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Review

In Vitro and *In Vivo* Genotoxicity Induced by Fullerene (C_{60}) and Kaolin

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Nanomaterials are being utilized for many kinds of industrial products, and the assessment of genotoxicity and safety of nanomaterials is therefore of concern. In the present study, we examined the genotoxic effects of fullerene (C_{60}) and kaolin using *in vitro* and *in vivo* genotoxicity systems. Both nanomaterials significantly induced micronuclei and enhanced frequency of sister chromatid exchange (SCE) in cultured mammalian cells. When ICR mice were intratracheally instilled with these nanomaterials, DNA damage of the lungs increased significantly that of the vehicle control. Formation of DNA adducts in the lungs of mice exposed to nanomaterials were also analyzed by stable isotope dilution LC-MS/MS. 8-Oxodeoxyguanosine and other lipid peroxide related adducts were increased by 2- to 5-fold in the nanomaterial-exposed mice. Moreover, multiple (four consecutive doses of 0.2 mg per animal per week) instillations of C_{60} or kaolin, increased *gpt* mutant frequencies in the lungs of *gpt* delta transgenic mice. As the result of mutation spectrum analysis, G:C to C:G transversions were commonly increased in the lungs of mice exposed to both nanomaterials. In addition, G:C to A:T was increased in kaolin-exposed mice. In immunohistochemical analysis, many regions of the lungs that stained positively for nitrotyrosine (NT) were observed in mice exposed to nanomaterials. From these observations, it is suggested that oxidative stress and inflammatory responses are probably involved in the genotoxicity induced by C_{60} and kaolin.

Key words: nanomaterials, genotoxicity, fullerene (C_{60}), kaolin, DNA adducts

Introduction

Recently, nanomaterials are being utilized for cosmetics and industrial products, and applications in medicine are under consideration. The assessment of genotoxicity and safety of nanomaterials is therefore of concern. One reason behind this is the asbestos crisis (1). Some nanomaterials are not only nano-sized particles, but also asbestos shape-like fibers, and the carcinogenic potential of such nanomaterials has attracted much attention over the years. Moreover, it is thought that nano-sized particles can be taken up in cells and cause intracellular damage (2,3). With this background, we here investigated induction of *in vitro* and *in vivo* genotoxicity using fullerene (C_{60}) and kaolin as examples. To clarify the mechanisms of mutations due to these nanomaterials, we analyzed the formation of DNA adducts in the lungs of mice after exposure. Here, we briefly summarize our data and also discuss mechanisms of genotoxicity induced by nanomaterials.

Size Distribution in Suspensions of Nanomaterials

The size distribution of nanomaterials used in the present study was analyzed by dynamic light scattering (DLS) as described previously (4). The most abundant sizes were at 234.1 ± 48.9 and 856.5 ± 119.2 nm for C_{60} and 357.6 ± 199.4 nm for kaolin, respectively.

In Vitro Genotoxicity Test

Micronucleus test: The micronucleus genotoxicity/clastogenicity test is widely used for assessment of environmental substances and medicinal chemicals. Here, we investigated the micronucleus inducing activity of C_{60} and kaolin using human lung carcinoma A549

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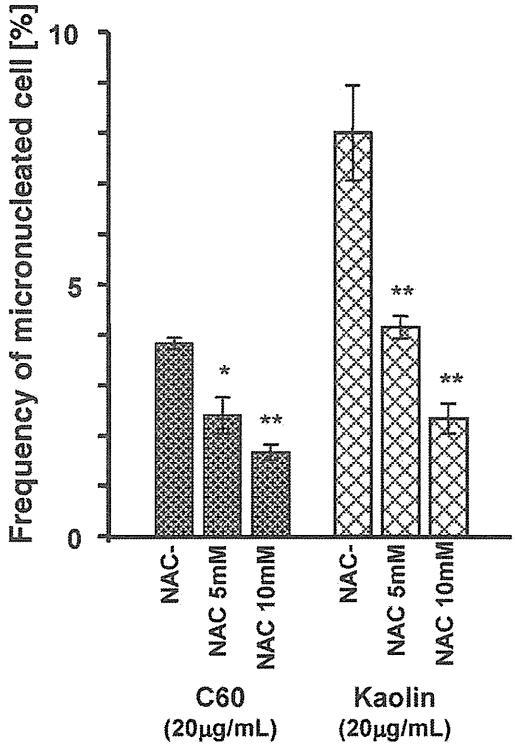

Fig. 1. Effects of anti-oxidative agents on the micronucleus inducing activity of nanoparticles. Values represent the means of three experiments ± SD. Asterisks (*, ** for $p<0.05$ and $p<0.01$, respectively) indicate significant differences from cells without NAC in the Student's t-test. Concentrations of nanoparticles in $\mu g/cm^2$ are given in parentheses.

cells (4). Six-hours treatment with 200 μg/mL kaolin caused growth inhibition of 60% whereas, C_{60} at the same concentration was without effect. C_{60} and kaolin particles both increased the number of micronucleated cells. The background frequency of micronucleated cells was 0.7% to 1.0%, and this rose to 10% and 5% with 200 μg/mL of C_{60} and kaolin, respectively, the increase being statistically significant in both cases. To investigate the effects of an anti-oxidative agent on the micronucleus induction, we conducted tests with or without *N*-acetyl cysteine (NAC) using Chinese hamster ovary CHO-AA8 cells. As shown in Fig. 1, the frequency of micronucleated cells was decreased significantly in the presence of NAC. With 20 μg/mL of C_{60} and kaolin for 6 h without NAC the results were 3.8% and 8%, respectively, but in the presence of 10 mM NAC these decreased to 1.7% and 2.3%. From this observation, oxidative stress might be involved in the genotoxicity induced by nanoparticles. Furthermore, it is known that photoexcited C_{60} produces reactive oxygen species (5) and in the present experiments, the cells and C_{60} were not shielded from visible light completely. Therefore, reactive oxygen species might contribute to micronucleus-induction in C_{60}-treated cells.

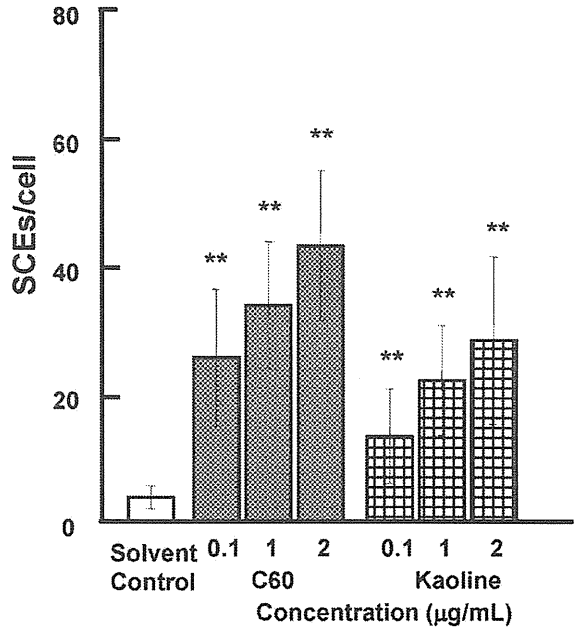

Fig. 2. Sister chromatid exchange (SCE) in CHO AA8 cells following treatment with C60 or kaolin for 1 h. The values represent the means of three experiments ± SD. Asterisks (**) indicate a significant difference ($p<0.01$) from control (treatment with 0.005% (v/v) Tween-80) cells in the Student's t-test.

On the other hand, biologically relevant features of kaolin are unclear and further studies will be required to elucidate genotoxic mechanisms.

Sister chromatid exchange (SCE) test: SCE is also used for mutagenic testing of many products. While the mechanisms responsible for SCE are not completely understood, they involve breakage of both DNA strands, followed by exchange of whole DNA duplexes. This occurs during the S phase and is efficiently induced by mutagens that form DNA adducts or that interfere with DNA replication. To investigate SCE inducing activity of nanoparticles, we examined CHO-AA8 cells following 1 h treatment with C_{60} and kaolin (Fig. 2). The SCE frequencies in cells treated with 2.0 μg/mL of C_{60} and kaolin were approximately 11 and 7 times higher than the control level, respectively ($P<0.01$ at 0.1 μg/mL or higher concentrations). C_{60} demonstrated stronger genotoxic/clastogenic potency than kaolin. Cozzi *et al.* earlier reported that H_2O_2-treatment produced reactive oxygen species and induced SCE in CHO cells, and antioxidants, such as ascorbic acid and β-carotene, reduced the frequency (6). In the present study, the results of the micronucleus test indicated involvement of reactive oxygen species so that they might contribute to SCE induction as well.

In Vivo Genotoxicity Test

Comet assay: The comet assay is known as a standard simple and sensitive technique for evaluation of

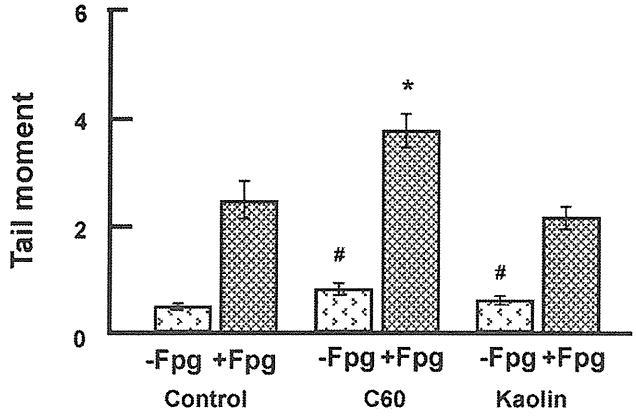

Fig. 3 DNA damage measured by comet assay in lungs of C57BL/6J mice intratracheally instilled with particles, with or without FPG treatment. Male mice were treated at a dose of 0.2 mg of particles per animal, and sacrificed 3 h after particle administration. The values represent the means of data for five animals ± SE. An asterisk (*) denotes $p < 0.01$ from that of control (+FPG) and a sharp (#) denotes $p < 0.01$ from that of control (-FPG) in a Dunnett's test after one-way ANOVA of Tail Moment.

DNA damage. The types of damage usually detected are single and double strand breaks. The pH (usually between neutral and alkaline pH) of the lysis condition can be adjusted depending upon the type of damage. Under alkaline conditions, AP sites and others where excision repair takes place are detected as DNA damage. We here evaluated DNA damage induced by particles using the comet assay under alkaline conditions. The values for DNA tail moment in the lungs with single-particle treatment at 0.2 mg/body for 3 h were measured, and DNA damage was significantly increased, around 2-fold, as compared with the vehicle control, and its intensity was C_{60} > kaolin. When we examined the effects of oxidation of purines, DNA damage was analyzed by formamidopyrimidin-glycosilase (FPG)-modified comet assay. DNA damage induced by kaolin was not changed, whereas DNA damage caused by C_{60} was elevated up to 1.7 fold compared with the vehicle control (Fig. 3). In addition, Jacobsen *et al.* also reported that C_{60} significantly increased the level of FPG sensitive sites/oxidized purines determined by the comet assay using the E1-Mutatrade markMouse lung epithelial cell line (7). From these findings, it seems that oxidative damage would be partly involved in the induction of DNA damage by C_{60}, although other changes responsible for DNA damage might be induced by kaolin.

Oxidative and lipid peroxide related DNA adduct formation: DNA adducts, formed by reactions with exogenous or endogenous agents, are known to induce gene mutations. Reactive oxygen species (ROS) are one type of endogenous agent that can produce oxidative DNA adducts such as 8-oxo-2′-deoxyguanosine (8-oxodG), a widely recognized and utilized biomarker of oxidative stress, and a major mutagenic lesion producing predominately G to T transversion mutations (8). In addition, ROS generate lipid hydroperoxides to yield heptanon-etheno (Hε)-adducts, such as HεdG, HεdA and HεdC via 4-oxo-2-nonenal (4-ONE) (9). These adducts can lead to mutations, if not repaired. We examined whether these oxidative and lipid peroxide related DNA adducts were induced in the lungs of mice by intratracheally instilled nanomaterials. 8-OxodG and three kinds of Hε-adducts were analyzed in the lungs of ICR mice 3, 24, 72 and 168 h after intratracheal instillation of 0.2 mg/body of C_{60} or kaolin, and quantified by the stable isotope dilution LC-MS/MS method described by Chou *et al.* (10). Compared with a vehicle control, DNA adduct levels were increased by about 2- to 5-fold in the lungs of mice 24 h after injection of nanoparticles (Fig. 4). The increases were time dependent until 72 h then gradually decreased within 168 h of injection (data not shown). Related to this, oxidative DNA damage was induced by intratracheal instillation of C_{60} or kaolin in the comet assay with FPG treatment, as described above. In addition, Folkmann *et al.* reported that oral gavage of C_{60} increased the levels of 8-oxodG in the liver and the lungs of F344 rats (11). Moreover, Tsurudome *et al.* described increased 8-oxodG levels induced by intratracheally instilled diesel exhaust particles in the lungs of F344 rats, and 8-oxoguanine DNA glycosylase 1 (OGG1) mRNA was also overexpressed (12). The decreased DNA adducts in the present study at 168 h may have been a result of a repair enzyme such as OGG1. This is the first observation that Hε-lipid peroxide related DNA adducts are increased by nanoparticles. Such adducts could clearly contribute to nanomaterial-induced DNA damage and mutation. Our findings suggest involvement of ROS generation, although differences between C_{60} and kaolin still require clarification.

***gpt* Mutations in the lungs of *gpt* transgenic mice:** Transgenic *gpt* delta mice are a useful model system for detecting both point mutations and large deletions (< 10 kb) (13). λEG10 transgenes carrying gpt (detection of point mutations) and *red*, *gam* (detection of deletion) genes have been integrated into mouse chromosome 17, and point mutations and deletions observed in any tissues can be detected as 6-thioguanine (6-TG) resistant colonies and Spi^- plaques, respectively. To examine *in vivo* mutagenicity of nanoparticles, *gpt* delta transgenic mice were exposed to C_{60} and kaolin at four different doses by intratracheal instillation, and *gpt* mutations were analyzed. The background *gpt* mutant frequency (MF) in lungs was $10.3 \pm 0.53 \times 10^{-6}$. MFs were significantly increased by 2 to 3-fold to $30.75 \pm 3.32 \times 10^{-6}$ ($p = 0.019$) for C_{60} and $19.30 \pm 4.82 \times 10^{-6}$ ($p = 0.002$) for kaolin (4).

Moreover, we examined the mutational characteris-

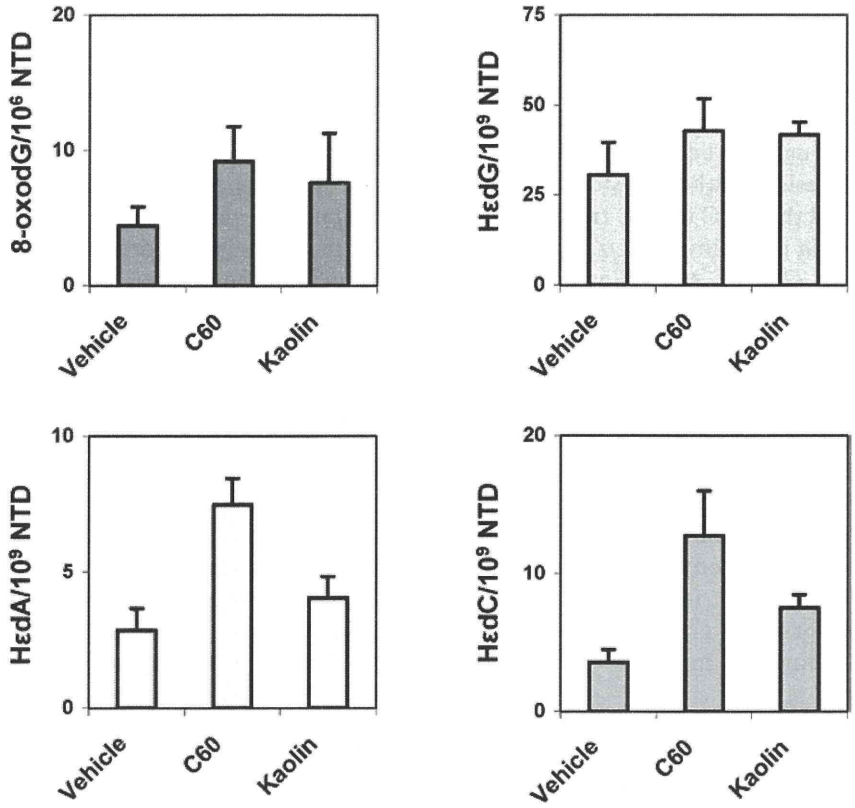

Fig. 4. Oxidative and lipid peroxide related DNA adduct formation in the lungs of ICR mice induced by nanoparticle exposure. DNA was extracted from lungs of mice 24 h after intratracheal instillation of 0.2 mg/body of C60 or kaolin, and digested enzymatically. Control animals were exposed to saline containing 0.05% Tween80. The 8-oxodG and 3 kinds of Hε-adducts were quantified by the stable isotope dilution LC-MS/MS method described by Chou *et al.* (10).

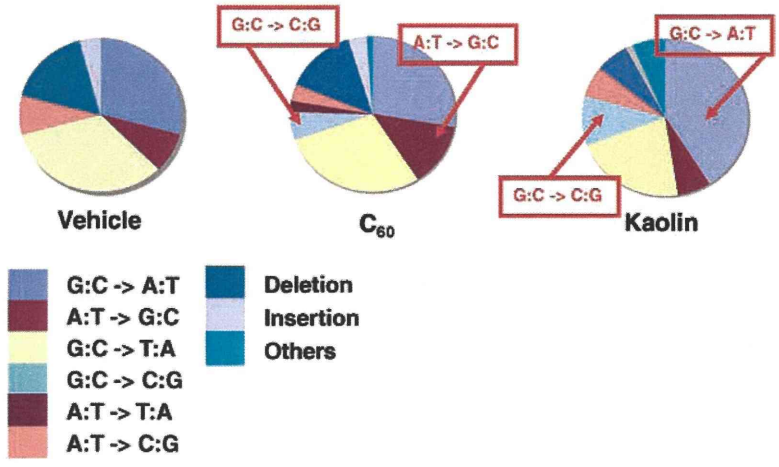

Fig. 5. Classification of *gpt* mutations from the lungs of control and nanoparticle treated mice.

tics induced by particles by PCR and DNA sequencing analysis of 6-TG resistant mutants. Classes of mutations found in the *gpt* gene are shown in Fig. 5. Interestingly, G:C to C:G transversions were increased in common with both particle treatments. Since these mutations were commonly increased regardless of the constituents (i.e., C_{60} is graphite and kaolin is aluminum silicate), the mechanisms might be the same. It has been reported that various oxidative stresses caused by sunlight, UV radiation, hydrogen peroxide and peroxy radicals frequently induce G:C to C:G transversions in various *in vitro* assay systems (14–17). Moreover, a variety of ox-

idative lesion products of guanine other than 8-oxodG, including imidazolone (Iz), oxazolone (Oz), spiroiminodihydantoin (Sp) and guanidinohydantoin (Gh), have been reported (18–24). Three such molecules, Oz, Sp and Gh are now thought to be key causes G to C transversions with translesion synthesis systems (22–25). Therefore, it is suggested that G:C to C:G transversions induced by C_{60} and kaolin could involve Oz, Sp and Gh formation. In addition, G:C to A:T transitions were also significantly increased by instillation of kaolin but not C_{60}. In general, G to A (or C to T) transitions have commonly been observed in spontaneous and chemically-induced mutants, and deamination of guanine or 5-methylcytosine might be involved. Burney *et al.* reported that nitric oxide induces DNA damage. NO· can form N_2O_3, and direct by this agent can lead to DNA deamination via diazonium ion formation (26). Moreover, nitric oxide is produced by activated macrophages in inflamed organs. In fact, test substance-phagocytized macrophages and granulomas were frequently observed in the lungs of mice (4).

Immunohistochemical Analysis of Inflammation Factors

In order to confirm enhancement of nitric oxide production by C_{60} and kaolin, we examined immunohistochemical staining of an inflammation factor, nitrotyrosine (NT), in the lungs of *gpt* delta mice treated with these nanoparticles using the same procedure reported previously (27) with minor modification. As shown in Fig. 6, the pattern of NT staining corresponded to the areas of inflammation within lung parenchyma. In the case of C_{60} exposure, many regions of the lungs stained positively (data not shown), and intense NT staining was localized in test substance-phagocytized macrophages and granulomas. Similarly, staining with NT antibodies was observed in macrophages and alveolar epithelial cells in the lungs of mice exposed to kaolin, although to a lesser extent as compared with C_{60}.

Conclusion

Our results clearly demonstrated that both *in vitro* and *in vivo* genotoxicity are induced by C_{60} and kaolin. However, the mechanisms have yet to be fully clarified, and oxidative stress might be at least partly involved. There are a number of ways in which reactive oxygen species (ROS) could be generated: i) nanoparticles might trigger ROS production by iron-catalysed Fenton reactions; ii) nanoparticles could accumulate in cells due to phagocytosis, then enhance the production of ROS by NADPH oxidase (28,29). Recently, innate immune activation through Nalp3 inflammasomes has been suggested to play an important role in pulmonary fibrotic disorders of silicosis and asbestosis (30,31). It has been reported that proinflammatory cytokines, such as interleukin 1β are key molecules for pneumoconiosis. At

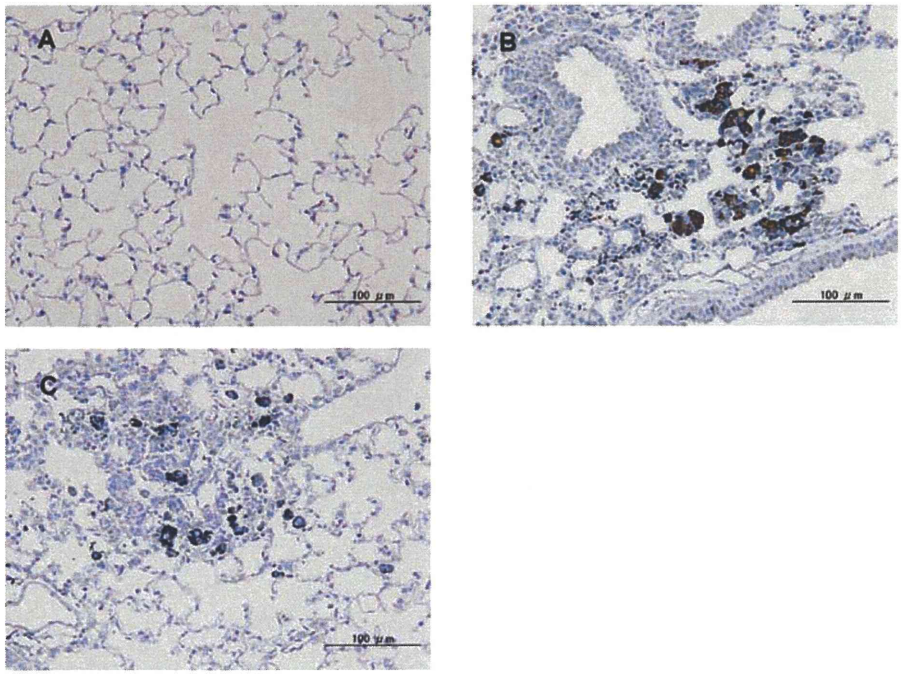

Fig. 6. Immunohistochemical localization of nitrotyrosine (NT). Since C_{60} is brown in color, we used an SG substrate kit (Vector Laboratories, USA) for peroxidase, with positive cells stained dark blue-gray. A: alveolar region in a control mouse, with no significant staining for NT. B: alveolar region in a mouse exposed to C_{60}, with positive macrophages phagocytizing test substance and epithelial cells. The brown colored material is C_{60}. C: alveolar region in a mouse exposed to kaolin. Note intense staining for NT in the granulomatous region.

present, no data are available for activation of the Nalp3 inflammasome pathway by C_{60} and kaolin. However, it is likely that both nanoparticles can activate in the same way as asbestos and silica, because oxidative stress was increased in the lungs of treated mice. Further studies of the mechanisms of genotoxicity are needed.

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Low-dose carcinogenicity of 2-amino-3-methylimidazo[4,5-*f*]quinoline in rats: Evidence for the existence of no-effect levels and a mechanism involving p21$^{Cip/WAF1}$

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The carcinogenicity of the low amounts of genotoxic carcinogens present in food is of pressing concern. The purpose of the present study was to determine the carcinogenicity of low doses of the dietary genotoxic carcinogen 2-amino-3-methylimidazo[4,5-*f*]quinoline (IQ) and to investigate mechanisms by which IQ exerts its carcinogenic effects. A total of 1595 male F344 rats were divided into seven groups and administered with IQ at doses of 0, 0.001, 0.01, 0.1, 1, 10 and 100 p.p.m. in the diet for 16 weeks. We found that IQ doses of 1 p.p.m. and below did not induce preneoplastic lesions in either the liver or the colon, while IQ doses of 10 and 100 p.p.m. induced preneoplastic lesions in both of these organs. These results demonstrate the presence of no-effect levels of IQ for both liver and colon carcinogenicity in rats. The finding that p21$^{Cip/WAF1}$ was significantly induced in the liver at doses well below those required for IQ mediated carcinogenic effects suggests that induction of p21$^{Cip/WAF1}$ is one of the mechanisms responsible for the observed no-effect of low doses of IQ. Furthermore, IQ administration caused significant induction of CYP1A2 at doses of 0.01–10 p.p.m., but administration of 100 p.p.m. IQ induced CYP1A1 rather than CYP1A2. This result indicates the importance of dosage when interpreting data on the carcinogenicity and metabolic activation of IQ. Overall, our results suggest the existence of no-effect levels for the carcinogenicity of this genotoxic compound. (*Cancer Sci* 2011; 102: 88–94)

Exposure to environmental carcinogens is one of the most significant causes of human cancers. Determination of the dose-response relationship between carcinogen exposure and induction of cancer is one of the most important areas of chemical risk assessment. Of particularly high priority is the cancer risk assessment of dietary carcinogens.

Heterocyclic amines (HCA) are well known dietary genotoxic carcinogens derived from cooked protein-rich foods such as meat and fish,[1–3] and the carcinogenicities of 2-amino-3,8-dimethylimidazo[4,5-*f*]quinoxaline (MeIQx), 2-amino-1-methyl-6-phenylimidazo[4,5-*b*]pyridine (PhIP) and 2-amino-3-methylimidazo[4,5-*f*]quinoline (IQ) have been widely investigated in various animal models. MeIQx induces cancers of the liver, zymbal gland, skin and clitoral gland in rats,[4] and caners of the liver and lung, and lymphoma and leukemia in mice.[5] PhIP induces colon cancers and mammary gland cancers in rats,[6] and lymphomas in mice.[7] IQ induces cancers of the liver, colon, mammary and zymbal glands in rats, caners of the liver, lung and forestomach in mice, and cancer of the liver in non-human primates.[8–10] MeIQx and PhIP are classified as category 2B compounds (possibly carcinogenic to humans) and IQ is classified as a category 2A compound (probably carcinogenic to humans) by the International Agency for Research on Cancer.[11] Therefore, although the concentrations of HCA in food are low, they constitute a potential hazard, and there is concern regarding the carcinogenic effects of low doses of these HCA.

Based on the view that even minute doses of a genotoxic carcinogen has the potential to produce irreversible deleterious genetic changes in the DNA of a target organ cell and the argument that if sufficient numbers of test animals are used the carcinogenic effect of a minute dose can be demonstrated, it is generally assumed that genotoxic carcinogens exert a non-threshold carcinogenic effect. However, the carcinogenicities of most genotoxic carcinogens are determined by experimental animal carcinogenicity studies using doses that are generally orders of magnitude higher than actual human exposure levels and the dose-response curves obtained are then extrapolated to zero using a non-threshold mathematical model. This approach, however, is being challenged as advancements in the understanding of the molecular mechanisms of carcinogenesis are being made and experimental evidence showing that genotoxic carcinogens do not exert mutagenic and carcinogenic effects at low doses accumulates.[12–19]

Previously, we demonstrated the existence of no-effect levels of MeIQx for both hepatocarcinogenicity and *in vivo* mutagenicity in various carcinogenesis models in different rat strains.[17,20–22] It has also been shown that low doses of PhIP do not exert either initiation or promotion activities in colon carcinogenesis in the rat.[23,24] However, little is known about the carcinogenic potential of low doses of IQ.

In addition, little is known about the mechanisms underlying the carcinogenicities of lower doses of HCA, but incorporation of mechanistic information is critical for quantitative cancer risk assessment. The purpose of the present study is to determine the relationship between administration of low doses of IQ and induction of preneoplastic lesions in the liver and colon in rats, and to investigate carcinogenic mechanisms of action of various doses of IQ by evaluating DNA-adduct formation, oxidative DNA damage and expression levels of genes involved in metabolic activation of IQ, cell proliferation and DNA damage repair in the liver.

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Materials and Methods

Chemical and diets. IQ was purchased from Nard Institute Ltd (Osaka, Japan) with a purity of 99.9%. Basal diets (powdered MF; Oriental Yeast Co., Tokyo, Japan) and the diets containing IQ were prepared once a month by Oriental Yeast Co.

Animals. A total of 1595 male F344 rats were supplied by Charles River Japan, Inc. (Hino, Shiga, Japan) and were used at 21 days of age. Animals were housed in polycarbonate cages (five per cage) in experimental animal rooms with a targeted temperature of 22 ± 3°C, relative humidity of 55 ± 5% and a 12-h light/dark cycle. Diet and tap water were available *ad libitum* throughout the study.

Experimental design. The animal experiment protocols were approved by the Institutional Animal Care and Use Committee of Osaka City University Medical School. Rats were randomized into seven groups, 245 rats in each of groups 1–6 and 125 rats in group 7. Since the levels of IQ in cooked foods are lower than those of MeIQx and PhIP,[11] IQ dosage and treatment duration in this study were the same as the previous low dose carcinogenicity studies with MeIQx and PhIP.[18,24] Animals were fed diets containing IQ as follows: 0 (group 1, control), 0.001 (group 2), 0.01 (group 3), 0.1 (group 4), 1 (group 5), 10 (group 6) and 100 p.p.m. (group 7) for 16 weeks. Fresh diet was supplied to the animals twice weekly. Bodyweights, food consumption and water intake were measured weekly.

Five rats in each group were killed at week 4 under ether anesthesia. At death, livers were snap frozen in liquid nitrogen and stored at −80°C for examination of IQ-DNA adducts and 8-hydroxy-2′-deoxyguanosine (8-OHdG) formation in the DNA. The remaining rats were killed at the end of week 16 under ether anesthesia for examination of the development of glutathione *S*-transferase placental form (GST-P) positive foci, which is a well-established preneoplastic lesion in the rat liver,[25,26] and aberrant crypt foci (ACF), which is a surrogate marker for preneoplastic lesions in the rat colon.[24,27,28] At death, livers were excised, weighed and then three slices each from the left lateral, medial and right lateral lobes were cut and placed in 10% phosphate-buffered formalin. The remaining liver tissues were snap frozen in liquid nitrogen and stored at −80°C for mRNA expression analysis. Following fixation, liver tissues were embedded in paraffin and processed for histopathological examination.

Examination of GST-P positive foci in the liver. Anti-rat GST-P polyclonal antibody (Medical and Biological Laboratories Co., Ltd, Nagoya, Japan) at a dilution of 1:1000 was used for immunohistochemical staining of GST-P. The GST-P-positive hepatocellular foci composed of two or more cells were counted under a light microscope.[17,18,20,22] Total areas of livers were measured using a color image processor IPAP (Sumica Technos, Osaka, Japan) and the number of GST-P-positive foci per square centimeter of liver tissue was calculated.

IQ-DNA adduct and 8-OHdG formation in livers. IQ-DNA adducts were measured by the ^{32}P-postlabeling method as described previously.[29,30] Levels of 8-OHdG formation in liver DNA were determined by high-performance liquid chromatography with electrochemical detection as previously described.[31]

TaqMan real-time quantitative PCR. The mRNA expression levels of genes involved in IQ metabolism (CYP1A1, CYP1A2 and CYP1B1), DNA damage repair (8-oxoguanine DNA glycosylase [Ogg1], growth arrest and DNA damage-inducible protein 45 [GADD45], AP endonuclease-1 [APE-1], MSH2 and MSH3) and cell cycle regulation (p53 and p21$^{Cip/WAF1}$ and proliferating cell nuclear antigen [PCNA]) were evaluated in the livers by TaqMan real-time quantitative PCR as described previously.[31] Sequence-specific primers and probes (Taqman Gene Expression Assay) were purchased from Applied Biosystems, Inc., Carlsbad, CA, USA. Beta-2-microglobulin (B2M) was used as an internal control.

Examination of ACF in colon. Formation of ACF was examined as described previously.[24] Although ACF consisting of four or more crypts are considered to be better predictors of colon tumor outcome in rats,[32] to ensure that all doses of IQ that have the potential to induce colon carcinogenesis were accounted for, doses of IQ that caused an increase of any size of ACF were considered to have the potential to induce colon carcinogenesis in the present study.[24]

Statistical analysis. All mean values are reported as mean ± SD. Statistical analyses were performed using the Statlight program (Yukms Co., Ltd, Tokyo, Japan). Homogeneity of variance was tested by the Bartlett test. Differences in mean values between the control and IQ-treated groups were evaluated by the 2-tailed Dunnett test when variance was homogeneous and the 2-tailed Steel test when variance was heterogeneous.[22,31] P values <0.05 were considered significant.

Results

General observation. All animals survived to the end of study without any apparent abnormal pathological features. The final average body and liver weights and IQ intake are summarized in Table 1. The final bodyweight of the 100 p.p.m. group was significantly lower than that of the 0 p.p.m. group. Absolute and relative liver weights were significantly decreased in the 0.1 and 1 p.p.m. groups and were significantly increased in the 100 p.p.m. group compared with the 0 p.p.m. group. There were no significant differences in either food or water consumption among groups (data not shown). The intake of IQ was proportional to the administered doses (Table 1). No tumors were found in any organs including the liver and colon in any of the groups.

Induction of GST-P-positive foci in the livers. No histopathological changes were observed in any of the IQ-treated groups.

Table 1. Body and organ weights, and IQ intake

Group	IQ (p.p.m.)	No. rats	Bodyweight (g)	Liver		Average IQ intake	
				Absolute weight (g)	Relative weight (%)	Daily intake (mg/kg b.w.)	Total (mg/kg b.w.)
1	0	240	331 ± 23	9.3 ± 1.7	2.8 ± 0.4	0	
2	0.001	240	332 ± 17	9.1 ± 1.4	2.8 ± 0.4	0.0001	0.008
3	0.01	240	331 ± 19	9.0 ± 1.5	2.8 ± 0.4	0.0007	0.08
4	0.1	240	331 ± 22	8.5 ± 1.2*	2.6 ± 0.3*	0.008	0.9
5	1	240	331 ± 17	8.5 ± 1.2*	2.6 ± 0.3*	0.08	8.7
6	10	240	330 ± 18	9.0 ± 1.3	2.7 ± 0.4	0.76	85.1
7	100	120	319 ± 19*	10.0 ± 1.6*	3.2 ± 0.4*	7.83	877.5

*Significantly different from group 1. IQ, 2-amino-3-methylimidazo[4,5-*f*]quinoline.

The number and size of GST-P-positive foci in rat livers at week 16 is summarized in Table 2. The total numbers of GST-P-positive foci per unit area in the livers in the groups administered 0.001–1 p.p.m. IQ did not differ from the control value (0 p.p.m. group), and no significant increases were observed in any size range of GST-P-positive foci in these groups. Significant increases in the total numbers of GST-P-positive foci per unit area in the liver were observed in the 10 and 100 p.p.m. groups compared with the control. The numbers of GST-P-positive foci composed of 2–4 cells and 5–10 cells in the 10 p.p.m. group and GST-P-positive foci of all sizes in the 100 p.p.m. group were significantly increased.

Formation of IQ-DNA adduct and 8-OHdG in liver DNA. Representative autoradiograms of IQ-DNA adducts in livers are shown in Figure 1. The levels of IQ-DNA adducts in the livers of the 0 and 0.001 p.p.m. IQ-treated groups were under the detectable limit at week 4 (Table 3). IQ-DNA adducts were detectable in the livers of rats administered 0.01 p.p.m. IQ, and adduct formation increased in a dose-dependent manner in groups administered higher doses of IQ. No significant differences in 8-OHdG levels were observed in the liver DNA between any of the groups administered IQ and the control group (Table 3).

Gene expression changes in the liver. Relative mRNA expression of IQ metabolizing genes CYP1A 1and CYP1A2, cell cycle genes PCNA and $p21^{Cip/WAF1}$, p53, and DNA repair genes APE-1 and GADD45 in the livers at week 16 is shown in Figure 2. CYP1A1 was significantly increased in the livers of rats treated with 100 p.p.m. IQ, but not in the lower doses of IQ. CYP1A2, on the other hand, was significantly increased in the 0.01–10 p.p.m. groups, but no significant change was observed in the 100 p.p.m. group. There was no significant difference in the CYP1B1 expression level among groups (data not shown).

A significant increase in PCNA was observed in the 100 p.p.m. group, but not in the groups administered lower doses of IQ, while the negative cell cycle regulator $p21^{Cip/WAF1}$ was significantly induced in the 0.01 p.p.m. group and maximally induced in the 100 p.p.m. group. The expression level of $p21^{Cip/WAF1}$ in the 100 p.p.m. group was significantly higher than in the 10 p.p.m. and lower dose groups. There were no significant changes in p53 expression levels in the IQ-treated groups.

APE-1 was significantly induced in the 10 and 100 p.p.m. groups and GADD45 was significantly induced in the 100 p.p.m. group. IQ had no effect on the expression of Ogg-1, MSH2 or MSH3 (data not shown).

Induction of ACF in the colon. The number and size of ACF in rat colons at week 16 is summarized in Table 4. In the 10 p.p.m. group, the number of ACF composed of one crypt was significantly increased compared with the control. In the 100 p.p.m. group, significant increases were observed in the numbers of all sizes of ACF. In contrast, in the groups administered 0.001–1 p.p.m. IQ, neither the number of any size ACF nor the total number of ACF differed from the control.

Discussion

Dose-response relationships for genotoxic carcinogens have been a topic of intense scientific and public debate. High doses of the genotoxic dietary carcinogen IQ have been demonstrated to induce liver and colon cancers in rats (300 p.p.m. in diet)[8] and liver cancers in nonhuman primates (10 mg/kg b.w./day).[10] However, as the concentrations of IQ in food are generally extremely low,[11] there is uncertainty regarding the carcinogenicity of the doses of IQ to which humans are exposed. The present study shows that IQ at doses of 1 p.p.m. (0.08 mg/kg body weight [b.w.]/day) and lower did not induce either GST-P-positive foci in the liver or ACF in the colon. Only in the groups administered higher doses of IQ, 10 p.p.m. (0.76 mg/kg b.w./day) and 100 p.p.m. (7.83 mg/kg b.w./day), were increases in GST-P-positive foci and ACF observed.

GST-P-positive foci and ACF are well-established preneoplastic lesions of the liver and colon, respectively, in rats. These lesions have been accepted as useful end-point markers in the assessment of carcinogenic effects of environmentally relevant concentrations of carcinogens as they can extend the range of observable effect levels.[24,26] Therefore, the results of the present study suggest the presence of no-effect levels of IQ for both liver and colon carcinogenicity in rats and indicate that the dose-response relationship for carcinogenicity of low dose IQ is nonlinear.

Several threshold mechanisms for genotoxic carcinogens have been suggested, including induction of detoxification processes, cell cycle delay, DNA repair, apoptosis and the suppression of neoplastically transformed cells by the immune system.[12,13,15,33] However, little *in vivo* evidence is available. To explore mechanisms underlying the carcinogenicity of low doses of IQ, we examined the relative mRNA expression of a panel of genes involved in cell proliferation, cell cycle regulation, DNA repair and IQ metabolic activation. We found that the cell proliferation marker PCNA was significantly increased only at a dose of 100 p.p.m., a dose that is carcinogenic. The cell cycle negative regulator $p21^{Cip/WAF1}$, on the other hand, was significantly induced at a dose of 0.01 p.p.m., a dose well below that which induced the formation of preneoplastic lesions. Furthermore, the finding that the levels of $p21^{Cip/WAF1}$ in the groups administered 10 p.p.m. and less were much lower than that of the group administered 100 p.p.m. implies that hepatocytes have adequate capacity to cope with the type of damage that is repaired by the $p21^{Cip/WAF1}$ pathway when exposed to low doses of IQ, but that the repair capacity of these hepatocytes, even in the presence of high $p21^{Cip/WAF1}$ expression, can be overwhelmed when the cell is subjected to very high doses of IQ. It is reasonable to suggest

Table 2. Development of GST-P-positive foci in the livers of rats administered IQ for 16 weeks

Group	IQ (p.p.m.)	No. rats	Size of GST-P positive foci				
			2–4	5–10	11–20	≥21	Total
1	0	240	0.09 ± 0.25	0.03 ± 0.11	0.02 ± 0.11	0.00 ± 0.02	0.15 ± 0.31
2	0.001	240	0.10 ± 0.24	0.04 ± 0.15	0.01 ± 0.07	0	0.16 ± 0.31
3	0.01	240	0.15 ± 0.47	0.07 ± 0.41	0.02 ± 0.22	0.02 ± 0.03	0.26 ± 1.30
4	0.1	240	0.10 ± 0.28	0.04 ± 0.15	0.01 ± 0.07	0.01 ± 0.08	0.15 ± 0.35
5	1	240	0.10 ± 0.25	0.04 ± 0.16	0.01 ± 0.06	0	0.14 ± 0.33
6	10	240	0.51 ± 0.65	0.19 ± 0.36*	0.02 ± 0.10	0.01 ± 0.11	0.74 ± 0.88*
7	100	120	26.23 ± 18.24*	23.81 ± 16.23*	19.25 ± 11.70*	18.74 ± 11.81*	88.03 ± 50.41*

*Significantly different from group 1. GST-P, glutathione *S*-transferase placental form positive foci; IQ, 2-amino-3-methylimidazo[4,5-*f*]quinoline.

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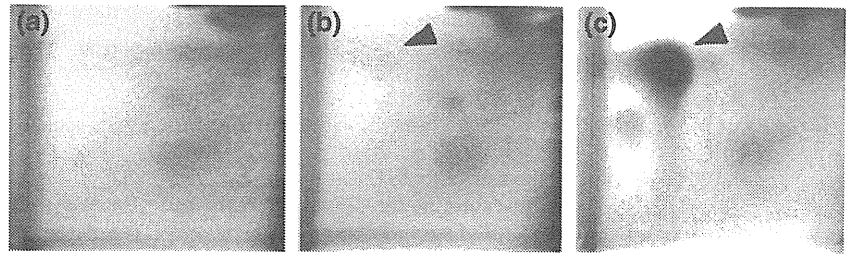

Fig. 1. Autoradiograms of 2-amino-3-methylimidazo-[4,5-*f*]quinoline (IQ)-DNA adducts the livers of 0 (a), 0.001 (b) and 100 (c) p.p.m. IQ-treated groups at week 4. Arrowheads indicate IQ-DNA adduct. The imaging plates were exposed for 3 h (a) and 24 h (b and c).

Table 3. IQ-DNA adduct and 8-OHdG formation in liver DNA

Group	IQ (p.p.m.)	No. rats	Adduct level ($\times 10^{-7}$ ntd)	8-OHdG ($\times 10^{-5}$ dG)
1	0	5	UDL	0.23 ± 0.07
2	0.001	5	UDL	0.25 ± 0.05
3	0.01	5	0.045 ± 0.02	0.24 ± 0.07
4	0.1	5	0.1 ± 0.004	0.32 ± 0.10
5	1	5	1.7 ± 0.07	0.24 ± 0.08
6	10	5	12.7 ± 0.07	0.22 ± 0.07
7	100	5	107.0 ± 0.07	0.23 ± 0.08

IQ, 2-amino-3-methylimidazo[4,5-*f*]quinoline; ntd, nucleotide; 8-OHdG: 8-hydroxy-2′-deoxyguanosine; UDL, under the detectable limit.

that suppression of cell cycle progression by $p21^{Cip/WAF1}$ followed by DNA repair is at least one of the mechanisms responsible for the observed no-effect of low doses of IQ in rats in the present model.

It is known that the vast majority of DNA damage is repaired by base excision repair (BER), nucleotide excision repair (NER) and mismatch repair (MMR).[34] APE-1 plays an essential role in the BER repair process by cleaving the phosphodiester backbone.[35] The activities of two different heterodimeric complexes, MSH2-MSH3 and MSH2-MSH6, belonging to the MMR system are mainly responsible for the post-replicative repair of mismatches.[36] We found that IQ significantly increased the expression levels of APE-1 but not MSH2 and MSH3 at doses of 10 and 100 p.p.m. in the liver. It has also been reported that IQ has no effect on expression of ERCC1, which is a key molecule in the NER process.[37] These findings suggest that BER rather than MMR or NER responds to IQ-induced DNA damage.

GADD45 is involved in a variety of growth regulatory mechanisms, including DNA repair, growth arrest and apoptosis.[38] It is induced by genotoxic and certain other cell stresses by p53-dependent and independent pathways.[39,40] GADD45 expression was significantly induced in the 100 p.p.m. group. The fact that significant induction of APE-1 and GADD45 was observed only at the highest doses of 10 and/or 100 p.p.m. indicate the IQ-induced DNA damage response is dose-dependent. Moreover, the fact that in the groups with low doses expression of APE-1 and GADD45 were not affected and that there was a significant but moderate induction of $p21^{Cip/WAF1}$ imply that normal physiological levels of these genes are sufficient to repair the DNA damage caused by low doses of IQ. However, the expression levels of these genes are all increased by higher carcinogenic doses of IQ. A reasonable explanation of the no-effect of low doses of IQ and the carcinogenicity of high doses of IQ is that carcinogenicity is the consequence of a disruption in the balance between DNA damage and repair and between abnormal cell proliferation and apoptosis or cell cycle regulation.

Our results show that p53 gene expression is not induced by administration of IQ. Furthermore, p53-deficient mice do not show higher susceptibility to IQ-induced liver carcinogenesis than wild type mice.[41] These results suggest that p53 does not have a significant impact on the carcinogenicity of IQ.

DNA adduct formation by metabolic activation of IQ is believed to play an important role in the carcinogenicity of IQ.[42] Formation of IQ-DNA adducts in the liver showed a linear dose-dependency and proved to be one of the most sensitive end-points for the detection of exposure to IQ. Adduct formation was detectable in groups administered far lower doses of IQ compared with detection of other end-points such as cell proliferation and preneoplastic lesion induction. That IQ-DNA adduct formation was not detected in the 0.001 p.p.m. group was most likely due to the detection limit of the assay. It should be noted that DNA adduct is a premutagenic lesion and not necessarily correlated to the frequencies of mutation and cancer induced by genotoxic compounds. For example, it is known that IQ forms DNA adducts in the kidneys and stomach of both rats and monkeys, but does not induce tumors in these organs.[43,44] Our present findings of a linear dose-response of IQ-DNA adduct formation and a nonlinear carcinogenic dose-response to IQ administration support the idea that IQ-DNA adducts do not necessarily lead to mutation and formation of cancerous lesions. Our results are also in line with previous results on HCA including MeIQx[1,18,45] and PhIP.[24] These results can be explained, at least in part, by the actions of gene products such as $p21^{Cip/WAF1}$, GADD45 and APE-1 and the other repair genes for DNA damage. Moreover, in the case of MeIQx, it has been suggested that formation of DNA adducts alone might not be sufficient to produce cancers and that the MeIQx-induced genetic alterations in the liver are enhanced by liver regeneration induced by high doses of MeIQx itself.[1] Therefore, while IQ-DNA adduct formation is important in IQ carcinogenicity, high levels of adduct formation are likely required and other factors such as cell proliferation can affect the balance between DNA damage and repair and lead to fixation of DNA mutations into the cell's genome.

It has been demonstrated *in vitro* that IQ is more efficiently metabolized and activated by CYP1A2 than by CYP1A1 or CYP1B1.[46] However, limited *in vivo* data are available. In a study by McPherson *et al.*[47], no significant induction in mRNA expression level or activity of either CYP1A1 or CYP1A2 were reported in the livers of rats receiving 300 p.p.m. IQ in the diet for 52 weeks, but these enzymes were significantly increased after daily administration of 20 mg/kg b.w. IQ by oral gavage for 3 days; in the average adult rat, a dose of 300 p.p.m. IQ in the diet is approximately equivalent to administration of 20 mg/kg b.w. IQ by oral gavage. The results of the present study revealed that IQ significantly induced CYP1A2 expression at doses from 0.01 to 10 p.p.m., but CYP1A2 was not induced in the 100 p.p.m. group. The lack of effect of 100 p.p.m. IQ on CYP1A2 expression is consistent with the results in rats receiving 300 p.p.m. IQ in the diet for 52 weeks.[47] Significant increases in CYP1A1 expression in the 100 p.p.m. group provide an alternative mechanism that can compensate for decreased CYP1A2 activity. However, as noted above, in apparent contrast to our results, in the study by McPherson *et al.*,[47] administration of 300 p.p.m. IQ over the course of 52 weeks did

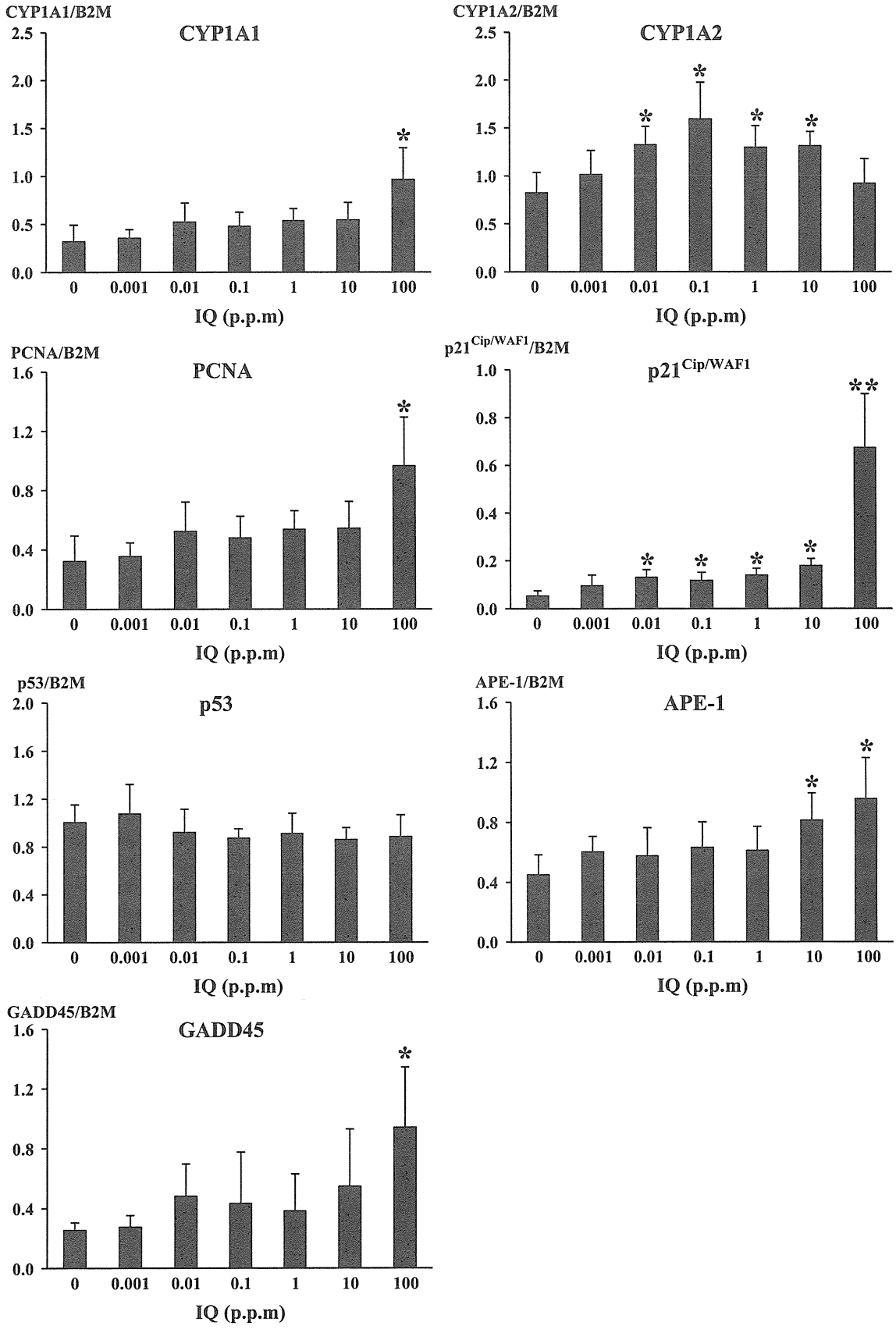

Fig. 2. Relative mRNA expression in the livers of rats at week 16. *Significantly different from 0 p.p.m. **Significantly different from all other groups. APE-1, AP endonuclease-1; B2M, beta-2-microglobulin; GADD45, growth arrest and DNA damage-inducible protein 45; PCNA, proliferating cell nuclear antigen.

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Table 4. Development of ACF in the colons of rats administered IQ for 16 weeks

Group	IQ (p.p.m.)	No. rats	Size of ACF				
			1	2	3	≥4	Total
1	0	240	0.08 ± 0.28	0.12 ± 0.32	0.06 ± 0.25	0.08 ± 0.29	0.33 ± 0.64
2	0.001	240	0.12 ± 0.36	0.08 ± 0.29	0.10 ± 0.32	0.09 ± 0.30	0.39 ± 0.69
3	0.01	240	0.15 ± 0.41	0.15 ± 0.42	0.06 ± 0.24	0.06 ± 0.24	0.43 ± 0.77
4	0.1	240	0.11 ± 0.33	0.11 ± 0.35	0.06 ± 0.25	0.08 ± 0.27	0.36 ± 0.63
5	1	240	0.15 ± 0.45	0.10 ± 0.30	0.10 ± 0.33	0.05 ± 0.23	0.41 ± 0.80
6	10	240	0.19 ± 0.48*	0.16 ± 0.41	0.07 ± 0.25	0.09 ± 0.40	0.50 ± 0.86
7	100	120	1.48 ± 1.46*	1.29 ± 1.51*	0.70 ± 0.93*	0.72 ± 1.01*	4.19 ± 3.34*

*Significantly different from group 1. ACF, aberrant crypt foci; IQ, 2-amino-3-methylimidazo[4,5-*f*]quinoline.

not induce CYP1A1. Therefore, it is reasonable to postulate that the dose-relationship between IQ and induction of CYP1A1 is not a simple dose-response. CYP1B1 does not appear to be involved in the metabolism of IQ at doses up to 100 p.p.m. in rats. The findings described above demonstrate the importance of taking into account dosage, duration and route of exposure in interpretation of the data on metabolic activation of IQ. Further studies on the dose-response relationships between chronic IQ exposure and the protein expression levels and activities of detoxifying enzymes, especially at doses relevant to human exposure, would provide further insight into the role of metabolic activation in IQ carcinogenicity.

Oxidative DNA damage does not appear to play a role in IQ-induced carcinogenesis. In the present study, no significant changes in 8-OHdG levels or Ogg1 expression levels in the livers of IQ-treated rats were observed. Our results are consistent with the recent findings in IQ-treated Big Blue rats that oxidative stress was not responsible for the initiation of IQ-induced carcinogenesis in the liver and colon.[37] In this respect, IQ is different from MeIQx, in which oxidative DNA damage plays an important role in liver carcinogenesis.[48]

In summary, the present study provides the first experimental data on the carcinogenicity of low doses of IQ in both the liver and colon of the test animal and compares the effect of IQ at the cellular level with its carcinogenic effect. Our findings support the idea that there is a practical threshold that should be considered when evaluating the risk of genotoxic carcinogens. To this end, further accumulation of data, especially mechanistic data, should be promoted to facilitate not only an understanding of the carcinogenic effects of low doses of genotoxic carcinogens but also to establish an accurate means of quantitative risk assessment.

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Disclosure Statement

The authors have no conflict of interest.

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—Review—

ナノマテリアルの慢性影響研究の重要性

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Importance of Researches on Chronic Effects by Manufactured Nanomaterials

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Manufactured nanomaterials are the most important substances for the nanotechnology. The nanomaterials possess different physico-chemical properties from bulk materials. The new properties may lead to biologically beneficial effects and/or adverse effects. However, there are no standardized evaluation methods at present. Some domestic research projects and international OECD programs are ongoing, in order to share the health impact information of nanomaterails or to standardize the evaluation methods. From 2005, our institutes have been conducting the research on the establishment of health risk assessment methodology of manufactured nanomaterials. In the course of the research project, we revealed that the nanomaterials were competent to cause chronic effects, by analyzing the intraperitoneal administration studies and carcinogenic promotion studies. These studies suggested that even aggregated nanomaterials were crumbled into nano-sized particles inside the body during the long-term, and the particles were transferred to other organs. Also investigations of the toxicokinetic properties of nanomaterials after exposure are important to predict the chronically targeted tissues. The long lasting particles/fibers in the particular tissues may cause chronic adverse effects. Therefore, focusing on the toxicological characterization of chronic effects was considered to be most appropriate approach for establishing the risk assessment methods of nanomaterials.

Key words——chronic toxicity; multi-wall carbon nanotube (MWCNT); fullerene

1. はじめに

近年，ナノテクノロジーの中心的な役割を担う物質としての産業用ナノマテリアルは，急速にその種類や生産量が増加しつつあるところであるが，新たに期待されているナノマテリアルの物理化学特性については，有効的な生理活性等に使用され得る特性を持つ反面，ヒト健康影響に対する懸念についても検証されるべきであると考えられている．つまり，ナノマテリアルを用いた技術や製品を社会的に受容するためには，安全性の検証を行うことが不可欠であると思われる．しかし，従来の一般的な化学物質とは異なる物理化学的特性は，その毒性評価においても従来とは異なる考え方を取り入れることも必要とされている．それゆえ，ナノマテリアルの特性を考慮した有害性評価手法の開発が急務となっている．また，国際的な枠組みにおいても，ナノマテリアルの安全性確認は，重要な問題として認識されており，OECD や ISO 等を中心として評価手法の国際的標準化に向けた取り組みが進行しているところでもある．本稿では，ナノマテリアルの安全性評価

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本総説は，日本薬学会第 130 年会シンポジウム S18 で発表したものを中心に記述したものである．

の確立に向けたこれらの取り組みに貢献してきたわれわれの研究成果の一部と，それらの研究結果から帰納的に導き出された慢性影響評価研究の重要性について論ずる．

2. ナノマテリアルのリスク評価法の確立における課題

一般的に，化学物質の健康影響評価（リスクアセスメント）の基本的なフレームは，有害性評価と曝露評価，及び各々の評価内容を比較・統合化する過程のリスク判定のステップから成り立っている．この基本的なフレーム自体は，ナノマテリアルの健康影響評価に適用できるものであると考えられる．[1-5] しかし，ナノマテリアルに特徴的な新たな物理化学的性質，特にサイズが生体内高分子と近いことや，高い表面活性のために凝集し易い性質を考慮すると，よりサイズの大きい通常のバルク化合物や完全に溶解した単一分子化合物とは，生体内挙動が異なることが予想され，同じ化学組成の化合物であってもその毒性発現部位や発現様式は異なることが予想される．つまり，体内動態［吸収 absorption，分布 distribution，代謝 metabolism，排泄 excretion (ADME)］情報は，一般の化学物質より重要な意味を持つと考えられる．

そこで，生体内での挙動を把握するためには，生体試料中で検出，同定・定量できる方法を確立しなくてはならない．一般にナノマテリアルの開発段階において，その性質を把握するための物理化学的測定法も同時に開発されているはずであるが，それらの手法は生体試料中に存在するナノマテリアルにそのまま適用できないことも多い．さらに，機器分析法による生体試料中での検出や定量が可能になったとしても，生体内で実際にナノの状態で存在しているのか，あるいは再凝集などはしていないかなど，標的組織における最終的な生体内反応に影響を及ぼすと考えられる実際のナノマテリアルの存在状態を把握するためには，最終的には，組織標本の電子顕微鏡などによる確認が必要となる．

一方，体内動態に影響を与える因子として，投与法を検討する必要もある．単独では凝集し易いナノマテリアルをそのまま曝露するということは，物理的に巨大となった粒子は体への吸収性が低く，ナノマテリアル自体の体内動態や懸念される有害性を検出することが困難になると考えられるためである．そのために曝露実験時におけるナノマテリアルの分散手法の開発が必要となる．職業曝露などの比較的大量のナノマテリアル曝露の安全性を評価するという観点からは，凝集したままの曝露にも意義があるかもしれないが，製品中への混入や環境中への排出を経由した，分散された曝露も想定されることは考慮すべきであると考えられる．

Figure 1 は，凝集したナノマテリアルが，生体に取り込まれた場合に想定される体内動態を模式図化したものである．ナノマテリアルの使用用途にも依存するが，製品中のナノマテリアルはポリマー等の他の高分子化合物等と混合された状態，あるいはナノマテリアルだけが単独で製品から解離していく状態を考慮しても，この凝集性のために，大きな粒子として曝露する可能性が高いものと想定される．急性的には，このサイズの大きくなった物質は生体に取り込まれることはほとんどなく，局所的な刺激を起こすような変化を除いては，生体内で有害性が惹起される可能性は低いものと考えられる．しかし，仮に凝集したナノマテリアルが長期間に渡って，吸収部位である肺胞や消化管，損傷皮膚などの局所に滞留したり，慢性的に曝露したりするケースを想定すると，時間経過とともに小さくなった凝集体の粒子を除去するために，マクロファージなどの食細胞による取り込みや，表面活性の高いナノマテリアル分子と生体成分との結合作用による侵食作用により，生体に少しずつ取り込まれることが想定される．もしも生体内に取り込まれたナノマテリアルと生体内成分との結合性が高い場合には，容易に生体外に排出されることはなく，特定の組織等へ蓄積し易くなり，慢性影響の可能性を検討する必要が出てくると想定できる．

3. 国立医薬品食品衛生研究所における取り組みの成果の概要

以上のナノマテリアル固有の検討課題を考慮して，われわれは 2005 年より厚生労働科学研究の化学物質リスク研究事業の枠組みの中で，ナノマテリアルの健康影響評価手法の開発に係わる研究を推し進めてきたところである．われわれは，これらの検討課題を解決するために，Fig. 2 に示すように 4 つの項目を中心に研究を行ってきた．これらの項目の中で，*in vivo* 研究については，比較的研究初期の段階から中心的に取り組んできた．その中で，繊維

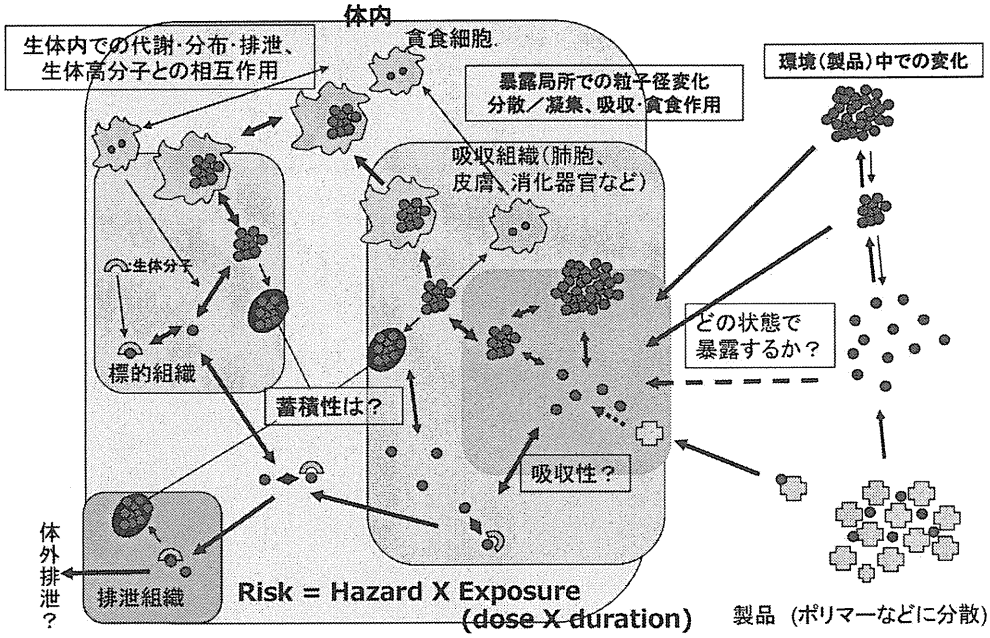


Fig. 1. The Estimated ADME Schema of Nanomaterials

***in vivo*試験法研究**

MWCNTのP53ヘテロ欠失マウスへのi.p.投与による中皮腫誘発性を確認
バイオマーカとしてマウスのメソセリン抗体の作成
一方、C60の腹腔内投与による慢性的影響として腎臓への影響を示唆
TiO_2と C60の気管内投与による発がんプロモーション作用の示唆

吸入試験法研究

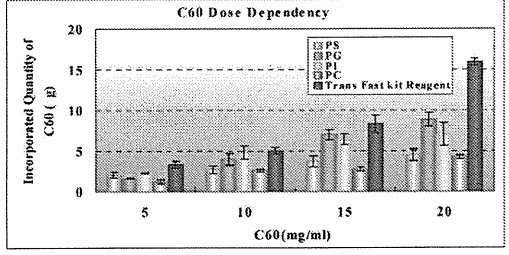
MWCNTのミスト暴露システムを開発
気管内投与時の分散性依存の発現様式差異を確認
リポソーム分散C60による気管内投与法を開発。

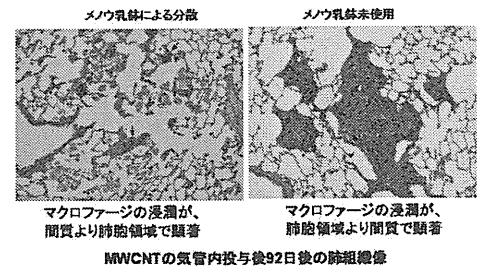
暴露測定法／動態解析研究

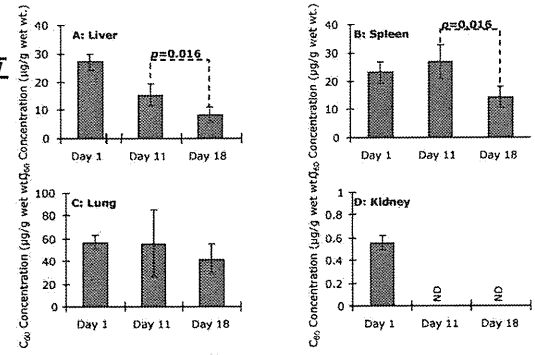
生体試料でのC60の定量的検出法との確立
静注後のC60の組織からの経時的消失検討
気管内投与後のMWCNTの肺及び肝臓での検出

***in vitro*試験法研究**

細胞培養系でのリポソーム等を用いた分散法の確立
→C60やTiO_2の遺伝毒性、細胞透過性、
神経系の細胞機能への影響、などへの適用







C_{60}のラットへの尾静脈投与(12.5 μg/kg)における体内分布
反復(4回)投与後の体内分布の経時変化

Fig. 2. The Overall Results of NIHS Projects for Nanomaterial Safety

長の長いタイプの多層型カーボンナノチューブ（MWCNT）が，中皮腫を誘発する可能性を持つことを確認した.[6] 上記の体内動態の重要性を考慮した概念からは，吸収性や体内分布について検証したのちに，慢性影響の可能性を検討することが論理的であるが，研究開始当時から，大量生産可能であった，酸化チタン（TiO_2）やフラーレン（C60），MWCNTについては，*in vivo* の慢性影響を先行して検討しておくべきであると判断した．特にその形状がアスベストに似ていたMWCNTについては，吸入曝露による有害影響が懸念されたが，MWCNTについての吸入曝露法が確立していない段階では，アスベストでも検証に使用されていた腹腔内投与による中皮腫誘発試験を行うこととした．

われわれの最初の実験は，アスベストで中皮腫の誘発時期が早くなることが知られているp53ヘテロノックアウトマウスへの腹腔内へ3 mg/mouseという高用量を投与することによって確認されたものであり，動物種の特異性や投与量の多さについて異論も指摘された．しかしその後の研究で，野生型の動物種であるF344ラットに対しても，同じMWCNTが中皮腫の誘発作用を持つことが確認された[7]ほか，投与量を1000分の1にまで少なくした実験においても中皮腫の起きることが示されている（投稿中）．

酸化チタンについては，雌ラットへの吸入曝露により発がん性のあることが示されているが，ナノサイズ化による発がん性の検証のために，気管内投与による肺がんのプロモーション作用の検討を行った．その結果，酸化チタンは，肺腺腫や乳腺腫に対してプロモーション作用を示し，その作用は，マクロファージから放出される炎症性因子であるMIP1αを介したものであることが示唆された.[8] 現在C60やMWCNTを用いたプロモーション作用の検討が進行中である．

一方，曝露手法の開発においては，ミスト法や粉体法によるMWCNTの吸入曝露システムの開発研究を進めているが，より簡易な手法として気管内投与のための適切な分散法の検討を行った．その結果，分散法の違いが肺の有害性発現様式に違いを引き起こすことを確認した.[9]

体内動態解析のために，生体試料中のC60やTiO_2の分析手法の開発や改良を行い，経口投与や気管内投与による体内吸収性について検討を行っている．現在のところ投与部位である消化管や肺以外で有意な検出量を確認できておらず，感度の向上に向けた研究を進めている．しかし，体内への吸収を前提にした解析として，C60の静脈内投与による解析を行ったところ，肝臓や脾臓，肺などへの分布を確認したが，腎臓への分布は極めて低いことが示された（投稿中）．その他，遺伝毒性や標的臓器などの毒性をスクリーニングするための *in vitro* 試験における培地等への分散法も検討対象としており，リポソームを用いたC60の分散法を確立した．

4. 慢性影響研究の重要性

ナノマテリアルの生体影響に関する情報はここ数年の活発な研究状況を反映して多くなりつつあるが，慢性影響に関する報告は依然その数が少ない状況である．一般の化学物質の有害性評価の常套手段として，変異原性試験や短期試験から情報を収集していくことは，必要なステップであり，OECDにおけるナノマテリアル作業グループの活動におけるスポンサーシッププログラムにおいても，加盟各国からの毒性試験情報として，短期試験を中心に収集されてきている．われわれの研究グループにおいても，これらの枠組みに対して，短期的な試験情報を中心に提供し始めている段階である．しかしMWCNTに関しては，研究初期から，短期毒性より長期毒性の方が懸念の強いことが，物性等の情報から推測されたところでもあり，その推定に基づいて，腹腔内投与の研究を最初にスタートさせた．腹腔内投与は，リスク評価の観点からは，曝露経路（吸入曝露）に伴う定量的な評価に問題のあるところであるが，最近の注目すべき研究として，分散剤で分散させたMWCNT（最高80 μgまで）をマウスに吸引させた研究や，MWCNT: 30 mg/m^{3}をマウスに単回吸入曝露した研究において，曝露後7–8週間目にMWCNTが胸膜に到達していたことが報告されている.[10,11] これらの研究結果は高用量の曝露による短期間の結果ではあるが，呼吸器を経由した曝露においてもMWCNTは胸膜（中皮）まで到達することを示唆しており，われわれの腹腔内投与による結果と合わせると，リスク評価の上でも重要な知見であると考えられる．

これらの腹腔内投与による中皮腫誘発能は，繊維状粒子による催腫瘍性のみを検出する系であり，短

いタイプやその他様々な形状の MWCNT における慢性毒性は別途検証する必要性がある．実際，われわれの行った腹腔内投与試験では，小さいサイズのナノチューブ繊維を含んだ細胞が腹膜の病変部のみならず，肝類洞内，又は肝葉間や腸間膜リンパ節の中にも認められ，体内に再分布することが示唆された（Fig. 3）.[6] さらに，SWCNT をマウスへ咽頭吸引させた実験では，一過性の急性症状の後に，炎症性細胞浸潤を伴わない間質の繊維化が認められている.[12] また，ApoE ノックアウトマウスを用いた実験では，タンパクカルボニル化活性の変化を伴うミトコンドリア DNA 障害と，アテローム性動脈硬化症の進行を増強することが示された.[13] MWCNT に関しても，マウスに MWCNT（200–400 μg）を気管内滴下した実験では，一過性の肺の炎症反応に加え，投与量に依存した血小板の活性化と凝固作用の活性化の促進が示唆されている.[14] また，MWCNT や SWCNT の気管内投与や経鼻投与により，アレルギー反応の増強反応が報告されている.[15–17] これらの結果が，カーボンナノチューブが直接体内循環に侵入した結果であるか，免疫細胞との接触を介した反応であるかを区別することは難しいが，曝露局所に留まらない全身作用の可能性を示している．われわれの酸化チタンの気管内投与による発がんプロモーション作用が，炎症因子により介在されたことは，これらの知見と同様の作用様式を示すものととらえることもできる．

以上の知見は，短期の試験だけでは検証することは困難であり，ナノマテリアルの有害性を確認するためには，長期の体内動態予測や慢性影響に関する研究が，重要なステップであることを示している．Figure 4 にスクリーニング試験や確定試験を開発するための手順についてまとめた．通常の化学物質については，その長い歴史の中で明らかとなった有害性に対して，それぞれの毒性発現様式に応じてスクリーニング試験が開発され，現在まで運用されてきている．特に変異原性試験は発がん性を予測する試験としての重要な役割を担っている．しかし，現時点ではナノマテリアルによる有害性影響が，これまでの研究経験の中で明らかとなった影響だけに留まるのかについては，まだ誰も判定できない状況である．これまでの一般化学物質に対応する有害性とスクリーニング試験を活用して進めていくと同時に，未知の影響を見極める最初のステップとして，少な

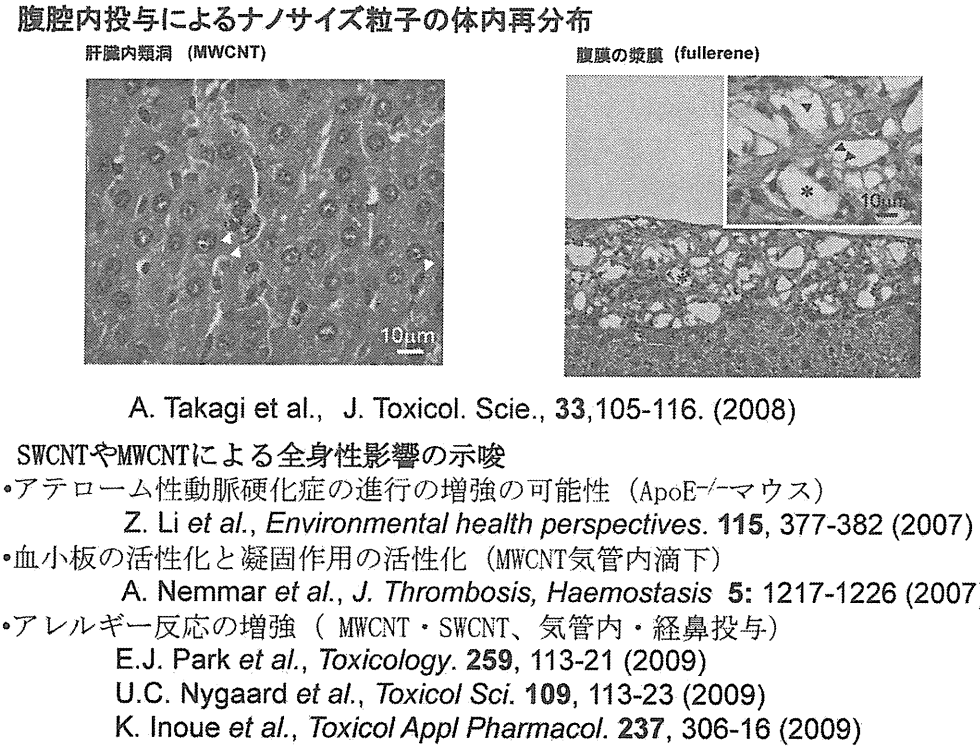


Fig. 3. The Suggestive Evidences for Systemic Toxicites by Nanomaterials