Table 1 Toxicity of NACRT

		0	1	2	3	4	G1–G4 (%)	G3–G4 (%)
Hematologic	Leucopenia	29	1	3	1	0	15	3.0
	Neutropenia	32	0	2	0	0	6.0	0
	Anemia	33	1	0	0	0	3.0	0
	Thrombocytopenia	34	0	0	0	0	0	0
Non-hematologic	Nausea	33	1	0	0	0	3.0	0
	Vomiting	33	0	1	0	0	3.0	0
	Anorexia	21	5	7	1	0	38	3.0
	Diarrhea	33	0	1	0	0	3.0	0
	Fatigue	27	0	6	1	0	21	3.0
	Weight loss	25	7	2	0	0	26.4	0
	Gastric ulcer	33	0	1	0	0	3.0	0
	DVT	33	0	1	0	0	3.0	0
	Skin rash	33	0	1	0	0	3.0	0
	Fever	26	8	0	0	0	23.5	0
	Stent trouble	28	0	0	7	0	20.8	20.8
No adverse events $n=15$								

Toxicity was graded according to Common Terminology Criteria for Adverse Events v4.0

DVT deep vein thrombosis

approximately 20% of patients are indicated for surgical resection. Even after "curative" resection, patients with pancreatic cancer face a 50–80% local recurrence rate and a 25–50% chance of developing distant metastases in the peritoneum and liver, resulting in an actual 5-year survival rate of approximately 10%.[1–5] Recently, some randomized studies have shown favorable results in pancreatic cancer patients who underwent curative resection followed by adjuvant therapy, reporting median survival times within the range of 20.1–23.6 months.[21–23] A systematic review and meta-analysis by Gillen et al. showed that an estimated median survival time of patients with resectable pancreatic cancer who underwent surgical resection following neo-adjuvant therapy was similar to those of patients who had adjuvant therapy.[24] Recently, a few centers have reported better actual survival rate in patients with pancreatic cancer who underwent surgical resection following NACRT. For example, the M.D. Anderson Cancer Center Group showed that the actual 5-year survival rate of patients after multidisciplinary management including surgical resection was 27%,[25] and in patients with resectable pancreatic head cancer who underwent surgical resection following preoperative gemcitabine-based chemoradiation, the actual 5-year survival rate was 36%.[26] Our previous study[9–11] demonstrated that the actual 5-year survival and disease-free survival rates in the pre-CRT group, who did not receive adjuvant chemotherapy, were significantly longer than in the surgery-alone group, in a sub-group analysis of patients who underwent curative resection. In fact, the actual survival curves in these studies demonstrated that a fall of the survival curve within 3 years after surgical resection, plateaued when it passed the 3-year mark. Thus, surgical resection following NACRT can be associated with improvement of long-term survival rate through good local disease control. In our previous study,

Table 2 Clinical background between NACRT and adjuvant groups

Parameter	Adjuvant ($n=36$)	NACRT ($n=30$)	p value
Sex (male/female)	25:11	15:15	0.133
Age (years)[a]	68 (51–81)	65.5 (36–79)	0.107
CA19–9 (U/ml)[a]	127 (6–1,729)	247 (1–2,232)	0.067
Diabetes mellitus (+/−)	10:26	11:19	0.596
Obstructive jaundice (+/−)	30:6	20:10	0.153
Albumin (g/dl)[a]	3.8 (1.9–4.4)	3.6 (2–4.3)	0.286
Hemoglobin (g/dl)[a]	12.1 (7.9–15.2)	11.4 (9.1–13.9)	0.142
Platelet count ($\times 10^4$)[a]	24 (12–43)	21 (13–40)	0.908
PR vs BR/UN	19:17/0	7:21/2	0.022
Stent exchange (+/−)	3:33	7:23	0.089

PR potentially resectable pancreatic cancer, *BR* borderline resectable pancreatic cancer, *UN* unresectable pancreatic cancer

[a]Values are median (range)

Table 3 Comparisons of surgical results between NACRT and adjuvant groups

Parameter	Adjuvant (n=36)	NACRT (n=30)	p value
Extent of blood loss (ml)	999 (324–5,238)	1,376 (438–3,853)	0.151
Op time (min)	514 (210–672)	531 (380–711)	0.146
Op type (PD/PpPD/DP/TP)	22:8:5:1	22:1:6:1	0.112
PV resection (+/−)	14:22	17:13[b]	0.216
CA/CHA resection (+/−)	0:36	2:28	0.203
Blood transfusion (none/auto/allo)	4:23:9	8:13:9	0.166
Location (Ph/Pbt)	31:5	22:8	0.227
Tumor size (mm)	32.5 (23–65)	30 (10–65)	0.341
Numbers of harvested LNs	26 (7–56)	33 (6–65)	0.340
Numbers of metastatic LNs	2 (0–19)	1 (0–25)	0.0363
Lymph node ratio[a]	0.07 (0–0.62)	0.02 (0–0.38)	0.032
N (−/+)	8:28	14:16	0.065
N 0/1:2/3	18:18	23:7	0.041
T 1/2:3/4	0:36	3:27	0.089
R0:1:2	21:13:2	28:2:0	0.005
Evans classification (IIA/IIB/III)	N/E	21:7:2	

TNM classification was defined by Japanese Pancreas Society

NACRT neo-adjuvant chemoradiation therapy, *Op* operation, *PD* pancreaticoduodenectomy, *PpPD* pylorus preserving pancreaticoduodenectomy, *DP* distal pancreatectomy, *TP* total pancreatectomy, *PV* portal vein, *CA* celiac axis, *CHA* common hepatic artery, *auto* autologous blood transfusion, *allo* allogeneic blood transfusion, *Ph* pancreatic head, *Pbt* pancreatic body and tail, *LN* lymph node, *R0* negative microscopic margin, *R1* positive microscopic margin, *R2* positive gross margin

[a] Lymph node ratio is calculated as number of metastatic lymph nodes/harvested lymph nodes

[b] One patient who underwent renal vein resection was included

approximately half the patients who underwent curative resection had disease recurrence at 1 year in both the NACRT and surgery-alone groups. NACRT followed by surgical resection did not have enough power to improve the short-term survival rate and the frequency of early liver metastases, which was one of the major post-operative recurrence sites.

Table 4 Comparison of morbidity and mortality between NACRT and adjuvant groups

Parameter	Adjuvant	NACRT	p value
Overall complication (+/−)	12:24	10:20	1.000
Mortality (+/−)	0:36	3:27	0.098
Re-operation/no re-operation	0:36	1:29	0.455
DGE (+/−)	3:33	2:28	1.000
POPF (+/−)	7:29	1:29	0.063
Grade A/B/C	4:3:0	1:0:0	0.245
Wound dehiscence (+/−)	4:32	6:24	0.492
Intra-abdominal abscess (+/−)	1:35	1:29	1.000
Cholangitis (+/−)	0:36	2:28	0.203
Pneumonia (+/−)	0:36	2:28	0.203
Bile leakage (+/−)	0:36	0:30	–
PPH (+/−)	0:36	0:30	–
Intractable ascites (+/−)	2:34	8:22	0.035
Diarrhea (+/−)	5:31	9:21	0.138

DGE delayed gastric emptying, *NACRT* neo-adjuvant chemoradiation therapy, *POPF* post-operative pancreatic fistula, *PPH* post-pancreatectomy hemorrhage

There were several limitations and issues with our previous study that we aimed to resolve in this present study, namely (1) approximately 20% of patients who received pre-operative CRT did not undergo surgical resection because of progressive disease, resulting in a median survival time of 5.5 months (unpublished data); (2) surgical resection followed by pre-operative CRT only did not improve the short-term results; and (3) the previous regimen of pre-CRT was not aggressive enough to achieve tumor shrinkage. Therefore, we introduced (1) staging laparoscopy before patient recruitment to the new regimen of NACRT, (2) standard adjuvant chemotherapy, and (3) full dose of S-1 (80 mg/m^2) and radiotherapy (50.4 Gy).

S-1 is an orally administered drug, which is a combination of tegafur, 5-chloro-2,4-dihydroxypyridine, and oteracil potassium. Very recently, the results of the gemcitabine and S-1 trial study (a randomized, prospective, open-label, three-arm, and phase III study) were presented to the public at the annual meeting of the American Society of Clinical Oncology 2011.[27] The results showed that oral S-1 provided

Table 5 Surgical results of neo-adjuvant chemoradiation therapy

Authors (reference number)	Year of publication	No of patients	Regimen of CRT	Resection rate; *n* (%)	Vascular resection rate; *n* (%)	R0 (%)	Negative LN mets rate (%)
White et al.[33]	2001	53	5-FU based (45 Gy)	28 (53)	2 (7)	71	19 (70)
Moutardier et al.[34]	2004	61	5-FU based (60 Gy)	40 (66)	5 (13)	95	30 (75)
Evans et al.[26]	2008	86	GEM (30 Gy)	64 (74)	13 (20)	89	40 (63)
Le Scodan et al.[35]	2009	41	5-FU based (50 Gy)	26 (63)	N/A	80.7	12 (46)
Ohigashi et al.[36]	2009	38	GEM (50.4 Gy)	31 (82)	17 (55)	97	28 (90)
Turrini et al.[37]	2010	34	Docetaxel-based (45)	17 (50)	N/A	100	13 (76)
Stokes et al.[38]	2011	40	Capecitabine (50 Gy)	16 (25)	4 (25)	88	13 (81)
Present study	–	34	S-1 (50 Gy)	30 (88)	17 (57)	94	14 (47)

CRT chemoradiation therapy, *5-FU* 5-fluorouracil, *GEM* gemcitabine, *LN* lymph node, *mets* metastasis, *N/A* not available

similar efficacy and tolerable toxicity to gemcitabine when used as first-line treatment for unresectable pancreatic cancer. The response rates of gemcitabine, S-1, and gemcitabine+S-1 were 13.3%, 21.0%, and 29.3%, respectively. In addition to the benefit of the oral drug on its own, the combination of S-1 and radiotherapy has been demonstrated to exert a synergistic effect against 5-FU-resistant cancer xenografts.[28,29] The response rate of CRT using S-1 was around 20% in phase I and II studies in patients with unresectable pancreatic cancer[12,30–32] As expected, our results showed that the response rate and disease control rate of NACRT using S-1 were 18% and 88.0%, respectively. To our knowledge, this is the first study of NACRT using S-1 and concurrent radiation for patients with resectable pancreatic cancer.

Despite the fact that we excluded patients with occult metastasis by using staging laparoscopy before study entry for NACRT, three of 34 patients had occult liver or peritoneal metastasis after NACRT. During the 9 weeks between study entry and surgical resection, approximately 10% of pancreatic cancer patients had progressive disease. In this study, the majority of patients who underwent NACRT using S-1 did not suffer from severe adverse effects, and 33 of 34 patients completed this regimen. However, seven of 34 (21%) NACRT patients required hospitalization because of cholangitis, resulting in a delay of the operation date. The primary endpoint of pathologically curative resection rate in this study showed a statistically significant difference in favor of NACRT over the adjuvant group, despite the fact that the NACRT group had a higher frequency of borderline resectable and unresectable pancreatic cancer cases. In this study, we pathologically examined the cut stump of the nerve plexus and retroperitoneal tissue independently in all cases, which was the main reason for a positive surgical margin (R1 resection). Moreover, there was a tendency for a higher rate of negative lymph node metastasis in the NACRT group than in the adjuvant group, but this did not reach statistical significance because of the small sample size. The frequency of N0 and N1 and number of metastatic lymph nodes in the NACRT group were significantly improved relative to those in adjuvant group. Most studies have reported that predictive factors for prognosis in patients with pancreatic cancer were pathologically curative resection and negative lymph node metastasis.[1–5] In addition to our results, some authors have reported the promising results of a higher rate of R0 resection (70–100%) and lower rate of metastastic lymph nodes (46–90%) in patients who underwent surgical resection following NACRT, as summarized in Table 5.[26,33–38]

We recognize that a limitation of our study was its prospective non-randomized design. Approximately 30% of patients were excluded before staging laparoscopy in the NACRT group due to no pathological evidence of pancreatic cancer, misdiagnosis of bile duct cancer, and so on. In contrast, 43 of 48 patients were included in the adjuvant group; the five excluded patients had T1/2 pancreatic cancer. Consequently, the frequency of borderline resectable pancreatic cancer in the NACRT group was significantly higher than in the adjuvant group. However, irrespective of this one important difference in the baseline characteristics between the two groups, the primary endpoint of this study was still reached.

Although there were no statistical differences in morbidity and mortality between the two groups, three in-hospital deaths were observed in the NACRT group. The common clinical features of these three patients were borderline resectable pancreatic cancer, grade 2 anorexia or fatigue during CRT, and other organ resection including vascular

resection. Gillen et al.[24] reported that in-hospital mortality after neo-adjuvant treatment and tumor resection was estimated at 2.2–6.0% in resectable patients and at 5.1–9.5% in non-resectable patients. In this present study, 23 of 30 (77%) patients in the NACRT group had borderline resectable and unresectable pancreatic cancer, and 17 of 30 (57%) patients underwent pancreatectomy with vascular resection. The patient population in the NACRT group had shifted to an advanced stage. Thus, special attention should be paid to patients with advanced pancreatic cancer who undergo this new type of surgical strategy.

In conclusion, in this study, NACRT using the orally administered drug S-1 resulted in a better response rate than was seen among the patients in the adjuvant group; it was also feasible and safe. Pancreatectomy after NACRT improved the rate of pathologically curative resection and reduced the number and extent of lymph node metastasis. A large-scale randomized controlled trial will be needed to confirm the clinical efficacy of NACRT.

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ORIGINAL ARTICLE

Activation of alpha-smooth muscle actin-positive myofibroblast-like cells after chemotherapy with gemcitabine in a rat orthotopic pancreatic cancer model

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Abstract
Background To investigate the behavior of activated pancreatic stellate cells (PSCs), which express alpha-smooth muscle actin (α-SMA), and pancreatic cancer cells in vivo, we examined the expression of α-SMA-positive myofibroblast-like cells in pancreatic cancer tissue after treatment with gemcitabine (GEM) using a Lewis orthotopic rat pancreatic cancer model.
Methods The effect of GEM on DSL-6A/C1 cell proliferation was determined by cell counting method. The orthotopic pancreatic cancer animals were prepared with DSL-6A/C cells, and treated with GEM (100 mg/kg/weekly, for 3 weeks). At the end of treatment, α-SMA expression, fibrosis, transforming growth factor (TGF)-β1 and vascular endothelial growth factor (VEGF) were evaluated by histopathological and Western blot analyses.
Results DSL-6A/C1 cell proliferation was significantly reduced by co-culturing with GEM in vitro. Survival time of pancreatic cancer animals (59.6 ± 13.4 days) was significantly improved by treatment with GEM (89.6 ± 21.8 days; $p = 0.0005$). Alpha-SMA expression in pancreatic cancer tissue was significantly reduced after treatment with GEM ($p = 0.03$), however, there was no significant difference in Sirius-red expression. Expression of VEGF was significantly reduced by GEM treatment, but the expression of TGF-β1 was not inhibited.
Conclusion GEM may suppress not only the tumor cell proliferation but also suppress PSCs activation through VEGF reduction.

Keywords Pancreatic cancer · Alpha-smooth muscle actin · Myofibroblast-like cell · Chemotherapy · Vascular endothelial growth factor

Introduction

Pancreatic cancer is a pathologically unique tumor that is composed of cancer cells and extremely dense desmoplasia containing extracellular matrix (ECM) protein, myofibroblast-like pancreatic stellate cells (PSCs), and inflammatory cells. Since their discovery in 1998, PSCs have been identified as the major source of ECM proteins found in chronic pancreatitis or pancreatic fibrosis in both experimental animals and humans [1–4]. Quiescent PSCs are activated by inflammatory cytokines or oxidative stress and transformed to myofibroblast-like cells (MCs), which express alpha-smooth muscle actin (α-SMA) [4, 5]. Activated PSCs show markedly increased ECM protein synthesis in response to various stimuli, such as cytokines and growth factors [5, 6].

There has been accumulating evidence of interaction between pancreatic cancer cells and PSCs. Pancreatic cancer cells induce PSC proliferation and ECM production. Conditioned medium from a pancreatic cancer cell line promotes the proliferation of PSCs [7]. Growth factors such as transforming growth factor (TGF)-β1, platelet-derived growth factor (PDGF), and fibroblast growth factor (FGF)-2 secreted by pancreatic cancer cells induce PSC activation [5]. However, most of these studies provided important evidence of stroma–tumor interactions through in vitro or subcutaneous tumor models.

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A recent study showed that activated PSCs may regulate the malignant behavior of pancreatic cancer cells [8]. Fujita et al. [9] investigated the significance of α-SMA expression in pancreatic cancer and the correlation between α-SMA mRNA levels and patient prognosis. Patients with high α-SMA expression had a significantly shorter survival. Thus, α-SMA-positive PSCs are thought to be strongly associated with growth and the microenvironment of pancreatic tumors.

The aim of this study was to investigate the relationship between activated PSCs and cancer cells in pancreatic tumors in vivo. We examined the expression of α-SMA, which is a marker of activated PSCs, in a clinically relevant rat orthotopic pancreatic cancer model after treatment with gemcitabine (GEM).

Materials and methods

Cell lines and culture

The rat ductal pancreatic adenocarcinoma cell line DSL-6A/C1 [10] was purchased from the American Type Culture Collection (Rockville, MD, USA), and cultured in Waymouth's MB 752/1 medium (Gibco, Grand Island, NY, USA). The cell culture medium was supplemented with 10 % heat-inactivated fetal bovine serum (FBS; Gibco), penicillin G (100 U/mL), and streptomycin (100 μg/mL). The cells were incubated in a humidified atmosphere of 5 % CO_2 at 37 °C.

DSL-6A/C1 cell proliferation assay

The effect of GEM on DSL-6A/C1 cell proliferation was determined by cell counting method. A total of 1×10^4 cells were incubated with or without GEM (Gemcitabine hydrochloride JD001, SYNCHEM OHG) at concentrations of 1.0 or 5.0 μM/L. The cell numbers relative to that on day 1 were counted on days 2, 4 and 7.

Laboratory animals

Five-week-old male Lewis rats (LEW/SsN Slc) weighing 100–150 g were purchased from Shimizu Laboratory Supplies Co., Ltd. (Kyoto, Japan). Animals were maintained in microisolator cages in a Specific Pathogen Free animal facility at the Kansai Medical University with autoclaved bedding, food, and water. The rats were maintained on a daily 12-h-light/12-h-dark cycle. All experiments were conducted in accordance with the national guidelines for the care and use of laboratory animals, and the experimental protocol was approved by the Animal Experimentation Committee, Kansai Medical University (No. 06-026).

Animal model of orthotopic pancreatic cancer

The Lewis rat orthotopic pancreatic cancer model was prepared as previously described, with modification [11]. A total of 10^6 DSL-6A/C1 cells were injected subcutaneously into the right flank of Lewis rats anesthetized with Isoflurane (Abbott Japan, Tokyo, Japan) inhalation. The subcutaneous tumors were excised under strict aseptic conditions, when they had reached a size of 15 mm in the largest diameter. The tumors were minced by a scalpel into small fragments of 1 mm^3 in size. Tumor recipient Lewis rats were also anesthetized with Isoflurane inhalation, and their abdomens were opened. Five small tissue pockets were prepared under a microscope in the pancreatic parenchyma as an implantation bed (OME-1000, Olympus, Tokyo, Japan). One tumor fragment was placed into each pancreatic tissue pocket in such a way that the tumor tissue was completely surrounded by pancreatic parenchyma. No sutures or fibrin glue were used to fix the tumor fragments to the recipient pancreas. The pancreas was relocated into the abdominal cavity, which was subsequently closed. Four weeks after orthotopic pancreatic tumor implantation, all animals were anesthetized, and tumor growth was confirmed by relaparotomy in the pancreatic parenchyma without distant metastasis.

Experimental model

All tumor-bearing animals were randomly divided into a control group (no treatment) and a GEM treatment group. The GEM treatment group animals were administered GEM (100 mg/kg) three times weekly through the penile vein. The control animals were administered saline.

Twelve animals in each group were followed for 120 days. Six in each group were sacrificed at 4 weeks for assays. At the time of sacrifice, pancreatic tumor tissue samples were fixed in 10 % buffered formalin for routine histopathology and immunohistochemistry and snap frozen for Western blot analysis.

Histopathological examinations

Routine histopathology

Initial hematoxylin and eosin staining was performed to choose tissue blocks that contained large enough areas of pancreatic carcinoma tissue. Normal pancreas and pancreatic carcinoma tissue were fixed in 10 % buffered formalin, embedded in paraffin, sectioned at 3 μm, and stained with hematoxylin and eosin.

Immunohistochemistry

Paraffin sections were rehydrated and washed in PBS for 5 min three times. Sections were incubated with 1 % H_2O_2 for 30 min to block endogenous peroxidases. To prevent nonspecific binding of antibody, sections were incubated for 30 min at room temperature with a blocking solution containing tris-buffered saline, 1 % bovine serum albumin, and 10 % goat serum. For immunohistochemical analysis, sections were incubated with primary antibody (monoclonal mouse anti-α-SMA antibody; Nichirei Bioscience, Tokyo, Japan) for 1 h at room temperature; monoclonal mouse anti-PCNA (Nichirei Bioscience, Tokyo, Japan) overnight at 4 °C, and then incubated with biotinylated secondary antibody at room temperature. The avidin–biotin–peroxidase immune complex was visualized with 3,3-diaminobenzidine tetrahydrochloride substrate chromogen system (DAKO, Botany, Australia). Sections were counterstained with Mayer's hematoxylin (Sigma) for 5 min.

Picrosirius red staining: The evaluation of fibrosis in the specimens of the four experimental groups was done by the Picrosirius red staining technique [12]. Picrosirius red staining can be used simply as a substitute for van Gieson's stain as a sensitive method for collagen staining. Following deparaffinization and hydration, the sections were stained with Sirius Red F3B (Alfa Aesar, MA, USA).

Quantitative color analysis for immunohistochemistry

The images were visualized with a Nikon ECLIPSE E1000 M microscope and photographed with a Nikon DIGITAL CAMERA DXM1200 (Nikon Corporation, Tokyo, Japan) using Lumina Vision software (version 2.2; Mitani Corporation, Tokyo, Japan). The area of positive immunostained regions per section was determined by computer assisted morphometry. A frame of 4 × 4 mm was marked over the carcinoma area for analysis. Tissues confined within this frame were then scanned automatically. Pictures were loaded individually onto the software interface and the color range of immunostained positive areas was evaluated. The results were expressed as the percentage of positive area in the total scanned surface and the means of the 12 analyzed photomicrographs per animal were calculated.

The number of PCNA-stained cancer cells was assessed by counting in a high-power field. The PCNA labeling index was determined as the percentage of the granulosa cell number with positively stained nuclei to the total cell number in the same fields [13]. The means of 12 randomly selected areas per section were calculated.

Western blot analysis of α-SMA, TGF-β1 and VEGF

Pancreatic tumor tissues (100 mg/rat) from control and experimental rats were minced and incubated on ice for 30 min in 1 mL of ice cold whole-cell lysate buffer (10 mM Tris–HCl, pH 7.4, containing 1 % Triton X-100, 0.5 % Nonidet P-40, 1 mM sodium orthovanadate, 1 mM phenylmethylsulfonylfluoride, and protease inhibitor cocktail, Roche Diagnostics, Mannheim, Germany). The minced tissue was homogenized (KINEMATICA POLYTRON PT1300D, Switzerland) and centrifuged at 16,000×*g* at 4 °C for 15 min. The proteins were then fractionated by SDS-PAGE, electrotransferred to PVDF membranes (Bio-Rad Lab., CA, USA), blotted with the following antibodies, and detected with an ECL blotting detection reagent (GE Healthcare Bio-sciences Corp., NJ, USA). Anti-α-SMA (diluted 1:1000), anti-TGF-β1 (Abcam, MA, USA) (diluted 1:2000) and anti-vascular endothelial growth factor (VEGF) (Santa Cruz Biotechnology, Inc., CA, USA) (diluted 1:200) were used as primary antibodies. β-Tubulin (internal control, CloneTUB2.1, Sigma, Saint Louis, USA) was used to verify equal loading. Experiments were repeated at least three times using different samples.

Statistical analysis

All experiments were performed three to five times. Data are presented as mean ± SEM. Data were analyzed for statistical significance by analysis of variance with post hoc Student's *t* test analysis. Differences in Kaplan–Meier survival curves were evaluated by the log-rank test. These analyses were performed with the assistance of a computer program (JMP 5 Software SAS Campus Drive, Cary, NC, USA). Differences were considered significant at $p < 0.05$.

Results

Effect of gemcitabine on DSL-6A/C1 cell proliferation

DSL-6A/C1 cells were cultured in complete medium to which was added 1.0 or 5.0 μM/L of GEM. DSL-6A/C1 cell proliferation was significantly reduced on day 7 in co-culture with 1.0 μM/L of GEM compared with the control ($p = 0.004$), and DSL-6A/C1 cell proliferation was almost completely blocked by 5.0 μM/L GEM (Fig. 1).

Orthotopic model of rat pancreatic cancer

Four weeks after orthotopic pancreatic tumor implantation, about 70 % of the animals had confirmed tumor growth in pancreatic parenchyma without visible distant metastasis

(Fig. 2a). The mean survival time of the control animals was 59.6 ± 13.4 days. Almost all the animals died from peritoneal dissemination with massive ascites. Histopathologic examination of the pancreatic tumors revealed moderately differentiated ductal-type adenocarcinomas (Fig. 2b).

Fibrosis in pancreatic tumor

Quiescent PSCs are activated and transformed to MCs, which express α-SMA and synthesize ECM. Alpha-SMA-positive MCs and fibrosis were histopathologically evaluated using anti-mouse-α-SMA antibody and Picrosirius red staining. Alpha-SMA-positive MCs and Picrosirius red-positive ECM were significantly increased in pancreatic tumor tissue compared with normal pancreas ($p = 0.005$) (Figs. 5d, 7c).

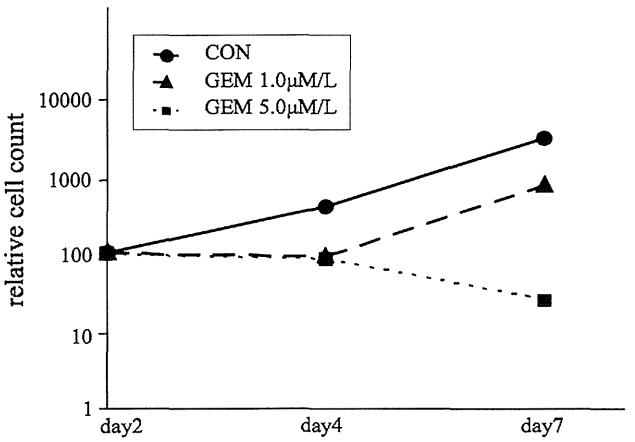

Fig. 1 The effect of GEM on DSL-6A/C1 cell proliferation. DSL-6A/C1 cell proliferation was significantly reduced in co-culture with GEM in a dose-dependent manner ($p = 0.004$). CON: without GEM (*black line*). GEM: 1.0 μM/L (*dashed line*) and 5.0 μM/L (*dotted line*) concentrations of GEM

In vivo anti-tumor effect of gemcitabine treatment

The mean animal survival time of the rat pancreatic cancer model (59.6 ± 13.4 days) was significantly improved by treatment with GEM (89.6 ± 21.8 days; $p = 0.0474$) (Fig. 3). The number of PCNA-stained cancer cells in pancreatic cancer tissue was significantly increased compared with normal pancreatic tissue (Fig. 4a, b). After treatment with gemcitabine, the number of PCNA-stained cancer cells was significantly reduced but still higher than that in normal pancreatic tissue (Fig. 4c).

Alpha-SMA-positive cells and extracellular matrix after gemcitabine treatment

Alpha-SMA immunoreactivity was markedly detected in the stroma of pancreatic cancer tissue. The immunohistochemistry showed that α-SMA expression in pancreatic cancer tissue was significantly reduced after treatment with GEM ($p = 0.03$) (Fig. 5). Moreover, Western blot analysis of α-SMA protein in pancreatic cancer tissue was also significantly reduced by the treatment ($p = 0.001$) (Fig. 6). However, there was no significant difference in Picrosirius red expression in pancreatic cancer tissue between control and GEM-treated animals (Fig. 7).

Growth factor expression in pancreatic tumor

Vascular endothelial growth factor (VEGF) and TGF-β were almost undetectable in normal pancreas by Western blot, but they were strongly expressed in pancreatic cancer tissue. However, the expression of VEGF was significantly reduced by GEM treatment ($p = 0.01$), whereas the expression of TGF-β was not inhibited ($p = 0.41$) (Fig. 8).

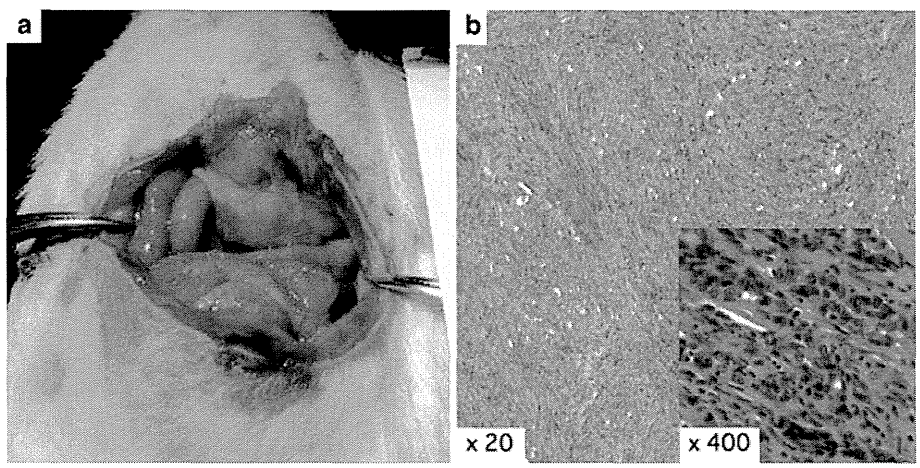

Fig. 2 Gross and microscopic features of orthotopic pancreatic cancer tissue. **a** Gross tumor nodules (4 weeks after implantation). **b** Pancreatic cancer tissue (4 weeks) (×20, ×400, H&E stain)

Discussion

During the last decade, various reports on the interaction between pancreatic cancer cells and PSCs have been published. Growth factors, such as FGF-2, TGF-β1, PDGF, and VEGF, which are secreted by pancreatic cancer cells, activate PSCs. Apte et al. [14] reported that exposure of PSCs to pancreatic cancer cell secretions in vitro resulted in PSC activation, as indicated by significantly increased cell proliferation and α-SMA expression. Furthermore, Bachem et al. [7] reported that pre-incubation of carcinoma cell supernatant with neutralizing antibodies against FGF-2, TGF-β1, and PDGF significantly reduced the PSCs stimulatory effect.

Although the above-mentioned studies provided important evidence of stroma–tumor interactions, most of the studies provided the evidence through subcutaneous tumor models or in vitro studies. Several studies using orthotopic pancreatic cancer models have been published in recent years. Most of these models used xenografted, immunoincompetent, or transgenic animals. The rat model that we used in this study was established by Hotz et al. [11], and they reported on a preclinical treatment study using this model [15]. This animal model is immunocompetent and clinically relevant, because syngeneic rat pancreatic cancer cells were orthotopically implanted. Our study showed that the DSL-6A/C1 cells were sensitive to GEM, and that the survival rate of this pancreatic cancer animal model was increased by treatment with GEM.

Interestingly, the expression of α-SMA in the pancreatic cancer tissue was significantly decreased by treatment with GEM. It is considered likely that the activity of α-SMA-positive MCs was inhibited, as demonstrated by the results of immunohistochemistry and Western blot. Because we initially grew the tumor in the subcutis before transplanting it into pancreas parenchyma in this pancreatic cancer model, MCs derived from the subcutis may have been included. However, as the tumor grew, the PSCs are also

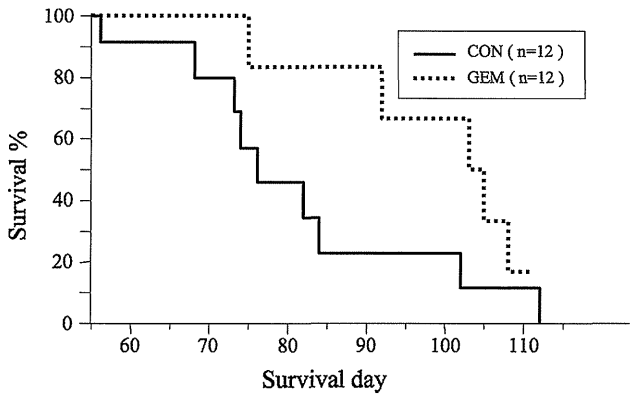

Fig. 3 Overall survival curve for rats with orthotopic pancreatic cancer. The mean animal survival time of the rat pancreatic cancer model (59.6 ± 13.4 days: *black line*) was significantly improved by treatment with GEM (89.6 ± 21.8 days: *dotted line*) ($p = 0.0474$)

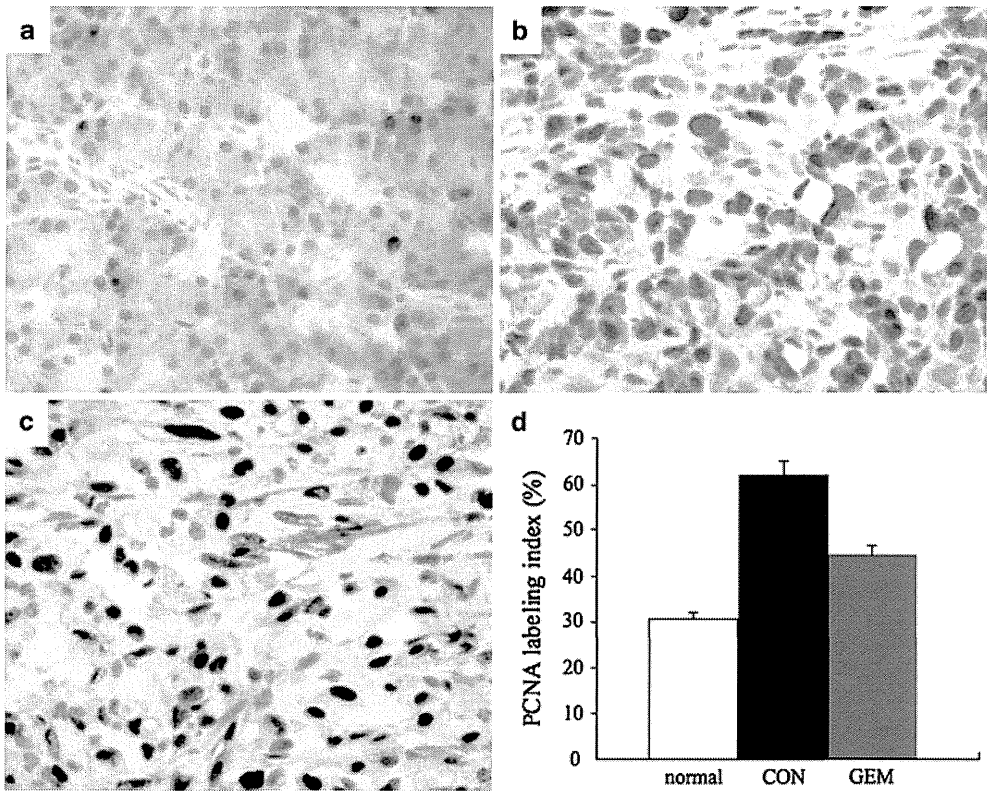

Fig. 4 Immunostaining for PCNA of the pancreatic tissue. PCNA (*brown*) showed the proliferating cells during the late G1 to S phase of the cell cycle. **a** Normal rat pancreatic tissue. **b** No treatment group (control). **c** GEM-treated group (×400). **d** PCNA labeling index shows that the number of PCNA-stained cancer cells after treatment with GEM (*gray bar*) was significantly reduced compared with the control (*black bar*), which still has a higher range than normal pancreatic tissue (*white bar*) (color figure online)

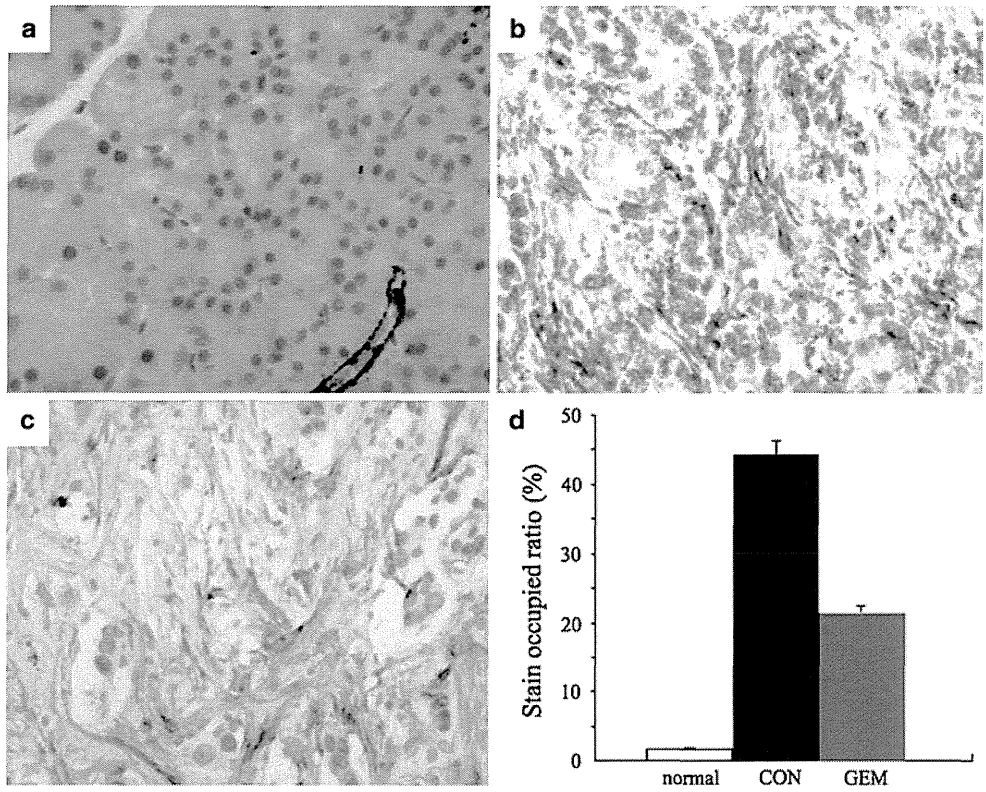

Fig. 5 Immunostaining for α-SMA of the pancreatic tissue. **a** Normal rat pancreatic tissue. **b** No treatment group (control). **c** GEM-treated group (×400). **d** Stain occupied ratio shows that α-SMA expression in pancreatic cancer tissue (*black bar*) was significantly reduced after treatment with GEM (*gray bar*) ($p = 0.03$)

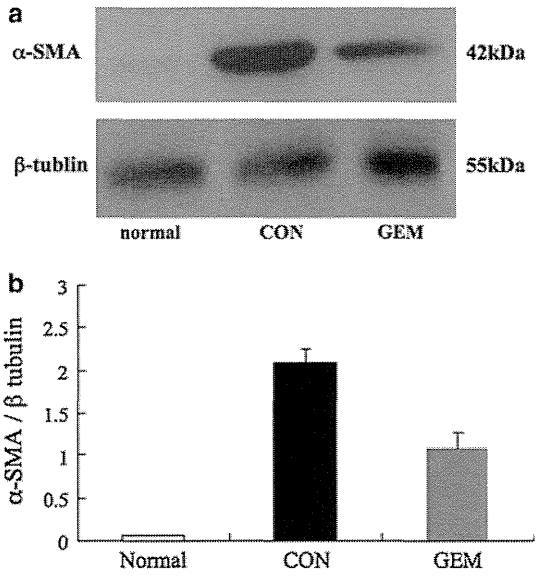

Fig. 6 Western blot analysis of α-SMA protein expression in pancreatic cancer tissue. **a** Western blot analysis of α-SMA protein. *Normal* normal rat pancreatic tissue; *CON* no treatment group (control); *GEM* GEM treated group. **b** Band intensity was quantified and expressed by ratio to β-tubulin ($n = 3$ for each group). Alpha-SMA protein of expression in pancreatic cancer tissue was significantly reduced by treatment with GEM ($p = 0.001$)

thought to migrate into the tumor from pancreatic parenchyma. Erkan et al. [16] have shown that human PSCs are practically resistant to GEM. We have been unable to discover any reports in Medline on direct GEM inhibition of rat PSC activation, but a main reason for this may be that the growth factor secretion by a tumor decreases when the growth of the tumor is being suppressed. We considered that inhibition of activation of α-SMA-positive MCs is not a direct effect of GEM.

Vascular endothelial growth factor plays an important role in tumor angiogenesis. Several reports have demonstrated that patients with pancreatic cancer showing high VEGF expression have significantly shorter survival than patients with lower VEGF expression [17–19]. In this study, we evaluated TGF-β and VEGF, which are well known growth- and PSC-activating factors secreted by pancreatic cancer cells. The expression of TGF-β was not inhibited, but VEGF expression was significantly reduced by GEM treatment. Although pancreatic cancer cells can secrete VEGF, PSCs is one of the principal sources of VEGF in pancreatic cancer tissue [20, 21]. In vitro, hypoxia increased PSC activity and doubled the secretion of periostin, type I collagen, fibronectin, and VEGF [22]. Therefore, we suggest that a decrease of VEGF expression in cancer tissue is caused by an anti-tumor effect of GEM and that inhibition of activation of α-SMA-positive MCs is a secondary effect of tumor cytokines.

Pancreatic cancer is a pathologically unique tumor that is composed of cancer cells and extremely dense desmoplasia containing ECM protein, activated PSCs, and inflammatory cells. Activated PSCs show markedly

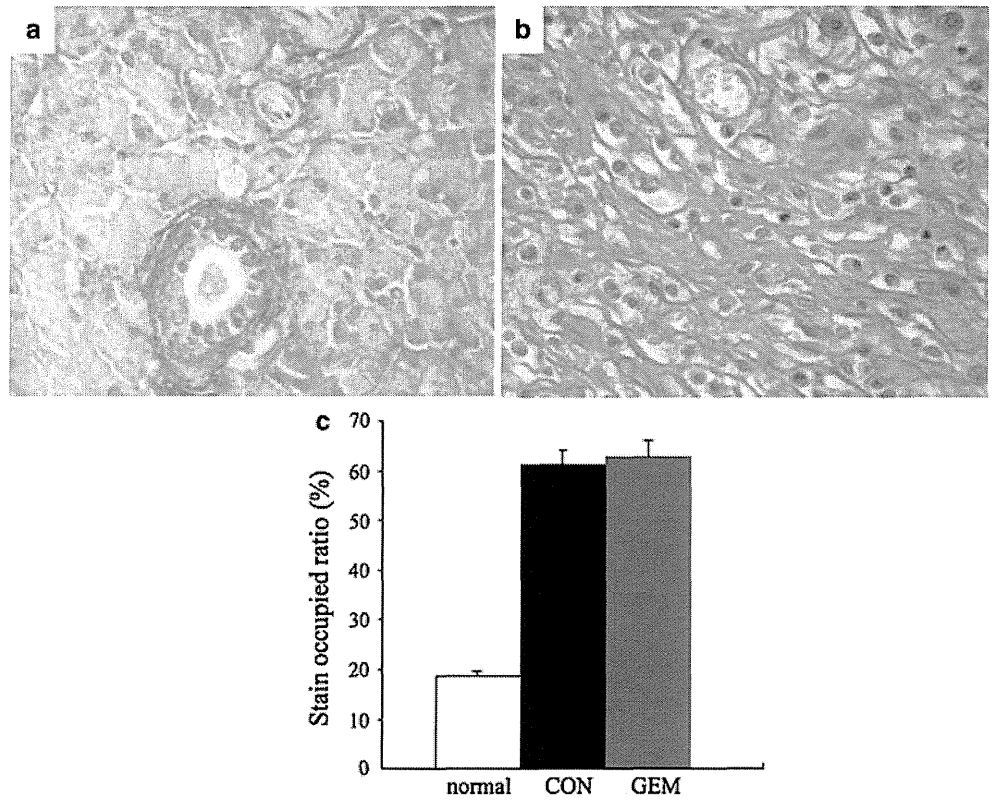

Fig. 7 Picrosirius red staining. **a** Normal pancreatic tissue. **b** Pancreatic cancer tissue (×400). **c** There was no significant difference in Picrosirius red expression in pancreatic cancer tissue between control (*black bar*) and GEM treated animals (*gray bar*)

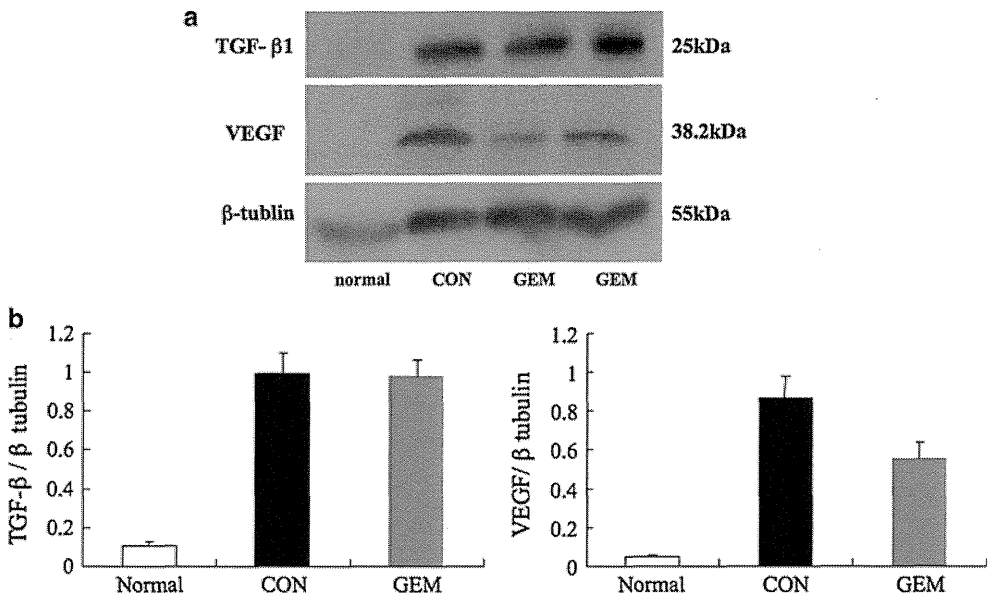

Fig. 8 Western blot analysis of VEGF and TGF-β protein expression in pancreatic cancer tissue. **a** The expression of TGF-β and VEGF in pancreatic cancer tissue. Each lane of GEM represents a different animal. *Normal* normal rat pancreatic tissue; *CON* no treatment group (control); *GEM* GEM treated group. **b** Band intensity was quantified and expressed by ratio to β-tubulin ($n = 3$ for each group). The expression of VEGF was significantly reduced by GEM treatment ($p = 0.01$), but the expression of TGF-β was not inhibited ($p = 0.41$)

increased ECM protein synthesis in response to various stimuli, such as cytokines and growth factors [5, 6], which results in pancreatic fibrosis and a hypoxic microenvironment. Our study showed that the activity of α-SMA-positive MCs in the pancreatic cancer tissue was inhibited by treatment with GEM, but the quantity of ECM did not change. Fibrosis due to chronic pancreatitis was irreversible, as shown in an earlier report [23], and also in our study. There was no change in ECM volume even if the activation of α-SMA-positive MCs, which play a pivotal role in fibrosis in pancreatic cancer tissue, was inhibited.

In summary, when pancreatic cancer growth was suppressed by GEM chemotherapy, activation of α-SMA-positive MCs in the pancreatic cancer tissue was inhibited, and the secretion of VEGF decreased but not secretion of TGF-β1. The decrease of VEGF secretion may be caused

by synergy between the suppression of the cancer cell proliferation and the suppression of activation of α-SMA-positive MCs. Even if the activity of α-SMA-positive MCs decreased, the fibrosis in the pancreatic cancer tissue was irreversible. In conclusion, GEM may not only suppress the tumor cell proliferation, but also favor the suppression of PSCs through VEGF reduction. We suggest that perhaps inhibition therapy of PSC activation in addition to chemotherapy might be a more effective strategy in pancreatic cancer treatment.

Conflict of interest None.

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ORIGINAL RESEARCH

Bevacizumab in the treatment of five patients with breast cancer and brain metastases: Japan Breast Cancer Research Network-07 trial

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Background: Brain metastases from breast cancer occur in 20%–40% of patients, and the frequency has increased over time. New radiosensitizers and cytotoxic or cytostatic agents, and innovative techniques of drug delivery are still under investigation.

Methods: Five patients with brain metastases who did not respond to whole-brain radiotherapy and then received bevacizumab combined with paclitaxel were identified using our database of records between 2011 and 2012. The clinicopathological data and outcomes for these patients were then reviewed.

Results: The median time to disease progression was 86 days. Of five patients, two (40%) achieved a partial response, two had stable disease, and one had progressive disease. In addition, one patient with brain metastases had ptosis and diplopia due to metastases of the right extraocular muscles. However, not only the brain metastases, but also the ptosis and diplopia began to disappear after 1 month of treatment. The most common treatment-related adverse events (all grades) were hypertension (60%), neuropathy (40%), and proteinuria (20%). No grade 3 toxicity was seen. No intracranial hemorrhage was observed.

Conclusion: We present five patients with breast cancer and brain metastases, with benefits from systemic chemotherapy when combined with bevacizumab.

Keywords: brain, bevacizumab, metastatic breast cancer

Introduction

Breast carcinoma is the most frequent neoplasia in the US, Europe, and even Japan.[1] Approximately 40%–45% of all patients with breast cancer will develop metastasis, and the mean survival time from the diagnosis of recurrence for these patients is 18–30 months.[2] Therefore, treatment of patients with metastatic breast cancer aims to prolong survival while relieving symptoms and maintaining a good quality of life.[1–4] Brain metastases from breast cancer occur in 20%–40% of patients, and the frequency has increased over time. As a treatment, the combination of surgery and whole-brain radiotherapy is well known and useful, but is still limited. New radiosensitizers and cytotoxic or cytostatic agents and innovative techniques of drug delivery are being investigated.[5]

Bevacizumab has selective activity against the vascular endothelial growth factor (VEGF)-A ligand and has proven to be efficacious when combined with paclitaxel.[6] It has been well documented that tumor blood vessels show increased vascular permeability and interstitial fluid pressure, decreased pericyte coverage, and increased occurrence of tumor hypoxia, further upregulating VEGF production. Therefore, inhibition of

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VEGF by bevacizumab will not only affect endothelial cells but also the tumor vasculature, suppressing new blood vessel growth and the existing vasculature.

Cautious use of bevacizumab has been recommended in patients at risk of bleeding and uncontrolled hypertension, as well as in patients with a history of arterial thrombotic events. Patients with central nervous system metastases have until recently been routinely excluded from bevacizumab trials, following a single case in 1997 of a 29-year-old patient with hepatocellular carcinoma who experienced a fatal cerebral hemorrhage from a previously undiagnosed brain metastasis in a Phase I study of bevacizumab.[7] However, bevacizumab recently gained accelerated approval from the US Food and Drug Administration for progressive primary brain tumors, with a low rate (approximately 3%) of intratumoral hemorrhage.[8,9] More recent studies showed that bevacizumab is safe in patients with brain metastases.[10–13] We present here the efficacy and side effects of bevacizumab for patients with breast cancer and brain metastases.

Materials and methods

From the Japan Breast Cancer Research Network database, we retrospectively identified five patients treated with bevacizumab-containing chemotherapy regimens for active central nervous system metastases. All patients had recurrent tumors after receiving radiation therapy. All patients received bevacizumab at a dose of 10 mg/kg by intravenous infusion every 2 weeks with concomitant paclitaxel. Paclitaxel 80 mg/m^2 was administered intravenously on days 1, 8, and 15 every 4 weeks. Dose reductions of paclitaxel from 80 to 60 mg/m^2 were performed as described previously.[14] Tumor response was determined by comparing measurements from consecutive magnetic resonance imaging (MRI) scans, as described elsewhere.[8] In brief, progressive disease was deemed to be present if a new lesion had occurred, if the MRI showed a >25% increase in fluid attenuated inversion recovery (FLAIR) or contrast-enhanced volume, or if the MRI scan showed an increase in tumor volume; partial response was defined as a >25% decrease in the enhanced lesion and FLAIR; and a complete response was defined as no detectable contrast enhancement and stable or improved FLAIR signal. Physical examination findings, tumor characteristics, number of treatment cycles, chemotherapy-related toxicities, and symptom severity were recorded every week. Toxicity was graded using the National Cancer Institute Common Terminology Criteria for Adverse Events version 3.0.

Results

Patient characteristics

The characteristics of the study population are presented in Table 1. The median age was 60 (range 39–71) years. Eastern Cooperative Oncology Group performance status was <3. All patients were pretreated with whole-brain radiotherapy. The median number of metastatic sites was three (range 1–5).

Efficacy

The patients were evaluable for response and toxicity. Of the five patients, two (40%) achieved a partial response, two had stable disease, and one had progressive disease. Representative data are shown in Figures 1–4. In addition, one patient with brain metastases had ptosis and diplopia due to metastases of the right extraocular muscles (Figures 4 and 5). However, not only the brain metastases but also the ptosis and diplopia began to disappear after 1 month of treatment (Figure 5). Median time to disease progression was 86 (range 30–135) days. Two patients (40%) were still alive at the last follow-up.

Safety

The most common treatment-related adverse events were grade 1/2 in intensity. Common toxicities were

Table 1 Patient characteristics

Patients (n = 5)	
Age, years, median (range)	60 (39–71)
Performance status	
0	0
1	2
2	2
Hormone status	
+	3
–	2
HER2 status	
+	1
–	4
Menopausal status	
Pre	2
Post	3
Type of prior chemotherapy	
Trastuzumab	1
Taxane	4
Anthracycline/taxane	1
Whole-brain radiotherapy	
+	5
–	0
Median number of brain metastatic sites (range)	3 (1–5)
Metastatic sites involved	
Bone/soft tissue	2
Visceral	2

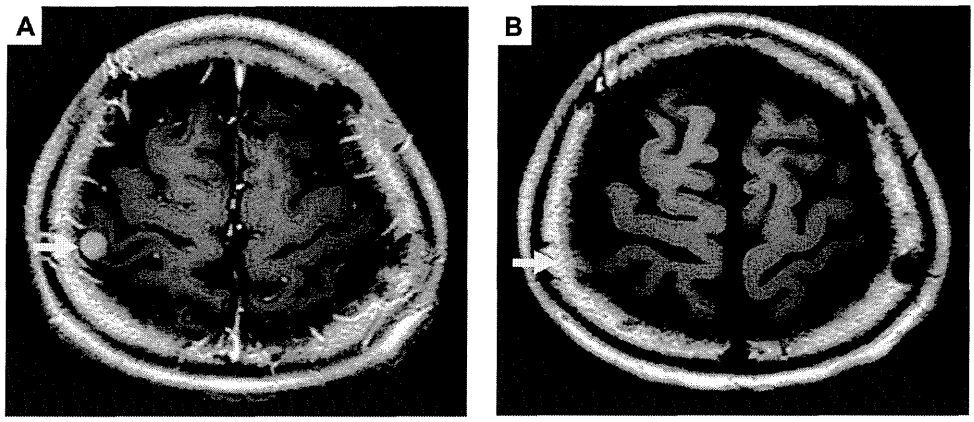

Figure 1 Pre (**A**) and post (**B**) treatment brain magnetic resonance imaging of metastatic tumor showing a partial response (arrows).

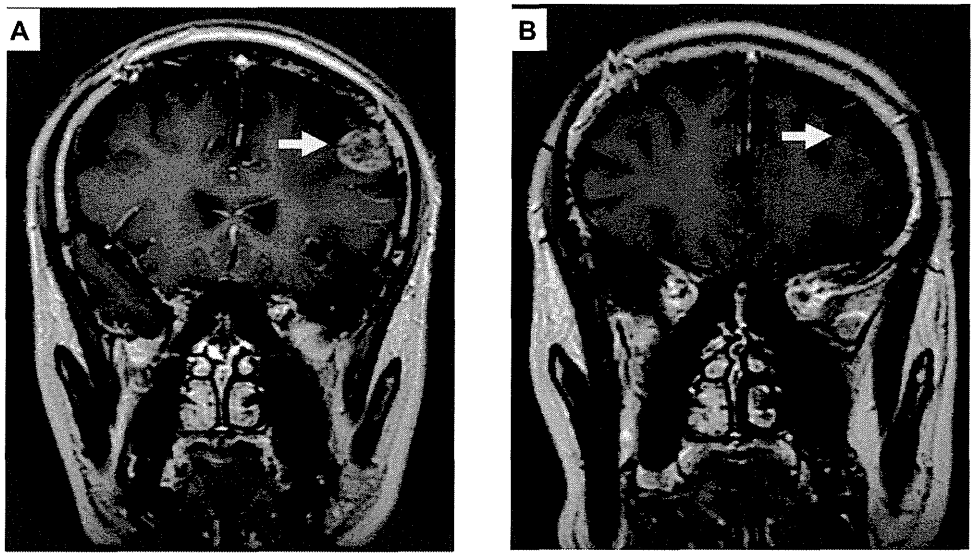

Figure 2 Pre (**A**) and post (**B**) treatment brain magnetic resonance imaging of metastatic tumor showing a complete response (arrows).

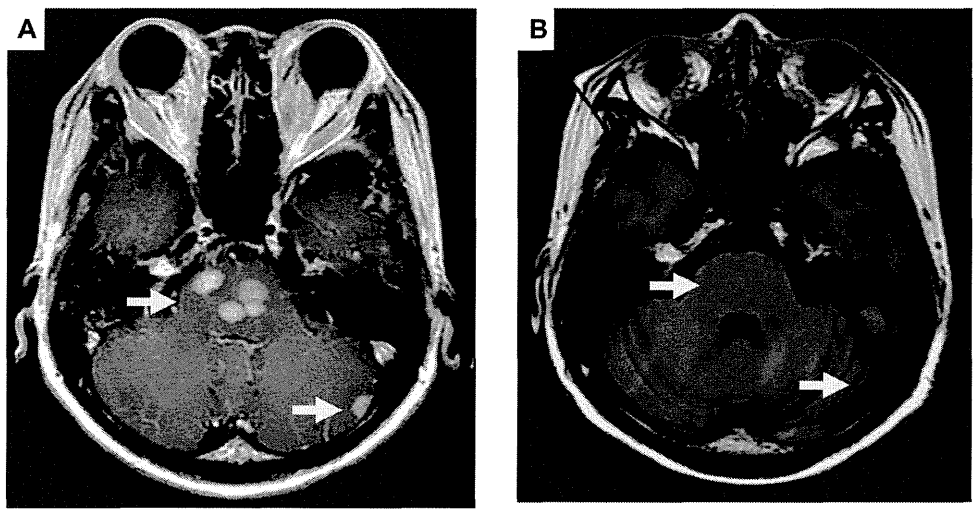

Figure 3 Pre (**A**) and post (**B**) treatment brain magnetic resonance imaging of metastatic tumor showing a partial response (arrows).

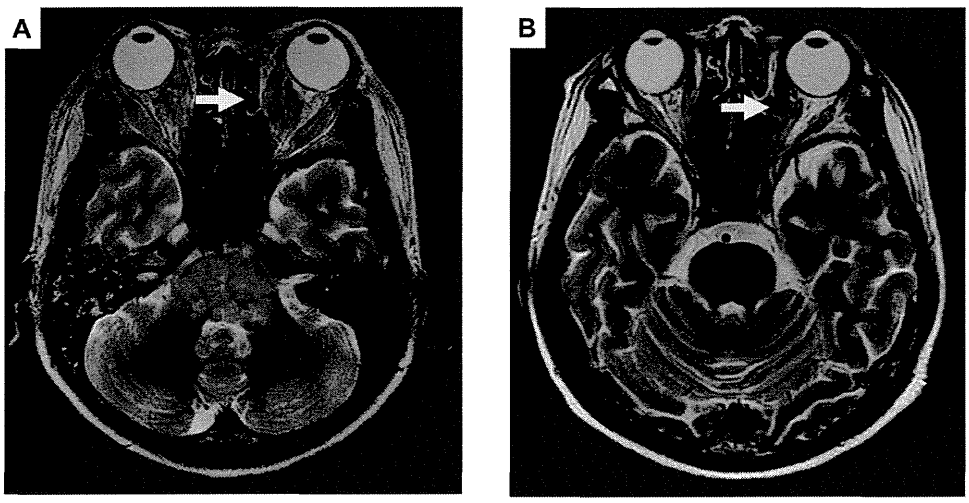

Figure 4 Pre (**A**) and post (**B**) treatment brain magnetic resonance imaging of metastases of right extraocular muscles showing a partial response (arrows).

grade 1 hypertension (60%), grade 1 neuropathy (40%), and proteinuria (20%). No grade 3 toxicity was seen. No intracranial hemorrhage was observed.

Discussion

The results of this multicenter retrospective study suggest that bevacizumab combined with a taxane is highly active and well tolerated by women with breast cancer and brain metastases who have failed whole-brain radiotherapy. Bevacizumab combined with a taxane yielded a 40% response rate. The median time to disease progression was 86 days. The results presented here are similar to those reported elsewhere.[12] The exact mechanism of bevacizumab in brain parenchymal disease is unknown; whether it is a result of direct effects on the tumor vasculature and/or the blood–brain barrier itself is unclear. Most drugs fail to enter the central nervous system because of the blood–brain barrier. This restriction particularly affects drugs that are not substrates for active transport into the central nervous system, hydrophilic molecules larger than 500 Da, and high molecular weight therapeutic modalities, such as monoclonal antibodies, antisense oligonucleotides, viral vectors, stem cells, and nanoparticles.[15] However, some studies have shown that VEGF may provide new opportunities for manipulating the permeability of the blood–brain barrier in vivo.[16,17] Further, previous studies in glioma models have demonstrated a fine balance between VEGF and angiopoietin-2, a proapoptotic factor in angiogenesis.[18] It has been noted that the blood–brain barrier is abnormal with tumors > 0.5 mm, and might affect the integrity of astrocytes and the endothelial cells of the blood–brain barrier.[19] Larger tumors result in an increased risk of ischemia, further disrupting the blood–brain barrier.[20] Additional studies are in progress to evaluate the role of bevacizumab in combination with chemotherapy in previously treated brain metastases originating from non-small-cell lung cancer and also in reducing central nervous system side effects after radiotherapy in patients with primary brain, melanoma, and head and neck cancer.[21,22] In the present study, bevacizumab and paclitaxel suppressed brain metastasis. Therefore, theoretically, there is a possibility that bevacizumab might cross the blood–brain barrier and penetrate brain tumors in sufficient concentrations to synergize with anticancer drugs.

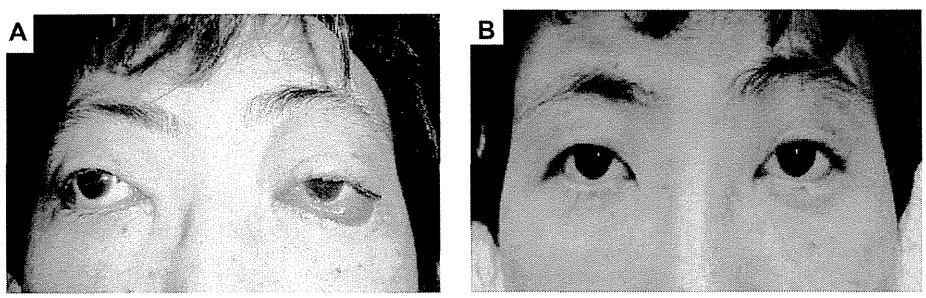

Figure 5 Facial features pre (**A**) and post (**B**) treatment.
Note: Ptosis and diplopia begin to disappear after 1 month of the treatment.

In the current study, the majority of adverse events were mild to moderate in intensity, and confirm the results of previous studies in similar patient populations.[11–13] Acute toxicities were quite mild and manageable. Hypertension and proteinuria were common, and neuropathy was managed with modification of the paclitaxel dose. Further, intracranial hemorrhage was not observed. The limitations of the present study include its retrospective nature and the small number of patients included. Nonetheless, the finding that bevacizumab has significant activity against breast cancer with brain metastasis is important.

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Disclosure

The authors report no conflicts of interest in this work.

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ORIGINAL ARTICLE

A Prospective Randomized Controlled Trial of Preoperative Whole-Liver Chemolipiodolization for Hepatocellular Carcinoma

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Abstract
Background We previously reported that preoperative chemolipiodolization of the whole liver is effective for reducing the incidence of postoperative recurrence and prolonging survival in patients with resectable hepatocellular carcinoma (HCC). The present randomized controlled trial was performed to evaluate the influence of preoperative transcatheter arterial chemoembolization (TACE) on survival after the resection of HCC.
Methods Operative results and long-term outcome were prospectively compared among 42 patients who received only selective TACE targeting the tumor (selective group), 39 patients who received TACE targeting the tumor plus chemolipiodolization of the whole liver (whole-liver group), and 43 patients without preoperative TACE or chemolipiodolization (control group).
Results There were no serious side effects of TACE or chemolipiodolization and the operative outcomes did not differ among the three groups. Even though preoperative TACE induced complete tumor necrosis, there were no significant differences in the pattern of intrahepatic recurrence or the time until recurrence among the three groups. There were also no significant differences in disease-free survival or overall survival among the three groups, even among patients with larger tumor size.
Conclusion These results indicate that preoperative selective TACE and whole-liver chemolipiodolization plus TACE do not reduce the incidence of postoperative recurrence or prolong survival in patients with resectable HCC.

Keywords Hepatocellular carcinoma · Preoperative chemolipiodolization · Whole liver · Hepatectomy · Randomized controlled trial

Introduction

Hepatocellular carcinoma (HCC) is the fifth most common cancer worldwide [1]. Although the majority of patients are still found in Asia and Africa, recent studies have shown that the incidence and mortality rate of HCC are rising in North America and Europe [2, 3]. There has been an increase in reports of non-surgical therapeutic options for small HCC, such as percutaneous ethanol injection therapy [4], microwave coagulation therapy [5], and percutaneous radiofrequency ablation (RFA) [6], but there is ongoing controversy regarding the best method of treating small tumors. In Japan, liver transplantation is not a practical option for most HCC patients, because the national health insurance scheme only covers transplantation for patients with decompensated cirrhosis whose tumors fit the Milan criteria. Resection is, therefore, generally the first-line treatment for patients with small tumors and underlying chronic liver disease, but the long-term survival rate after

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potentially curative resection of HCC is still unsatisfactory because of the high rate of recurrence [7]. To improve prognosis, it is important to prevent the recurrence of HCC after its initial resection, but standard therapy for intrahepatic metastasis has not yet been developed.

With various improvements in interventional radiology, transcatheter arterial chemoembolization (TACE) has become an increasingly important palliative treatment for HCC. Initially, TACE was only performed for unresectable HCC, as well as for some early tumors that were extremely difficult to resect. More recently, TACE has been used as preoperative adjuvant therapy in patients who have resectable HCC with the hope that it may improve survival [8–13]. Based on the current evidence, however, preoperative TACE is not routinely recommended for patients undergoing hepatectomy to treat resectable HCC [14–16], and TACE may be contraindicated in patients with cirrhosis because it can lead to the progressive deterioration of liver function [14]. Whether preoperative TACE can improve the long-term survival of HCC patients is still unclear, and there have been only three randomized controlled trials evaluating the influence of preoperative TACE on survival [15, 17, 18]. We previously reported that preoperative chemolipiodolization of the entire liver is effective for reducing the incidence of postoperative recurrence and for prolonging survival in patients with resectable HCC [19]. Accordingly, the present randomized controlled trial was conducted to better assess the influence of preoperative TACE combined with whole-liver chemolipiodolization on survival after the resection of HCC.

Patients and Methods

Patients

Between January 2004 and June 2007, 124 patients with HCC underwent curative hepatic resection at our institution. A curative operation was defined as the resection of all detectable tumors. The eligibility criteria for inclusion in this study were as follows: (1) age 20–80 years; (2) a preoperative diagnosis of HCC with no previous treatment; (3) no other malignancies; (4) Child–Pugh score A or B; (5) leukocyte count $\geq$3,000/mm^3; (6) hemoglobin level $\geq$9.5 g/dl; (7) platelet count $\geq$50,000/mm^3; (8) serum creatinine level <1.2 mg/dl; (9) total bilirubin <2.0 mg/dl; (10) local nodular disease without extrahepatic metastasis; and (11) Eastern Cooperative Oncology Group (ECOG) performance status 0–1 [20]. The etiology of HCC (HCV-related or other [HBV-related or non-B, non-C-related]) and the size of the tumor on imaging were taken into consideration when dividing patients into the three groups. The sample size was estimated based on our previously reported 3-year disease-free survival rates in selective and whole-liver groups, being 25 and 60%, respectively [19]. We needed 37 patients in each group for a type I error rate of 5% and a type II error rate of 20% with a two-tailed test. Among the 124 patients, TACE was performed preoperatively in 81. Patients were randomized to receive chemolipiodolization with gelatin sponge (equal to TACE) targeting the tumor (selective group, $n = 42$), chemolipiodolization with gelatin sponge (equal to TACE) targeting the tumor plus chemolipiodolization without gelatin sponge for the non-cancerous liver (whole-liver group, $n = 39$), or no preoperative TACE (control group, $n = 43$). The study protocol was explained to all patients, and they understood that they would be randomly selected for one of the above three groups. All patients gave written informed consent to participation in the trial. They were randomized by the envelope method and were informed of the result of the randomization before angiography. All operations were performed by the same surgeon, who had experience of over 700 hepatic resections. The protocol for this study was approved by the ethics committee of Kansai Medical University. The primary outcome measures were disease-free survival rate and overall survival rate. Secondary outcome measures included procedure-related complications and hospital mortality (Fig. 1).

Chemolipiodolization

A catheter was selectively inserted into the right or left hepatic artery, a segmental artery, or a subsegmental artery by Seldinger's method. In the selective group, TACE was performed via the right hepatic artery in 16 patients, the left hepatic artery in 10 patients, a segmental artery in 9 patients, and a subsegmental artery in 7 patients. In the whole-liver group, TACE (i.e., chemolipiodolization with gelatin sponge) was performed via the right hepatic artery in 18 patients and the left hepatic artery in 13 patients to target the tumor, while chemolipiodolization alone was performed on the non-cancerous side via the left or right hepatic artery. In a further 8 patients, TACE was performed via a right or left subsegmental artery to target the tumor and chemolipiodolization of the non-cancerous liver was performed via the right and left hepatic arteries as the catheter was withdrawn. The selective group was treated with epirubicin (Farmorubicin) at a mean ($\pm$ standard deviation [SD]) dose of 47.0 $\pm$ 17.8 mg, iodized oil (Lipiodol) at a mean volume of 3.8 $\pm$ 2.1 ml, and gelatin sponge particles. In the whole-liver group, epirubicin (28.1 $\pm$ 5.5 mg), Lipiodol (2.9 $\pm$ 1.4 ml), and gelatin sponge particles were used to treat the tumor, while only epirubicin (22.2 $\pm$ 6.2 mg) and Lipiodol (1.9 $\pm$ 0.8 ml) were infused into the non-cancerous liver. In the control group, only angiography was performed.