

Figure 2. *S6 kinase-1 (S6K1) and S6K2 expression in cancer cell lines. A: The mRNA expression levels of S6K1 and S6K2 were determined using real-time RT-PCR. Rel. mRNA: Relative mRNA expression levels (target genes/GAPD $\times 10^3$). B: Western blot analysis for S6K1 and S6K2 protein. The arrows indicate the target proteins. β -actin was used as an internal control.*

percentages of the pathological stages were as follows: stage I, 8%; stage II, 12%, stage III, 29% and stage IV, 51%. Intestinal-type gastric cancer was observed in 42% (89/213) of the histologically-examined gastric carcinomas. The patient age, sex, pathological stage, and histology were not significantly associated with *S6K2*-amplification (Table I). The characteristics of the *S6K2*-amplified gastric carcinomas are summarized in Table II.

Finally, we examined the prognostic impact of *S6K2* amplification on OS after surgery. Although the sample size was relatively small ($n=108$), *S6K2* amplification was associated with a significantly shorter OS, compared with non-amplified cases, among patients with stage IV gastric cancer (log rank test, $p=0.02$; Figure 3C). Thus, *S6K2* amplification may be a novel prognostic factor for gastric cancer.

Discussion

Accumulating evidence demonstrates that the level of phosphorylation of S6K1 protein as detected using immunohistochemistry, which reflects activated mTORC1-S6K1 signaling, is increased in various types of cancer, including breast, lung, melanoma, hepatocellular carcinoma and glioma (3). Most of these studies showed that an increased phospho-S6K1 level was positively correlated with a poor prognosis, such as nodal metastasis and overall survival (3, 13-14). Therefore, the activation and overexpression of S6K1 enhances the malignant potential of the cancer cells. Regarding the gene amplification of *S6K1*, amplification is mostly observed in breast cancer, with a detection frequency of 6% to 14% using a Southern blot

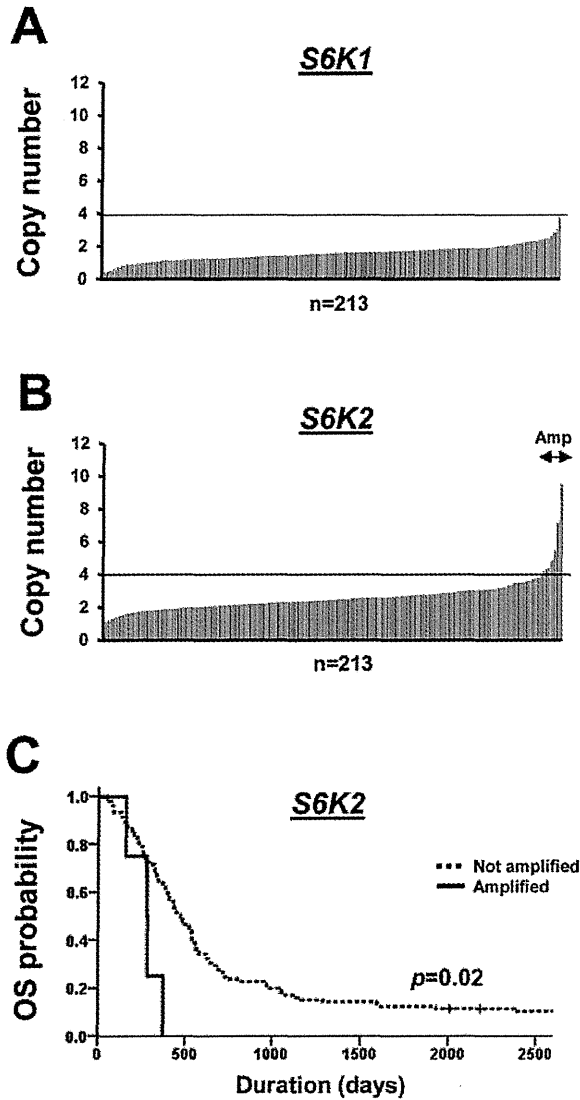


Figure 3. Gene amplification of *S6K1* and *S6K2* in clinical gastric cancer samples. *S6K1* (A) amplification and *S6K2* (B) amplification were determined using the DNA copy number assay for 213 gastric cancer samples. *S6K2* amplification over four copies was observed in 10 cases. C: Kaplan-Meier curves for overall survival (OS) in patients with stage IV gastric cancer. Patients were divided into two groups according to the *S6K2* amplification. Amp, Gene amplification.

analysis, CGH, and fluorescence *in situ* hybridization (15-19). Thus, breast cancer is widely recognized to harbor a *S6K1* amplification in around 10% of samples. For other types of cancers, *S6K1* amplification was observed in three out of 16 (19%) medulloblastomas, in seven out of 38 (18%) diffuse large B-cell lymphoma combinations, and in two out of 17 (8%) ovarian cancer cell lines (20-22). In this study,

Table I. *S6 kinase-2 (S6K2) amplification and clinicopathological features in gastric cancer.*

		S6K2 amplification			p-Value
		Total n=213	+ n=10	- n=203	
Age (years)	Range	31-91	45-75	31-91	0.94
	Median	63	63	63	
Gender	Male	147	7	140	0.78
	Female	66	3	63	
pStage	I	18	0	18	0.70*
	II	25	3	22	
	III	61	3	58	
	IV	108	4	104	
	Unknown	1	0	1	
Histology	Tub1	32	1	31	0.66**
	Tub2	44	1	43	
	Pap	5	0	5	
	Muc	8	1	7	
	Sig	11	2	9	
	Poor1	25	0	25	
Poor2	88	5	83		

Tub, Tubular adenocarcinoma; Pap, papillary adenocarcinoma; Muc, mucinous adenocarcinoma; Sig, signet ring-cell carcinoma; Poor, poorly-differentiated adenocarcinoma; pStage, pathological stage. *Comparison between pStage I+II and III+IV. **Comparison between intestinal (Tub1, Tub2, Pap and Muc) and others. p-Values were calculated using the *t*-test for age and the χ^2 test for the other variables.

we found that the 44As3 gastric cancer cell line harbored *S6K1* amplification, but amplification was not detected in clinical gastric cancer samples, suggesting that *S6K1* amplification is relatively rare in gastric cancer.

A limited number of studies have focused on the molecular and biological function of *S6K2*, despite the great number of studies that have examined *S6K1* to date. Similarly, the dysregulation of *S6K2* in cancer remains largely unknown. A recent study demonstrated that *S6K2* amplification was observed in nine out of 207 (4%) breast carcinomas, whereas the *S6K1* amplification was observed in 22 out of 206 (11%) (23). In addition, the *S6K2* amplification was correlated with a high mRNA expression level and was associated with a poor prognosis (23). Of note, another study from the same research group showed that the chromosomal region of 11q13, which includes the *S6K2* gene, was frequently co-amplified with the chromosomal region of 8p12, which includes another key downstream molecule of mTOR signaling, the eukaryotic translation initiation factor 4E binding protein-1 gene (24). Further study on such co-amplification is needed to understand the dysregulation of mTOR signaling. In addition, *S6K2* amplification may alter sensitivity to mTOR inhibitors, and thus further studies are warranted.

Table II. Summary of patients with *S6 kinase-2 (S6K2)*-amplified gastric cancer.

No.	Age	Gender	Macroscopic type*	Histology	pStage	OS (days)	S6K1 (CN)	S6K2 (CN)
1	45	M	2	Poor2	IIIa	394	1.5	4.1
2	64	M	4	Sig	IV	283	2.5	4.2
3	63	F	0-IIc	Sig	IIIb	4732+	1.8	4.3
4	52	M	2	Tub2	II	3935+	1.2	4.4
5	64	M	3	Tub1	II	1907	1.8	4.8
6	66	M	3	Poor2	IV	157	1.9	4.9
7	71	F	3	Poor2	II	835	1.3	5.5
8	75	M	3	Poor2	IV	280	2.0	7.1
9	73	M	4	Poor2	IV	373	1.7	7.3
10	57	F	3	Muc	IIIb	2742	1.6	9.5

No., Sample number; pStage, pathological stage; OS, overall survival; CN, copy number; *classification is based on the definitions of the Japanese Research Society for Gastric Cancer; + for OS, indicates the patient was still alive at the time of writing. Tub, Tubular adenocarcinoma; Muc, mucinous adenocarcinoma; Sig, signet ring-cell carcinoma; Poor, poorly-differentiated adenocarcinoma; pStage, pathological stage.

In conclusion, we found that the *S6K2* amplification was observed in gastric cancer at a frequency of 4.7%, and its amplification was related to a poor outcome. Our results may provide an insight into the dysregulation of mTOR signaling in gastric cancer.

Conflicts of Interest

None.

Acknowledgements

We thank Miss Tomoko Kitayama and Miss Hideko Morita for their technical assistance. This study was supported by the Third-Term Comprehensive 10-Year Strategy for Cancer Control and a Grant-in-Aid for Cancer Research from the Ministry of Health, Labour and Welfare.

References

- 1 Bittoni A, Maccaroni E, Scartozzi M, Berardi R and Cascinu S: Chemotherapy for locally advanced and metastatic gastric cancer: state of the art and future perspectives. *Eur Rev Med Pharmacol Sci* 14: 309-314, 2010.
- 2 Fujii M, Kochi M and Takayama T: Recent advances in chemotherapy for advanced gastric cancer in Japan. *Surg Today* 40: 295-300, 2010.
- 3 Pópulo H, Lopes JM and Soares P: The mTOR Signalling Pathway in Human Cancer. *Int J Mol Sci* 13: 1886-1918, 2012.
- 4 Sarbassov DD, Ali SM, Kim DH, Guertin DA, Latek RR, Erdjument-Bromage H, Tempst P and Sabatini DM: Rictor, a novel binding partner of mTOR, defines a rapamycin-insensitive and raptor-independent pathway that regulates the cytoskeleton. *Curr Biol* 14: 1296-1302, 2004.
- 5 Nojima H, Tokunaga C, Eguchi S, Oshiro N, Hidayat S, Yoshino K, Hara K, Tanaka N, Avruch J and Yonezawa K: The mammalian target of rapamycin (mTOR) partner, raptor, binds the mTOR substrates p70 S6 kinase and 4E-BP1 through their TOR signaling (TOS) motif. *J Biol Chem* 278: 15461-15464, 2003.

- 6 Magnuson B, Ekim B and Fingar DC: Regulation and function of ribosomal protein S6 kinase (S6K) within mTOR signaling networks. *Biochem J* 441: 1-21, 2012.
- 7 Pearce LR, Komander D and Alessi DR: The nuts and bolts of AGC protein kinases. *Nat Rev Mol Cell Biol* 11: 9-22, 2010.
- 8 Faivre S, Kroemer G and Raymond E: Current development of mTOR inhibitors as anticancer agents. *Nat Rev Drug Discov* 5: 671-688, 2006.
- 9 Dancey J: mTOR signaling and drug development in cancer. *Nat Rev Clin Oncol* 7: 209-219, 2010.
- 10 Matsumoto K, Arao T, Hamaguchi T, Shimada Y, Kato K, Oda I, Taniguchi H, Koizumi F, Yanagihara K, Sasaki H, Nishio K and Yamada Y: *FGFR2* gene amplification and clinicopathological features in gastric cancer. *Br J Cancer* 106: 727-732, 2012.
- 11 Matsumoto K, Arao T, Tanaka K, Kaneda H, Kudo K, Fujita Y, Tamura D, Aomatsu K, Tamura T, Yamada Y, Saijo N and Nishio K: mTOR signal and hypoxia-inducible factor-1 alpha regulate CD133 expression in cancer cells. *Cancer Res* 69: 7160-7164, 2009.
- 12 Furuta K, Arao T, Sakai K, Kimura H, Nagai T, Tamura D, Aomatsu K, Kudo K, Kaneda H, Fujita Y, Matsumoto K, Yamada Y, Yanagihara K, Sekijima M and Nishio K: Integrated analysis of whole-genome exon array and array-comparative genomic hybridization in gastric and colorectal cancer cells. *Cancer Sci* 103: 221-227, 2012.
- 13 Dobashi Y, Suzuki S, Kimura M, Matsubara H, Tsubochi H, Imoto I and Ooi A: Paradigm of kinase-driven pathway downstream of epidermal growth factor receptor/AKT in human lung carcinomas. *Hum Pathol* 42: 214-226, 2011.
- 14 Zhou L, Huang Y, Li J and Wang Z: The mTOR pathway is associated with the poor prognosis of human hepatocellular carcinoma. *Med Oncol* 27: 255-261, 2010.
- 15 Couch FJ, Wang XY, Wu GJ, Qian J, Jenkins RB and James CD: Localization of PS6K to chromosomal region 17q23 and determination of its amplification in breast cancer. *Cancer Res* 59: 1408-1411, 1999.
- 16 Bärlund M, Forozan F, Kononen J, Bubendorf L, Chen Y, Bittner ML, Thorhorst J, Haas P, Bucher C, Sauter G, Kallioniemi OP and Kallioniemi A: Detecting activation of ribosomal protein S6 kinase by complementary DNA and tissue microarray analysis. *J Natl Cancer Inst* 92: 1252-1259, 2000.

- 17 Wu GJ, Sinclair CS, Paape J, Ingle JN, Roche PC, James CD and Couch FJ: 17q23 amplifications in breast cancer involve the *PAT1*, *RAD51C*, *PS6K*, and *SIGmalB* genes. *Cancer Res* 60: 5371-5375, 2000.
- 18 Adem C, Soderberg CL, Hafner K, Reynolds C, Slezak JM, Sinclair CS, Sellers TA, Schaid DJ, Couch F, Hartmann LC and Jenkins RB: *ERBB2*, *TBX2*, *RPS6KB1*, and *MYC* alterations in breast tissues of BRCA1 and BRCA2 mutation carriers. *Genes Chromosomes Cancer* 41: 1-11, 2004.
- 19 Bärlund M, Monni O, Kononen J, Cornelison R, Torhorst J, Sauter G, Kallioniemi OLLI-P and Kallioniemi A: Multiple genes at 17q23 undergo amplification and overexpression in breast cancer. *Cancer Res* 60: 5340-5344, 2000.
- 20 Ehrbrecht A, Müller U, Wolter M, Hoischen A, Koch A, Radlwimmer B, Actor B, Mincheva A, Pietsch T, Lichter P, Reifemberger G and Weber RG: Comprehensive genomic analysis of desmoplastic medulloblastomas: Identification of novel amplified genes and separate evaluation of the different histological components. *J Pathol* 208: 554-563, 2006.
- 21 Zhao MY, Auerbach A, D'Costa AM, Rapoport AP, Burger AM, Sausville EA, Stass SA, Jiang F, Sands AM, Aguilera N and Zhao XF: Phospho-p70S6K/p85S6K and cdc2/cdk1 are novel targets for diffuse large B-cell lymphoma combination therapy. *Clin Cancer Res* 15: 1708-1720, 2009.
- 22 Watanabe T, Imoto I, Kosugi Y, Ishiwata I, Inoue S, Takayama M, Sato A and Inazawa J: A novel amplification at 17q21-23 in ovarian cancer cell lines detected by comparative genomic hybridization. *Gynecol Oncol* 81: 172-177, 2001.
- 23 Pérez-Tenorio G, Karlsson E, Waltersson MA, Olsson B, Holmlund B, Nordenskjöld B, Fornander T, Skoog L and Stål O: Clinical potential of the mTOR targets S6K1 and S6K2 in breast cancer. *Breast Cancer Res Treat* 128: 713-723, 2011.
- 24 Karlsson E, Waltersson MA, Bostner J, Pérez-Tenorio G, Olsson B, Hallbeck AL and Stål O: High-resolution genomic analysis of the 11q13 amplicon in breast cancers identifies synergy with 8p12 amplification, involving the mTOR targets S6K2 and 4EBP1. *Genes Chromosomes Cancer* 50: 775-787, 2011.

Received November 26, 2012
Revised December 26, 2012
Accepted January 3, 2013

