

Table 3. Continued.

CAS	Chemical name	log P Pred.	KATE class name	log (1/EC ₅₀ [mM])		Domain		"reactive" ¹⁴
				Pred.	Meas.	log P ²	C ³	
117-80-6	2,3-Dichloro-1,4-naphthoquinone	2.65	Halides1	3.06	4.13	O	X	Michael
1484-13-5	9-Vinylcarbazole	4.19	Conjugated systems1 (ClogP)	4.03	5.51	O	X	Michael
569-64-2	CI Basic Green 4	0.8	Amines aromatic or phenols5	0.56	3.45	X	X	Michael
101-20-2	3,4,4'-Trichlorodiphenylurea	4.9	Unclassified	2.71	4.3	O	X	acylating
89-40-7	4-Nitrophthalimide	1.12	Nitrobenzenes	0.45	0.84	X	X	acylating
86-73-7	Fluorene	4.02	Hydrocarbons aromatic	2.03	2.53	O	C (1)	
88-05-1	2,4,6-Trimethylaniline	2.72	Amines aromatic or phenols3	2.11	1.05	O	C (1)	
88-75-5	<i>o</i> -Nitrophenol	1.91	Amines aromatic or phenols4	1.06	0.7	O	C (1)	
91-17-8	Bicyclo[4.4.0]decane	4.2	Hydrocarbons aliphatic	2.22	2.78	O	C (1)	
91-17-8	Bicyclo[4.4.0]decane	4.2	Neutral organics	2.05	2.78	O	C (1)	
95-78-3	2,5-Dimethylaniline	2.17	Amines aromatic or phenols5	1.31	0.83	O	C (1)	
95-87-4	2,5-Xylenol	2.61	Amines aromatic or phenols4	1.46	1.37	O	C (1)	
100-63-0	Hydrazine, phenyl-	0.79	Hydrazines (ClogP)	2.14	3.83	O	C (1)	
108-87-2	Methylcyclohexane	3.59	Hydrocarbons aliphatic	1.81	2.47	O	C (1)	
108-87-2	Methylcyclohexane	3.59	Neutral organics	1.63	2.47	O	C (1)	
111-78-4	1,5-Cyclooctadiene	3.73	Hydrocarbons aliphatic	1.91	2.09	O	C (1)	
111-78-4	1,5-Cyclooctadiene	3.73	Neutral organics	1.73	2.09	O	C (1)	
129-00-0	Pyrene	4.93	Hydrocarbons aromatic	2.58	3.62	O	C (1)	
143-08-8	1-Nonanol	3.3	Alcohols or ethers aliphatic	1.06	1.4	O	C (1)	
143-08-8	1-Nonanol	3.3	Neutral organics	1.43	1.4	O	C (1)	
615-58-7	2,4-Dibromophenol	3.29	Amines aromatic or phenols4	1.85	2.08	O	C (1)	
634-93-5	2,4,6-Trichloroaniline	3.01	Amines aromatic or phenols3	2.15	1.66	O	C (1)	
716-79-0	1H-Benzimidazole, 2-phenyl-	3	Hydrocarbons aromatic	1.41	1.67	O	C (1)	
764-13-6	2,5-Dimethylhexa-2,4-diene	3.95	Hydrocarbons aliphatic	2.05	1.42	O	C (1)	
764-13-6	2,5-Dimethylhexa-2,4-diene	3.95	Neutral organics	1.88	1.42	O	C (1)	
924-41-4	1,5-Hexadien-3-ol	1.48	Conjugated systems2	1.22	0.58	O	C (1)	
2219-82-1	6- <i>tert</i> -Butyl- <i>o</i> -cresol	3.97	Amines aromatic or phenols4	2.24	1.49	O	C (1)	
2243-62-1	1,5-Naphthalenediamine	1.34	Amines aromatic or phenols1	2.55	1.62	O	C (1)	
4798-44-1	1-Hexen-3-ol	1.61	Conjugated systems2	1.25	-0.32	O	C (1)	
3295-94-1	Allyl <i>n</i> -hexyl ether	3.37	Conjugated systems2	1.62	1.77	O	C (1)	
120-95-6	2,4-Di- <i>tert</i> -pentylphenol	6.31	Amines aromatic or phenols4	3.59	3.29	X	C (1)	
693-98-1	2-Methylimidazole	0.61	Hydrocarbons aromatic	-0.04	-0.39	X	C (1)	
1116-76-3	Tri- <i>n</i> -octylamine	10.35	Secondary and tertiary amines	1.58	4.13	X	C (1)	

4418-61-5	5-Aminotetrazole	-3.41	Hydrazines (ClogP)	1.34	0.72	X	C (1)
7212-44-4	1,6,10-Dodecatrien-3-ol, 3,7,11-trimethyl-	5.68	Conjugated systems2	2.12	2.36	X	C (1)
16245-79-7	4-Octylbenzenamine	5.06	Amines aromatic or phenols3	2.42	3.84	X	C (1)
89-63-4	4-Chloro-2-nitroaniline	2.66	Amines aromatic or phenols3	2.1	1.61	O	C (2)
89-64-5	4-Chloro-2-nitrophenol	2.55	Unclassified	1.45	1.34	O	C (2)
96-96-8	2-Nitro- <i>p</i> -anisidine	2.1	Amines aromatic or phenols3	2.03	1.58	O	C (2)
101-80-4	4,4'-Diaminodiphenyl ether	2.22	Amines aromatic or phenols3	2.04	2.31	O	C (2)
827-52-1	Cyclohexylbenzene	4.81	Hydrocarbons aromatic	2.51	2.64	O	C (2)
843-55-0	1,1-Bis (4-hydroxyphenyl)-cyclohexane	5.48	Amines aromatic or phenols4	3.11	2.17	O	C (2)
2100-42-7	2-Chlorohydroquinonedimethylether	2.8	Unclassified	1.58	1.01	O	C (2)
78-51-3	Tri- <i>n</i> -butoxyethyl phosphate	3	Esters phosphate	4.02	1.08	X	C (2)
100-55-0	3-Hydroxymethylpyridine	-0.11	Hydrocarbons aromatic	-0.48	-0.79	X	C (2)
101-02-0	Triphenyl phosphite	6.62	Unclassified	3.63	2.84	X	C (2)
57-14-7	<i>N,N</i> -Dimethylhydrazine	-1.19	Hydrazines (ClogP)	1.76	1.66	O	X
374-25-4	1-Butene, 4-bromo-3-chloro-3,4,4-trifluoro-	3.78	Halides2	3.01	1.95	O	X
873-32-5	<i>o</i> -Chlorobenzonitrile	2.18	Amides and imides	1.05	0.88	O	X
948-65-2	2-Phenylindole	3.82	Hydrocarbons aromatic	1.9	2.81	O	X
2840-28-0	3-Amino-4-chlorobenzoic acid	1.6	Amines aromatic or phenols3	1.96	1.4	O	X
16691-43-3	3-Amino -5-mercapto-1H-1,2,4-triazole	0.31	Hydrazines (ClogP)	2.05	2.34	O	X
57500-00-2	Furfuryl methyl disulfide	2.95	Disulfides (ClogP)	2.35	2.71	O	X
63-74-1	Sulphanilamide	-0.55	Amines aromatic or phenols5	-0.19	1.12	X	X
90-30-2	1- (<i>N</i> -phenylamino)-naphthalene	4.47	Amines aromatic or phenols5	2.58	2.93	X	X
123-46-6	Ethanaminium, 2-hydrazino- <i>N,N,N</i> -trimethyl-2-oxo-, chloride	-5.29	Hydrazines (ClogP)	0.98	-0.24	X	X
462-08-8	3-Aminopyridine	-0.11	Amines aromatic or phenols5	0.05	1.12	X	X
497-18-7	Carbonohydrazide	-3.73	Hydrazines (ClogP)	1.28	0.98	X	X
504-24-5	4-Aminopyridine	-0.11	Amines aromatic or phenols5	0.05	0.8	X	X
504-29-0	2-Aminopyridine	0.53	Amines aromatic or phenols5	0.41	0.43	X	X
4418-61-5	5-Aminotetrazole	-3.41	Primary amines	-0.7	0.72	X	X
19715-19-6	3,5-Di- <i>tert</i> -butylsalicylic acid	6.06	Unclassified	3.33	1.89	X	X

Note: ¹When a chemical belongs to more than one QSAR class, all the predicted data are adopted.

²O: In-domain of log *P*-judgement, X: out-of-domain of log *P*-judgement.

³C (1): In-domain of *C*-judgement, defined as: (1) all substructures of a test chemical are found in reference chemicals in the class.

C (2): In-domain of *C*-judgement, defined as: (2) all substructures of a test chemical are in reference chemicals in either *Neutral organics* or the class.

X: Out-of-domain of *C*-judgement.

⁴Names of the reactive mechanistic domains are given in the text.

Table 4. Numbers (%¹) of chemicals characterised by underestimation (Under-), overestimation (Over-) and acceptable criterion (Acceptable) for fish toxicity.

Reactive domain	KATE domain					
	all	C(2)	C(1)	log P	log P & C(2)	log P & C(1)
<i>Reactive</i>						
Under-	4 (14.8)	2 (16.7)	2 (20)	3 (15.8)	1 (11.1)	1 (14.3)
Over-	2 (7.4)	1 (8.3)	1 (10)	1 (5.3)	1 (11.1)	1 (14.3)
Acceptable	21 (77.8)	9 (75)	7 (70)	15 (78.9)	7 (77.8)	5 (71.4)
<i>Non-reactive</i>						
Under-	5 (13.5)	4 (12.5)	4 (15.4)	3 (10)	2 (6.9)	2 (8.7)
Over-	3 (8.1)	1 (3.1)	0 (0)	1 (3.3)	1 (3.4)	0 (0)
Acceptable	29 (78.4)	27 (84.4)	22 (84.6)	26 (86.7)	26 (89.7)	21 (91.3)

Notes: ¹For example, the proportion of 'Under-' is defined as [(Number of under-)/(Number of under- + over- + Acceptable in the same reactive and KATE domains)].

Under-: Numbers (%) of chemicals for which toxicity was underestimated. Underestimation was defined as [calculated log (1/LC₅₀) - measured log (1/LC₅₀)] < -1.0.

Over-: Numbers (%) of chemicals for which toxicity was overestimated. Overestimation was defined as [calculated log (1/LC₅₀) - measured log (1/LC₅₀)] > 1.0.

Acceptable: Absolute errors between predicted and measured toxicities were less than 1.0.

Columns *all*, *C(2)*, *C(1)* and *log P* are defined in the footnote to Table 1.

Table 5. Numbers (%) of chemicals characterised by underestimation (Under-), overestimation (Over-) and acceptable criterion (Acceptable) for *Daphnia* toxicity.

Reactive domain	KATE domain					
	all	C(2)	C(1)	log P	log P & C(2)	log P & C(1)
<i>Reactive</i>						
Under-	7 (24.2)	2 (15.4)	1 (11.2)	6 (31.6)	2 (25)	1 (16.7)
Over-	3 (10.4)	1 (7.7)	1 (11.2)	2 (10.6)	1 (12.5)	1 (16.7)
Acceptable	19 (65.6)	10 (77)	7 (77.8)	11 (57.9)	5 (62.5)	4 (66.7)
<i>Non-reactive</i>						
Under-	7 (12.3)	4 (9.8)	4 (13)	2 (5.2)	2 (6.3)	2 (8)
Over-	6 (10.6)	3 (7.4)	2 (6.5)	3 (7.7)	2 (6.3)	2 (8)
Acceptable	44 (77.2)	34 (83)	25 (80.7)	34 (87.2)	28 (87.5)	21 (84)

Notes: *Under-*: Numbers (%) of chemicals for which toxicity was underestimated. Underestimation was defined as [calculated log (1/EC₅₀) - measured log (1/EC₅₀)] < -1.0.

Over-: Numbers (%) of chemicals for which toxicity was overestimated. Overestimation was defined as [calculated log (1/EC₅₀) - measured log (1/EC₅₀)] > 1.0.

Acceptable: Absolute errors between predicted and measured toxicities were less than 1.0.

Columns *all*, *C(2)*, *C(1)* and *log P* are defined in the footnote to Table 1.

after application of the log *P*-judgement, but it did increase after use of the *C*-judgement. In contrast, the proportion of "non-reactive" chemicals meeting the acceptable criterion did not increase with the use of the *C*-judgement at the *C(1)* rather than the *C(2)* level, but it did increase after the log *P*-judgement was applied. Unlike the fish end-point, the log *P*- and *C*-judgements did not guarantee that the predicted values of *Daphnia* toxicity

Table 6. Root mean square errors (RMSEs) for chemical toxicities according to the KATE domains, with respect to the reactive mechanistic domains.

End-point	Reactive domain	KATE domain					
		all	C(2)	C(1)	log P	log P & C(2)	log P & C(1)
Fish	Not considered ¹	1.22	0.95	0.96	1.10	0.90	0.88
	Reactive	1.59	1.37	1.32	1.56	1.50	1.48
	Non-reactive	0.84	0.74	0.77	0.66	0.59	0.59
Daphnia	Not considered	0.97	0.86	0.82	0.77	0.72	0.73
	Reactive	1.12	0.71	0.68	0.93	0.83	0.73
	Non-reactive	0.89	0.91	0.85	0.67	0.69	0.73

Notes: ¹ 'Not considered' shows the RMSE without considering the existence of the reactive mechanistic domains ('reactive' + 'non-reactive').

Columns all, C(2), C(1) and log P are defined in the footnote to Table 1.9

met the acceptable criterion when the chemicals were classified by reactive domain (either "reactive" or "non-reactive"). Although there was a small difference between log P with C(2) and log P with C(1) domains, the RMSEs became less than 1.0 with the in-domain of the C-judgement when the chemicals were categorised in "reactive" or "non-reactive" domains (Table 6). In other words, when these judgements were considered, chemicals in the "reactive" group did not always have a higher RMSE than those in the "non-reactive" group.

Poor efficiency was revealed in the SB or pro-SB reactive domains (Figure 2). The predicted *Daphnia* toxicities [$\log(1/EC_{50})$] of the chemicals that had these reactive substructures showed a difference of less than 1.0 from the measured toxicities. As shown in the case of the aldehyde substructure, $-CH=O$, in Figures 2(a)–(f), six SB chemicals were classified in the *aldehyde* class by KATE. The last of these, a pro-SB chemical (Figure 2(g)), was classified in the *secondary and tertiary amines* class by KATE. The pro-SB domains were thus inappropriate as additional information for *Daphnia* toxicity predictions for KATE. This was different from the fish toxicity prediction. One reason for the inconsistency between fish and *Daphnia* end-points is that *Daphnia* is a completely different organism from fish; consequently the "reactive" domain discussed is not always applicable to both *Daphnia* and fish.

3.3 Outliers

Our third focus is on the outliers among the "non-reactive" chemicals and the KATE log P- and C(1)-judgements (Figures 3 and 4). In the present article a chemical is considered to be an outlier if the KATE predicted toxicity differs by more than 1.0 $\log(1/LC_{50})$ or $\log(1/EC_{50})$ from the measured toxicity. The chemical structures of the outliers indicated that it was necessary to reconsider the fish and *Daphnia* toxicity predictions in the class that included aromatic amines and heterocyclics (Figures 3(a) and 3(b), and 4(a) and 4(b)). Additionally, the *Daphnia* toxicity predictions indicated that the presence of a pyrene structure (Figure 4(c)) was significant for its inclusion in the pro-S_N2 reaction mechanism, which for technical reasons is not defined by the current FITS. In addition, 1-hexen-3-ol (Figure 4(d)) was classified in the KATE *conjugated systems2* class, whose toxicity would not be predicted by only using log P descriptors [14].

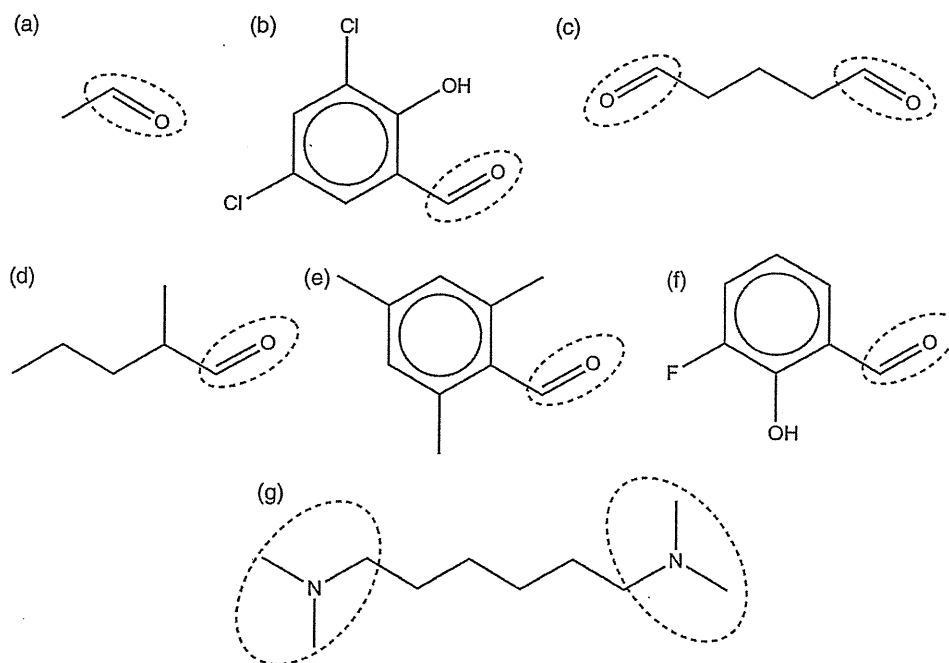


Figure 2. Chemical structure of (a) aldehyde, (b) 3,5-dichlorosalicylaldehyde, (c) glutaraldehyde, (d) methylvaleraldehyde, (e) 2,4,6-trimethylbenzaldehyde, (f) 3-fluoro-2-hydroxybenzaldehyde, and (g) *N,N,N',N'*-tetramethylhexamethylenediamine. The KATE predicted *Daphnia* toxicities, $\log(1/EC_{50})$, had less than 1.0 difference from the measured toxicities. Reactive substructures are indicated by dotted lines: SB (a–f) and pro-SB (g) reactive mechanisms.

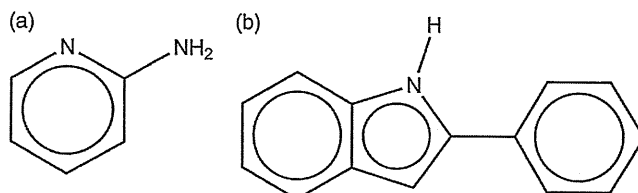


Figure 3. Outlier chemicals in the KATE fish toxicity prediction: (a) 2-aminopyridine, and (b) 2-phenylindole.

3.4 Applicability to KATE

Schultz and Cronin [48] have summarised both the essential and the desirable characteristics of ecotoxicity QSARs. They concluded that QSARs should be developed through the interaction of a group of multidisciplinary experts, since no single expert can construct a complete system to define toxicity in both fish and *Daphnia*. No one is able to categorise untrained or new chemicals, whose properties have yet to be established.

The QSAR models in KATE, even if they apply expert knowledge to the classification rules, have revealed certain drawbacks since their publication. For example, acrylonitrile is an alpha-beta unsaturated nitrile and due to the reactivity of other alpha-beta unsaturated

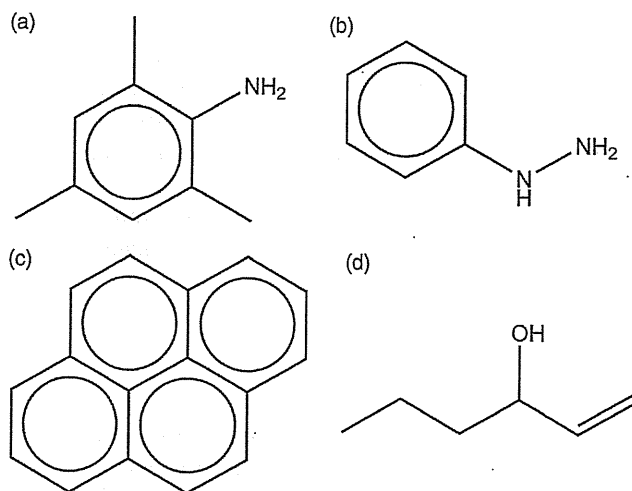


Figure 4. Outlier chemicals in the KATE fish toxicity prediction: (a) 2,4,6-trimethylaniline, (b) hydrazine, (c) pyrene, and (d) 1-hexen-3-ol.

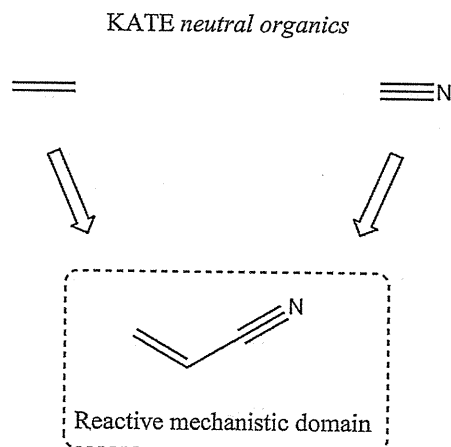


Figure 5. Acrylonitrile (centre), including chemical substructures within the “reactive” mechanistic domain [18,19]. Due to the existence of the ethane (right) and hydrogen cyanide (left) structures, this chemical was classified in *neutral organics* in KATE.

compounds is therefore expected to be toxic to fish. However, acrylonitrile itself is an “untrained” substructure for the purposes of the current version of KATE and is categorised in the *neutral organics* class, since both ethylene ($\text{CH}_2=\text{CH}_2$) hydrogen cyanide (HCN) are categorised as *neutral organics* (Figure 5). Neither the KATE classification nor *C*-judgement can filter the reactive substructure, “acrylonitrile”. The patterns introduced by Enoch et al. can thus reinforce chemical space information in the KATE system. This means that the classification rules of KATE can be reviewed and updated using information from other end-points. This procedure may play a corresponding role to QSAR development by the interaction of a range of multidisciplinary experts.

Moreover, SMARTS patterns may use a defined substructure to settle the *C*-judgement, especially for the fish end-point. But careful selection is required when the SMARTS patterns are applied to the *Daphnia* end-point. In such a situation the defined substructure seems to include the concept of "structural alerts". However, the structural alerts for aquatic toxicity QSARs [49] are normally defined in such a way as to discriminate between excess toxicity and narcotic effect levels [50,51]. The predicted toxicity discussed here was not from a QSAR for narcotic toxicity, and as a result of the KATE classification rules the QSAR models in the KATE system include discrimination from narcotic toxicity.

4. Conclusions

We have demonstrated the effectiveness of the reactive domains defined by skin sensitisation [16] applied to the ecotoxicity system, KATE. In predictions of fish toxicity, under identical judgements the group of chemicals with "reactive" substructures always had higher RMSEs than those with "non-reactive" substructures; this was confirmed by external validation of the current KATE system. The proposed reactive mechanism of skin sensitisation was valid for predicting fish toxicity using KATE. In the case of *Daphnia* on the other hand, the group of chemicals with "reactive" substructures did not always have higher RMSEs. One reason for this is that *Daphnia* organisms are very different in type from fish organisms: toxic action in *Daphnia* might not always correspond either to toxic action in fish nor to skin sensitisation potential. Even though there were some exceptions, the reactive domains discussed can be employed to improve the prediction accuracy in KATE. The presence of outliers in the test sets show, however, that the KATE classification needs to be reconsidered, especially in the case of chemicals containing nitrogen atoms.

We have previously concluded that QSAR development for KATE should in future focus on the reactivity of the chemicals and include multi-regression analysis [14]. As regards the latter, Hidaka et al. [52] have tested quantum chemical parameters, such as dipole, HOMO and LUMO, for the ecotoxicity prediction by using multiple regression analysis (MLR) and partial least squares regression (PLS) analyses based on the *Daphnia* dataset gathered by the Japan MoE [33]. However Hidaka et al. were unable to exclude the unnecessary descriptors [52].

As regards reactivity, the present study has demonstrated that reaction mechanistic domains of toxic action in terms of skin sensitisation offer useful complementary information for prediction of acute aquatic ecotoxicity, particularly at the fish end-point. Ultimately, the reliability of toxicity prediction is improved by the refined *C*-judgement rule, which utilises the classification of the substructures for skin sensitisation. The results of the validation study will help to improve prediction accuracy in the next version of the KATE system.

Acknowledgements

KATE was researched and developed by the Research Centre for Environmental Risk at the National Institute for Environmental Studies (NIES) between 2004 and 2009 under contract to the Japan MoE.

We are grateful to the US EPA for permission to use KOWWIN in "KATE on PAS", the standalone version of the KATE system. We also wish to thank Drs S.J. Enoch and M.T.D. Cronin for providing the raw data for the SMARTS strings [25].

References

- [1] *Report of the World Summit on Sustainable Development: Johannesburg, South Africa, 26 August–4 September 2002 (with corrigendum)*, United Nations, 2003. Available at <http://www.johannesburgsummit.org/html/documents/documents.html>
- [2] *Parliament Adopts REACH: New EU Chemicals Legislation and New Chemicals Agency*, 2006. Available at: <http://www.europarl.europa.eu/sides/getDoc.do?language=EN&type=IM-PRESS&reference=20061213IPR01493>
- [3] *Act on the Evaluation of Chemical Substances and Regulation of their Manufacture, etc. (CSCL)*, 2010. Available at: http://www.meti.go.jp/policy/chemical_management/english/cscl.html
- [4] *OECD Quantitative Structure–Activity Relationships [(Q)SARs] Project*. Available at: http://www.oecd.org/document/23/0,3343,en_2649_34379_33957015_1_1_1_1,00.htm
- [5] T. Netzeva, P. Manuela, and W. Andrew, *Review of Data Sources, QSARs and Integrated Testing Strategies for Aquatic Toxicity*, European Commission Joint Research Centre, 2007. Available at: http://ecb.jrc.ec.europa.eu/DOCUMENTS/QSAR/EUR_22943_EN.pdf
- [6] *OECD Environment Health and Safety Publications Series on Testing and Assessment, No. 69, Guidance Document on the Validation of (Quantitative) Structure–Activity Relationships [(Q)SAR] Models*, Paris, 2007. Available at <http://www.oecd.org/dataoecd/55/35/38130292.pdf>
- [7] T.I. Netzeva, A.P. Worth, T. Aldenberg, R. Benigni, M.T.D. Cronin, P. Gramatica, J.S. Jaworska, S. Kahn, G. Klopman, C.A. Marchant, G. Myatt, N. Nikolova-Jeliazkova, G.Y. Patlewicz, R. Perkins, D.W. Roberts, T.W. Schultz, D.T. Stanton, J.J.M. van de Sandt, W.D. Tong, G. Veith, and C.H. Yang, *Current status of methods for defining the applicability domain of (quantitative) structure–activity relationships: Report and recommendations of ECVAM Workshop 52, ATLA*, *Altern. Lab. Anim.* 33 (2005), pp. 155–173.
- [8] L. Eriksson, J. Jaworska, A.P. Worth, M.T.D. Cronin, R.M. McDowell, and P. Gramatica, *Methods for reliability and uncertainty assessment and for applicability evaluations of classification- and regression-based QSARs*, *Environ. Health Persp.* 111 (2003), pp. 1361–1375.
- [9] J. Jaworska, N. Nikolova-Jeliazkova, and T. Aldenberg, *QSAR applicability domain estimation by projection of the training set in descriptor space: A review*, *ATLA*, *Altern. Lab. Anim.* 33 (2005), pp. 445–459.
- [10] T.I. Netzeva, A.G. Saliner, and A.P. Worth, *Comparison of the applicability domain of a quantitative structure–activity relationship for estrogenicity with a large chemical inventory*, *Environ. Toxicol. Chem.* 25 (2006), pp. 1223–1230.
- [11] R.S. Boethling and J. Costanza, *Domain of EPI suite biotransformation models*, *SAR QSAR Environ. Res.* 21 (2010), pp. 415–443.
- [12] O.G. Mekenyan, S.D. Dimitrov, T.S. Pavlov, and G.D. Veith, *A systematic approach to simulating metabolism in computational toxicology: I. The TIMES heuristic modelling framework*, *Curr. Pharm. Des.* 10 (2004), pp. 1273–1293.
- [13] S. Dimitrov, G. Dimitrova, T. Pavlov, N. Dimitrova, G. Patlewicz, J. Niemela, and O. Mekenyan, *A stepwise approach for defining the applicability domain of SAR and QSAR models*, *J. Chem. Inf. Model.* 45 (2005), pp. 839–849.
- [14] A. Furuhashi, T. Toida, N. Nishikawa, Y. Aoki, Y. Yoshioka, and H. Shiraishi, *Development of an ecotoxicity QSAR model for the KASHINHOU Tool for Ecotoxicity (KATE) system, March 2009 version*, *SAR QSAR Environ. Res.* 21 (2010), pp. 403–413.
- [15] KATE: *It should be noted that the prediction results provided by the KATE system do not guarantee satisfactory prediction accuracy. It should be regarded as a tool for obtaining reference*

- figures for the extent of the ecotoxicity impact of a chemical substance. Furthermore, the results may not be used for MoE submissions in compliance with the CSCL, 2009. Available at <http://kate.nies.go.jp>
- [16] A.O. Aptula, G. Patlewicz, and D.W. Roberts, *Skin sensitization: Reaction mechanistic applicability domains for structure–activity relationships*, *Chem. Res. Toxicol.* 18 (2005), pp. 1420–1426.
- [17] J.L.M. Hermens, *Electrophiles and acute toxicity to fish*, *Environ. Health Perspect.* 87 (1990), pp. 219–225.
- [18] A.O. Aptula and D.W. Roberts, *Mechanistic applicability domains for nonanimal-based prediction of toxicological end points: General principles and application to reactive toxicity*, *Chem. Res. Toxicol.* 19 (2006), pp. 1097–1105.
- [19] D.W. Roberts, G. Patlewicz, P.S. Kern, F. Gerberick, I. Kimber, R.J. Dearman, C.A. Ryan, D.A. Basketter, and A.O. Aptula, *Mechanistic applicability domain classification of a local lymph node assay dataset for skin sensitization*, *Chem. Res. Toxicol.* 20 (2007), pp. 1019–1030.
- [20] S.D. Dimitrov, L.K. Low, G.Y. Patlewicz, P.S. Kern, G.D. Dimitrova, M.H.I. Comber, R.D. Phillips, J. Niemela, P.T. Bailey, and O.G. Mekenyan, *Skin sensitization: Modeling based on skin metabolism simulation and formation of protein conjugates*, *Int. J. Toxicol.* 24 (2005), pp. 189–204.
- [21] I.R. Jowsey, D.A. Basketter, C. Westmoreland, and I. Kimber, *A future approach to measuring relative skin sensitising potency: A proposal*, *J. Appl. Toxicol.* 26 (2006), pp. 341–350.
- [22] C. Grindon, R. Combes, M.T.D. Cronin, D.W. Roberts, and J.F. Garrod, *An integrated decision-tree testing strategy for skin sensitisation with respect to the requirements of the EU REACH legislation*, *ATLA, Altern. Lab. Anim.* 35 (2007), pp. 683–697.
- [23] G. Patlewicz, A.O. Aptula, D.W. Roberts, and E. Uriarte, *A minireview of available skin sensitization (Q)SARs/expert systems*, *QSAR Comb. Sci.* 27 (2008), pp. 60–76.
- [24] R.J. Vandebriel and H. van Loveren, *Non-animal sensitization testing: State-of-the-art*, *Crit. Rev. Toxicol.* 40 (2010), pp. 389–404.
- [25] S.J. Enoch, J.C. Madden, and M.T.D. Cronin, *Identification of mechanisms of toxic action for skin sensitisation using a SMARTS pattern based approach*, *SAR QSAR Environ. Res.* 19 (2008), pp. 555–578.
- [26] Daylight Chemical Information Systems Inc., *Daylight Theory Manual: 4. SMARTS® – A Language for Describing Molecular Patterns*. Available at <http://www.daylight.com/dayhtml/doc/theory/theory.smarts.html>
- [27] D. Weininger, *SMILES, A chemical language and information system: 1. Introduction to methodology and encoding rules*, *J. Chem. Inf. Comp. Sci.* 28 (1988), pp. 31–36.
- [28] Y. Yoshioka, *Know-how in the development of a program for QSAR platform, PAS: Part 1. Extract the component from the SMILES notation (in Japanese)*, *Jpn. J. Environ. Toxicol.* 11 (2008), pp. 33–40.
- [29] Y. Yoshioka, *Know-how in the development of a program for QSAR platform, PAS: Part 2. Drawing method of chemical structure (in Japanese)*, *Jpn. J. Environ. Toxicol.* 12 (2009), pp. 107–112.
- [30] Y. Yoshioka, *Know-how in the development of a program for QSAR platform, PAS: Part 3. An extraction method of partial structure (in Japanese)*, *Jpn. J. Environ. Toxicol.* 12 (2009), pp. 113–122.
- [31] R.L. Lipnick, D.E. Johnson, J.H. Gilford, C.K. Bickings, and L.D. Newsome, *Comparison of fish toxicity screening data for 55 alcohols with the quantitative structure–activity relationship predictions of minimum toxicity for nonreactive nonelectrolyte organic compounds*, *Environ. Toxicol. Chem.* 4 (1985), pp. 281–296.
- [32] R.L. Lipnick, *Narcosis, electrophile and proelectrophile toxicity mechanisms: Application of SAR and QSAR*, *Environ. Toxicol. Chem.* 8 (1989), pp. 1–12.
- [33] MoE, *Japan ecotoxicity test data*, 2009. Available at <http://www.env.go.jp/chemi/sesaku/02e.pdf>

- [34] US EPA fathead minnow database. Available at http://www.epa.gov/med/Prods_Pubs/fathead_minnow.htm
- [35] C.L. Russom, S.P. Bradbury, S.J. Broderius, D.E. Hammermeister, and R.A. Drummond, *Predicting modes of toxic action from chemical structure: Acute toxicity in the fathead minnow (Pimephales promelas)*, Environ. Toxicol. Chem. 16 (1997), pp. 948–967.
- [36] T.I. Netzeva, M. Pavan, and A.P. Worth, *Review of (quantitative) structure–activity relationships for acute aquatic toxicity*, QSAR Comb. Sci. 27 (2008), pp. 77–90.
- [37] OECD, *OECD Environment Monographs No. 58, Report of the OECD workshop on quantitative structure activity relationships (QSARs) in aquatic effects assessment*, Paris, 1992. Available at <http://www.oecd.org/dataoecd/19/27/35220377.pdf>
- [38] H.J.M. Verhaar, C.J. van Leeuwen, and J.L.M. Hermens, *Classifying environmental pollutants: 1. Structure–activity relationships for prediction of aquatic toxicity*, Chemosphere 25 (1992), pp. 471–491.
- [39] M.T.D. Cronin and J.C. Dearden, *QSAR in toxicology: 1. Prediction of aquatic toxicity*, Quant. Struct.–Act. Rel. 14 (1995), pp. 1–7.
- [40] H.J.M. Verhaar, E.U. Ramos, and J.L.M. Hermens, *Classifying environmental pollutants: 2. Separation of class 1 (baseline toxicity) and class 2 ('polar narcosis') type compounds based on chemical descriptors*, J. Chemom. 10 (1996), pp. 149–162.
- [41] H.J.M. Verhaar, J. Solbe, J. Speksnijder, C.J. van Leeuwen, and J.L.M. Hermens, *Classifying environmental pollutants: Part 3. External validation of the classification system*, Chemosphere 40 (2000), pp. 875–883.
- [42] S.P. Bradbury, C.L. Russom, G.T. Ankley, T.W. Schultz, and J.D. Walker, *Overview of data and conceptual approaches for derivation of quantitative structure–activity relationships for ecotoxicological effects of organic chemicals*, Environ. Toxicol. Chem. 22 (2003), pp. 1789–1798.
- [43] ECOSAR, 2009. Available at <http://www.epa.gov/oppt/newchems/tools/21ecosar.htm>
- [44] G.D. Veith and S.J. Broderius, *Rules for distinguishing toxicants that cause type-I and type-II narcosis syndromes*, Environ. Health Perspect. 87 (1990), pp. 207–211.
- [45] KOWWIN. Available at <http://www.epa.gov/opptintr/exposure/pubs/episuite.htm>
- [46] EPISuite. Available at <http://www.epa.gov/oppt/exposure/pubs/episuite.htm>
- [47] E.M. Hulzebos and R. Posthumus, *(Q)SARs: Gatekeepers against risk on chemicals?*, SAR QSAR Environ. Res. 14 (2003), pp. 285–316.
- [48] T.W. Schultz and M.T.D. Cronin, *Essential and desirable characteristics of ecotoxicity quantitative structure–activity relationships*, Environ. Toxicol. Chem. 22 (2003), pp. 599–607.
- [49] P.C. von der Ohe, R. Kühne, R.U. Ebert, R. Altenburger, M. Liess, and G. Schütürmann, *Structural alerts: A new classification model to discriminate excess toxicity from narcotic effect levels of organic compounds in the acute daphnid assay*, Chem. Res. Toxicol. 18 (2005), pp. 536–555.
- [50] G.D. Veith, D.J. Call, and L.T. Brooke, *Structure toxicity relationships for the fathead minnow, Pimephales promelas: Narcotic industrial chemicals*, Can. J. Chem. 40 (1983), pp. 743–748.
- [51] C.J. van Leeuwen, P.T.J. van der Zandt, T. Aldenberg, H.J.M. Verhaar, and J.L.M. Hermens, *Application of QSARs, extrapolation and equilibrium partitioning in aquatic effects assessment: 1. Narcotic industrial pollutants*, Environ. Toxicol. Chem. 11 (1992), pp. 267–282.
- [52] S. Hidaka, H. Shiraiishi, Y. Ohmayu, H. Yamasaki, K. Okamoto, N. Kawashita, T. Yasunaga, and T. Takagi, *Ecotoxicity prediction using 3D descriptors*, J. Comput.-Aided Chem. 11 (2010), pp. 11–18.

Estimation of the Respiratory Ventilation Rate of Preschool Children in Daily Life Using Accelerometers

Junko Kawahara

Research Center for Environmental Risks, National Institute for Environmental Studies, Tsukuba, Japan

Shigeho Tanaka

Health Promotion and Exercise Program, National Institute of Health and Nutrition, Tokyo, Japan

Chiaki Tanaka

Department of Health Science, College of Health and Welfare, J.F. Oberlin University, Tokyo, Japan

Yuki Hikihara

Faculty of Engineering, Chiba Institute of Technology, Narashino, Japan

Yasunobu Aoki and Junzo Yonemoto

Research Center for Environmental Risks, National Institute for Environmental Studies, Tsukuba, Japan

ABSTRACT

Inhalation rate is an essential factor for determining the inhaled dose of air pollutants. Here, accelerometers were used to develop regression equations for predicting the minute ventilation rate (\dot{V}_E) to estimate the daily inhalation rate in young children. Body acceleration and heart rate were measured in 29 Japanese preschool children (6 yr of age) during nine different levels of activities (lying down, sitting, standing, playing with plastic bricks, walking, building with blocks, climbing stairs, ball tossing, and running) using the Actical omnidirectional accelerometer, the ActivTracer triaxial accelerometer, and a heart rate monitor. Measurements were calibrated against the \dot{V}_E measured by the Douglas bag method. ActivTracer accelerometer measurements gave a strong correlation with \dot{V}_E (Pearson's $r = 0.913$), which was marginally

stronger than that for the Actical counts ($r = 0.886$) and comparable to the correlation between heart rate and logarithmic \dot{V}_E ($r = 0.909$). According to the linear regression equation, the \dot{V}_E for lying down, sitting, standing, playing with plastic bricks, walking, and running was overestimated by 14–60% by the Actical and by 14–37% by the ActivTracer. By comparison, for building with blocks, climbing stairs, and ball tossing, the \dot{V}_E was underestimated by 19–23% by the Actical and by 13–18% by the ActivTracer. When these three activities were excluded, a stronger correlation was found between the \dot{V}_E and ActivTracer measurements ($r = 0.949$); this correlation was 0.761 for the three excluded activities. Discriminant analysis showed that the ratio between vertical and horizontal acceleration obtained by the ActivTracer could discriminate walking from building with blocks, climbing stairs, and ball tossing with a sensitivity of 75%. The error in estimating \dot{V}_E was considerably improved for the ActivTracer measurements by the use of two regression equations developed for each type of activity.

IMPLICATIONS

Respiratory ventilation rate is an essential factor for assessing the health risk from air pollutants because it allows the dose of air pollutants delivered to the respiratory system to be determined. When establishing standards or criteria related to the management of health risks from hazardous environmental pollutants, the particular vulnerability of young children to environmental pollutants and their pattern of exposure should be considered. However, there are limited data on the respiratory ventilation rate of young children in daily life. To help address this problem, accelerometers were used to develop regression equations for predicting the \dot{V}_E to estimate the daily inhalation rate in young children.

INTRODUCTION

Respiratory ventilation rate is an essential factor for determining the daily inhaled dose of air pollutants, which is important information for the establishment of health criteria or guideline values for air quality. In recent years, the particular vulnerability of children to environmental pollutants and age-related differences in exposure have become a concern in health risk assessment and management.¹⁻³ In Japan, an inhalation rate of $15 \text{ m}^3 \cdot \text{day}^{-1}$ for a 50-kg adult is commonly used to estimate the daily inhaled dose of air pollutants.⁴ However, no standard inhalation rate value has been established for children. To

adequately assess adverse health risks to children, the inhalation rate of young children in daily life needs to be determined.

The doubly labeled water (DLW) method and the time-activity-ventilation method are experimental approaches that have been used to estimate daily inhalation rates during the daily activities of free-living individuals.⁵⁻¹¹ The DLW method is the most accurate method to measure the average total energy expenditure (EE) of people over several days and is based on the rates of disappearance of a dose of water containing the stable isotopes ²H and ¹⁸O. Although the DLW approach provides an accurate estimation of the daily inhalation rate, it is an expensive method and requires the collection of urine samples for 2-3 weeks. By comparison, the time-activity-ventilation method estimates the volume of air inhaled by a person over a defined period of time as a time-weighted average of the minute ventilation rate (\dot{V}_E) for each level of activity during the period. The daily inhalation rate using this approach is expressed by the following equation:

$$\text{Daily inhalation rate} = \left[\sum_{i=1}^k \bar{\dot{V}}_{Ei} \times t_i \right] \times 10^{-3} \quad (1)$$

where *Daily inhalation rate* is the 24-hr inhalation rate ($\text{m}^3 \cdot \text{day}^{-1}$), $\bar{\dot{V}}_{Ei}$ is the mean \dot{V}_{Ei} for each activity i ($\text{L} \cdot \text{min}^{-1}$), t_i is the time spent at each activity i ($\text{min} \cdot \text{day}^{-1}$), i is activity during the day, k is the number of activities, and 10^{-3} is a conversion factor. The critical issue in this approach is the need to accumulate adequate data for \dot{V}_E during short-term activities throughout the day. However, there have been quite limited experimental data on \dot{V}_E for children, especially for moderate-to-vigorous levels of physical activity and the habitual activities of daily life. The absence of data for children has been a source of error in estimations of daily inhalation rate by the time-activity-ventilation method. For example, Brochu¹² reported that the time-activity-ventilation approach can lead to an overestimation of the daily inhalation rates in 5- to 12-yr-old children by 75% because of the lack of \dot{V}_E data for children during different levels of activities in daily life.

Another limitation of the time-activity-ventilation method is the availability of suitable techniques to obtain the intensity and duration of physical activity of young children during daily life. Self-report activity diaries and heart rate monitoring are the traditional tools used to determine the level of physical activity for estimating the \dot{V}_E of free-living people. The disadvantages of self-reporting are that it tends to be subjective and is not practical for the assessment of young children, who are unable to accurately self-estimate and record their physical activity levels.¹³⁻¹⁵ Even if an observer records the activities of children, it is still not possible to accurately measure the intensity and duration of the activities of children, which can vary within a few seconds.^{16,17} Heart rate is a valid predictor of \dot{V}_E ,¹⁸ but its limitations include interruption of heart rate measurement by electrode displacement and discomfort from the electrodes, which can reduce the

compliance of children and their parents with the study requirements.¹⁵ To overcome these issues in monitoring the activity of children, an accelerometer could be a practical alternative for measuring the intensity, frequency, and duration of physical activity of free-living people. The advantages of an accelerometer-based approach are that it is noninvasive, it is not affected by the wearing of clothes, and it can provide data on a second time scale over long periods of time without interfering with normal activities. Many studies have examined the reliability and validity of these devices in children under laboratory and field conditions and determined that they provide valid measures of EE and oxygen (O_2) consumption in children.¹⁹⁻³¹

The primary goal of this study was to develop a regression equation to predict \dot{V}_E from accelerometer measurements to estimate the daily inhalation rate in young children. First evaluated was the validity of accelerometer measurements to predict \dot{V}_E by calibrating the Actical omniaxial accelerometer and the ActivTracer triaxial accelerometer against the \dot{V}_E measured at different levels of activity by the Douglas bag method. The performance of the accelerometers was also evaluated by comparing the accelerometer measurements with heart rate.

METHODS

Participants and Study Design

The study presented here was conducted from December 2006 through March 2007. The study population comprised 29 Japanese preschool children (16 boys and 13 girls) 6 yr of age (mean age, 6.5 ± 0.2 years) who were recruited from a single kindergarten class thanks to the cooperation of the administrative personnel. The children's mean height was 115.4 cm (SD, 5 cm; range, 103.4-124.6 cm) and mean body weight was 20.6 kg (SD, 2.4 kg; range, 17-26.3 kg); these data were obtained at the start of the experiment. The \dot{V}_E associated with nine different levels of physical activity were measured in a laboratory setting. Body acceleration and heart rate were measured by using activity and heart rate monitors and these measures were calibrated against the \dot{V}_E values. Written informed consent was obtained from the parents of each participant. The ethics committee of the National Institute (Tsukuba, Ibaraki, Japan) approved the experimental protocol for Environmental Studies.

Procedure

The experiment was conducted in a recreation room of the kindergarten from 2:00 to 6:00 p.m. Room temperature was set between 23 and 25 °C throughout the experiment to avoid the influence of cold temperature on the metabolism of the participants.^{32,33} All participants had fasted for at least 2 hr before the experiment to avoid an increase in metabolic rate.³³ At the start of the experiment, participants first lay on a floor mat without movement for at least 30 min to allow their metabolic rate to stabilize. After this equilibrating period, expired breath was collected for the resting state over 10 min. The participants then sequentially performed eight activities: (1) sitting in a chair, (2) standing, (3) playing with plastic bricks, (4) building with blocks, (5) walking, (6) climbing stairs (up and down), (7) tossing a ball, and (8) running. When the participants were sitting or standing, they were

asked to keep calm and watch a cartoon video for 10 min. The children played for 15 min with the plastic bricks while sitting in a chair at a desk. When building with blocks, participants were allowed to move freely within a 1.9- by 2.4-m area of free space for 5 min; this activity included standing, squatting, walking, and lifting and carrying blocks. For climbing stairs, participants repeated climbing up and down four steps for 2 min. For ball tossing, a handball was thrown to the participant from a set distance every 7 sec for 3 min; participants were asked to catch the ball and throw it back. For walking, participants were instructed to walk continuously at their normal pace along a straight course of 10 m for 4 min. Running was performed for 3 min under the same conditions as walking, but participants were encouraged to maintain a speed of more than $100 \text{ m} \cdot \text{min}^{-1}$.³⁴ The average speed was $69 \pm 6.4 \text{ m} \cdot \text{min}^{-1}$ for walking and $133 \pm 17 \text{ m} \cdot \text{min}^{-1}$ for running. The activities were chosen because they are commonly performed by children and cover a wide range of activity levels on the basis of observations in the kindergarten and because they were thought not to interrupt the breath sampling described below.

The \dot{V}_E during the nine different activities was measured by the Douglas bag method. During each activity, participants wore facemasks to collect expired breath. Expired breath was collected through tubes (2.5-cm inner diameter \times 80-cm length) into the 50-L Douglas bag near the end of each activity, after O_2 consumption ($\dot{V}\text{O}_2$) had stabilized. Previous studies have reported that $\dot{V}\text{O}_2$ and heart rate almost reach a plateau 1–2 min after the start of a 5-min endurance run in children 5 yr of age.^{35,36} The durations of the activities and sampling periods are summarized in Table 1.

Instruments

Body acceleration was recorded throughout the experiment by using an Actical accelerometer ($27 \times 28 \times 10 \text{ mm}$, 17 g, Mini-mitter, Inc.) and an ActivTracer accelerometer (type AC-210A, $48 \times 67 \times 16 \text{ mm}$, 57 g, GMS, Inc.). These two acceleration monitors were selected because of superior technical support and the following reasons. The Actical was selected because it was small, lightweight, and was considered to be suitable for invasive monitoring of activities of young children. The ActivTracer was selected because triaxial accelerometers

Table 1. Time periods for performance of physical activity and sampling of exhaled breath.

Activity	Time for Performance (min)	Time for Breath Sampling (min)
Lying	30	10
Sitting	10	5
Standing	7	5
Playing plastic bricks	10	5
Building with blocks	5	2
Walking	4	2
Climbing stairs	3	1
Ball tossing	3	1
Running	3	1

have been reported to capture the principal acceleration component, which varies considerably with type of activity (e.g., between arm work and walking)^{21,22} and would thus more accurately measure the free-living activities of children than uniaxial accelerometers. This notion is supported by a previous report that the output from a triaxial accelerometer provides a stronger correlation with $\dot{V}\text{O}_2$ than that of a uniaxial accelerometer.²³

The Actical accelerometer has an omnidirectional sensor that is sensitive to movements in all directions and is most sensitive to vertical movement when mounted on the hip. The sensor detects movements in the 0.5- to 3.2-Hz frequency range and generates an analog voltage. The voltage is amplified, filtered, and then digitized by an analog-to-digital converter to create a digital value. The process is repeated 32 times per second (32 Hz), or every 31.25 msec, and the 32 digitized values in 1 sec are summed. The resulting 1-sec value is divided by 4 and then added to an accumulated activity value for the time intervals (epoch) specified by the user. The Actical accelerometer provides the measurements in terms of activity counts.

The ActivTracer accelerometer contains a triaxial accelerometer that detects movements in the anteroposterior (x-axis), mediolateral (y-axis), and vertical (z-axis) directions. The sensor detects the movements in each of the three directions every 40 msec with a band-pass filter of 0.3–100 Hz. The measurement range of the sensor is 0 to $\pm 4 \text{ G}$ for the vertical direction and 0 to $\pm 2 \text{ G}$ for the anteroposterior and mediolateral directions, and the resolution is 0.002 G. The output measure of the ActivTracer is the average of absolute values for acceleration (mG) in each axis direction and a synthesis of acceleration in three directions (xyz, vector synthetic) for the time interval defined by the user. Note that the outputs of the two accelerometers are not comparable because each provides acceleration measurement through a different data processing algorithm.

During the experiment, participants wore the two accelerometers on the hip by using a belt (Figure 1). Heart rate was also monitored (ActiHR, Mini-mitter, Inc.) to compare the validity of the measurement for predicting \dot{V}_E with the accelerometer measurement. The sensor and lead of the heart rate monitor were attached to the participant's chest using disposable electrode pads in accordance with the manufacturer's instructions. Acceleration and heart rate were monitored every 15 sec.

The volume of expired breath was measured with a dry gas flow meter (Shinagawa Company). To measure $\dot{V}\text{O}_2$ and carbon dioxide (CO_2) production by participants, the O_2 and CO_2 concentrations in expired breath were analyzed by using a portable gas analyzer (AR-1, ARCO System, Inc.), which was calibrated and verified with outdoor air before each sample analysis.

Data Analysis

The volume of expired breath measured at ambient temperature and pressure and saturated with water vapor (ATPS condition) was converted to the volume at standard temperature and pressure, under dry conditions (STPD condition). With the exception of the data for running, if the respiratory quotient (which is expressed as the ratio between



Figure 1. Children wearing Actical and ActivTracer accelerometers on their hips with a belt. Heart rate was monitored at the same time using an ActiHR attached to the participant's chest. Dashed lines indicate the location of the sensor and lead of the heart rate monitor under the clothing.

CO_2 production [$\dot{V}\text{CO}_2$] and $\dot{V}\text{O}_2$) was outside of the range 0.75–0.99, the \dot{V}_E data were excluded from the analysis because the participant was in a state of hyperventilation or the exhaled breath had leaked from the facemask during breath sampling. Mean height, body weight, and \dot{V}_E values were compared between boys and girls by using the Student's *t* test. For comparison of acceleration among multiple activities, one-way analysis of variance (ANOVA) was conducted, followed by the Scheffe test for the individual comparison if the result of ANOVA was significant.

Linear regression analysis was performed to develop equations to predict \dot{V}_E from the synthetic acceleration from the accelerometers and heart rates with all of the participants and activities combined. The regression equations were presented as follows:

$$y = a + b \times x \quad (2)$$

where y is \dot{V}_E as an independent variable and x is the synthetic acceleration or heart rate as a dependent variable. The \dot{V}_E for each activity and participant was estimated from the linear regression equation. To evaluate

the validity, the \dot{V}_E estimated from activity counts measured by the Actical and ActivTracer and from heart rate measurements were individually compared with the \dot{V}_E measured by the Douglas bag method. The percentage differences were then calculated for each activity using a technique developed by Bland and Altman to assess the agreement between two methods of measurement.³⁷

Using the vertical and horizontal acceleration measured by the ActivTracer, linear discriminant analysis was performed to identify variables that strongly discriminate between the different types of activities. This analysis was used to dichotomize the type of activity into two groups, group 0 and group 1, and group was included in the analysis as a categorical dependent variable. The vertical activity counts (z) and the ratio between vertical and horizontal acceleration ($z/[x^2 + y^2]^{0.5}$) were included as independent variables. The significance of the discriminant function was evaluated with Wilks' λ . All statistical analyses were performed using SPSS software version 15 for Windows (SPSS, Inc.). Statistical significance was set at $P < 0.05$.

The physical activity ratio (PAR), which is the energy cost of an activity per unit time, was calculated to classify \dot{V}_E according to the intensity of physical activity on the basis of energy consumption. The PAR was calculated by dividing the EE ($\text{kcal} \cdot \text{min}^{-1}$) for a specific activity by the estimated basal metabolic rate (BMR).^{38,39} EE was calculated from the $\dot{V}\text{O}_2$ ($\text{L} \cdot \text{min}^{-1}$) and $\dot{V}\text{CO}_2$ ($\text{L} \cdot \text{min}^{-1}$) as described previously.⁴⁰ BMR was calculated by dividing the EE during the resting state by 1.1 on the assumption that the thermic effect of food is 10% of the BMR.³² PAR was categorized as sedentary ($\text{PAR} < 1.5$), light ($1.5 \leq \text{PAR} < 3$), moderate ($3 \leq \text{PAR} < 6$), and vigorous ($6 \leq \text{PAR}$) according to values given in the literature for the energy cost of physical activities.²⁶ All data in the text, tables, and figures are presented as mean \pm 1 SD.

RESULTS

The \dot{V}_E (STPD), $\dot{V}\text{O}_2$ (STPD), accelerometer measurements, and heart rates of participants during the nine physical activities are summarized in Tables 2 and 3. The \dot{V}_E ranged from $0.18 \pm 0.02 \text{ L} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$ when the participant was lying down to $1.07 \pm 0.29 \text{ L} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$ during running. There were no gender-associated differences ($P > 0.05$, *t* test) in the height or weight of participants or in \dot{V}_E during the various activities.

Table 2. Summary of \dot{V}_E and $\dot{V}\text{O}_2$ at STPD and PAR during nine different activities.

Activity	n	\dot{V}_E (STPD)		$\dot{V}\text{O}_2$ (STPD) $\text{L} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$	PAR
		$\text{L} \cdot \text{min}^{-1}$	$\text{L} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$		
Lying	27	3.9 ± 0.5	0.18 ± 0.02	0.006 ± 0.001	—
Sitting	24	4.4 ± 0.6	0.21 ± 0.02	0.007 ± 0.001	1.15 ± 0.77
Standing	25	4.5 ± 0.6	0.22 ± 0.03	0.007 ± 0.001	1.19 ± 0.21
Playing plastic bricks	28	5.1 ± 0.7	0.25 ± 0.03	0.008 ± 0.001	1.37 ± 0.36
Building with blocks	28	10.5 ± 1.7	0.51 ± 0.09	0.017 ± 0.003	2.98 ± 0.68
Walking	24	9.7 ± 1.5	0.47 ± 0.06	0.016 ± 0.002	2.80 ± 0.48
Climbing stairs	28	16.5 ± 3.3	0.81 ± 0.12	0.028 ± 0.004	4.79 ± 0.83
Ball tossing	28	15.4 ± 3.1	0.75 ± 0.12	0.026 ± 0.004	4.55 ± 0.89
Running	28	22.8 ± 4.8	1.07 ± 0.29	0.032 ± 0.005	5.74 ± 0.23

Table 3. Summary of accelerometer measurements and heart rate during nine different activities.

Activity	Accelerometer Measurements						Heart Rate (beats · min ⁻¹)	
	Actical (counts · 15 sec ⁻¹)		ActivTracer (mG · 15 sec ⁻¹)			n		
	n	Synthetic	n	Synthetic	Vertical			Horizontal
Lying	27	2 ± 4	27	7 ± 5	2 ± 2	6 ± 4	27	98 ± 9
Sitting	27	1 ± 3	28	15 ± 7	3 ± 2	11 ± 6	28	101 ± 10
Standing	29	5 ± 9	29	19 ± 11	6 ± 5	15 ± 8	29	102 ± 22
Playing with plastic bricks	27	23 ± 38	27	38 ± 16	7 ± 4	31 ± 13	25	111 ± 9
Building with blocks	26	217 ± 133	26	205 ± 57	116 ± 49	128 ± 26	26	134 ± 16
Walking	24	523 ± 176	24	341 ± 70	241 ± 62	179 ± 31	25	131 ± 13
Climbing stairs	24	772 ± 160	24	441 ± 78	321 ± 69	217 ± 34	24	162 ± 12
Ball tossing	26	673 ± 210	26	426 ± 94	293 ± 91	223 ± 29	26	161 ± 16
Running	26	1960 ± 370	25	1024 ± 126	855 ± 118	386 ± 70	27	177 ± 14

Synthetic acceleration ranged from 1 ± 3 counts · sec⁻¹ during sitting to 1960 ± 370 counts · sec⁻¹ during running when measured with the Actical accelerometer and from 7.4 ± 5.2 during lying down to 1024 ± 126 mG · sec⁻¹ during running with the ActivTracer accelerometer. The ActivTracer detected body acceleration in all participants during all nine activities. By comparison, the Actical detected body acceleration in only 78% of participants when lying down, 61% during sitting, 90% during standing, 75% while playing with plastic blocks, and in 100% of participants during all other activities. A significant difference was observed among the accelerations for resting, sitting, standing, and building with blocks when measured with the ActivTracer, whereas no significant difference was observed among the accelerations for these four activities when measured with Actical (ANOVA, $P < 0.05$). Heart rates ranged from 98 ± 9 beats · min⁻¹ when the participant was lying down to 177 ± 14 beats · min⁻¹ during running.

Calibration

There was a strong correlation between the experimentally measured \dot{V}_E using the Douglas bag method and the synthetic acceleration measured by the Actical and ActivTracer; Pearson's correlation coefficients were $r =$

0.886 ($n = 223$, $P < 0.001$) for the Actical and 0.913 ($n = 222$, $P < 0.001$) for the ActivTracer. Heart rates showed a high linear correlation with the logarithmic \dot{V}_E ($r = 0.909$, $n = 223$, $P < 0.001$).

The mean percentage differences between experimentally measured \dot{V}_E and that estimated from synthetic acceleration and heart rates were calculated using linear regression models (Table 4). The synthetic acceleration overestimated \dot{V}_E from lying down through playing with plastic bricks by 24–60% for the Actical and by 14–37% by the ActivTracer. \dot{V}_E values for walking and running were overestimated by 11–20% for the Actical and by 11–24% for the ActivTracer. By comparison, the Actical underestimated the \dot{V}_E values for building with blocks, ball tossing, and stair climbing by 19–23%, and the ActivTracer underestimated these values by 13–18%. When predicted from heart rates, \dot{V}_E for lying down through to playing with plastic bricks was overestimated by 14–19%, whereas that for walking through to running was underestimated by 2–11%, with the exception of ball tossing, which was within an overestimation of 1%. The overall mean differences between the observed and predicted \dot{V}_E were greater for the Actical ($13\% \pm 33\%$) and ActivTracer ($9\% \pm 24\%$) than for those calculated from heart rates ($4\% \pm 26\%$).

Table 4. Difference between observed and estimated \dot{V}_E from Actical and ActivTracer accelerometer measurements and heart rate using a single regression equation.

Activity	Actical		ActivTracer ^a		Heart Rate	
	Difference (L · kg ⁻¹ · min ⁻¹)	Percent Difference	Difference (L · kg ⁻¹ · min ⁻¹)	Percent Difference	Difference (L · kg ⁻¹ · min ⁻¹)	Percent Difference
Lying	0.11 ± 0.02	60.0 ± 22.5	0.06 ± 0.02	36.6 ± 19.0	0.03 ± 0.04	18.7 ± 23.5
Sitting	0.09 ± 0.02	43.0 ± 17.6	0.05 ± 0.02	24.6 ± 15.3	0.03 ± 0.05	13.8 ± 25.1
Standing	0.08 ± 0.03	37.3 ± 19.8	0.04 ± 0.03	21.8 ± 17.6	0.04 ± 0.06	19.7 ± 32.6
Playing plastic bricks	0.05 ± 0.03	24.1 ± 17.1	0.03 ± 0.03	14.1 ± 14.1	0.03 ± 0.04	13.9 ± 17.4
Building with blocks	-0.13 ± 0.04	-22.7 ± 13.9	-0.09 ± 0.07	-15.0 ± 12.1	-0.06 ± 0.10	-11.4 ± 19.4
Walking	0.06 ± 0.07	14.1 ± 14.4	0.09 ± 0.07	20.4 ± 15.1	-0.05 ± 0.10	-10.2 ± 20.3
Climbing stairs	-0.16 ± 0.11	-18.7 ± 11.0	-0.15 ± 0.11	-17.5 ± 12.3	-0.02 ± 0.16	-2.2 ± 20.9
Ball tossing	-0.15 ± 0.11	-18.7 ± 12.4	-0.11 ± 0.12	-13.5 ± 13.9	0.01 ± 0.19	0.6 ± 24.3
Running	0.08 ± 0.25	10.7 ± 24.9	0.08 ± 0.21	10.9 ± 20.9	-0.04 ± 0.27	-3.1 ± 23.9

Notes: ^aResults of estimation using a single regression equation are presented.

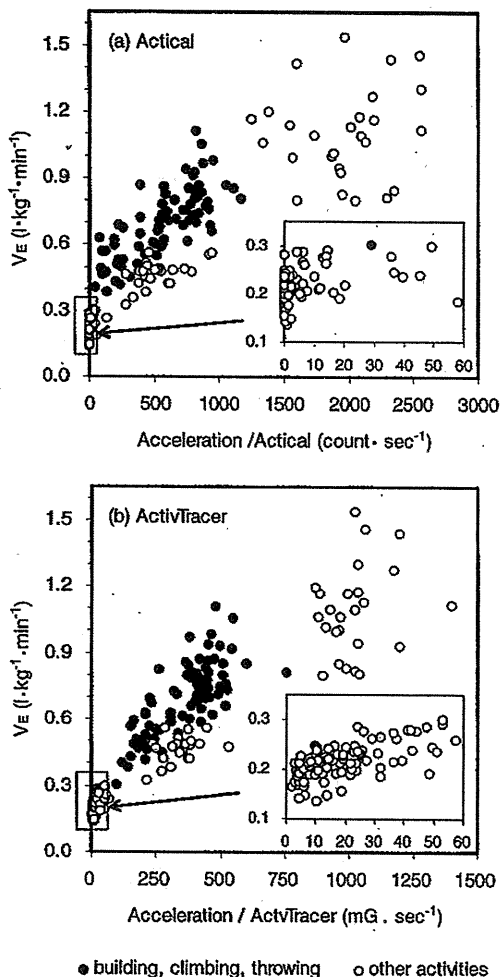


Figure 2. Scatterplots of \dot{V}_E vs. accelerometer measurement (vector synthesis): (a) Actical and (b) ActivTracer. Activities were divided into two groups: (1) building, climbing, and throwing and (2) all other activities.

Additional regression analysis showed that when building with blocks, ball tossing, and stair climbing were excluded, the correlation between \dot{V}_E and synthetic acceleration was improved, with $r = 0.949$ ($n = 146$, $P < 0.001$, $y = 0.0004x + 0.22$) for the Actical and $r = 0.962$ ($n = 145$, $P < 0.001$, $y = 0.00086x + 0.20$) for the ActivTracer. For the group of activities including building with blocks, ball tossing, and stair climbing, the correlation between \dot{V}_E and synthetic acceleration was $r = 0.781$ ($n = 77$, $P < 0.001$, $y = 0.0004x + 0.44$) for the Actical and $r = 0.761$ ($n = 77$, $P < 0.001$, $y = 0.00094x + 0.35$) for the ActivTracer (Figure 2).

Activity Type by Using Vertical and Horizontal Activity Counts

The linear discriminant analysis showed a significant difference between walking and the group of activities comprising building with blocks, stair climbing, and ball tossing (Wilks' $\lambda = 0.897$, $P < 0.005$). The discriminant function obtained was

$$f = -3.31 + 5.98 \times (z/[x^2 + y^2]^{0.5}) - 0.017 \times z \quad (3)$$

The standardized canonical coefficient was greater for the ratio between vertical and horizontal acceleration than for vertical acceleration alone, indicating that the ratio was a more significant discriminator than the vertical acceleration. The sensitivity of discrimination was 75% for group 1 (walking) and 70% for group 0 (other activities). The correct classification rate was 75% for walking, as category 1, and was 60, 64, and 89% for the category 0 activities building with blocks, stair climbing, and ball tossing, respectively.

The \dot{V}_E for each activity was recalculated in two steps for data obtained with the ActivTracer. When the synthetic acceleration of a given activity was between 96 and 754 $mG \cdot sec^{-1}$ (the range of accelerometer measurements by the ActivTracer from building with blocks through to ball tossing), the activity was classified as walking or other activities using the previously described discriminant function. The \dot{V}_E then was estimated from the synthetic acceleration measured by the ActivTracer using the regression equation developed for the group of activities to which the activity of interest was classified. As a result, the mean difference between observed and estimated \dot{V}_E was +10% for lying down, +1% for sitting, -1% for standing, -6% for playing with plastic bricks and building with blocks, +16% for walking, -12% for stair climbing, -0.3% for ball tossing, and +1% for running (Figure 3). The overall mean percentage difference was $0.2\% \pm 19\%$ ($-0.01 \pm 0.12 L \cdot kg^{-1} \cdot min^{-1}$).

Classification of \dot{V}_E

The mean PAR was 1.16 ± 0.07 for sitting, 1.20 ± 0.11 for standing, 1.38 ± 0.20 for playing with plastic bricks, 2.79 ± 0.48 for walking, 3.00 ± 0.56 for building with

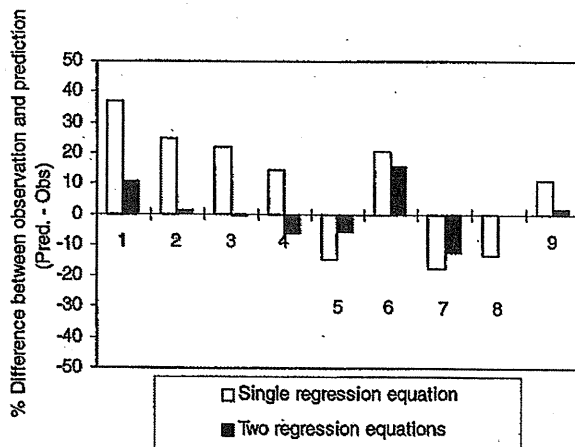


Figure 3. Percent difference between \dot{V}_E measured with a Douglas bag and that predicted from synthetic accelerometer measurements by the ActivTracer during nine different activities. Percent differences using the two regression equations were compared with those using the single regression equation. Activity: 1 = lying down, 2 = sitting, 3 = standing, 4 = playing with plastic bricks, 5 = building with blocks, 6 = walking, 7 = stair climbing, 8 = ball tossing, and 9 = running.

blocks, 4.84 ± 0.68 for climbing stairs, 4.54 ± 0.91 for ball tossing, and 5.79 ± 1.05 for running. The PAR indicates a sedentary state for sitting, standing, and playing with plastic bricks; a light-to-moderate level of physical activity for building with blocks and walking; moderate physical activity for stair climbing and ball tossing; and moderate-to-vigorous physical activity for running. The thresholds corresponding to PAR values of 1.5, 3, and 6, which are the cutpoints between sedentary-to-light, light-to-moderate, and moderate-to-vigorous levels of physical activity intensity were 132, 478, and 1783 counts \cdot sec $^{-1}$ for the Actical measurements and 71, 412, and 1093 mG \cdot sec $^{-1}$ (category 1) and 31, 218, and 593 mG \cdot sec $^{-1}$ (category 0) for the ActivTracer measurements.

DISCUSSION

In the study presented here, mean \dot{V}_E values were obtained for Japanese preschool children 6 yr of age during the resting state and during sedentary through to vigorous physical activity. The weight and height of the participants in this study were similar to those reported for 6-yr-old children by the Ministry of Health, Labor, and Welfare, Japan.⁴¹ The \dot{V}_E values presented here for sitting and standing, which were equivalent to 5.00 ± 0.64 and 5.23 ± 0.67 L \cdot min $^{-1}$, respectively, at body temperature, ambient pressure, and saturated with water vapor (BTPS) were consistent with the \dot{V}_E values of 4.98 ± 0.56 and 5.48 ± 0.44 L \cdot min $^{-1}$ (BTPS) in children 4–6 yr of age reported by Honda.⁴²

The accelerometer measurements were also validated for estimating the \dot{V}_E of young children during daily life. To the authors' knowledge, this study is the first to use accelerometer-based activity measurements to estimate \dot{V}_E for different levels of activity. There was a strong positive linear correlation between synthetic acceleration from the accelerometers and \dot{V}_E from the resting state through to vigorous physical activity, indicating that accelerometers can be used to estimate \dot{V}_E .

The comparison between the Actical and ActivTracer accelerometers presented here revealed smaller percentage differences between observed and predicted values with the ActivTracer, especially from resting through to light levels of physical activity. This result suggests that the ActivTracer more accurately measures physical activity and hence \dot{V}_E than does the Actical. This difference is largely because of the differences in the ability of the Actical and ActivTracer to sense body acceleration. The ActivTracer detected body acceleration during resting and sedentary activities in all participants and revealed significant differences among the accelerations for resting, sitting, standing, and building with blocks. In contrast, the Actical failed to detect body movement in all participants during these activities and could not detect differences in acceleration among the activities. On the basis of the correlation coefficients for the single linear regression model, heart rate was a better predictor of \dot{V}_E than the synthetic acceleration measurements from the accelerometers, but there was poor agreement between observed and predicted \dot{V}_E from the resting state through to light levels of activity.

The primary limitation of accelerometers for estimating \dot{V}_E is that a single regression model does not accurately predict the \dot{V}_E associated with activities during daily life. This study showed that the \dot{V}_E for building with blocks, stair climbing, and ball tossing was underestimated by using the synthetic acceleration from the Actical and ActivTracer accelerometers. The experimentally observed \dot{V}_E was significantly higher for these three activities than for walking, although activity counts for all of these activities varied similarly. This variability in the relationship between \dot{V}_E and synthetic acceleration resulted in marked differences between observed data and estimated values derived using the single regression model. Similar results have been reported in studies validating the use of accelerometer-based activity monitors for determining EE.^{43,44} The underlying explanation is that static work or upper body activities such as lifting, carrying, and throwing objects results in increased EE or $\dot{V}O_2$,^{45–49} and thus an increase in \dot{V}_E , without a proportional increase in body movement. This discrepancy cannot be captured by synthetic acceleration alone.

It was shown that the type of activity can be discriminated by using three-dimensional motion data obtained from a triaxial accelerometer. Discriminant analysis revealed that the ratio between vertical and horizontal acceleration and vertical acceleration alone were reasonably effective in discriminating walking from playing with plastic bricks, climbing stairs, and ball tossing as a group but were less effective in discriminating between playing with plastic bricks and climbing stairs. The study presented here also demonstrated that prediction of \dot{V}_E can be improved through discrimination of the type of activity using the three-dimensional motion data and use of one of two regression models specific for the type of activity. Using this approach, the percentage difference between observed and predicted \dot{V}_E was improved for each activity by an average of 14% and was even smaller than those predicted from heart rate (Table 4 and Figure 3).

This approach for improving the accuracy of estimating \dot{V}_E is essential for the more accurate assessment of the short-term or daily dose of air pollutants through inhalation, especially from the resting state through to light physical activity, because people tend to spend the greatest proportion of time at these levels of activity.^{50,51} For example, for lying down, the 37% overestimation in \dot{V}_E resulting from the single regression equation with all activities combined is estimated to result in an overestimation of 8.6% in daily inhalation volume given the assumption that in 1 day children spend 10.2 hr in a resting state; 4 hr in a sedentary state; and 8.3, 1.3, and 1.2 hr in light, moderate, and vigorous levels of physical activity, respectively.⁵² By comparison, the 10% overestimation in \dot{V}_E for lying down calculated by the two regression equations is estimated to result in only a 2% overestimation in daily inhalation volume using the same assumptions.

Misclassification of the activity type by the discriminant function still leads to inaccurate estimations of \dot{V}_E for walking and climbing stairs. Using the assumption described above, a 16% error in the estimation for light-level activities (e.g., walking) is roughly equivalent to 7%

of the total daily inhalation volume. However, an overestimation of \dot{V}_E can be canceled out, at least in part, by underestimation of V_E for other light-to-moderate activities (e.g., building with blocks and stair climbing). Underestimation of \dot{V}_E for climbing stairs will not cause considerable error because generally people spend only brief amounts of time in this type of activity during daily life.

The study presented here demonstrates that accelerometer measurements are a valid predictor of \dot{V}_E in children and overcome some of the limitations of traditional approaches. The traditional approach using self-report activity diaries has limited objectivity and accuracy in quantifying the intensity of physical activity of free-living people. For example, Bender¹⁶ conducted a study of 65 children aged 10–16 yr together with their parents and reported a poor correlation between proxy records of children's physical activity by parents and the metabolic equivalent value on the basis of accelerometer measurements as an objective measure of physical activity ($r \leq 0.383$). In another study, Lichtman⁵³ reported that the actual daily EE, measured by the DLW method, was overestimated by 50% in self-report activity diaries for adults. The inherent inability of self-reports to accurately quantify the intensity of physical activity could thus result in errors in estimating the daily inhalation rate. The validity of accelerometers in estimating \dot{V}_E was estimated by comparing measurements from them with measurements from the Douglas bag method, and it was found that the correlation between accelerometer measurement and \dot{V}_E is strong and that the error in estimating \dot{V}_E using the regression equations derived from ActivTracer measurements is between –12% and +16% depending on the activity. Also, this difference was smaller than those estimated from the heart rate measurements.

Standards for categorizing \dot{V}_E have not been defined for children. Here, \dot{V}_E was classified in relation to the level of physical activity for children in terms of the PAR. Differences in this classification structure between studies might lead to differences in the \dot{V}_E corresponding to the activity level of interest and the estimates of the time spent in activities of different intensity categories. A previous study⁵⁴ presented V_E corresponding to the level of physical activity intensity described in terms of metabolic equivalents (METs), which are expressed as a multiple of the resting metabolic rate for an activity. One MET is generally considered equivalent to the resting $\dot{V}O_2$ of 3.5 mL · kg⁻¹ · min⁻¹ in adults. However, in children, the resting metabolic rate is known to exceed 3.5 mL · kg⁻¹ · min⁻¹.^{26,55} Puyau et al.²⁴ reported that use of the adult default value resulted in significant differences between the observed PAR and estimated MET and that this difference was greater in younger children and at higher levels of EE. Thus, use of the adult default value would inaccurately reflect the level of intensity of physical activities in young children.

One of the shortcomings of this study is the small number of subjects. Further studies with a larger study group would more accurately represent the population of 6-yr-old children in Japan. Repeated measurements of \dot{V}_E are also needed to obtain a more reliable assessment of \dot{V}_E in young children. The conclusions presented here are based on nine different levels of physical activity in a

laboratory setting and provide strong groundwork for further analysis and research. Additional studies covering a more comprehensive range of activities are needed to confirm the validity of this approach for prediction of \dot{V}_E and to better understand the factors that contribute to variability in accelerometer measurements.

ACKNOWLEDGMENTS

The authors thank Seiko Yamaguchi for her technical advice on V_E measurements and Naoko Yoshida, Mai Yamanishi, Makiko Yamamoto, Mamiko Iwasaki, and Satomi Hara for their assistance with the laboratory work. The authors thank all of the participants, their parents and teachers, and the administration of the kindergarten for their kind participation and support. The Ministry of Environment of Japan supported this study from 2005 through 2006. The contents of this article are solely the responsibility of the authors and do not necessarily represent official views of the ministry. The peer reviewer provided excellent comments that resulted in an improved paper.

REFERENCES

1. Strategic Approach to International Chemical Management (SAICM), Report of the International Conference on Chemicals Management on the Works of Its First Session; available at http://www.chem.unep.ch/ICCM/meeting_docs/lccm1_77%20Report%20E.pdf (accessed April 19, 2010).
2. A Framework for Assessing Health of Environmental Exposures to Children; EPA/600/R-05/093F; U.S. Environmental Protection Agency; National Center for Environmental Assessment; Washington, DC, 2006; available at <http://www.epa.gov/ncea> (accessed April 19, 2010).
3. Principle for Evaluating Health Risks in Children Associated with Exposure to Chemicals; Environmental Health Criteria 237; World Health Organization; Geneva, Switzerland 2006.
4. Report on Committee for Setting Goal for Reducing the Level of Dioxin in the Atmosphere; Japan Environment Agency; Tokyo, Japan, 1997.
5. Brochu, P.; Robitaille, D.; Francois, J.; Brodeur, J. Physiological Daily Inhalation Rates for Free Living Individuals Aged 1 Month to 96 Years Using Data from Doubly Labeled Water Measurements: A Proposal for Air Quality Criteria, Standard Calculations and Health Risk Assessment; *Hum. Ecol. Risk Assess.* 2007, 12, 675-701.
6. Splier, C.E.; Little, D.E.; Trim, S.C.; Johnson, T.R. Linn; W.S.; Hackney, J.D. Activity Patterns in Elementary and High School Students Exposed to Oxidant Pollution; *J. Expo. Anal. Environ. Epidemiol.* 1992, 2, 277-293.
7. Allan, M.; Richardson, G.M. Probability Density Functions Describing 24-Hour Inhalation Rates for Use in Human Health Risk Assessment; *Hum. Ecol. Risk Assess.* 1998, 4, 379-408.
8. Layton, D.W. Metabolically Consistent Breathing Rates for Use in Dose Assessment; *Health Phys.* 1993, 64, 23-36.
9. McCurdy, T. Conceptual Basis for Multi-Route Intake Dose Modeling Using an Energy Expenditure Approach; *J. Expo. Anal. Environ. Epidemiol.* 2000, 10, 86-97.
10. Exposure Factors Handbook; U.S. Environmental Protection Agency; National Center for Environmental Assessment; Office of Research and Development; Washington, DC, 1997.
11. Child-Specific Exposure Factors Handbook; EPA/600/R-06/096F; U.S. Environmental Protection Agency; National Center for Environmental Assessment; Office of Research and Development; Washington, DC, 2008.
12. Brochu, P.; Robitaille, D.; Francois, J.; Brodeur, J. Physiological Daily Inhalation Rates for Free Living Individuals Aged 2.6 Month to 96 Years Based on Doubly Labeled Water Measurements: Comparison with Time-Activity-Ventilation and Metabolic Energy Conversion Estimates; *Hum. Ecol. Risk Assess.* 2006, 12, 736-761.
13. Sallis, J.F. Self Report Measures of Children's Physical Activity; *J. Sch. Health.* 1991, 61, 215-219.
14. Coleman, K.J.; Saelens, B.E.; Wiedrich-Smith, M.D.; Finn, J.D.; Epstein, L.H. Relationships between TriTrac-R3D Vectors; Heart Rate, and Self Report in Obese Children; *Med. Sci. Sports Exerc.* 1997, 29, 1535-1542.
15. Terblanche, A.P.S.; Ozkaynak, H.; Spengler, J.D.; Butler, D.A. Relationship between Self Reported Activity Levels and Actual Heart Rates in Teenagers; *J. Air & Waste Manage. Assoc.* 1991, 41, 942-946.

16. Bender, J.M.; Brownson, R.C.; Elliott, M.B.; Haire-Joshu, D.L. Children's Physical Activity: Using Accelerometers to Validate a Parent Proxy Record; *Med. Sci. Sports Exerc.* **2005**, *37*, 1409-1413.
17. Bailey, R.C.; Olson, J.; Pepper, S.L.; Porszasz, J.; Barstow, T.J.; Cooper, D.M. The Level and Tempo of Children's Physical Activities: An Observational Study; *Med. Sci. Sports Exerc.* **1995**, *27*, 1033-1041.
18. Adams, W.C. *Measurement of Breathing Rate and Volume in Routinely Performed Daily Activities*; Contract No. A033-205, Final Report; California Environmental Protection Agency; Air Resources Board: Sacramento, CA, 1993.
19. Welk, G.J. Measurement Issues in the Assessment of Physical Activity in Children; *Res. Q. Exerc. Sport.* **2000**, *71*, 59-73.
20. Meljer, G.A.; Westerterp, K.R.; Koper, H.; Hoor, F.T. Assessment of Energy Expenditure by Recording Heart Rate and Body Acceleration; *Med. Sci. Sports Exerc.* **1989**, *21*, 343-347.
21. Bouten, C.V.; Westerterp, K.R.; Verduin, V.; Janssen, J.D. Assessment of Energy Expenditure for Physical Activity Using a Triaxial Accelerometer; *Med. Sci. Sports Exerc.* **1994**, *26*, 1516-1523.
22. Eston, R.G.; Rowlands, A.V.; Ingledew, D.K. Validity of Heart Rate, Pedometer and Accelerometry for Predicting Energy Cost of Children's Activities; *J. Appl. Physiol.* **1998**, *84*, 362-371.
23. Ott, A.E.; Pate, R.R.; Trost, S.G.; Ward, D.S.; Saunders, R. The Use of Uniaxial and Triaxial Accelerometers to Measure Children's "Free Play" Activity; *Pediatr. Exerc. Sci.* **2000**, *12*, 360-370.
24. Puyau, M.R.; Adolph, A.L.; Vohra, F.A.; Butte, N.F. Validation and Calibration of Physical Activity Monitors in Children; *Obes. Res.* **2002**, *10*, 150-157.
25. Rodriguez, G.; Beghin, L.; Michaud, L.; Moreno, L.A.; Turck, D.; Gottrand, F. Comparison of the TriTrac-R3D Accelerometer and a Self-Report Activity Diary with Heart-Rate Monitoring for the Assessment of Energy Expenditure in Children; *Br. J. Nutr.* **2002**, *87*, 623-631.
26. Puyau, M.R.; Adolph, A.L.; Vohra, F.A.; Zakeri, I.; Butte, N.F. Prediction of Activity Energy Expenditure Using Accelerometers in Children; *Med. Sci. Sports Exerc.* **2004**, *36*, 1625-1631.
27. Pfeiffer, K.A.; McIver, K.L.; Dowda, M.; Almeida, M.J.; Pate, R.R. Validation and Calibration of the Actical Accelerometer in Preschool Children; *Med. Sci. Sports Exerc.* **2006**, *38*, 152-157.
28. Trost, G.; Ward, D.S.; Moorehead, S.M.; Watson, P.D.; Riner, W.; Bruke, J.R. Validity of the Computer Science and Applications (CSA) Activity Monitor in Children; *Med. Sci. Sports Exerc.* **1998**, *30*, 629-633.
29. Plasqui, G.; Joosen, A.M.; Kester, A.D.; Westerterp, K.R. Measuring Free Living Energy Expenditure and Physical Activity with Tri-Axial Accelerometry; *Obes. Res.* **2005**, *13*, 1363-1369.
30. Freedson, P.S.; Pober, D.; Janz, K.F. Calibration of Accelerometer Output for Children; *Med. Sci. Sports Exerc.* **2005**, *37*, S523-S530.
31. Freedson, P.; Miller, K. Objective Monitoring of Physical Activity Using Motion Sensors and Heart Rate; *Res. Q. Exerc. Sport.* **2000**, *71*, 21-29.
32. Kobayashi, S. *Nutrition Standards and Recommendations: Recent Trends in RDA, RDI and Dietary Guidelines and Their Background*; Kenpakusya: Tokyo, Japan, 1997; pp 86-96.
33. Kashiwazaki, H.; Dejlma, Y.; Suzuki, T. Influence of Upper and Lower Thermoneutral Room Temperatures on Fasting and Post-Prandial Resting Metabolism under Different Outdoor Temperature; *Eur. J. Clin. Nutr.* **1990**, *44*, 405-413.
34. Yoshizawa, S. *Aerobic Capacity in Young Children*; Kyorin-Shoin: Tokyo, Japan, 2002; pp 61-71.
35. Yoshizawa, S.; Honda, H.; Urushihara, M.; Nakamura, N. The Study on Aerobic Work Capacities of Preparatory School Children III. *Jpn. J. Phys. Fitness Med.* **1981**, *30*, 73-85.
36. Hatano, Y.; Ono, M.; Miyazaki, Y. Heart Rate Response and Work Intensity on Treadmill Running in Preschool Children; *Rep. Res. Center Phys. Ed.* **1981**, *9*, 127-136.
37. Bland, J.M.; Altman, D.G. Statistical Methods for Assessing Agreement between Two Methods of Clinical Measurement; *Lancet* **1986**, *1*, 308-310.
38. *Human Energy Requirements. Report of a Joint FAO/WHO/UNU Expert Consultation*; Food and Nutrition Technical Report Series I; U.N. Food and Agriculture Organization: Geneva, Switzerland, 2004; pp 35-52.
39. *Dietary Reference Values for Food Energy and Nutrients for the United Kingdom*; Report on Health and Social Subjects 41; 16th Impression; Department of Health; The Stationery Office: London, United Kingdom, 2007; pp 15-38.
40. Weir, J.B. New Methods for Calculating Metabolic Rate with Special Reference to Protein Metabolism; *J. Physiol.* **1949**, *109*, 1-9.
41. *The National Health and Nutrition Survey in Japan, 2003*; Ministry of Health, Labor, and Welfare of Japan: Tokyo, Japan, 2006; pp 318-319.
42. Honda, H.; Watanabe, N.; Ito, K.; Nakamura, N.; Yoshizawa, S. Changes in the Effects of 18 Month Endurance Run Training on Aerobic Work Capacity in Young Children; *Jpn. J. Phys. Fitness Med.* **1995**, *44*, 251-266.
43. Hendelman, D.; Miller, K.; Baggett, C.; Debold, E.; Freedson, P. Validity of Accelerometry for the Assessment of Moderate Intensity Physical Activity in the Field; *Med. Sci. Sports Exerc.* **2000**, *32*, S442-S449.
44. Tanaka, C.; Tanaka, S.; Kawahara, J.; Midorikawa, T. Triaxial Accelerometry for Assessment of Physical Activity in Young Children; *Obesity* **2007**, *15*, 1233-1244.
45. Debusk, R.F.; Valdez, R.; Houston, N.R.N.; Haskell, W. Cardiovascular Responses to Dynamic and Static Effort Soon after Myocardial Infarction; *Circulation* **1978**, *58*, 368-375.
46. Toner, M.M.; Sawka, M.N.; Levine, L.; Pandolf, K. Cardiorespiratory Responses to Exercise Distributed between the Upper and Lower Body; *J. Appl. Physiol.* **1983**, *54*, 1403-1407.
47. Vander, L.B.; Franklin, B.A.; Wrisley, D.; Rubenfire, M. Cardiorespiratory Responses to Arm and Leg Ergometry in Woman; *Phys. Sports Med.* **1984**, *12*, 101-106.
48. Hangeman, F.C.; Lawrence, R.A.; Mansfield, M.C. A Comparison of Energy Expenditure during Rowing and Cycling Ergometry; *Med. Sci. Sports Exerc.* **1988**, *20*, 479-488.
49. Pendergast, D.R. Cardiovascular, Respiratory, and Metabolic Responses to Upper Body Exercise; *Med. Sci. Sports Exerc.* **1989**, *21*, S121-S125.
50. *2005 NHK National Time Use Survey*; NHK Broadcasting Culture Research Institute: Tokyo, Japan, 2006; available at http://www.nhk.or.jp/bunken/book/book_data/bookdata_06020701.html (accessed April 19, 2010).
51. Hubal, E.A.C.; Sheldon, E.A.C.; Burke, J.M.; McCurdy, T.R.; Barry, M.R.; Rigas, M.L.; Zartarian, V.G.; Freeman, N.C.G. Children's Exposure Assessment: A Review of Factors Influencing Children's Exposure, and the Data Available to Characterize and Assess that Exposure; *Environ. Health Perspect.* **2000**, *108*, 475-486.
52. Kawahara, J.; Tanaka, C.; Tanaka, S.; Aoki, Y.; Yonemoto, J. Estimation of Daily Inhalation Rate of Preschool Children by Using Tri-Axial Accelerometer; *Epidemiology* **2008**, *19*, S139.
53. Lichtman, S.W.; Pisarska, K.; Berman, E.R.; Pestone, M.; Dowling, H.; Weisel, H.; Heshka, S.; Matthews, D.E.; Heymsfield, S.B. Discrepancy between Self-Reported and Actual Caloric Intake and Exercise in Obese Subjects; *New Engl. J. Med.* **1992**, *327*, 1893-1898.
54. *Methods for the Derivation of Inhalation Reference Concentrations and Application of Inhalation Dosimetry*; EPA/600/8-90/066F; U.S. Environmental Protection Agency: Washington, DC, 1994.
55. *Dietary Reference Intakes for Energy, Carbohydrate, Fiber, Fat, Fatty Acids, Cholesterol, Protein, and Amino Acids*; National Academy: Washington, DC, 2005; pp 880-935.

About the Authors

Dr. Junko Kawahara is a research fellow at the Research Center for Environmental Risk at the National Institute for Environmental Studies in Tsukuba, Ibaraki, Japan. Dr. Shigeo Tanaka is a project leader of the Project for Energy Metabolism on Health Promotion and Exercise Program of the National Institute of Health and Nutrition in Tokyo, Japan. Dr. Chiaki Tanaka is an associate professor in the Department of Health Science at the College of Health and Welfare of J.F. Oberlin University in Tokyo, Japan. Dr. Yuki Hikiyama is a researcher in the Faculty of Engineering at Chiba Institute of Technology in Narashino, Japan. Dr. Yasunobu Aoki is General Manager of the Environmental Health Risk Research Section at the Research Center for Environmental Risk at the National Institute for Environmental Studies. Dr. Junko Yonemoto is Deputy Director of the Research Center for Environmental Risk at the National Institute for Environmental Studies. Please address correspondence to: Junko Kawahara, Research Center for Environmental Risk, National Institute for Environmental Studies, 16-2 Onogawa, Tsukuba, Ibaraki 305-8506, Japan; phone: +81-29-850-2695; fax: +81-29-850-2920; e-mail: jkawa@nies.go.jp.

ファルマシア

別刷