

Figure 2. Representative splicing mutations in our study. (A) C-to-T substitution at nucleotide position 6453 produced a 6-nucleic acid sequence (c.6452-6457) (underlined) identical to that of IVS52+1 to +6 (gtgagt) (underlined), and this was recognized as the splicing donor site. The following nucleotides of exon 52 from this point were deleted in the mutant allele. (B) The fifth nucleotide at the beginning of intron 11 was substituted from G to A (c.IVS11+5G>A). Resequencing of complementary DNA revealed that the latent splice donor site within exon 11 (underlined) was activated and became a new splice donor site and created the frame-shift mutation. (C) The last nucleotide of intron 34 was substituted from G to A (c.IVS34-1G>A). Resequencing FBN1 complementary DNA obtained from the peripheral blood demonstrated the splicing aberration, resulting in the deletion of 11 nucleotides of exon 35.

satisfactory, although their aortas dilated gradually later on. One patient met the revised Ghent criteria after genetic analysis of FBN1. Although further studies are warranted to clarify the properties and usefulness of the novel Ghent criteria, genetic testing is more important, although it is not mandatory. In such a setting, a high-throughput resequencing method such as the present microarray-based resequencing system can be a powerful tool for making an accurate diagnosis of MS.

Acknowledgments: We would like to express our sincere gratitude to the patients and their families for agreeing to

participate in this work and to all the doctors in the MS clinic for their valuable advice. We also thank Wu Zhenghong, Eri Kasukawa, Ryohei Kataoka, Keiko Hori, Yumiko Fujimaki, and Takako Kawamura for their excellent technical assistance.

- Dietz HC, Pyeritz RE. Mutations in the human gene for fibrillin-1 (FBN1) in the Marfan syndrome and related disorders. *Hum Mol Genet* 1995;4:1799-1809.
- De Paepe A, Devereux RB, Dietz HC, Hennekam RC, Pyeritz RE. Revised diagnostic criteria for the Marfan syndrome. Am J Med Genet 1996;62:417–426.

- Akutsu K, Morisaki H, Takeshita S, Ogino H, Higashi M, Okajima T, Yoshimuta T, Tsutsumi Y, Nonogi H, Morisaki T. Characteristics in phenotypic manifestations of genetically proved Marfan syndrome in a Japanese population. Am J Cardiol 2009;103:1146-1148.
- Loeys BL, Dietz HC, Braverman AC, Callewaert BL, De Backer J, Devereux RB, Hilhorst-Hofstee Y, Jondeau G, Faivre L, Milewicz DM, Pyeritz RE, Sponseller PD, Wordsworth P, De Paepe AM. The revised Ghent nosology for the Marfan syndrome. J Med Genet 2010; 47:476-485
- Collod-Béroud G, Le Bourdelles S, Ades L, Ala-Kokko L, Booms P, Boxer M, Child A, Comeglio P, De Paepe A, Hyland JC, Holman K, Kaitila I, Loeys B, Matyas G, Nuytinck L, Peltonen L, Rantamaki T, Robinson P, Steinmann B, Junien C, Béroud C, Boileau C. Update of the UMD-FBN1 mutation database and creation of an FBN1 polymorphism database. *Hum Mutat* 2003;22:199-208.
- 6. Faivre L, Collod-Beroud G, Loeys BL, Child A, Binquet C, Gautier E, Callewaert B, Arbustini E, Mayer K, Arslan-Kirchner M, Kiotsekoglou A, Comeglio P, Marziliano N, Dietz HC, Halliday D, Beroud C, Bonithon-Kopp C, Claustres M, Muti C, Plauchu H, Robinson PN, Adès LC, Biggin A, Benetts B, Brett M, Holman KJ, De Backer J, Coucke P, Francke U, De Paepe A, Jondeau G, Boileau C. Effect of mutation type and location on clinical outcome in 1,013 probands with Marfan syndrome or related phenotypes and FBN1 mutations: an international study. Am J Hum Genet 2007;81:454–466.
- Cutler DJ, Zwick ME, Carrasquillo MM, Yohn CT, Tobin KP, Kashuk C, Mathews DJ, Shah NA, Eichler EE, Warrington JA, Chakravarti A. High-throughput variation detection and genotyping using microarrays. Genome Res 2001;11:1913–1925.
- Warrington JA, Shah NA, Chen X, Janis M, Liu C, Kondapalli S, Reyes V, Savage MP, Zhang Z, Watts R, DeGuzman M, Berno A, Snyder J, Baid J. New developments in high throughput resequencing and variation detection using high-density microarrays. *Hum Mutat* 2002;19:402–409.
- Takahashi Y, Seki N, Ishiura H, Mitsui J, Matsukawa T, Kishino A, Onodera O, Aoki M, Shimozawa N, Murayama S, Itoyama Y, Suzuki Y, Sobue G, Nishizawa M, Goto J, Tsuji S. Development of a highthroughput microarray-based resequencing system for neurological disorders and its application to molecular genetics of amyotrophic lateral sclerosis. Arch Neurol 2008;65:1326–1332.
- Roman MJ, Devereux RB, Kramer-Fox R, O'Loughlin J. Two-dimensional echocardiographic aortic root dimensions in normal children and adults. Am J Cardiol 1989;64:507–512.
- 11. Sakai H, Visser R, Ikegawa S, Ito E, Numabe H, Watanabe Y, Mikami H, Kondoh T, Kitoh H, Sugiyama R, Okamoto N, Ogata T, Fodde R, Mizuno S, Takamura K, Egashira M, Sasaki N, Watanabe S, Nishimaki S, Takada F, Nagai T, Okada Y, Aoka Y, Yasuda K, Iwasa M, Kogaki S, Harada N, Mizuguchi T, Matsumoto N. Comprehensive genetic analysis of relevant four genes in 49 patients with Marfan syndrome or Marfan-related phenotypes. Am J Med Genet 2006;140: 1719–1725.
- Mizuguchi T, Collod-Beroud G, Akiyama T, Abifadel M, Harada N, Morisaki T, Allard D, Varret M, Claustres M, Morisaki H, Ihara M, Kinoshita A, Yoshiura K, Junien C, Kajii T, Jondeau G, Ohta T, Kishino T, Furukawa Y, Nakamura Y, Niikawa N, Boileau C, Matsu-

- moto N. Heterozygous TGFBR2 mutations in Marfan syndrome. *Nat Genet* 2004;36:855–860.
- Nijbroek G, Sood S, McIntosh I, Francomano CA, Bull E, Pereira L, Ramirez F, Pyeritz RE, Dietz HC. Fifteen novel FBN1 mutations causing marfan syndrome detected by heteroduplex analysis of genomic amplicons. Am J Hum Genet 1995;57:8–21.
- 14. Mátyás G, Alonso S, Patrignani A, Marti M, Arnold E, Magyar I, Henggeler C, Carrel T, Steinmann B, Berger W. Large genomic fibrillin-1 (FBN1) gene deletions provide evidence for true haploin-sufficiency in Marfan syndrome. *Hum Genet* 2007;122:23–32.
- Rommel K, Karck M, Haverich A, von Kodolitsch Y, Rybczynski M, Müller G, Singh KK, Schmidtke J, Arslan-Kirchner M. Identification of 29 novel and nine recurrent fibrillin-1 (FBN1) mutations and genotype-phenotype correlations in 76 patients with Marfan syndrome. Hum Mutat 2005;26:529-539.
- Liu WO, Oefner PJ, Qian C, Odom RS, Francke U. Denaturing HPLC-identified novel FBN1 mutations, polymorphisms, and sequence variants in Marfan syndrome and related connective tissue disorders. Genet Test 1997;1:237–242.
- Pepe G, Giusti B, Evangelisti L, Porciani MC, Brunelli T, Giurlani L, Attanasio M, Fattori R, Bagni C, Comeglio P, Abbate R, Gensini GF. Fibrillin-1 (FBN1) gene frameshift mutations in Marfan patients: genotype-phenotype correlation. Clin Genet 2001;59:444-450.
- Loeys B, De Backer J, Van Acker P, Wettinck K, Pals G, Nuytinck L, Coucke P, De Paepe A. Comprehensive molecular screening of the FBN1 gene favors locus homogeneity of classical Marfan syndrome. Hum Mutat 2004;24:140-146.
- Loeys BL, Schwarze U, Holm T, Callewaert BL, Thomas GH, Pannu H, De Backer JF, Oswald GL, Symoens S, Monouvrier S, Roberts AE, Faravelli F, Greco MA, Pyeritz RE, Milewicz DM, Coucke PJ, Cameron DE, Braverman AC, Byers PH, De Paepe AM, Dietz HC. Aneurysm syndromes caused by mutations in the TGFb receptor. N Engl J Med 2006;355:788-798.
- 20. Guo DC, Pannu H, Tran-Fadulu V, Papke CL, Yu RK, Avidan N, Bourgeois S, Estrera AL, Safi HJ, Sparks E, Amor D, Ades L, McConnell V, Willoughby CE, Abuelo D, Willing M, Lewis RA, Kim DH, Scherer S, Tung PP, Ahn C, Buja LM, Raman CS, Shete SS, Milewicz DM. Mutations in smooth muscle alpha-actin (ACTA2) lead to thoracic aortic aneurysms and dissections. Nat Genet 2007;39: 1488-1493.
- Zhu L, Vranckx R, Khau Van Kien P, Lalande A, Boisset N, Mathieu F, Wegman M, Glancy L, Gasc JM, Brunotte F, Bruneval P, Wolf JE, Michel JB, Jeunemaitre X. Mutations in myosin heavy chain 11 cause a syndrome associating thoracic aortic aneurysm/aortic dissection and patent ductus arteriosus. *Nat Genet* 2006;38:343–349.
- 22. van de Laar IM, Oldenburg RA, Pals G, Roos-Hesselink JW, de Graaf BM, Verhagen JM, Hoedemaekers YM, Willemsen R, Severijnen LA, Venselaar H, Vriend G, Pattynama PM, Collée M, Majoor-Krakauer D, Poldermans D, Frohn-Mulder IM, Micha D, Timmermans J, Hilhorst-Hofstee Y, Bierma-Zeinstra SM, Willems PJ, Kros JM, Oei EH, Oostra BA. Wessels MW, Bertoli-Avella AM. Mutations in SMAD3 cause a syndromic form of aortic aneurysms and dissections with early-onset osteoarthritis. Nat Genet 2011;43:121–126.

Expert Opinion

- 1. Introduction
- 2. Reagents
- 3. The effects on disease models
- 4. Expert opinion

Novel IκB kinase inhibitors for treatment of nuclear factor-κB-related diseases

Jun-ichi Suzuki[†], Masahito Ogawa, Susumu Muto, Akiko Itai, Mitsuaki Isobe, Yasunobu Hirata & Ryozo Nagai

[†]University of Tokyo, Graduate School of Medicine, Department of Advanced Clinical Science and Therapeutics, Tokyo, Japan

Introduction: NF-κB is a key regulator of inflammation and immunity in cancer development. The IκB kinase (IKK) is a multisubunit complex containing catalytic subunits termed IKK- α , - β and - γ . It is well known that many pro-inflammatory stimuli require the IKK- β subunit for NF-κB activation.

Areas covered: NF-κB affects the progression of inflammation-related diseases, such as myocardial ischemia, bronchial asthma, arthritis, cancer and other diseases. We review the characteristics and effects of these inhibitors on inflammatory and other diseases.

Expert opinion: Various synthesized IKK inhibitors have been developed and they will be used clinically in the near future.

Keywords: chemical compounds, inflammation, IκB kinase, NF-κB

Expert Opin. Investig. Drugs (2011) 20(3):395-405

1. Introduction

NF-KB is known to be a key factor in the regulation of inflammation [1] and immunity in cancer development [2]. The IKB kinase (IKK) is a multisubunit complex containing catalytic subunits termed IKK-α, -β and -γ [3]. To date, two major NF-KB activation pathways have been elicited based on the ligand interaction with surface receptors. Canonical signaling depends on IKK-γ and -β and induces the transcription of genes that regulate inflammation and cell survival. In contrast, non-canonical NF-KB activation is mostly involved in the regulation of B-cell development [4]. Although both pathways can affect development of inflammation, most of our knowledge relates to the canonical pathway. This pathway is triggered by infections and pro-inflammatory cytokines which activate IKK complex. This complex is composed of two catalytic subunits, IKK-α and -β, and a regulatory subunit, IKK-γ (NF-κB essential modulator; NEMO). The IKK complex phosphorylates NF-KB-bound IKBs, thereby targeting them for proteasomal degradation and liberating NF-κB dimers that are composed of REL-A (known as p65), REL and p50 subunits to enter the nucleus and mediate transcription of target genes. This reaction mostly depends on the catalytic subunit IKK-β, which carries out IκB phosphorylation. The non-canonical pathway involves the upstream kinase NF-KB-inducing kinase leading to the phosphorylation and processing of p100 in response to certain members of the TNF family. The two pathways switch on different gene sets and, therefore, mediate different immune functions. The contribution of the canonical pathway to acute inflammation and cell-survival mechanisms is well accepted, and sustained NF-κB activation in various malignancies has been described. Owing to the variety of target genes of the canonical pathway, which include those encoding mediators of inflammation, cytokines, chemokines, proteases and inhibitors of apoptosis (Figure 1), it has been proposed that canonical NF-KB activation might link inflammation to tumor promotion and progression [5].



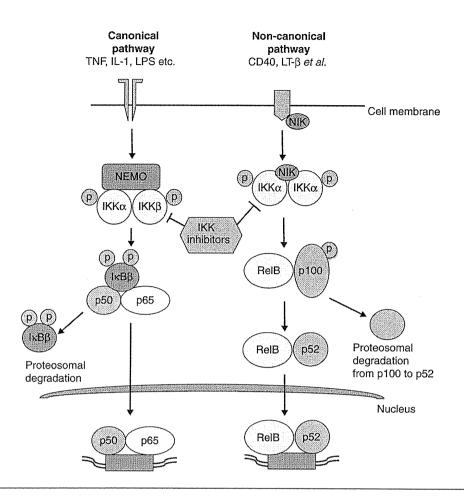


Figure 1. Canonical and non-canonical pathways of NF-KB.

Gene-targeting experiments revealed that many proinflammatory stimuli required IKK- β subunit for NF- κ B activation [6]. Thus, IKK-deficient mice died in the embryonic or perinatal periods (IKK- α^{-l-} , - β^{-l-} or - γ^{-l-}) demonstrating their critical role [7,8]. Dominant-negative IKK- β blocks NF- κ B-dependent transcription, while IKK- α plays a role only in response to certain stimuli and in limited cells. Recently, novel synthesized chemical compounds that act as IKK inhibitors have been developed. They are the phosphorylation inhibitors of I κ B that act via inhibition of IKK- α and/or - β . To clarify the effects of the inhibitors, we review previous articles on inflammatory diseases.

2. Reagents

IMD-0354 (N-(3,5-bis-trifluoromethyl-phenyl)-5-chloro-2-hydroxy-benzamide) [9,10] and IMD-0560 (N-(2,5-bis-trifluoromethyl-phenyl)-5-bromo-2-hydroxy-benzamide) [11] were developed as novel IKK inhibitors. They are the phosphorylation inhibitors of IkB that act via inhibition of IKK- β . IMD-0354 (molecular mass, 384.1, Figure 2A) was molecular-designed, synthesized and provided by the Institute of Medicinal Molecular Design, Inc. (Tokyo, Japan). The 3D

structure of a kinase domain of IKK-B was constructed by homology modeling with protein kinase A as a template. The structure of active IKK-β was estimated by referring to a model of IKK regulation. The molecular structure of IMD-0354 was designed by analyzing a binding mode of aspirin to IKK-β. To investigate the characteristics of IMD-0354, we performed a NF-κB-IKK-β reporter assay with the constitutively active mutant IKK-β. IMD-0354 inhibited the activated expression of NF-kB in a dosedependent manner in HepG2 cells that were transfected with pFLAG-CMV-IKKh (S177E/S181E) vector. IMD-0354 also decreased the levels of cytosolic phospho-IkBa in TNF-a stimulated cardiomyocytes in a dose-dependent manner. The kinetics of NF-kB translocation was consistent with the degree of phosphorylation of cytosolic IKBa. The translocation was blocked markedly by treatment with IMD-0354. TNF-α-induced production of IL-1α and monocyte chemoattractant protein-1 (MCP-1) from cultured cardiomyocytes were reduced significantly by IMD-0354. Taken together, IMD-0354 inhibits IKK-β, resulting in the blockade of IκBα phosphorylation.

Another IKK inhibitor, IMD-0560 (molecular mass 428.1), was also synthesized and provided by the Institute of Medical

Figure 2. Chemical structures of the chemical compounds A, IMD-0354; B, BMS-345541; C, PS-1145; D, SC-514; E, ACHP and F, Bay 65 – 1942.

CN

 NH_2

Molecular Design using the same methodology of IMD-0354. IMD-0560 inhibited the activated expression of NF- κ B in HEK293T cells transfected with the p-FLAG-CMV-IKK- β (S177E/S181E) vector in a dose-dependent manner. Further, pretreatment with IMD-0560 dose-dependently suppressed the DNA binding activity of NF- κ B. IMD-0560 also suppressed the nuclear translocation of NF- κ B and phosphorylation of I κ B α induced by TNF- α in fibroblast-like synoviocytes (FLS). Further, this compound suppressed the production of inflammatory cytokines and chemokines; it also inhibited the proliferation of FLS without showing cellular toxicity [11,12].

BMS-345541, 4(2'-aminoethyl)amino-1,8-dimethylimidazo(1,2- α) quinoxaline, was reported as a highly selective inhibitor of IKK that inhibits NF- κ B-dependent transcription of pro-inflammatory cytokines (Figure 2B). BMS-345541 was identified as a selective inhibitor of the catalytic subunits of IKK (IKK- β IC₅₀ = 0.3 μ M, IKK- α IC₅₀ = 4 μ M). This inhibitor appears to bind to an unidentified allosteric-binding site of the IKK catalytic subunits. The compound failed to inhibit a panel of 15 other kinases and selectively inhibited the stimulated phosphorylation of I κ B in cells. It also failed to affect c-Jun and STAT3 phosphorylation, as well as MAPK-2 activation in cells. The compound has good pharmacokinetic

NH HCI

characteristics (oral bioavailability 100%, intravenous half-life 2.2 h), which makes it particularly well suited for use in investigating the utility of IKK inhibitors in disease models [13].

PS-1145, N-(6-chloro-9H-β-carbolin-8-yl) nicotinamide, was also reported (Figure 2C) [14]. The compound PS-1145 was tested for its ability to block the phosphorylation of IκΒα in HeLa cells following TNF-α stimulation. Immunoblot analysis showed a dose-dependent inhibition of phosphorylated IκΒα. It then evaluated the effects of the compound on NF-κB activation by measuring DNA binding activity after TNF-α treatment in the same HeLa cells. EMSA showed a dose-dependent inhibition of NF-κB activation by the compound. Consistent with the inhibition of NF-κB activation, this compound also blocks the transcription of intracellular adhesion molecule (ICAM)-1 in HUVEC primary cultures [15]. PS-1145 inhibited TNF-α production with an IC50 of 4.7 μM/l [16].

SC-514, 5-(thien-3-yl)-3-aminothiophene-2-carboxamide, was identified as another selective IKK inhibitor (Figure 2D). This compound does not inhibit other IKK isoforms or other serine-threonine and tyrosine kinases. SC-514 inhibits the native IKK complex or recombinant human IKK- α /- β heterodimer and IKK- β homodimer. IKK- β inhibition by SC-514 is selective, reversible and competitive with ATP. SC-514 has several interesting effects; it does not inhibit the phosphorylation and activation of the IKK complex, and delays but does not completely block IkB α phosphorylation and degradation. Thus, the effect of SC-514 on cytokine gene expression is a combination of inhibiting IkB α phosphorylation/degradation affecting NF-kB nuclear import/export, as well as the phosphorylation and transactivation of p65 [17].

ACHP, 2-amino-6-[2-(cyclopropylmethoxy)-6-hydroxyphenyl] -4-piperidin-4-yl-nicotinonitrile, was developed and evaluated as a potent inhibitor for IKK- α and - β (Figure 2E). When massive screening was conducted, ACHP was found to have specific inhibitory action on IKK- α and - β . The IC₅₀ values for IKK- α and - β are 8.5 and 250 nmol/l, respectively, measured by *in vitro* kinase assays. ACHP also showed good aqueous solubility and cell permeability, thus, demonstrating high bioavailability in mice and rats [18,19].

Bay 65 – 1942, $\{7-[2-(cyclopropylmethoxy)-6-hydroxy-phenyl]-5-[(3S)-3-piperidinyl]-1,4-dihydro-2Hpyrido[2,3-d] [1,3]oxazin-2-one hydrochloride}, was composed as an IKK-<math>\beta$ inhibitor (Figure 2F). Through competitive inhibition of ATP at the IKK- β subunit, Bay 65 – 1942 prevented the phosphorylation of IkB α by the IKK complex [20].

AS602868 is an anilinopyrimidine derivative and ATP competitor, which inhibits IKK- β with an IC₅₀ = 62 nmol/l) (K_i = 20 nmol/l). The compound showed some inhibitory effect on JNK2. In a series of tests on different cell lines, AS602868 was shown to block phosphorylation of IKB and subsequent NF-KB activation [21].

NEMO binding domain (NBD) peptide was reported as an IKK inhibitor [22]. The specific molecular mechanisms of NEMO-IKK interactions involve a C-terminal hexapeptide core sequence present on both IKK- α and - β . Peptides corresponding to the IKK- β NBD have been found to disrupt the association of recombinant NEMO with recombinant IKK- α or - β in vitro. The cell-permeable versions of these peptides blocked NF- κ B activation in various cellular and animal models of inflammation [23].

It is well known that some natural products, such as polyphenols, have various effects on IKK inhibition. Pan *et al.* investigated the inhibition of IKK activity in lipopolysaccharide (LPS)-activated murine macrophages by various polyphenols including (-)-epigallocatechin-3-gallate (EGCG) and theaflavin-3,3'-digallate (TF-3). TF-3 inhibited IKK activity in activated macrophages more strongly than the other polyphenols. TF-3 strongly inhibited both IKK- α and - β activity and prevented the degradation of IKB α and IKB β in activated macrophage cells. The results suggest that the inhibition of IKK activity by TF-3 could occur by a direct effect on IKKs or on upstream events in the signal transduction pathway [24]. Yang *et al.* also revealed that EGCG was a potent IKK inhibitor because EGCG inhibited phosphorylation of IKB α and decreased IKK activity in cytosolic extracts [25].

3. The effects on disease models

3.1 Cardiovascular diseases

Myocardial ischemia reperfusion injury is related closely to inflammatory reactions. Morishita et al. revealed that inhibition of NF-κB using decoy oligodeoxynucleotides (ODNs) reduced the extent of myocardial infarction following reperfusion [26]. Thus, we investigated the efficacy of IKB phosphorylation blockade using IMD-0354 in a rat myocardial ischemia/reperfusion injury model [9]. Treatment with IMD-0354 resulted in a significant reduction of the infarction area: area at risk ratio and the preservation of fractional shortening. Histology showed that accumulation of polymorphonuclear neutrophils in the area at risk decreased significantly. In vitro study revealed that IMD-0354 inhibited nuclear translocation of NF- κ B induced by TNF- α in cultured cardiomyocytes. IMD-0354 caused a significant reduction of chemokine (MCP-1) production in a concentration-dependent manner compared with vehicletreated cells. Therefore, we concluded that inhibition of nuclear translocation of NF-κB by IMD-0354 could provide an effective approach to attenuate ischemia/reperfusion injury through chemokine suppression. Ventricular remodeling after myocardial infarction is also related to inflammatory reactions. Thus, we studied the effects of IMD-0354 [6] and IMD-0560 [11] in a rat myocardial infarction model. IMD-0354 or IMD-0560 administration reduced plasma brain natriuretic peptide levels after myocardial infarction. Either IMD-0354 or IMD-0560 treatment preserved left ventricular fractional shortening after infarction. Histology

showed that IMD-0354 significantly reduced myocardial macrophage infiltration and fibrosis. Western blot revealed that IMD-0354 suppressed MCP-1 expression in non-infarcted myocardial samples. *In gel* and *in situ* zymography clarified that either IMD-0354 or IMD-0560 treatment significantly suppressed MMP-9 activity in the non-infarcted myocardium. Therefore, we concluded that both IMD-0354 and IMD-0560 significantly affect the prevention of heart failure that is induced by ventricular remodeling after myocardial ischemia via altered MMP activation.

BMS-345541 was tested for its ability to suppress graft rejection in a murine heterotopic cardiac allograft model by Townsend *et al.* The compound did not prolong graft survival when administered at 50 mg/kg as a single agent. However, graft survival was significantly increased when it was administered with a suboptimal dose of cytotoxic T-lymphocyte antigen-4 immunoglobulin or cyclosporine A compared with either agent alone. They concluded that BMS-345541 may serve as a novel adjunctive therapy for the prevention of graft rejection [27].

SC-514 examined the effects using rat aortic smooth muscle cells. SC-514 treated cells significantly reduced iNOS induction, NF- κ B DNA binding and I κ B α loss. The results suggest that IKK- β plays a predominant, selective role in the regulation of NF- κ B-dependent induction of iNOS in smooth muscle cells [28]. SC-514 inhibited all forms of recombinant human IKK- α , including rhIKK- β homodimer, rhIKK- α /- β heterodimer, as well as the constitutively active form of rhIKK- β with comparable IC50 values in the 3 – 12 μ M range [29].

Moss *et al.* also reported that Bay 65 – 1942 can provide both acute and chronic cardioprotection and offers a clinically accessible target for preventing cardiac injury following ischemia reperfusion [30].

We demonstrated that tea catechins suppressed several cardiovascular diseases. We performed oral administration of catechins into murine and rat models of cardiac transplantation [31], myocarditis [32], myocardial ischemia [33] and atherosclerosis [34] to reveal the effects of catechins on the inflammation-induced ventricular and arterial remodeling. From our results, catechins are potent agents for the treatment and prevention of inflammation-related cardiovascular diseases, as they are critically involved in the suppression of pro-inflammatory signaling pathways.

3.2 Lung injury

NF-κB plays a key role in the progression of lung injury. Matsuda *et al.* revealed that NF-κB decoy ODNs prevented acute lung injury in mice [35]. Thus, we examined the effects of IMD-0354 to attenuate bleomycin-induced pulmonary fibrosis in mice [36]. IMD-0354 reduced the collagen content and fibrotic scores in the mice that received bleomycin. The bronchoalveolar lavage demonstrated that the proportions of neutrophils and lymphocytes decreased in mice treated with IMD-0354. The results suggested that IMD-0354 might be

useful to ameliorate inflammation in the lungs induced by chemical injury.

BMS-345541 is also known to affect acute lung injury. Everhart et al. administered BMS-345541 to determine whether intervention in the NF-kB pathway could prevent progression of lung injury in the LPS pump model. They revealed that treatment with BMS-345541 reduced lung NF-κB activation, concentration of pro-inflammatory cytokines and chemokines in lung lavage, neutrophil influx and lung edema. Therefore, they concluded that sustained NF-κB activation correlates with severity of lung injury and that BMS-345541 is beneficial to suppress lung inflammation [37]. PS-1145 also tested the effects using human ASM cells and pulmonary epithelial cells in vitro. As observed in human ASM cells, PS-1145 reduced expression of several adhesion molecules, cytokines and chemokines [38]. Similarly, PS-1145 reduced NF-KB-dependent transcription induced by IL-1 β and TNF- α in primary pulmonary epithelial cells [39]. Chapoval et al. revealed the in vivo effects of PS-1145 using IL-13 transgenic mice. While IL-13 induced tissue inflammation, fibrosis and alveolar remodeling, PS-1145 inhibited lung inflammatory and structural cell apoptosis with suppression of caspase activation [40].

3.3 Arthritis

Rheumatoid arthritis (RA) is affected by NF- κ B activation. Tomita *et al.* showed that NF- κ B decoy ODN suppressed the severity of collagen-induced arthritis in rats [41]. Thus, we evaluated the effect of IMD-0560 on collagen type II-induced arthritis in mice [12]. In this investigation, IMD-0560 suppressed the nuclear translocation of NF- κ B and phosphorylation of I κ B induced by TNF- α . In addition, this compound suppressed the production of inflammatory cytokines, including IL-6 and -8. IMD-0560 was effective against collagen-induced arthritis in mice via suppression of pro-inflammatory cytokines. Thus, we concluded that IMD-0560 could be a new therapeutic agent for RA.

BMS-345541 intensively evaluated the effects on RA. McIntyre *et al.* revealed that BMS-345541 is efficacious against collagen-induced arthritis in mice. BMS-345541 reduced the incidence of disease, inhibiting clinical signs of disease. Histological evaluation of the joints showed that BMS-345541 blocked inflammation and joint destruction. Transcription levels of IL-1 in the joints were also inhibited in the mice that received BMS-345541 [42]. Pattoli *et al.* also examined whether BMS-345541 directly inhibits cytokine-induced metalloproteinase expression and cartilage degradation. BMS-345541 inhibited IL-1-dependent expression of MMP-1, -3 and -13 in chondrosarcoma cells. Thus, BMS-345541 blocks collagen degradation through suppression of metalloproteinase expression [43].

Jimi et al. also revealed that the NBD peptide inhibited RANKL-stimulated NF-KB activation and osteoclastogenesis both in vitro and in vivo. This peptide significantly reduced the severity of collagen-induced arthritis in mice by reducing

levels of TNF- α and IL-1 β , abrogating joint swelling and reducing destruction of bone and cartilage [44].

3.4 Bronchial asthma

It is well known that NF-κB plays a critical role in induction of allergic airway inflammation. Desmet *et al.* revealed that NF-κB inhibition using decoy ODN was associated with strong attenuation of allergic lung inflammation, airway hyper-responsiveness [45]. Thus, we generated ovalbuminsensitized mice which had allergic airway inflammation and hyper-responsiveness. Administration of IMD-0354 ameliorated airway hyper-responsiveness and reduced the numbers of bronchial eosinophils in the mice. The total numbers of bronchial eosinophils and IgE production were reduced by treatment with IMD-0354 [46]. Thus, IMD-0354 has therapeutic potential for bronchial asthma.

BMS-345541 inhibited TNF-α-induced expression of IL-6, -8 and eotaxin dose-dependently in the airway smooth muscle (ASM) cells as Keslacy *et al.* revealed [47]. Goto *et al.* also investigated the effect of BMS-345541 using human ASM cells. They demonstrated that treatments with TNF-α and IL-13 induced a translocation of NF-κB to nuclei in ASM cells. However, co-incubation with BMS-345541 markedly inhibited the translocation of NF-κB [48].

PS-1145 reduced the expression of inflammatory factors including adhesion molecules, cytokines and chemokines on ASM cells, suggesting that the IKK inhibitor may be of considerable benefit in inflammatory airways diseases, particularly in severe asthma as Catley *et al.* reported [38].

Bay 65 – 1942 inhibited cockroach allergen-induced airway inflammation and hyper-reactivity in mice. It also efficiently abrogated leukocyte trafficking induced by carrageenan in mice or by ovalbumin in a rat model of airway inflammation [49].

3.5 Skin disorders

NF-KB activation on disease severity in allergic disorders is well known. Nakamura et al. showed that ointment containing NF-KB decoy ODN prevented atopic dermatitis in a mouse model [50]. Thus, we examined the relevance of IMD-0354 for atopic dermatitis by its topical application [51]. To investigate the in vivo efficacy, IMD-0354 ointment was applied to mice with severe dermatitis. Histological examinations revealed that the hyperplasia of keratinocytes and infiltration of inflammatory cells were significantly reduced in the skin of IMD-0354-treated mice. IMD-0354 suppressed the proliferation of various immunocompetent cells, IgE production from splenic B cells and IgE-mediated activation of mast cells. Therefore, we concluded that IMD-0354 might provide an alternative therapeutic strategy for the treatment of atopic dermatitis. So far, the effects of other IKK inhibitors on atopic dermatitis have not yet been reported; further investigation in this area is needed. di Meglio et al. showed that the NBD peptide significantly inhibited edema formation and cellular infiltration in inflamed mouse paws.

This anti-inflammatory activity was most likely due to inhibition of expression of pro-inflammatory mediators, such as TNF- α and COX-2, in inflamed tissues [52].

3.6 Malignant diseases

Development and progression of cancers, such as lymphoma and leukemia, and some epithelial cancers are known to be regulated by constitutive NF-KB activity [53,54]. Thus, inhibition of NF-KB may offer promise as a therapeutic approach for the treatment of tumors via manipulation of desired target genes. Kawamura et al. reported that NF-KB decoy ODN inhibited hepatic metastasis of reticulosarcoma in mice through a decrease in transactivation of important NF-KBdriven genes [55]. Regarding the contribution of NF-KB in carcinogenesis, IKK inhibition might have a therapeutic potential against cancers. We have reported that IMD-0354 suppressed the growth of human breast cancer cells by arresting cell cycles and inducing apoptosis. In the cells incubated with IMD-0354, cell cycle was arrested at the G0-G1 phase and apoptotic cells were increased. The expression of some cell cycle regulatory molecules and antiapoptotic molecules was suppressed in cells treated with IMD-0354. Daily administration of IMD-0354 inhibited tumor expansion in immunodeficient mice into which cancer cells were transplanted. We concluded that inhibition of NF-KB activity using IMD-0354 might have a therapeutic role in the treatment of human breast cancers.

BMS-345541 investigated the effects on several malignant diseases, such as melanoma, lymphoma, neuroblastoma and others [56-59]. Yang et al. revealed that BMS-345541 treatment resulted in the reduction of NF-KB activity, chemokine secretion by cultured melanoma cells and melanoma cell survival. The effect of BMS-345541 on tumor cell growth was through mitochondria-mediated apoptosis based on the reduced ratio of Bcl-2 per Bax. Thus, the mechanisms of antitumor effect of BMS-345541 are downregulation of IKK activity that results in mitochondria-mediated apoptosis of tumor cells because the programmed cell death is highly regulated by NF-κB signaling. Therefore, IKK may serve as a potential target for melanoma therapy [56]. Roué et al. reported that BMS-345541 decreased cellular-FLIP expression and allowed mantle cell lymphoma cells to undergo the TNF-related apoptosis-inducing ligand (TRAIL)-mediated apoptosis. They concluded that the combination of TRAIL stimulation and IKK inhibition as a new approach to MCL therapy [57]. Ammann et al. also revealed that BMS-345541 significantly enhances TRAIL-induced apoptosis, pointing to an antiapoptotic function of NF-KB in TRAIL-mediated apoptosis in neuroblastoma cells [58].

PS-1145 also tested the effects on malignant diseases, such as myeloma, lymphoma, prostate cancer, pancreatic cancer, breast cancer and others [60-69]. In myeloma, Hideshima et al. revealed that PS-1145 blocked TNF- α -induced NF- κ B activation in the tumor cells through inhibition of I κ B phosphorylation and degradation of I κ B α , respectively.

Moreover, PS-1145 blocks the protective effect of IL-6 against apoptosis. TNF-α-induced ICAM-1 expression on myeloma cells was also inhibited by PS-1145. Moreover, PS-1145 inhibits both IL-6 secretion from bone marrow stromal cells (BMSCs) triggered by multiple myeloma cell adhesion and proliferation of myeloma cells adherent to BMSCs. They also clarified the pathophysiology IKK inhibition in myeloma cells using a JNK-specific inhibitor SP600125. PS-1145 inhibits SP600125-induced NF-κB activation and blocks the protective effect of SP600125 against apoptosis [60,61]. Akiyama et al. revealed that PS-1145 blocks telomerase activity [62] and cell migration [63] in the myeloma cells. In solid tumor cells, Yemelyanov et al. found that PS1145 induced apoptosis and inhibited cell proliferation in prostate cancer cells. In addition, they found that incubation with PS1145 inhibited the invasion activity of highly invasive prostate cancer cells in an invasion chamber assay [66].

Bay 65 – 1942 induced growth suppression and death in ells of imatinib- or dasatinib-resistant forms of chronic myelogenous leukemia as Duncan *et al.* showed [70]. Lounnas *et al.* revealed that a solid IKK inhibitor AS602868 had a promising new therapeutic potential for the treatment of imatinib-resistant chronic myeloid leukemia patients. Because the mutation escapes all currently used Bcr-Abl inhibitors, it is likely to become a major clinical problem as it is associated with a poor clinical outcome [71]. Other IKK inhibitors, such as SC-514 [72] and ACHP [73,74], also have antitumor effects. Because Bednarski *et al.* revealed that IKK plays a critical role in NF-κB-mediated chemoresistance in response to doxorubicin [75], IKK inhibition may serve as a potential effect in combinational strategies to improve chemotherapeutic response.

3.7 Liver diseases

Because Ogushi *et al.* showed that NF-κB decoy ODN prevented fatal liver failure in a murine model [76], IKK inhibitors may prevent various liver diseases. Beraza *et al.* showed that AS602868 efficiently prevented liver steatosis and inflammation and improved antioxidant response. All the effects contributed to attenuation of the non-alcoholic-steatohepatitis progression, as evidenced by lower hepatocyte apoptosis and early stages of liver fibrosis [77].

3.8 Neurological diseases

Dasgupta *et al.* showed that the NBD peptides are antineuroinflammatory and that NBD peptides may have a therapeutic effect in neuroinflammatory disorders such as MS [78]. Acharyya *et al.* demonstrated that a specific pharmacological inhibition of IKK resulted in improved pathology and muscle function in *mdx* mice, which is a model of Duchenne muscular dystrophy [79].

3.9 The potential negative effects of IKK inhibitors

Many papers have reported that IKK inhibition has some potential negative effects. It is well known that NF-κB plays

an important role in immunity to infection. Genetic studies using animal models demonstrated the critical role of NF-kB in host defenses against pathogens. Three human primary immunodeficiencies associated with impaired NF-κB signaling were also reported [80]. Therefore, pharmacological IKK inhibition may damage defense systems against bacteria and fungi infection. The relationship between NF-kB and cancer development has also been reported. While the use of NSAIDs, which inhibit activation of NF-κB, reduced the incidence of cancers and lymphomas, some reports showed that NSAIDs might increase the risk of pancreatic cancer or non-Hodgkin's lymphoma. Thus, these relationships are very complicated because NF-κB activation can have either positive or negative, indirect, secondary effects on tumor development. NF-KB usually promotes cell survival that results in decreased cell proliferation, thereby its negative effect on tumor development [2]. Thus, IKK inhibitors may promote cancer development in some cases. Maeda et al. revealed that deletion of the gene encoding IKK-β in the cells resulted in a marked increase in tumor number, size, growth rate and aggressiveness [81]. Chen et al. also revealed that IKK inhibition prevented systemic inflammation but increased local injury following intestinal ischemia reperfusion [82]. These results showed the dual function of the IKK system, which is responsible for both tissue protection and systemic inflammation, and underscore the caution that should be exerted when using IKK inhibitors.

3.10 Clinical trials

IMD-1041, which is a prodrug of IMD-0354, specifically inhibits IKK-\$\beta\$ in vivo and in vitro [83]. Because this compound is an investigational drug, it is not yet on the market. To prove the effect of IMD-1041 on the treatment of chronic obstructive pulmonary disease (COPD), the Institute of Medicinal Molecular Design started the interventional, randomized, placebo-controlled and double-blind clinical trial entitled 'A Phase IIa, Proof of Concept Study to Evaluate the Reduction in Inflammatory Biomarkers and Assess Airway Function Following Administration of IMD-1041 in Patients With COPD' from 2009 (ClinicalTrials.gov Identifier: NCT00883584). The purpose of this study is to see if IMD-1041 has the ability to reduce inflammatory derived symptoms and airway remodeling by looking at changes in chemical levels in the blood and sputum. Sanofi-Aventis has also started a clinical trial using an IKK inhibitor (SAR113945) in patients with knee osteoarthritis (Clinical-Trials.gov Identifier: NCT01113333). Although these results have not yet been analyzed, potent IKK inhibitors will be available in the near future.

4. Expert opinion

We have reviewed the effects of novel synthesized IKK inhibitors on inflammatory diseases in this article. Because NF-κB plays a critical role in inflammation, IKK inhibition has the

Novel IKK inhibitors for treatment of NF-κB-related diseases

potential to prevent and treat the cardiovascular, pulmonary, allergic, malignant and other diseases.

To date, various synthesized IKK inhibitors, IMD-0354, IMD-0560, BMS-345541, PS-1145, SC-514, ACHP, Bay 65 - 1942, AS602868 and others, have been reported. However, direct comparison of the effects among the compounds on the same diseases has yet to be elucidated. For example, we have revealed that the IKK inhibitor IMD-0354 had significant effects when used to treat myocardial ischemia, pulmonary fibrosis, bronchial asthma, atopic dermatitis and breast cancer. However, while other IKK inhibitors demonstrated significant effects on liver, colon and neurological disorders, we have not yet elucidated the IMD-0354 effect. On the other hand, other synthesized IKK inhibitors (AS602868 and NDB peptides) have yet to be examined on cardiovascular diseases. Thus, we should perform further comparative analysis to validate the effects using the same experimental disease models.

Further, we have not yet compared the effects between the novel compounds and conservative products such as corticosteroids and NSAIDs. Corticosteroids are known to be potent anti-inflammatory agents and suppressors of cytokine production. The anti-inflammatory effects of corticosteroids are mediated through inhibition of NF-κB activation. Although corticosteroids have not proven to be beneficial in clinical studies on patients with some diseases, specific inhibition of IKK may have superior effects on these diseases compared to corticosteroids. Aspirin is one of the most commonly used NSAIDs because of its ability to inhibit COX activity. It has been reported that NF-κB activation and its associated gene expressions were suppressed by the aspirin supplementation

through the inhibition of phosphorylation and degradation of $I\kappa B\alpha$ via the IKK pathway. Although corticosteroids and NSAIDs are known to have adverse effects, they have been broadly used in clinical settings for a long time. Because we need specific IKK inhibitors without detrimental effects in clinical settings, we have to clarify the superior effects of the new compounds in comparison to the other conservative compounds, including corticosteroids and NSAIDs.

Finally, we have to evaluate the adverse effects of the new compounds. Although the deletion of the gene encoding IKK-β in the cells resulted in a marked increase of carcinogenesis, there has been no report to demonstrate the adverse results by IKK inhibitors in vivo. Because the carcinogenesis should be evaluated using several factors, such as tumor number, size, growth rate, invasion to other tissues and remote metastasis, the IKK inhibitory effects against malignant diseases should be evaluated using several experimental models. It was also reported that IKK inhibition increased local tissue injury following intestinal ischemia reperfusion. However, there has been no report to demonstrate similar results of ischemia reperfusion injury in other solid organ systems. These adverse effects show the complexity of the IKK system, which is responsible for both local and systemic immunity. Therefore, further investigation is needed to expand the strategy of specific IKK inhibition for clinical applications.

Declaration of interest

The authors declare no conflict of interest and have received no payment in preparation of this manuscript.

Bibliography

- Ghosh S, Hayden MS. New regulators of NF-kappaB in inflammation.
 Nat Rev Immunol 2008;8:837-4
- Karin M, Greten FR. NF-kappaB: linking inflammation and immunity to cancer development and progression. Nat Rev Immunol 2005;5:749-59
- Karin M, Ben-Neriah Y. Phosphorylation meets ubiquitination: the control of NF-kappaB activity. Annu Rev Immunol 2000;18:621-63
- Bollrath J, Greten FR. IKK/NF-kappaB and STAT3 pathways: central signalling hubs in inflammation-mediated tumour promotion and metastasis. EMBO Rep 2009:10:1314-19
- Oeckinghaus A, Ghosh S. The NF-kappaB family of transcription factors and its regulation. Cold Spring Harb Perspect Biol 2009;1:a000034
- Ghosh S, Karin M. Missing pieces in the NF-kappaB puzzle. Cell 2002;109:S81-96
- Gerondakis S, Grossmann M, Nakamura Y, et al. Genetic approaches in mice to understand Rel/NF-kappaB and IkappaB function: transgenics and knockouts. Oncogene 1999;18:6888-95
- Caamano J, Hunter CA. NF-kappaB family of transcription factors: central regulators of innate and adaptive immune functions. Clin Microbiol Rev 2002;15:414-29
- Onai Y, Suzuki J, Kakuta T, et al.
 Inhibition of IkappaB phosphorylation in cardiomyocytes attenuates myocardial ischemia/reperfusion injury.
 Cardiovasc Res 2004;63:51-9
- Onai Y, Suzuki J, Maejima Y, et al. Inhibition of NF-kappaB improves left ventricular remodeling and cardiac dysfunction after myocardial infarction. Am J Physiol Heart Circ Physiol 2007;292:H530-8
- Wakatsuki S, Suzuki J, Ogawa M, et al. A novel IKK inhibitor suppresses heart failure and chronic remodeling after myocardial ischemia via MMP alteration. Expert Opin Ther Targets 2008;12:1469-76
- 12. Okazaki Y, Sawada T, Nagatani K, et al. Effect of nuclear factor-kappaB inhibition on rheumatoid fibroblast-like synoviocytes and collagen induced arthritis. J Rheumatol 2005;32:1440-7

- Burke JR, Pattoli MA, Gregor KR, et al. BMS-345541 is a highly selective inhibitor of IkappaB kinase that binds at an allosteric site of the enzyme and blocks NF-kappaB-dependent transcription in mice. J Biol Chem 2003;278:1450-6
- Hideshima T, Hayashi T, Chauhan D, et al. Biologic sequelae of c-Jun NH(2)-terminal kinase (JNK) activation in multiple myeloma cell lines. Oncogene 2003;22:8797-801
- Castro AC, Dang LC, Soucy F, et al. Novel IKK inhibitors: beta-carbolines. Bioorg Med Chem Lett 2003;13:2419-22
- 16. Lam LT, Davis RE, Pierce J, et al. Small molecule inhibitors of IkappaB kinase are selectively toxic for subgroups of diffuse large B-cell lymphoma defined by gene expression profiling. Clin Cancer Res 2005;12:28-40
- Kishore N, Sommers C, Mathialagan S, et al. A selective IKK-2 inhibitor blocks NF-kappaB-dependent gene expression in interleukin-1 beta-stimulated synovial fibroblasts. J Biol Chem 2003;278:32861-71
- Murata T, Shimada M, Sakakibara S, et al. Discovery of novel and selective IKK-beta serine-threonine protein kinase inhibitors. Part 1. Bioorg Med Chem Lett 2003;13:913-18
- Murata T, Shimada M, Sakakibara S, et al. Synthesis and structure-activity relationships of novel IKK-beta inhibitors. Part 3: orally active anti-inflammatory agents. Bioorg Med Chem Lett 2004;14:4019-22
- 20. Ziegelbauer K, Gantner F, Lukacs NW, et al. A selective novel low-molecular-weight inhibitor of IkappaB kinase-beta (IKK-beta) prevents pulmonary inflammation and shows broad anti-inflammatory activity. Br J Pharmacol 2005;145:178-92
- Frelin C, Imbert V, Griessinger E, et al. AS602868, a pharmacological inhibitor of IKK2, reveals the apoptotic potential of TNF-alpha in Jurkat leukemic cells. Oncogene 2003;22:8187-94
- 22. May MJ, D'Acquisto F, Madge LA, et al. Selective inhibition of NF-kappaB activation by a peptide that blocks the interaction of NEMO with the IkappaB

- kinase complex. Science 2000:289:1550-4
- Baima ET, Guzova JA, Mathialagan S, et al. Novel insights into the cellular mechanisms of the anti-inflammatory effects of NF-kappaB essential modulator binding domain peptides. J Biol Chem 2010;285:13498-506
- 24. Pan MH, Lin-Shiau SY, Ho CT, et al. Suppression of lipopolysaccharide-induced nuclear factor-kappaB activity by theaflavin-3,3'-digallate from black tea and other polyphenols through down-regulation of IkappaB kinase activity in macrophages.

 Biochem Pharmacol 2000;59:357-67
- 25. Yang F, Oz HS, Barve S, et al. The green tea polyphenol (-)-epigallocatechin-3-gallate blocks nuclear factor-kappaB activation by inhibiting IkappaB kinase activity in the intestinal epithelial cell line IEC-6. Mol Pharmacol 2001;60:528-33
- 26. Morishita R, Sugimoto T, Aoki M, et al. In vivo transfection of cis element 'decoy' against nuclear factor-kappaB binding site prevents myocardial infarction. Nat Med 1997;3:894-9
- 27. Townsend RM, Postelnek J, Susulic V, et al. A highly selective inhibitor of IkappaB kinase, BMS-345541, augments graft survival mediated by suboptimal immunosuppression in a murine model of cardiac graft rejection. Transplantation 2004;77:1090-4
- 28. Gomez AB, MacKenzie C, Paul A, et al. Selective inhibition of inhibitory kappaB kinase-beta abrogates induction of nitric oxide synthase in lipopolysaccharide-stimulated rat aortic smooth muscle cells. Br J Pharmacol 2005;146:217-25
- Kishore N, Sommers C, Mathialagan S, et al. A selective IKK-2 inhibitor blocks NF-kappaB-dependent gene expression in interleukin-1 beta-stimulated synovial fibroblasts. J Biol Chem 2003;278:32861-71
- Moss NC, Stansfield WE, Willis MS, et al. IKKbeta inhibition attenuates myocardial injury and dysfunction following acute ischemia-reperfusion injury. Am J Physiol Heart Circ Physiol 2007;293:H2248-53

Expert Opin. Investig. Drugs (2011) 20(3)

Novel IKK inhibitors for treatment of NF-kB-related diseases

- Suzuki J, Ogawa M, Sagesaka YM, et al.
 Tea catechins attenuate ventricular
 remodeling and graft arterial diseases in
 murine cardiac allografts. Cardiovasc Res
 2006;69:272-9
- Suzuki J, Ogawa M, Futamatsu H, et al.
 Tea catechins improve left ventricular
 dysfunction, suppress myocardial
 inflammation, fibrosis, and alter cytokine
 expression in rat autoimmune
 myocarditis. Eur J Heart Fail
 2007;9:152-9
- Suzuki J, Ogawa M, Maejima Y, et al.
 Tea catechins attenuate chronic
 ventricular remodeling after myocardial
 ischemia in rats. J Mol Cell Cardiol
 2007;42:432-40
- 34. Suzuki J, Ogawa M, Izawa A, et al. Dietary consumption of green tea catechins attenuate hyperlipidemia-induced atherosclerosis and systemic organ damage in mice. Acta Cardiologica 2005;60:271-6
- Matsuda N, Hattori Y, Jesmin S, et al. Nuclear factor-kappaB decoy oligodeoxynucleotides prevent acute lung injury in mice with cecal ligation and puncture-induced sepsis. Mol Pharmacol 2005;67:1018-25
- Inayama M, Nishioka Y, Azuma M, et al. A novel IkappaB kinase-beta inhibitor ameliorates bleomycin-induced pulmonary fibrosis in mice. Am J Respir Crit Care Med 2006;173:1016-22
- Everhart MB, Han W, Sherrill TP, et al. Duration and intensity of NF-kappaB activity determine the severity of endotoxin-induced acute lung injury. J Immunol 2006;176:4995-5005
- 38. Catley MC, Sukkar MB, Chung KF, et al. Validation of the anti-inflammatory properties of small-molecule IkappaB Kinase (IKK)-2 inhibitors by comparison with adenoviral-mediated delivery of dominant-negative IKK1 and IKK2 in human airways smooth muscle.

 Mol Pharmacol 2006;70:697-705
- Newton R, Holden NS, Catley MC, et al. Repression of inflammatory gene expression in human pulmonary epithelial cells by small-molecule IkappaB kinase inhibitors. J Pharmacol Exp Ther 2007;321:734-42
- Chapoval SP, Al-Garawi A, Lora JM, et al. Inhibition of NF-kappaB activation reduces the tissue effects of transgenic IL-13. J Immunol 2007;179:7030-41

- 41. Tomita T, Takeuchi E, Tomita N, et al. Suppressed severity of collagen-induced arthritis by in vivo transfection of nuclear factor kappaB decoy oligodeoxynucleotides as a gene therapy. Arthritis Rheum 1999;42:2532-42
- 42. McIntyre KW, Shuster DJ, Gillooly KM, et al. A highly selective inhibitor of IkappaB kinase, BMS-345541, blocks both joint inflammation and destruction in collagen-induced arthritis in mice. Arthritis Rheum 2003;48:2652-9
- 43. Pattoli MA, MacMaster JF, Gregor KR, et al. Collagen and aggrecan degradation is blocked in interleukin-1-treated cartilage explants by an inhibitor of IkappaB kinase through suppression of metalloproteinase expression. J Pharmacol Exp Ther 2005;315:382-8
- Jimi E, Aoki K, Saito H, et al. Selective inhibition of NF-kappaB blocks osteoclastogenesis and prevents inflammatory bone destruction in vivo. Nat Med 2004;10:617-24
- Desmet C, Gosset P, Pajak B, et al. Selective blockade of NF-kappaB activity in airway immune cells inhibits the effector phase of experimental asthma. J Immunol 2004;173:5766-75
- 46. Sugita A, Ogawa H, Azuma M, et al. Antiallergic and anti-inflammatory effects of a novel IkappaB kinase beta inhibitor, IMD-0354, in a mouse model of allergic inflammation. Int Arch Allergy Immunol 2008;148:186-98
- 47. Keslacy S, Tliba O, Baidouri H, et al. Inhibition of tumor necrosis factor-alpha-inducible inflammatory genes by interferon-gamma is associated with altered nuclear factor-kappaB transactivation and enhanced histone deacetylase activity. Mol Pharmacol 2007;71:609-18
- 48. Goto K, Chiba Y, Misawa M.
 IL-13 induces translocation of
 NF-kappaB in cultured human bronchial
 smooth muscle cells. Cytokine
 2009;46:96-9
- Ziegelbauer K, Gantner F, Lukacs NW, et al. A selective novel low-molecular-weight inhibitor of IkappaB kinase-beta (IKK-beta) prevents pulmonary inflammation and shows broad anti-inflammatory activity.
 Br J Pharmacol 2005;145:178-92
- Nakamura H, Aoki M, Tamai K, et al. Prevention and regression of atopic dermatitis by ointment containing

- NF-kB decoy oligodeoxynucleotides in NC/Nga atopic mouse model. Gene Ther 2002;9:1221-9
- Tanaka A, Muto S, Jung K, et al.
 Topical application with a new
 NF-kappaB inhibitor improves atopic dermatitis in NC/NgaTnd mice.
 J Invest Dermatol 2007;127:855-63
- di Meglio P, Ianaro A, Ghosh S.
 Amelioration of acute inflammation by systemic administration of a cell-permeable peptide inhibitor of NF-kappaB activation. Arthritis Rheum 2005;52:951-8
- 53. Tanaka A, Muto S, Konno M, et al.
 A new IkappaB kinase beta inhibitor
 prevents human breast cancer progression
 through negative regulation of cell cycle
 transition. Cancer Res 2006;66:419-26
- 54. Tanaka A, Konno M, Muto S, et al. A novel NF-kappaB inhibitor, IMD-0354, suppresses neoplastic proliferation of human mast cells with constitutively activated c-kit receptors. Blood 2005;105:2324-31
- 55. Kawamura I, Morishita R, Tsujimoto S, et al. Intravenous injection of oligodeoxynucleotides to the NF-kappaB binding site inhibits hepatic metastasis of M5076 reticulosarcoma in mice. Gene Ther 2001;8:905-12
- 56. Yang J, Amiri KI, Burke JR, et al. BMS-345541 targets inhibitor of kappaB kinase and induces apoptosis in melanoma: involvement of nuclear factor kappaB and mitochondria pathways. Clin Cancer Res 2006;12:950-60
- 57. Roue G, Perez-Galan P, Lopez-Guerra M, et al. Selective inhibition of IkappaB kinase sensitizes mantle cell lymphoma B cells to TRAIL by decreasing cellular FLIP level. J Immunol 2007;178:1923-30
- 58. Ammann JU, Haag C, Kasperczyk H, et al. Sensitization of neuroblastoma cells for TRAIL-induced apoptosis by NF-kappaB inhibition. Int J Cancer 2009;124:1301-11
- Katdare M, Efimova EV, Labay E, et al. Diverse TNFalpha-induced death pathways are enhanced by inhibition of NF-kappaB. Int J Oncol 2007;31:1519-28
- Hideshima T, Chauhan D,
 Richardson P, et al. NF-kappaB as a therapeutic target in multiple myeloma.
 J Biol Chem 2002;277:16639-47

- Hideshima T, Hayashi T, Chauhan D, et al. Biologic sequelae of c-Jun NH(2)-terminal kinase (JNK) activation in multiple myeloma cell lines. Oncogene 2003;22:8797-801
- Akiyama M, Hideshima T, Hayashi T, et al. Cytokines modulate telomerase activity in a human multiple myeloma cell line. Cancer Res 2002;62:3876-82
- 63. Tai YT, Podar K, Mitsiades N, et al. CD40 induces human multiple myeloma cell migration via phosphatidylinositol 3-kinase/AKT/NF-kappaB signaling. Blood 2003;101:2762-9
- 64. Roychowdhury S, Baiocchi RA, Vourganti S, et al. Selective efficacy of depsipeptide in a xenograft model of Epstein-Barr virus-positive lymphoproliferative disorder. J Natl Cancer Inst 2004;96:1447-57
- Lam LT, Davis RE, Pierce J, et al. Small molecule inhibitors of IkappaB kinase are selectively toxic for subgroups of diffuse large B-cell lymphoma defined by gene expression profiling. Clin Cancer Res 2005;11:28-40
- 66. Yemelyanov A, Gasparian A,
 Lindholm P, et al. Effects of IKK
 inhibitor PS1145 on NF-kappaB
 function, proliferation, apoptosis and
 invasion activity in prostate carcinoma
 cells. Oncogene 2006;25:387-98
- 67. Domingo-Domenech J, Oliva C,
 Rovira A, et al. Interleukin 6, a nuclear
 factor-kappaB target, predicts resistance
 to docetaxel in hormone-independent
 prostate cancer and nuclear
 factor-kappaB inhibition by PS-1145
 enhances docetaxel antitumor activity.
 Clin Cancer Res 2006;12:5578-86
- 68. Khanbolooki S, Nawrocki ST, Arumugam T, et al. Nuclear factor-kappaB maintains TRAIL resistance in human pancreatic cancer cells. Mol Cancer Ther 2006;5:2251-60
- 69. Singh S, Shi Q, Bailey ST, et al. Nuclear factor-kappaB activation: a molecular therapeutic target for estrogen receptor-negative and epidermal growth factor receptor family receptor-positive

- human breast cancer. Mol Cancer Ther 2007:6:1973-82
- Duncan EA, Goetz CA, Stein SJ, et al. IkappaB kinase beta inhibition induces cell death in Imatinib-resistant and T315I Dasatinib-resistant BCR-ABL+ cells. Mol Cancer Ther 2008;7:391-7
- Lounnas N, Frelin C, Gonthier N, et al. NF-kappaB inhibition triggers death of imatinib-sensitive and imatinib-resistant chronic myeloid leukemia cells including T315I Bcr-Abl mutants. Int J Cancer 2009;125:308-17
- 72. Choo MK, Sakurai H, Kim DH, et al. A ginseng saponin metabolite suppresses tumor necrosis factor-alpha-promoted metastasis by suppressing nuclear factor-kappaB signaling in murine colon cancer cells. Oncol Rep 2008:19:595-600
- Sanda T, Iida S, Ogura H, et al. Growth inhibition of multiple myeloma cells by a novel IkappaB kinase inhibitor. Clin Cancer Res 2005;11:1974-82
- Sanda T, Asamitsu K, Ogura H, et al. Induction of cell death in adult T-cell leukemia cells by a novel IkappaB kinase inhibitor. Leukemia 2006;20:590-8
- 75. Bednarski BK, Ding X, Coombe K, et al. Active roles for inhibitory kappaB kinases alpha and beta in nuclear factor-kappaB-mediated chemoresistance to doxorubicin. Mol Cancer Ther 2008;7:1827-35
- Ogushi I, Iimuro Y, Seki E, et al. Nuclear factor kappaB decoy oligodeoxynucleotides prevent endotoxin-induced fatal liver failure in a murine model. Hepatology 2003;38:335-44
- 77. Beraza N, Malato Y, Vander Borght S, et al. Pharmacological IKK2 inhibition blocks liver steatosis and initiation of non-alcoholic steatohepatitis. Gut 2008;57:655-63
- 78. Dasgupta S, Jana M, Zhou Y, et al.
 Antineuroinflammatory effect of
 NF-kappaB essential modifier-binding
 domain peptides in the adoptive transfer
 model of experimental allergic

- encephalomyelitis. J Immunol 2004;173:1344-54
- Acharyya S, Villalta SA, Bakkar N, et al. Interplay of IKK/NF-kappaB signaling in macrophages and myofibers promotes muscle degeneration in Duchenne muscular dystrophy. J Clin Invest 2007;117:889-901
- Puel A, Picard C, Ku CL, et al. Inherited disorders of NF-kappaB-mediated immunity in man. Curr Opin Immunol 2004:16:34-41
- 81. Maeda S, Kamata H, Luo JL, et al. IKKbeta couples hepatocyte death to cytokine-driven compensatory proliferation that promotes chemical hepatocarcinogenesis. Cell 2005;121:977-90
- 82. Chen LW, Egan L, Li ZW, et al.
 The two faces of IKK and NF-kappaB inhibition: prevention of systemic inflammation but increased local injury following intestinal ischemia-reperfusion.
 Nat Med 2003;9:575-81
- 83. Fukuda S, Horimai C, Harada K, et al.
 Aldosterone-induced kidney injury is
 mediated by NF-kappaB activation.
 Clin Exp Nephrol. 2010.
 [Epub ahead of print]

Affiliation

Jun-ichi Suzuki^{†1}, Masahito Ogawa¹, Susumu Muto², Akiko Itai², Mitsuaki Isobe³, Yasunobu Hirata¹ & Ryozo Nagai⁴ [†]Author for correspondence ¹University of Tokyo, Graduate School of Medicine, Department of Advanced Clinical Science and Therapeutics, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8655, Japan Tel: +81 3 5800 9116; Fax: +81 3 5800 9182; E-mail: junichisuzuki-circ@umin.ac.jp ²Institute of Medicinal Molecular Design, Inc. Tokyo, Japan ³Tokyo Medical and Dental University, Department of Cardiovascular Medicine, Tokyo, Japan ⁴University of Tokyo, Department of Cardiovascular Medicine, Tokyo, Japan

研究

マルファン症候群では歯周病は極めて高頻度に認められる

 青木美穂子
 今井
 靖
 藤田
 大司
 小川
 直美

 加藤
 昌義
 西村
 敬史
 鈴木
 淳一
 平田
 恭信

 永井
 良三

呼 吸 と 循 環 第59巻 第9号 別刷 2011年9月15日 発行

医学書院



マルファン症候群では歯周病は極めて 高頻度に認められる*

青木美穂子¹ 今井 靖 藤田 大司 小川 直美 加藤 昌義 西村 敬史 鈴木 淳一 平田 恭信 永井 良三

要旨

マルファン症候群は、骨格異常、眼異常、心血管異常など多くの器官に病変を引き起こす常染色体優性遺伝の全身性結合組織疾患である。以前より口腔内所見として、高口蓋、歯列不正などが知られている。近年、諸外国においてマルファン症候群と歯周病との関係が注目されてきており、日本人におけるマルファン症候群の実態調査として Ghent 基準陽性 20 名のマルファン症候群症例につき歯周病罹患状態を評価した。現在歯数は 27 歯とほぼ保たれていたが、歯周ボケットの深さ (PD) は 2.815±0.624 mm、PD 測定部位での出血の有無(BOP) は 11.567±8.394%、地域歯周疾患指数(CPI) は中等度・重度に該当するコード 3、4の症例が 15 名(75%) も認められた。以上よりマルファン症候群では、中等度から重度の歯周病が高頻度に認められマルファン症候群における歯周組織の脆弱性が示唆された。

キーワード マルファン症候群、歯周病、地域歯周疾患指数(CPI)

マルファン症候群は、1896年にパリの小児科医Antoine Marfan により初めて報告された常染色体優性遺伝性の疾患である¹⁾. 全身において骨格異常,限異常,心血管異常など多くの器官に病変を引き起こし、また、口腔においては高口蓋,歯列不正,歯の形態異常などがみられることが知られている^{2,3)}. 今日その診断には、Ghent の基準⁴⁾を採用することが一般的であり,骨格異常や限異常,心血管異常といった多彩な病態の表現型ごとに設定された大基準と小基準および家族歴や遺伝的要素を加味したものとなっている.

近年、諸外国においてマルファン症候群と歯周病との関係が注目されている⁵⁾.以前より国内ではマルファン症候群を有する顎変形症症例に対する外科処置の報告は散見されるものの、マルファン症候群の口腔内所見に関する報告は少なく、また骨格系に関する表現型は同じマルファン症候群であっても欧米人と日本人では相違点が少なくないことが知られている⁶⁾.そこで、今回マルファン症候群の口腔内の状態を把握する目的で歯周病罹患状態を調査し、またマルファン症

候群の表現型と歯周病所見との関係にも注目し検討したので、文献的考察を加えて報告する。

■ 対象と方法

東京大学マルファン症候群専門外来を受診し、Ghent 基準においてマルファン症候群と診断された症例で、本研究の主旨に同意が得られた患者 20 名(男性11 名、女性 9 名、平均年齢 35.7 歳)を対象とした.

患者には事前に研究の目的を十分に説明し、同意を 書面で確認後、口腔内診査を行った。本研究は、東京 大学医学部研究倫理審査委員会で承認を得た。

1. 歯周組織の評価

各対象者について現在歯数をはじめ以下の項目について歯周組織検査を実施した.

1) Probing Depth (PD)

歯周病の現在の進行度を表すため歯周ポケットの深さを測定した。カラーコードポケット探針(PCP-11, Hu-Friedy 社製)を用い、約 20g 前後の力で 1 点法にて測定した。被験者の 1 歯あたりの平均値を mm 単位で算出した。

0452-3458/11/¥500/論文/JCOPY

^{*} The High Prevalence of Periodontitis in Patients with Marfan Syndrome(2011年6月6日受付)

¹ 東京大学医学部附属病循環器内科(〒113-8655 東京都文京区本郷 7-3-1) Mieko Aoki, Yasushi Imai, Daishi Fujita, Naomi Ogawa, Masayoshi Kato, Hiroshi Nishimura, Jun-ichi Suzuki, Yasunobu Hirata, Ryozo Nagai: Department of Cardiovascular Medicine, University of Tokyo Hospital

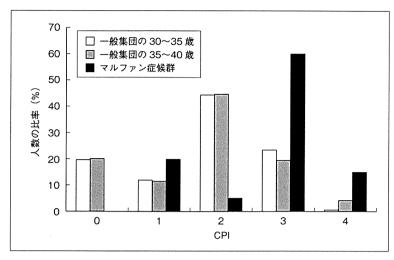


図1 本研究対象者と同年代男性およ び女性との間での CPI の比較

2) Bleeding on Probing (BOP)

歯周ポケット内の現在の炎症を調べるため PD 測定 部位での出血の有無を測定し、被験者の全被験歯に対する検出率(%)を算出した.

3) 歯の動揺度

Miller ら⁷の方法により0度~3度の4段階で測定し全被験歯に対する平均値を算出した。

4) Community Periodontal Index (CPI)

1982年に Ainamo ら⁸⁾が WHO の提案として発表した地域歯周疾患指数 CPI にて歯周組織の評価を行った. 口腔内を 6 群に分割し, それぞれの分画の代表歯を被験歯として評価した.

2. マルファン症候群の表現型と歯周病所見との関

マルファン症候群の診断基準である Ghent 基準の 各臓器所見, すなわち骨格系, 眼, 心血管系, 肺, 皮膚, 硬膜のどの表現型, あるいは表現型の合計数と歯 周病罹患状態との間に相関があるか否かを検証した.

3. 統計解析

統計処理は、一元配置分散分析を用いた(SPSS11.0 J for Windows, SPSS Japan. 東京). 特に指示がなければ p 値が 0.05 未満のものを有意とし、全体として有意差を認めたものは post hoc 解析を追加した.

■ 結果

1. 歯周病所見

20 名の現在歯数の平均は 27 歯であった. PD は 2.815±0.624 mm, BOP は 11.567±8.394%, 動揺度 はすべての症例において 0 であった. さらに CPI code は, CPI code0 の者が 0 名, code1 もしくは

code2 の者(歯肉炎)5名(25%), code3 の者(軽~中等度歯周炎)が12名(60%), code4 の者(中~重度歯周炎)が3名(15%)であった。すなわち4 mm 以上の歯周ポケットを有する者(CPI=3または4の者)は15名(75%)と非常に高頻度であった(CPI 2.70 ± 0.98)(図1)

このことは平成 17 年歯科疾患実態調査による報告における 30~35歳(CPI:1.73±1.05), 35~40歳(CPI:1.76±1.09)の年齢層(本研究の対象集団の平均年齢は35歳)と比較して統計的に明らかな有意差をもってCPI が高値を示している。一元配置分散分析にて3 群を比較すると p 値 = 0.001, post hoc 解析(Scheffe 法)にてわれわれの症例と 30~35歳, 35~40歳の一般集団と比較して p<0.001と有意に CPI の値が高値であることが示された。

2. 表現型

20 名にみられた表現型は心血管系が 20 名(100%) と全症例に認められた. 次いで皮膚が 12 名(60%), 眼が 11 名(55%)となった(表 1).

3. 表現型と歯周病所見との関係

表現型の数と PD の比較を行った結果,表現型 3 つでは PD が 2.808 mm,表現型 4 つでは PD が 2.819 mm,表現型 5 つでは PD が 2.822 mm と表現型が多くなるにつれて PD は深くなる傾向であったが,両者の間に有意差は認めなかった。さらに表現型の数と BOP の比較を行った結果,表現型 3 つでは BOP が 12.81%,表現型 4 つでは BOP が 10.98%,表現型 5 つでは BOP が 10.23% と表現型と BOP の間に有意な差はみられなかった。

考察

マルファン症候群は5,000人~10,000人に1人の確率で発症するといわれている⁹. 特徴的な表現型として,クモ状指,側弯症,後弯症,胸郭変形,バルサルバ洞を含め大動脈弁逆流,大動脈解離,水晶体亜脱臼,硬膜拡張などが挙げられる. 本症例でもバルサルバ洞を含む上行大動脈の拡大は全症例においてみられた. また約半分に眼症状がみられた.

治療にあたってはβ遮断薬,アンジオテンシンⅡ受容体拮抗薬による血圧のコントロール,運動制限,妊娠出産時の厳格な管理,大動脈径の定期的な評価と人工血管置換術などが挙げられる。このように多臓器に表現型を呈する全身疾患であり、集学的な検査および治療体制が必要とされる。そのため当院では、診療科の枠を越えて循環器内科、心臓外科、小児科、整形外科、眼科、放射線科、臨床ゲノム情報部・診療部がチーム体制を作り、マルファン症候群専門外来を開設して対応している100.

歯科的な特徴として,下顎後退症,高口蓋,口蓋垂裂,口蓋正中部の偏位,舌の奇形,歯列不正,歯の先 天欠如,形態異常や形成不全などが挙げられる.

近年, マルファン症候群に有意に歯周病罹患率が高 いことが指摘されている5.しかしマルファン症候群 の口腔内所見の報告は少なく、特に国内において歯周 病罹患状態に関した報告は皆無に近いのが現状であ る. 今回の結果, 本症例では PD が 4 mm 以上の部位 を有する者(CPI=3 または4の者)は15例(75%)で あった. これは平成17年歯科疾患実態調査11)による と 35~39 歳で 23.7% であり、全国調査に比較して非 常に高いことが明らかになった。また、今回の結果で は CPI の最も多い値は CPI が 3 であったのに対し、 平成 17 年歯科疾患実態調査によると CPI2 が最も多 く,マルファン症候群は歯周炎が重度の傾向を示し た. このように高頻度に認められる歯周病は、あわせ て存在する心臓弁膜疾患(大動脈弁閉鎖不全, 僧帽弁 逸脱症など)において口腔内細菌を起因菌とする感染 性心内膜炎の発症母地となり得るとともに、最近では このような口腔内の慢性炎症によって大動脈解離や拡 大といった血管病変の進行に寄与する可能性も十分に 考えられる.

マルファン症候群の原因として 1991 年に 15q21.1 に座位を有する FBN1 遺伝子が発見された $^{12.13}$. その後 2004 年には TGFBR2 遺伝子 14 , さらには TGFBR1 が新たにマルファン症候群の原因遺伝子として特定され、最近ではフィブリリン異常と $TGF-\beta$ シグナルとの関連性がマルファン症候群の病態生理に

表 1 本症例における各表現型

	大基準	小基準	合計
骨格系症状	3 例	2 例	5 例(25%)
眼症状	11 例	0	11 例(55%)
心血管系症状	19 例	1例	20 例 (100%)
肺症状	_	4 例	4例(20%)
皮膚症状	_	12 例	12例 (60%)
硬膜拡張	8 例		8例 (40%)

重要であることが明らかになりつつある。FBNI遺伝 子は全身の結合組織の構成要素となる主要蛋白のフィ ブリリンをコードする. フィブリリンは歯周組織の歯 根膜にも存在する. 歯周組織は歯の支持組織で, セメ ント質、歯根膜、歯槽骨、歯肉の一部によって構成さ れている. 特に歯根膜は特殊化した線維性結合組織で あり、フィブリリンを主成分とする微細線維が集まっ て構成されたオキシタラン線維から成る. オキシタラ ン線維は歯根膜以外にも血管外膜、神経上皮、神経周 膜、腱などほとんどの結合組織に存在する。 歯根膜で のオキシタラン線維は、 歯根を歯軸方向に三次元的に 囲み、しばしば血管やリンパ管の複合体に終わるか近 接している.機能は脈管周囲や圧力のかかる部分に分 布していることから, 脈管の機械的支持と血流調整作 用が考えられている。また、歯の萌出方向をガイドし ているという報告もある。 よってフィブリリンの異 常は、オキシタラン線維の正常な働きを阻害する、す なわち、FBNI 遺伝子の異常は歯根膜の機能異常を来 している可能性があり、マルファン症候群における歯 周病の重症度と関係があるかもしれない.また, TGFBR1 および2遺伝子はTGF-BI またはⅡ型受容 体をコードしており、この異常は結合組織の脆弱性を 引き起こすといわれているが、歯周組織との関連性は 不明である.

マルファン症候群の表現型である眼症状や心血管系 異常と歯周病との関連に関する報告は検索する限りで はみられず、本研究でも明確な示唆は得られなかった が、今後さらに症例を重ねることで他臓器の表現型と の関連性について検証が可能と考える.

マルファン症候群は突然死の恐れのある予後不良な病気と認識されていたが、最近の治療成績の向上およびマルファン症候群の早期診断により予後は改善している。これは一方ではマルファン症候群の長期生存を意味し、今後ますますこれらの患者が歯科を受診する機会が増加することが予想される。よってマルファン症候群の口腔症状を理解し、歯周病のマネージメントを行うことは患者のQOLの維持の点からも急務であ

る.

圖 結 語

本研究から、マルファン症候群の患者は中等度から 重度の歯周病に罹患している確率が非常に高く、歯周 組織の脆弱性が示唆された。

文 献

- Marfan AB: A case of congenital deformation of the four limbs-especially fingers and toes-characterized by long bones (in French). Bull Mem Soc Med Hop Paris 13: 220-226, 1986
- Beighton P, De Paepe A, Danks D, et al: International nosology of heritable disorders of connective tissue. Am J Med Genet 29: 581-594, 1988
- 3) De Coster PJA, Martens LCM, De Paepe A: Oral manifestations of patients with Marfan syndrome: a case-control study. Oral Med Oral Pathol Oral Radiol Endod 93: 564-572, 2002
- De Paepe A, Devereux RB, Dietz HC, et al: Revised diagnostic criteria for the Marfan syndrome. Am J Med Genetics 62:417-426, 1996
- Straub AM, Grahame R, Scully C, Tonetti MS: Severe periodontitis in Marfan's syndrome: a case report. J Periodontol 73: 823-826, 2002
- Akutsu K, Morisaki H, Takeshita S, et al: Characteristics in phenotypic manifestations of genetically proved Marfan syndrome in a Japanease population. Am J Cardiol 103: 1146-1148, 2009
- 7) Miller SC: Textbook of Periodontia. 3rd ed. Blakiston Co Inc, Philadelphia, pp 125-212, 1950
- 8) Ainamo J, Barmes D, Beagrie G, et al: Development of the World Health Organization (WHO) community periodontal index of treatment needs (CPITN). Int Dent J 32:281-291, 1982
- Gray JR: Ascertainment and severity of Marfan syndrome in Scottish population. J Med Genet 31:51-54, 1994
- 10) 今井 靖, 小川直美, 西村敬史, 他:東京大学医学部 附属病院におけるマルファン症候群専門外来:包括的 な診療体制の実践 呼と循 57:1099-1103, 2009
- 11) 歯科疾患実態調査報告解析検討委員会編:平成17年 歯科疾患実態調査,口腔保健協会,東京,2007
- 12) Lee B. Godfrey M. Vitale E. et al: Linkage of Marfan syndrome and a phenotypically related disorder to two different fibrillin genes. Nature 352: 330-334, 1991

- 13) Dietz HC, Cutting G, Pyeritz R, et al: Marfan syndrome caused by a recurrent de novo missense mutation in the fibrillin gene. Nature 352:337-339, 1991
- 14) Mizuguchi T. Collod-Beroud G, Akiyama T, et al: Heterozugous TGFBR2 mutations in Marfan syndrome. Nat Genet 004:36:855-860
- 15) 矢嶋俊彦,敦賀英知,入江一元:歯周組織の弾性線維、日歯周誌46:175-184,2004

Summary

The High Prevalence of Periodontitis in Patients with Marfan Syndrome

by

Mieko Aoki¹, Yasushi Imai, Daishi Fujita, Naomi Ogawa, Masayoshi Kato, Hiroshi Nishimura, Jun-ichi Suzuki, Yasunobu Hirata, Ryozo Nagai

from

1 Department of Cardiovascular Medicine, University of Tokyo Hospital

Marfan syndrome is a connective tissue disorder with autosomal dominant inheritance.

The disease affects mainly the skeletal, cardiovascular, and ocular systems. Patients with this syndrome often demonstrate oral and maxillofacial manifestations including highly arched palate with crowding of teeth. In order to evaluate the clinical characteristics in Japanese Marfan syndrome patients, we evaluated the periodontal status of those patients who were diagnosed as Marfan syndrome according to the Ghent nosology (n = 20). The results showed that the number of teeth present was 27. Probing pocket depth were 2.815±0.624 mm, bleeding on probing $11.567 \pm 8.394\%$, and percentages of CPI (community periodontal index) codes 3 or 4 75%. Our results demonstrate the significantly high prevalence of severe periodontitis in patients with Marfan syndrome. The connective tissue disorder in Marfan syndrome may also increase susceptibitity to inflammatory breakdown of periodontal tissue.

Key words Marfan syndrome, periodontitis, CPI

Case Reports

Diagnostic Efficacy of Coronary CT Angiography as a Follow-up Modality for Procedure-Related Coronary Dissection

Eriko Hasumi, MD, Hiroshi Iwata, MD, Kan Saito, MD, Katsuhito Fujiu, MD, Jiro Ando, MD, Yasushi Imai, MD, Hideo Fujita, MD, Yasunobu Hirata, MD, and Ryozo Nagai, MD

SUMMARY

Procedure-related coronary dissection is associated with an increased risk of major adverse cardiovascular events after percutaneous coronary intervention (PCI). In most patients with such an iatrogenic complication, further PCI or bypass surgery aimed at complete revascularization is performed. Moreover, conventional coronary angiography has been used as a standard modality in the follow-up of such patients. The present report describes a 70 year old female patient who was complicated by catheter-related extensive coronary dissection in the right coronary artery (RCA) when treated for an acute myocardial infarction. Although RCA flow was insufficient, we decided against revascularization and followed her medically without additional revascularization procedures. Her clinical course had been uneventful for 4 years. However, symptoms of effort angina developed and re-examinations were performed at approximately 5 years after the myocardial infarction. Although conventional coronary angiography failed to show the culprit lesion responsible for the angina symptoms, the superior spatial resolution of the coronary CT angiography clearly identified significant progression of the stenotic lesion in the true lumen of the dissected RCA. Thus, coronary CT angiography might be considered as a possible first-line follow-up modality in patients with procedure-related coronary dissection. (Int Heart J 2011; 52: 240-242)

Key words: PCI-related coronary dissection, Coronary CT angiography, Evaluation of true lumen, Coronary stenosis

Procedure-related coronary dissection is one of the life-threatening complications of percutaneous coronary intervention (PCI) and it is associated with an increasing risk of adverse outcomes. Most patients complicated by coronary dissection are followed by conventional coronary angiography. However, as coronary CT angiography is less invasive and superior in the visualization of the three-dimensional structure of the complex vasculature in dissected coronary arteries, its use may be appropriate in the follow-up of such patients.

CASE REPORT

A 70 year-old female who had a history of medically treated hypertension and dyslipidemia was admitted to our hospital complaining of worsening chest discomfort on effort. Five years before admission, she was admitted to another hospital due to severe chest pain at rest. She was diagnosed as having acute ST-segment elevation myocardial infarction (STEMI) with ST-segment elevation in leads II, III, and aVF in a 12-lead electrocardiogram and had decreased motion in the inferior wall of the left ventricle in echocardiography. Since severe stenosis (90%) in the proximal portion of the RCA was

revealed by emergent coronary angiography, although there was no significant stenosis in the left coronary artery (LCA), she was moved to subsequent rescue PCI at the same hospital. A 7 French guiding catheter (Judkins-right shape) was engaged in the RCA and a 0.014 inch soft-tip guide wire was used to pass through the culprit lesion that was located at a severe angulation in the proximal portion of the RCA (Figure 1a, arrow). However, soon after starting the procedure, spiral and

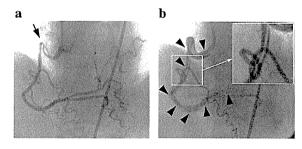


Figure 1. Angiographical findings before (a) and after (b) complicating coronary dissections in RCA. a: severe stenosis in proximal portion of RCA (arrow), b: coronary dissections from the ostium to posterior descending artery of RCA (arrow heads).

From the 1 Department of Cardiovascular Medicine, The University of Tokyo Hospital, Tokyo, Japan.

This study was supported by the Japan Society for the Promotion of Science (JSPS) through its "Funding Program for World-Leading Innovative R&D on Science and Technology (FIRST Program)".

Address for correspondence: Hiroshi Iwata, MD, Department of Cardiovascular Medicine, The University of Tokyo Hospital, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8655, Japan.

Received for publication February 22, 2011.

Revised and accepted April 1, 2011.

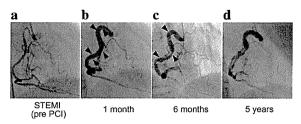


Figure 2. Changes in angiographical findings before (a), at 1 month (b), 6 months (c), and 5 years (d) after complicating coronary dissections with intracoronary tears (arrow heads).

extensive coronary dissection in the RCA emerged (Figure 1b, arrow heads). Intracoronary tears were clearly visualized by coronary angiography. Although much effort was expended to identify the true coronary lumen using guide wires, it was unsuccessful due to severe narrowing in the true lumen induced by compression of the false lumen. Since flow in the RCA was not sufficient (TIMI grade 2) at the end of the procedure, an intra-aortic balloon pumping (IABP) device was placed for 3 days. Thallium stress scintigraphy at chronic phase demonstrated the deterioration in the viability of the posterior wall of the left ventricle. However, as the patient was asymptomatic and her hemodynamics were stable, it was decided that no further procedures would be conducted in consideration of procedural risk. Coronary angiograms at 1 month (Figure 2b) and 6 months (Figure 2c) still demonstrated an intracoronary tear (Figure 2b, 2c, arrow heads), as well as insufficient right coronary flow accompanied by an extremely complex coronary vasculature of the true and false lumens.

However, her clinical course had been generally uneventful and asymptomatic for over 4 years, although coronary computer tomography (CT) angiography performed 4 years later demonstrated the narrowing of the true lumen in the RCA (Figure 3a arrow head). Five years after the onset of STEMI, symptoms of effort angina had gradually developed over a 1 month period. Since she was referred to our hospital, we performed detailed and comprehensive follow-up examinations. Conventional coronary angiography failed to clarify the apparent difference from the angiography findings at 6 months after STEMI with complex three-dimensional structure characterized by vascular screws of the true and false lumens in the RCA (Figure 2c and d). On the contrary, the curved planar reconstruction method of coronary CT clearly demonstrated the stenotic lesion responsible for the symptoms. The cross-sectional view revealed significant progression in narrowing of the true lumen, which was separated by an extensive intimal flap in the RCA (Figure 3, arrows), in comparison with the findings of the CT before development of symptoms. After careful consideration regarding the risks and benefits of RCA revascularization, it was decided the patient would be treated with maximal antianginal agents, such as a nitrate, a beta-blocker, and a potassium channel opener. Consequently, the chest symptoms were brought under control and her clinical course has been uneventful for 3 years without a need for rehospitalization.

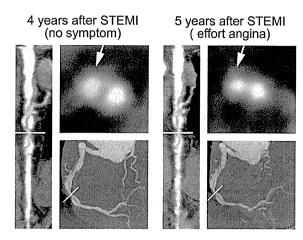


Figure 3. Coronary CT angiography successfully identified the lesion (lines) where the true lumen stenosis of dissected RCA had progressed for a year (arrows).

DISCUSSION

Despite rapid progress in the development of devices and techniques, PCI procedures can still induce life-threatening complications. Coronary dissection induced by PCI is associated with an increased risk of major adverse cardiovascular events. The frequency of PCI-related coronary dissection in the recent drug-eluting stent era has been reported to be 1.2-9.2%.1) Huber, et al described the relationship between morphological complexity in accordance with the classification of the National Heart, Lung, and Blood Institute (NHLBI) and in-hospital adverse outcomes.2) In addition to extensive manipulation of devices or contrast infusion, established risk factors of procedure-related coronary dissection include the use of Amplatz guiding catheters and coronary artery anatomical anomalies.³⁾ The gold standard for the treatment of coronary dissection is to pass a guide wire through the true coronary lumen and to secure coronary flow by expanding that with a balloon followed by complete coverage with a stent(s).4 However, coronary bypass graft surgery should be considered without delay in cases where it is extremely difficult or impossible to pass a guide wire due to severe narrowing or closure of the true lumen and serious consequences caused by residual and ongoing ischemia of the target vessel can be predicted.

In the present case, a procedure-related spiral dissection covering almost the entire length of the RCA was complicated by engaging a guiding catheter for the treatment of STEMI. According to the angiographical classification of procedure-related coronary dissection, the present case was classified into the group in which major adverse events, such as additional revascularization procedures or in-hospital myocardial infarction were found in more than 50% of cases. ²⁾ This case was indeed accompanied by severe stenosis or closure of the true lumen and this resulted in unsuccessful revascularization followed by myocardial infarction. Because the patient was hemodynamically stable with IABP and medications, further revascularization was not performed. At almost 5 years later, symptoms of effort angina gradually developed and reevaluation of the coronary artery was performed. While the extremely