



Fig. 2. Blue native polyacrylamide gel electrophoresis (BN-PAGE) analysis of liver respiratory chain enzymes showed markedly decreased protein expression of complex I, while the protein bands of complex II, III, and IV were comparable to the control (N) samples.

Hughes; yet, studies have not progressed because of technical difficulties. More recently, complex I deficiency was regarded as the most common energy generation disorder. The manifestations range from typical mitochondrial diseases, such as Leigh syndrome, to obscure conditions such as slow regression or intractable secretory diarrhea [4].

Complex II activity has been shown to be more labile than complex I when measuring respiratory chain enzymes in patients with a wide range of metabolic disorders, liver failure, or liver disease [5]. In the present case, only complex I activity was very low; this indicates primary complex I deficiency rather than a secondary effect of influenza A infection. Complex I includes seven mitochondrial DNA-encoded subunits and at least 39 nuclear-encoded subunits. In our case, no mutation was detected in the mitochondrial DNA (mtDNA). The detection rate for mutations in mitochondrial or nuclear DNA in complex I deficiency is as small as 20% [6,7].

In the present case, complex I was deficient only in the liver, not in fibroblasts. Mitochondrial respiratory complex disorders can show clinical and biochemical tissue specificity [2,4,6,8,10]. For this reason, it is difficult to diagnose by suspension cells or serum enzyme assays. The possible mechanisms of tissue specificity are tissue-specific subunits of complex I [9], the ratio between normal and mutant mtDNA in a specific tissue [7], and tissue differences in RNA processing [10]. To our knowledge, very few cases with liver-specific complex I deficiency have been reported [2,8]. These reported cases had chronic neurological symptoms such as epilepsy, hypotonia, or developmental regression, with the exception of one case that had severe cardiomyopathy in early

infancy [2]. There was one case without evidence of liver dysfunction [8]. Clinically there was no definite difference from usual Co I deficiency. One reason for the small number of cases is that the liver is not the prime diagnostic tissue. Respiratory chain complex deficiency is usually confirmed by tissue biopsy. Muscle is usually the prime diagnostic tissue, and cultured skin fibroblasts are also often analyzed [10]. False-negative diagnostic results may occur because the liver is not examined.

This case was determined to be complex I deficiency by BN-PAGE Western blotting and determination of enzyme activities. This is the first report of respiratory chain complex I deficiency in influenza encephalopathy. We suggest there may be many undiagnosed cases of this metabolic disorder. Here, we described a healthy child, who had never been suspected of having any disease, diagnosed with a metabolic disorder after acute encephalopathy with subsequent death. Future studies are needed to focus on the development of a method to detect this inborn metabolic disorder before onset.

References

- [1] Yao D, Mizuguchi H, Yamaguchi M, Yamada H, Chida J, Shikata K, et al. Thermal instability of compound variants of carnitine palmitoyltransferase II and impaired mitochondrial fuel utilization in influenza-associated encephalopathy. *Hum Mutat* 2008;29:718–27.
- [2] Kirby DM, Crawford M, Cleary MA, Dahl HM, Dennett X, Tourburn DR. Respiratory chain complex I deficiency. An underdiagnosed energy generation disorder. *Neurology* 1999;52:1255–64.
- [3] Van Coster R, Smet J, George E, De Meirleir L, Seneca S, Van Hove J, et al. Blue native polyacrylamide gel electrophoresis: a powerful tool of oxidative phosphorylation defects. *Pediatr Res* 2001;50:658–65.
- [4] Murayama K, Nagasaka H, Tsuruoka T, Omata Y, Horie H, Tregoning S, et al. Intractable secretory diarrhea in a Japanese boy with mitochondrial respiratory chain complex I deficiency. *Eur J Pediatr* 2009;168:297–302.
- [5] Hui J, Kirby DM, Thorburn DR, Boneh A. Decreased activities of mitochondrial respiratory chain complexes in non-mitochondrial respiratory chain diseases. *Dev Med Child Neurol* 2006;48:132–6.
- [6] Thorburn DR, Sugiana C, Salemi R, Kirby DM, Worgan L, Ohtake A, et al. Biochemical and molecular diagnosis of mitochondrial respiratory chain disorders. *Biochim Biophys Acta* 2004;1659:121–8.
- [7] Rötig A, Lebon S, Zinovieva E, Mollet J, Sarzi E, Bonnefont JP, et al. Molecular diagnostics of mitochondrial disorders. *Biochim Biophys Acta* 2004;1659:129–35.
- [8] Panetta J, Gibson K, Kirby DM, Thorburn DR, Boneh A. The importance of liver biopsy in the investigation of possible mitochondrial respiratory chain disease. *Neuropediatrics* 2005;36:256–9.
- [9] Clay VJ, Ragan CI. Evidence for the existence of tissue specific isoenzymes of mitochondrial NADH dehydrogenase. *Biochem Biophys Res Commun* 1988;157:1423–8.
- [10] Bindoff LA, Howell N, Poulton J, McCullough DA, Morten KJ, Lightowlers RN, et al. Abnormal RNA processing associated with a novel tRNA mutation in mitochondrial DNA. A potential disease mechanism. *J Biol Chem* 1993;268:19559–64.

