High prevalence of early repolarization in short QT syndrome

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BACKGROUND Short QT syndrome (SQTS) is characterized by an abnormally short QT interval and sudden death. Due to the limited number of cases, the characteristics of SQTS are not well understood. It has been reported recently that early repolarization is associated with idiopathic ventricular fibrillation and the QT interval is short in patients with early repolarization.

OBJECTIVE The purpose of this study was to study the association between early repolarization and arrhythmic events in SQTS.

METHODS The study consisted of three cohorts: SQTS cohort (N = 37), control cohort with short QT interval and no arrhythmic events (N = 44), and control cohort with normal QT interval (N = 185). ECG parameters were compared among the study cohorts.

RESULTS Heart rate, PR interval, and QRS duration were similar among the three study cohorts. Early repolarization was more common in the SQTS cohort (65%) than in the short QT control cohort (30%) and the normal QT control cohort (10%). Duration from T-wave peak to T-wave end was longer in the SQTS cohort than in the short QT control cohort, although QT and corrected QT intervals were similar. In the SQTS cohort, there were more males among patients with arrhythmic events than in those with a family history but without arrhythmic events. In multivariate models, early repolarization was associated with arrhythmic events in the SQTS cohort. ECG parameters including QT and QTc intervals were not associated with arrhythmic events in the SQTS cohort.

CONCLUSION There is a high prevalence of early repolarization in patients with SQTS. Early repolarization may be useful in identifying risk of cardiac events in SQTS.

KEYWORDS Arrhythmia; Electrocardiogram; QT interval; Repolarization; Sudden death

ABBREVIATIONS **QTc** = corrected QT interval; **SQTS** = short QT syndrome

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Introduction

The short QT syndrome (SQTS) is characterized by an abnormally short QT interval and increased risk of ventricular fibrillation and sudden death.[1,2] Similar to other arrhythmia syndromes, such as long QT syndrome and Brugada syndrome,[3] SQTS is a genetically heterogeneous disease, and, to date, five responsible genes encoding different ion channels have been identified.[3–7] Some inherited arrhythmia syndromes may share genetic backgrounds that result in overlapping arrhythmia phenotypes.[3]

Although early repolarization is generally considered benign,[8] it has been reported recently that early repolarization is associated with increased risk for sudden cardiac death in patients with idiopathic ventricular fibrillation.[9–12] Haissaguerre et al[9] reported that, among patients with idiopathic ventricular fibrillation, the QT interval was shorter in patients with early repolarization than in those without, suggesting an association between early repolarization and QT interval shortening. Evidence that mutations in calcium channel genes are associated with Brugada-type ST-segment elevation and abnormally short QT intervals further suggests a relationship between early phase repolarization abnormalities and short QT interval.[4] Here we report on our

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study of the prevalence of early repolarization and its association with arrhythmic events in SQTS.

Methods

This cooperative study consisted of three cohorts. (1) *SQTS cohort* included patients with SQTS referred to our institutions and patients with SQTS from previous reports. The diagnosis of SQTS was made if a patient with a short QT interval [corrected QT interval (QTc) by Bazett formula ≤330 ms] had an arrhythmic event including documented ventricular fibrillation, resuscitated sudden cardiac death, and syncope and/or had a family history of SQTS, or if a patient with a short QT interval (QTc ≤360 ms) had mutations in ion channel genes responsible for SQTS.[3,13] We searched in the electronic databases PubMed, EMBASE, and Cochrane for all published studies that examined patients with SQTS. The search was limited to the end of June 2009. Published studies were considered eligible if they included clinical characteristics of the patients and ECGs. All ECGs from patients reported in the literature were reanalyzed. Electrophysiologic study was performed in patients with SQTS based on the indication of each institution. (2) *Control cohort with short QT interval* (QTc ≤330 ms) *and no arrhythmic events* was selected from among 86,068 consecutive ECGs stored on the ECG database at Niigata University Medical and Dental Hospital from May 7, 2003 to July 2, 2009. Subjects who did not have arrhythmic events or cardiovascular disease and were not taking any medication were included in this cohort. (3) *Control cohort with normal QT interval* was also selected from the ECG database. This cohort consisted of subjects who were matched to the SQTS cohort for gender and age. Subjects who had normal QT interval (360–440 ms) and did not have cardiovascular disease or were not taking any medication were included in this cohort. Subjects with Brugada-type ST-segment elevation were excluded from all study cohorts.[3,9]

QT intervals were measured on lead V_2 with the tangent methods for determination of QT_{end} using a semi-automated digitizing program with electronic calipers by an experienced observer blinded to the clinical details of all subjects included in this study.[14,15] Early repolarization was defined as elevation of the J point noted as either as QRS slurring or notching ≥0.1 mV in more than two leads.[9]

Differences in parameters were analyzed using multivariable logistic regression models when SQTS cohort and control cohort with short QT interval were compared and analyzed using conditional logistic regression models when SQTS cohort and control cohort with normal QT interval were compared. All statistical analyses were performed with SPSS (version 12.0, SPSS, Inc., Chicago, IL, USA). Two-sided $P < .05$ was considered significant. Values are expressed as mean ± SD. The study protocol was approved by the Ethics Committee of Niigata University School of Medicine. To determine interobserver variability, a second observer made independent blinded QT interval determinations of all study subjects with short QT interval.

Results

Thirty-seven patients with SQTS were identified: 12 from our institutions and 25 reported in the literature,[2,5,6,14,16–25] Forty-four control subjects with short QT interval and 185 control subjects with normal QT interval also were identified (Table 1). The SQTS cohort consisted of 25 (68%) patients with symptoms, including 14 with cardiac arrest (3 sudden death, 11 resuscitated) and 11 with syncope. Genetic screening identified mutations in ion channels in 7 (41%) of 17 probands who were genetically screened (2 *KCNQ1*, 4 *KCNH2*, 1 *KCNJ2*). Among patients in our institutions and those reported in the literature, there was no difference with regard to gender, age, prevalence of family history, QT or QTc interval, or inducibility of ventricular tachyarrhythmia by electrical programmed stimulation.

Heart rate, PR interval, and QRS duration in the SQTS cohort were not different among patients in either the short QT control cohort or the normal QT control cohort (Table 1). QT and corrected QT intervals were shorter in the SQTS and short QT control cohorts than in the normal QT control cohort. Early repolarization occurred in 24 (65%) patients with SQTS (Figure 1). Interobserver variability between two investigators was 8.6 ms (95% confidence interval −0.5 to 17.7 ms) for QT interval and 9.0

Table 1 ECG parameters of study cohorts

	Patients with SQTS (N = 37)	Subjects with short QTc (N = 44)	Versus subjects with short QTc*		Subjects with normal QTc† (N = 185)	Versus subjects with normal QTc	
			OR (95% CI)	*P* value		OR (95% CI)	*P* value
Male gender [N (%)]	27 (73)	34 (77)	2.84 (0.72–11.2)	.14	135 (73)	—	—
Age (years)	30 ± 19	47 ± 23	1.05 (1.02–1.08)	.001	30 ± 19	—	—
Heart rate (bpm)	69 ± 393	65 ± 398	1.00 (1.00–1.01)	.3	70 ± 327	1.00 (1.00–1.00)	0.70
PR interval (ms)	138 ± 19	153 ± 38	1.01 (0.99–1.03)	.54	143 ± 24	0.99 (0.97–1.01)	0.18
QRS interval (ms)	86 ± 7	84 ± 8	0.97 (0.91–1.04)	.38	85 ± 7	1.01 (0.96–1.06)	0.74
QT interval (ms)	286 ± 36	286 ± 15	0.99 (0.97–1.01)	.28	367 ± 36	0.97 (0.96–0.98)	<0.001
QTc (ms)	308 ± 29	299 ± 21	0.98 (0.96–1.00)	.06	399 ± 24	0.97 (0.97–0.98)	<0.001

CI = confidence interval; OR = odds ratio; QTc = corrected QT interval; SQTS = short QT syndrome.

*Models were adjusted for gender and age.

†Gender and age were matched between patients with SQTS and subjects with normal QT interval.

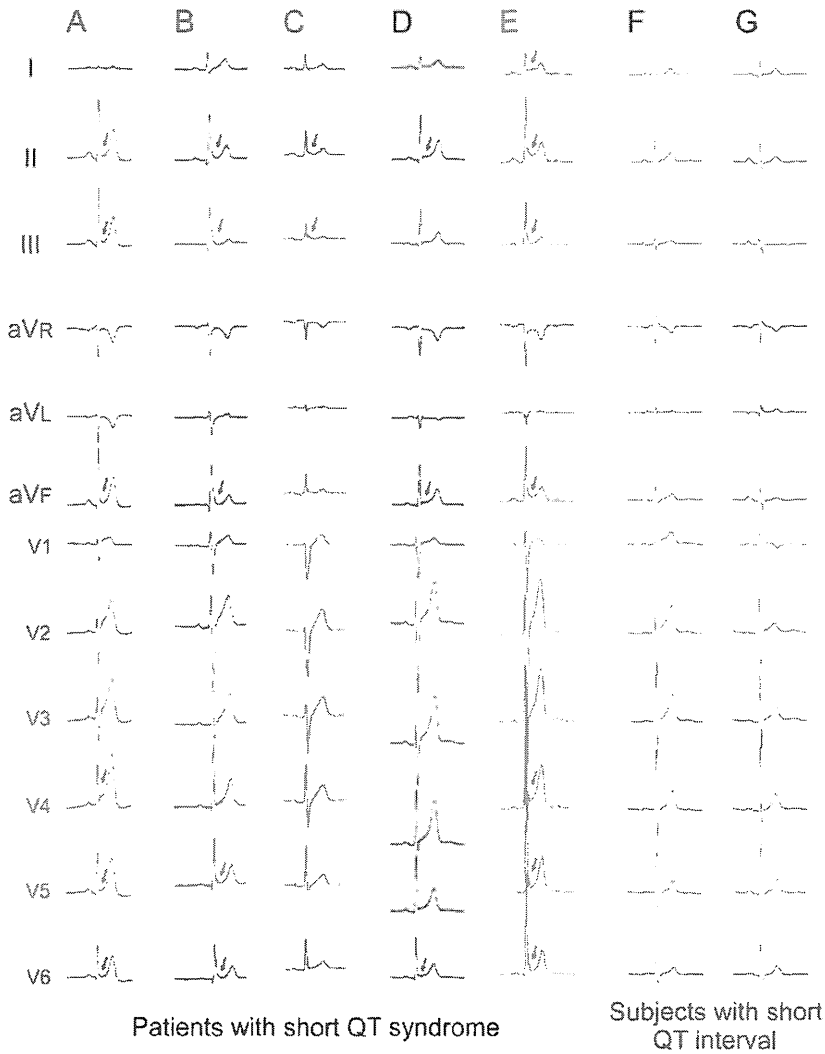

Figure 1 Early repolarization in short QT syndrome. ECGs were recorded from patients with short QT syndrome (**A**: 61-year-old woman; **B**: 30-year-old man; **C**: 38-year-old man; **D**: 31-year-old man; **E**: 22-year-old man) and control subjects with a short QT interval (**F**: 23-year-old man; **G**: 44-year-old woman). In each patient with short QT syndrome, early repolarization was evident in the inferolateral leads (*arrows*).

ms (95% confidence interval −0.6 to 18.7 ms) for QTc interval. The frequency of early repolarization was not different between patients in our institutions and those reported in the literature. Early repolarization was present in the inferior leads (II, III, aVF) in 9 patients, in the lateral leads (I, aVL, V_4–V_6) in 6 patients, and in both the inferior and lateral leads in 9 patients. Of 10 probands with early repolarization genetically screened, mutations were identified in 3 patients (1 *KCNQ1*, 2 *KCNH2*). Early repolarization was more common in the SQTS cohort than in the short QT control and normal QT control cohorts (Figure 2).

The association of early repolarization with arrhythmic events then was studied in patients with SQTS. In the SQTS cohort, there were more males among patients with arrhythmic events than among those with a family history but without arrhythmic events (Table 2). In multivariate models adjusted for gender and age, early repolarization was associated with arrhythmic events, although ECG parameters including QT and QTc intervals were not associated with arrhythmic events. Early repolarization remained associated with arrhythmic events after adjustment for age, gender, and QTc interval (P = .001). Electrophysiologic study performed in 18 patients with SQTS revealed no difference in inducibility of ventricular tachyarrhythmia between patients with arrhythmic events (73%) and those without arrhythmic events (71%).

QT interval parameters were compared between SQTS and short QT control cohorts because some of the parameters recently have been associated with SQTS.[26] Interval from T-wave peak to T-wave end (T_{peak} to T_{end}) was longer in the SQTS cohort than in the short QT control cohort even after heart rate correction using the Bazett formula, whereas QT interval, QTc interval, and interval from Q-wave to T-wave peak (QT_{peak}) were not different between the two cohorts (Table 3). Ratio of T_{peak} to T_{end} per QT was larger in the SQTS cohort than in the short QT control cohort.

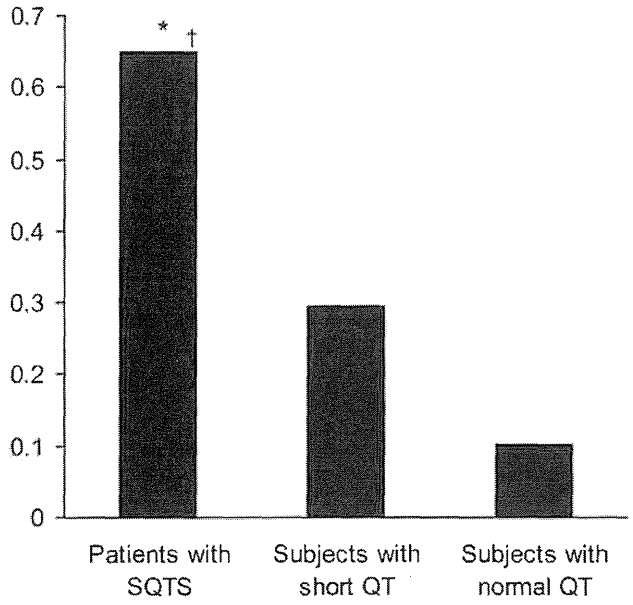

Figure 2 Frequency of early repolarization. Odds ratios (95% confidence intervals) for early repolarization in patients with short QT syndrome (SQTS) were 5.64 (1.97–16.15) and 16.58 (7.2–38.21) versus subjects with short QT interval and those with normal QT interval, respectively. *P = .001 vs subjects with short QT interval. †P <.001 vs subjects with normal QT interval.

Discussion

SQTS is a recently discovered, very rare disease with an increased risk of sudden death.[2] Due to the limited number of cases, the characteristics of SQTS are not well understood. Therefore, we conducted a cooperative analysis of ECGs from patients with SQTS in our institutions and those reported in the literature and found that early repolarization is common in SQTS.

Early repolarization is a common ECG finding. It is present in 1% to 13% of the general population and usually is considered as a normal variant due to its benign long-term prognosis.[8,11,27–29] However, increasing evidence suggests that early repolarization is associated with arrhythmia.[9,27,30–34] Since 1985, we and other investigators have reported an association between early repolarization (or late depolarization) and sudden cardiac death.[30–32] A multicenter study including our institution recently showed that early repolarization is present in one third of patients with idiopathic ventricular fibrillation.[9] Early repolarization is associated with increased risk of sudden cardiac arrest in idiopathic ventricular fibrillation, and the amplitude of early repolarization increases before development of arrhythmic events.[9,10] In Brugada syndrome, which is characterized by J-wave and ST-segment elevation in the right precordial leads on ECG and sudden cardiac death,[3] early repolarization in the inferolateral leads is not uncommon and is associated with arrhythmic events,[34] although another report has shown negative results.[33] In our study, early repolarization in the inferolateral leads was frequently found in SQTS and, more importantly, was associated with arrhythmic events in SQTS. In addition to arrhythmia syndromes unassociated with structural heart disease, a high frequency of early repolarization in arrhythmogenic right ventricular dysplasia/cardiomyopathy has been reported.[27]

It has been suggested that SQTS and idiopathic ventricular fibrillation share clinical characteristics.[23] Short QT interval is frequently found in idiopathic ventricular fibrillation,[23] and QT interval is relatively short in patients with idiopathic ventricular fibrillation who have early repolarization.[9] Spontaneous and inducible ventricular fibrillation can be initiated by short-coupled premature ventricular beat in SQTS and idiopathic ventricular fibrillation.[21,35,36] The efficacy of isoproterenol and quinidine has been reported for both arrhythmia syndromes,[21,37] although the arrhythmogenic effects of isoproterenol in an experimental model of SQTS have been reported.[38] Our study showing an association of early repolarization with SQTS further supports the presence of common arrhythmogenic substrates in SQTS and idiopathic ventricular fibrillation.

A precise mechanism for ventricular fibrillation in SQTS is not known, but characteristic ECG abnormalities may reflect arrhythmogenicity. A prior study showed that the interval from T-wave peak to T-wave end is relatively long in SQTS, and our study replicated the results.[26] T-wave peak to T-wave end interval is considered to reflect transmural dispersion of repolarization, and relative prolongation of the interval in SQTS may indicate a high vulnerability to ventricular fibrillation.[39] An experimental model of SQTS

Table 2 Characteristics of SQTS patients with and those without arrhythmic events

	Patients with arrhythmic events (N = 25)	Patients without arrhythmic events (N = 12)	OR (95% CI)	P value
Male gender [N (%)]	21 (84)	6 (50)	10.44 (0.85–127.48)	.07
Age (years)	30 ± 19	23 ± 18	1.05 (0.99–1.12)	.13
Heart rate (bpm)	69 ± 393	76 ± 473	1.00 (1.00–1.01)	.38
PR interval (ms)	138 ± 19	134 ± 18	0.99 (0.95–1.04)	.84
QRS interval (ms)	86 ± 7	85 ± 10	0.93 (0.82–1.07)	.31
QT interval (ms)	286 ± 36	271 ± 40	1.00 (0.97–1.03)	.75
QTc (ms)	308 ± 29	306 ± 33	0.98 (0.94–1.02)	.33
Early repolarization [N (%)]	22 (88)	2 (17)	46.53 (4.52–478.79)	.001

CI = confidence interval; OR = odds ratio; QTc = corrected QT interval; SQTS = short QT syndrome.
Models were adjusted for gender and age.

Table 3 ECG parameters for study cohorts with short QT interval

	Patients with SQTS	Subjects with short QTc	OR (95% CI)	*P* value
QT_{peak} (ms)	211 ± 37	222 ± 19	0.99 (0.98–1.01)	.37
Corrected QT_{peak}	226 ± 32	234 ± 24	0.99 (0.98–1.01)	.56
T_{peak} to T_{end} (ms)	81 ± 21	67 ± 13	1.08 (1.03–1.13)	<.001
Corrected T_{peak} to T_{end}	89 ± 28	72 ± 17	1.05 (1.02–1.09)	.002
QT_{peak}/QT ratio (%)	27 ± 6	22 ± 4	0.83 (0.73–0.94)	.004

Models were adjusted for gender and age.
CI = confidence interval; OR = odds ratio; QTc = corrected QT interval; SQTS = short QT syndrome.

provides evidence that increased transmural dispersion of repolarization under short QT interval conditions results in ventricular tachyarrhythmia.[38] A tall peaked T wave is one of the characteristic ECG abnormalities in SQTS,[1] but the amplitude of the T wave is not different between patients with SQTS and subjects with short QT interval and no arrhythmic events, suggesting that a tall T wave is associated with a short QT interval but is not associated with arrhythmogenicity.[26] In SQTS, characteristic ECG abnormities are also found in the early repolarization phase. In patients with SQTS, the ECG shows a very short J-point to T-wave peak interval and no flat ST segment.[26] In our study, early repolarization was frequently found in SQTS and was associated with arrhythmic events. Whether the inferolateral J-point elevation reflects late depolarization or early repolarization is controversial, but this pattern has been considered repolarization because of slower inscription, spontaneous changes occurring concurrently with ST segment but not with QRS complexes, and absence of late potentials on signal-averaged ECG.[9,40] Taken together, the finding suggest that abnormalities in the early phase of repolarization create the arrhythmogenic substrate in SQTS.

Sex hormone and gender difference have an important role in the arrhythmia syndromes.[41–43] It is well known that the QT interval is affected by sex hormones, and the QT interval is longer in women than men.[44] Female gender is a risk factor for development of ventricular tachyarrhythmias in both congenital and acquired long QT syndrome.[41,42] On the other hand, Brugada syndrome is more prevalent in men than in women, and the male hormone testosterone is reported to contribute to male predominance in Brugada syndrome.[43] In this study, male gender was associated with arrhythmic events in SQTS and short QT interval was frequently found in men, suggesting a role of sex hormones in SQTS opposite to that in long QT syndrome. Recent evidence that the QT interval can be shortened by anabolic androgenic steroids and testosterone further supports this hypothesis.[45,46]

SQTS is a genetically heterogeneous disease with five responsible genes encoding ion channels: *KCNQ1*, *KCNH2*, *KCNJ2*, *CACNA2D1*, and *CACNB2b*.[3,4] An increase in outward current by gain-of-function mutations in potassium channels or a decrease in inward current by loss of function mutations in calcium channels may be responsible for SQTS.[3,4] Early repolarization was found in patients with mutations in *KCNQ1* and *KCNH2* and in those without mutations in the known genes, suggesting a heterogeneous genetic background for the association between short QT interval and early repolarization. To date, mutations in calcium channel genes (*CACNA2D1* and *CACNB2b*) have been identified in three probands with Brugada syndrome associated with a short QT interval, but early repolarization is not present in the inferolateral leads in any of them.[4] A recent study has identified a mutation in *KCNJ8*, an initial responsible gene for idiopathic ventricular fibrillation associated with early repolarization.[47] Although there are some similarities in phenotype between SQTS and idiopathic ventricular fibrillation with early repolarization, a common genetic background has not been identified.

Conclusion

Our study showed a high prevalence of early repolarization in patients with SQTS and an association of early repolarization with arrhythmic events. Early repolarization may be a useful marker for risk stratification of cardiac arrest in SQTS, although further investigation with longitudinal follow-up is required to evaluate our results.

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KCNE2 modulation of Kv4.3 current and its potential role in fatal rhythm disorders

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BACKGROUND The transient outward current I_{to} is of critical importance in regulating myocardial electrical properties during the very early phase of the action potential. The auxiliary β subunit *KCNE2* recently was shown to modulate I_{to}.

OBJECTIVE The purpose of this study was to examine the contributions of *KCNE2* and its two published variants (M54T, I57T) to I_{to}.

METHODS The functional interaction between Kv4.3 (α subunit of human I_{to}) and wild-type (WT), M54T, and I57T *KCNE2*, expressed in a heterologous cell line, was studied using patch-clamp techniques.

RESULTS Compared to expression of Kv4.3 alone, co-expression of WT *KCNE2* significantly reduced peak current density, slowed the rate of inactivation, and caused a positive shift of voltage dependence of steady-state inactivation curve. These modifications rendered Kv4.3 channels more similar to native cardiac I_{to}. Both M54T and I57T variants significantly increased I_{to} current density and slowed the inactivation rate compared with WT *KCNE2*. Moreover, both variants accelerated the recovery from inactivation.

CONCLUSION The study results suggest that *KCNE2* plays a critical role in the normal function of the native I_{to} channel complex in human heart and that M54T and I57T variants lead to a gain of function of I_{to}, which may contribute to generating potential arrhythmogeneity and pathogenesis for inherited fatal rhythm disorders.

KEYWORDS Cardiac arrhythmia; M54T variation; I57T variation; *KCNE2*; Kv4.3; Sudden cardiac death

ABBREVIATIONS CHO = Chinese hamster ovary; **HERG** = human ether-a-go-go related gene; **WT** = wild type

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Introduction

Classic voltage-gated K^+ channels consist of four pore-forming (α) subunits that contain the voltage sensor and ion selectivity filter[1,2] and accessory regulating (β) subunits.[3] *KCNE* family genes encode several kinds of β subunits consisting of single transmembrane-domain peptides that co-assemble with α subunits to modulate ion selectivity, gating kinetics, second messenger regulation, and the pharmacology of K^+ channels. Association of the *KCNE1* product minK with the α subunit Kv7.1 encoding *KCNQ1* forms the slowly activating delayed rectifier K^+ current I_{Ks} in the heart.[4,5] In contrast, association of the *KCNE2* product MiRP1 with the human ether-a-go-go related gene (HERG) forms the cardiac rapid delayed rectifier K^+ current I_{Kr}.[6]

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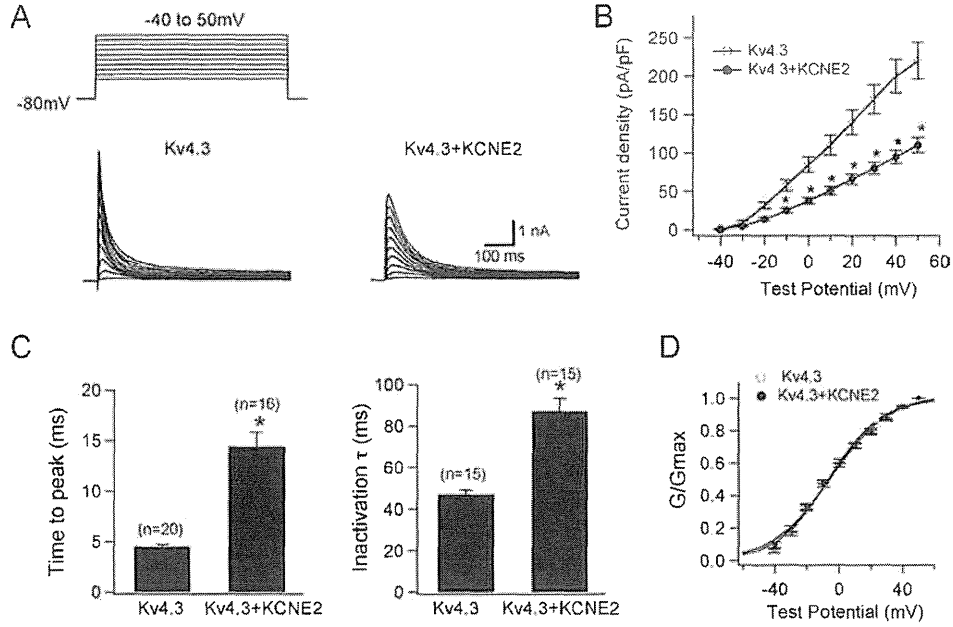

Figure 1 *KCNE2* co-expression with Kv4.3 produces smaller I_{to}-like currents with slower activation/inactivation kinetics. **A:** Representative current traces recorded from Chinese hamster ovary (CHO) cells expressing Kv4.3 (**left**) and Kv4.3 + *KCNE2* (**right**). As shown in the **inset** in panel A, depolarizing step pulses of 1-second duration were introduced from a holding potential of −80 mV to potentials ranging from −40 to +50 mV in 10-mV increments. **B:** Current–voltage relationship curve showing peak current densities in the absence and presence of co-transfected *KCNE2* (*P <.05 vs Kv4.3). **C:** Bar graphs showing the kinetic properties of reconstituted channel currents: time to peak of activation course (**left**) and inactivation time constants (**right**) measured using test potential to +20 mV (*P <.05 vs Kv4.3). Numbers in parentheses indicate numbers of experiments. **D:** Normalized conductance–voltage relationship for peak outward current of Kv4.3 and Kv4.3 + *KCNE2* channels.

Abbott et al reported that three *KCNE2* variants (Q9E, M54T, I57T) caused a loss of function in I_{Kr} and thereby were associated with the congenital or drug-induced long QT syndrome.[6,7] However, the reported QTc values in two index patients with M54T and I57T variants, both located in the transmembrane segment of MiRP1, were only mildly prolonged (390–500 ms and 470 ms).[6] We recently identified the same missense *KCNE2* variant, I57T, in which isoleucine was replaced by threonine at codon 57, in three unrelated probands showing a Brugada type 1 ECG. These findings are difficult to explain on the basis of a loss of function in I_{Kr}, thus leading us to explore other mechanisms.

Recent studies have demonstrated that interaction between α and β subunits (*KCNE*s) of voltage-gated K^+ channel is more promiscuous; for example, MiRP1 has been shown to interact with Kv7.1,[8–10] HCN1,[11] Kv2.1,[12] and Kv4.2.[13] These studies suggest that MiRP1 may also co-associate with Kv4.3 and contribute to the function of transient outward current (I_{to}) channels.[14] Indeed, a recent study reported that I_{to} is diminished in *kcne2* (–/–) mice.[15]

In the human heart, I_{to} currents are of critical importance in regulating myocardial electrical properties during the very early phase of the action potential and are thought to be central to the pathogenesis of Brugada-type ECG manifestations.[16] Antzelevitch et al demonstrated that a gain of function in I_{to} secondary to a mutation in *KCNE3* contributes to a Brugada phenotype by interacting with Kv4.3 and thereby promoting arrhythmogenicity.[14]

We hypothesized that mutations in *KCNE2* may have similar actions and characterize the functional consequences of interaction of wild-type (WT) and two mutant (I57T, M54T) MiRP1 with Kv4.3[17,18] using heterologous co-expression of these α and β subunits in Chinese hamster ovary (CHO) cells.

Methods

Heterologous expression of hKv4.3 and β subunits in CHO cells

Full-length cDNA fragment of *KCNE2* in pCR3.1 vector[10] was subcloned into pIRES-CD8 vector. This expression vector is useful in cell selection for later electrophysiologic study (see below). Two *KCNE2* mutants (M54T, I57T) were constructed using a Quick Change II XL site-directed mutagenesis kit according to the manufacturer's instructions (Stratagene, La Jolla, CA, USA) and subcloned to the same vector. Two *KCNE2* mutants were fully sequenced (ABI3100x, Applied Biosystems, Foster City, CA, USA) to ensure fidelity. Full-length cDNA encoding the short isoform of human Kv4.3 subcloned into the pIRES-GFP (Clontech, Palo Alto, CA, USA) expression vector was kindly provided by Dr. G.F. Tomaselli (Johns Hopkins University). Full-length cDNA encoding Kv channel-interacting protein (*KCNIP2*) subcloned into the PCMV-IRS expression vector was a kind gift from Dr. G.-N. Tseng (Virginia Commonwealth University). *KCND3* was transiently transfected into CHO cells together with *KCNE2* (or M54T or I57T) cDNA at equimolar ratio (*KCND3* 1.5 μg,

Table 1 Effects of *KCNE2* on Kv4.3 and Kv4.3 + KChIP2b

Parameter	Kv4.3	Kv4.3 *KCNE2*	Kv4.3 KChIP2b	Kv4.3 KChIP2b *KCNE2*
Current density at +20 mV (pA/pF)	142.0 ± 16.0 (n = 12)	66.0 ± 6.6* (n = 12)	191.5 ± 33.8 (n = 15)	77.8 ± 5.9† (n = 20)
Steady-state activation ($V_{0.5}$ in mV)	−6.5 ± 2.1 (n = 9)	−5.5 ± 1.7 (n = 11)	−7.5 ± 1.7 (n = 8)	−7.4 ± 1.4 (n = 8)
Steady-state inactivation ($V_{0.5}$ in mV)	−46.0 ± 1.3 (n = 10)	−40.8 ± 1.7* (n = 8)	−49.8 ± 1.4 (n = 7)	−44.5 ± 1.9† (n = 7)
τ of inactivation at +20 mV (τ_{inact} in ms)	47.3 ± 2.0 (n = 15)	87.2 ± 6.2* (n = 15)	47.5 ± 2.2 (n = 15)	66.6 ± 3.5† (n = 15)
Time to peak at +50 mV (TtP in ms)	4.5 ± 0.2 (n = 20)	14.4 ± 1.4* (n = 16)	4.1 ± 0.2 (n = 15)	6.1 ± 0.5† (n = 21)
τ of recovery from inactivation (ms)	419.6 ± 18.8 (n = 6)	485.6 ± 74.8 (n = 6)	89.2 ± 5.3 (n = 6)	60.2 ± 6.9† (n = 6)

*Significantly different from Kv4.3.
†Significantly different from Kv4.3 + KChIP2b.

KCNE2 1.5 μg) using Lipofectamine (Invitrogen Life Technologies, Carlsbad, CA, USA) according to the manufacturer's instructions. In one set of experiments, we also co-transfected equimolar levels of KChIP2b (*KCND3* 1.5 μg, *KCNE2* 1.5 μg, *KCNIP2* 1.5 μg). The transfected cells were then cultured in Ham's F-12 medium (Nakalai Tesque, Inc., Kyoto, Japan) supplemented with 10% fetal bovine serum (JRH Biosciences, Inc., Lenexa, KS, USA) and antibiotics (100 international units per milliliter penicillin and 100 μg/mL streptomycin) in a humidified incubator gassed with 5% CO_2 and 95% air at 37°C. The cultures were passaged every 4 to 5 days using a brief trypsin-EDTA treatment. The trypsin-EDTA treated cells were seeded onto glass coverslips in a Petri dish for later patch-clamp experiments.

Electrophysiologic recordings and data analysis

After 48 hours of transfection, a coverslip with cells was transferred to a 0.5-mL bath chamber at 25°C on an inverted microscope stage and perfused at 1 to 2 mL/min with extracellular solution containing the following (in mM): 140 NaCl, 5.4 KCl, 1.8 $CaCl_2$, 0.5 $MgCl_2$, 0.33 NaH_2PO_4, 5.5 glucose, and 5.0 HEPES; pH 7.4 with NaOH. Cells that emitted green fluorescence were chosen for patch-clamp experiments. If co-expressed with *KCNE2* (or its mutants), the cells were incubated with polystyrene microbeads precoated with anti-CD8 antibody (Dynabeads M450, Dynal, Norway) for 15 minutes. In these cases, cells that emitted green fluorescence and had attached beads were chosen for electrophysiologic recording. Whole-cell membrane currents were recorded with an EPC-8 patch-clamp amplifier (HEKA, Lambrecht, Germany), and data were low-pass filtered at 1 kHz, acquired at 5 kHz through an LIH-1600 analog-to-digital converter (HEKA), and stored on hard disk using PulseFit software (HEKA). Patch pipettes were fabricated from borosilicate glass capillaries (Narishige, Tokyo, Japan) using a horizontal microelectrode puller (P-97, Sutter Instruments, Novato, CA, USA) and the pipette tips fire-polished using a microforge. Patch pipettes had a resistance of 2.5 to 5.0 MΩ when filled with the following pipette solution (in mM): 70 potassium aspartate, 50 KCl, 10 KH_2PO_4, 1 $MgSO_4$, 3 Na_2-ATP (Sigma, Japan, Tokyo), 0.1 Li_2-GTP (Roche Diagnostics GmbH, Mannheim, Germany), 5 EGTA, and 5 HEPES (pH 7.2).

Cell membrane capacitance (C_m) was calculated from 5 mV-hyperpolarizing and depolarizing steps (20 ms) applied from a holding potential of −80 mV according to Equation 1[19]:

$$C_m = \tau_c I_0/\Delta V_m(1 - I_\infty/I_0), \quad (1)$$

where τ_c = time constant of capacitance current relaxation, I_0 = initial peak current amplitude, ΔV_m = amplitude of voltage step, and I_∞ = steady-state current value. Whole-cell currents were elicited by a family of depolarizing voltage steps from a holding potential of −80 mV. The difference between the peak current amplitude and the current at the end of a test pulse (1-second duration) was referred to as the transient outward current. To control for cell size variability, currents were expressed as densities (pA/pF).

Steady-state activation curves were obtained by plotting the normalized conductance as a function of peak outward potentials. Steady-state inactivation curves were generated by a standard two-pulse protocol with a conditioning pulse of 500-ms duration and obtained by plotting the normalized current as a function of the test potential. Steady-state inactivation/activation kinetics were fitted to the following Boltzmann equation (Eq. 2):

$$Y(V) = 1/(1 + \exp[(V_{1/2} - V)/k]), \quad (2)$$

where Y = normalized conductance or current, $V_{1/2}$ = potential for half-maximal inactivation or activation, respectively, and k = slope factor.

Data relative to inactivation time constants, time to peak, and mean current levels were obtained by using current data recorded at +50 mV or +20 mV. Recovery from inactivation was assessed by a standard paired-pulse protocol: a 400-ms test pulse to +50 mV (P1) followed by a variable

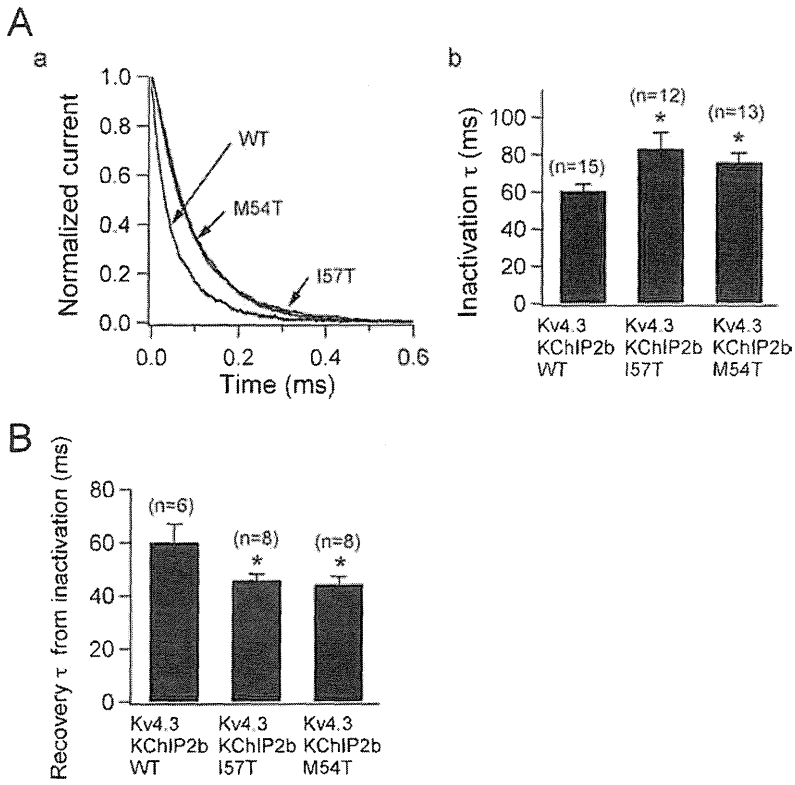

Figure 5 Two *KCNE2* variants slow inactivation kinetics and accelerate recovery from inactivation. **A, a:** Three current traces obtained from Chinese hamster ovary (CHO) cells transfected with wild-type (WT), I57T, and M54T *KCNE2* variant co-expressed with Kv4.3 and KChIP2b. Traces, which are normalized and superimposed, show that the variants slow inactivation. **A, b:** Time constants of decay at +20 mV for WT and variant *KCNE2* (*P <.05 vs Kv4.3 + KChIP2b + WT). Numbers in parentheses indicate numbers of observations. **B:** Time constants of recovery from inactivation recorded using a double-pulse protocol (*P <.05 vs Kv4.3 + KChIP2b + WT). Numbers in parentheses indicate numbers of observations.

Figure 5A shows the three traces depicted in Figure 4B normalized to their peak current level. This representation shows that the time course of inactivation of the two variant currents is slowed. The current decay was fitted by Equation 3 and the time constants (at +20 mV) summarized in Figure 5A, panel b. Finally, Figure 5B shows that the time constants of recovery of the two mutant channels from inactivation were significantly reduced. Thus, compared to WT *KCNE2*, recovery of reconstituted Kv4.3 + KChIP2b channels from inactivation was significantly accelerated with both I57T and M54T mutants.

Discussion

Kv4.3/KChIP2/MiRP1 complex can recapitulate the native I_{to}

In the present study, co-expression of WT *KCNE2* produced changes in kinetic properties (Figures 1–3 and Table 1) that led to close recapitulation of native cardiac I_{to}.[28,29] Notably, in addition to causing a positive shift of steady-state inactivation (Figure 2), *KCNE2* co-expression hastened the recovery of Kv4.3 + KChIP2b channels from inactivation (Figure 3). These modifications rendered Kv4.3 + KChIP2b channels more similar to native cardiac I_{to}, suggesting that *KCNE2* may be an important component of the native I_{to} channel complex. In contrast to a previous observation in HEK293 cells,[21] *KCNE2* co-expression decreased the current density of Kv4.3 and Kv4.3 + KChIP2b channel current in the present study, which seems to be a more reasonable result as the native I_{to} density reportedly was smaller in isolated human heart.[28] *KCNE2* co-expression has also been shown to reduce the density of Kv7.1[8,9] and HERG[6,7] channels.

Similar to the result of Deschenes and Tomaselli,[21] we failed to observe an overshoot during recovery from inactivation when *KCNE2* was co-expressed with Kv4.3 (Figure 3A), which is in contrast to the report of another group.[13] However, co-expression of *KCNE2* with Kv4.3 + KChIP2 channels produced an overshoot (Figure 3B), consistent with the report of Wettwer's group.[25] Wettwer et al also found that other *KCNE* subunits either were ineffective or induced only a small overshoot in CHO cells. Therefore, both MiRP1 and KChIP2 subunits are sufficient and necessary to recapitulate native I_{to} in the heart. Considering that the overshoot phenomenon has been described only in human ventricular I_{to} channels of the epicardial but not endocardial region,[28] these results may further implicate participation of MiRP1 and KChIP2 in the I_{to} channel complex in epicardium.

KCNE2 variants may alter the arrhythmogenic substrate by modulating I_{to}

Heterologous expression in CHO cells was conducted to examine the functional effects of I57T and M54T variants on Kv4.3 + KChIP2 channels. Both I57T and M54T

KCNE2 variants significantly (1) increased peak transient outward current density (Figure 4), (2) slowed the decay of the reconstituted I_{to} (Figure 5A), and (3) accelerated its recovery from inactivation (Figure 5B). Both variants thus caused an important gain of function in human I_{to}. These sequence changes may play a role in modulating I_{to} and thereby predispose to some inherited fatal rhythm disorders.

Functional effects on I_{to} induced by I57T and M54T resemble each other, increasing I_{to} density and accelerating its recovery from inactivation. The gain of function in I_{to} opposes the fast inward Na^+ currents during phase 0 of the action potential, leading to all or none repolarization at the end of phase 1 and loss of the epicardial action potential dome, thus promoting phase 2 reentry and fatal ventricular arrhythmias.[30]

Another *KCNE2* variant (M54T) associated with fatal arrhythmias was first identified in a woman who had a history of ventricular fibrillation and varied QT intervals.[6] It is possible that her arrhythmia was also related to a gain of function in I_{to} secondary to this variation in *KCNE2*. Interestingly, the I57T variant has been reported to produce a loss of function of HERG or Kv7.1 channels, thereby predisposing to long QT syndrome,[6,8] indicating that the same *KCNE2* variant could cause two different cardiac rhythm disorders, similar to long QT syndrome and Brugada syndrome caused by *SCN5A* mutations.[31,32]

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Neurally Mediated Syncope as a Cause of Syncope in Patients With Brugada Electrocardiogram

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Neurally Mediated Syncope in Brugada Syndrome. ***Introduction:*** **Patients with type 1 Brugada electrocardiogram (ECG) and an episode of syncope are diagnosed as symptomatic Brugada syndrome; however, all episodes of syncope may not be due to ventricular tachyarrhythmia.**

Methods and Results: **Forty-six patients with type 1 Brugada ECG (all males, 51 ± 13 years, 29 spontaneous, 17 Ic-drug induced), 20 healthy control subjects (all males, 35 ± 11 years), and 15 patients with suspected neurally mediated syncope (NMS; 9 males, 54 ± 22 years) underwent the head-up tilt (HUT) test. During the HUT test, 12-lead ECGs were recorded in all patients, and the heart rate variability was investigated in some patients. Sixteen (35%) of 46 patients with Brugada ECG, 2 (10%) of 20 control subjects, and 10 (67%) of 15 patients with suspected NMS showed positive responses to the HUT test. Although no significant differences were observed in HUT-positive rate among Brugada patients with documented VT (7/14; 50%), syncope (5/19; 26%) and asymptomatic patients (4/13; 31%), the HUT-positive rate was significantly higher in patients with documented VT (50%) and those with VT or no symptoms (11/27, 41%) compared to that in control subjects (10%) ($P < 0.05$). Augmentation of ST-segment amplitude (≥0.05 mV) in leads V1-V3 was observed in 11 (69%) of 16 HUT-positive patients with Brugada ECG during vasovagal responses, and was associated with augmentation of parasympathetic tone following sympathetic withdrawal.**

Conclusion: **Thirty-five percent of patients with Brugada ECG showed vasovagal responses during the HUT test, suggesting that some Brugada patients have impaired balance of autonomic nervous system, which may relate to their syncopal episodes.** *(J Cardiovasc Electrophysiol, Vol. 21, pp. 186-192, February 2010)*

autonomic nervous system, Brugada syndrome, head-up tilt test, syncope, sudden death

Introduction

Brugada syndrome is characterized by ST-segment elevation in the right precordial leads V1 through V3 and an episode of ventricular tachyarrhythmia (VT) in the absence of structural heart disease.[1-3] In patients with Brugada syndrome, syncopal episodes are generally thought to be due to VT; however, all episodes of syncope may not be owing to VT events. Neurally mediated syncope (NMS) is 1 of the causes of syncope in general population, and it refers to a reflex response that some triggering factors give rise to arterial vasodilatation associated with relative or absolute bradycardia.[4] In general, the overall prognosis in patients with NMS is quite favorable.[4] On the other hand, the precise cause of syncope in patients with Brugada syndrome is difficult to determine. Therefore, the therapeutic strategy for Brugada patients with syncope is often problematic. The aim of this study was to evaluate the possibility of NMS as a cause of syncope in patients with Brugada electrocardiogram (ECG).

Methods

Patients Population

The study population consisted of 46 consecutive patients with type 1 Brugada ECG who were admitted to the National Cardiovascular Center, Suita, Japan, between May 2004 and March 2006 (all males, ages 26 to 77; mean 51 ± 13 years, 29 spontaneous, 17 Ic-drug induced), 20 healthy control subjects (all males, 35 ± 11 years), and 15 patients suspected of NMS (9 males, 54 ± 22 years). Ethical approval was obtained from the Institutional Review Committee of our hospital, and all patients and control subjects gave their informed, written consent before participation. The control subjects and the patients with suspected NMS showed no structural heart diseases, normal physical examination results, and normal 12-lead ECGs, and received no drug treatment affecting the sympathetic nervous system. Type 1 Brugada ECG was defined as a coved type ST-segment elevation of ≥ 0.2 mV at

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J point observed in more than 1 of the right precordial leads (V1 to V3) in the presence or absence of a sodium channel blocker.[2]

Head-Up Tilt Test

The HUT test was performed in the afternoon after 4 hours of fasting in a quiet and comfortable room equipped for cardiopulmonary resuscitation. All patients were allowed to lie on an electrically controlled tilt table an intravenous line containing 5% dextrose was inserted into 1 arm, and allowed to rest in supine position for at least 10 minutes. A positive HUT test was defined by the development of syncope or presyncope associated with relative bradycardia (≥20% decrease in heart rate compared with baseline) or hypotension (systolic blood pressure <80 mmHg). Presyncope was defined as the induction of symptoms of imminent syncope, and syncope was defined as sudden transient loss of consciousness. Positive response to the HUT test was classified into 3 types owing to hemodynamic status, such as vasodepressor type (hypotension without significant bradycardia), cardioinhibitory type (bradycardia without associated hypotention), and mixed type (hypotension followed by bradycardia).[4] At first, we performed passive tilt (Control-Tilt) at an angle of 70 degrees for 30 minutes. When Control-Tilt was negative, sublingual nitroglycerin (NTG) spray 0.3 mg was administered, and the test was continued for 15 minutes (NTG-Tilt). The endpoint of each tilt test was the time when patients showed positive responses or the completion of HUT-protocol.

Parameters Measured During the Head-Up Tilt Test

Heart rate and blood pressure

Heart rate was monitored, and cuff blood pressure was measured by electrosphygmomanometry with a microphone placed over the brachial artery to detect Korotkoff sounds every minute (STBP-780, Colin Electronics, Komaki, Japan) in all patients during the HUT test.

ST-segment amplitude in the right precordial leads

Twelve-lead ECGs were recorded every 1 minute during the HUT test, and the changes of ST-segment amplitude in the right precordial leads (V1-V3) were analyzed (ML-6500, Fukuda-denshi, Tokyo, Japan) in all patients during the HUT test.

Heart rate variability

Six-lead ECGs from the Task Force Monitor (CNSystem, Graz, Austria)[5–7] were measured for beat-to-beat heart rate and consecutive R-R intervals in 10 patients with Brugada ECG (4 documented VT, 5 syncopal episode only, and 1 asymptomatic), 9 control subjects, and 5 patients with suspected NMS. The heart rate variability (HRV) was investigated by a power spectral analysis delineating the low-frequency component (LF; 0.04–0.15 Hz) and the high-frequency component (HF; 0.15–0.40 Hz).[8] We analyzed the normalized unit of the HF components (%) calculated automatically (HF/power spectral density-very low-frequency component [0–0.04 Hz] × 100)[8,9] and the LF/HF ratio. The HF indicates the tone of the parasympathetic nervous system, and the LF/HF ratio indicates the sympathovagal balance.

Statistical Analysis

Numerical values were expressed as means ± SD unless otherwise indicated. Comparisons of parameters between 2 groups were made using the unpaired Student *t*-test. Comparisons of parameters among 3 groups were made with a one-way analysis of variance (ANOVA), followed by the Scheffe's multiple-comparison test. Categorical variables were compared using a chi-square analysis using the Yate's correction or Fisher exact test if necessary. An overall chi-square test for a 2 × n table was performed when comparisons involved >2 groups. A P-value < 0.05 was considered significant.

Results

Clinical Characteristics

The clinical characteristics of 46 patients with Brugada ECG and 15 patients with suspected NMS are shown in Table 1. The patients with Brugada ECG were divided into 3 groups: (1) 14 patients with documented VT; (2) 19 patients with syncopal episodes only; and (3) 13 asymptomatic patients. No significant differences were observed in age, incidence of spontaneous type 1 ECG, family history of sudden cardiac death (SCD), induced ventricular fibrillation during electrophysiologic study (EPS), and *SCN5A* mutation. Implantable cardioverter-defibrillator (ICD) was implanted more frequently in patients with documented VT. The triggers of VT and/or syncope are also shown in Table 1. Seventy-nine percent of VT episodes occurred during sleep or at rest in patients with documented VT ($P < 0.0001$ vs the patients with syncopal episodes only and suspected NMS). On the other hand, in patients with syncopal episodes only, 15% of syncopal episodes occurred after urination, 21% during standing, and 21% after drinking alcohol, which seemed to be similar patterns in patients with suspected NMS. Based on the clinical description of the syncopal events, 16 (84%) of 19 Brugada patients with syncopal episodes were suspected to have NMS. Syncopal episodes seemed to be due to VT in 1 of the remaining 3 patients.

Positive Response to the Head-Up Tilt Test

Comparison of the positive responses to the HUT test between 46 patients with Brugada ECG and 20 control subjects along with 15 patients with suspected NMS are shown in Table 2. Sixteen (35%) of 46 patients with Brugada ECG showed positive responses. Positive responses were developed in 1 (2%) of 46 patients during Control-Tilt and in 15 (33%) of 45 patients during NTG-Tilt, and the mixed type was predominant (94%). In patients with Brugada ECG, there were no significant differences in the incidence of positive responses among patients with documented VT (50%), those with syncopal episodes only (26%), and asymptomatic patients (31%). No significant differences were observed in the type of positive responses between the 3 groups. The mixed type was predominant (100%, 100%, and 75%, respectively), and cardioinhibitory type was not observed in all 3 groups. Two (10%) of 20 control subjects and 10 (67%) of 15 patients with suspected NMS showed positive responses. The HUT-positive rate was not significantly different between all 46 patients with Brugada ECG, 20 control subjects and 15 subjects with suspected NMS (35% vs 10% vs 67%);

TABLE 1

Clinical Characteristics of Patients with Brugada Electrocardiogram and Suspected NMS

	Documented VT (n = 14)	Syncopal Episodes only (n = 19)	Asymptomatic (n = 13)	Suspected NMS (n = 15)
Age (years)	50 ± 15	51 ± 12	52 ± 14	54 ± 22
Spontaneous type 1 ECG	10 (71)	9 (47)	10 (77)	–
Family history of SCD	4 (29)	4 (21)	4 (31)	–
Induced VF during EPS	10/12 (83)	15/18 (83)	8/11 (73)	–
SCN5A mutation	1 (7)	3 (16)	0 (0)	–
ICD implantation	14 (100)	13 (68)*	7 (54)*	–
Triggers of syncope				
During sleeping or at rest	11 (79)	1 (5)*	–	0*
After urination	0	3 (15)	–	1 (7)
Prolonged standing at attention	0	4 (21)	–	4 (27)
After drinking alcohol	0	4 (21)	–	6 (40)
After meal	1 (7)	0	–	0
After exertion	0	2 (11)	–	2 (13)
After sudden unexpected pain	0	2 (11)	–	0
During driving	0	1 (5)	–	0
Others	2 (14)	2 (11)	–	2 (13)

Values are mean ± SD for age, and expressed as frequency (%). *P < 0.05 vs documented VT group. ECG = electrocardiogram; EPS = electrophysiological study; ICD = implantable cardioverter-defibrillator; NMS = neurally mediated syncope; SCD = sudden cardiac death; VT = ventricular tachyarrhythmias; VF = ventricular fibrillation.

however, the HUT-positive rate was significantly higher in 14 patients with documented VT (50%) and 27 patients with VT or no symptoms (41%) compared to that in control subjects (10%) (P = 0.03, P = 0.04, respectively). The HUT-positive rate in 19 Brugada patients with syncopal episodes (26%) was significantly lower than that in 15 patients with suspected NMS (P = 0.04), although the syncopal episodes in 84% of the 19 patients were suspected to be due to NMS. Positive responses to the HUT test were more frequency observed in 15 patients with suspected NMS compared to those in 20 control subjects (10/15 vs 2/20; P < 0.001).

Comparison of the clinical characteristics between 16 HUT-positive patients and 30 HUT-negative patients with Brugada ECG were shown in Table 3. No significant differences were observed in cardiac events, such as documented VT or syncope. Furthermore, there were no significant differences in the clinical characteristics, such as age, spontaneous type 1 ECG, a family history of SCD, inducibility of ventricular fibrillation during EPS, *SCN5A* mutation, and ICD implantation.

Response of Heart Rate and ST-Segment Amplitude

In patients with Brugada ECG, the heart rate was increased by 12 ± 9 beats/min during Control-Tilt, and by 24 ± 14 beats/min during NTG-Tilt. As the heart rate was increased, decrease of ST-segment amplitude of ≥ 0.05 mV from baseline in the right precordial leads was observed in 11 (24%) of 46 patients during Control-Tilt (−0.14 ± 0.08 mV), and in 19 of 45 (42%) patients during NTG-Tilt (−0.15 ± 0.10 mV) (Fig. 1C). However, augmentation of ST-segment amplitude of ≥ 0.05 mV in the right precordial leads was observed just before and after positive responses to the HUT test in 11 (69%) of 16 HUT-positive patients (0.10 ± 0.06 mV) (Figs. 1D and E). These significant ST-segment augmentation was observed in 1 patient during Control-Tilt (documented VT), and 10 patients during NTG-Tilt (5 documented VT, 2 syncopal episodes only, 3 asymptomatic), respectively. On the other hand, augmentation of the ST-segment amplitude of ≥ 0.05 mV was 2 (7%) of 30 HUT-negative patients during NTG-Tilt (1 documented VT, 1 syncopal episodes only). As a result, the average ST-segment augmentation was

TABLE 2

Responses to Head-Up Tilt Test in Patients with Brugada Electrocardiogram, Control Subjects, and Patients with Suspected NMS

	All (n = 46)	Documented VT (n = 14)	Syncopal Episodes Only (n = 19)	Asymptomatic (n = 13)	Brugada ECG with VT or No Symptoms (n = 27)	Control Subjects (n = 20)	Suspected NMS (n = 15)
Age (years)	51 ± 13*	50 ± 15*	51 ± 12*	52 ± 14*	51 ± 14*	35 ± 11	54 ± 22*
Positive response	16 (35)	7 (50)*	5 (26)†	4 (31)	11 (41)*	2 (10)	10 (67)*
Control-tilt	1/46 (2)	1/14 (7)	0/19 (0)	0/13 (0)	1/27 (4)	0/20 (0)	0/15 (0)
NTG-tilt	15/45 (33)†	6/13 (46)*	5/19 (26)†	4/13 (31)	10/26 (38)	2/20 (10)	10/15 (67)*
Type of positive response							
Vasodepressive	1/16 (6)	0	0	1/4 (25)	1/11 (9)	0	1/10 (10)
Cardioinhibitory	0	0	0	0	0	0	0
Mixed	15/16 (94)	7/7 (100)	5/5 (100)	3/4 (75)	10/11 (91)	3 (100)	9/10 (90)

Values are expressed as frequency (%). *P < 0.05 vs control subjects, †P < 0.05 vs suspected NMS. ECG = electrocardiogram; NMS = neurally mediated syncope; NTG = nitroglycerin; VT = ventricular tachyarrhythmias.

TABLE 3

Comparison of Clinical Characteristics Between Head-up Tilt-Positive Patients and Head-up Tilt-Negative Patients

	HUT-Positive (n = 16)	HUT-Negative (n = 30)	P-value
Age (years)	52 ± 13	50 ± 14	0.58
Documented VT	7 (44)	7 (23)	0.15
Syncope only	5 (31)	14 (47)	0.49
Asymptomatic	4 (25)	9 (30)	0.99
Spontaneous type 1 ECG	11 (69)	18 (60)	0.79
Family history of SCD	4 (25)	8 (27)	1.0
Induced VF during EPS	13/15 (87)	20/26 (77)	0.72
SCN5A mutation	1 (6)	3 (10)	1.0
ICD implantation	14 (88)	24 (80)	0.82

Values are expressed as frequency (%). ECG = electrocardiogram; EPS = electrophysiological study; HUT = head-up tilt test; ICD = implantable cardioverter-defibrillator; SCD = sudden cardiac death; VT = ventricular tachyarrhythmias; VF = ventricular fibrillation.

significantly larger in 16 HUT-positive patients than in 30 HUT-negative patients at similar heart rate (0.06 ± 0.06 mV vs −0.04 ± 0.06 mV, $P < 0.0001$). No ventricular arrhythmias were induced during the HUT test in any patients with Brugada ECG. The ST-segment augmentation was not observed during the HUT test in any control subjects (−0.02 ± 0.02 mV, $P < 0.0001$ vs 16 HUT-positive Brugada patients) and patients with suspected NMS (−0.02 ± 0.04 mV, $P < 0.001$ vs 16 HUT-positive Brugada patients; Fig. 2).

Heart Rate Variability and ST-segment Amplitude

Positive responses during NTG-Tilt were observed in 4 (40%) of 10 patients with Brugada ECG, in 1 (11%) of 9 control subjects, and in 4 (80%) of 5 patients with suspected NMS in whom the HRV was monitored. The autonomic activities in a representative NTG-Tilt-positive patient with Brugada ECG and those with suspected NMS are shown in Figure 3A and B, respectively. Before positive responses to the HUT test, sympathetic activity (LF/HF ratio) dramatically increased; and then, sympathetic withdrawal occurred immediately. Thereafter, parasympathetic nerve activity (the normalized unit of the HF components) gradually increased. The similar pattern of augmented parasympathetic nerve activity following sympathetic withdrawal during positive responses to the HUT test was observed in all 9 HUT-positive patients. The patterns of HRV were not different among the HUT-positive patients with Brugada ECG, the HUT-positive control subjects, and the HUT-positive patients with suspected NMS. In 3 (75%) of 4 HUT-positive patients with Brugada ECG, the LF/HF ratio decreased and the HF component increased gradually toward the maximum ST-segment elevation just before and after positive response for the HUT test (Fig. 3A), but ST-segment was decreased in patients with NMS (Fig. 3B).

Discussion

In this study, 35% of patients with Brugada ECG showed vasovagal responses during the HUT test regardless of the presence VT or syncope. The HUT test was also positive in 41% among only Brugada patients with documented VT or no symptoms. During vasovagal response, ST-segment augmentation in the right precordial leads (V1-V3) was observed in 11 (69%) of 16 HUT-positive patients with Brugada ECG, but no ventricular arrhythmias were induced in any HUT-positive patients.

Neurally Mediated Syncope as a Cause of Syncope in Brugada Syndrome

Several case reports have described patients exhibiting clinical phenotype of both Brugada syndrome and NMS.[10-12]

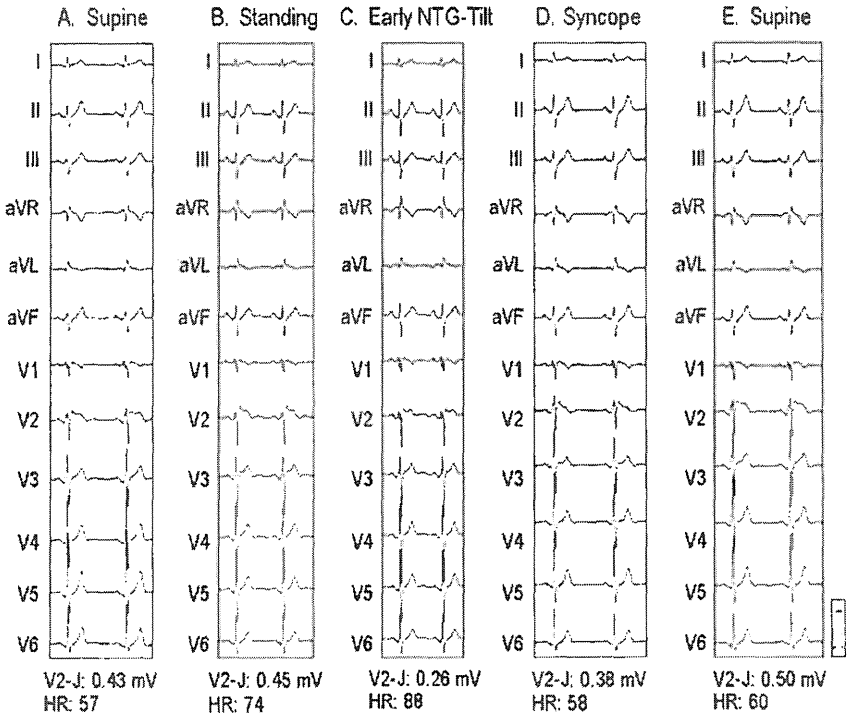

Figure 1. *The12-leads electrocardiogram (ECG) during head-up tilt test in a representative nitroglycerin (NTG)-Tilt-positive patient with type 1 Brugada ECG at supine position (A), at standing position (B), at early phase of NTG-Tilt (C), at syncope (D), and at supine position following syncope (E). The ST-segment elevation was decreased from 0.45 mV to 0.26 mV at early phase of NTG-Tilt as the heart rate was increased (C), while it was augmented to 0.38 mV at syncope (D), and to 0.50 mV at supine position following syncope (E).*

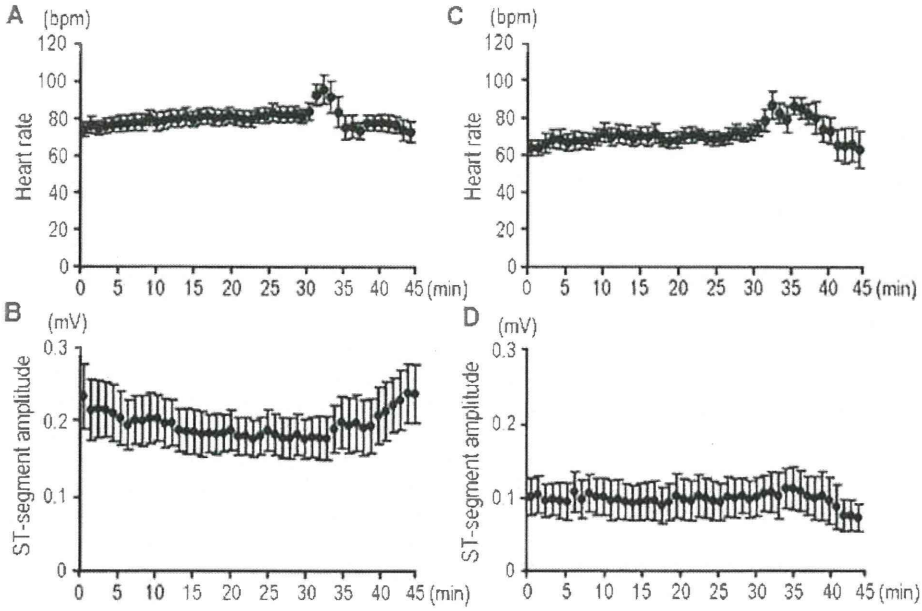

Figure 2. *Response of the heart rate and ST-segment amplitude during the head-up tilt (HUT) test in 16 HUT-positive patients with Brugada electrocardiogram (ECG) (A, B) and in 10 HUT-positive patients with suspected neurally mediated syncope (NMS) (C, D). At first, the passive tilt (Control-Tilt) was performed for 30 minutes (0–30 minutes). When Control-Tilt was negative, nitroglycerin tilt was continued for 15 minutes (30–45 minutes). The responses of heart rate during positive responses to the HUT test were similar in patients with Brugada ECG (A) to those in patients with suspected NMS (C). In patients with Brugada ECG, ST-segment in lead V2 was augmented before and after positive responses to the HUT test (B), but not in those with suspected NMS (D).*

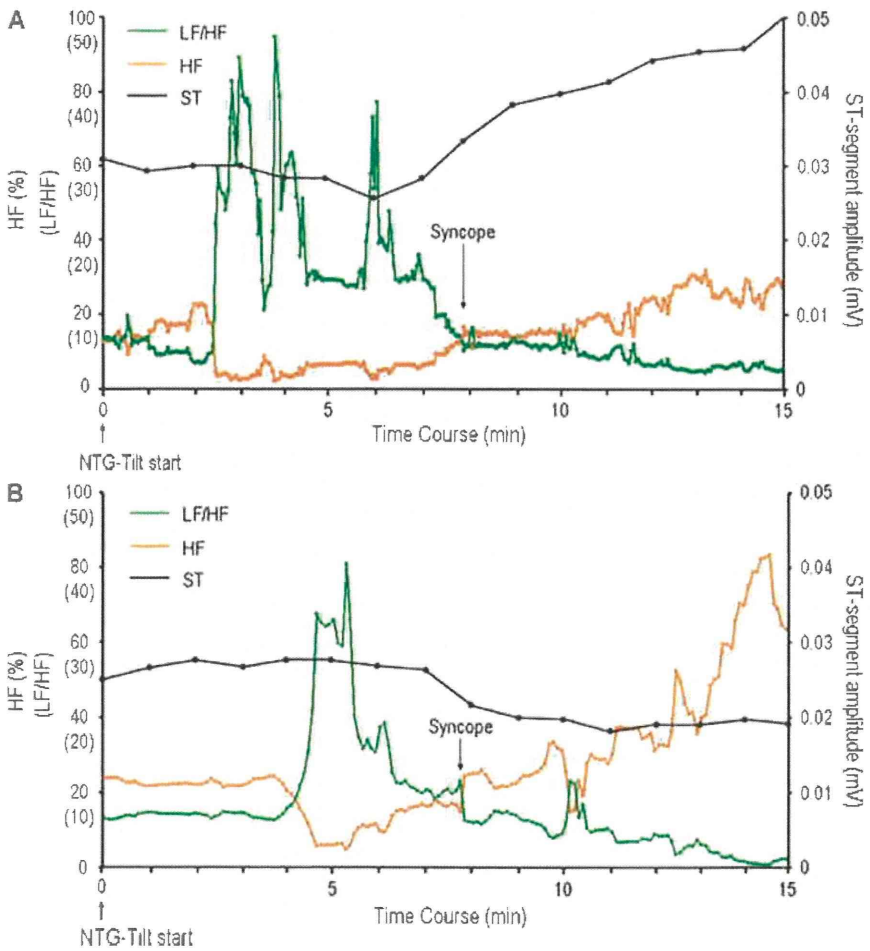

Figure 3. *Autonomic responses during head-up tilt (HUT) test. The autonomic activities in a representative nitroglycerin (NTG)-Tilt-positive patient with type 1 Brugada electrocardiogram (ECG) (A) and those in a representative NTG-Tilt-positive patient with suspected NMS (B). Before tilt-induced syncope, sympathetic activity (LF/HF ratio) dramatically increased; and then, sympathetic withdrawal occurred immediately. Thereafter, parasympathetic nerve activity (the normalized unit of the HF components) gradually increased. In the HUT-positive patient with Brugada ECG, ST-segment augmentation in lead V2 was observed just before and after positive responses, and the LF/HF ratio decreased and the HF component increased gradually toward the maximum ST-segment elevation (A). In contrast, in the HUT-positive patient with suspected NMS, ST-segment amplitude in lead V2 was decreased gradually after positive responses (B).*

It is well known that the autonomic nervous system plays an important role on the arrhythmogenesis of Brugada syndrome. Previous studies showed that the withdrawal of sympathetic activity and the sudden rise in vagal activity was an important triggering factor of ventricular fibrillation.[13-15] Similarly, it has been presumed that parasympathetic tone increase during NMS events in patients with Brugada ECG. Recent basic study showed that *SCN5A*, a major responsible gene in Brugada patients, is expressed not only in the myocardial cells but also in intracardiac ganglia.[16] Makita *et al.* also demonstrated a novel nonsense mutation in *SCN5A* gene in a patient with Brugada syndrome who had been diagnosed as NMS.[17] These results suggested that the abnormal regulation or imbalance of autonomic nervous system may exist regardless of the presence or absence of cardiac events in patients with Brugada ECG.

ST-Segment Elevation in the Precordial Leads During the HUT Test in Patients with Brugada ECG

In Brugada syndrome, spontaneous augmentation of ST-segment elevation occurred along with an increase in vagal activity, especially just before and after the occurrence of ventricular fibrillation.[14] The ST-segment elevation is also known to be modulated by exercise,[18] pharmacological interventions that interact with automatic nervous activities,[19] or taking meals associated with glucose-induced insulin levels.[20] In this study, ST-segment augmentation in the right precordial leads was observed just before and after positive responses to the HUT test in two-thirds (69%) of the HUT-positive patients with Brugada ECG but only in 7% of the HUT-negative patients. In patients with Brugada ECG, the preceding increase of sympathetic nerve activity during the HUT test may cause augmentation of ICa-L, resulting in attenuation of ST-segment elevation.[19] Subsequent augmentation of parasympathetic nerve activity during the HUT test may decrease of ICa-L, and increase Ito, thus augmenting ST-segment amplitude.

Clinical Implication

The second consensus report suggested that symptomatic patients displaying type 1 Brugada ECG (either spontaneous or after class Ic drugs) who present with aborted sudden death should undergo ICD implantation.[3] ICD implantation is also recommended in patients with syncope, seizure, or nocturnal agonal respiration, after noncardiac causes of these symptoms have been carefully ruled out.[3] Needless to say, the ECG recording during syncope is the only convincing way to rule in or out VT during syncope, and only clinical judgment can be used to guide diagnostic and therapeutic decisions. However, in patients with Brugada syndrome, there is an abnormal regulatory imbalance of the autonomic nervous system that may be a common denominator to both syncope and ventricular fibrillation.

Limitations

The control subjects were significantly younger than patients with Brugada ECG or those with suspected NMS. However, it is reported that the positive rate of NTG-Tilt in the elderly was comparable to that seen in younger subjects.[21] Therefore, lower incidence of positive rate of the HUT test in the control subjects than that in the other 2 groups was not due to the relevant difference of age. The incidence of spontaneous type 1 ECG and the positive rate of the HUT test are smaller in Brugada patients with syncope episodes only than in those with documented VT or asymptomatic patients; however, statistical significance was not observed between the 3 groups.

Conclusions

Thirty-five percent of patients with Brugada ECG showed vasovagal responses during the HUT test. The HUT test was also positive in 41% among only Brugada patients with documented VT or no symptoms. During vasovagal response, ST-segment augmentation in the right precordial leads was observed in 69% of the HUT-positive Brugada patients, but no ventricular arrhythmias were induced. These data suggest that some Brugada patients have impaired balance of autonomic nervous system, which may relate to their syncopal episodes. Additional studies including a large number of subjects are needed to validate our findings and possibly evaluate the role of the HUT test in risk stratification of patients with Brugada ECG.

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Structural Heterogeneity in the Ventricular Wall Plays a Significant Role in the Initiation of Stretch-Induced Arrhythmias in Perfused Rabbit Right Ventricular Tissues and Whole Heart Preparations

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Rationale: Mechanical stress is known to alter the electrophysiological properties of the myocardium and may trigger fatal arrhythmias when an abnormal load is applied to the heart.

Objective: We tested the hypothesis that the structural heterogeneity of the ventricular wall modulates globally applied stretches to create heterogeneous strain distributions that lead to the initiation of arrhythmias.

Methods and Results: We applied global stretches to arterially perfused rabbit right ventricular tissue preparations. The distribution of strain (determined by marker tracking) and the transmembrane potential (measured by optical mapping) were simultaneously recorded while accounting for motion artifacts. The 3D structure of the preparations was also examined using a laser displacement meter. To examine whether such observations can be translated to the physiological condition, we performed similar measurements in whole heart preparations while applying volume pulses to the right ventricle. At the tissue level, larger stretches (≥20%) caused synchronous excitation of the entire preparation, whereas medium stretches (10% and 15%) induced focal excitation. We found a significant correlation between the local strain and the local thickness, and the probability for focal excitation was highest for medium stretches. In the whole heart preparations, we observed that such focal excitations developed into reentrant arrhythmias.

Conclusions: Global stretches of intermediate strength, rather than intense stretches, created heterogeneous strain (excitation) distributions in the ventricular wall, which can trigger fatal arrhythmias. (*Circ Res*. 2010;106:176-184.)

Key Words: stretch-induced arrhythmia ■ mechanoelectric feedback ■ optical mapping

Alterations to the mechanical state of the myocardium affect its electrophysiological properties, a phenomenon termed mechanoelectric feedback (MEF).[1,2] MEF is considered to play a significant role in the genesis of cardiac rhythm disturbances in various disease states, such as myocardial infarction and heart failure, in which myocardial tissues are subjected to abnormal loading conditions.[3–5] This speculation is supported by previous observations that in myocardial infarction, ventricular ectopic excitations are initiated by acute stretches of the border zone between the infarct and the normal myocardium.[6–8] A more definite causality is suspected in the etiology of commotio cordis, where sudden death occurs owing to a nonpenetrating chest wall impact in the absence of injury to the ribs, sternum, and heart.[9,10] Using anesthetized juvenile swine, Link et al[10] found that ventricular fibrillation can be produced by a baseball strike, and examined the effects of the phase, strength and speed of the strike for the induction of arrhythmias.

To elucidate the mechanisms underlying MEF and related arrhythmias, extensive studies have been carried out using various preparations from various species, including rabbits, lambs and dogs.[11–13] Stretch-activated channels (SACs) have been regarded as the most likely candidates for the primary transducers of mechanical stress.[14–16] Although such findings at the molecular level can account for changes in the action potential duration, amplitude, effective refractory period and resting potential induced by mechanical interventions at the cellular level, we still face a huge gap between these laboratory findings and clinical arrhythmias observed at the organ level. In this context, Franz et al[17] investigated the effects of increases in ventricular volume and pressure on epicardial monophasic action potentials in both isolated cross-circulated hearts and

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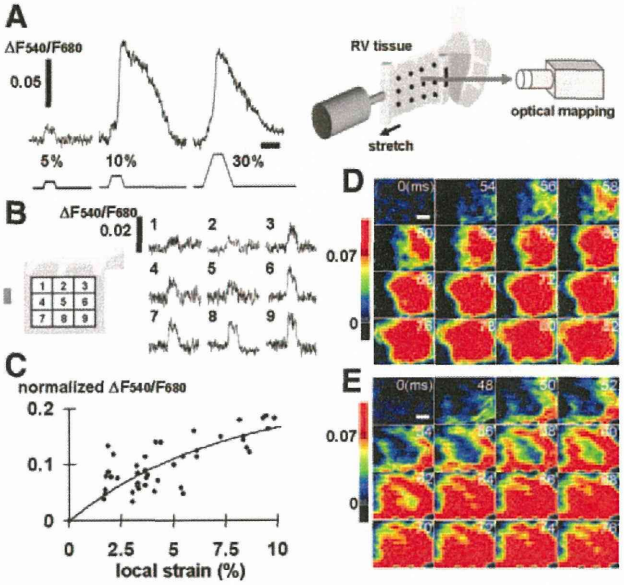

Figure 1. Alterations in the electric response in a cardiac tissue. A, Ratiometric optical signals ($\Delta F_{540}/F_{680}$) in response to 5%, 10%, and 30% stretches from left to right. Scale bar: 100 ms. B, Spatiotemporal pattern of the depolarizations (typical optical signals in each segment) in response to a 5% stretch. C, Relationship between the changes in the normalized optical signals and the local strain under the excitation threshold (n=5). The smooth curve through the data points was fitted with a nonlinear regression model. D and E, Representative action potentials and optical maps in response to 10% and 30% stretches, respectively. The stretch starts at 0 ms. Scale bar: 4 mm.

in situ canine hearts to clearly demonstrate the manifestation of MEF. However, these volume and/or pressure alterations do not allow detailed evaluation of the changes in myocardial stress or strain, which are believed to be the keys for establishing a link between the macroscopic and microscopic phenomena.

To elucidate how the cellular responses to stretches lead to arrhythmias in the heart, we focused on the morphology of tissue preparations and its role in the modulation of the electric responses. We developed an experimental set-up in which controlled uniaxial stretches were applied to crystalline perfused rabbit ventricular walls while monitoring the local strain. The use of optical transmembrane potential mapping combined with a tissue tracking technique enabled us to examine the relationship between local strain and excitation of the myocardium. By applying acute stretches of varying amplitudes, we demonstrate that global stretches applied to the ventricular wall tissue can create strain dispersion in the heterogeneous structure of the ventricular wall and that mechanical insults of intermediate, rather than intense, strength induce focal excitation, thus potentially triggering fatal arrhythmias. Finally, using whole heart preparations, we confirm that only medium stretches of the myocardium can evoke spiral wave formation.

Methods

Japanese white rabbits weighing 2.4 to 2.9 kg were used. The distribution of strain and the transmembrane potential were simultaneously recorded while applying an acute stretch to right ventricle (RV) tissue preparations. The 3D structure of the preparations was also examined. Similar measurements were conducted in whole heart preparations while applying acute volume pulses to the RV.

An expanded Methods section is available in the Online Data Supplement at http://circres.ahajournals.org.

Non-standard Abbreviations and Acronyms

MEF	*mechanoelectric feedback*
SAC	stretch-activated channel
RV	right ventricle

Results

Effect of the Stretch Amplitude on Excitation of the Tissue

To elucidate the relationship between the electric response and the stretch level, we measured the optical transmembrane potential signals of stretched tissues. Figure 1 shows representative transmembrane potential signals in response to stretches of varying amplitudes. When a uniaxial stretch with a small amplitude (5%) was applied, the myocardial tissue was depolarized but an action potential did not develop (Figure 1A, left). The distribution of these depolarizations was heterogeneous and the amplitudes of these depolarizations had a positive dependence on the local strains (n=5) (Figure 1B and 1C). However, above a certain level of amplitude (≥10%), we observed focal excitation (development of an action potential in less than 4 segments of 9 blocks) (Figure 1A, middle; Figure 1D). A larger stretch (30%) only induced multiple occurrences of excitation in the tissue (Figure 1E). Figure 2A shows the relationship between the probability of tissue excitation (development of an action potential in at least one locus within the tissue) and the amplitude of the stretch applied (global strain). We found a fairly abrupt transition in the tissue responses to a uniaxial stretch (n=7). Specifically, excitation was rare when the amplitude was small (5%), but its rate increased with stretches in the medium range (10% and 15%) to reach 100% (sure observation) in response to large stretches (20%, 25% and 30%).

The use of a trapezoidal command with constant rates of rise and fall necessarily made the entire duration of the stretch longer for larger stretches, which may thus have led to modulation of the responses of the myocardium through different mechanisms. To exclude these possibilities, we applied stretches of varying amplitudes while keeping the entire duration constant at 50 ms. We found similar responses, thereby indicating that the amplitude rather than the duration is the major determinant of stretch-induced activation of the myocardium (Online Figure V, A). We also confirmed that stretches applied during the action potentials could modulate their shapes, and sometimes found stretch-activated depolarizations followed by premature ventricular contractions (Online Figure V, B).

Relationship Between Stretch-Induced Excitation and Epicardial Local Strain

We also evaluated the relevance between stretch-activated excitation and epicardial local strain (n=7). To compare the