

Figure 1 (a) Abnormal brain vessels in MMD. The dotted circle indicates the X-ray field of cerebral angiography (left panel). Normal structures of the right internal carotid artery (ICA), anterior cerebral artery (ACA) and middle cerebral artery (MCA) are illustrated (middle panel). The arrowheads indicate abnormal collateral vessels appearing like a puff of smoke in the angiogram of an individual with MMD (right panel). Note that ACA and MCA are barely visible, because of the occlusion of the terminal portion of the ICA. (b) Manhattan plot of the 785 720 SNPs used in the genome-wide association analysis of MMD patients. Note that the SNPs in the 17q25-ter region reach a significance of $P < 10^{-8}$.

MATERIALS AND METHODS

Affected individuals

Genomic DNA was extracted from blood and/or saliva samples obtained from members of the families with MMD (Supplementary Figure 1), MMD patients with no family history and control subjects. All of the subjects were Japanese. MMD was diagnosed on the basis of guidelines established by the Research Committee on Spontaneous Occlusion of the Circle of Willis of the Ministry of Health and Welfare of Japan. This study was approved by the Ethics Committee of Tohoku University School of Medicine. Total RNA samples were purified from leukocytes using an RNeasy mini kit (Qiagen, Hilden, Germany) and used as templates for cDNA synthesis with an Oligo (dT)₂₀ primer and SuperScript II reverse transcriptase according to the manufacturer's instructions (Invitrogen, Carlsbad, CA, USA).

Linkage analysis

For the linkage analysis, DNA samples were genotyped for 36 microsatellite markers within five previously reported MMD loci using the ABI 373A DNA Sequencer (Applied Biosystems, Foster City, CA, USA). Pedigrees and haplotypes were constructed with the Cyrillic version 2.1 software (Oxfordshire, UK). Multipoint analyses were conducted using the GENEHUNTER 2 software (http://www.broadinstitute.org/ftp/distribution/software/genehunter/). Statistical analysis was performed with SPSS version 14.0J (SPSS, Tokyo, Japan).

Genome-wide and locus-specific association studies

A genome-wide association study was performed using a group of 72 MMD patients, which consisted of 64 patients without a family history of MMD and 8 probands of MMD families. The Illumina Human Omni-Quad 1 chip (Illumina, San Diego, CA, USA) was used for genotyping, and single-nucleotide polymorphisms (SNPs) with a genotyping completion rate of 100% were used for further statistical analysis (785 720 out of 1 140 419 SNPs). Genotyping data

from 45 healthy Japanese controls were obtained from the database at the International HapMap Project web site. The 785 720 SNPs were statistically analyzed using the PLINK software (http://pngu.mgh.harvard.edu/~purcell/plink/index.shtml). For a locus-specific association study, we used 63 DNA samples consisting of 58 non-familial MMD patients and 5 probands of MMD families. A total of 384 SNPs within chromosome 17q25-ter were genotyped (Supplementary Table 1), using the GoldenGate Assay and a custom SNP chip (Illumina). Genotyping data for 45 healthy Japanese were used as a control. Case—control single-marker analysis, haplotype frequency estimation and significance testing of differences in haplotype frequency were performed using the Haploview version 3.32 program (http://www.broad.mit.edu/mpg/haploview/).

Mutation detection

Mutational analyses of RNF213 and FLJ35220 were performed by PCR amplification of each coding exon and putative promoter regions, followed by direct sequencing. Genomic sequence data for the two genes were obtained from the National Center for Biotechnology Information web site (http://www.ncbi.nlm.nih.gov/) for design of exon-specific PCR primers. RNF213 cDNA fragments were amplified from leukocyte mRNA for sequencing analysis. Sequencing of the PCR products was performed with the ABI BigDye Terminator Cycle Sequencing Reaction Kit using the ABI 310 Genetic Analyzer. Identified base changes were screened in control subjects. Statistical difference of the carrier frequency of each base change was estimated by Fisher's exact test (the MMD group vs the control group).

Quantitative PCR

MTC Multiple Tissue cDNA Panels (Clontech Laboratory, Madison, WI, USA) were the source of cDNAs from human cell lines, adult and fetal tissues. Mononuclear cells and polymorphonuclear cells were isolated from the fresh peripheral blood of healthy human adults using Polymorphprep (Cosmo Bio,

Carlsbad, CA, USA). T and B cells were isolated from the fresh peripheral blood of healthy human adults using the autoMACS separator (Militeny Biotec, Bergisch Gladbach, Germany). Total RNA was isolated from these cells with the RNeasy Mini Kit (Qiagen) following the manufacturer's instructions. We reverse transcribed 100 ng samples of total RNA into cDNAs using the High Capacity cDNA Reverse Transcription Kit (Applied Biosystems). Quantitative PCRs were performed in a final volume of 20 µl using the FastStart TaqMan Probe Master (Rox) (Roche, Madison, WI, USA), 5 µl of cDNA, 10 µm of RNF- or GAPDH-specific primers and 10 µM of probes (Universal ProbeLibrary Probe #80 for RNF213 and Roche Probe #60 for GAPDH). All reactions were performed in triplicate using the ABI 7500 Real-Time PCR system (Applied Biosystems). Cycling conditions were 2 min at 50°C and 10 min at 95°C, followed by 40 cycles of 15 s at 95°C and 60 s at 60°C. Real-time PCR data were analyzed by the SDS version 1.2.1 software (Applied Biosystems). We evaluated the relative level of RNF213 mRNA by determining the C_T value, the PCR cycle at which the reporter fluorescence exceeded the signal baseline. GAPDH mRNA was used as an internal reference for normalization of the quantitative expression values.

Multiplex PCR

MTC Multiple Tissue cDNA Panels (Clontech) were the source of human cell lines and cDNAs from human adult and fetal tissues. Multiplex PCRs were performed in a final volume of $20\,\mu$ l using the Multiplex PCR Master Mix (Qiagen), $2\,\mu$ l of cDNA, a $2\,\mu$ M concentration of RNF213 and a $10\,\mu$ M concentration of GAPDH-specific primers. The samples were separated on a 2% agarose gel stained with ethidium bromide. Cycling conditions were 15 min at 94°C, followed by 30 cycles of 30 s at 94 °C, 30 s at 57 °C and 30 s at 72 °C. For normalization of the expression levels, we used GAPDH as an internal reference for each sample.

In situ hybridization (ISH) analysis

Paraffin-embedded blocks and sections of mouse tissues for ISH were obtained from Genostaff (Tokyo, Japan). The mouse tissues were dissected, fixed with Tissue Fixative (Genostaff), embedded in paraffin by proprietary procedures (Genostaff) and sectioned at 6 µm. To generate anti-sense and sense RNA probes, a 521-bp DNA fragment corresponding to nucleotide positions 470-990 of mouse Rnf213 (BC038025) was subcloned into the pGEM-T Easy vector (Promega, Madison, WI, USA). Hybridization was performed with digoxigenin-labeled RNA probes at concentrations of 300 ng ml⁻¹ in Probe Diluent-1 (Genostaff) at 60 °C for 16h. Coloring reactions were performed with NBT/BCIP solution (Sigma-Aldrich, St Louis, MO, USA). The sections were counterstained with Kernechtrot stain solution (Mutoh, Tokyo, Japan), dehydrated and mounted with Malinol (Mutoh). For observation of Rnf213 expression in activated lymphocytes, 10-week-old Balb/c mice were intraperitoneally injected with 100 µg of keyhole limpet hemocyanin and incomplete adjuvant and sacrificed in 2 weeks. The spleen of the mice was removed for Hematoxylin-eosin staining and ISH analyses.

RESULTS

Using 20 Japanese MMD families, we reevaluated the linkage mapped previously to five putative MMD loci. No locus with significant linkage, Lod score >3.0 or NPL score >4.0 was confirmed (Supplementary Figure 2). We conducted a genome-wide association study of 72 Japanese MMD cases. Single-marker allelic tests comparing the 72 MMD cases and 45 controls were performed for 785720 SNPs using χ^2 statistics. These tests identified a single locus with a strong association with MMD ($P < 10^{-8}$) on chromosome 17q25-ter (Figure 1b), which is in line with the latest mapping data of a MMD locus. ¹⁶ The SNP markers with $P < 10^{-6}$ are listed in Table 1. To confirm this observation, we performed a locus-specific association study. A total of 384 SNP markers (Supplementary Table 1) were selected within the chromosome 17q25-ter region and genotyped in a set of 63 MMD cases and 45 controls. The SNP markers demonstrating a high association with MMD ($P < 10^{-6}$) were clustered in a 151-kb region from base position 75 851 399-76 003 020 (SNP No.116-136 in

Table 1 A genome-wide association study of Japanese MMD patients and controls

					Risk allele/	Risk allele	Risk allele				95% confide	95% confidence interval
	SNP	Chromosome	Base position	Gene	non-risk allele	frequency in MMD	frequency in controls	κ^2	P-value	Odds ratio	Lower	Upper
7	rs11870849	17	76025668	RNF213	1/C	0.4792	0.1111	33.55	6 95F-09	7 36	3 530	16.24
2	rs6565681	17	75963089	RNF213	A/G	0.7361	0.3667	31.35	2.35E 03	4.819	2.002	13.34
ന	rs7216493	17	75941953	RNF213	G/A	0.75	0.3889	30,39	3.53E-08	4 715	2,733	0.400 c1c a
4	rs7217421	17	75850055	RNF213	A/G	0.6667	0.3	29.86	4 64E-08	4.666	2,642	0.010
2	rs12449863	17	75857806	RNF213	C/T	0.6667	0.3	29.86	4.64F-08	4.666	2,0,2	0.63/
9	rs4890009	17	75926103	RNF213	G/A	0.8819	0.5778	28.5	9 38F-08	5.459	2,042	0.507
7	SNP17-75933731	17	75933731	RNF213	G/A	0.8819	0.5778	28.5	9.38F-08	5.455 7.458	2.831	10.327
_∞	rs7219131	17	75867365	RNF213	1/C	0.6667	0.3111	28.11	1.15E-07	0:438	2,631	7707
6	rs6565677	17	75932037	RNF213	T/C	0.7431	0.3977	27.43	1.63F-07	4378	2.017	7.7.54
10	rs4889848	17	75 969 256	RNF213	C/T	0.75	0.4111	26.99	2.05E-07	4.297	2,473	7 990
11	rs7224239	17	75969771	RNF213	A/G	0.8681	0.5667	26.99	2.05E-07	5.03	2.659	9.529
Athron	Abbraulations, MAND manage disease CND single	- II- Olyo										

testing 1140419 SNPs on the Human Omni-Quad 1chip (Illumina, San Diego, CA, USA) was performed in 72 Japanese MMD cases. Ists the 11 SNP markers with a significance of $P<10^{-6}$. study disease; SNP, single-nucleotide polymorphism.A genome-wide association ϵ cases and controls were performed using χ^2 statistics for all markers. This ' the MMD, tests t iations: allelic t

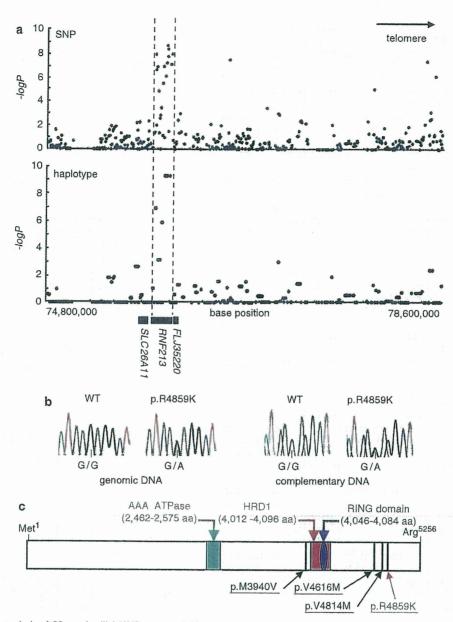


Figure 2 (a) Association analysis of 63 non-familial MMD cases and 45 control subjects. Statistical significance was evaluated by the χ^2 -test. SNP markers with a strong association with MMD ($P < 10^{-6}$) clustered in a 161-kb region (base position 75851399–76012838) indicated by two dotted lines (upper panel), which included the entire region of RNF213 (lower panel). Haplotype analysis revealed a strong association ($P = 5.3 \times 10^{-10}$) between MMD and a single haplotype located within RNF213. (b) Sequencing chromatograms of the identified MMD mutations. The left panel shows the sequences of an unaffected individual and a carrier of a p.R4859K heterozygous mutation. The right panel indicates the sequencing chromatograms of the leukocyte cDNA obtained from an unaffected individual and an individual with MMD who carries the p.R4859K mutation. Note that both wild-type and mutant alleles were expressed in leukocytes. (c) The structure of the RNF213 protein. The RNF213 protein contains three characteristic structures, the AAA-superfamily ATPase motif, the RING motif and the HMG-CoA reductase degradation motif. The positions of four mutations identified in MMD patients are underlined, including one prevalent mutation (red) and three private mutations (black).

Supplementary Table 1); this entire region was within the *RNF213* locus (Figure 2a). A single haplotype determined by seven SNPs (SNP Nos.130–136 in Supplementary Table 1) that resided in the 3′ region of *RNF213* was strongly associated with MMD onset ($P=5.3\times10^{-10}$). Analysis of the linkage disequilibrium block indicated that this haplotype was not in complete linkage disequilibrium with any other haplotype in this region (Supplementary Figure 3). These results strongly suggest that a founder mutation may exist in the 3′ part of *RNF213*.

Mutational analysis of the entire coding and promoter regions of RNF213 and FLJ35220, a gene 3' adjacent to RNF213, revealed that 19 of the 20 MMD families shared the same single base substitution, c.14576G>A, in exon 60 of RNF213 (Figure 2b and Table 2). This nucleotide change causes an amino-acid substitution from arginine⁴⁸⁵⁹ to lysine⁴⁸⁵⁹ (p.R4859K). The p.R4859K mutation was identified in 46 of 63 non-familial MMD cases (73%), including 45 heterozygotes and a single homozygote (Table 3). Both the wild-type and the p.R4859K mutant alleles were co-expressed in leukocytes



Table 2 Nucleotide changes with amino-acid substitutions identified in the sequencing analysis of RNF213 and FLJ35220

			Genotype	e (allele)			
Gene	Ехоп	Nucleotide change³ (amìno-acid substitution)	Non-familial cases	Control subjects	P <i>-value</i> ^b	χ² (df=1) °	Odds ratio (95% CI)
RNF213	29	c.7809C>A (p.D2603E)	2/63 (2/126)	15/381 (15/762)	0.77	0.09	0.80 (0.2–3.6)
RNF213	41	c.11818A>G (p.M3940 V)	1/63 (1/126)	0/388 (0/776)	0.01	6.17	ND
RNF213	41	c.11891A>G (p.E3964G)	4/63 (4/126)	3/55 (4/110)	0.84	0.04	1.2 (0.3–5.5)
RNF213	52	c.13342G>A (p.A4448T)	4/63 (4/126)	2/53 (2/106)	0.53	0.39	1.7 (0.3–9.8)
RNF213	56	c.13846G>A (p.V4616 M)	1/63 (1/126)	0/388 (0/776)	0.01	6.17	1.7 (0.9–3.8) ND
RNF213	59	c.14440G>A (p.V4814 M)	1/63 (1/126)	0/388 (0/776)	0.01	6.17	ND
RNF213	60	c.14576G>A (p.R4859 K)	46/63 (47/126)	6/429 (6/858)	1.2×10 ⁻⁴³	298.1	190.8 (71.7–507.9)
FLJ35220		None		11 122 (0,000)	1,2,10	250.1	190.6 (/1./-507.9)

Abbreviations: ND, not determined; SNP, single-nucleotide polymorphism

Table 3 Association of the p.R4859K (c.14576G > A) mutation with MMD

			Genotype	
	Total	wt/wt (%)	wt/p.R4859K (%)	p.R4859K/ p.R4859 K (%) ^d
Members of 19 N	MMD famil	iesª		
Affected	42	0	39 (92.9)	3 (7.1)
Not affected	28	15 (53.6)	13 (46.4)	0
Individuals withou	ut a family	history of MMD	ı,c	
Affected	63	17 (27.0)	45 (71.4)	1 (1.6)
Not affected	429	423 (98.6)	. 6 (1.4)	0

in three patients heterozygous for the p.R4859K mutation (Figure 2b), excluding the possible instability of the mutant RNF213 mRNA. Additional missense mutations, p.M3940V, p.V4616M and p.V4814M, were detected in three non-familial MMD cases without the p.R4859K mutation (Figure 2c). These mutations were not found in 388 control subjects and were detected in only one patient, suggesting that they were private mutations (Table 2). No copy number variation or mutation was identified in the RNF213 locus of 12 MMD patients using comparative genome hybridization microarray analysis (Supplementary Figure 4). In total, 6 of the 429 control subjects (1.4%) were found to be heterozygous carriers of p.R4859K. Therefore, we concluded that the p.R4859K mutation increases the risk of MMD by a remarkably high amount (odds ratio=190.8 (95% confidence interval=71.7-507.9), $P=1.2\times10^{-43}$) (Table 3). It was recently reported that an SNP (ss161110142) in the promoter region of RPTOR, which is located ~150 kb downstream from RNF213, was associated with MMD. 17 Genotyping of the SNP in RPTOR showed that the RNF213 p.R4859K mutation was more strongly associated with MMD than ss161110142 (Supplementary Figure 1).

RNF213 encodes a protein with 5256 amino acids harboring a RING (really interesting new gene) finger motif, suggesting that it

functions as an E3 ubiquitin ligase (Figure 2c). It also has an AAA ATPase domain, which is characteristic of energy-dependent unfoldases. 18 To our knowledge, RNF213 is the first RING finger protein known to contain an AAA ATPase domain. The expression profile of RNF213 has not been previously fully characterized. We performed a quantitative reverse transcription PCR analysis in various human tissues and cells. RNF213 mRNA was highly expressed in immune tissues, such as spleen and leukocytes (Figure 3a and Supplementary Figure 5). Expression of RNF213 was detected in fractions of both polymorphonuclear cells and mononuclear cells and was found in both B and T cell fractions (Supplementary Figure 6). A low but significant expression of RNF213 was also observed in human umbilical vein endothelial cells and human pulmonary artery smooth muscle cells. Cellular expression was not enhanced in tumor cell lines, compared with leukocytes. In human fetal tissues, the highest expression was observed in leukocytes and the thymus (Supplementary Figure 6E). The expression of RNF213 was surprisingly low in both adult and fetal brains. Overall, RNF213 was ubiquitously expressed, and the highest expression was observed in immune tissues.

We studied the cellular expression of Rnf213 in mice. The ISH analysis of spleen showed that Rnf213 mRNA was present in small mononuclear cells, which were mainly localized in the white pulps (Figures 3b-g). The ISH signals were also detected in the primary follicles in the lymph node and in thymocytes in the medulla of the thymus (Supplementary Figure 7). To study Rnf213 expression in activated lymphocytes we immunized mice with keyhole limpet hemocyanin, and examined Rnf213 mRNA in spleen by ISH analysis. Primary immunization with keyhole limpet hemocyanin antigen revealed that the expression of Rnf213 in the secondary follicle is as high as in the primary follicle in the lymph node (Supplementary Figure 8). In an E16.5 mouse embryo, expression was observed in the medulla of the thymus and in the cells around the mucous palatine glands (Supplementary Figure 9). These findings suggest that mature lymphocytes in a static state express Rnf213 mRNA at a higher level than do their immature counterparts.

DISCUSSION

We identified a susceptibility locus for MMD by genome-wide and locus-specific association studies. Further sequencing analysis revealed a founder missense mutation in RNF213, p.R4859K, which was tightly associated with MMD onset. Identification of a founder mutation in individuals with MMD would resolve the following recurrent

Nucleotide numbers of RNF213 cDNA are counted from the A of the ATG initiator methionine codon (NCBI Reference sequence, NP_065965.4).

bP-values were calculated by Fisher's exact test.

*Genotypic distribution (carrier of the polymorphism vs non-carrier).

Abbreviations: MMD, moyamoya disease. ^aEntire distribution, χ^2 =29.4, P=4.2×10⁻⁷. ^bEntire distribution, χ^2 =298.2, P=1.8×10⁻⁶⁵.

^cGenotypic distribution (p.R4859K carrier vs non-carrier), χ^2 =298.1, P=1.2×10⁻⁴³, odds

ratio=190.8 (95% Cl=71.7–507.9).

The age of onset and initial symptoms of the four homozygotes were comparable to those of

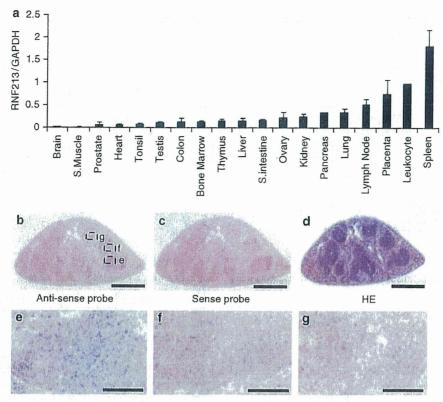


Figure 3 Expression of human RNF213 and murine Rnf213. (a) RT-PCR analysis of RNF213 mRNA in various human tissues. The expression levels of *RNF213* mRNA in various adult human tissues were evaluated by quantitative PCR using *GAPDH* mRNA as a control. The signal ratio of *RNF213* mRNA to *GAPDH* mRNA in each sample is shown on the vertical axis. (b–g) *In situ* hybridization (ISH) analysis of Rnf213 mRNA in mouse spleen. Specific signals for *Rnf213* mRNA were detected by ISH analysis with the anti-sense probe (b) but not with the sense probe (c). Hematoxylin–eosin staining of the mouse spleen (d). Signals for the *Rnf213* mRNA were observed in small mononuclear cells, which were mainly localized in the white pulps (dotted square, e) and partially distributed in the red pulps (dotted squares, f and g). Panels e, f and g show the high-magnification images of the corresponding fields in panel b. Scale bars, 1 mm (b–d) and 50 μm (e–g).

questions:2,19 (i) why is MMD more prevalent in East Asia than in Western countries? The carrier frequency of p.R4859K in Japan is 1/72 (Table 2). In contrast, we found no p.R4859K carrier in 400 Caucasian controls (data not shown). Furthermore, no mutation was identified in five Caucasian patients with MMD after the full sequencing of RNF213. These results suggest that the genetic background of MMD in Asian populations is distinct from that in Western populations and that the low incidence of MMD in Western countries may be attributable to a lack of the founder RNF213 mutation. (ii) Is unilateral involvement a subtype of MMD or a different disease?² We collected DNA samples from six patients with unilateral involvement and found a p.R4859K mutation in four of them (data not shown), suggesting that bilateral and unilateral MMD share a genetic background. (iii) Is pre-symptomatic diagnosis of MMD possible? In the present study, MMD never developed in the 15 mutation-negative family members in the 19 MMD families with the p.R4859K mutation (Table 3 and Supplementary Figure 1), suggesting the feasibility of presymptomatic diagnosis or exclusion by genetic testing.

How the mutant RNF213 protein causes MMD remains to be elucidated. The expression of *RNF213* was more abundant in a subset of leukocytes than in the brain, suggesting that blood cells have a function in the etiology of MMD. This observation agrees with a previous report that MMD patients have systemic angiopathy.²⁰

Recent studies have suggested that the postnatal vasculature can form through vasculogenesis, a process by which endothelial progenitor cell are recruited from the splenic pool and differentiate into mature endothelial cells. Levels of endothelial progenitor cells in the peripheral blood are increased in MMD patients. RNF213 may be expressed in splenic endothelial progenitor cells and mutant RNF213 might dysregulate the function of the endothelial progenitor cells. Further research is necessary to elucidate the role of RNF213 in the etiology of MMD.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGEMENTS

We thank all of the patients and their families for participating in this study. We also thank Dr Hidetoshi Ikeda at the Department of Neurosurgery, Tohoku University School of Medicine and Drs Toshiaki Hayashi and Reizo Shirane at the Department of Neurosurgery, Miyagi Children's Hospital, Sendai, Japan for patient recruitment. We are grateful to Ms Kumi Kato for technical assistance. This study was supported by grants from the Ministry of Education, Culture, Sports, Science and Technology, Japan and by the Research Committee on Moyamoya Disease of the Ministry of Health, Labor and Welfare, Japan.



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Supplementary Information accompanies the paper on Journal of Human Genetics website (http://www.nature.com/jhg)

Supplementary Table 1. Genotyping of 384 SNPs within chromosome 17q25-ter in MMD patients.

No	SNP Name	Position	minor allele frequency	-logP (SNP)	-logP (Haplotype)***	gene
1	rs897595	74814739	0.23	0.54	0.60	
2	rs4790005	74820377	0.48	0.61	0.60	
3	rs4790007	74825355	0.49	0.56	0.56	
4	rs1869932	74830160	0.27	0.68	0.56	
5	rs897597	74833704	0.34	0.24		
6	rs4789887	74838408	0.35	0.16		
7	rs4790013	74846331	0.40	0.30		
8	rs8075376	74862253	0.16	0.18		
9	rs897600	74878104	0.38	1.59		
10	rs751848	74885572	0.35	0.24		
11	rs2034860	74891235	0.19	0.03		
12	rs9912528	74899145	0.18	0.21	0.98	
13	rs7225663	74902740	0.25	0.88		
14	rs897587	74907220	0.14	0.29	0.98	
15	rs2377405	74915689				
16			0.27	0.23 **		
	rs872016	74923774	0.19			
17	rs971626	74928535	0.48	0.71		
18	rs1007464	74932660	0.43	0.41		
19	rs4790037	74935348	0.42	1.07		
20	rs2137774	74947906	0.10	0.05		
21	rs884025	74965704	0.50	0.09		
22	rs871739	74973490	0.21	0.36		
23	rs7213580	74979416	n/a*	**		
24	rs4790051	74991920	n/a*	**		
25	rs211788	74995852	0.40	0.03		
26	rs9902874	75006934	n/a*	**		
27	rs11868921	75019989	n/a*	**		
28	rs12451031	75020299	n/a*	**		
29	rs7216806	75050112	0.46	0.66		
30	rs3935352	75257677	n/a*	**		
31	rs4074469			**		
		75269306	n/a*	**		
32	rs7208711	75286482	n/a*			
33	rs4555183	75294840	0.22	0.06		
34	rs6565697	75304120	n/a*	**		
35	rs8072313	75313636	n/a*	**		
36	rs8072274	75316196	0.46	0.06		
37	rs6565475	75316941	n/a*	**		
38	rs8074728	75319640	0.43	**		
39	rs11657217	75323934	0.21	0.80		
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41	rs8076446	75338179	0.28	0.79		
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45	rs6565539	75363556	0.06	0.11	0.00	
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53	rs1285251	75424418	0.32	**		
54	rs2289728	75426449	0.37	1.92	1.82	
55	rs3764374	75429891	0.18	0.32	-	
56	rs1622986	75438157	0.22	0.36		
57	rs1696756	75442568	0.31	1.48	1.48	
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59	rs8078624	75459394			1.48	
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62	rs4889796	75471233	0.28	1.87	1.87	
63	rs1285260	75477911	0.41	0.01		
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65	rs3843732	75486138	0.41	0.33		
66	rs4493093	75489976	0.29	0.15		
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	rs1285285	75496304	() 4×			
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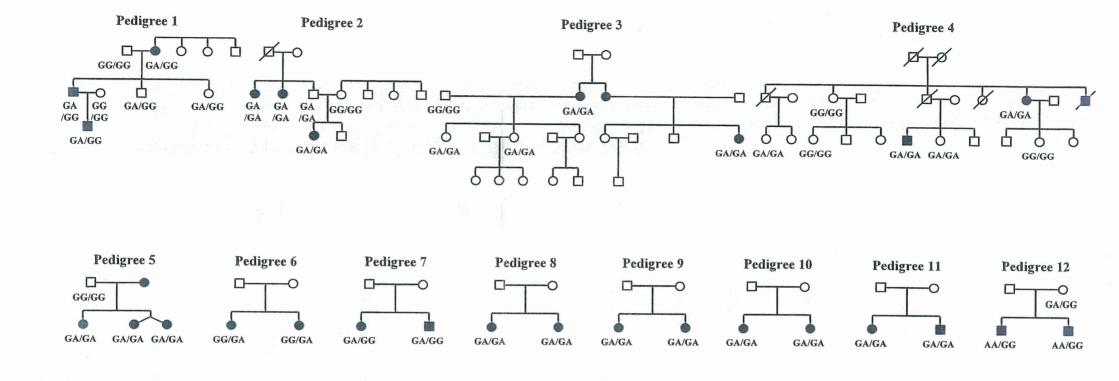
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74	rs1285293	75539371		0.06		
75	rs7210391	75547666	0.13	0.75		
76	rs4889940	75553402	0.48	0.96		
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78	гѕ935200	75563386	0.39	**		
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79	rs1115834	75564609	0.43			
80	rs4441315	75568048	0.49	0.93		
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82	rs11150827	75582679	0.28	0.06	0.01	
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86	rs2289529	75636840	0.15	0.38	• 0.34	
87	rs2289531	75638317	0.14	0.35	0.34	
88	rs2289533	75638689	0.28	0.20	0.5 .	
89	rs9319623	75669201	0.42	0.81		
90	rs715041	75673027	0.26	2.17	2.64	
91	rs1561811	75675223	0.26	0.73	2.64	
92	rs4889954	75680189	0.25	0.00	2.64	
93	rs2361701	75681212	0.30	1.54	2.64	
94	rs2304854	75688157	0.49	0.42	2.64	
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96	rs1800307	75700466	0.22	2.23	2.64	
97	rs2304836	75701441	0.23	**		
98	rs8132	75707948	0.15	0.64		
99	rs7211079	75722132	0.50	0.45	0.38	
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119	rs7219131	75867365	0.30	6.82	6.93	within RNF213
120	rs11869363	75881354	0.49	4.79		within RNF213
121	rs9905727	75887409	0.15	1.34	3.15	within RNF213
122	rs8066993	75894625	0.40	2.54	3.15	within RNF213
123	rs8081176	75898582	0.41	2.77	3.15	within RNF213
124	rs9674807	75900772	0.42	**		within RNF213
125	rs7501761	75904780	0.47	3.40	3.15	within RNF213
126	rs4890008	75920214	0.05	1.11		within RNF213
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127	rs8074015	75920875	0.41	5.41	5.89	within RNF213
128	rs4890010	75930774	0.38	6.82	5.89	within RNF213
129	rs11150856	75937477	0.07	0.93		within RNF213
130	rs8067292	75948435	0.31	6.01	9.28	within RNF213
131	rs8070106	75959041	0.34	7.12	9.28	within RNF213
132	rs6565681	75963089	0.36	8.56	9.28	within RNF213
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136	rs4603608	76003020	0.35	7.00	9.28	
137	rs4077240	76012838	0.38	7.88		within FLJ35220
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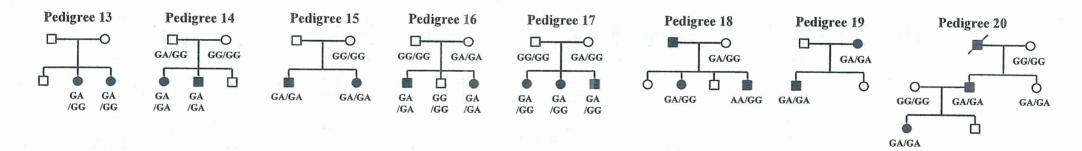
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154	rs1485330	76255417	0.31	0.10	0.76
155	rs7217223	76260258	0.28	0.61	0.76
156	rs4255830	76263825	0.40		
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	rs4889875	76266095	0.40	0.85	0.76
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160	rs6565478	76306599	0.36	0.98	0.98
161	rs4969230	76340856	0.47	0.37	
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164	rs4969429	76379814	0.17	2.04	
165	rs734338	76396935	0.39	0.20	
166	rs2048753	76403883	0.34	0.11	0.82
167	rs2589133	76408071	0.40	0.01	0.82
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195	rs1399571	76567065	0.23	0.10	0.20
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208	rs11657991	76644343	n/a*	**	
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217	rs11664	76697457	0.43	1.10	1.22
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220	rs9900420	76725990 76733197	0.15	0.50	1.54
221 222	rs4969259 rs4969405	76737591	0.42 0.49	1.67 0.93	1.54 1.54
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224	rs7209950	76747354	0.26	0.03	0.88
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246 247	rs899288 rs7216513	77001454 77010284	0.14 0.13	0.32 0.17	
248	rs12601728	77010284	n/a*	U.I./ **	
249	rs4969441	77021631	0.27	0.15	
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256	rs8079717	77083169	0.26	0.19	0.28
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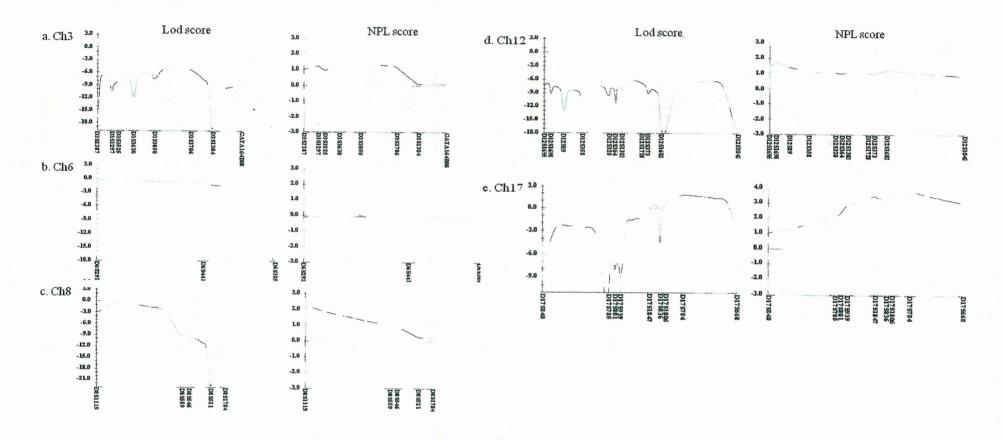
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	309	rs11654140	77925540	n/a*	**	
	310	rs3935179	77935545	0.48	**	
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	313 314	rs7211306	77960642	0.07	0.04	
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	356 357	rs629246	78350827	0.41	0.74	0.68
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358	rs3744165	78383731	0.09	0.02	0.68	
359	rs7225515	78389072	0.44	0.62	0.68	
360	rs7219521	78395034	0.49	0.41	0.68	
361	rs4986117	78402394	0.23	0.90	0.68	
362	rs6502007	78412566	0.33	0.85	0.80	
363	rs733342	78415369	0.23	1.05	0.80	
364	rs8067926	78431877	0.32	2.08		
365	rs4986129	78446843	0.03	7.28		
366	rs1551625	78470842	0.23	1.22		
367	rs1078334	78477536	0.43	2.46		
368	rs3785512	78479817	0.14	0.47	2.38	
369	rs898095	78483927	0.36	3.10	2.38	
370	rs3785521	78489058	0.27	2.55	2.38	
371	rs1001865	78508277	0.23	0.71	2.38	
372	rs9303016	78517996	0.17	1.13		
373	rs7209936	78519504	0.47	1.21		
374	rs1551628	78528901	0.33	6.03		
375	rs6502033	78541807	0.49	0.73		
376	rs12601298	78552287	0.27	0.04		
377	rs9893868	78564747	0.38	**		
378	rs7222550	78578613	0.23	0.79		
3 7 9	rs4986140	78583049	0.46	1.56		
380	rs9890099	78587132	0.43	0.51		
381	rs967825	78593728	0.48	0.18	1.22	
382	rs7224733	78598059	0.23	1.22	1.22	
383	rs6502040	78605474	0.31	0.44	1.22	
384	rs3935099	78609338	0.32	1.30		



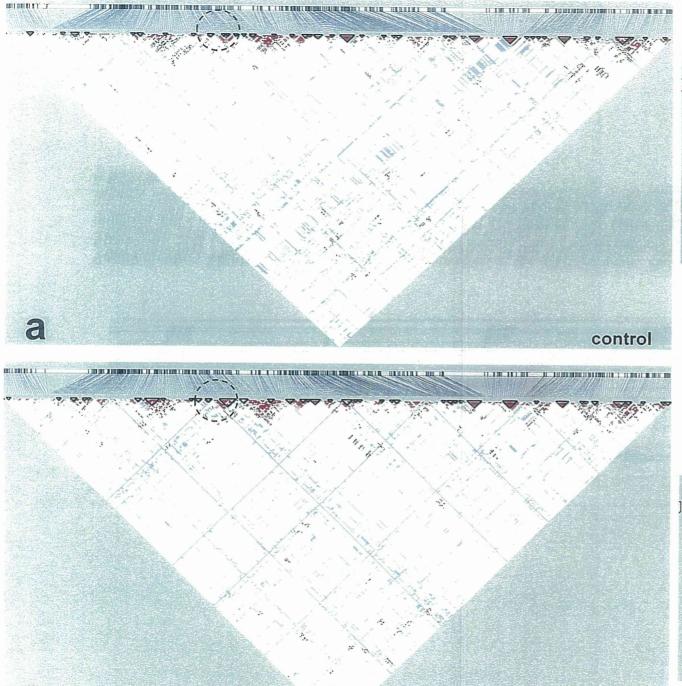


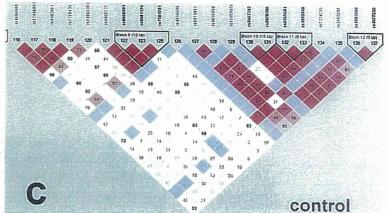
Supplementary Figure 1. MMD families and genotypes of c.14719G>A (p.R4859K) mutation in RNF213 and ss161110142 at position -1490 from the transcription start site in RPTOR. All MMD pedigrees but pedigree 6 had the p.R4859K mutant allele. Note that MMD never developed in family members without the p.R4859K mutant allele in p.R4859K mutation-positive family. A homozygote of p.R4859K mutation was found in families 12 and 18.



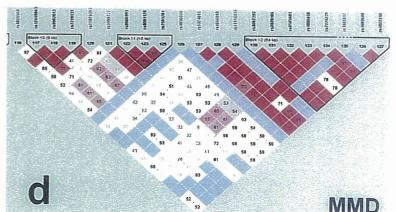
Supplementary Figure 2. Linkage analysis of 20 Japanese MMD families

Twenty MMD families were studied by genotyping of the microsatellite markers, which were previously used for identification of five candidate chromosome loci for MMD genes. Highest linkage score was observed at the microsatellite maker D17S784 on chromosome 17q25, the Lod score 2.4 and the NPL score 3.8, which was suggestive but not definitive.

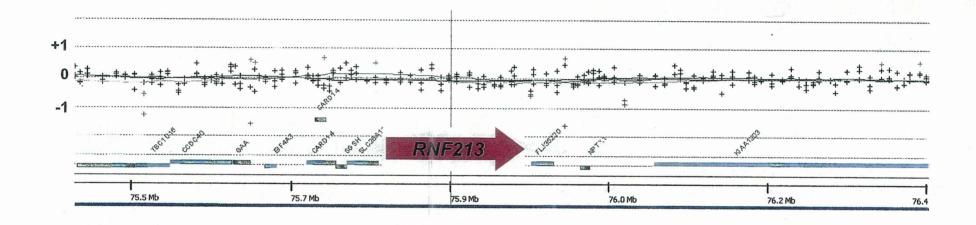




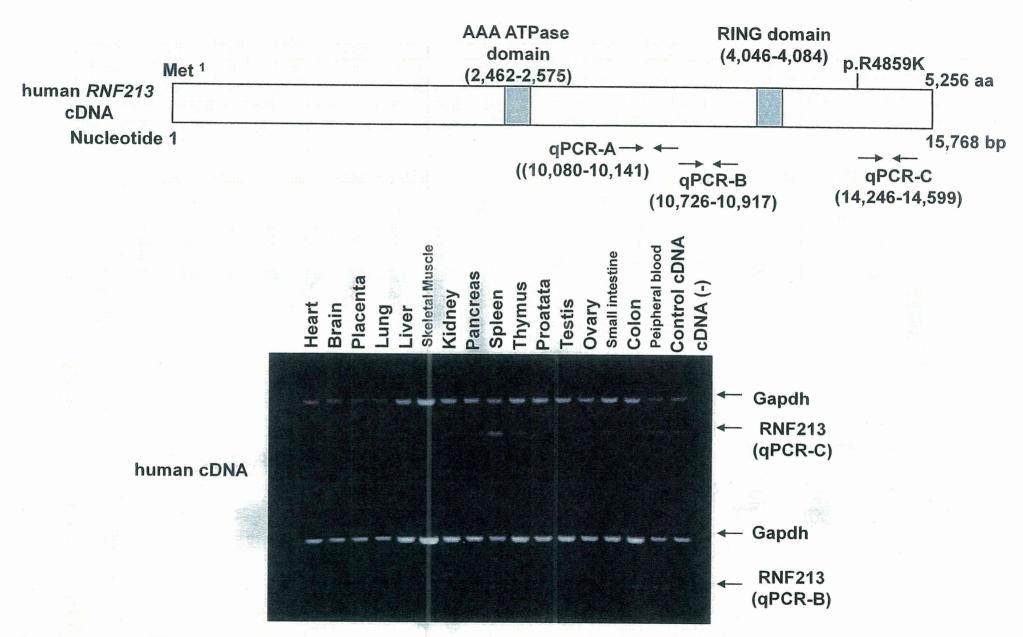
Supplementary Figure 3. Structure of the linkage disequilibrium (LD) block in Japanese control subjects (a) and patients with MMD (b). Dotted circles in panels a and b indicate the regions of the higher magnification images in panels c and d, respectively.



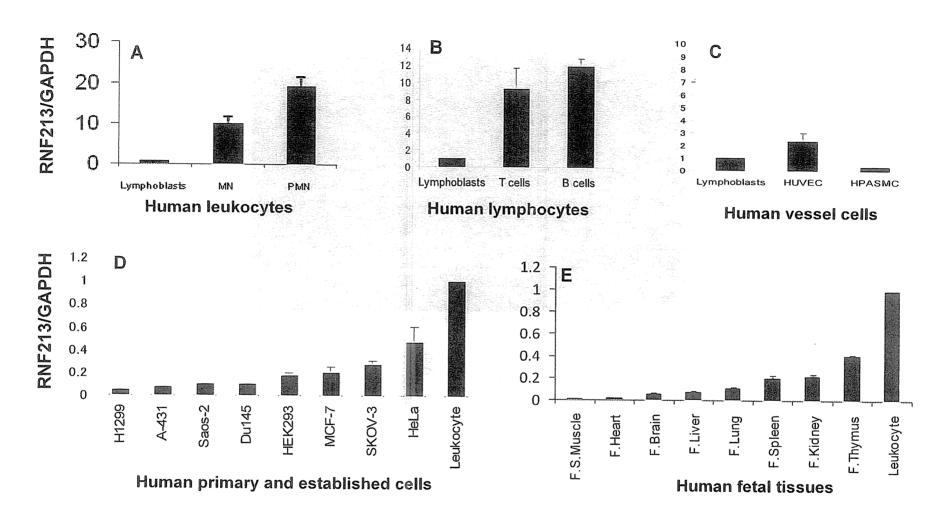
MMD



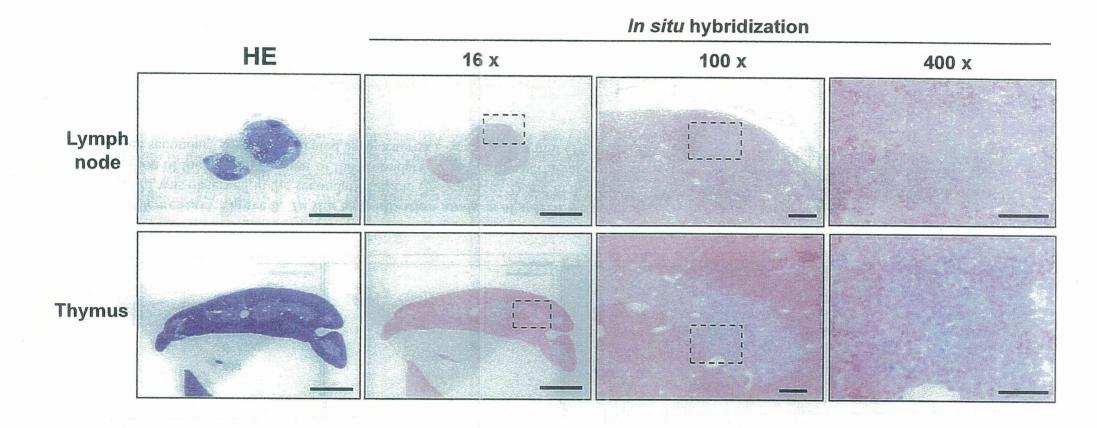
Supplementary Figure 4. The CGH microarray analysis of the RNF213 locus. The array CGH analysis with Human Genome Microarray 244k (Agilent Technology, Germany) was performed in 12 MMD patients, which included 8 patients with p.R4859K mutation and 4 patients without the mutation. The vertical axis indicates \log_2 (ratio of patient/control signal intensity). No copy number variation or mutation within the RNF213 locus was detected in any of the patients. Results of three patients with p.R4859K mutation were shown in the panel.



Supplementary Figure 5. Schematic illustration of RNF213 cDNA, including the location of primers used in RT-PCR analysis. Multiplex PCR analysis which amplify 5' part and 3' part of the RNF213 cDNA was performed. Relatively higher expression was observed in spleen, peripheral blood and thymus. Result of qPCR-A was shown in Fig.3a. Relative expression pattern was similar among PCR product from qPCR-A, qPCR-B and qPCR-C sets.

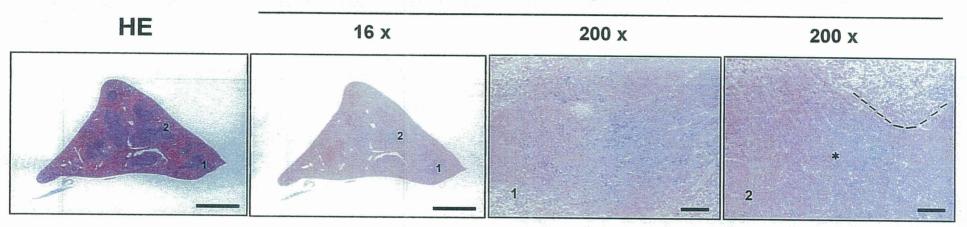


Supplementary Figure 6. RT-PCR analysis of RNF213 in various human cells and fetal tissues. Expression levels of RNF213 mRNA was evaluated by the quantitative PCR using GAPDH RNA as the control in human leukocytes (A), human lymphocytes (B), human vessel cells (C), human primary and established cells (D) and human fetal tissues (E). The signal ratio of RNF213 mRNA and GAPDH RNA in each sample is shown in the vertical axis. MN, mononuclear cells; PMN, polymorphonuclear cells; HUVEC, Human Umbilical Vein Endothelial Cells; HPASMC, Human Pulmonary Artery Smooth Muscle Cells.



Supplementary Figure 7. In situ hybridization analysis of lymph node and thymus in mice. Small round cells expressing Rnf213 mRNA was observed in the primary follicles in the lymph node. Specific signals for Rnf213 mRNA was also detected in thymocytes in the medulla of the thymus. Dotted squares indicate the fields of the higher magnification images. HE stands for hematoxylin-eosin staining. Scale bars, 1 mm (16x), 100 μ m (100x), 50 μ m (400x).

In situ hybridization



Supplementary Figure 8. In situ hybridization analysis of spleen from adult mice immunized with KLH. The expression of Rnf213 was observed in the secondary follicle (indicated as 2) and in the primary follicle (indicated as 1). Dotted lines indicated the location of the germinal center of the secondary follicles. The germinal center was represented less signal intensity than the other region of the secondary follicle (indicated as an asterisk). Scale bars, 1 mm (16x), 50 µm (200x).