities comparable to their affinities for the dopamine D2 receptors (Kongsamut et al. 2002). Further research will be required to clarify the role of

KCNH2 in the pathophysiology of schizophrenia. This research might potentially lead to new targets for antipsychotic medications.

There were several limitations to this study. We examined only M30 in the *KCNH2* gene, based on evidence that the variant predicts cognition, brain structure and function, and the gene expression level. We did not examine other markers of *KCNH2* gene or other genes to identify the association between those phenotype and schizophrenia. The lack of such association makes it unclear whether our results are directly linked with M30 or with other polymorphisms in linkage disequilibrium with this genetic variant. In addition, the neurocognitive tests batteries used in this study measure several complex functions (such as executive functions), not only associated with the one gene. Large number of researches show significant importance of genes connected with dopaminergic neurotransmitter system and other genes may interact with dopaminergic system (Tan et al. 2008b). Further study to investigate not only the single marker M30 but also these SNP/gene interactions is required.

Acknowledgments

We thank all individuals who participated in this study. We also appreciate Daniel R. Weinberger for critical comments on the manuscript. This work was supported in part by Grants-in-Aid from the Japanese Ministry of Health, Labor and Welfare (H18-kokoro-005, H19-kokoro-002, Comprehensive Research on Disability Health and Welfare, and the Research Committee of System Development for Clinical Trials in Psychiatry and Neurology), the Japanese Ministry of Education, Culture, Sports, Science and Technology (18689030), CREST of JST, and Japan Foundation for Neuroscience and Mental Health.

Statement of Interest

None to declare.

References

- Atalar F, Acuner TT, Cine N, Oncu F, Yesilbursa D, Ozbek U, et al. 2010. Two four-marker haplotypes on 7q36.1 region indicate that the potassium channel gene *HERG1* (*KCNH2*, *Kv11.1*) is related to schizophrenia: a case control study. *Behav Brain Funct* 6:27.
- Berrettini WH. 2005. Genetic bases for endophenotypes in psychiatric disorders. *Dialogues Clin Neurosci* 7:95-101.
- Cardno AG, and Gottesman, II. 2000. Twin studies of schizophrenia: from bow-and-arrow concordances to star wars Mx and functional genomics. *Am J Med Genet* 97:12-17.
- Chen WJ, Liu SK, Chang CJ, Lien YJ, Chang YH, Hwu HG. 1998. Sustained attention deficit and schizotypal personality features

- in nonpsychotic relatives of schizophrenic patients. *Am J Psychiatry* 155:1214–1220.
- Cornblatt BA, Risch NJ, Faris G, Friedman D, Erlenmeyer-Kimling L. 1988. The Continuous Performance Test, identical pairs version (CPT-IP): I. New findings about sustained attention in normal families. *Psychiatry Res* 26:223–238.
- Green MF, Nuechterlein KH, Gold JM, Barch DM, Cohen J, Essock S, et al. 2004. Approaching a consensus cognitive battery for clinical trials in schizophrenia: the NIMH-MATRICES conference to select cognitive domains and test criteria. *Biol Psychiatry* 56:301–307.
- Green MF. 2006. Cognitive impairment and functional outcome in schizophrenia and bipolar disorder. *J Clin Psychiatry* 67(Suppl 9):3–8; discussion 36–42.
- Hashimoto R, Hashimoto H, Shintani N, Chiba S, Hattori S, Okada T, et al. 2007. Pituitary adenylate cyclase-activating polypeptide is associated with schizophrenia. *Mol Psychiatry* 12:1026–1032.
- Huffaker SJ, Chen J, Nicodemus KK, Sambataro F, Yang F, Mattay V, et al. 2009. A primate-specific, brain isoform of *KCNH2* affects cortical physiology, cognition, neuronal repolarization and risk of schizophrenia. *Nat Med* 15:509–518.
- Husted JA, Lim S, Chow EW, Greenwood C, Bassett AS. 2009. Heritability of neurocognitive traits in familial schizophrenia. *Am J Med Genet B Neuropsychiatr Genet* 150B:845–853.
- Kongsamut S, Kang J, Chen XL, Roehr J, Rampe D. 2002. A comparison of the receptor binding and HERG channel affinities for a series of antipsychotic drugs. *Eur J Pharmacol* 450:37–41.
- Lezak MD. 1995. *Neuropsychological assessment*. 3rd ed. New York: Oxford University Press.
- Meyer-Lindenberg A, Weinberger DR. 2006. Intermediate phenotypes and genetic mechanisms of psychiatric disorders. *Nat Rev Neurosci* 7:818–827.
- Nuechterlein KH, Barch DM, Gold JM, Goldberg TE, Green MF, Heaton RK. 2004. Identification of separable cognitive factors in schizophrenia. *Schizophr Res* 72:29–39.
- Nuechterlein KH, Green MF, Kern RS, Baade LE, Barch DM, Cohen JD, et al. 2008. The MATRICS Consensus Cognitive Battery, part 1: test selection, reliability, and validity. *Am J Psychiatry* 165:203–213.
- O'Donovan MC, Craddock N, Norton N, Williams H, Peirce T, Moskva V, et al. 2008. Identification of loci associated with schizophrenia by genome-wide association and follow-up. *Nat Genet* 40:1053–1055.
- Ohi K, Hashimoto R, Yasuda Y, Kiribayashi M, Iike N, Yoshida T, et al. 2009a. TATA box-binding protein gene is associated with risk for schizophrenia, age at onset and prefrontal function. *Genes, Brain and Behavior* 8:473–480.
- Ohi K, Hashimoto R, Yasuda Y, Yoshida T, Takahashi H, Iike N, et al. 2009b. Association study of the G72 gene with schizophrenia in a Japanese population: a multicenter study. *Schizophr Res* 109:80–85.
- Ohi K, Hashimoto R, Yasuda Y, Yoshida T, Takahashi H, Iike N, et al. 2010. The chitinase 3-like 1 gene and schizophrenia: evidence from a multi-center case-control study and meta-analysis. *Schizophr Res* 116:126–132.
- Posthuma D, de Geus EJ, Boomsma DI. 2001. Perceptual speed and IQ are associated through common genetic factors. *Behav Genet* 31:593–602.
- Sekiyama R, Iwase M, Takahashi H, Nakahachi T, Takahashi K, Ikezawa K, et al. 2008. Perception of emotional and neutral facial expression correlates with social functioning in chronic schizophrenia. *Seishin Igaku* 50:337–344.
- Snitz BE, Macdonald AW, Carter CS. 2006. Cognitive deficits in unaffected first-degree relatives of schizophrenia patients: a meta-analytic review of putative endophenotypes. *Schizophr Bull* 32:179–194.
- Sugishita M. 2001. *Japanese Wechsler Memory Scale-Revised*. Tokyo: Nihonbunkakagakusha.
- Sumiyoshi C, Sumiyoshi T, Matsui M, Nohara S, Yamashita I, Kurachi M, et al. 2004. Effect of orthography on the verbal fluency performance in schizophrenia: examination using Japanese patients. *Schizophr Res* 69:15–22.
- Tan HY, Callicott JH, Weinberger DR. 2008a. Intermediate phenotypes in schizophrenia genetics redux: is it a no brainer? *Mol Psychiatry* 13:233–238.
- Tan HY, Nicodemus KK, Chen Q, Li Z, Brooke JK, Honea R, et al. 2008b. Genetic variation in *AKT1* is linked to dopamine-associated prefrontal cortical structure and function in humans. *J Clin Invest* 118:2200–2208.
- Tsuang M. 2000. Schizophrenia: genes and environment. *Biol Psychiatry* 47:210–220.
- Wechsler D. 1997. *Manual for the Wechsler Adult Intelligence Scale-III*. San Antonio, TX: The Psychological Corporation.

Supplementary material available online

Supplementary Table I. Demographic variables for subjects included in the neurocognitive battery analysis.

Supplementary Figure 1. A sensitivity analysis of M30.

Open

Variants of the *RELA* Gene are Associated with Schizophrenia and their Startle Responses

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The pathogenesis of schizophrenia is thought to involve aberrant immune and inflammatory responses. Nuclear factor kappa B (NF- κ B) has important roles in the immune and inflammatory responses. The *v-rel avian reticuloendotheliosis viral oncogene homolog A (RELA)* gene encodes the major component of the NF- κ B complex. We genotyped four single-nucleotide polymorphisms (SNPs) in the *RELA* gene and performed a gene-based association analysis using 1224 patients with schizophrenia and 1663 controls. We found significant associations of three SNPs (rs11820062: $p = 0.00011$, rs2306365: $p = 0.0031$, and rs7119750: $p = 0.0080$) with schizophrenia and stronger evidence for association in a multi-marker sliding window haplotype analysis (the lowest $p = 0.00006$). The association between this gene and schizophrenia was evident in male subjects but not in female subjects, when separately analyzed by gender. *In silico* genotype-gene expression analysis using web database and the WGAViewer software revealed that these three schizophrenia-associated SNPs might be related to *RELA* mRNA expression in immortalized B-lymphocytes. *In silico* analysis also suggested the putative promoter SNP, rs11820062, might disrupt the consensus transcription factor binding sequence of the androgen receptor. The impact of four *RELA* polymorphisms on pre-pulse inhibition (PPI) was investigated in 53 patients with schizophrenia. We provided evidence that at risk genotypes of three SNPs were associated with deficits in PPI; however, there was no effect of the one non-risk SNP on PPI. These findings suggest that variants of the *RELA* gene are associated with risk for schizophrenia and PPI deficits in a Japanese population. *Neuropsychopharmacology* (2011) **36**, 1921–1931; doi:10.1038/npp.2011.78; published online 18 May 2011

Keywords: schizophrenia; *v-rel avian reticuloendotheliosis viral oncogene homolog A (avian) (RELA)*; NF- κ B; pre-pulse inhibition (PPI); single-nucleotide polymorphism; gene expression

INTRODUCTION

Schizophrenia is a common and complex psychiatric disease. The lifetime morbidity rate is 0.5–1.0% across distinct populations. Family, twin, and adoption studies of schizophrenia have indicated that there are strong genetic factors with an estimated heritability of 80% (Cardno and

Gottesman, 2000; Tsuang, 2000). Although genes implicated in the pathogenesis of schizophrenia have been found through intense research efforts, eg association studies of candidate gene approach, genomewide association studies (GWAS), copy number variation (CNV) studies, and pedigree studies (Harrison and Weinberger, 2005; Cichon *et al*, 2009), the exact genetic factors of this complex disease remain to be explained. Recent polygenic component analysis in GWAS studies demonstrated the less effect between different populations, eg Europeans and African-Americans, Europeans, and Japanese, compared with the effects between two European populations (Purcell *et al*, 2009; Ikeda *et al*, 2010). As it may be due to aggregate differences in allele frequencies and patterns of linkage disequilibrium and/or population specific risk for

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Received 12 November 2010; revised 4 March 2011; accepted 6 April 2011

schizophrenia, association studies using multiple populations are expected.

Alterations in cytokine expression in schizophrenia have been extensively studied. The levels of cytokines, such as interleukin-1 β (IL-1 β), IL-1 receptor antagonist (IL-1RA), IL-6, and tumor necrosis factor- α (TNF- α), are increased in the plasma and cerebrospinal fluid of patients with schizophrenia (Naudin *et al.*, 1997; Potvin *et al.*, 2008). Nuclear factor-kappa B (NF- κ B), a prototypical transcription factor, regulates the expression of cytokines. Conversely, NF- κ B can be activated by pro-inflammatory cytokines, including IL-1 β and TNF- α , and in response to various cell stressors. Many genes have been shown to be responsive to NF- κ B (Baeuerle, 1991), including genes involved in survival/apoptosis, immune and inflammatory responses, and cell differentiation. In addition, the NF- κ B-responsive genes *IL2*, *IL6*, *TNF- α* , major histocompatibility complex (MHC), Bcl-2 family members, Calbindin, and ICAM-1 have been reported to be associated with schizophrenia (Jarskog *et al.*, 2000, 2004; Schwarz *et al.*, 2000; Benes *et al.*, 2001, 2003; Potvin *et al.*, 2008; Woo *et al.*, 2008; Purcell *et al.*, 2009; Shi *et al.*, 2009; Song *et al.*, 2009; Stefansson *et al.*, 2009).

NF- κ B is present in synaptic terminals and serves as a regulator of neuronal plasticity, which is activated by the activity of neuronal circuits (Mattson *et al.*, 2000). The NF- κ B complex is inhibited by the I κ B complex, which inactivates NF- κ B by sequestering it in the cytoplasm (Huxford and Ghosh, 2009). After the phosphorylation of serine residues on the I κ B proteins, I κ B dissociates from and activates the NF- κ B complex. The activated NF- κ B complex translocates into the nucleus and binds to regulatory elements in target genes. Constitutively, activated NF- κ B is detected mostly in glutamatergic neurons, whereas NF- κ B in glia has a lower basal activity and is heavily inducible (Kaltschmidt and Kaltschmidt, 2009). Knockout of a subunit of NF- κ B or inhibition of NF- κ B by super-repressor I κ B in neuron of mice resulted in defects in learning and memory and the loss of neuroprotection (Kaltschmidt and Kaltschmidt, 2009). It has been shown that activation of NF- κ B prevents neuronal apoptosis in various cell types. H₂O₂ increased NF- κ B activation and dopamine D2 receptor expression (Larouche *et al.*, 2008), suggesting that NF- κ B may participate in the psychopathology of schizophrenia through its effect on the neurotransmitter system. The association between cytokine expression and NF- κ B activation has been reported in schizophrenia (Song *et al.*, 2009). These findings support the hypothesis that alterations in cytokines and NF- κ B, which cause abnormal inflammatory responses, might contribute to the pathogenesis of schizophrenia.

Although three components of NF- κ B, NFKB1, NFKB2, and NFKB3, were not in major locus in schizophrenia (OMIM181500: <http://www.ncbi.nlm.nih.gov/omim>), *NFKB3* located on chromosome 11q13 showed a suggestive linkage to schizophrenia in a family-based linkage disequilibrium analysis in a Japanese population (Yamada *et al.*, 2004). The v-rel avian reticuloendotheliosis viral oncogene homolog A (*RELA*) gene (OMIM164014; alternative names include nuclear factor kappa-B, subunit 3 (*NFKB3*), transcription factor NFKB3, NFKB, p65 subunit, and nuclear factor of kappa light chain gene enhancer in

B cells 3) encodes the major subunit of the NF- κ B protein complex, are abundantly expressed in neurons and glia (Kaltschmidt and Kaltschmidt, 2009). Mice lacking *RELA* showed a learning deficit in the spatial version of the radial arm maze (Meffert *et al.*, 2003), indicating the critical involvement of the *RELA* gene in memory function, which may be related to pathophysiology of memory dysfunction in schizophrenia. Microarray Expression Data in UCSC Genome Bioinformatics Site (<http://genome.ucsc.edu/>) showed moderate mRNA expression of the *RELA* gene and the components of I κ B (*NFKBIA* and *NFKBIB*) in human brain and prominent expression in prefrontal cortex. Altered expression of these genes in postmortem brain of patients with schizophrenia was not found in the pooled gene expression data in the Stanley Genomic Medical Research Institute Online Genomics Database (<https://www.stanleygenomics.org>). In this study, we first investigated the association between the *RELA* gene and schizophrenia in a Japanese population, and then performed *in silico* genotype-gene expression analysis.

Impaired sensorimotor gating is considered to be a common psychophysiological feature of schizophrenia that could theoretically lead to a variety of severe defects in perception, attention, and thinking (Braff and Geyer, 1990). Pre-pulse inhibition (PPI) of the acoustic startle reflex (ASR) is the most common psychophysiological index of sensorimotor gating. PPI is emerging as an important intermediate phenotype for schizophrenia (Braff and Light, 2005), because it has high heritability (Anokhin *et al.*, 2003) and PPI deficits have been found in high-risk subjects (Cadenhead *et al.*, 2000). It has been hypothesized that the maternal immune response to infection may influence fetal brain development and lead to schizophrenia (Brown and Derkits, 2010). Prenatal immune challenge by bacterial endotoxin lipopolysaccharide (LPS) or polyriboinosinic-polyribocytidylic acid (poly I:C) resulted in deficits in PPI (Cardon *et al.*, 2010; Romero *et al.*, 2010). Thus, it has been hypothesized that there are genetic variants that are related to PPI deficits in patients with schizophrenia. Genetic variations in the serotonin-2A receptor, Catechol O-methyltransferase and neuregulin-1 genes have been associated with PPI in schizophrenia (Quednow *et al.*, 2008, 2010; Hong *et al.*, 2008). Thus, we also analyzed the association between the identified SNPs in the *RELA* gene and PPI in patients with schizophrenia.

MATERIALS AND METHODS

Subjects

The subjects of our genetic association study consisted of 1224 patients with schizophrenia (50.9% male (623/601), mean age \pm SD: 46.2 \pm 15.0 years) and 1663 healthy controls (46.5% male (773/890), mean age \pm SD: 46.9 \pm 20.7 years). The mean age did not differ significantly between the groups ($Z = -1.10$, $p = 0.27$), while the sex ratio differed significantly between the groups ($\chi^2 = 5.51$, $p = 0.019$). All the subjects were biologically unrelated Japanese individuals. Patients were recruited at the National Center Hospital of Neurology and Psychiatry, Showa University School of Medicine, Fujita Health University School of Medicine and Osaka University Graduate School of

Medicine. Cases were recruited from both outpatients and inpatients at the hospitals. Each patient with schizophrenia had been diagnosed by at least two trained psychiatrists according to the criteria of the *Diagnostic and Statistical Manual of Mental Disorders*, fourth edition (DSM-IV), on the basis of unstructured clinical interviews. When the diagnosis of two trained psychiatrists was discordant, they started to discuss the diagnosis. When the disputes about the diagnosis were resolved and the patient was diagnosed as schizophrenia, we included the patients. When the disputes were not resolved by discussion or the patients were not diagnosed as schizophrenia, we excluded the patients. Controls, including hospital and institutional staff, were recruited through local advertisements. Psychiatrically healthy controls were evaluated using unstructured interviews to exclude individuals who had current or past contact with psychiatric services, who had received psychiatric medication or who were not Japanese. We did not assess the controls for the family history of mental disorders, such as schizophrenia, bipolar disorder, or major depressive disorder. The ethnicity was determined by the self-report and it was not confirmed by genetic analyses.

Data for the PPI analysis were available for 53 patients with schizophrenia (56.6% males (30/23), mean age \pm SD: 39.1 \pm 13.2 years). The subjects included in the PPI analysis met additional criteria. All subjects were recruited at Osaka University, and subjects were excluded from this study if they had neurological or medical conditions that could potentially affect the central nervous system, such as atypical headaches, head trauma with loss of consciousness, chronic lung disease, kidney disease, chronic hepatic disease, thyroid disease, active cancer, cerebrovascular disease, epilepsy, seizures, substance-related disorders, or mental retardation. None of the subjects reported history of any known hearing impairment and all participants were able to clearly detect 70 dB noise. Written informed consent was obtained from all subjects after the procedures had been fully explained. This study was performed in accordance with the World Medical Association's Declaration of Helsinki and approved by the institutions' ethical committees (National Center Hospital of Neurology and Psychiatry, Showa University School of Medicine, Fujita Health University School of Medicine and Osaka University Graduate School of Medicine).

SNP Selection and Genotyping

Venous blood was drawn from the subjects, and genomic DNA was extracted from whole blood according to standard procedures. We used the Tagger program (Paul de Bakker, <http://www.broad.mit.edu/mpg/tagger>) of the Haploview software (ver. 4.1) (Barrett *et al*, 2005) to select SNPs in the HapMap database (release #22/phase II, April 2007, www.hapmap.org, population: Han Chinese in Beijing (CHB) + Japanese in Tokyo (JPT); minor allele frequencies (MAFs) of more than 0.05) that covered the *RELA* gene spanning 8.5 Kb (5'-flanking regions including approximately 2 kb from the first exon and approximately 0.5 kb downstream (3') from the last exon; HapMap database contig number chr11: 65178000.65189000). The criterion for detecting tag SNPs was an r^2 threshold greater than 0.80 in

'pair-wise tagging only' mode. Two tag SNPs were selected (rs2306365 and rs11820062) by the Tagger program among five SNPs in the HapMap database in the *RELA* gene region. We also searched putative functional SNPs, which are located in exons, exon-intron boundaries and putative promoter regions (5'-flanking region including approximately 2 kb from the first exon and 3' region approximately 1 kb from first exon). We only find one SNP (rs11820062) fulfilled the criteria, which was already selected by Tagger program. Because these two SNPs were located in the 5' region, we added two SNPs (rs11568300 and rs7119750) on the 3' region in this gene for better coverage (2.2 kb per SNP) (Figure 1). The four selected SNPs (rs7119750 (SNP1), rs11568300 (SNP2), rs2306365 (SNP3), and rs11820062 (SNP4)) in the *RELA* gene were genotyped using the TaqMan 5'-exonuclease allelic discrimination assay (Applied Biosystems, Foster City, CA) as described previously (Hashimoto *et al*, 2006, 2007; Ohi *et al*, 2009). The positions of the four SNPs analyzed in the present study are indicated in Figure 1. Primer and probe sequences for detection of the SNPs are as follows: SNP1: forward primer 5'-GCTCAGGCTCAATCCCTCTCTA-3', reverse primer 5'-CCTACAGGCTGGGTCAGATG-3', probe 1 VIC-ACTGCCAA CACCC, probe 2 FAM-CACTGCTAACACCC; SNP2: forward primer 5'-GGTGTGCGCAGAGAAGCA-3', reverse primer 5'-CCTTCTCCATGCAGCTGTCT-3', probe 1 VIC-CACACT GGCTCCG, probe 2 FAM-CACAGTGGCCTCCG; SNP3: forward primer 5'-GCCAAGAAAACAGGCGATCAG-3', reverse primer 5'-CCTCCTCTAGGACTTGTGTTTTCAC-3', probe 1 VIC-CCCTCCCAGTGCAGAG, probe 2 FAM-CCT CCCAGCGCAGAG; SNP4: forward primer 5'-CGCATCTG ATTCACTTCTCTCTCT-3', reverse primer 5'-AATCAGG GCCTGTTGACTTTTCTT-3', probe 1 VIC-CTCCCTCAAT TTTCTT, probe 2 FAM-TCCCTCAGTTTTCTT.

In Silico Analysis to Identify SNPs Associated with *RELA* Expression (eQTLs)

To identify whether the SNPs in the *RELA* gene might be expression quantitative trait loci (eQTLs), we used GeneVar (<http://www.sanger.ac.uk/humgen/genevar/>). Genotype and gene lymphoblastoid expression data from multiple HapMap ethnic samples such as Japanese, Han Chinese, Utah residents with Northern and Western European ancestry from the CEPH collection, and Yoruban in Ibadan, Nigeria are deposited in GeneVar. Users could not access the original genotype, gene expression and demographic data in each individual; however, users are able to analyze the association between genotype and gene expression by the WGAViewer software as described by the group that developed GeneVar (Stranger *et al*, 2007). WGAViewer software is able to perform correlational analysis with number of allele as a continuous variable (allele dose effect: 1/1 = 0, 1/2 = 1, 2/2 = 2). However, any other statistical models such as categorical analysis such as dominant and recessive are not able to perform by the software. We searched for potential transcription factor binding sites in the sequence that included SNP4 with the Patch 1.0 pattern search program and the TRANSFAC 6.0 public site (<http://www.gene-regulation.com/cgi-bin/pub/programs/patch/bin/patch.cgi>).

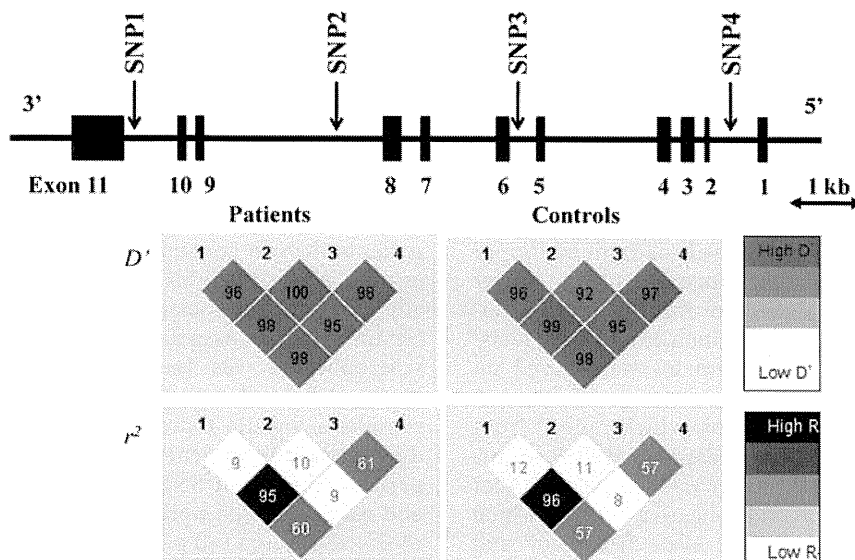


Figure 1 Genomic structure of *RELA*, including locations of the four SNPs studied and linkage disequilibrium of these four SNPs in the patient and control groups. The genomic structure of *RELA* is based on an entry in the Entrez Gene database (National Center for Biotechnology Information). The locations of the SNPs analyzed in this study are indicated with arrows. The distances of the exons–introns and intermarkers are drawn to scale. The linkage disequilibrium (LD) between pairwise SNPs, using D' and r^2 values, are shown at the bottom of the gene structure map separately for 1224 cases and 1663 controls. High levels of LD are represented by increasing gray scale intensity from 0 to 100, as shown by the bars.

Startle Response Measurement

A computerized human startle response monitoring system (Startle Eyeblink Reflex Analysis System Map1155SYS, NIHONSANTEKU, Osaka, Japan) was used to measure PPI. The methods for the startle paradigm, eyeblink acquisition, scoring parameters, and the procedure are described in detail elsewhere (Takahashi *et al*, 2008, 2010; Moriwaki *et al*, 2009). The startle paradigm was a total of 44 trials that consisted of three blocks with a continuous 70 dB sound pressure level (SPL) background white noise. Pulse stimuli consisted of broadband white noises at 115 dB SPL with an instantaneous rise/fall time of 40 ms. The pre-pulse stimuli were also broadband white noises with an instantaneous rise/fall time of 20 ms presented at three different intensities (82, 86, and 90 dB SPL). The lead interval (from pre-pulse onset to pulse onset) was 120 ms. In block 1, the startle response for the pulse alone trial (PA trial) was recorded six times. Block 2 consisted of PA trials or trials of pulse with pre-pulse at the three different intensities (PP trials) performed eight times for each condition. Block 3 was the same as block 1 to observe the habituation phenomenon. All trials were presented in a fixed pseudorandom order and were separated by inter-trial intervals of 15–25 s (20 s on average). The session lasted approximately 20 min, including 5 min of acclimation to the background noise. The following startle measures were calculated: (i) for the acoustic startle reflex, the average eyeblink amplitude of startle response to PA trials in block 1; (ii) habituation of the startle response during the session, calculated as the percentage of amplitude reduction between blocks 1 and 3 with the formula ((1–average eyeblink amplitude of the startle response in block 3/average eyeblink amplitude of the startle response in block 1) \times 100); and (iii) PPI82, PPI86, PPI90, the pre-pulse inhibitions at

intensities of 82 dB, 86 dB, and 90 dB SPL, respectively. The PPI for each intensity level was calculated as the percentage of the amplitude reduction between the PA and PP trials in block 2 with the following formula: (1–average eyeblink amplitude of the startle response in the PP trials in block 2/average eyeblink amplitude of the startle response in the PA trials in block 2) \times 100.

Statistical Analyses

Differences in clinical characteristics between patients and controls or between genotype groups were analyzed using the χ^2 -test for categorical variables and the Mann–Whitney *U*-test for continuous variables using the PASW Statistics 18.0. software (SPSS Japan, Tokyo, Japan). Statistical analyses for genetic association were performed using the SNPalyze v5.1.1 Pro software (DYNACOM, Yokohama, Japan). A logistic regression analysis (forced entry method) was conducted to examine the independent association of the sex (1: male, 2: female) and each genotype (0:M/M, 1:M/m, 2:m/m; M:major allele, m:minor allele) on the categorical diagnosis of schizophrenia (0: control, 1: patient). Sex and genotype statuses were included in the model as independent variables, and diagnosis was included as dependent variables. Deviation from the Hardy-Weinberg equilibrium (HWE) was tested separately in cases and controls using χ^2 -tests for goodness of fit. The allelic and genotypic distributions of *RELA* polymorphisms between patients and controls were analyzed using χ^2 -tests. The number of effective independent SNPs assayed was estimated to correct for multiple testing by the spectral decomposition method of Nyholt using the SNPSpD software (Nyholt, 2004). Haplotype frequencies were estimated by the maximum likelihood method using the genotyping data and the

expectation-maximization algorithm. Rare haplotypes found in less than 3% of both patients and controls were excluded from the association analysis. We performed 10 000 permutations for the most significant tests to determine the empirical significance. We used a 2–4-window fashion analysis. Pairwise linkage disequilibrium (LD) analyses, expressed by D' and r^2 , were applied to detect the intermarker relationships in each group using the Haploview 4.1 software (<http://www.broad.mit.edu/mpg/haploview/contact.php>). We performed *post hoc* power calculations using the Power Calculator for Two Stage Association Studies (<http://www.sph.umich.edu/csg/abecasis/CaTS> (Skol et al, 2006)). Power estimates were based on the allele frequencies in patients, which ranged from 0.43 (SNP1) to 0.45 (SNP4); the odds ratios, which ranged from 1.16 (SNP1) to 1.23 (SNP4), for each associated SNP in this study and an alpha level of 0.05. Power was calculated under a prevalence of 0.01 using a multiplicative model, which assumed varying degrees of marker allele frequency and odds ratios. The effects of the RELA genotypes on the PPI in patients with schizophrenia were analyzed by one-way analysis of covariance (ANCOVA) to adjust for possible confounding factors (gender, current smoking status, and age) using the PASW Statistics 18.0 software (Swerdlow et al, 2008). Standardized effect sizes were calculated using Cohen's d method (<http://www.uccs.edu/faculty/lbecker>). All p -values reported are two tailed. Statistical significance was defined as $p < 0.05$.

RESULTS

Genetic Association Analysis

The genotype and allele frequencies of four SNPs located in the RELA gene are summarized in Table 1. The genotyping call rates were 96.4% (SNP1), 99.3% (SNP2), 96.7% (SNP3), and 97.5% (SNP4). No deviation from HWE was detected in the cases or controls ($p > 0.05$). Significant differences in the genotype and allele frequencies between patients and controls were observed for SNP1, SNP3, and SNP4 (SNP1: genotype $\chi^2 = 7.1$, $p = 0.028$, allele $\chi^2 = 7.0$, $p = 0.0080$; SNP3: genotype $\chi^2 = 8.6$, $p = 0.014$, allele $\chi^2 = 8.8$, $p = 0.0031$; SNP4: genotype $\chi^2 = 14.7$, $p = 0.00064$, allele

$\chi^2 = 14.9$, $p = 0.00011$). These associations remained significant even after the SNPSpD correction for multiple SNP tests, except for the genotypic association for SNP1 (the effective number of independent marker loci: 2.76: corrected p values, SNP1: genotype 0.078, allele 0.0021; SNP3: genotype 0.039, allele 0.0086; SNP4: genotype 0.0018, allele 0.00030). The frequencies of the C allele of SNP1, the G allele of SNP3 and the T allele of SNP4 were higher in patients than in controls. There was no allelic or genotypic association between SNP2 and schizophrenia. We additionally performed association analyses in males and females, separately. The association between the RELA gene and schizophrenia was found in males but not in females (Table 2). As the gender ratio was not matched in this sample, a logistic regression analysis (dependent variable: diagnosis, independent variables: genotype and gender) was performed for four SNPs. This analysis showed that the sex and each genotype were significant predictors for diagnosis of schizophrenia (all $p < 0.05$, except for genotype of SNP2 ($p = 0.19$) (Supplementary Table S1). Diagnosis of schizophrenia was significantly predicted by sex and genotype (SNP1, SNP3, and SNP4), respectively.

Two-four SNP sliding window haplotype analysis revealed significant association of this gene with schizophrenia (the lowest global $p = 0.00040$, SNP3-SNP4) (Table 3). The differences in the detailed haplotype frequencies between cases and controls are shown in Table 3 (lowest $p = 0.00006$, G-C haplotype compared with other haplotypes of SNP3-SNP4). The LD relationships between markers are provided in Figure 1. As the strong LD pattern observed in patients with schizophrenia was nearly identical to that among our controls and the JPT, CEU, and YRI HapMap samples, the strong LD patterns in this gene were likely to be common among ethnic groups.

In Silico Genotype-Expression Analysis

We analyzed the associations between the SNPs and the expression levels of the RELA gene in lymphoblasts in GeneVar database (Table 4). Unfortunately, data concerning SNP2 was not available in this database. We found that there were significant correlations between the RELA gene expression and all three SNPs associated with schizophrenia

Table 1 Genotypic and Allelic Distributions for SNPs in the RELA Gene Between Patients with Schizophrenia and Controls

Marker	M/m	Location	SCZ			CON			Genotypic p value (df = 2)	SCZ	CON	Allelic p value (df = 1)	OR (95%CI)	
			M/M	M/m	m/m	M/M	M/m	m/m						
SNP1	10728386	C/T	Intron 10	0.32	0.48	0.19	0.29	0.48	0.23	0.028	0.43	0.47	0.008	0.87 (0.78–0.96)
SNP2	10730962	G/C	Intron 8	0.78	0.21	0.0091	0.76	0.23	0.01	0.45	0.12	0.13	0.22	0.90 (0.77–1.06)
SNP3	10733141	G/A	Intron 5	0.32	0.48	0.20	0.28	0.48	0.24	0.014	0.44	0.48	0.0031	0.85 (0.77–0.95)
SNP4	10735731	C/T	Intron 1	0.31	0.48	0.21	0.36	0.47	0.16	0.00064	0.45	0.40	0.00011	1.23 (1.11–1.37)

Abbreviations: CI, confidence interval; CON, controls; M, major allele; m, minor allele; MAF, minor allele frequency; OR, odds ratio; SNP, single-nucleotide polymorphism; SCZ, patients with schizophrenia.

All the alleles are represented according to the minus strand DNA sequence.

P values < 0.05 are in bold and underlined.

^adb SNP build 129.